# scDCCA: Deep contrastive clustering for single-cell RNA-seq data based on auto-encoder network

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#### **Abstract**

- ► Task: Cell clustering in scRNA-seq analysis.
- Technical Challenges:
  - High dimensionality, noise, and significant sparsity of scRNA-seq data.
  - Limitations of previous methods.

#### Challenge Key Insights:

- Intrinsic properties of cells.
- Relationship among cells.

#### ► Technical Contributions:

- Denoising Zero-Inflated Negative Binomial (ZINB) model-based auto-encoder.
- Dual contrastive learning module for pairwise proximity of cells.
- Joint feature learning and clustering in an end-to-end manner.

#### Experiment:

- Outperform 8 methods on 14 real datasets.
- ▶ Metrics: Accuracy, generalizability, scalability, and efficiency.
- Cell visualization and biological analysis.



## Introduction: Background and Challenges

- Task and Application:
  - Characterize cell types in scRNA-seq.
- ► Technical Challenges for Previous Methods:
  - ► Methods without prior information:
    - SIMLR: Multiple kernel.
    - Seurat: Louvain algorithm on KNN.
    - SHARP: Ensemble random projection-based.
    - scHFC: Fuzzy C Mean and Gath-Geva algorithms.
  - Limitations of these methods:
    - ▶ Dimension reduction (e.g., PCA) may lose vital information.
    - Similarity matrices fail to reflect cell similarity.
    - High computational and time cost.

#### Introduction: Advanced Methods and Limitations

- Deep Clustering Methods:
  - scGMAI, scCCESS: Auto-encoder, minimize MSE.
  - DCA: Auto-encoder with ZINB.
  - scDeepCluster: Combines DCA and DEC.
- Focus on data only, neglecting cell relationships.
- ► Cell Relations Considered Methods:
  - Graph-based methods:
    - ► GraphSCC, scDSC, scGNN.
    - Limitation: Mixed clustering results.
  - Contrast-sc:
    - Ignores the characteristics of the data itself.
    - Impact clustering performance as dividing into 2 stages: embedding and clustering

### Introduction: Technical Contributions of scDCCA

- Dual contrastive learning module to acquire pairwise cell proximity.
- Denoising ZINB auto-encoder for intrinsic feature representation.
- ▶ End-to-end training with clustering optimization.

#### Method

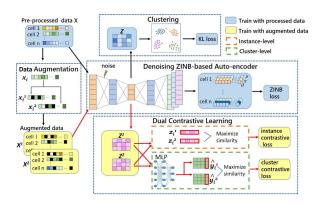


Figure: scDCCA

#### Method Overview

#### **▶** Four Components:

- Data augmentation for contrastive learning.
- Denoising ZINB-based auto-encoder.
- Dual contrastive learning module.
- Clustering module.

#### ► Training Process:

Pre-train with auto-encoder to capture clustering-friendly embeddings.

$$L_{pre-train} = \min (L_{ZINB} + \alpha L_{ins})$$

Fine-tune the embeddings with the cluster-level contrastive loss (Lcc) and cluster data with Kullback–Leibler (KL) loss (Ldec) to achieve intra-cluster compactness and inter-cluster separation.

$$L_{train} = \min (L_{ZINB} + \beta L_{cc} + \gamma L_{dec})$$

#### Data Processing:

Log normalization and gene selection using SCANPY.



## Denoising Auto-Encoder Embedding Module

- ► Corruption with Gaussian noise: Add random Gaussian noise to the input to enhance feature robustness.
- ▶ Encoder-Decoder: The encoder maps input to a latent space, and the decoder reconstructs it. This learns essential features.
- ► Loss Function: Minimize reconstruction loss using the ZINB loss with three parameters: mean, dispersion, and coefficient:

$$L = -\log P(\text{data}|\text{model parameters}).$$

$$L_{ZINB} = \sum_{ij} -\log \left(ZINB\left(x_{ij}|\pi_{ij}, \mu_{ij}, \theta_{ij}\right)\right)$$

## Data Augmentation Module

- ▶ Purpose: Enhance contrastive learning by diversifying training data without labeled examples.
- Positive/Negative Pairs: Construct positive pairs (similar) and negative pairs (dissimilar).
- ▶ **Augmentation:** For each input cell  $x_i$ , create two augmented views  $x_i^1$  and  $x_i^2$  by randomly masking genes.

## **Dual Contrastive Learning Module**

- ▶ Cell Relationships: Learn relationships among cells to improve intra-cluster compactness and inter-cluster separation.
- ▶ Loss in Latent Space: Compute losses at both instance and cluster levels in the ZINB latent space to optimize clustering.

## Experiment

- Comparison: scDCCA outperforms 8 state-of-the-art methods.
- Batch size effect:
  - Suitable batch size for each dataset is related to its sample size.
- Ablation Studies:
  - Instance-level and cluster-level contrastive learning both critical.
- Biological Analysis:
  - Cell annotation, KEGG pathway, and DEG analysis.
- Robustness: down-sampling and change ratio of masked genes
- Scalability and Efficiency:
  - Handles large datasets.
  - Reasonable runtime and memory usage.



## Experiment: Comparisons on 3 metrics

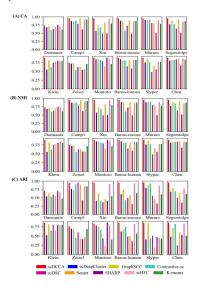


Figure: Clustering performance

## Experiment: Comparisons with cell visualization

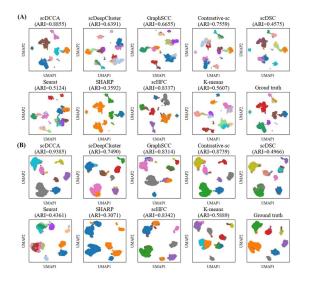


Figure: Cell visualization

## Experiment: Batch size effect

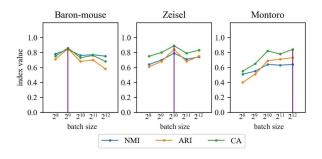


Figure: Batch size effect

## Experiment: Ablation Study

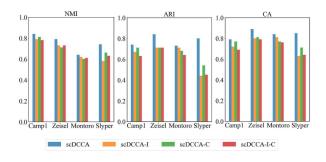


Figure: Ablation Study

## **Experiment: Robustness**

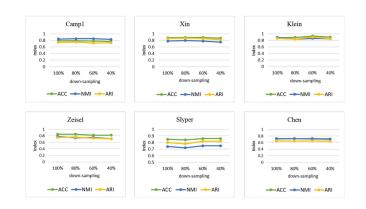


Figure: Robustness experiment

## Experiment: Biological analysis

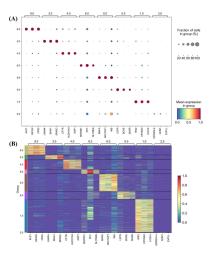


Figure: Biological analysis

#### Limitations

- Only uses genes as features, ignoring gene relationships.
- Future work:
  - Integrate gene relationships.
  - Explore GNN for higher-order structural information.