sirDAG:

Surrogate Intervention Recovery of a Directed Acyclic Graph (DAG)

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1 Overview

This vignette describes how to use R/sirDAG package to estimate high-dimensional directed acyclic graphs (DAGs) for gene expression with surrogate intervention data such as cis-eQTL SNP genotype data. We first estimate a skeleton from gene expression data using PenPC algorithm and then orient the edges in the skeleton using sirDAG algorithm. For the whole procedure we need to download three packages.

```
> library(PEN) # Installed from PEN_1.01.tar.gz
> library(PenPC) # Installed from PenPC_1.0.tar.gz
> library(sirDAG)
```

2 Example

We illustrate the usage of sirDAG package using simulated data under the scenario of p = 100, n = 30, ER model for the true DAG with pE = 2/(p-1) and multiple eQTLs per gene (total no. of eQTLs across all genes are 200, m = 200).

```
> load("simul_p100n30r27pE2_ER_M.Rdata")
> names(simul)
```

- [1] "A"
- [2] "B"
- [3] "Y"
- [4] "X"
- [5] "G"
- [6] "Gy"
- [7] "cross"
- [8] "markers.nms"
- [9] "allqtls"
- [10] "pheno.names"

The list simul includes objects:

- $p \times p$ matrix A for the gene-gene coefficients
- $p \times m$ matrix B for the SNP-gene coefficients
- $n \times p$ matrix Y for gene expression data
- $n \times m$ matrix X for SNP genotype data

- graphNEL object G that includes the augmented DAG for gene expression and eQTL
- graphNEL object Gy that includes the DAG for gene expression only
- cross, markers.nms, and allqtls are used for simulation studies for QDG algorithm (Neto et. al., 2008)

Before fitting DAG, we scale the gene expression and genotype data.

```
> n=30
> p=100
> datX = simul$X
> datY = simul$Y
> meandat = apply(datX,2,mean)
> normdat = sqrt(rowSums((t(datX)-meandat)^2)/n)
> datX= scale(datX,meandat,normdat)
> meandat = apply(datY,2,mean)
> normdat = sqrt(rowSums((t(datY)-meandat)^2)/n)
> datY= scale(datY,meandat,normdat)
```

2.1 Skeleton Estimation

We estimate the skeleton of gene expression using PenPC algorithm. In the first step of PenPC, neighborhood selections for all nodes are performed to uncover Markov blanket.

```
> coef = ne.PEN(dat=datY,nlambda=100,ntau=10,V=1:p,order=TRUE,verbose=FALSE)
> # Set edge weight
> edgeWeights = matrix(apply(abs(cbind(c(coef),c(t(coef)))),1,max),ncol=p)
Then we perform partial correlation tests for the p-value cutoff α = 0.01 to eliminate the false positive edges.
```

2.2 Edge orientation

We orient the undirected edges in the skeleton using sirDAG-MAP or sirDAG-NG.

By default, we perform sirDAG-MAP. If we set NormalGamma=T, sirDAG-NG is performed with predetermined set of hypoerparameters of the prior, nu, delta, and gg, where gg is set to be the sample size n for the unit g-prior.

```
> fit.sirDAG_NG = directionPosterior(gInput=gSkel,B=simul$B,datX=datX,datY=datY
+ ,exhaustive.cut=10,no.T=2^10,verbose=F,acyclic=T
+ ,bf.cut1=1,bf.cut2=1,Cluster=T
+ ,NormalGamma=T,nu=1,delta=1,gg=n)
```

The output values includes posterior for posterior probabilities for each edge directions, g for the reuslting DAG (or PDAG), OK is to check whether the edge orientation is performed.

```
> names(fit.sirDAG_MAP)
```

```
[1] "posterior" "g"
```

[3] "OK"

> names(fit.sirDAG_NG)

```
[1] "posterior" "g"
```

[3] "OK"

We can disply the networks by the plot function using igraph package.

