# Report

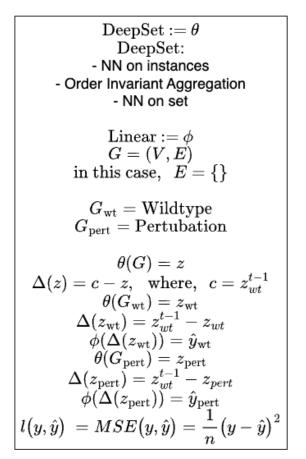
### **Experimental Summary**

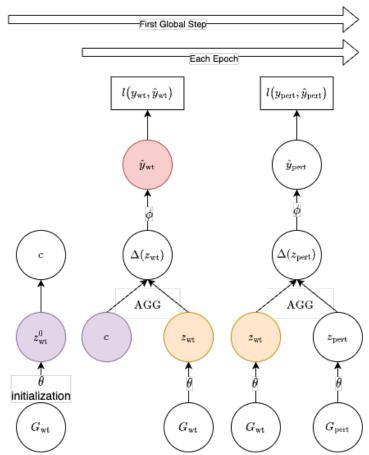
- Total data size is 1.3e7, but we train on 1.3e6 for faster training.
- All models use features from the FungalLM, I refer to these are as FungalCRE features since they describe the Fungal Cis-Regulatory-Elements. This model embeds sequences to size 768.
  - downstream are embeddings of the 300 bp downstream of the gene, (more precisely the CDS). This captures the terminator, and has been shown to be predictive of gene expression.
  - upstream are embeddings of the 1003 bp upstream of the gene, (more precisely the CDS). This captures the promoter region and should contain information about transcription factor binding. They have also shown to be predictive of gene expression.

### Motivation

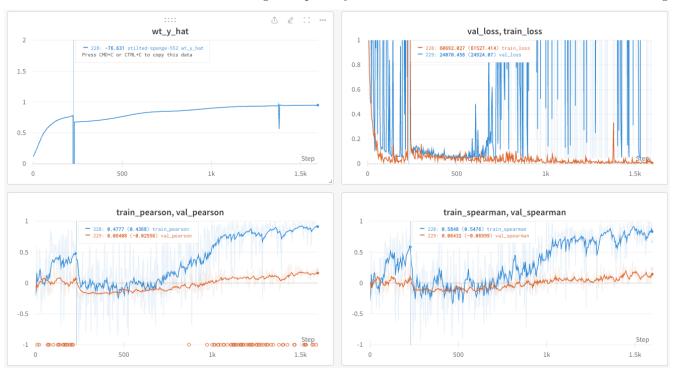
- Previous model development has shown that gene expression of pertubed strains (deletion mutants), can be used to predict strain growth with high correlations. Collecting gene expression data is expensive, if we can predict phenotypes (in this simple case growth), with features that can be computed from genomic sequence alone, this would be very useful. We would like to start with growth as there are good benchmarks but there are other global labels that we would like to predict. For instance cell morphology  $((e_1, e_2))$  of an ellipse).
- We model data structure as a graph, because we might do do inference at different levels of the cell. For instance, we have expression data paired with growth data for a gene deletions. We could train the node level on the expression data, the global level on the fitness data, and see if we can do any inference on expression for different gene deletions. Capturing major trends would be sufficient. One important caveat is that to maintain the ability to do any inference on nodes, or to backpropagate through nodes, we need a representation for the entire genome, not just the nodes that were removed. This poses a difficult challenge because the engineering edits to genomes are often small perturbations to the entire graph, we are talking 1-3 nodes in a 6000 node graph. To account for this I've attempted to learn a wildtype different embedding.

### Wildtype Difference Embedding





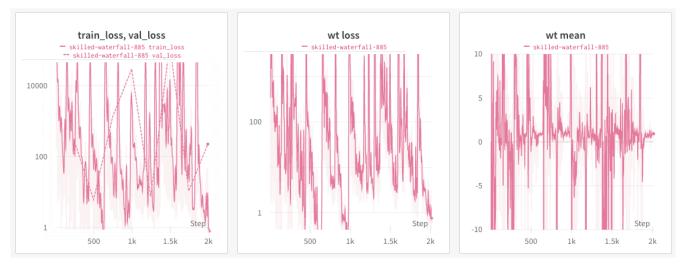
This model has a lot of instabilities in training. The primary issue seems to be the stabilization of the wt embedding.



When there is stabilization in the wt\_y\_hat the model starts to learn some and correlation starts to rise. As soon as we get another instability the model has to reset.

### Wildtype Difference Embedding - Gradient Norm Clipping

To try stabilize learning we have tried adding gradient norm clipping.

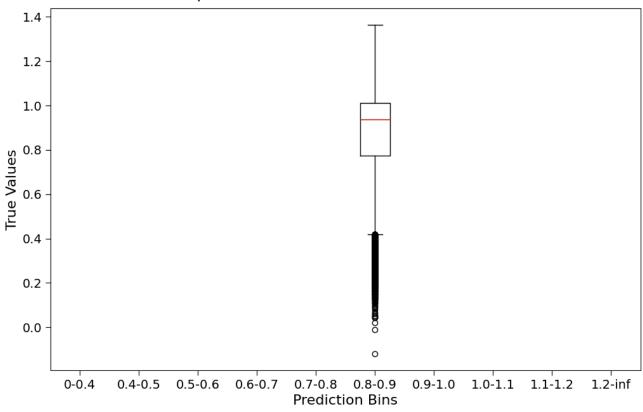


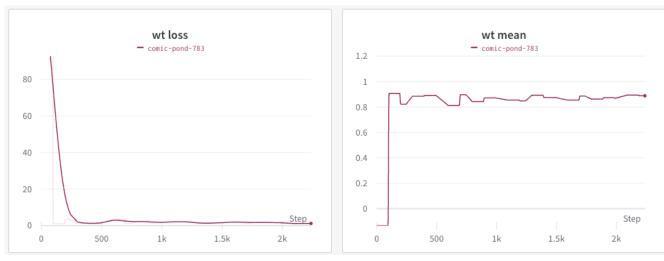
The wt\_mean is the mean of a batch of wildtype mutants. I started using a batch to avoid conditional logic of skipping BatchNorm for single mutant embeddings. This fixes some of the automation of using DDP with torch lightning. wt\_mean = 1, so we need the stabilization around 1. Everytime there is spiking wt\_mean, there is corresponding spiking in the overall loss.

### Wildtype Difference Embedding - Layer Norm

We have also tried using LayerNorm instead of BatchNorm. LayerNorm stabilizing the learning, but the model just learns to predict the mean of the dataset  $\sim 0.85$ . We can see this in the boxplot and the wt mean.

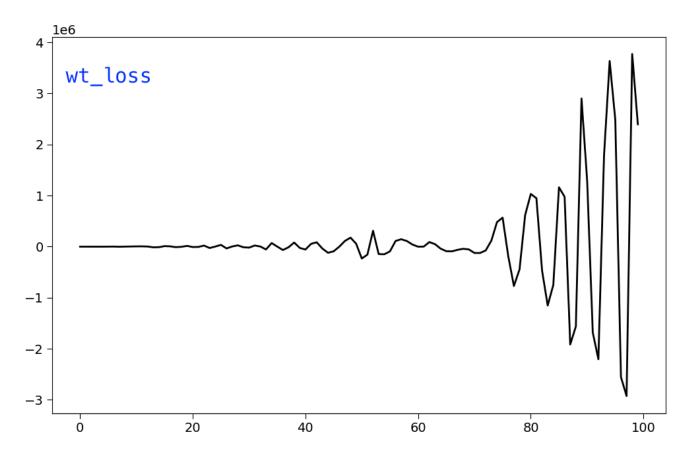
## Box plot of True Values for each Prediction Bin





### Wildtype Difference Embedding - Overfit on Mean

The wt\_loss is MSE for just the wt. In the training loop I have attempted to overfit the wt, before moving on to training on the rest of the data. This should solve the stabilization issue, but this is unstable. I'm thinking there might be an issue with the way I am trying to learn the difference embeddings, and there there is probably a better way.

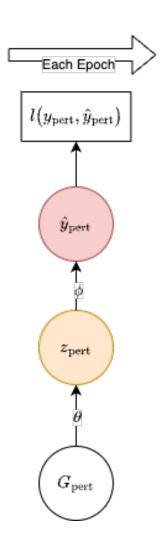


## Perturbation Nodes Benchmark Model

To make sure this task is even possible, I have built some benchmark models, which I think are interesting in their own right. This model is trained to predict fitness from the embeddings of the perturbation graph alone. I still use a Deep set for this.

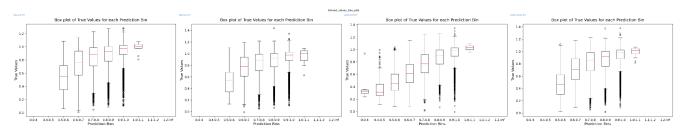
 $egin{aligned} \operatorname{DeepSet} &:= \theta \\ \operatorname{DeepSet} &: \\ \text{- NN on instances} \\ \text{- Order Invariant Aggregation} \\ \text{- NN on set} \end{aligned}$ 

$$egin{aligned} ext{Linear} &:= \phi \ G &= (V, E) \ ext{in this case}, \ E &= \{\} \ G_{ ext{pert}} &:= ext{embedding of nodes removed} \ heta(G) &= z \ heta(z_{ ext{pert}}) &= \hat{y}_{ ext{pert}} \ lig(y, \hat{y}ig) &= rac{1}{n}ig(y - \hat{y}ig)^2 \end{aligned}$$

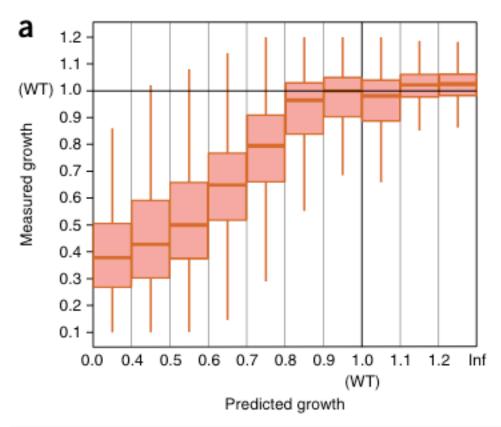


#### Perturbation Nodes Benchmark Model - Downstream Only

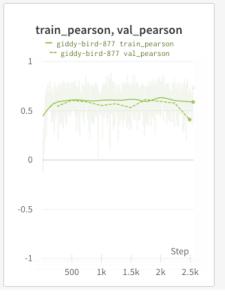
• This model was only trained on downstream CRE. This was initally a mistake, but it turned out to look better than the model trained on both downstream + upstream.

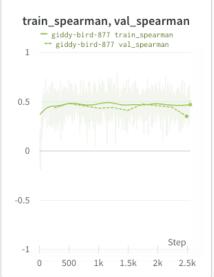


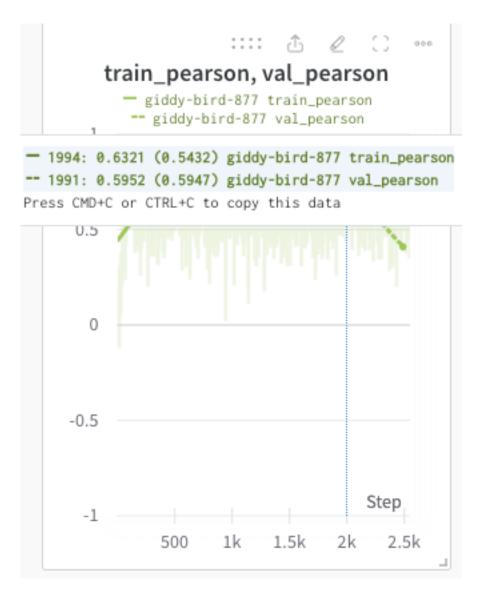
These panels are starting to look comparable to the DCell benchmark model that train on the entire 1.3e7 dataset using the gene ontology visible neural network.











If we look at  $R^2 = 0.6^2 = 0.36$ , so the embeddings of the downstream DNA sequences that are deleted from the genome can explain 36% of the variance in growth.

### Perturbation Nodes Benchmark Model - Downstream and Upstream Comparison

Models Trained on downstream and downstream + upstream.

