# Golden Gate Assembly

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#### 1 Introduction

Pending...

#### 2 Protocol

### 2.1 Reaction System

As an example, here, we use BsaI (type IIs restriction enzyme) in this reaction system. Below is an typical reaction system which is 20  $\mu$ L.

Components	Volume
Fragments	$40~\mathrm{fmol}$ or $100~\mathrm{ng}$ ea. $^{(1)}$
BsaI	10 U (2 $\mu$ L)
T4 ligase	$5~{\rm U}~(0.5~\mu L)^{~(2)}$
$10 \times$ T4 Ligase Buffer	$2~\mu { m L}$
$ddH_2O$	Up to 20 $\mu L$

Table 1: Golden Gate reaction system

#### Note:

- (1) When do  $1 \sim 2$  insertions reaction, it is convenient adding 100 ng of each fragment into reaction. If performing multiple (> 3 insertions) ligation, we recommend adding 40 fmol of each fragment.
- (2) If assembling more than 3 fragments, high concentration of T4 ligase is necessary. I recommend 20 U (for NEB CEU, 1 U = 200 CEU).
- (3) For convenience, we often prepare 10  $\mu$ L reactions and the volume of each enzyme is 1  $\mu$ L.

#### 2.2 Ligation Procedure

If performing one fragment insertion ligation, incubating reaction system 30 min at 37 °C. When insertions are more than 2, we recommend using thermocycler: 37 °C for 2 min, the 16 °C for 3 min, repeat for 10 - 100 cycles, depending on the number of insertions (10 cycles per insertion). Following, the reaction is heated to 50 °C for 30 min, 30 min at 37 °C , which is for digesting non-specific ligation events, completely. Last, the reaction is incubated 5 min at 80 °C for inactivating the reaction enzyme.

(Optionally, 50  $\mu$ L system containing 1  $\mu$ L T4 DNA ligase (2, 000, 000 U/ $\mu$ L), 2  $\mu$ L Type IIs restriction enzyme and totaling  $\sim 1\mu$ g DNA, is recommended. After reaction, 1  $\mu$ L additional restriction enzyme is added, and the assembly is incubated for 1 h at 27 °C. Assembly is then purified for electroporation.) The 10  $\mu$ L reaction is transformed to 50  $\mu$ L chemically competent cell. Then stalled cell in sawed ice for 30 min, and allowed to recover for an hour in SOC medium.

# 3 Design

## 3.1 Primer Design

The flank sequences of primer for introducing enzymatic cutting site into double ends of insertions are collected in Table 2.

Enzyme Name	Sequence
BsaI BbsI	GCATTA <u>GGTCTC</u> CNNNN AAGTGC <u>GAAGAC</u> CANNNN
BfuAI	GGCAAT <u>ACCTGC</u> GTGANN NN
BtgZI	CGAATG <u>GCGATG</u> TTG TACTGCCNNNN
SapI BsmBI	GGAATC <u>GCTCTTC</u> CNNN ATAGCG <u>CGTCTC</u> CNNNN

Table 2: Flank Sequence of Primer, the underlined nucleotides are denoted as the enzyme recognition sequence, and the sequence before recognition sequence is protection sequence, and the base N is denote as the sticky end.