# Interpretable Machine Learning Approach - Elucidate Messenger RNA Subcellular Localization Codes

COMP 680: Mining Biological Sequences — Mathieu Blanchette, Winter 2017

Mansha Imtiyaz (260712985) mansha.imtiyaz@mail.mcgill.ca

Atefeh Mohajeri at.mohajeri@gmail.com

Abstract—In this report we introduce the work of Ribeiro et al. [1] which presents a model-agnostic interpretation tool called LIME. LIME enables us to explain individual predictions of a Machine Learning technique by learning an interpretable model locally around the prediction. We develop a pipeline for predicting the localization patterns of mRNAs in Drosophila melanogaster embryos, and use LIME to identify RBPs involved in this process.

#### I. INTRODUCTION

Subcellular localization of mRNAs have been known as a mechanism for locally producing proteins. Nevertheless, its only in recent studies that subcellular localization has emerged as the main mechanism underlying the localization of proteins in many polarized cells. Lecuyer et al [2] has discovered that in Drosophila embryo, more than 70% of the 2314 expressed transcripts were sub cellularly localized. All fundamental developmental cell processes such as asymmetric cell division or embryonic patterning highly depend on cell polarization, which in turn depends on cytoplasmic mRNA localization.

There are several reasons why transporting mRNA might be preferable to transporting proteins. It reduces the cost of transport since one mRNA can produce several proteins once it has reached the target region. Also, transporting mRNA, prevents the corresponding proteins from unwanted interactions on their way to the target site. Another benefit of the sub cellular localization of mRNA is facilitating production of proteins with high concentration which is beneficial for integration of proteins in macromolecular complexes.

There are three distinct mechanisms involved in the process of distribution of mRNAs within cells: localized protection from degradation, uniform diffusion coupled with local entrapment and directed transport. We are interested in the third mechanism. RNA binding proteins (RBPs) play an important role in the active transport of mRNA. RBPs bind to target mRNAs through cis-localization elements (or zipcodes), and make a complex that transports the mRNA. Furthermore, the structure of an RNA molecule plays key roles in determining the accessibility of these binding sites. Despite their importance in gene regulation, the binding sites for most RBPs involved in mRNA localization are not well characterized.

In this report, we develop a pipeline for predicting the localization patterns of mRNAs in Drosophila melanogaster embryos, and use interpretable machine learning model to identify RBPs involved in this process. We present the work of Ribeiro *et al.* [1] who propose LIME, an explanation technique which attempts to provide interpretable explanations for predictions of *any* classifier.

LIME approaches the problem by learning an *interpretable model* (called the *explainer*) in the vicinity (in feature space) of the prediction generated by the ML model. This report employs LIME to reproduce the results presented by the authors and to further evaluate the methodology on the given dataset.

#### II. METHOD OVERVIEW

In this section, we introduce the notion of *explanation* and formalize its definition by outlining the algorithm discussed in [1]. A high-level description of the LIME pipeline is given in Figure 1. LIME stands for Local Interpretable Model-Agnostic Explanations.

# A. Overview of an Explainer

There are four important desired characteristics of a Posthoc explainer. First and foremost, it must be interpretable, meaning that it should provide a qualitative understanding between the input and the output. Note that the notion of interpretability directly depends on the target audience, as one type of explainer could be more easily interpreted by machine learning experts than by laymen. Second, it should conform to the notion of *local fidelity*. This implies that we require an explainer to be locally faithful in order for it to be meaningful. Put differently, it must comply to how the model behaves in the vicinity of the instance being predicted. It should be noted that local fidelity does not imply global fidelity. Third, the explainer should be model-agnostic, i.e., it should treat any ML model as a black box. This ensures that the tool will be able to explain both the contemporary state-of-the-art models as well as future, more complex classifiers. Finally, we want an explainer to provide a global perspective on how the model behaves. This enables us to evaluate the performance of a model and conduct comparisons between different models before real-world deployments.

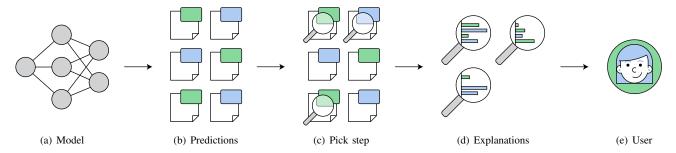


Fig. 1. LIME PIPELINE. A complex model (a) is used to predict labels from a given dataset (b). Then, an explainer is learned to choose representative instances (c) to explain. The explainer provides explanations (d) to the user, who takes the final decision (e).

# B. An Algorithm for Explaining

More formally, an *explainer* is defined as a model  $g \in \mathcal{G}$  where  $\mathcal{G}$  is the set of potentially interpretable models such as linear models and decision trees. Given an original instance  $x \in \mathbb{R}^d$  to be explained, we first convert it to its *interpretable representation*. An *interpretable representation* is any representation that is intuitive for humans, *i.e.*, it should conform to the notion of *decomposability*. LIME denotes this representation as a binary vector  $\varphi(x) = \widehat{x} \in \{0,1\}^{\widehat{d}}$ . In the case of text classification this denotes the presence or absence of a word while for image classification it represents presence or absence of a contiguous patch of pixels (a superpixel).

Though models like linear models and decision trees are considered interpretable, a large dimensional model can lead to the degradation in its simulability. Hence the authors define  $\Omega(g)$  to be a measure of complexity of the explainer. This can be the depth of a tree for decision trees, or the number of nonzero weights for a linear model.

Now, let  $f:\mathbb{R}^d\to\mathcal{C}$  be a classifier over a set of classes  $\mathcal{C}$ , to be explained. Further let  $\pi_x(z)$  be a proximity metric between x and z that encapsulates the notion of locality around x. Given these definitions, we set the notion of local fidelity by defining a loss function  $L(f,g,\pi_x)$  which represents the explainer's faithfulness to the original model in the locality of the x as defined by  $\pi_x(z)$ . To obtain an interpretable explanation, we minimize this loss function while also ensuring simulability by keeping  $\Omega(g)$  low. This is given by

$$\xi(x) = \underset{g \in \mathcal{G}}{\operatorname{arg \, min}} \ L(f, g, \pi_x) + \Omega(x). \tag{1}$$

To approximate L, we sample around  $\widehat{x}$  to get a set of perturbed instances  $\widehat{z} \in \{0,1\}^d$ . The perturbed samples  $\widehat{z}$  are then transformed to the original representation of the instance to get  $z \in \mathbb{R}^d$ . This is done by letting  $\widehat{z}$  act as a binary mask over x. f(z) now denotes the labels generated by the original classifier for the perturbed samples. Assuming we let  $\pi_x(z) = \exp(-d(x,z)^2/\sigma^2)$  be an exponential kernel with variance  $\sigma^2$  defined on some distance function d (e.g., cosine distance for text), then the locality-aware loss L is given by

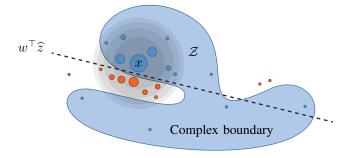


Fig. 2. INTUITION BEHIND LIME. Points are sampled around x and a new, simpler decision boundary is learned as an explanation.

$$L(f, g, \pi_x) = \sum_{z, \widehat{z} \in \mathcal{Z}} \pi_x(z) (f(z) - g(\widehat{z}))^2,$$
 (2)

where  $\mathcal{Z}$  is the perturbed samples set with associated labels. LIME currently defines g as a linear classifier, that is,  $g(\widehat{z}) = w^{\top}\widehat{z}$  for some weight vector w. This definition can however be extended to include other explainers such as decision trees. Figure 2 illustrates the above procedure with a linear explainer.

As mentioned earlier, in the case of text classification the interpretable representation is denoted by the presence or absence of a word. Hence the complexity parameter  $\Omega(x)$  is specified by a limit K on the number of words to be accounted in the computation. In the of case images the authors use superpixels as the interpretable representation. Using the same notion the complexity parameter  $\Omega(x)$  now sets a limit on the number of superpixels to be considered during computation. This renders the Equation 1 intractable. Authors address this problem by introducing the K-Lasso algorithm, where  $\Omega(x)$  is approximated by selecting K features using the  $L_1$ -regularization (Lasso) and then learning the weights via least squares procedure. This describes the formulation of an explainer  $\xi(x) = (g \circ f)(x)$ .

The second part of the algorithm consists of providing the user with a global representation of a model. This is achieved via judiciously selecting multiple instances as representation of the model to be presented to the user. Put briefly, a Submodular Pick (SP) algorithm is defined to choose a diverse,

representative set of explanations to assist in understanding how a model behaves globally. This essentially translates into a max-coverage problem of size B (which denoted the user budget) which can be well-approximated by a greedy algorithm, offering a constant-factor approximation guarantee of 1-1/e to the optimum. For further details of the SP algorithm the reader is suggested to refer to [1].

# C. Algorithm Implementation

LIME is an open-source project available at [3]. Experiments are conducted by modifying the given framework in order to accommodate different classifiers and datasets. For text classification, the cosine distance for d is used and perturbations to documents are performed by randomly removing words. For image classification, the  $L^2$  (Euclidean) distance is employed, and perturbed instances are created by segmenting them into contiguous superpixels and randomly removing patches using [4]. This process is depicted in Figure 3.





(a) Original image

(b) Superpixel segments

Fig. 3. SUPERPIXEL ALGORITHM. Each of the n=30 patches corresponds to an interpretable component for LIME.

TensorFlow [5] is employed to easily obtain complex deep convolutional neural network architectures. For all other classifiers, scikit-learn [6] modules are used.

#### III. LIME ON BREAST CANCER TUMOR DATA SET

We begin with evaluation of LIME to explain predictions made on medical data using the breast cancer dataset from scikit-learn, which a copy of UCI ML Breast Cancer Wisconsin (Diagnostic) Dataset [7]. Medical applications such as this serves as a major motivation for explanation tools like LIME. It is often necessary to inspect the reasons of a particular prediction generated by a ML model considering the sensitive nature of the application. We illustrate such a scenario with the suggested evaluation.

The features are computed from a digitized image of a fine needle aspirate (FNA) of a breast mass. These describe characteristics of the cell nuclei present in the image scan. The dataset includes 569 instances and two classes: *malignant* and *benign*, with 212 and 357 instances, respectively. Ten core attributes are used as detailed in Table I. The mean, standard error, and "worst" or largest (mean of the three largest values) of these features are computed for each instance, amounting to a total of 30 features.

TABLE I UCI Breast Cancer Dataset Core Attributes

FEATURE	DESCRIPTION
Radius	Mean of distances from center to points
	on the perimeter
Texture	Standard deviation of grayscale values
Perimeter	Distance around the nuclear border
Area	Area of outlined nucleus
Smoothness	Local variation in radius lengths
Compactness	Perimeter <sup>2</sup> / area $-1.0$
Concavity	Severity of concave portions of the contour
Concave points	Number of concave portions of the contour
Symmetry	Symmetry of the outlined nucleus
Fractal dimension	Coastline approximation $-1.0$

Random Forests with default parameters and n=500 trees is used to model the data. Random Forests are chosen due to their complex nature of decision logic. This is a good example of the types of models LIME will have to explain in the field. We also use a second classifier, a Gaussian Naive Bayes classifier, to model the data. The training and test sets are divided in a ratio of 80/20. For explaining individual test cases, predicted instances for both classifiers are obtained and an explainer is learned on them to analyze which features are being used to generate the prediction.

The results for breast cancer tumors show that the algorithm is capable of identifying relevant features that contribute to a prediction. Here, *malignant* and *benign* cases are taken individually and an explainer is learned on the predictions. The results are shown in Figure 4.

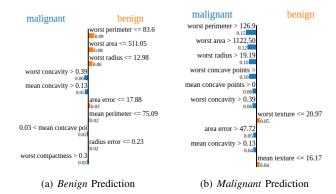


Fig. 4. EXPLAINING BENIGN AND MALIGNANT CASES.

For both the instances, worst concavity adds weight to a malignant prediction (> 0.39) but in the benign case, other features (worst perimeter, area, radius) add more weight to the categorization task.

One of the assumptions of using the explainer is that you need expert/background knowledge in order to interpret the results. When comparing the results with these in the literature [8], it was found that the results obtained are akin to implying that if used in a medical setting, providing explanations can increase the acceptance of automated systems.

#### IV. PREDICTING LOCALIZATION PATTERNS OF MRNAS

In this section, we describe the process to identify RBPs in predicting the localization patterns of mRNAs.

### A. Data Set Description

We ran our predictor on 6 different data sets containing 5 classes - Table II. Out of these, one dataset was imbalanced which resulted in high accuracy but low precision/recall. The rest of datasets contains 300 feature vectors and were balanced for both classes. A feature vector (n1, n2, , nm) is made for each gene, where coordinates correspond to number of times each RBP binding site is present in paired/unpaired regions.

Labels 0 and 1 indicate whether the RNAs bind or not. To make the predictions, training and validation set were divided in 80/20 for each of the classes.

TABLE II CLASSES IN THE DATASET PROVIDED

CLASSES		
Cortical Nuclei		
Perinuclear around pole cell nuclei		
RNA islands		
Yolk plasm enrichment		
Pole plasm		

#### B. Prediction

Random Forest Classifier was used and the results obtained on each of the dataset are given in Table III. Also, the ROC curve for all are given in the Appendix. As seen in the table, even though the accuracy was good for imbalanced dataset, recall was just 29.5%. For this reason, we run LIME on only balanced datasets, so that there is no bias in explaining the predictions.

TABLE III
ACCURACY, PRECISION AND RECALL

CLASSES	ACCURACY	PRECISION	RECALL
Unbalanced Cortical Nuclei	94.3	91.9	29.5 [6]
Cortical Nuclei	83.3	79.5	81.8
Perinuclear around pole cell nuclei	98.8	97.5	100
RNA islands	84.6	79.2	85.1
Yolk Plasm enrichment	79.2	74.9	85.8
Pole Plasm	80.12	75.8	86.28

# C. Explaining the Prediction - Identifying RBPs

To identify RBP's, we run explainer on individual instances. For this report, we are focusing on Cortical Nuclei class only. One of the observations was that the key feature vectors which motivated the prediction had quite less weight (e-50) for this dataset unlike other datasets which had a positive weight in the range 0-10 for each of the key feature vectors.

Everytime the explainer was run, top features changed (due to negligible weight). In order to get meaningful results, we ran the loop 10 times for Cortical nuclei class on the true positive predictions and made a note of the features which were repeated as shown in Table IV, We identify these as RBPs essential for making the prediction in this process.

TABLE IV
KEY RBPS IDENTIFIED FOR SOME OF THE TRUE POSITIVE PREDICTIONS
ON CORTICAL NUCLEI CLASS

Name	RBP's identified	
FBtr0075329-apiMel4.Group1	Rna15, ybx2-a, SNRPA	
FBtr0075329-dm6.chr3L	SRSF2, SNRPA	
FBtr0075329-droBip2.KB463974	SNRPA, SNRPA, SRSF1, SRSF2, SNRPA	
FBtr0075329-droEug2.KB465257	SRSF2, SNRPA, ybx2-a	
FBtr0075329-droKik2.KB459701	SNRPA , SNRPA, SNRPA	
FBtr0075329-droMoj3.scaffold_6680	SRSF2, SRSF7, ELAVL2, SRSF2, ybx2-a	
FBtr0075329-droPer1.super_29	SRSF7, SNRPA	

The list of key feature vectors for all true positive cases which were selected by Lime algorithm have been attached in the appendix for all true positives of class Cortical Nuclei.

# V. FUTURE WORK

Next steps would be to compare the key RBPs identified by this process with that of Lasso classifier and to see if we can find some interesting findings in that.

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#### **APPENDIX**

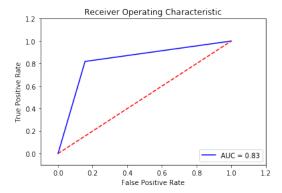


Fig. 5. ROC CURVE FOR CORTICAL NUCLEI CLASS

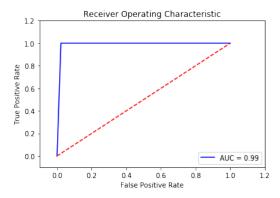


Fig. 6. ROC Curve For Perinuclear around pole cell nuclei Class

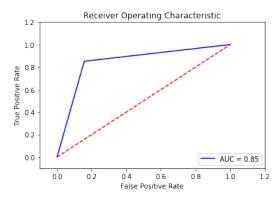


Fig. 7. ROC CURVE FOR RNA ISLAND CLASS

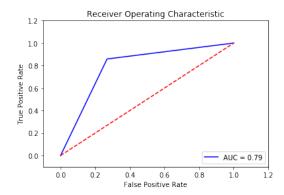


Fig. 8. ROC CURVE FOR YOLK PLASM ENRICHMENT CLASS

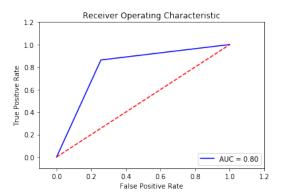


Fig. 9. ROC CURVE FOR POLE PLASM CLASS

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0.00', 0.0)
ID: 118
Name: FBtr0088676-droKik2.KB459872
[('285 SNRPA <= 0.00', 0.0), ('263 IGF2BP1 <= 0.00', 0.0), ('284 SNRPA <=
0.00', \overline{0}.0), ('283\_SNRPA <= 0.00', \overline{0}.0)]
ID: 122
Name: FBtr0088676-droSuz1.KI420922
[('249 \text{ ybx2-a} \le 0.00', 0.0)]
ID: 124
Name: FBtr0088676-droWil2.CH963719
[('249 ybx2-a <= 0.00', 0.0), ('278 SRSF7 <= 0.00', 0.0), ('279 SRSF2 <=
0.00', 0.0), ('274 \text{ NOVA2} \le 0.00', \overline{0.0})]
ID: 127
Name: FBtr0080389-dm6.chr2L
[('263__IGF2BP1 <= 0.00', 0.0), ('266__Rna15 <= 0.00', 0.0)]
ID: 135
Name: FBtr0080389-droFic2.KB457472
[('256__SRSF2 <= 0.00', 0.0), ('285__SNRPA <= 0.00', 0.0), ('284__SNRPA <=
0.00', 0.0), ('283 SNRPA \le 0.00', 0.0)]
ID: 140
[('267__Rna15 <= 0.00', 0.0), ('258__su(s) <= 0.00', 0.0), ('256__SRSF2 <= 0.00', 0.0)]
ID: 141
Name: FBtr0080389-droPse3.chr4 CH379060 3 random
[('256__SRSF2 <= 0.00', 0.0), ('274__NOVA2 <= 0.00', 0.0), ('278__SRSF7 <=
0.00', 0.0), ('266\_Rna15 \le 0.00', 0.0)]
ID: 144
Name: FBtr0080389-droSim1.chr2L
[('283 SNRPA <= 0.00', 0.0)]
Name: FBtr0080389-droVir3.scaffold 12963
[('258\_su(s) \le 0.00', 0.0), ('26\overline{6}\_Rna15 \le 0.00', 0.0), ('249\_ybx2-a \le 0.00', 0.0)]
0.00', 0.0)]
ID: 150
Name: FBtr0300090-droSec1.super 20
[('256 SRSF2 <= 0.00', 0.0), ('263 IGF2BP1 <= 0.00', 0.0), ('275 ELAVL2
\leftarrow 0.00', 0.0), ('279\_SRSF2 \leftarrow 0.00', 0.0)]
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ID: 153
Name: FBtr0075167-droAlb1.JH856983
[('258 \text{ su}(s) \le 0.00', 0.0), ('263 \text{ IGF2BP1} \le 0.00', 0.0), ('279 \text{ SRSF2} \le 0.00', 0.0)]
0.00', 0.0)
ID: 154
Name: FBtr0075167-droAna3.scaffold 13337
[('256\_SRSF2 \le 0.00', 0.0), ('274 NOVA2 \le 0.00', 0.0), ('249 ybx2-a \le 0.00', 0.0)]
0.00', 0.0)
ID: 155
Name: FBtr0075167-droBia2.KB462646
[('285\_SNRPA <= 0.00', 0.0), ('289\_SRSF1 <= 0.00', 0.0), ('284\_SNRPA <= 0.00', 0.0)]
0.00', 0.0), ('283 SNRPA <= 0.00', 0.0)]
ID: 162
Name: FBtr0075167-droKik2.KB459821
[('256\_SRSF2 \le 0.00', 0.0), ('249\_ybx2-a \le 0.00', 0.0), ('284\_SNRPA \le 0.00', 0.0)]
0.00', \overline{0}.0), ('283 \text{ SNRPA} <= 0.00', \overline{0}.0)]
ID: 163
Name: FBtr0075167-droMir2.chrXR
[('285 SNRPA <= 0.00', 0.0), ('283 SNRPA <= 0.00', 0.0), ('289 SRSF1 <=
0.00', 0.0), ('284 \text{ SNRPA} \le 0.00', 0.0), ('266 \text{ Rna15} \le 0.00', 0.0)]
ID: 165
Name: FBtr0075167-droPer1.super 20
[('274 \text{ NOVA2} \le 0.00', 0.0)]
ID: 166
Name: FBtr0075167-droPse3.chrXR CH379069 3 random
[('256 SRSF2 \le 0.00', 0.0), ('279 SRSF2' \le 0.00', 0.0), ('249 ybx2-a \le 0.00', 0.0)]
0.00', 0.0)]
ID: 172
Name: FBtr0075167-droYak3.chr3L
[('256 SRSF2 <= 0.00', 0.0), ('283 SNRPA <= 0.00', 0.0), ('274 NOVA2 <=
0.00', 0.0)
ID: 173
Name: FBtr0075218-dm6.chr3L
[('256 SRSF2 <= 0.00', 0.0), ('284 SNRPA <= 0.00', 0.0), ('283 SNRPA <=
0.00', 0.0)
ID: 175
Name: FBtr0075218-droAna3.scaffold 13337
[('256__SRSF2 <= 0.00', 0.0), ('274__NOVA2 <= 0.00', 0.0), ('279__SRSF2 <=
0.00', 0.0), ('283\_SNRPA <= 0.00', 0.0)]
ID: 177
Name: FBtr0075218-droBip2.KB464219
[('256 SRSF2 <= 0.00', 0.0), ('279 SRSF2 <= 0.00', 0.0), ('283 SNRPA <=
0.00', 0.0)]
ID: 179
Name: FBtr0075218-droEre2.scaffold 4784
[('278 SRSF7 \leftarrow 0.00', 0.0), ('279\_SRSF2 \leftarrow 0.00', 0.0), ('266\_Rna15 \leftarrow
0.00', 0.0)
ID: 185
Name: FBtr0075218-droMoj3.scaffold 6680
[('267\_Rna15 \le 0.00', 0.0), ('249\_ybx2-a \le 0.00', 0.0), ('256 SRSF2 \le 0.00', 0.0)]
0.00', \overline{0.0}]
ID: 191
Name: FBtr0075218-droSuz1.KI420009
[('278\_SRSF7 \leftarrow 0.00', 0.0), ('283\_SNRPA \leftarrow 0.00', 0.0)]
ID: 193
Name: FBtr0075218-droYak3.chr3L
[('285__SNRPA <= 0.00', 0.0), ('278__SRSF7 <= 0.00', 0.0), ('284__SNRPA <=
0.00', 0.0), ('283 SNRPA <= 0.00', 0.0)]
ID: 195
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Name: FBtr0077832-droAna3.scaffold 12916
[('258 \text{ su}(s) \le 0.00', 0.0), ('278 \text{ SRSF7} \le 0.00', 0.0), ('283 \text{ SNRPA} \le 0.00', 0.0)]
0.00', 0.0)]
ID: 196
Name: FBtr0077832-droBia2.KB462402
ID: 197
Name: FBtr0077832-droBip2.KB464241
[('285__SNRPA <= 0.00', 0.0), ('284__SNRPA <= 0.00', 0.0), ('283__SNRPA <=
0.00', \overline{0}.0)1
ID: 202
Name: FBtr0077832-droGri2.scaffold 15252
[('267 \text{ Rna}15 \leftarrow 0.00', 0.0), ('256 \text{ SRSF2} \leftarrow 0.00', 0.0), ('279 \text{ SRSF2} \leftarrow
0.00', 0.0), ('249_ybx2-a \le 0.00', 0.0)]
ID: 204
Name: FBtr0077832-droMir2.chr4
[('258 \text{ su}(s) \le 0.00', 0.0), ('256 \text{ SRSF2} \le 0.00', 0.0), ('278 \text{ SRSF7} \le 0.00', 0.0)]
0.00', 0.0), ('275 \text{ ELAVL2} \le 0.00', 0.0), ('249 \text{ ybx2-a} \le 0.00', 0.0)]
ID: 205
Name: FBtr0077832-droMoj3.scaffold 6500
[('256\_SRSF2 \leftarrow 0.00', 0.0), ('283\_SNRPA \leftarrow 0.00', 0.0), ('249\_ybx2-a \leftarrow 0.00', 0.0)]
0.00', 0.0), ('284 \text{ SNRPA} \le 0.00', 0.0), ('285 \text{ SNRPA} \le 0.00', 0.0)]
ID: 206
Name: FBtr0077832-droPer1.super 1
[('263 \text{ IGF2BP1} \le 0.00', 0.0), ('284 \text{ SNRPA} \le 0.00', 0.0)]
ID: 207
Name: FBtr0077832-droPse3.chr4 CH379060 3 random
[('256\_SRSF2 \leftarrow 0.00', 0.0), \overline{('263\_IGF2BP1 \leftarrow 0.00', 0.0), ('279 SRSF2 \leftarrow 0.00', 0.0)]
0.00', 0.0), ('249_ybx2-a \le 0.00', 0.0)]
ID: 209
Name: FBtr0077832-droSec1.super 121
[('274\_NOVA2 \le 0.00', 0.0), ('283\_SNRPA \le 0.00', 0.0)]
ID: 210
Name: FBtr0077832-droSim1.chr2L
[('249_ybx2-a \le 0.00', 0.0)]
ID: 212
Name: FBtr0077832-droVir3.scaffold 12963
[('256__SRSF2 <= 0.00', 0.0), ('284__SNRPA <= 0.00', 0.0), ('279__SRSF2 <=
0.00', 0.0), ('283 SNRPA <= 0.00', 0.0)]
ID: 214
Name: FBtr0077832-droYak3.chr2L
[('249\_ybx2-a \le 0.00', 0.0)]
ID: 216
Name: FBtr0343860-dm6.chr2L
0.00', 0.0), ('249 \text{ ybx2-a} \le 0.00', 0.0), ('283 \text{ SNRPA} \le 0.00', 0.0)]
ID: 218
Name: FBtr0343860-droAna3.scaffold 12916
[('256 \text{ SRSF2} \leftarrow 0.00', 0.0), ('279 \text{ SRSF2} \leftarrow 0.00', 0.0), ('249 \text{ ybx2-a} \leftarrow 0.00', 0.0)]
0.00', \overline{0}.0)
ID: 219
Name: FBtr0343860-droBia2.KB462819
[('258\_su(s) \le 0.00', 0.0), ('263\_IGF2BP1 \le 0.00', 0.0), ('256\_SRSF2 \le 0.00', 0.0)]
0.00', 0.0), ('279\_SRSF2 \le 0.00', 0.0), ('266\_Rna15 \le 0.00', 0.0),
('267\_Rna15 \le 0.\overline{00}', 0.0), ('249\_ybx2-a \le 0.\overline{00}', 0.0), ('278\_SRSF7 \le 0.\overline{00}', 0.0)
0.00', 0.0), ('274\_NOVA2 \le 0.00', 0.0)]
ID: 220
Name: FBtr0343860-droBip2.KB464214
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[('249 \text{ ybx2-a} \leftarrow 0.00', 0.0), ('284 \text{ SNRPA} \leftarrow 0.00', 0.0), ('283 \text{ SNRPA} \leftarrow
0.00', 0.0)
ID: 222
Name: FBtr0343860-droEre2.scaffold 4845
[('258 \text{ su}(s) \le 0.00', 0.0), ('278 \text{ SRSF7} \le 0.00', 0.0), ('266 \text{ Rna15} \le 0.00', 0.0)]
0.00', 0.0)
ID: 224
Name: FBtr0343860-droFic2.KB457391
[('267 \text{ Rna}15 \le 0.00', 0.0), ('258 \text{ su}(s) \le 0.00', 0.0), ('283 \text{ SNRPA} \le 0.00', 0.0)]
0.00', \overline{0}.0)
ID: 231
Name: FBtr0343860-droSec1.super 7
[('256 SRSF2 \le 0.00', 0.0), ('249 ybx2-a \le 0.00', 0.0), ('283 SNRPA \le 0.00', 0.0)]
0.00', 0.0)]
ID: 232
Name: FBtr0343860-droSim1.chr2L
[('266 Rna15 <= 0.00', 0.0), ('275 ELAVL2 <= 0.00', 0.0), ('283 SNRPA <=
0.00', \overline{0}.0)
ID: 238
Name: FBtr0086180-anoGam1.chr2L
[('256__SRSF2 <= 0.00', 0.0), ('278__SRSF7 <= 0.00', 0.0), ('285__SNRPA <=
0.00', 0.0), ('283_SNRPA \le 0.00', 0.0)]
ID: 244
Name: FBtr0086180-droEre2.scaffold 4929
[('285 SNRPA <= 0.00', 0.0), ('283 SNRPA <= 0.00', 0.0), ('284 SNRPA <=
0.00', 0.0), ('266 \text{ Rna15} \leftarrow 0.00', 0.0)]
ID: 245
Name: FBtr0086180-droEug2.KB465221
[('267 Rna15 <= 0.00', 0.0), ('256 SRSF2 <= 0.00', 0.0), ('275 ELAVL2 <=
0.00', 0.0)
ID: 246
Name: FBtr0086180-droFic2.KB457417
[('258\_su(s) \le 0.00', 0.0), ('256\_SRSF2 \le 0.00', 0.0), ('284\_SNRPA \le 0.00', 0.0)]
0.00', 0.0), ('283 SNRPA <= 0.00', 0.0)]
ID: 256
Name: FBtr0086180-droSuz1.KI420922
[('284 SNRPA \le 0.00', 0.0), ('283 SNRPA \le 0.00', 0.0)]
ID: 257
Name: FBtr0086180-droVir3.scaffold 10324
[('258 \text{ su}(s) \le 0.00', 0.0), ('279 \text{ SRSF2} \le 0.00', 0.0), ('283 \text{ SNRPA} \le 0.00', 0.0)]
0.00', 0.0)
ID: 262
Name: FBtr0089395-droAlb1.JH862323
[('285__SNRPA <= 0.00', 0.0), ('274__NOVA2 <= 0.00', 0.0), ('284__SNRPA <=
0.00', 0.0), ('289 \text{ SRSF1} \le 0.00', 0.0), ('283 \text{ SNRPA} \le 0.00', 0.0)]
ID: 265
Name: FBtr0089395-droBip2.KB464214
[('267\_Rna15 \le 0.00', 0.0), ('258\_su(s) \le 0.00', 0.0), ('283\_SNRPA \le 0.00', 0.0)]
0.00', 0.0)]
ID: 270
Name: FBtr0089395-droGri2.scaffold 15126
[('258 \text{ su}(s) \le 0.00', 0.0), ('275 \text{ ELAVL2} \le 0.00', 0.0), ('274 \text{ NOVA2} \le 0.00', 0.0)]
0.00', 0.0)
ID: 277
Name: FBtr0089395-droSec1.super 7
[('258 \text{ su}(s) \le 0.00', 0.0), ('249 \text{ ybx2-a} \le 0.00', 0.0)]
ID: 289
Name: FBtr0086301-droBia2.KB462718
[('266 \text{ Rna}15 \leq 0.00', 0.0)]
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ID: 291
Name: FBtr0086301-droEle2.KB458561
[('256\_SRSF2 \le 0.00', 0.0), ('249\_ybx2-a \le 0.00', 0.0)]
ID: 296
Name: FBtr0086301-droKik2.KB459686
[('256__SRSF2 <= 0.00', 0.0), ('263__IGF2BP1 <= 0.00', 0.0), ('275__ELAVL2
\leq 0.00^{\circ}, 0.0)
ID: 297
Name: FBtr0086301-droMir2.chr3
[('267 \text{ Rna}15 \le 0.00', 0.0), ('275 \text{ ELAVL}2 \le 0.00', 0.0)]
ID: 298
Name: FBtr0086301-droMoj3.scaffold 6496
[('258\_su(s) \le 0.00', 0.0), ('256\_SRSF2 \le 0.00', 0.0), ('283\_SNRPA \le 0.00', 0.0)]
0.00', 0.0), ('249_ybx2-a \le 0.00', 0.0), ('285 SNRPA \le 0.00', 0.0),
('284\_SNRPA <= 0.00', 0.0)]
ID: 301
Name: FBtr0086301-droRho2.KB451266
[('284 SNRPA <= 0.00', 0.0), ('266 Rna15 <= 0.00', 0.0)]
Name: FBtr0086301-droSec1.super 1
[('256\_SRSF2 \leftarrow 0.00', 0.0), ('274\_NOVA2 \leftarrow 0.00', 0.0)]
ID: 304
Name: FBtr0086301-droVir3.scaffold 12875
[('258\_su(s) \le 0.00', 0.0), ('283 SNRPA \le 0.00', 0.0), ('256 SRSF2 \le 0.00', 0.0)]
0.00', 0.0)]
ID: 307
Name: FBtr0086301-musDom2.KB857006
[('267__Rna15 <= 0.00', 0.0), ('278__SRSF7 <= 0.00', 0.0), ('275__ELAVL2 <=
0.00', 0.0), ('279 \text{ SRSF2} \le 0.00', 0.0)]
ID: 312
Name: FBtr0087939-droBia2.KB462460
[('267__Rna15 <= 0.00', 0.0), ('256__SRSF2 <= 0.00', 0.0), ('274__NOVA2 <=
0.00', \overline{0}.0)
ID: 314
Name: FBtr0087939-droFic2.KB457278
[('258 \text{ su(s)} <= 0.00', 0.0), ('263 \text{ IGF2BP1} <= 0.00', 0.0), ('266 \text{ Rna15} <= 0.00', 0.0)]
0.00', 0.0), ('274 \text{ NOVA2} \leq 0.00', 0.0)]
ID: 320
Name: FBtr0344852-dm6.chr2L
[('284 SNRPA <= 0.00', 0.0), ('283 SNRPA <= 0.00', 0.0)]
Name: FBtr0344852-droBip2.KB464229
[('258\_su(s) \le 0.00', 0.0), ('283\_SNRPA \le 0.00', 0.0), ('256\_SRSF2 \le 0.00', 0.0)]
0.00', 0.0), ('249 \text{ ybx2-a} <= 0.00', 0.0)]
ID: 330
Name: FBtr0344852-droMoj3.scaffold 6500
[('256 SRSF2 <= 0.00', 0.0), ('274 NOVA2 <= 0.00', 0.0)]
ID: 331
Name: FBtr0344852-droSec1.super 7
[('283 \text{ SNRPA} \leftarrow 0.00', 0.0), ('266 \text{ Rna15} \leftarrow 0.00', 0.0)]
ID: 332
Name: FBtr0344852-droSim1.chr2L
[('267 Rna15 \le 0.00', 0.0)]
ID: 340
Name: FBtr0079168-droEle2.KB458409
[('249\_ybx2-a \le 0.00', 0.0), ('279\_SRSF2 \le 0.00', 0.0), ('266 Rna15 \le 0.00', 0.0)]
0.00', 0.0)
ID: 341
Name: FBtr0079168-droEre2.scaffold 4929
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ID: 342
Name: FBtr0079168-droEug2.AFP002004873
[('249 ybx2-a <= 0.00', 0.0), ('284 SNRPA <= 0.00', 0.0), ('283 SNRPA <=
0.00', 0.0)]
ID: 343
Name: FBtr0079168-droFic2.KB457397
[('256 SRSF2 <= 0.00', 0.0), ('263 IGF2BP1 <= 0.00', 0.0), ('275 ELAVL2
\leftarrow 0.00^{\circ}, 0.0), ('249 \text{ ybx2-a} \leftarrow 0.00^{\circ}, 0.0)]
ID: 347
Name: FBtr0079168-droPer1.super 10
[('267_Rna15 <= 0.00', 0.0), ('256_SRSF2 <= 0.00', 0.0), ('263_IGF2BP1 <=
0.00', 0.0)
ID: 350
Name: FBtr0079168-droSim1.chr2L
[('278 SRSF7 \le 0.00', 0.0)]
ID: 351
Name: FBtr0079168-droSuz1.KI419149
[('256 SRSF2 \le 0.00', 0.0)]
ID: 352
Name: FBtr0079168-droVir3.scaffold 12963
[('258\_su(s) \le 0.00', 0.0), ('256\_SRSF2 \le 0.00', 0.0), ('283\_SNRPA \le 0.00', 0.0)]
0.00', 0.0), ('267_Rna15 <= 0.00', 0.0), ('249_ybx2-a <= 0.00', 0.0),
('278 \text{ SRSF7} \le 0.\overline{00}', 0.0)]
ID: 353
Name: FBtr0079168-droWil2.CH963857
[('279\_SRSF2 \leftarrow 0.00', 0.0), ('283\_SNRPA \leftarrow 0.00', 0.0), ('289\_SRSF1 \leftarrow
0.00', 0.0), ('285\_SNRPA \le 0.00', 0.0), ('284\_SNRPA \le 0.00', 0.0),
('274 \text{ NOVA2} \le 0.\overline{00}', 0.0)]
ID: 356
Name: FBtr0300085-dm6.chrX
[('256 \text{ SRSF2} \leftarrow 0.00', 0.0), ('278 \text{ SRSF7} \leftarrow 0.00', 0.0)]
ID: 357
Name: FBtr0083417-anoGam1.chr2L
[('249 \text{ ybx2-a} \leftarrow 0.00', 0.0), ('275 \text{ ELAVL2} \leftarrow 0.00', 0.0), ('284 \text{ SNRPA} \leftarrow
0.00', 0.0), ('283 SNRPA <= 0.00', 0.0)]
ID: 361
Name: FBtr0083417-droAna3.scaffold 13340
[('263__IGF2BP1 <= 0.00', 0.0), ('266__Rna15 <= 0.00', 0.0)]
ID: 362
Name: FBtr0083417-droBia2.KB462558
[('285\_SNRPA <= 0.00', 0.0), ('284\_SNRPA <= 0.00', 0.0), ('289\_SRSF1 <= 0.00', 0.0)]
0.00', 0.0), ('283 SNRPA <= 0.00', 0.0)]
ID: 371
Name: FBtr0083417-droSec1.super 12
[('249\_ybx2-a \le 0.00', 0.0), ('256\_SRSF2 \le 0.00', 0.0)]
ID: 374
Name: FBtr0070233-droAna3.scaffold 13117
[('249 ybx2-a <= 0.00', 0.0), ('266 Rna15 <= 0.00', 0.0), ('284 SNRPA <=
0.00', \overline{0.0}), ('283 \text{ SNRPA} \leftarrow 0.00', \overline{0.0})]
ID: 375
Name: FBtr0070233-droBia2.KB462463
[('279__SRSF2 <= 0.00', 0.0), ('284__SNRPA <= 0.00', 0.0), ('283__SNRPA <=
0.00', \overline{0}.0)
ID: 377
Name: FBtr0070233-droEle2.KB458387
[('256__SRSF2 <= 0.00', 0.0), ('266__Rna15 <= 0.00', 0.0), ('275__ELAVL2 <=
0.00', 0.0), ('249 ybx2-a <= 0.00', 0.0)]
ID: 381
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Name: FBtr0070233-droMir2.chrXR
[('285 SNRPA <= 0.00', 0.0), ('283 SNRPA <= 0.00', 0.0), ('279 SRSF2 <=
0.00', 0.0), ('266 \text{ Rna15} \leq 0.00', 0.0)]
ID: 382
Name: FBtr0070233-droPer1.super 11
[('258_su(s) <= 0.00', 0.0), ('275_ELAVL2 <= 0.00', 0.0), ('279_SRSF2 <=
0.00', 0.0), ('256 \text{ SRSF2} \leq 0.00', 0.0)]
ID: 384
Name: FBtr0070233-droRho2.KB451367
[('283 SNRPA <= 0.00', 0.0), ('289 SRSF1 <= 0.00', 0.0), ('285 SNRPA <=
0.00', 0.0), ('278\_SRSF7 \le 0.00', 0.0), ('284\_SNRPA \le 0.00', 0.0),
('274 \text{ NOVA2} \le 0.00', 0.0)]
ID: 389
Name: FBtr0334286-dm6.chr3R
[('275__ELAVL2 <= 0.00', 0.0), ('284__SNRPA <= 0.00', 0.0)]
ID: 391
Name: FBtr0334286-droAna3.scaffold 13340
[('266 \text{ Rna}15 \leq 0.00', 0.0)]
ID: 392
Name: FBtr0334286-droBia2.KB462814
ID: 393
Name: FBtr0334286-droBip2.KB464355
[('267 Rna15 <= 0.00', 0.0), ('256 SRSF2 <= 0.00', 0.0), ('263 IGF2BP1 <=
0.00', 0.0), ('275 \text{ ELAVL2} <= 0.00', 0.0)]
ID: 394
Name: FBtr0334286-droEle2.KB457684
[('256 SRSF2 \le 0.00', 0.0), ('274 NOVA2 \le 0.00', 0.0), ('249 ybx2-a \le 0.00', 0.0)]
0.00', 0.0)
ID: 396
Name: FBtr0334286-droEug2.KB464661
[('258\_su(s) \le 0.00', 0.0), ('283\_SNRPA \le 0.00', 0.0), ('275\_ELAVL2 \le
0.00', 0.0), ('284 SNRPA <= 0.00', 0.0), ('266 Rna15 <= 0.00', 0.0)]
ID: 397
Name: FBtr0334286-droFic2.KB457554
[('279 SRSF2 \le 0.00', 0.0)]
ID: 401
Name: FBtr0334286-droMoj3.scaffold 6540
[('249 \text{ ybx2-a} \leftarrow 0.00', 0.0), ('283 \text{ SNRPA} \leftarrow 0.00', 0.0)]
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