

Code exploration

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Abstract

Related to paper “Beyond p-values: A phase II dual-criterion design with statistical significance and clinical relevance” [Roychoudhury et al. \(2018\)](#).

1 Introduction

Proof-of-concept (POC) in Phase II trials is important in investigating the efficacy of an experimental drug. It will influence the decision of whether continuing or not continuing the development of this drug.

Dual-criterion design in frequentist and Bayesian applications are discussed.

Three generic phase II designs are reviewed:

1. Standard design

For comparative treatment and control trials, it puts forward criteria expressed as error rates:

Type I error control and Power (correctly reject H_0 when it is false).

Control type I error and maximize power.

Type I error: $\mathbb{P}(\text{reject } H_0 | H_0 \text{ is true}) = \alpha$

Type II error: $\mathbb{P}(\text{not reject } H_0 | H_0 \text{ is false}) = \beta$.

Limitation: statistical significant only guarantees evidence to reject “No effect”, but is not sufficient for clinical perspective. Also, it always result in success or failure according to statistical significance.

Increasing the sample size increases the power for effects better than null.

2. Dual-criterion design

Considers both the statistical significance and the effect estimate.

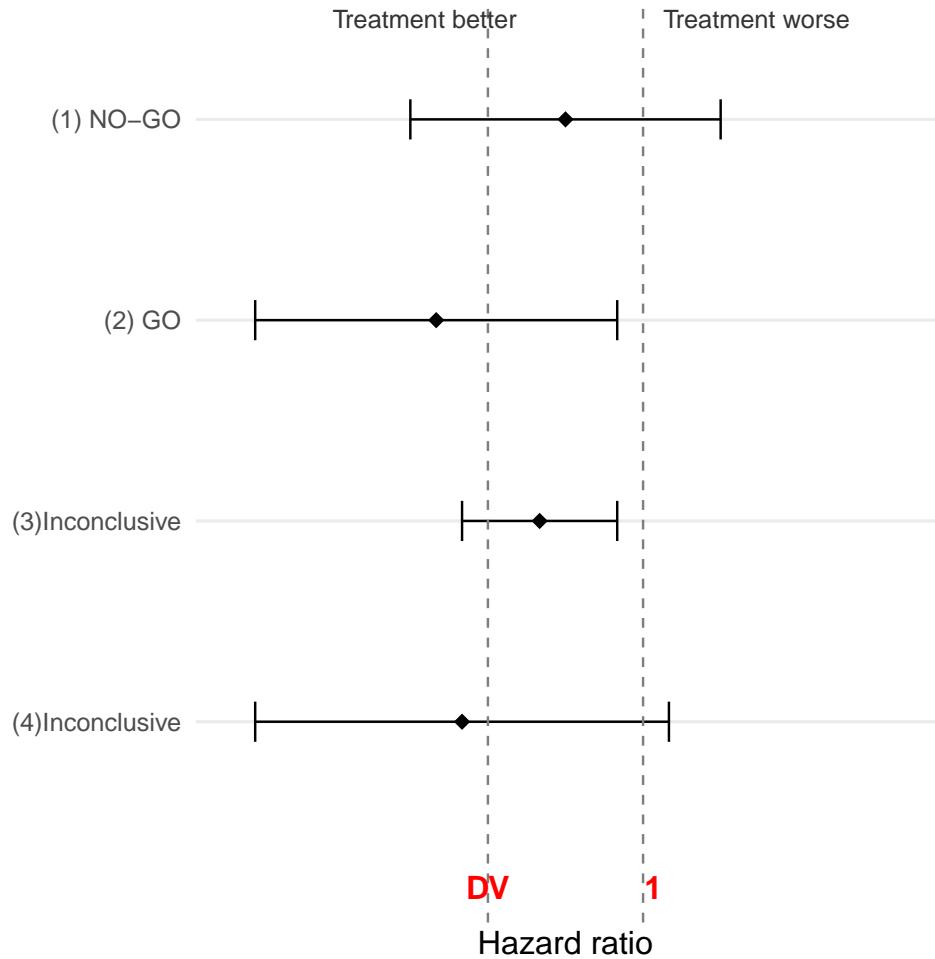
Required inputs: type I error control (null hypothesis and type I error α) and a decision value (DV). The DV is same as the target difference in Fisch’s paper. It is the minimal effect estimate needed for trial success (if higher than this value with moderate confidence, then GO).

By considering both, we have both statistical significance and guarantees a sufficiently large effect estimate.

The dual-criterion is more demanding, the resulting power of study is less compared to standard design.

Power is only increased for values superior to the DV since inferior values are clinically irrelevant.

Decisions for dual-criterion design:



3. Precision design

Doesn't rely on error rates. When null hypothesis or other benchmark values cannot be determined, this can be an option. It requires sufficiently precise effect estimate.

Precision = $\frac{TP}{TP+FP}$. High Precision means that when the model predicts a positive outcome, it is very likely to be correct.

1.1 Hazard ratio and log hazard ratio

Consider hazard function in survival analysis, it describes the risk of failing. We consider hazard ratio between experimental drug and control as the outcome of interest. Hazard ratio(HR) less than 1 means the drug is better than the control. We want to have smaller hazard and hazard ratio, so that the drug is more effective.

In the paper, the log hazard ratio (log HR) is used instead of the hazard ratio. This could be because of the following reasons.

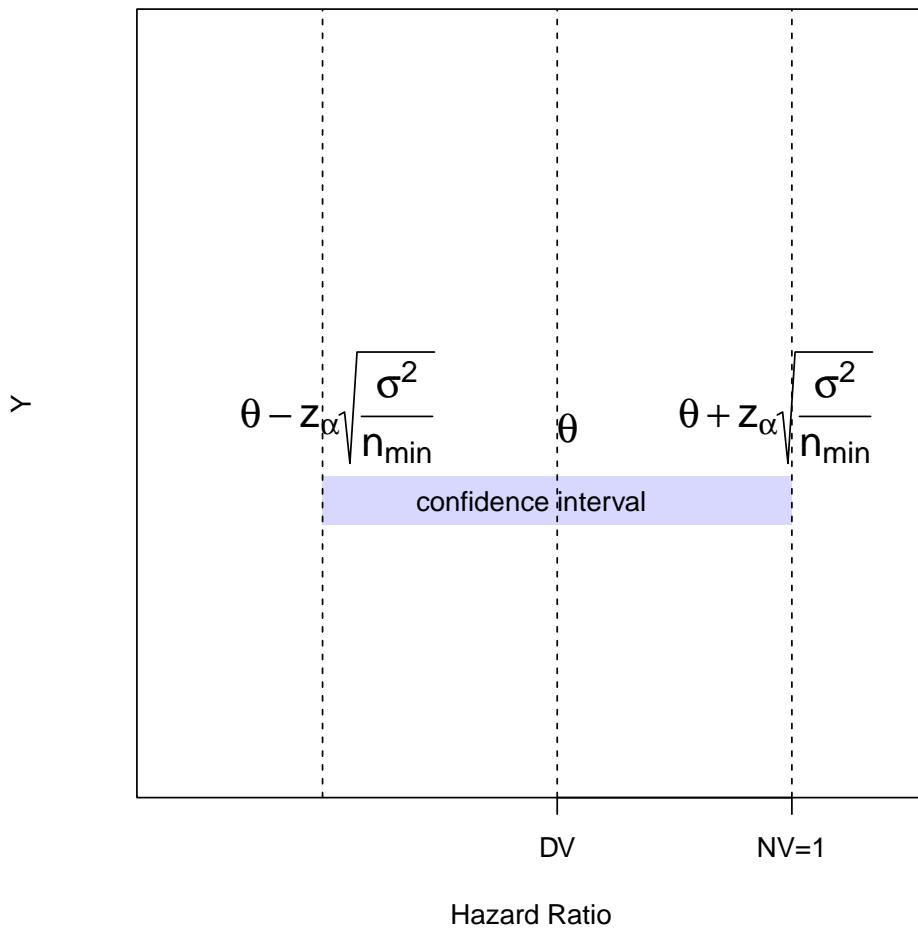
Firstly, according to the Central Limit Theorem, as the sample size increases, the distribution of the estimator (the log HR) approaches a normal distribution. Also, the HR itself is a ratio of hazard rates and is positively skewed, meaning it doesn't naturally fit a normal distribution. Taking the logarithm of the HR stabilizes its variance, transforming the skewed HR distribution into one that is more symmetric and closer to normal. Moreover, this approximate normality of log HR is useful in sample size calculation, which is discussed in the next session.

1.2 Sample size

Given the significance level α , the null value (NV), and the decision value (DV), we can calculate the minimum sample size (for normally distributed data):

$$n_{\min} = \frac{\sigma^2 \times z_\alpha^2}{(NV - DV)^2}$$

where σ is the outcome standard deviation, and takes the value 2 under equal randomization for the standard normal approximation to time-to-event data. The z_α is the $100(1 - \alpha)\%$ quantile of the standard normal distribution. The n_{\min} gives the minimum sample size that implies statistical significance if the effect estimate equals the DV. This value is calculated under the situation that both criterion are just satisfied. As illustrated in the below graph, when the effect estimate $\theta = DV$, and the lower bound of the confidence interval just touches the NV so that statistical significance is reached, we have the minimum sample size. Notice that when sample size equals the minimum sample size, the width of the confidence interval $z_\alpha \sqrt{\frac{\sigma^2}{n_{\min}}}$ equals NV-DV, so there will be no “Inconclusive” decisions. When the sample size is larger, the confidence interval becomes narrower, then an “Inconclusive” decision will occur.



1.3 Operating characteristics

The operating characteristics are the type I error and power of the clinical trial design.

For dual-criterion designs, the power at the DV is approximately 50%, so that if the true parameter equals the DV, there is roughly equal chance that the effect estimate lies on either side of the DV. Having 50% at the DV does not mean the study is under-powered.

1.4 Reproduce Figure 1

In Figure 1, the two plots illustrate the operating characteristics of dual-criterion designs with 309 and 420 events. The number 309 is the minimum sample size calculated under the example conditions $\sigma = 2$, $\alpha = 2.5\%$, log hazard ratios NV = $\log(1)$, DV = $\log(0.8)$.

$$n_{\min} = \frac{2^2 \times z_{0.025}^2}{(\log 1 - \log 0.8)^2} = 308.594 \approx 309$$

The probability of making a “GO” decision is the probability of the estimate smaller than the DV (i.e. clinical relevance) while the NV is outside the confidence interval (i.e. statistical significance). The “NO-GO” decision is made when the estimate is larger than the DV, and the NV is inside the confidence interval. The “Inconclusive” decisions are made if neither “GO” nor “NO-GO” is satisfied. The probability of “Inconclusive” decision is hence 1 minus the probability of “GO” and “NO-GO” decisions.

Below code presents the process of obtaining Figure 1 in the paper [Roychoudhury et al. \(2018\)](#). An extra notice here is the calculation of the cutting value for statistical significance when the sample size is larger than n_{\min} . As we mentioned earlier, when $n > n_{\min}$, the confidence interval is shorter, so the cutting value `cut.ssig` of the “NO-GO” decision should be a value between DV and NV such that $z_{\alpha} \sqrt{\frac{\sigma^2}{n}} = NV - cut.ssig$. So in this case, the $cut.ssig = \log(1) - z_{\alpha} \sqrt{\frac{\sigma^2}{n}}$.

```
> # Sequence of true hazard ratios in log scale
> t.d <- log(seq(0.5, 1, 0.01))
> # Left panel (n = 309)
> n1 <- 309
> sd1 <- sqrt((2^2) / n1) # standard deviation
> cut.ssig1 <- log(0.8) # cutting point for statistical significance
> cut.crel1 <- log(0.8) # cutting point for clinical relevance
> pp.go1 <- pnorm(cut.crel1, t.d, sd1) # probability of GO decision
> pp.ngo1 <- 1 - pnorm(cut.ssig1, t.d, sd1) # probability of NO-GO decision
> pp.intd1 <- 1 - pp.go1 - pp.ngo1 # probability of inconclusive decision
> df1 <- data.frame(HazardRatio = exp(t.d), GO = pp.go1,
+                     NOGO = pp.ngo1, Inconclusive = pp.intd1)
> # Right panel (n = 420)
> n2 <- 420
> sd2 <- sqrt((2^2) / n2) # standard deviation
> cut.ssig2 <- log(1) - qnorm(0.975) * sqrt(2^2 / 420) # statistical significance
> cut.crel2 <- log(0.8) # clinical relevance
> pp.go2 <- pnorm(cut.crel2, t.d, sd2) # probability of GO decision
```

```

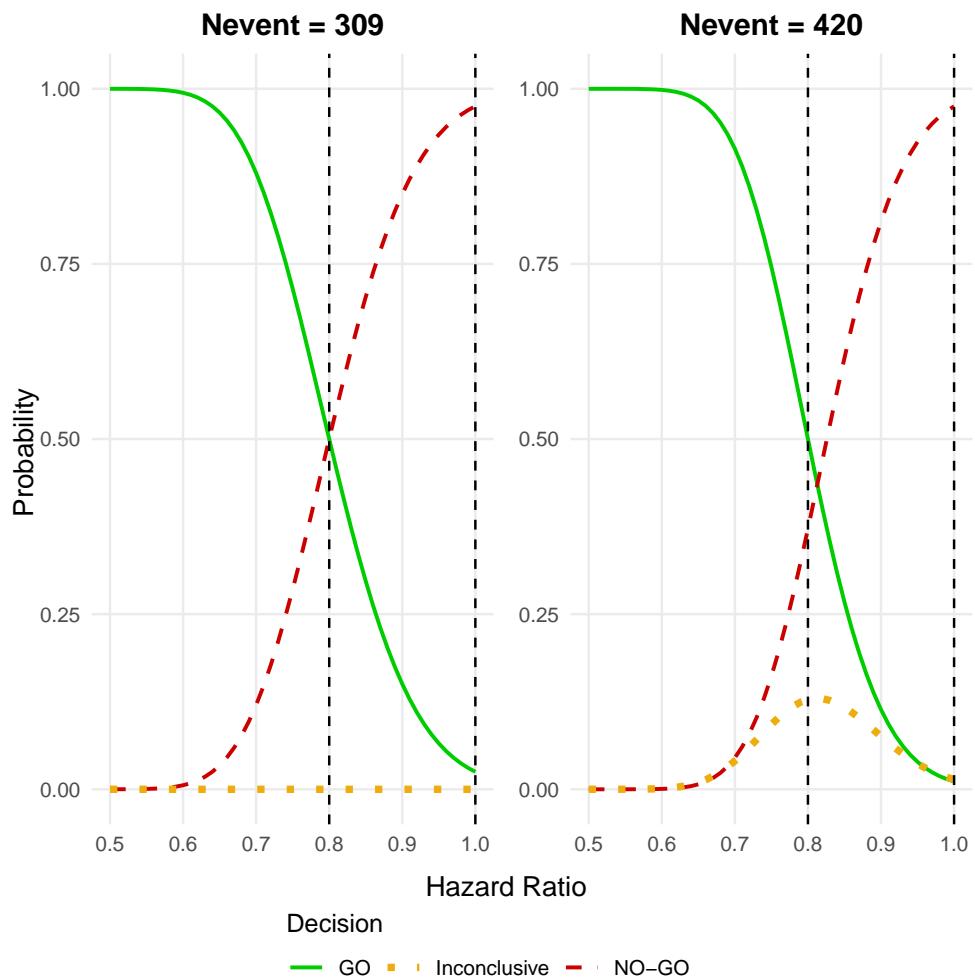
> pp.ngo2 <- 1 - pnorm(cut.ssig2, t.d, sd2) # probability of NO-GO decision
> pp.intd2 <- 1 - pp.go2 - pp.ngo2 # probability of inconclusive decision
> df2 <- data.frame(HazardRatio = exp(t.d), GO = pp.go2,
+                      NOGO = pp.ngo2, Inconclusive = pp.intd2)
> # Define the line colors and types
> line_colors <- c("GO" = "green3", "Inconclusive" = "darkgoldenrod2", "NO-GO" = "red3")
> line_types <- c("GO" = "solid", "Inconclusive" = "dotted", "NO-GO" = "dashed")
> # Plot for n = 309
> p1 <- ggplot(df1, aes(x = HazardRatio)) +
+   geom_line(aes(y = GO, color = "GO", linetype = "GO"), size=0.8) +
+   geom_line(aes(y = NOGO, color = "NO-GO", linetype = "NO-GO"), size=0.8) +
+   geom_line(aes(y = Inconclusive, color = "Inconclusive",
+                  linetype = "Inconclusive"), size=1.5) +
+   geom_vline(xintercept = c(0.8, 1.0), linetype = "dashed", color = "black") +
+   labs(title = "Nevent = 309") +
+   scale_color_manual(values = line_colors) +
+   scale_linetype_manual(values = line_types) +
+   theme_minimal() +
+   theme(
+     legend.position = "none",
+     plot.title = element_text(hjust = 0.5, face = "bold"),
+     panel.grid.minor = element_blank(),
+     axis.title.y = element_blank(), # Remove individual y-labels
+     axis.title.x = element_blank()
+   )
> # Plot for n = 420
> p2 <- ggplot(df2, aes(x = HazardRatio)) +
+   geom_line(aes(y = GO, color = "GO", linetype = "GO"), size=0.8) +
+   geom_line(aes(y = NOGO, color = "NO-GO", linetype = "NO-GO"), size=0.8) +
+   geom_line(aes(y = Inconclusive, color = "Inconclusive",
+                  linetype = "Inconclusive"), size=1.5) +
+   geom_vline(xintercept = c(0.8, 1.0), linetype = "dashed", color = "black") +
+   labs(title = "Nevent = 420") +
+   scale_color_manual(values = line_colors) +
+   scale_linetype_manual(values = line_types) +
+   theme_minimal() +
+   theme(
+     legend.position = "none",
+     plot.title = element_text(hjust = 0.5, face = "bold"),
+     panel.grid.minor = element_blank(),
+     axis.title.y = element_blank(), # Remove individual y-labels
+     axis.title.x = element_blank()
+   )

```

```

> # Combine the plots into a single figure without individual y-axis labels
> combined_plots <- plot_grid(p1, p2, ncol = 2, align = 'hv', rel_widths = c(1, 1))
> # Extract and create a shared legend
> legend <- get_legend(
+   p1 + theme(legend.position = "right") +
+   guides(color = guide_legend(title = "Decision", nrow = 1),
+         linetype = guide_legend(title = "Decision", nrow = 1)))
> # Add a shared y-axis label using grid.arrange
> final_plot <- grid.arrange(
+   arrangeGrob(combined_plots,
+               left = textGrob("Probability", rot = 90, vjust = 1.2),
+               bottom = textGrob("Hazard Ratio", just = "centre")),
+   legend = legend,
+   ncol = 1,
+   heights = c(10, 1)
+ )
> # Print the final plot
> print(final_plot)

```



2 Example 1: A randomized PoC design with time-to-event data

Randomized, double-blind, RCT. Patients were randomized equally to: (experimental drug + standard care) OR (standard care only).

Primary outcome of interest (or called “endpoint”) is the progression-free survival (PFS), which is the time when the disease or cancer do not get worse. The endpoint was assessed with a *log-rank test* and *Cox regression* with treatment as a covariate.

- *log-rank test*: compare the survival distributions of two or more groups. It tests the hypothesis that there is no difference in survival (or time-to-event) between the two groups. If the log-rank test indicates a significant difference, it suggests the treatment affects how long patients live without their disease worsening.
- *Cox regression*: estimate the hazard ratio between two groups, which tells us the relative risk of disease progression in the treatment group compared to the control group. If the HR < 1, it suggests that the new treatment delays disease progression better than the control treatment.

As for the DV, HR=0.7 was deemed necessary to be clinically meaningful. Values larger than 0.7 are unsatisfactory to clearly justify further development of the drug.

So the dual-criterion is:

1. Statistical significance: one-sided p-value of log-rank test ≤ 0.1 .
2. Clinical relevance: estimated HR from Cox regression ≤ 0.70 .

2.1 Reproduce Table 3

Here we attempt to reproduce the values in Table 3.

For the first sub-table, similar to Section 1.4, when $n > n_{\min}$, the cutting value $\hat{\theta}$ (`cut.ssig`) of the “NO-GO” decision is given by $\text{cut.ssig} = \log(1) - z_{\alpha} \sqrt{\frac{\sigma^2}{n}}$. These correspond to $\hat{\theta} > 0.736 \neq 0.7$ from the paper.

For the second sub-table, $n = n_{\min}$.

For the third sub-table, it requires one-sided type I error of 0.1 and power of 0.9 for HR=0.5. So the cutting value for clinical relevance is chosen to satisfy these requirements. One thing to be answered is why the authors used 0.901 as the power for HR=0.5, rather than 0.9.

For the forth sub-table, it requires one-sided type I error of 0.1 and power of 0.8 for HR=0.5. The sample size $n = 38 < n_{\min}$. The cutting value of the decisions are calculated in a similar way as in Section 1.4. One thing to be answered here is that the current calculation returns the same values in the table, but the $\hat{\theta}$ is slightly different from 0.659 as claimed in the paper. Also, the power at HR=0.5 is 0.804, rather than 0.8.

For sub-table five, it used a type I error of 0.2 and a power of 0.9, with sample size $n = 38 < n_{\min}$. The cutting value of the decisions are calculated in a similar way as in Section 1.4. However the question to be answered is the same with that in sub-table 4.

```
> # minimum sample size  
> n.min <- ceiling((4*pnorm(0.9)^2)/(log(1)-log(0.7))^2) # n.min = 52  
> # a sequence of true log(HR).
```

```

> t.d <- log(seq(0.5, 1, 0.1))
> # Dual-criterion design: alpha=0.1, DV=log(0.7), n=70
> n1 <- 70
> sd1 <- sqrt((2^2)/n1) # standard deviation
> cut.ssig <- log(1)-qnorm(0.9)*sqrt(2^2 / n1) # statistical significance
> cut.crel <- log(0.7) # critical relevance
> pp.go1 <- pnorm(cut.crel, t.d, sd1)
> pp.ngo1 <- 1- pnorm(cut.ssig, t.d, sd1)
> pp.intd1 <- 1 -pp.go1 - pp.ngo1
> subtable1 <- matrix(data=round(c(exp(t.d), pp.go1,pp.ngo1,pp.intd1),3), ncol=4)
> # Dual-criterion design: alpha=0.1, DV=0.7, n=52
> n2 <- 52
> sd2 <- sqrt((2^2)/n2)
> cut.ssig <- log(0.7)
> cut.crel <- log(0.7)
> pp.go2 <- pnorm(cut.crel, t.d, sd2)
> pp.ngo2 <- 1-pnorm(cut.ssig, t.d, sd2)
> pp.intd2 <- 1 -pp.go2 - pp.ngo2
> subtable2 <- matrix(data=round(c(exp(t.d), pp.go2,pp.ngo2,pp.intd2),3), ncol=4)
> # Dual-criterion design: alpha=0.1, beta=0.1, n=55
> n3 <- 55
> sd3 <- sqrt((2^2)/n3)
> cut.ssig <- qnorm(0.901,log(0.5),sd3) # reason for using 0.901 rather than 0.9?
> cut.crel <- qnorm(0.901,log(0.5),sd3)
> pp.go3 <- pnorm(cut.crel, t.d, sd3)
> pp.ngo3 <- 1-pnorm(cut.ssig, t.d, sd3)
> pp.intd3 <- 1 -pp.go3 - pp.ngo3
> subtable3 <- matrix(data=round(c(exp(t.d), pp.go3,pp.ngo3,pp.intd3),3), ncol=4)
> # Dual-criterion design: alpha=0.1, beta=0.2, n=38
> n4 <- 38
> sd4 <- sqrt((2^2)/n4)
> cut.ssig <- log(1)-qnorm(0.9)*sqrt(2^2 / n4) # different from 0.659
> cut.crel <- log(1)-qnorm(0.9)*sqrt(2^2 / n4)
> pp.go4 <- pnorm(cut.crel, t.d, sd4)
> pp.ngo4 <- 1-pnorm(cut.ssig, t.d, sd4)
> pp.intd4 <- 1 -pp.go4 - pp.ngo4
> subtable4 <- matrix(data=round(c(exp(t.d), pp.go4,pp.ngo4,pp.intd4),3), ncol=4)
> # Dual-criterion design: alpha=0.2, beta=0.1, n=38
> n5 <- 38
> sd5 <- sqrt((2^2)/n5)
> cut.ssig <- log(1)-qnorm(0.8)*sqrt(2^2 / n5)
> # How is the power guaranteed to be 0.1? This is only using alpha=0.2.
> cut.crel <- log(1)-qnorm(0.8)*sqrt(2^2 / n5)

```

```

> pp.go5 <- pnorm(cut.crel, t.d, sd5)
> pp.ngo5 <- 1-pnorm(cut.ssig, t.d, sd5)
> pp.intd5 <- 1 -pp.go5 - pp.ngo5
> subtable5 <- matrix(data=round(c(exp(t.d), pp.go5, pp.ngo5, pp.intd5), 3), ncol=4)
> # Combine subtables by rows
> combined_table <- rbind(subtable1, subtable2, subtable3, subtable4, subtable5)
> # Convert to data frame for better kable support
> combined_table <- as.data.frame(combined_table)
> kable(combined_table, format = "latex", align = "c", booktabs = TRUE,
+       col.names = c("True HR", "GO", "NO-GO", "Inconclusive")) %>%
+   kable_styling(full_width = FALSE, position = "center",
+                 latex_options = c("hold_position", "scale_down")) %>%
+   add_header_above(c("Reproduced Table 3" = 4)) %>%
+   group_rows("Subtable 1", 1, 6) %>%
+   group_rows("Subtable 2", 7, 12) %>%
+   group_rows("Subtable 3", 13, 18) %>%
+   group_rows("Subtable 4", 19, 24) %>%
+   group_rows("Subtable 5", 25, 30)

```

Reproduced Table 3				
True HR	GO	NO-GO	Inconclusive	
Subtable 1				
0.5	0.920	0.053	0.027	
0.6	0.740	0.196	0.063	
0.7	0.500	0.417	0.083	
0.8	0.288	0.636	0.076	
0.9	0.147	0.800	0.054	
1.0	0.068	0.900	0.032	
Subtable 2				
0.5	0.887	0.113	0.000	
0.6	0.711	0.289	0.000	
0.7	0.500	0.500	0.000	
0.8	0.315	0.685	0.000	
0.9	0.182	0.818	0.000	
1.0	0.099	0.901	0.000	
Subtable 3				
0.5	0.901	0.099	0.000	
0.6	0.729	0.271	0.000	
0.7	0.516	0.484	0.000	
0.8	0.324	0.676	0.000	
0.9	0.186	0.814	0.000	
1.0	0.100	0.900	0.000	
Subtable 4				
0.5	0.804	0.196	0.000	
0.6	0.615	0.385	0.000	
0.7	0.428	0.572	0.000	
0.8	0.276	0.724	0.000	
0.9	0.169	0.831	0.000	
1.0	0.100	0.900	0.000	
Subtable 5				
0.5	0.902	0.098	0.000	
0.6	0.768	0.232	0.000	
0.7	0.602	0.398	0.000	
0.8	0.439	0.561	0.000	
0.9	0.303	0.697	0.000	
1.0	0.200	0.800	0.000	

3 Example 2: A single-arm PoC design with binary data

Experimental drug in Chinese patients with non-small-cell lung cancer.

Primary endpoint is objective response rate (ORR), which quantifies the preliminary efficacy of the experimental drug.

Prior: minimally informative unimodal beta prior distribution $Beta(0.0811, 1)$, which has mean 0.75.

NV is set to 7.5% rather than 0, because of the absence of a comparator (in single arm trials).

DV is set to be $10\% + 7.5\% = 17.5\%$.

So the dual-criterion is:

1. Bayesian statistical significance: $\mathbb{P}(ORR \geq 7.5\% | data) \geq 0.95$
2. Clinical relevance: Posterior median $\geq 17.5\%$

The minimal sample size was 22. Final sample size 25.

Null hypothesis: there is no effect of the drug, i.e. ORR=7.5%

$$\mathbb{P}(\text{type I error}) = \mathbb{P}(\text{reject } H_0 | H_0 \text{ is true}) = \mathbb{P}(\text{reject } H_0 | ORR \leq 7.5\%)$$

$$\mathbb{P}(\text{type II error}) = \mathbb{P}(\text{not reject } H_0 | H_0 \text{ is false}) = \mathbb{P}(\text{reject } H_0 | ORR = \text{response rate})$$

Table 4 results show that this dual-criterion design is a three-outcome design with desirable properties.

3.1 Reproduce Figure 2

3.2 Reproduce Table 4

References

Roychoudhury, S., Scheuer, N., and Neuenschwander, B. (2018). Beyond p-values: A phase II dual-criterion design with statistical significance and clinical relevance. *Clinical trials (London, England)*, 15(5):452–461.

Computational details

```
> cat(paste(Sys.time(), Sys.timezone(), "\n"))
```

2024-11-07 22:13:39.296199 Europe/Zurich

```
> sessionInfo()
```

R version 4.4.0 (2024-04-24)

Platform: aarch64-apple-darwin20

Running under: macOS Sonoma 14.4

Matrix products: default

BLAS: /Library/Frameworks/R.framework/Versions/4.4-arm64/Resources/lib/libRblas.0.dylib

LAPACK: /Library/Frameworks/R.framework/Versions/4.4-arm64/Resources/lib/libRlapack.dylib; LAP

locale:

```
[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
```

time zone: Europe/Zurich

tzcode source: internal

attached base packages:

```
[1] grid      stats     graphics  grDevices utils      datasets  methods  
[8] base
```

other attached packages:

```
[1] latex2exp_0.9.6  kableExtra_1.4.0 gridExtra_2.3    cowplot_1.1.3  
[5] ggplot2_3.5.1   knitr_1.48
```

loaded via a namespace (and not attached):

```
[1] gtable_0.3.5      dplyr_1.1.4       compiler_4.4.0    tidyselect_1.2.1  
[5] xml2_1.3.6       stringr_1.5.1     systemfonts_1.1.0 scales_1.3.0  
[9] fastmap_1.2.0     R6_2.5.1        labeling_0.4.3    generics_0.1.3  
[13] tibble_3.2.1      munsell_0.5.1    svglite_2.1.3    pillar_1.9.0  
[17] rlang_1.1.4       utf8_1.2.4       stringi_1.8.4    xfun_0.44  
[21] viridisLite_0.4.2 cli_3.6.2       withr_3.0.0      magrittr_2.0.3  
[25] digest_0.6.35    rstudioapi_0.16.0 lifecycle_1.0.4   vctrs_0.6.5  
[29] evaluate_0.24.0   glue_1.7.0       farver_2.1.2    fansi_1.0.6  
[33] colorspace_2.1-0 rmarkdown_2.27   tools_4.4.0      pkgconfig_2.0.3  
[37] htmltools_0.5.8.1
```