

The Elizabeth H.
and James S. McDonnell III

**MCDONNELL
GENOME INSTITUTE**

at Washington University



Washington
University in St. Louis

SCHOOL OF MEDICINE

PMBIO Module 03

Align. Alignment algorithms,
visualization, and QC

Malachi Griffith, Obi Griffith, Zachary Skidmore, Huiming Xia
Introduction to bioinformatics for DNA and RNA sequence
analysis (IBDR01)

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Glasgow



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Learning objectives of module 03: Align

- **Key concepts:** Sequence alignment algorithms, BAM files, genome viewers, alignment quality assessment
- Compare and contrast DNA vs. RNA sequence alignment strategies
- Perform alignment of sequence data using a few popular alignment algorithms
- Explore the BAM file format and learn approaches for summarizing, filtering, and otherwise manipulating BAM files
- Learn to use a genome viewer
- Perform a quality assessment using the aligned data

Alignment algorithms...

SAM/BAM/CRAM files represent sequence alignments

- The specification
 - <http://samtools.sourceforge.net/SAM1.pdf>
- The SAM format consists of two sections:
 - Header section
 - Used to describe source of data, reference sequence, method of alignment, etc.
 - Alignment section
 - Used to describe the read, quality of the read, and nature alignment of the read to a region of the genome
- BAM/CRAM are compressed versions of SAM.
 - BAM compressed using lossless BGZF format
 - CRAM compressed further using knowledge of reference. May or may not be lossless
- BAM/CRAM files are usually ‘indexed’
 - A ‘.bai’ file will be found beside the ‘.bam’ file
 - Indexing aims to achieve fast retrieval of alignments

A BAM file is divided in header and alignment sections

Example SAM/BAM header section (abbreviated)

```
mrgiffitt@linux270 ~$ samtools view -H /gscmnt/gc13001/info/model_data/2891632684/build136494552/alignments/136080019.bam | grep -P "SN:22|HD|RG|PG"
```

```
@HD      VN:1.4      SO:coordinate  
@SQ      SN:22      LN:51304566      UR:ftp://ftp.ncbi.nih.gov/genbank/genomes/Eukaryotes/vertebrates_mammals/Homo_sapiens/GRCh37/special_requests/GRCh37-lite.fa.gz      AS:GRCh37-lite      M5:a718acaa6135fdca8357d5bfe94211dd      SP:Homo sapiens  
@RG      ID:2888721359      PL:illumina      PU:D1BA4ACXX.3      LB:H_KA-452198-0817007-cDNA-3-lib1      PI:365      DS:paired end      DT:2012-10-03T19:00:00-0500      SM:H_KA-452198-0817007      CN:WUGSC  
@PG      ID:2888721359      VN:2.0.8      CL:tophat --library-type fr-secondstrand --bowtie-version=2.1.0  
@PG      ID:MarkDuplicates      PN:MarkDuplicates      PP:2888721359      WN:1.85(exported)      CL:net.sf.picard.sam.MarkDuplicates INPUT=[/gscmnt/gc13001/info/build_merged_alignments/merged-alignment-blade10-2-5.gsc.wustl.edu-jwalker-15434-136080019/scratch-ILG6Y/H_KA-452198-0817007-cDNA-3-lib1-2888360300.bam] OUTPUT=/gscmnt/gc13001/info/build_merged_alignments/merged-alignment-blade10-2-5.gsc.wustl.edu-jwalker-15434-136080019/scratch-ILG6Y/H_KA-452198-0817007-cDNA-3-lib1-2888360300-post_dup.bam METRICS_FILE=/gscmnt/gc13001/info/build_merged_alignments/merged-alignment-blade10-2-5.gsc.wustl.edu-jwalker-15434-136080019/staging-ljUJS/H_KA-452198-0817007-cDNA-3-lib1-2888360300.metrics REMOVE_DUPLICATES=false ASSUME_SORTED=true MAX_FILE_HANDLES_FOR_READ_ENDS_MAP=95000 TMP_DIR=[/gscmnt/gc13001/info/build_merged_alignments/merged-alignment-blade10-2-5.gsc.wustl.edu-jwalker-15434-136080019/scratch-ILG6Y] VALIDATION_STRINGENCY=SILENT MAX_RECORDS_IN_RAM=500000 PROGRAM_RECORD_ID=MarkDuplicates PROGRAM_GROUP_NAME=Mark Duplicates MAX_SEQUENCES_FOR_DISK_READ_ENDS_MAP=50000 SORTING_COLLECTION_SIZE_RATIO=0.25 READ_NAME_REGEX=[a-zA-Z0-9]+:(0-9)+:[(0-9)+]:[(0-9)+]:[(0-9)+]:[(0-9)+]* OPTICAL_DUPLICATE_PIXEL_DISTANCE=100 VERBOSITY=INFO QUIET=false COMPRESSION_LEVEL=5 CREATE_INDEX=false CREATE_MD5_FILE=false
```

```
mrgiffitt@linux270 ~$
```

Example SAM/BAM alignment section (only 10 alignments shown)

[illegible]

BAM header section provides general information about alignment strategy

- Used to describe source of data, reference sequence, method of alignment, etc.
- Each section begins with character '@' followed by a two-letter record type code. These are followed by two-letter tags and values
 - @HD The header line
 - VN: format version
 - SO: Sorting order of alignments
 - @SQ Reference sequence dictionary
 - SN: reference sequence name
 - LN: reference sequence length
 - SP: species
 - @RG Read group
 - ID: read group identifier
 - CN: name of sequencing center
 - SM: sample name
 - @PG Program
 - PN: program name
 - VN: program version

BAM alignment section provides details for each read alignment

Col	Field	Type	Regex/Range	Brief description
1	QNAME	String	[!-?A-~]{1,255}	Query template NAME
★ 2	FLAG	Int	[0,2 ¹⁶ -1]	bitwise FLAG
3	RNAME	String	* [!-()+-<>-~] [!-~]*	Reference sequence NAME
4	POS	Int	[0,2 ²⁹ -1]	1-based leftmost mapping POSition
5	MAPQ	Int	[0,2 ⁸ -1]	MAPping Quality
★ 6	CIGAR	String	* ([0-9]+[MIDNSHPX=])+	CIGAR string
7	RNEXT	String	* = [!-()+-<>-~] [!-~]*	Ref. name of the mate/next segment
8	PNEXT	Int	[0,2 ²⁹ -1]	Position of the mate/next segment
9	TLEN	Int	[-2 ²⁹ +1,2 ²⁹ -1]	observed Template LENgth
10	SEQ	String	* [A-Za-z=.]+	segment SEQUENCE
11	QUAL	String	[!-~]+	ASCII of Phred-scaled base QUALity+33

1	QNAME	e.g.	HWI-ST495_129147882:1:2302:10269:12362 (QNAME)
2	FLAG	e.g.	99
3	RNAME	e.g.	1
4	POS	e.g.	11623
5	MAPQ	e.g.	3
6	CIGAR	e.g.	100M
7	RNEXT	e.g.	=
8	PNEXT	e.g.	11740
9	TLEN	e.g.	217
10	SEQ	e.g.	CCTGTTTCTCCACAAAGTGTTTACTTTTGGATTTTTGCCAGTCTAACAGGTGAAGCCTGGAGATTCTTATTAGTGATTGGGCTGGGCCTGGCCATGT
11	QUAL	e.g.	CCCCFFFFHHHHHJJJIFIJJJJJJJJJJJHIJJJJJJJJJJJGGHHIHJIJJJJJJJJJGHGGIJJJJJIIJEEHHHHFFFCDCDDDDDDDB@ACDD

BAM flags describe several alignment properties in a single number

- <http://broadinstitute.github.io/picard/explain-flags.html>
- 12 bitwise flags describing the alignment
- These flags are stored as a binary string of length 11 instead of 11 columns of data
- Value of '1' indicates the flag is set. e.g. 00100000000
- All combinations can be represented as a number from 1 to 2048 (i.e. $2^{11}-1$). This number is used in the BAM/SAM file. You can specify 'required' or 'filter' flags in samtools view using the '-f' and '-F' options respectively

Bit	Description
1	0x1 template having multiple segments in sequencing
2	0x2 each segment properly aligned according to the aligner
4	0x4 segment unmapped
8	0x8 next segment in the template unmapped
16	0x10 SEQ being reverse complemented
32	0x20 SEQ of the next segment in the template being reverse complemented
64	0x40 the first segment in the template
128	0x80 the last segment in the template
256	0x100 secondary alignment
512	0x200 not passing filters, such as platform/vendor quality controls
1024	0x400 PCR or optical duplicate
2048	0x800 supplementary alignment


Note that to maximize confusion, each bit is described in the SAM specification using its hexadecimal representation (i.e., '0x10' = 16 and '0x40' = 64).

CIGAR strings similarly describe the entire alignment in as few characters as possible

Op	BAM	Description
M	0	alignment match (can be a sequence match or mismatch)
I	1	insertion to the reference
D	2	deletion from the reference
N	3	skipped region from the reference
S	4	soft clipping (clipped sequences present in SEQ)
H	5	hard clipping (clipped sequences NOT present in SEQ)
P	6	padding (silent deletion from padded reference)
=	7	sequence match
X	8	sequence mismatch

- The CIGAR string is a sequence of base lengths and associated ‘operations’ that are used to indicate which bases align to the reference (either a match or mismatch), are deleted, are inserted, represent introns, etc.
- e.g. 81M859N19M
 - A 100 bp read consists of: 81 bases of alignment to reference, 859 bases skipped (an intron), 19 bases of alignment

Genome browsers - Ensembl

 [BLAST/BLAT](#) | [BioMart](#) | [Tools](#) | [Downloads](#) | [Help & Documentation](#) | [Blog](#) | [Mirrors](#)

Search: for


Go


e.g. [BRCA2](#) or [rat 5:62797383-63627669](#) or [rs699](#) or [coronary heart disease](#)


Browse a Genome

Ensembl is a genome browser for vertebrate genomes that supports research in comparative genomics, evolution, sequence variation and transcriptional regulation. Ensembl annotate genes, computes multiple alignments, predicts regulatory function and collects disease data. Ensembl tools include BLAST, BLAT, BioMart and the Variant Effect Predictor (VEP) for all supported species.

Favourite genomes

 **Human**
GRCh38.p10

 **Mouse**
GRCm38.p5

 **Zebrafish**
GRCz10


[Edit favourites](#)

All genomes

-- Select a species --

[View full list of all Ensembl species](#)


Find a Data Display



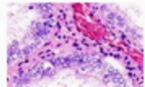
Not sure how to find the data visualisation you need? With our new [Find a Data Display](#) page, you can choose a gene, region or variant and then browse a selection of relevant visualisations

Try it now!

Variant Effect Predictor




Gene expression in different tissues



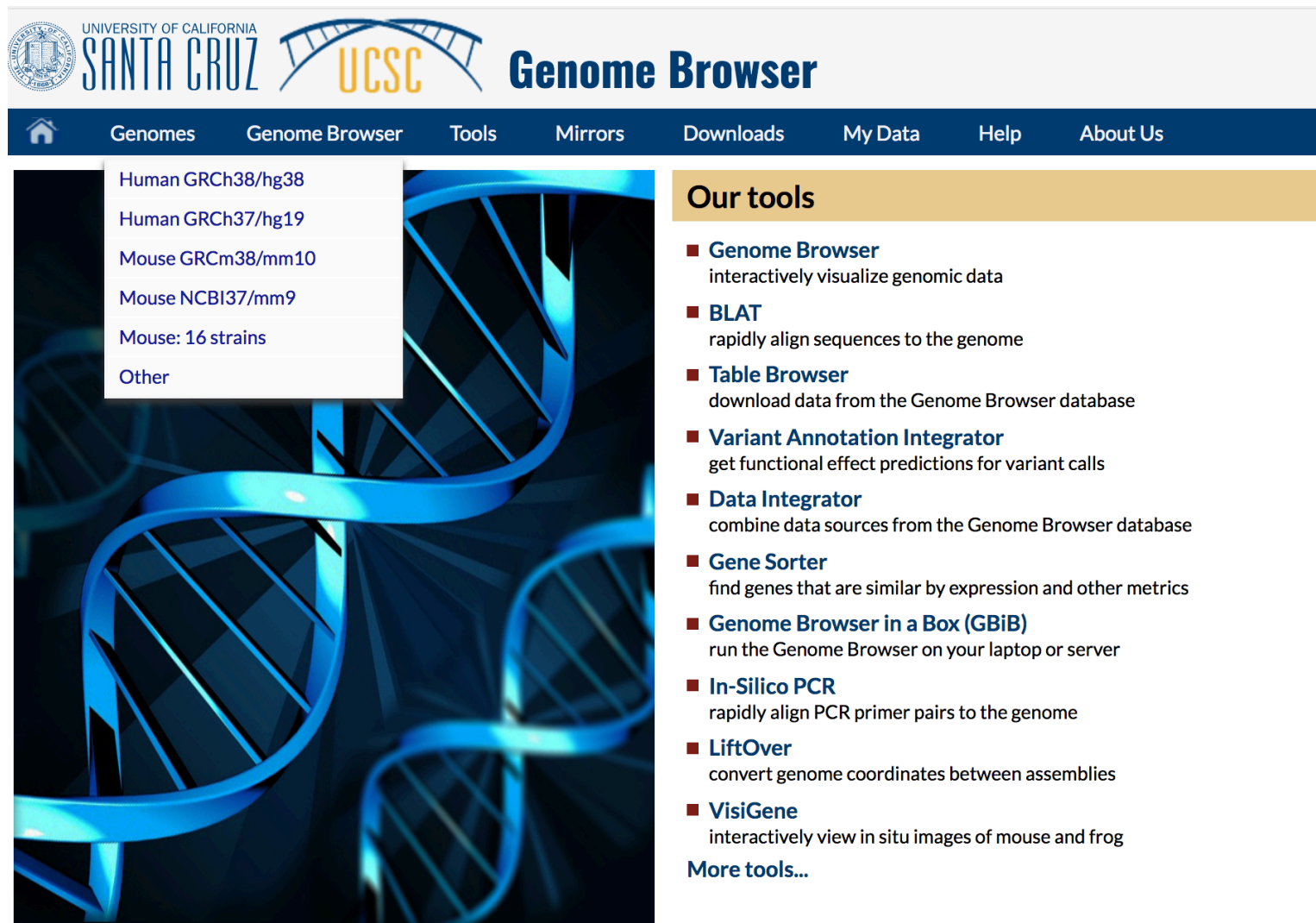
Retrieve gene sequence

```
GCCTGACTTCGGGTGG:
GGGCTTGTGGCGGAGC
GCGCCTCTGCTGCGCCT
AGGGGACAGATTTGTGA
CACCTCTGGAGCGGTT
CCCACTCCAGCGTGGCG
```

Compare genes across species



Genome browsers - UCSC



The screenshot shows the UCSC Genome Browser homepage. At the top is the University of California Santa Cruz logo and the UCSC Genome Browser title. Below is a navigation bar with links: Home, Genomes, Genome Browser, Tools, Mirrors, Downloads, My Data, Help, and About Us. A dropdown menu is open under 'Genomes', listing: Human GRCh38/hg38, Human GRCh37/hg19, Mouse GRCm38/mm10, Mouse NCBI37/mm9, Mouse: 16 strains, and Other. To the right, a section titled 'Our tools' lists various genomic tools with brief descriptions.

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Genome Browser

Home Genomes Genome Browser Tools Mirrors Downloads My Data Help About Us

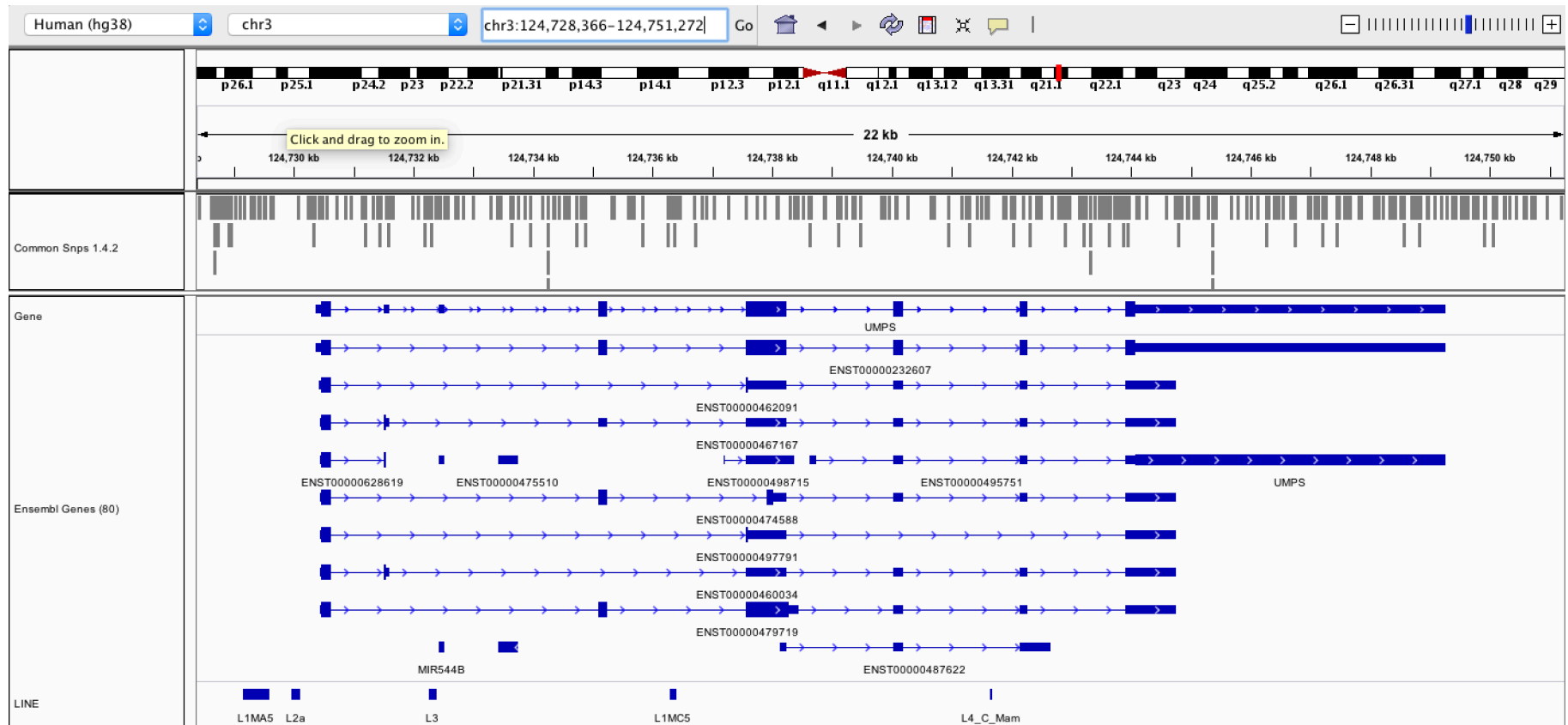
Human GRCh38/hg38
Human GRCh37/hg19
Mouse GRCm38/mm10
Mouse NCBI37/mm9
Mouse: 16 strains
Other

Our tools

- **Genome Browser**
interactively visualize genomic data
- **BLAT**
rapidly align sequences to the genome
- **Table Browser**
download data from the Genome Browser database
- **Variant Annotation Integrator**
get functional effect predictions for variant calls
- **Data Integrator**
combine data sources from the Genome Browser database
- **Gene Sorter**
find genes that are similar by expression and other metrics
- **Genome Browser in a Box (GBiB)**
run the Genome Browser on your laptop or server
- **In-Silico PCR**
rapidly align PCR primer pairs to the genome
- **LiftOver**
convert genome coordinates between assemblies
- **VisiGene**
interactively view in situ images of mouse and frog

More tools...

Genome browsers - IGV



Alignment QC ...