

Panic in the Disko Bay: Estimating Greenland Bowhead Whale Detection and Abundance with Distance Sampling

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Project 1: Distance Sampling



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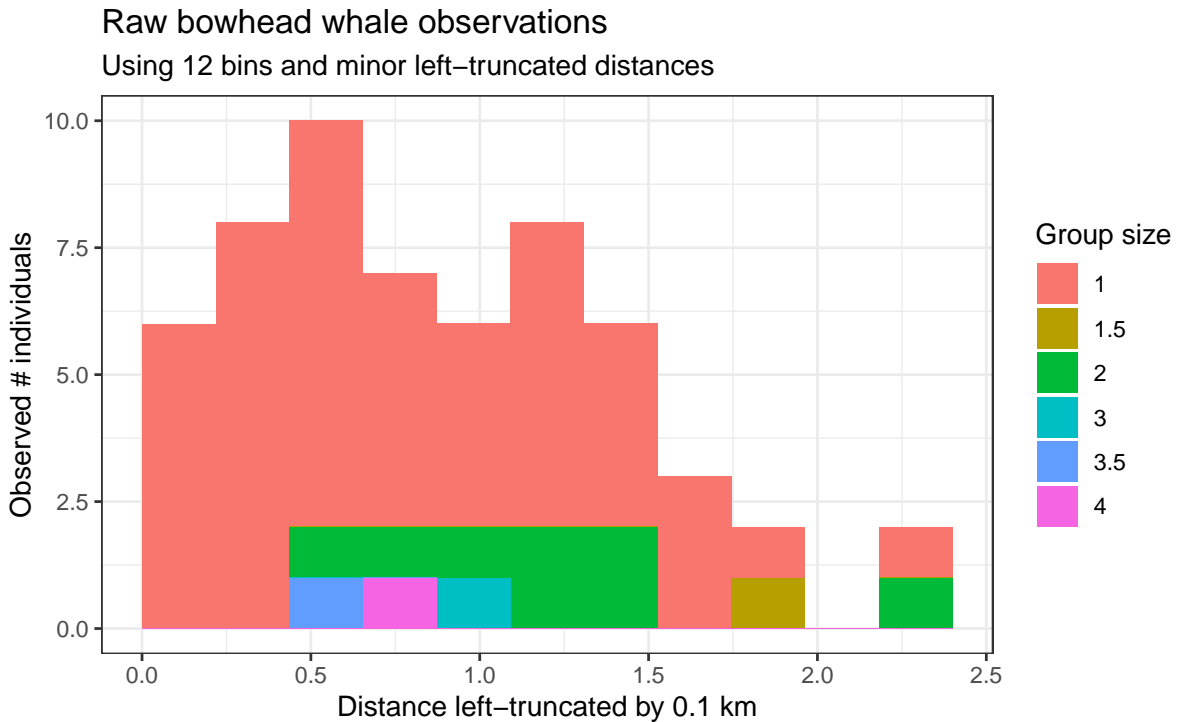
Abstract

Introduction

In this report, we aim to use distance sampling methods to estimate the abundance of Bowhead whales in Disko Bay. As the data show some observer avoidance, we also wish to explore different approaches used to deal with this issue.

Methods

Visual aerial surveys were conducted along 41 systematically placed east-west transects along the West Greenland coast, with a combined length of 4,445km. There were two observation platforms which each recorded the declination angle using inclinometers and the times of the first sighting and when the whale passed abeam, which were used to calculate the perpendicular and forward distances respectively. Both observers also recorded Beaufort sea state and group size, and if there were any differences in size or declination between the observers the mean was taken. The distances were left-truncated to 100m as the view was obscured close to the transect line [1]. In total there were 58 valid observations.

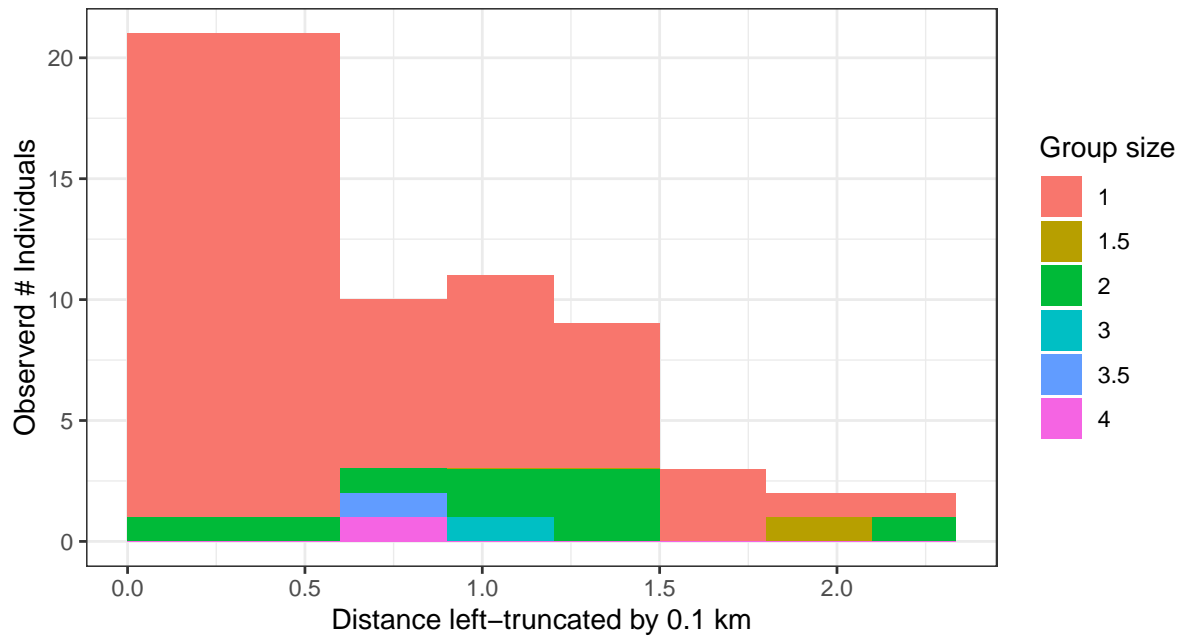


An initial plot of the detection distances suggests there could be some line avoidance, as more whales were detected further away from the plane than on the transect, even after the left-truncation. This could violate the assumption in distance sampling that animals are distributed independently with respect to the transect, or the assumption that all animals on the line are detected, depending on whether the whales are hiding or moving away [2]. Equally, this could be stochastic variation as there is only a difference of 4 observations between the 0-0.2 bin and the 0.4-0.6 bin. We considered 3 approaches to deal with this problem. Firstly, treating the difference in detection as random variation and fitting detection functions as normal. This preserves all the observations but will result in overestimating the detection probability (therefore underestimating abundance) if detection is not certain on the line. Another option is to bin the distances into 0-0.6km, 0.6-0.9km, ..., 2.1-2.4km categories so

that there are more observations in the first group than the others. This assumes the whales moved away from the observers, so those that would have been detected on the transect were instead seen further away. This method deals well with the potentially violated assumptions, but in grouping the data we lose information.

Binned bowhead whale observations

Using 7 custom bins with minor left-truncated distances



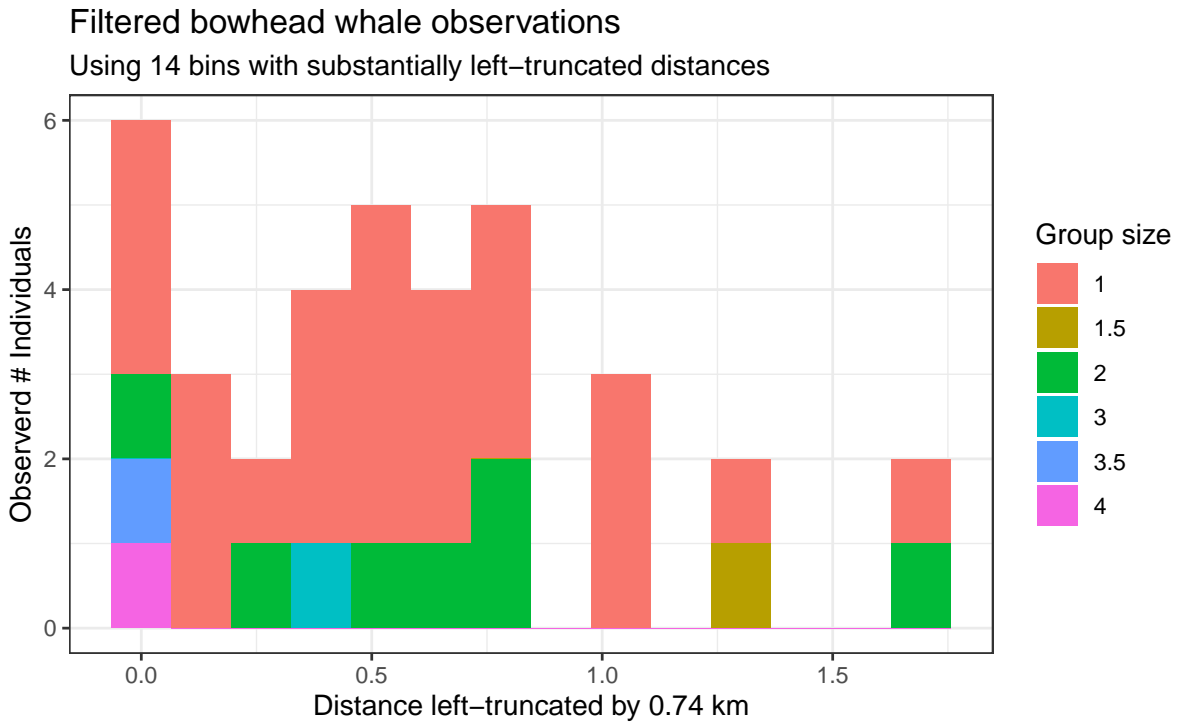
The final option we considered is to left-truncate the distances up to the peak in detections at 0.64km. This method ensures we have the monotonic decline in detections with increasing distance, but again loses information and assumes that detection is certain 0.64km away from the plane. The monotonic decrease in detection allowed complex models with multiple covariates and adjustment terms to be fitted successfully, which didn't happen for the other methods. However, given the small sample size these models likely suffer from over-fitting.

Table 1: Estimated abundance from best-performing models (by AIC)

Detection.Function	Data	Size.Covariate	Estimate	SE	Lower.CI	Upper.CI	AIC
Half Normal	Base	Yes	212.07	65.62	112.13	401.06	86.03
Half Normal	Base	No	228.82	71.45	122.00	429.14	86.32
Half Normal	Binned	Yes	212.84	66.14	112.32	403.31	197.58
Half Normal	Binned	No	229.54	72.22	121.94	432.07	197.91
Hazard Rate	Truncated	Yes	1803.36	1346.22	461.56	7045.88	27.20
Half Normal	Truncated	No	519.51	90.74	363.25	743.00	28.43
Half Normal	Truncated	Yes	608.75	154.26	356.93	1038.20	28.73

Table 2: Estimates (and errors) of size on detectability

Model	Size.Coefficient	SE	Significant.
Base HN + size	6.16	5046.40	No
Binned HN + size	5.64	305.31	No
Truncated HN + size	-0.51	0.26	Yes



We used the Distance package to fit models using a maximum likelihood approach. For each data set (unedited, binned, truncated) we fitted half-normal and hazard rate detection functions with all combinations of adjustment terms (cosine, Hermite, polynomial) and covariates (group size and Beaufort sea state). We used AIC to compare between models fitted to the same data set.

Results

Given raw data set the best model selected by AIC was half-normal detection function with size covariate and no adjustments. It estimates whale abundance as 212 with confidence interval (112.13, 401.06). However, size is not significant, therefore the model without covariates gives

similar estimate of whale abundance (229) with confidence interval (112.13, 401.06).

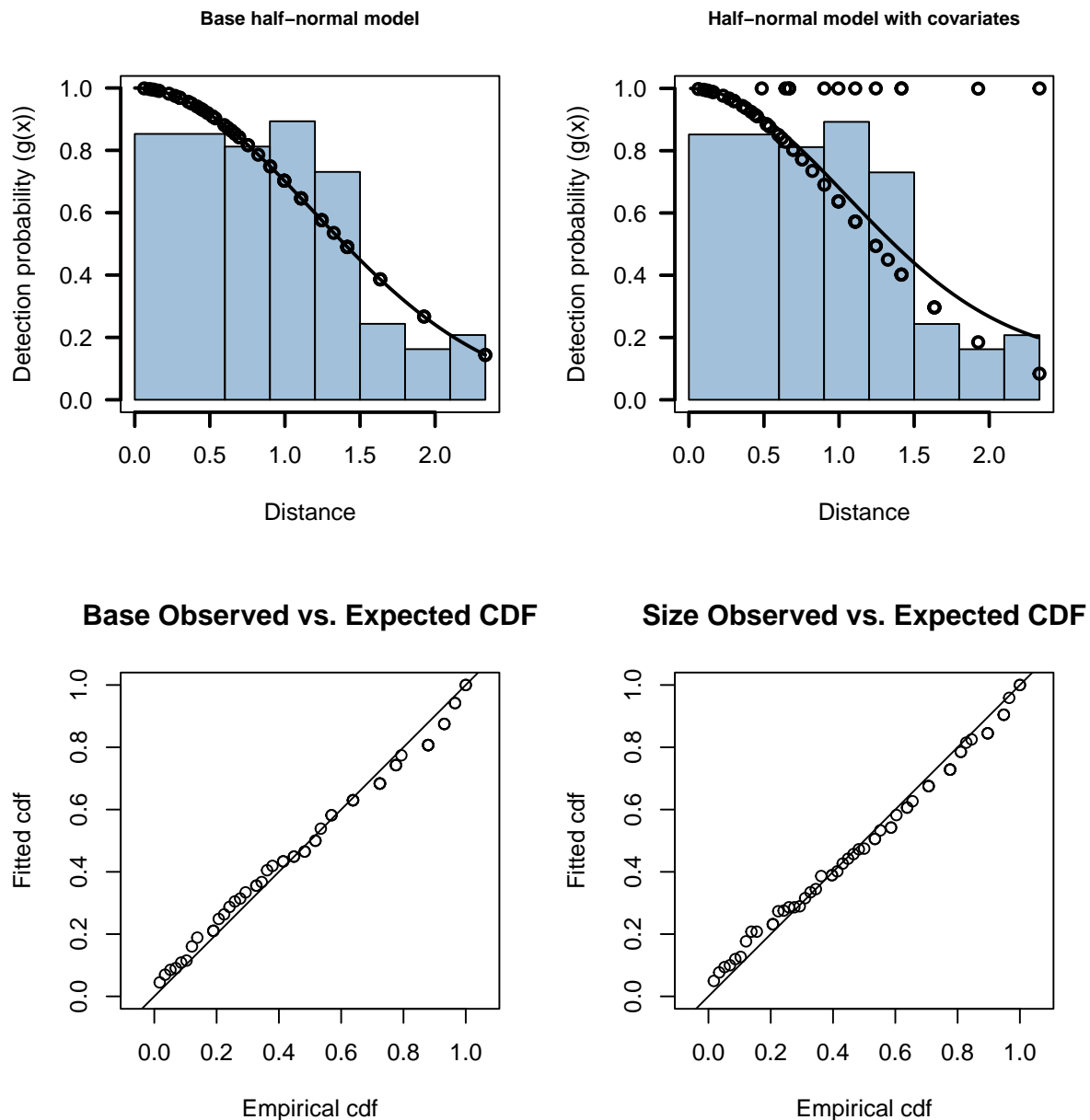


Figure 1: Detection functions using original data

When using binning, AIC selects half-normal detection function with no adjustments and size. Like the model using raw data, it gives estimate of abundance of 213 and confidence interval (112.32, 403.31). Size is not significant.

The model chosen using truncated data is quite different. The best model is hazard rate with size covariate, with estimate of 1803 whales and confidence interval (461.56, 7045.88). It is worth noting that size is significant. Nonetheless, given huge standard error, there's a lot of uncertainty, therefore it is likely that the model largely overestimates.

Even though detection function using truncated data has lowest AIC, it overestimates whale abundance. The reason might be limited to sample size and extrapolation based on the area we have no data about. Therefore, it might be best to use half-normal detection function with

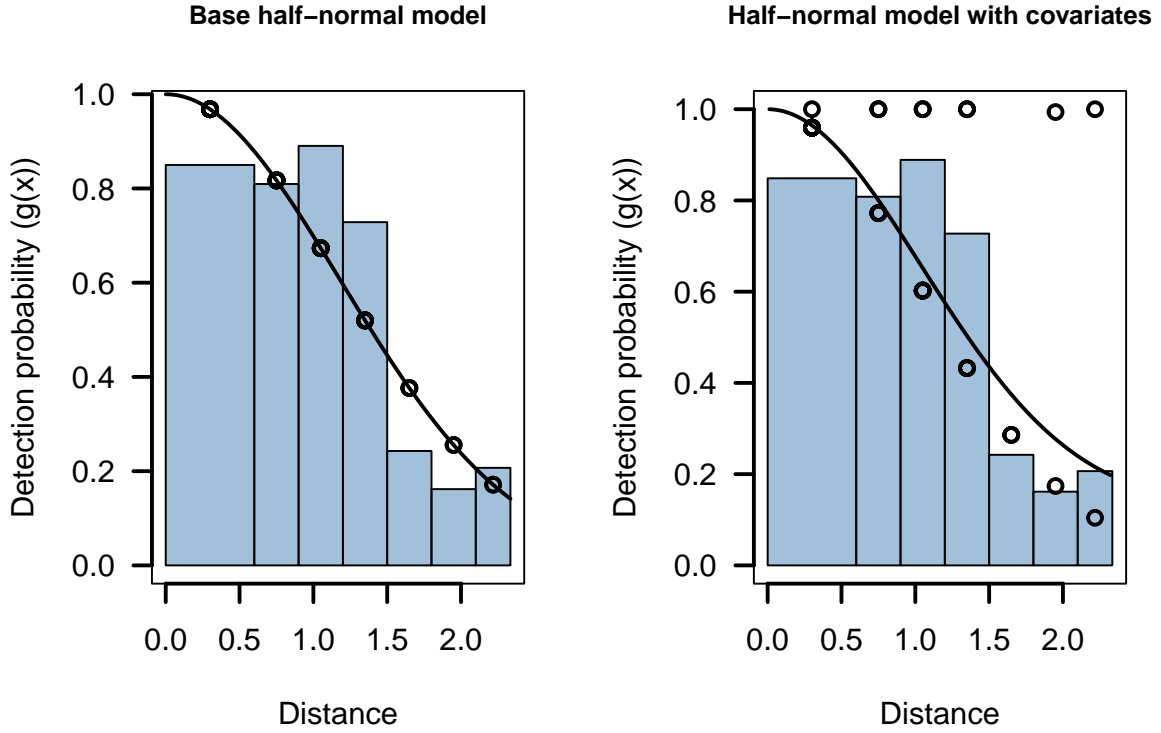


Figure 2: Detection functions using binned data

size covariate to get the best estimate of whale abundance.

Discussion

While conducting survey, there are several assumptions that had been violated. The area beneath the aircraft is not visible to surveyors, therefore, we cannot assume that objects on the transect line are detected with certainty. To account for this, passive monitoring such as cameras to count whales on the transect line could be used. Moreover, since whales are never stationary, as they run or hide when noticing observers, assumption of object detection at their initial location is violated. This results in less observations within close distance and we are likely underestimating the true whale abundance due to imperfect detection. To address this availability bias, hidden Markov model could be used [3]. HMM copes with a small data by fitting a more restrictive model, which incorporates information on how the aircraft moves.

Violation of these assumptions bias estimates of whale abundance – models will underestimate the real number of whales; therefore, truncation of data is needed. However, truncation reduces survey area by 30% and since data is quite unbalanced and sample size is small, it is not a great approximation of how whale abundance would look like in real life and any variation seen in estimates is due to stochasticity.

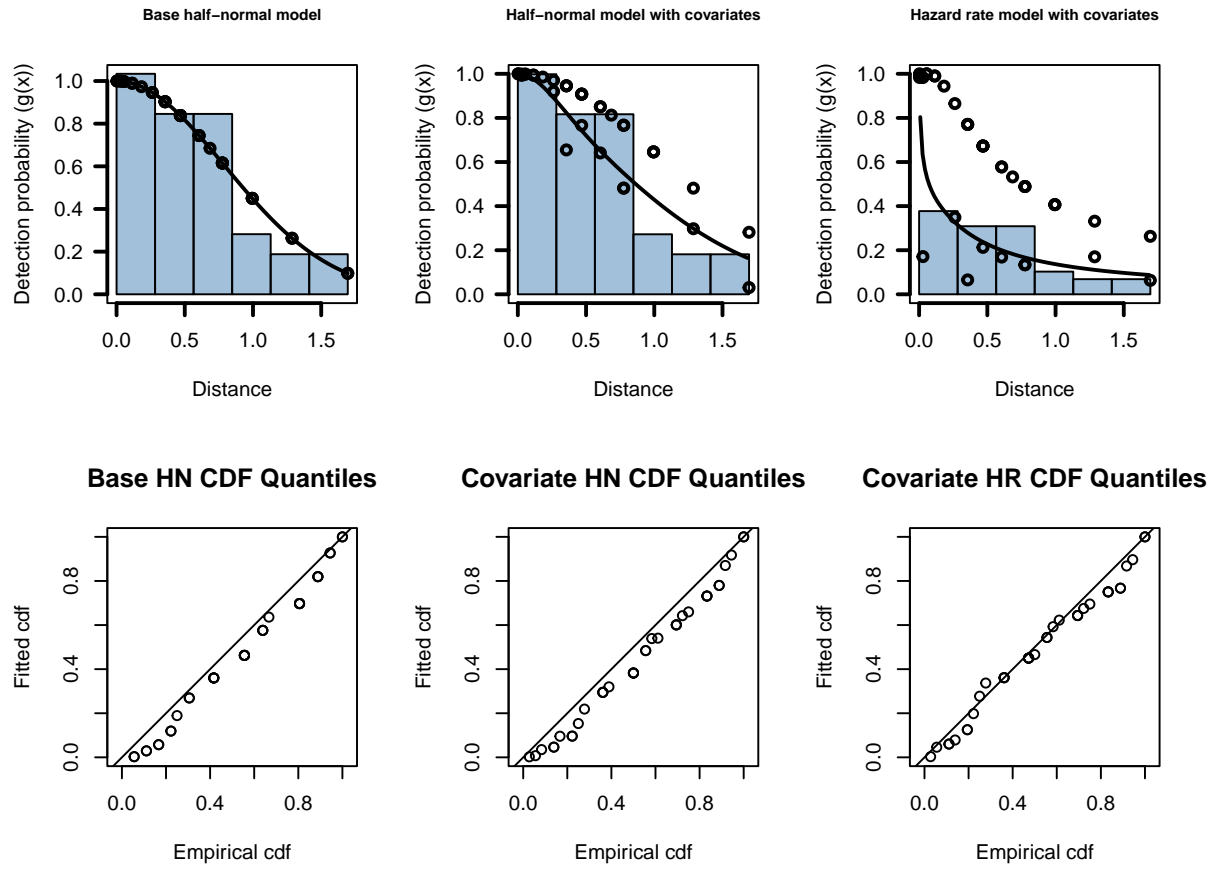


Figure 3: Detection functions using truncated data

Code Appendix

```
# import packages and data
library(statsecol)
library(tidyverse)
library(viridisLite)
library(ggbridges)
library(Distance)
data(bowhead_LT)

# raw data plot
# looks like hazard rate would be the best fit here
ggplot(data = bowhead_LT) +
  geom_histogram(aes(x = distance, fill = as.factor(size)), bins = 21) +
  labs(x = "Distance truncated by 0.1 km",
       y = "Observerd # individuals",
       title = "Raw bowhead whale observations",
       subtitle = "Using 21 bins and minor left-truncated distances",
       fill = "Group size") +
  theme_bw()

# aggressive binning
# definitely half-normal is the better fit here
histBreaks2 <- c(0, 0.6, 0.9, 1.2, 1.5, 1.8, 2.1, max(bowhead_LT$distance,
                                                       na.rm = TRUE))

ggplot(data = bowhead_LT) +
  geom_histogram(aes(x = distance, fill = as.factor(size)),
                breaks = histBreaks2) +
  labs(x = "Distance truncated by 0.1 km",
       y = "Observerd # Individuals",
       title = "Binned bowhead whale observations",
       subtitle = "Using 7 custom bins with minor left-truncated distances",
       fill = "Group size") +
  theme_bw()

#truncate to get decreasing observations with distance
bowhead_LT_Trunc = bowhead_LT %>% filter(distance >= 0.64) %>%
  mutate(distance = distance - 0.64)

# plot the new data
# both half-normal and hazard rate appear possible here
ggplot(data = bowhead_LT_Trunc) +
  geom_histogram(aes(x = distance, fill = as.factor(size)), bins = 14) +
  labs(x = "Distance left-truncated by 0.74 km",
       y = "Observerd # Individuals",
       title = "Filtered bowhead whale observations",
       subtitle = "Using 14 bins with substantially left-truncated distances",
       fill = "Group size") +
  theme_bw()
```



```

conversion.factor <- convert_units(distance_units = "kilometre",
                                   effort_units = "kilometre",
                                   area_units = "square kilometre")

# using model fits from separate alternatives,
# we'll only fit no-adjustment HN, HR, and size covariates
# ds() will automatically pick the low AIC version (adjustment = NULL)
baseRawHN <- ds(data = bowhead_LT, key = "hn",
               convert_units = conversion.factor)
baseRawHR <- ds(data = bowhead_LT, key = "hr",
               convert_units = conversion.factor)

# sometimes does not converge
sizeRawHN <- ds(data = bowhead_LT, key = "hn", adjustment = NULL,
               convert_units = conversion.factor,
               formula = ~size)
sizeRawHR <- ds(data = bowhead_LT, key = "hr", adjustment = NULL,
               convert_units = conversion.factor,
               formula = ~size)

# size + half-normal is the best fit on the raw data
summarize_ds_models(baseRawHN,
                   baseRawHR,
                   sizeRawHN,
                   sizeRawHR, output = "plain")

par(mfrow = c(2,2))
plot(baseRawHN, which=2, pl.col = adjustcolor("steelblue",0.5), border=NULL,
     lwd = 2,
     breaks = histBreaks2,
     ylab = "Detection probability (g(x))", xlab = "Distance", las=1,
     main = "Base half-normal model")

# covariate model assumes perfect detection of large-sized groups
# this is a quirk of the limited data
# only Region 2 has any within-group variation in size
plot(sizeRawHN, which=2, pl.col = adjustcolor("steelblue",0.5),border=NULL,
     lwd = 2,
     breaks = histBreaks2,
     ylab = "Detection probability (g(x))", xlab = "Distance", las=1,
     main = "Half-normal model with covariates")

# both models have solid GOF
# covariate model is slightly better from visual QQ inspection
gof_ds(baseRawHN, main="Base Observed vs. Expected CDF", ks = TRUE)
gof_ds(sizeRawHN, main="Covariate Observed vs. Expected CDF", ks = TRUE)

# repeat for binned data
baseBinHN <- ds(data = bowhead_LT, key = "hn",
               cutpoints = histBreaks2,
               convert_units = conversion.factor)
baseBinHR <- ds(data = bowhead_LT, key = "hr",
               cutpoints = histBreaks2,
               convert_units = conversion.factor)

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# often does not converge
sizeBinHN <- ds(data = bowhead_LT, key = "hn", adjustment = NULL,
               cutpoints = histBreaks2,
               convert_units = conversion.factor,
               formula = ~size)
sizeBinHR <- ds(data = bowhead_LT, key = "hr", adjustment = NULL,
               cutpoints = histBreaks2,
               convert_units = conversion.factor,
               formula = ~size)

#binned data also prefers models in the same order as unbinned
summarize_ds_models(baseBinHN,
                   baseBinHR,
                   sizeBinHN,
                   sizeBinHR, output = "plain")

# binned models have a very similar fit (if not identical) to the raw data
par(mfrow = c(1,2))
plot(baseBinHN, which=2, pl.col = adjustcolor("steelblue",0.5),border=NULL,
     lwd = 2,
     ylab = "Detection probability (g(x))", xlab = "Distance", las=1,
     main = "Base half-normal model")
# covariate model still assumes perfect detection of large-sized groups
# still a quirk of the data due to between-strata variation
plot(sizeBinHN, which=2, pl.col = adjustcolor("steelblue",0.5),border=NULL,
     lwd = 2,
     ylab = "Detection probability (g(x))", xlab = "Distance", las=1,
     main = "Half-normal model with covariates")
# GOF produce output but cannot generate plots
# bootstrapping impossible so KS test doesn't happen
gof_ds(baseBinHN, main="Base Observed vs. Expected CDF", ks = TRUE)
gof_ds(sizeBinHN, main="Covariate Observed vs. Expected CDF", ks = TRUE)

# finally repeat for truncated data
# truncated data prefers complex fits w/ more covariates and adjustments
baseTruncHN <- ds(data = bowhead_LT_Trunc, key = "hn",
                 convert_units = conversion.factor)
baseTruncHR <- ds(data = bowhead_LT_Trunc, key = "hr",
                 convert_units = conversion.factor)
sizeTruncHN <- ds(data = bowhead_LT_Trunc, key = "hn", adjustment = NULL,
                 convert_units = conversion.factor,
                 formula = ~size)
sizeTruncHR <- ds(data = bowhead_LT_Trunc, key = "hr", adjustment = NULL,
                 convert_units = conversion.factor,
                 formula = ~size)

# truncated data shows very different rankings
# hazard rate is preferred w/ size covariate
# then basic hanf-normal, then size + HN, finally base HR
summarize_ds_models(baseTruncHN,
                   baseTruncHR,

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sizeTruncHN,
sizeTruncHR, output = "plain")

# looks like the binned models have exactly the same fit as the raw data
par(mfrow = c(2,3))
plot(baseTruncHN, which=2, pl.col = adjustcolor("steelblue",0.5),border=NULL,
     lwd = 2,
     ylab = "Detection probability (g(x))", xlab = "Distance", las=1,
     main = "Base half-normal model")
# covariate model no longer assumes perfect detection of large-sized groups
plot(sizeTruncHN, which=2, pl.col = adjustcolor("steelblue",0.5),border=NULL,
     lwd = 2,
     ylab = "Detection probability (g(x))", xlab = "Distance", las=1,
     main = "Half-normal model with covariates")
plot(sizeTruncHR, which=2, pl.col = adjustcolor("steelblue",0.5),border=NULL,
     lwd = 2,
     ylab = "Detection probability (g(x))", xlab = "Distance", las=1,
     main = "Hazard rate model with covariates")
# covariate model still slightly better from visual QQ plot
gof_ds(baseTruncHN, main="Base Half-Normal Observed vs. Expected CDF",
      ks = TRUE)
gof_ds(sizeTruncHN, main="Covariate Half-Normal Observed vs. Expected CDF",
      ks = TRUE)
gof_ds(sizeTruncHR, main="Covariate Hazard Rate Observed vs. Expected CDF",
      ks = TRUE)

# output the models for consistency and processing speed
save(baseRawHN, baseRawHR, sizeRawHN, sizeRawHR,
     baseBinHN, baseBinHR, sizeBinHN, sizeBinHR,
     baseTruncHN, baseTruncHR, sizeTruncHN, sizeTruncHR,
     file="df-models.RData")

# break out abundance under both models by region
# generate abundance estimates for the best raw data models
region_table <- unique(bowhead_LT[,c("Region.Label", "Area")])
sample_table <- unique(bowhead_LT[,c("Region.Label",
                                     "Sample.Label",
                                     "Effort")])
observation_table <- unique(bowhead_LT[,c("object",
                                           "Region.Label",
                                           "Sample.Label")])

# raw model
baseRawHN_N <- dht(model = baseRawHN$ddf,
                  region.table = region_table,
                  sample.table = sample_table,
                  obs.table = observation_table)

# covariate model
# lower overall SE driven by Regions 2 and 9
# Regions 3, 11, 12, and 15 have increased SE
# abundance decreases in regions 2 and 9 while increasing elsewhere
# Regions 2 and 9 are the only sub-regions with size > 1 observations

```

```

sizeRawHN_N <- dht(model = sizeRawHN$ddf,
  region.table = region_table,
  sample.table = sample_table,
  obs.table = observation_table)
# repeat abundance estimation process for remaining models of interest
# base binned model
baseBinHN_N <- dht(model = baseBinHN$ddf,
  region.table = region_table,
  sample.table = sample_table,
  obs.table = observation_table)
# size covariate binned data model
sizeBinHN_N <- dht(model = sizeBinHN$ddf,
  region.table = region_table,
  sample.table = sample_table,
  obs.table = observation_table)
# base truncated data model
baseTruncHN_N <- dht(model = baseTruncHN$ddf,
  region.table = region_table,
  sample.table = sample_table,
  obs.table = observation_table)
# size covariate truncated data model
sizeTruncHN_N <- dht(model = sizeTruncHN$ddf,
  region.table = region_table,
  sample.table = sample_table,
  obs.table = observation_table)

# check variance using bootstraps rather than delta method
# delta-method approximation assumes independence between
# uncertainty in the detection function and variability in encounter rate

# takes ~3 minutes on my computer, likely longer for most
# check or delete the "cores = " if you don't have 10 cores on your computer
est.boot <- bootdht(model=sizeRawHN, flatfile=bowhead_LT,
  summary_fun=bootdht_Nhat_summarize,
  convert_units=conversion.factor, nboot=999, cores=10)

alpha <- 0.05
# the lower bound of our estimate is very poorly constrained
# 95% CI between 0 and 276
bootci <- quantile(est.boot$Nhat, probs = c(alpha/2, 1-alpha/2),
  na.rm=TRUE)

# plot generated bootstraps
par(mfrow = c(1,1))
hist(est.boot$Nhat, nc=30,
  main="Distribution of bootstrap estimates\nwithout model uncertainty",
  xlab="Estimated abundance")
abline(v=bootci, lwd=2, lty=2)

# repeat bootstraps w/ resample by region rather than by transect
# our upper CI increases slightly but the lower CI stays at 0
# est.bootStrata <- bootdht(model=bowhead.hn.null.size, flatfile=bowhead_LT,

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#           summary_fun=bootdht_Nhat_summarize,
#           resample_strata = TRUE,
#           convert_units=conversion.factor, nboot=999, cores=10)
#bootciStrata <- quantile(est.bootStrata$Nhat, probs = c(alpha/2, 1-alpha/2),
#           na.rm=TRUE)

# this following section is for transparency on full model fitting
# attempted with the original, minorly truncated data
# play around with some adjustment terms with the size model
# adjustment terms generally aren't included in covariates for whatever reason
# Cosine(2) is preferred but never monotonic so that's not it
#bowhead.hn.cos.size <- ds(data = bowhead_LT, key = "hn", adjustment = "cos",
#           convert_units = conversion.factor,
#           formula = ~size,
#           nadj = c(1,2,3,4))
# Hermite(4) is preferred but is non-monotonic; Hermite(4,6) is not as bad
#bowhead.hn.herm.size <- ds(data = bowhead_LT, key = "hn", adjustment = "herm",
#           convert_units = conversion.factor,
#           formula = ~size,
#           nadj = c(2,3,4))
# Poly(4,6) ends up being non-monotonic, so leaving it at Polynomial(4)
#bowhead.hn.poly.size <- ds(data = bowhead_LT, key = "hn", adjustment = "poly",
#           convert_units = conversion.factor,
#           formula = ~size,
#           nadj = c(1,2,3,4))
# now repeat for half-normal
# non-monotonic and  $g(x) > 1$  with a cosine adjustment
#bowhead.hr.cos.size <- ds(data = bowhead_LT, key = "hr", adjustment = "cos",
#           convert_units = conversion.factor,
#           formula = ~size,
#           nadj = c(1,2,3,4))
#Hermite(4,6) surprisingly matches everything
#bowhead.hr.herm.size <- ds(data = bowhead_LT, key = "hr", adjustment = "herm",
#           convert_units = conversion.factor,
#           formula = ~size,
#           nadj = c(2,3,4))
#Polynomial adjustments don't match monotonicity
#bowhead.hr.poly.size <- ds(data = bowhead_LT, key = "hr", adjustment = "poly",
#           convert_units = conversion.factor,
#           formula = ~size,
#           nadj = c(1,2,3,4))

```

References

- [1] S. L. Rekdal *et al.*, “Trends in bowhead whales in west greenland: Aerial surveys vs. Genetic capture-recapture analyses,” *Marine Mammal Science*, vol. 31, pp. 133–154, Jul. 2014, doi: [10.1111/mms.12150](https://doi.org/10.1111/mms.12150).
- [2] S. T. Buckland, E. A. Rexstad, T. A. Marques, and C. S. Oedekoven, *Distance sampling: Methods and applications*. Springer International Publish, 2015.
- [3] D. L. Borchers, W. Zucchini, M. P. Heide-Jørgensen, A. Cañadas, and R. Langrock, “Using hidden markov models to deal with availability bias on line transect surveys,” *Biometrics*, vol. 69, pp. 703–713, Jul. 2013, doi: [10.1111/biom.12049](https://doi.org/10.1111/biom.12049).