

Are Alchemical Free Energy Calculations Reproducible?

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Abstract

Alchemical free energy calculations are an increasingly important modern simulation technique. Contemporary Molecular Dynamics software such as AMBER, CHARMM, GROMACS and SOMD include support for the method. Implementation details vary among those codes but users expect reliability and reproducibility, i.e. a simulation must yield a comparable free energy within statistical bounds regardless of code used. *Relative* alchemical free energy (RAFE) simulation is increasingly used to support molecular design activities, yet the reproducibility of the methodology has been less well tested than its absolute counterpart. Here we present the results for RAFE calculations for

hydration free energies of a set of small organic molecules and show that free energies can be satisfactorily reproduced with aforementioned codes. However, this requires attention to detail and package-specific protocols. The benchmarks and protocols reported here should be useful for the community to validate new and future versions of free energy calculations software.

1 Introduction

The free energy is a fundamental function of thermodynamics as it explains how processes in nature evolve. The equilibrium balance of products and reactants in a hypothetical chemical reaction can be immediately determined from the knowledge of the free energy difference of reactants and products and their concentrations. The free energy landscape of a given system, however, can be very complicated and rugged with barriers which impose limits on how fast the process can take place. It is therefore of little surprise that the determination of free energy changes is of utmost importance in the natural sciences, e.g. for binding and molecular association, solvation and solubility, protein folding and stability, partition and transfer, and design and improvement of force fields.

The calculation of free energies via molecular simulations¹⁻⁵ has been particularly attractive as it promises to circumvent certain limitations of experimental approaches. Specifically, processes can be understood at the atomic level and there is the potential that computational techniques can be more cost and time effective, especially if they can predict the properties of new molecules before their synthesis. Thus, a multitude of methods have been devised to make reversible work estimates accessible through computation.¹⁻⁵ However, the reliability of estimates is still very much a matter of concern.^{2,6}

Here we are interested in *alchemical* free energy methods because they are firmly rooted in statistical thermodynamics and thus give asymptotically correct free energy estimates, i.e. they are correct for a given potential energy function in the limit of sufficient simulation time.^{1,7-9} The method has been applied in various forms for several decades now since the

early days of computer simulation.^{10–15} The method has gained renewed attention in recent years — concomitant with improvements in computer hardware design — within the traditional equilibrium framework^{16–18} and also increasingly in combination with non-equilibrium techniques.^{19–21} The name comes from the nonphysical intermediates that often need to be created to obtain reliable estimates of free energy differences between physical end states and because parts or all of a molecule may appear or disappear in a transformation. In the context of force field methods the transformation takes place in parameter space, i.e. the various force field parameters are varied by scaling. This can be a particularly efficient approach as it does not require sampling of diffusive motions, avoid crossing prohibitively large energy barriers if pathways are not well chosen and is easier to automate.

Alchemical free energy simulations rely on the concept of thermodynamic cycles.¹⁴ As the free energy is a state function, the sum of free energy changes computed around any closed cycle must be zero. This also implies that the reversible work can be computed along conveniently chosen legs of the cycle, even if the cycle is artificial. For example, in Fig. 1 the relative free energy of hydration can be computed along the vertical legs, that is, following the physical process of moving a molecule from the gas phase to the liquid phase, or along the horizontal legs in a non-physical but computationally more efficient alchemical calculation.

Absolute (standard) alchemical free energy calculation has been of particular interest for many years.^{16–19,21,23} *Absolute* here really means that the equilibrium constant of a physical reaction, e.g. binding and dissociation, can be calculated directly by completely decoupling or annihilating a whole molecule from its environment and the term is mostly being used to discriminate against techniques usually referred to as *relative* (see below). We emphasize that the “absolute” approach will still result in a *relative* free energy between the state where the solute fully interacts with its environment and the state where it does not. Decoupling means the scaling of the non-bonded *inter*-molecular interactions between the perturbed group (all atoms that differ in at least one force field parameter between the end states) and its environment. We distinguish this from “annihilation” which also scales the *intra*-



Figure 1: The thermodynamic cycle to compute the relative free energy of hydration $\Delta\Delta G_{\text{hydr}} = \Delta G_{\text{sol}} - \Delta G_{\text{vac}} = \Delta G'' - \Delta G'$. The example is for the ethanol \leftrightarrow methanol transformation. Alchemical simulations are performed along the non-physical horizontal legs while vertical legs illustrate the physical process of moving a molecule from the vacuum to the solution. The latter is also accessible through absolute alchemical free energy simulation, see e.g. Ref. 22.

molecular interactions in addition to the inter-molecular interactions. These schemes may require two simulations along the opposite edges of a quadrilateral thermodynamic cycle but approaches that produce the reversible work directly in one simulation have been proposed too.^{24,25}

Relative alchemical free energy (RAFE) calculations mutate one molecule into another. RAFEs have proven useful for instance to rank sets of related molecules according to their binding affinity for a given receptor. This approach has recently gained increased traction in the context of relative free binding energies between small molecules, e.g. drug or lead like molecules and biomolecules.^{26,27}

RAFEs are readily implemented^{28,29} by making use of the single topology method.^{15,28,30} Single topology means that there is only one representation of the molecule to be mutated, also implying a single set of coordinates. Thus, atom types are directly transformed into the new type, typically by linearly scaling the force field parameters. In typical implementations, disappearing/appearing atoms need to be balanced with “dummy” atoms to ensure constant

number of atoms in both end states. Dummy atoms have no non-bonded interactions in the end state but retain the bonded terms of the original atom to avoid complications with unbound atoms³⁰ (see also wandering ligand problem, discussed below). However this need not be. For instance, the AMBER implementation is a special case as it does not require the user to define dummy atoms explicitly, i.e. as atoms with coordinates and zero end state parameters in the topology, but only to mark the disappearing and appearing atoms in the control file of the MD engine. The contributions of bonded terms that involve at least one dummy atom will not be factored into the free energy as it is assumed that those contributions will perfectly cancel out in the thermodynamic cycle.^{31,32}

The single topology approach²⁸ requires topological and structural similarity. But also chemical similarity is also of importance; e.g. chirality and binding modes where the relative three dimensional arrangement of groups in space must be taken into account. Furthermore, ring breaking is technically challenging²⁷ but it has also been shown that this can be done in certain circumstances.^{31,32}

When the two molecules are structurally dissimilar or do not spatially overlap, the dual topology method^{28,30} can be applied to compute relative free energies. In this approach *all* atoms of the end states are duplicated and thus both sets are present at all times but do not interact with each other. Only non-bonded interactions need to be scaled such that the disappearing end state corresponds to an ideal gas molecule.³⁰ This, however, comes with additional complications as two independent molecules can drift apart and so suffer from the “wandering” ligand problem as in absolute transformations.^{16–18} Technically, a dual topology calculation is the same as two absolute calculations run simultaneously in opposite directions. It has been shown though that with the introduction of special restraints or constraints this can be a viable option.^{33–35} Restraints between corresponding atoms, thus emulating the single topology approach, can also be used without affecting the free energy.³⁵ A recent alternative⁶ considered molecules with a common core where all atom types are the same. The charges that would be typically different in individual parameterization due to the local

chemistry were made equal. This means that the core does not need to be duplicated and thus is not included in the mutation. But all the chemistry is now exclusively handled by, possibly, just a few non-core atoms such that this approach may only be of limited use.

A covalent link, e.g. as in side-chain mutation simulations, provides a natural restraint such that dual topology simulations can be applied without further problems. Modern MD software, e.g. AMBER,³⁶ CHARMM,³⁷ GROMACS,³⁸ GROMOS³⁹ and SOMD,^{40,41} often provides a hybrid single/dual topology approach i.e. the user can specify which part of a perturbed group should be handled by which method.³²

As alluded to above, reliability is a principal matter of concern. In particular, we need to ensure reproducibility of free energy results among computer codes. To the best of our knowledge this has not been systematically tested yet for a set of different MD packages. However, there have been some recent efforts to test *energy* reproducibility across packages⁴² — a necessary but not sufficient prerequisite. Another study went further and also compared densities across packages, revealing a variety of issues.⁴³ For free energies, given a predefined force field and run-time parameters we ought to be able to obtain comparable free energy results within statistical convergence limits, though this has not yet been tested.

In practice, we have the problem, however, that the methodologies used in one MD program are not always present in another package, or the same functionality is provided via different algorithms (e.g. algorithms for pressure and temperature scaling, integrators, etc). Nevertheless, it is critical that free energy changes computed with different simulation software should be reproducible within statistical error, as this otherwise limits the transferability of potential energy functions, and the relevance of properties computed from a molecular simulation with a given package. This is especially important as the community increasingly combines or swaps different simulation packages within workflows aimed at addressing challenging scientific problems.^{44–48}

In this work we present the results of relative hydration free energies of a set of small organic molecules (see Fig. 2). Solvation free energies have a wide range of uses and vari-

ous methods exist to compute them.⁴⁹ They are also needed for calculations of a variety of important physical properties, and to calculate binding free energies where the simulation in solution (see Fig. 1) is combined with a mutation of the molecule bound to a partner.⁴⁹ A large database of hydration free energies computed from alchemical free energy (AFE) simulations, FreeSolv, has been presented recently⁵⁰ and was just updated.²² Here, we focus on the reproducibility of RAFF with the simulation programs AMBER, CHARMM, GROMACS and SOMD. We will discuss the reversible work results obtained with these packages and make recommendations regarding simulation protocols, setup procedures and analysis techniques. We will also deliberate on what needs to be done to progress the field, both from a usability perspective as well as from the view point of code development.

2 Methods

We will compare relative free energies calculated via simultaneous parameter change simulations — our “unified protocol” — and separated parameter change simulations — our “split protocol”— to relative free energy calculations obtained from two absolute hydration free energies — our “absolute protocol”. The unified protocol changes partial charges, van der Waals parameters, and bond parameters simultaneously along the alchemical path so that charge interactions and van der Waals interactions are varying simultaneously, while the split protocol separates the transformation of the van der Waals parameters from the charge transformation so that electrostatic interactions are modified separately from van der Waals interactions. The order in which this has to be done is detailed in section 2 of the SI. In the split protocol, the scaling of the bonded terms can be combined with either transformation.

2.1 Alchemical Free Energy Implementations

We begin by examining the differences in the alchemical free energy implementations of the four MD codes we consider — AMBER, CHARMM, GROMACS and SOMD. One key

difference is in the softcore functions^{51,52} implemented in each code as summarised in section 1 of the SI. Softcore functions are used to avoid numerical and thus stability problems of the conventional van der Waals and Coulombic potentials⁵³ as they have singularities at zero distance (vertical asymptotes). Attempting to modify interactions by linearly scaling back these interaction potentials as a function of an interaction parameter, λ , causes these functions to increasingly behave like hard-sphere potentials as $\lambda \rightarrow 0$. This means that there is a discontinuous change between $\lambda = 0$ where particles can overlap, and $\lambda = \epsilon$ (where ϵ is very small) where particles still behave like hard though minuscule spheres. This can lead to strongly fluctuating forces/energies and to severe instabilities in the integrator, as well as numerical errors in post processing analyses even when simulations do terminate normally.^{51–53}

Another difference is how the codes scale force field parameters (“parameter scaling”) and/or the energy (“energy scaling”).³⁰ In the former case each parameter is scaled individually, e.g. in the case of a harmonic bond or angle term, the force constant and the equilibrium distance/angle are scaled. In the latter case, the total energy is scaled, all at once or, equivalently for each individual force field contribution. The two approaches are not mathematically equivalent and so the pathway through state space or alchemical space is different. It should be noted, however, that modification of Lennard-Jones interactions is handled through softcore functions in modern codes (as is, in some cases, that for electrostatic interactions) and thus parameter scaling^{28,54} is now only used for bonded terms and sometimes electrostatic interactions, at least in modern codes.

One more important issue is whether the code allows constraints to be applied to bonds which change bond lengths in a transformation e.g. C–H to C–C. Changes in bond length need to account for the associated change in the free energy. These and other details will be outlined below.

AMBER. This code is strictly dual topology and all terms are energy-scaled. The code allows, however, mapping of atoms in a single topology fashion and computes these non-softcore atoms by linearly scaling the energy and forces for each atom in the pair. The perturbed group must be entirely duplicated, i.e. for `sander` this means two topology files with one end state each, and for `pmemd` both end states in one topology file. The softcore potential applies to any atom chosen by the user which may include atoms that actually have an equivalent in the other state. Non-softcore atoms must still match 1:1 in both states. Explicit dummy atoms are not needed as the code will only compute bonded contributions for “real” atoms and thus ignores bonded energies involving dummy atoms. We will call this the “implicit dummy protocol”. The code cannot handle bond length changes involving a constraint. There is only one global λ for parameter transformation. Protocols that couple only some parameters (split protocols, see below) must be emulated through careful construction of topologies by keeping force field parameters that are not being modified constant along λ .

CHARMM. The PERT module duplicates the topology similar to `sander` but mapped atoms are given in the topology only once. The module requires balancing with explicit dummy atoms. All terms are energy-scaled. The PSSP softcore potential is applied to *all* atoms in the perturbed group (see section 1 in the SI). The code can handle constraints of changing bond lengths in the perturbed group but this may cause incorrect results with PSSP softcores (Stefan Boresch, private communication). There is only one global λ for parameter transformation, however, the scripting facilities in CHARMM allow run time modification of topologies e.g. by setting charges or vdW parameters to arbitrary values.

GROMACS. This code uses a single topology description. Bonded terms are strictly parameter-scaled, which requires proper balancing of multi-term dihedrals i.e. each individual term in the Fourier series must have an equivalent in both end states. If the term does not exist it must be created with parameters zeroing its energy. The softcore potential applies to

dummy atoms only determined from atoms having zero vdW parameters in the end states. The code allows changing bond lengths involving constraints within the perturbed group but this can lead to instabilities and wrong results (Michael Shirts, private communication). There are separate λ s for vdW, Coulomb and bonded parameters (and some other) which allows easy implementation of split protocols.

SOMD. This code uses a single topology description. The final state is constructed at run time from the initial state with a “patch” (list of force field parameters to be modified). All dummy atoms needed to describe the transformation must be present in the initial state. Bond and angle terms are parameter-scaled while the dihedral term is energy-scaled. The softcore potential applies to atoms that become dummy atoms in one end-state. Dummy atoms are specified by a keyword in the patch file. The code cannot handle constraints of changing bond lengths in the perturbed group. There is only one global λ for parameter scaling. Separated protocols (see below) must be emulated through careful construction of the patch file. SOMD is software built by linking of the Sire⁴⁰ and OpenMM⁴¹ molecular simulation libraries.

2.2 RAFF Setup

The setup for all relative free energy simulations has been carried out with the tool FESetup⁴⁷ (version 1.2). FESetup is a perturbed topology writer for AMBER, CHARMM, GROMACS, SOMD and also NAMD⁵⁵ (within the limits of the dual topology approach). The tool makes use of a maximum common substructure search algorithm to automatically compute atoms that can be mapped, i.e. atoms that have a direct relationship to an equivalent atom in the other state – atoms undergoing atom type conversion or modification. The only current limit is that rings are required to be preserved.³¹ With this strategy a maximal single topology description is achieved: any atom that does not match will be made a dummy atom. FESetup allows equilibration of the solvated simulation systems and ensures

that “forward” and “backward” simulations have the same number of total atoms. The masses of perturbed hydrogen atoms are set to the mass of the heavy atom for SOMD. The other codes use the atom masses of the initial state (AMBER, CHARMM?) or allow the user to define how masses vary as a function of lambda (GROMACS). The tool creates all input files with control parameters, topologies and coordinates as required for RAFE simulations. Full details on FESetup can be found in Ref. 47.

Figure 2 shows all 18 transformation considered in this study including “forward” and “backward” mutations. In the limit of sufficient sampling, RAFE simulations do not have a directionality with respect to the coupling (order) parameter λ . But to test for possible discrepancies we have run simulations in both directions. As we shall discuss in the Results section we do see differences in some cases.

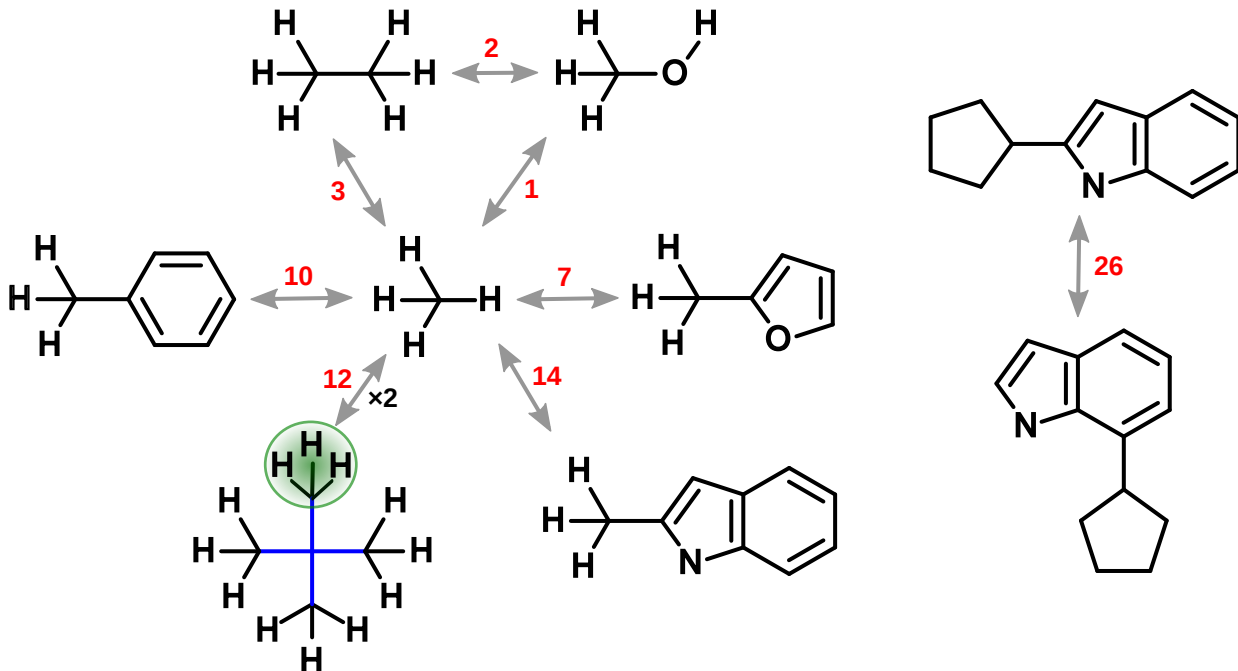


Figure 2: The thermodynamic cycles considered in this study. To compute the free energy of hydration, all pair-wise transformations have to be carried out once in solution and once in vacuum. Green and blue colours in neopentane show two alternative mappings for methane. The numbers in red denote the number of dummy atoms.

The ethane \rightarrow methanol transformation is traditionally regarded as a standard test for RAFE simulations.^{15,56} The other transformations are centered around mutations from and

to methane, and are meant to mimic components of typical transformations that could be attempted in the context of e.g. protein-ligand binding calculations. The 2-cyclopentanylindole to 7-cyclopentanylindole (2-CPI to 7-CPI in our notation) transformation has been added to include both deletion as well as insertion of sub-parts of the perturbed group in one transformation, an aspect not tested by the other transformations. For neopentane \rightarrow methane we point out that there are two alternative mappings possible, see Figure 2. One mapping has methane matched to a terminal methyl (green) and the other one has the methane carbon matched with the central carbon in neopentane (blue). The first approach will be called “terminally mapped” and the second one “centrally mapped”.

2.3 RAFF Simulation Protocols

One of the major concerns of this reproducibility study is to ensure consistency in the applied protocols. This is complicated by the fact that a given MD software may employ a wide range of methods and algorithms that may not be available in other MD software. For example, pressure and temperature scaling, integrators and other algorithms can be very different. It is also unclear if and how implementation details can affect results, in particular see subsection 2.1 discussing the implementation details of alchemical free energy simulation in code.

In this study we consider at a set of simple organic molecules (see Figure 2). As the focus here is on probing for reproducibility among various MD packages, we chose fairly small, rigid and neutral molecules to minimise statistical sampling errors, and avoid difficulties with charged particles.^{57,58} The force field was chosen to be GAFF⁵⁹ (version 1.8), utilising AM1/BCC charges^{60,61} for the solute and TIP3P⁶² for the solvent. Charges were computed with the `antechamber` program and missing bonded and vdW terms were generated with the `parmchk2` program, both from the AmberTools16 distribution. The quality of free energies of various small molecule force fields has been discussed elsewhere, see e.g. Refs. 63,64.

While the MD packages principally allow a “one-step” transformation,⁶⁵ that is, both van

der Waals and Coulombic softcore potentials vary simultaneously, it can be more efficient to carry out a split protocol.^{66,67} In such a protocol the charges are transformed linearly between the end states followed by a mutation of the van der Waals parameters using a softcore potential^{51,52} (see section 1 in the SI for details) on the vdW term only. It is important to note that in the split protocol charges have to be switched off before vdW parameters (and vice versa for the transformation in opposite direction) to avoid collapse of other atoms, e.g. solvents, onto a “naked” charge⁶⁵ see section 2 in the SI.

All simulations were started from simulation boxes prepared by FESetup.⁴⁷ It should be noted, however, that in constructing the system steric overlaps between the solute and the solvent may happen. This is because each unperturbed solute is independently equilibrated and the perturbed system combined from those two. The number of atoms are always chosen to be the same for forward and backward setups by using the larger box of the two unperturbed systems. Thus, in transformation from a smaller to a larger solute, water molecules may be in particularly close proximity to the solute. The production simulations were run at 298.15 K and 1.0 bar.

AMBER. We made use of AMBER16 for all free energy calculation. The starting coordinates were usually taken directly from the pre-equilibrated setup step but no further λ specific equilibration was carried out, i.e. RAFE MD simulations were started with new velocities appropriate for the final simulation temperature. In a very few cases it was necessary to use coordinates from the end of the simulation at a nearby λ state because of simulation instabilities. This happened in transformations with a larger number of dummy atoms. Water hydrogens (TIP3P) were constrained with SHAKE. None of the atoms in the perturbed group were constraint and hence the time step set to 1 fs. We also tested a alternative protocol with SHAKE also on bonds that do not change during transformation and a time step of 2 fs (see SOMD protocol below). The temperature was controlled through a Langevin thermostat with a friction constant of 2.0 ps^{-1} and pressure rescaling through a

Monte Carlo barostat with 100 steps between isotropic volume change attempts. Long-range electrostatics in solution was handled with PME and a cutoff of 8.0 Å for both Coulomb and vdW interactions. In vacuum no cutoff was used.

CHARMM. The CHARMM c40b1 molecular dynamics package was used to carry out all relative hydration free energy simulations. 21 evenly spaced windows were used and all windows were run for 1.5 ns with a timestep of 1 fs. Most windows used the same pre-equilibrated configuration, but a few windows were started with their own equilibration. Constant temperature and pressure control was done with the Berendsen weak coupling method, with a compressibility of $4.63 \times 10^{-5} \text{ atm}^{-1}$ and temperature and pressure coupling constants of 5.0 ps^{-1} . A 12.0 Å atom-based cutoff was used for non-bonded interactions and Particle Mesh Eward (PME) with order of 6 was used to calculate electrostatic forces along with periodic boundary condition. SHAKE was used to constrain water hydrogens only. The PERT module in CHARMM was used to handle alchemical transformations and PSSP softcore potential functions were used to avoid numerical instabilities.

GROMACS. We used GROMACS 4.6.7 to carry out simulations for the relative hydration free energies ($\Delta\Delta G_{\text{hydr}}$). Each transformation had its Gibbs free energy calculated: (i) in a single topology approach in which van der Waals energy terms were changed separately from the electrostatic and bonded components; (ii) in a single topology approach in which bonded, van der Waals, and electrostatic terms are changed together; and (iii) via the difference between two absolute calculations. In the first two cases, each alchemical transformation was described by 31 and 16 states, respectively, and simulated for 4.2 ns with time steps of 1.0 fs in water and vacuum. Atomic masses were not changed along the alchemical path; their change affect kinetic energy only and do not contribute to the free energy change on this case. We used the Langevin integrator implemented in GROMACS with a default friction coefficient of $1.0 \text{ ps}/m_{\text{atom}}$, where m_{atom} is the the mass of the atom. Absolute hydration free energies were calculated from 5 ns Langevin dynamics at 298.15 K. We used the 20-step alchemical

protocol where charge coupling and van der Waals coupling were dealt with separately along the path.^{22,50} No bond constraints were used. A Parrinello–Rahman barostat with $\tau_p = 10$ ps and compressibility equal to $4.5 \times 10^{-5} \text{ bar}^{-1}$. We used two methods to calculate electrostatic interactions: Particle Mesh Ewald (PME) and charge group-based Reaction Field with a dielectric of 78.3, as implemented in the software. We set the electrostatic and non-bonded cutoffs to 10.0 Å; a switch was applied to the latter starting at 9.0 Å. Non-bonded interactions in the vacuum legs of the simulation were calculated to a cutoff of 50.0 Å. PME calculations were of order 6 and had a tolerance of 1.0×10^{-6} , with a grid spacing of 1.0 Å. All transformations required the use of softcore potentials to avoid numerical problems in the free energy calculation. We chose the 1–1–6 softcore potential for Lennard-Jones terms ($\alpha=0.5$ and $\sigma=0.3$) and used the default softcore Coulomb implementation in paths where charges, van der Waals, and bonded terms were modified together, but no soft core potentials were applied to Coulomb interactions when electrostatic interactions were modified separately.

SOMD. All simulations were carried out with SOMD from the Sire 2016.1 release^{40,41} Each alchemical transformation was divided into 17 evenly spaced windows and simulated for 2 ns each both in water and in vacuum. A velocity-Verlet integrator was employed with a 2 fs time step, constraining only bonds involving hydrogens which are not alchemically transformed. This approach will be called the “unperturbed H bond constraint protocol”. Referring to eq. S1 the softcore parameters were set to default values for all the transformations, specifically $n = 0$ for Coulombic interactions and $\alpha = 2.0$ for the Lennard-Jones potential.³³ A shifted atom-based Barker–Watts reaction field⁶⁸ with a dielectric constant of 78.3 was adopted for the solution phase simulations with a cutoff of 10 Å. A similar cutoff was used for Lennard-Jones interactions. Temperature control was achieved with the Andersen thermostat⁶⁹ with a coupling constant of 10 ps⁻¹. A Monte Carlo barostat assured pressure control, with isotropic box edge scaling moves attempted every 25 time steps. The reaction

field was not employed in the vacuum legs, where a Coulombic potential without cutoff was used. This leads to an inconsistent description of the intramolecular electrostatic interactions of the solute in the solvated and vacuum phases. To maintain a consistent description of intramolecular energetics across vacuum and water legs, a free energy correction term ΔG_c was evaluated as detailed in Ref. 70). The ΔG_c term was obtained via post-processing of the end state trajectories of each water phase simulation, using the Zwanzig relationship:⁷¹

$$\Delta G_c = -\beta^{-1} \ln \langle \exp [-\beta (U_{ic,nc}(\mathbf{r}) - U_{ic,sim}(\mathbf{r}))] \rangle_{sim} \quad (1)$$

where $U_{ic,nc}(\mathbf{r})$ is the solute intramolecular electrostatic-no cutoff potential that depends on the coordinates \mathbf{r} of the solute and is given by Coulomb’s law computed without cutoffs. $U_{ic,sim}(\mathbf{r})$ is the intramolecular electrostatic potential term as computed in the simulation with the shifted atom-based Barker-Watts Reaction Field cutoff.

To compute absolute free energy of hydration a double annihilation technique^{16,23,70} was adopted. Initially the solute atoms’ partial charges are turned off both in water and in vacuum (discharging step), giving a discharging free energy change ΔG_{solv}^{elec} and ΔG_{vac}^{elec} respectively. Secondly, the molecule is fully decoupled from the surrounding environment by switching off the van der Waals terms (vanishing step), giving a ΔG_{solv}^{vdW} free energy change in water phase and ΔG_{vac}^{vdW} free energy change in vacuum. Next, the absolute free energy of hydration ΔG_{wat} is computed as:

$$\Delta G_{wat} = (\Delta G_{solv}^{elec} + \Delta G_{solv}^{vdW}) - (\Delta G_{vac}^{elec} + \Delta G_{vac}^{vdW}) \quad (2)$$

The same setup for relative free energy calculations was followed to compute absolute free energy of hydration. Thus, 17 evenly spaced λ windows were used and simulations were run for 2 ns employing a velocity-Verlet integrator with 2 fs time step, constraining only the hydrogen bonds which are not alchemically transformed.

2.4 Free Energy Estimations

In this work we primarily focus on TI as this is supported by all the tested MD packages “out-of-the-box”, whereas BAR and MBAR are not. Equation 3 computes the free energy as

$$\Delta G = \int_{\lambda=0}^{\lambda=1} \left\langle \frac{\mathcal{H}(\mathbf{q}, \mathbf{p}; \lambda)}{\partial \lambda} \right\rangle_{\lambda} d\lambda \quad (3)$$

where $\mathcal{H}(\mathbf{q}, \mathbf{p}; \lambda)$ is the Hamiltonian as a function of the coordinate vectors \mathbf{q} and the momentum vectors \mathbf{p} , and parametric dependence on the coupling parameter λ . The angle brackets denote the ensemble average of the gradient of the Hamiltonian with respect to λ at a given λ value. An AFE simulation is typically carried out in a series of equilibrium simulations at discrete values of λ but the gradient can also be evaluated with a continuously varying coupling parameter as a function of the simulation time. The free energy is finally computed through a suitable numerical integration method.

Results from additional estimators will be given where available. We have used the alchemical analysis tool⁷² for analysis of all free energies. This tool provides various estimators such as TI, TI with cubic splines, BAR and MBAR. All data was sub-sampled to eliminate correlated data.

All RAFF simulations were run in triplicate in forward as well as backward direction for a total of 6 simulations per mutation. The final hydration free energy $\Delta\Delta G_{\text{hydr}}$ was computed as the average for each direction separately. For comparison we have also calculated the absolute (standard) hydration free energies for all molecules in Figure 2.

To estimate the reliability and convergence of the results, the standard error of the mean (SEM) has been calculated. The SEM is defined as

$$\text{err}(\Delta\Delta G_{\text{hydr}}) = \frac{\sigma}{\sqrt{n}} \quad (4)$$

where σ is the sample standard deviation and n is the size of the uncorrelated sample. The SEM for component free energies is combined as

$$\text{err}(\text{combined}) = \sqrt{\sum_i \sigma_i^2}. \quad (5)$$

which is appropriate if the property to be computed is a sum of contributions.

We also make use of the mean absolute error MAE (also called mean unsigned error, MUE) to compare data sets.

$$\text{MAE} = \frac{1}{n} \sum_{i=1}^n |y_i - x_i| \quad (6)$$

where n is the total number of samples, y_i and x_i are the i -th datum to be compared.

3 Results

We will use absolute hydration free energies here as our standard point of comparison because for the present dataset these are easily calculated with high precision²² and are considerably simpler to set up and implement than relative calculations. Prior work actually compared calculated absolute hydration free energies across different codes with significant success⁷³ (see Fig. 7 in the reference), further supporting this view.

Table 1 summarizes results for relative free energies of hydration obtained from absolute transformations which are given in more detail in Table S1. The table shows the data from simulations with the recommended protocol for each MD code, as discussed in detail in the following subsections. The $\Delta\Delta G_{\text{hydr}}$ obtained with the various MD packages in this way agree quite well, although some larger deviations are apparent as well. GROMACS predicts a smaller $\Delta\Delta G_{\text{hydr}}$ for the methanol transformations by about 0.2 kcal mol⁻¹. Similarly, there are some discrepancies in the toluene, 2-methylfuran and 2-methylindole cases where it is mostly CHARMM that produces somewhat smaller $\Delta\Delta G_{\text{hydr}}$ with 0.4 kcal mol⁻¹ difference between the smallest and highest values. The largest deviation can be found for the largest transformation (2-CPI to 7-CPI) with the AMBER result higher by 0.3–0.5 kcal mol⁻¹ than any other MD package.

Table 1: Comparing relative free energies of hydration for various MD packages as obtained from the absolute protocol.

transformation		AMBER ^a	CHARMM ^a	GROMACS ^b	SOMD ^a
ethane	methane	-0.02 ± 0.01	-0.08 ± 0.01	-0.04 ± 0.01	-0.05 ± 0.02
methanol	methane	6.20 ± 0.01	6.15 ± 0.02	5.95 ± 0.01	6.21 ± 0.06
ethane	methanol	-6.22 ± 0.01	-6.23 ± 0.02	-5.98 ± 0.01	-6.26 ± 0.05
toluene	methane	3.19 ± 0.01	3.00 ± 0.01	3.16 ± 0.01	3.07 ± 0.03
neopentane	methane	-0.13 ± 0.02	-0.18 ± 0.02	-0.14 ± 0.01	-0.19 ± 0.06
2-methylfuran	methane	2.96 ± 0.02	2.79 ± 0.01	2.95 ± 0.01	2.90 ± 0.03
2-methylindole	methane	8.72 ± 0.01	8.36 ± 0.02	8.79 ± 0.02	8.57 ± 0.03
2-CPI	7-CPI	0.39 ± 0.04	-0.10 ± 0.04	0.02 ± 0.05	0.08 ± 0.14

^aunified protocol.

^bsplit protocol with PME.

Table 2 shows the MAD between SOMD, GROMACS, AMBER and CHARMM. The SOMD code produces figures that agree the most with all other MD packages (MAD ca. $0.1 \text{ kcal mol}^{-1}$), and the MAD reaches just under $0.2 \text{ kcal mol}^{-1}$ for CHARMM/GROMACS, and CHARMM/AMBER.

Table 2: Mean Absolute Deviations (MAD) (kcal mol^{-1}) between relative free energies obtained with the absolute protocol for the SOMD, GROMACS, AMBER and CHARMM packages.

Package	GROMACS	AMBER	CHARMM
SOMD	0.12	0.09	0.08
GROMACS		0.13	0.17
AMBER			0.16

Having established the predictive value from absolute transformations we now turn to computing $\Delta\Delta G_{\text{hydr}}$ from relative mutations. Table 3 summarizes the results for the four MD packages. Again the data is from the recommended protocol (see detailed discussions in the following subsections).

CHARMM tends to show smaller relative free energies for a number of transformations: toluene, neopentane, 2-methylfuran and 2-methylindole. SOMD displays smaller $\Delta\Delta G_{\text{hydr}}$ in the methanol and toluene transformations. The largest discrepancy, however, is in the

Table 3: Comparison of relative free energies of hydration for various MD packages as obtained from relative transformations via unified or split protocols. Signs of the backward transformation have been reverted to correspond to the forward transformation.

		AMBER ^a		CHARMM ^b	GROMACS ^a	SOMD ^b
		implicit ^c	explicit ^c			
ethane	methane	0.02 ± 0.01	-0.13 ± 0.02	-0.11 ± 0.02	-0.04 ± 0.02	-0.01 ± 0.05
methane	ethane	0.00 ± 0.03	-0.19 ± 0.03	-0.06 ± 0.01	-0.02 ± 0.01	-0.04 ± 0.02
methanol	methane	6.19 ± 0.01	6.20 ± 0.02	6.17 ± 0.01	6.20 ± 0.01	5.99 ± 0.05
methane	methanol	6.20 ± 0.03	6.15 ± 0.01	6.20 ± 0.01	6.20 ± 0.01	5.96 ± 0.04
ethane	methanol	-6.20 ± 0.01	-6.27 ± 0.01	-6.26 ± 0.01	-6.19 ± 0.01	-6.09 ± 0.03
methanol	ethane	-6.20 ± 0.01	-6.25 ± 0.01	-6.29 ± 0.01	-6.19 ± 0.01	-6.09 ± 0.02
toluene	methane	3.24 ± 0.02	3.39 ± 0.02	2.93 ± 0.02	3.21 ± 0.01	2.89 ± 0.09
methane	toluene	3.42 ± 0.03	3.52 ± 0.03	2.96 ± 0.02	3.20 ± 0.01	3.06 ± 0.02
neopentane ^d	methane	0.32 ± 0.04	-0.03 ± 0.06	-0.44 ± 0.01	-0.15 ± 0.02	-0.20 ± 0.05
methane ^d	neopentane	0.25 ± 0.03	-0.07 ± 0.03	-0.33 ± 0.02	-0.16 ± 0.05	-0.13 ± 0.05
neopentane ^e	methane	-0.13 ± 0.01	-0.12 ± 0.02	-0.65 ± 0.02	-0.14 ± 0.01	-0.11 ± 0.01
methane ^e	neopentane	-0.13 ± 0.03	-0.12 ± 0.03	-0.49 ± 0.02	-0.18 ± 0.03	-0.10 ± 0.06
2-methylfuran	methane	3.09 ± 0.01	3.10 ± 0.01	2.76 ± 0.03	2.93 ± 0.05	2.92 ± 0.05
methane	2-methylfuran	3.10 ± 0.03	3.15 ± 0.03	2.76 ± 0.02	2.96 ± 0.01	2.83 ± 0.03
2-methylindole	methane	8.78 ± 0.03	8.78 ± 0.04	8.34 ± 0.01	8.73 ± 0.03	8.64 ± 0.06
methane	2-methylindole	9.14 ± 0.02	9.13 ± 0.03	8.42 ± 0.02	8.74 ± 0.01	8.67 ± 0.08
2-CPI ^f	7-CPI	0.36 ± 0.03	0.63 ± 0.06	-0.01 ± 0.01	-0.03 ± 0.03	-0.11 ± 0.07
7-CPI ^f	2-CPI	0.34 ± 0.05	0.50 ± 0.03	0.03 ± 0.01	-0.20 ± 0.04	-0.01 ± 0.08

^asplit protocol.

^bunified protocol.

^cusing either the implicit or the explicit dummy atom approach.

^dcentral mapping.

^eterminal mapping.

^fpartial re/discharge i.e. only the charges of the appearing and the disappearing 5-rings are switched.

neopentane transformation with central mapping where AMBER with implicit dummy atoms is about $0.5 \text{ kcal mol}^{-1}$ higher and CHARMM about $0.3 \text{ kcal mol}^{-1}$ lower than the other two codes. The terminal mapped neopentane case reveals AMBER to be in line with GROMACS and SOMD while CHARMM’s results are essentially the same as in the central mapped transformations. AMBER deviates also quite strongly from the other codes in the cyclopentanyl indole cases.

The MADs of the relative free energy simulations are presented in Table 4. They tend to be larger than the MADs from the absolute simulations (Table 2) and reach $0.3 \text{ kcal mol}^{-1}$ for CHARMM compared with AMBER and GROMACS.

Table 4: MAD (in kcal mol^{-1}) comparing relative free energies from relative simulations between SOMD, GROMACS, AMBER and CHARMM.

Package	GROMACS	AMBER	CHARMM
SOMD	0.18	0.23	0.19
GROMACS		0.14	0.31
AMBER			0.32

We can also compute the cycle closure error from Table 5 for the closed cycle ethane \rightarrow methanol \rightarrow methane \rightarrow ethane (see Figure 2). For the implicit dummy simulation with AMBER we calculate a cycle error for $\Delta\Delta G_{\text{hydr}}$ of $(0.069 \pm 0.041) \text{ kcal mol}^{-1}$ and for the explicit dummy simulation the error is $(-0.016 \pm 0.047) \text{ kcal mol}^{-1}$.

3.1 AMBER

Using AMBER for RAFE simulations has revealed several problems with the implementation. Some bugs were identified and the developers have fixed those for AMBER16, e.g. energy minimization in `sander` led to diverged coordinates for mapped atoms. For a single topology description, however, it is necessary to have the same coordinates. Other issues are that vacuum simulations can only be carried out with the `sander` program because `pmemd` cannot handle AFE simulations in vacuum at the moment. This will, however, be rectified in future

versions.⁷⁴ A disadvantage of `sander` is that it cannot be used to simulate the λ end points⁷⁵ such that the TI gradients need to be extrapolated (minimum and maximum allowed λ s are 0.005 and 0.995).

Also, `sander` considers the whole system as the perturbed region while `pmemd` restricts this to a user chosen atom selection. This has obvious implications for performance.⁷⁵

We also found that, in contrast to the other three codes, AMBER does not yield correct relative free energies with the unified protocol, i.e. when all force field parameters are scaled simultaneously (see Table S2). This appears to be a problem when more than a few dummy atoms are involved, while the unified protocol works for the smaller transformations (refer to Figure 2). The split RAFF protocol and absolute free energies, however, are very close to the other MD packages as demonstrated in Table 5 below.

End point geometries appear to be another issue with AMBER simulations in both solution and vacuum. This is most obvious in the neopentane \rightarrow methane test case with central mapping (see RAFF Setup and Figure 1). As shown in Figure S3, the methane end state exhibits incorrect distances between the carbon and the four attached hydrogens of approximately 1.23 Å. This value is about 1.12 Å for the terminal dummy atoms in the other test cases but still higher than the expected 1.09 Å on average. Figure S3 demonstrates how this depends on the number of dummy atoms immediately surrounding the central atom.

We also compare free energies obtained from the implicit dummy approach in AMBER with results from explicit dummy atom simulations and results from absolute transformations, see Tables 1 and 3. The relative simulations have been carried out with the split protocol while the absolute simulations used a unified protocol throughout. SHAKE was explicitly deactivated for all bonds in the perturbed region in these protocols. Table 5 shows selected results for transformations with SHAKE enabled for all bonds to hydrogens except those bonds that change bond length during transformation.

The time step has been increased from 1 fs as used in the other three protocols to 2 fs. As the results are essentially the same as the non-SHAKE simulations, this SHAKE protocol

Table 5: Comparing AMBER results for simulations with various split protocols. The emphasis is here on the data with SHAKE enabled and a time step of 2 fs (last column). Implicit, explicit and absolute protocols had SHAKE disabled and a time step of 1 fs. Signs of the backward transformation have been reverted to correspond to the forward transformation.

transformation		implicit $\Delta\Delta G$	explicit $\Delta\Delta G$	absolute ΔG	SHAKE ^a $\Delta\Delta G$
ethane	methanol	-6.20 ± 0.01	-6.27 ± 0.01	-6.22 ± 0.01	-6.18 ± 0.01
methanol	ethane	-6.20 ± 0.01	-6.25 ± 0.01		
toluene	methane	3.24 ± 0.02	3.39 ± 0.02	3.19 ± 0.01	3.27 ± 0.03
methane	toluene	3.42 ± 0.03	3.52 ± 0.03		
neopentane ^b	methane	0.32 ± 0.04	-0.03 ± 0.06	-0.13 ± 0.02	0.35 ± 0.02
methane ^b	neopentane	0.25 ± 0.03	-0.07 ± 0.03		
neopentane ^c	methane	-0.13 ± 0.01	-0.12 ± 0.02		
methane ^c	neopentane	-0.13 ± 0.03	-0.12 ± 0.03		

^aimplicit dummy atom protocol with $\delta t = 2$ fs and SHAKE on all H-bonds except perturbed bonds.

^bcentral mapping.

^cterminal mapping.

appears to be a viable solution to increase the performance of RAFF simulations. We have repeated this protocol with AMBER in response to the good results obtained with SOMD using this implementation. From a practical point of view, AMBER uses an *atom* based mask for bond SHAKEs such that the mask must be set for the hydrogens in question while the same is not possible for their non-H counter-part in the other state because *all* bonds emanating from this atom would be affected.

In general, the free energies computed with each approach are in good agreement with each other and with the results of the other MD packages (Tables 1 and 3). There are, however, a few notable deviations. Neopentane \rightarrow methane with central mapping differs from the result with terminal mapping by about $0.4 \text{ kcal mol}^{-1}$. The terminal mapping and the free energies from the explicit dummy simulations are, however, consistent with the absolute transformations (Table 1). We also observe a systematic deviation between forward and backward vacuum transformations in the 2-methylindole simulation (see Table S3). The discrepancy is consistently $0.2\text{--}0.4 \text{ kcal mol}^{-1}$ for each λ step of the vdW plus bonded transformation with both implicit and explicit dummy atoms.

3.2 CHARMM

1.bug in TI gradient accumulation in parallel runs (does not affect serial?, does not affect EXP)

2.cannot handle LRC: test with larger cutoffs and/or LRC correction with arbitrary, single structure; check

<http://pubs.acs.org/doi/abs/10.1021/jp0735987>

3.3 GROMACS

GROMACS has some run input options which can simplify the procedure for setting up free energy calculations. Specifically, `couple-moltype` implicitly defines the initial and final states by giving a special tag to a molecule and controls whether intramolecular interactions of the tagged molecule are retained or not along the alchemical path. It should be used in absolute free energy calculations to tag the molecule which will be decoupled from the rest of the system. Using this in relative calculations is possible, but will result in unintended behavior and errors. `couple-lambda0` and `couple-lambda1` control the interactions of the molecule specified by `couple-moltype` with its surroundings. The entries `vdw-lambdas` and `fep-lambdas` define the lambda schedule. The former indicates the value of the λ vector component that modifies van der Waals interactions for each state, while the latter changes all λ vector components that are not specified in the `.mdp` file. For instance, in split protocol simulations, these entries are sets such that the components of the energy are modified in different stages. If the transformation involves particle deletion (“forward process”), `fep-lambdas` is set to change charges and bonds before `vdw-lambdas` changes van de Waals components. If the process involves particle insertion (“backward process”) we reverse the roles. In this work, `mass-lambdas` were all set to zero to not included mass changes in the free energy. Unified protocols set all λ vectors the same.

Table 6 lists the relative free energies obtained from GROMACS simulations. Relative free energies are in good agreement with each other and with $\Delta\Delta G_{\text{hydr}}$ obtained from the other software used in this study (cmp. Tables 1 and 3). A noteworthy exception is the difference between the unified and split results of methane \rightarrow methanol and its reverse

Table 6: Relative hydration free energies obtained from GROMACS simulations in $kcal \cdot mol^{-1}$. Signs of the backward transformation have been reverted to correspond to the forward transformation.

transformation	split ^a		unified ^b		absolute ^c	
	RF $\Delta\Delta G$	PME $\Delta\Delta G$	RF $\Delta\Delta G$	PME $\Delta\Delta G$	RF $\Delta\Delta G$	PME $\Delta\Delta G$
ethane	-0.025 \pm 0.005	-0.035 \pm 0.020	-0.017 \pm 0.003	-0.030 \pm 0.001	-0.06 \pm 0.01	-0.04 \pm 0.01
methane	-0.01 \pm 0.02	-0.02 \pm 0.01	0.046 \pm 0.020 ^d	0.01 \pm 0.02		
methanol	6.163 \pm 0.006	6.197 \pm 0.004	7.30 \pm 0.02	7.380 \pm 0.007	5.77 \pm 0.01	5.95 \pm 0.01
methane	6.168 \pm 0.005	6.199 \pm 0.008	7.09 \pm 0.02	7.17 \pm 0.02		
ethane	-6.123 \pm 0.007	-6.185 \pm 0.006	-7.117 \pm 0.005	-7.21 \pm 0.02	-5.83 \pm 0.01	-5.98 \pm 0.01
methanol	-6.124 \pm 0.005	-6.193 \pm 0.004	-7.338 \pm 0.004	-7.404 \pm 0.004		
toluene	3.22 \pm 0.01	3.211 \pm 0.006	3.229 \pm 0.008	3.22 \pm 0.01	2.97 \pm 0.01	3.16 \pm 0.01
methane	3.25 \pm 0.01	3.20 \pm 0.01	3.22 \pm 0.01	3.211 \pm 0.001		
neopentane ^e	-0.103 \pm 0.008	-0.15 \pm 0.02	-0.08 \pm 0.02	-0.18 \pm 0.03	-0.18 \pm 0.01	-0.14 \pm 0.01
methane ^e	-0.11 \pm 0.02	-0.16 \pm 0.05	0.00 \pm 0.03	-0.18 \pm 0.03		
neopentane ^f	-0.116 \pm 0.007	-0.13 \pm 0.01	-0.14 \pm 0.01	-0.14 \pm 0.01		
methane ^f	-0.10 \pm 0.03	-0.18 \pm 0.03	-0.089 \pm 0.007	-0.15 \pm 0.02		
2-methylfuran	2.986 \pm 0.006	2.930 \pm 0.050	3.05 \pm 0.01	3.00 \pm 0.01	2.87 \pm 0.01	2.95 \pm 0.01
methane	3.007 \pm 0.004	2.96 \pm 0.01	3.056 \pm 0.006	3.01 \pm 0.01		
2-methylindole	8.71 \pm 0.02	8.73 \pm 0.03	8.73 \pm 0.01	8.80 \pm 0.03	8.44 \pm 0.02	8.79 \pm 0.02
methane	8.73 \pm 0.03	8.74 \pm 0.01	8.30 \pm 0.02	8.77 \pm 0.04		
2-CPI	-0.07 \pm 0.02	-0.03 \pm 0.03	-0.10 \pm 0.05	-0.2 \pm 0.1	-0.02 \pm 0.05	0.02 \pm 0.02
7-CPI	-0.12 \pm 0.06	-0.20 \pm 0.04	-0.04 \pm 0.06	-0.14 \pm 0.09		

^aresults obtained from alchemical transformations with electrostatic and bonded scaling separate from vdW parameter change.

^bresults obtained from alchemical transformation with all parameters scaling together.

^cresults obtained from absolute free energy calculations.

^dinverted sign

^ecentral mapping

^fterminal mapping

process, which was investigated with additional split protocol simulations using Coulomb softcore potentials (Table 7).

Table 7: Relative hydration free energies of methanol \rightarrow methane and methane \rightarrow methanol transformations without and with the use of Coulomb softcore potentials from GROMACS. Signs of the backward transformation have been reverted to correspond to the forward transformation. The complete version of this table is in the SI.

transformation		split		split+sc		absolute	
		RF $\Delta\Delta G$	PME $\Delta\Delta G$	RF $\Delta\Delta G$	PME $\Delta\Delta G$	RF $\Delta\Delta G$	PME $\Delta\Delta G$
methanol	methane	6.163 ± 0.006	6.197 ± 0.004	7.32 ± 0.03	7.42 ± 0.04	5.77 ± 0.01	5.95 ± 0.01
methane	methanol	6.168 ± 0.005	6.199 ± 0.008	7.14 ± 0.03	7.21 ± 0.03		

We noticed a difference of approximately $1.5 \text{ kcal mol}^{-1}$ between the split protocol without Coulomb softcore potentials and both protocols that use it. Our data suggests that softcore use in electrostatic interactions requires adjustments in the λ -distance between states in the rapidly varying part of the $\partial\mathcal{H}/\partial\lambda$, see Figure S7 A test with combining the bonded terms with the vdW transformation did not change this result. Thus, we find that the split protocol without Coulomb softcore potentials is the most effective way to calculate relative free energy calculations. We also find that there is no significant difference in calculated free energies depending on the choice of Particle Mesh Ewald or Reaction Field for calculation of long-range electrostatic interactions.

One peculiarity of the software worth mentioning is that relative free energy simulations will crash if a hydrogen alchemically becomes a heavy atom if the simulation employs hydrogen bond constraining algorithms such as SHAKE or LINCS. Successful simulations require turning off the constraint and decreasing the time step. Alternative protocols that require some scripting and changes in the topology file could be pursued in the future. For instance 2-fs constraints protocols similar to those used in SOMD or AMBER in this study could be implemented via the definition of a new atom type for the hydrogen atoms that will be alchemically changed, or the use of `constraint = none` and of explicit constraints in the topology file.

3.4 SOMD

Fig. S4 compares relative free energy of hydration $\Delta\Delta G$ according to the protocol with unperturbed H bond constraints, with relative $\Delta\Delta G$ obtained from two absolute free energy calculations. Table 3 summarizes all the computed relative free energy of hydration for the dataset in Fig. 2. A very good agreement is observed between both methodologies ($R^2=0.99 \pm 0.01$ and $\text{MAD} = (0.10 \pm 0.03) \text{ kcal mol}^{-1}$), highlighting internal consistency within SOMD.

To achieve this level of reproducibility within SOMD it was crucial to pay close attention to constraints. Specifically, bonds that involve unperturbed hydrogen atoms are constrained. Bonds involving hydrogen atoms that are perturbed to a heavy elements are unconstrained. Additionally the atomic mass of the perturbed hydrogen atom is set to the mass of the heavy atom it is perturbed to. Bonds involving hydrogen atoms that are perturbed to another hydrogen atom type are constrained.

This protocol suppresses high frequency vibrations in flexible bonds involving hydrogen atoms, thus enabling a time step of 2 fs, whilst giving essentially negligible errors due to the use of constraints for perturbed bonds. This is apparent from the comparison with the absolute hydration free energy calculations. Additionally, the protocol yields relative hydration free energy extremely similar ($\text{MAE} = 0.09 \text{ kcal mol}^{-1}$) to those computed from simulations where no constraints are applied on the solutes and a timestep of 1 fs is used (See Figure S6).

By contrast, a protocol that constrains all bonds in a solute leads to significant differences with the absolute hydration free energies. For instance neopentane \rightarrow methane (centrally mapped) gives a RAFE $\Delta\Delta G=(2.04 \pm 0.01) \text{ kcal mol}^{-1}$ whereas the absolute hydration free energy calculations give $\Delta\Delta G=(-0.19 \pm 0.06) \text{ kcal mol}^{-1}$ as shown in tab. S6 and fig. S6.

This discrepancy occurs because in the SOMD implementation, the energies of constrained bonds are not evaluated, but the calculation of the energies of the solute at perturbed λ values is carried out using the coordinates of the reference λ trajectory. This leads

to a neglect of contributions of the bonded term (and associated coupled terms) to the free energy change. The effect is more pronounced for perturbations that feature a large change in equilibrium bond lengths, such as those where a hydrogen atom is perturbed to/from a heavy atom. This problem has already been discussed by Pearlman⁷⁶ and Boresch,^{30,56} as a missing bond-stretching term, which introduce a free energy offset estimated as:

$$\Delta G = RT \ln \left(\frac{r_i}{r_f} \right) \quad (7)$$

where r_i and r_f are the initial and final bond lengths, respectively.

The reaction fields implemented in SOMD and GROMACS differ somewhat (atom-based shifted Barker Watts⁶⁸ vs group based switched Barker Watts), but nevertheless SOMD and GROMACS RF produce comparable results with a MAD of 0.18 kcal mol⁻¹.

Overall, the SOMD free energy estimations are in good agreement with the other MD packages, as the MAE suggests (see Table XX). Reaction field and PME results are in good agreement. In particular, the unperturbed hydrogen bonds constraint protocol allows a timestep of 2 fs. All SOMD RAFF simulations were carried out with simultaneous transformation of Lennard-Jones, charges, and bonded terms. This suggests that the failure of the GROMACS “unified protocol” in some instances may be due to differences in the softcore Coulomb implementations.

For the methane \rightarrow neopentane transformations SOMD yields consistent results between central and terminal mappings, as shown in Table S5 while GROMACS RF displays a $\Delta\Delta G$ of (-0.20 ± 0.01) kcal mol⁻¹ for the centrally mapped transformation and $\Delta\Delta G = (-0.05 \pm 0.02)$ kcal mol⁻¹ for the terminally mapped transformation. The SOMD RF calculations show a MAD = 0.18 kcal mol⁻¹ and MAD = 0.23 kcal mol⁻¹ with GROMACS PME and AMBER respectively, similar to the MAD obtained with GROMACS RF.

4 Discussion

We have studied reproducibility under the view point of using a set of different MD packages with the same force field.

3.recommended protocols

4.protocols to avoid

5.lessons learned

6.We can't do convergence tests so only discuss and explain why not done: different lambda schedules, number of windows, length of each windows; codes have different speeds ("time to answer"). Instead focus is on low errors to establish if codes are different or not.

7.SHAKE/LINCS issues with changing bond lengths

8.2-cyclopentanylindole to 7-cyclopentanylindole: better to go through intermediates?

9.what do we need to progress the field e.g. automation to make things easy but also consistent and thus more reproducible (FESetup also for reproducibility); GPU: GROMACS, SOMD but not AMBER (yet) and CHARMM; alternative softcore functions?; sampling?; force field improvements?; analysis?

10.developer notes: constraints, both appearing/disappearing; lambda paths for AMBER (relative), absolute: crgmask requires vacuum corr if split protocol

11.further investigation required: binding RAFEs?

ToDo

P.

1. bug in TI gradient accumulation in parallel runs (does not affect serial?, does not affect EXP) 24
2. cannot handle LRC: test with larger cutoffs and/or LRC correction with arbitrary, single structure; check <http://pubs.acs.org/doi/abs/10.1021/jp0735987> 24
3. recommended protocols 29
4. protocols to avoid 29

5. lessons learned	29
6. We can't do convergence tests so only discuss and explain why not done: different lambda schedules, number of windows, length of each windows; codes have different speeds ("time to answer"). Instead focus is on low errors to establish if codes are different or not.	29
7. SHAKE/LINCS issues with changing bond lengths	29
8. 2-cyclopentanylindole to 7-cyclopentanylindole: better to go through intermediates?	29
9. what do we need to progress the field e.g. automation to make things easy but also consistent and thus more reproducible (FESetup also for reproducibility); GPU: GROMACS, SOMD but not AMBER (yet) and CHARMM; alternative softcore functions?; sampling?; force field improvements?; analysis?	29
10. developer notes: constraints, both appearing/disappearing; lambda paths for AM- BER (relative), absolute: crgmask requires vacuum corr if split protocol	29
11. further investigation required: binding RAFEs?	29

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References

- (1) Hansen, N.; Van Gunsteren, W. F. Practical aspects of free-energy calculations: A review. *Journal of Chemical Theory and Computation* **2014**, *10*, 2632–2647.
- (2) Pohorille, A.; Jarzynski, C.; Chipot, C. Good Practices in Free-Energy Calculations. *The Journal of Physical Chemistry B* **2010**, *114*, 10235–10253, PMID: 20701361.
- (3) Gallicchio, E.; Levy, R. M. In *Computational chemistry methods in structural biology*; Christov, C., Ed.; Advances in Protein Chemistry and Structural Biology; Academic Press, 2011; Vol. 85; pp 27 – 80.
- (4) Lu, X.; Fang, D.; Ito, S.; Okamoto, Y.; Ovchinnikov, V.; Cui, Q. QM/MM free energy simulations: recent progress and challenges. *Molecular Simulation* **2016**, *42*, 1056–1078, PMID: 27563170.
- (5) Rickman, J. M.; LeSar, R. Free-Energy Calculations in Materials Research. *Annual Review of Materials Research* **2002**, *32*, 195–217.
- (6) Wan, S.; Knapp, B.; Wright, D. W.; Deane, C. M.; Coveney, P. V. Rapid, Precise, and Reproducible Prediction of Peptide–MHC Binding Affinities from Molecular Dynamics That Correlate Well with Experiment. *Journal of Chemical Theory and Computation* **2015**, *11*, 3346–3356, PMID: 26575768.

- (7) Beveridge, D. L.; Dicapua, F. M. Free Energy Via Molecular Simulation: Applications to Chemical and Biomolecular Systems. *Annual Review of Biophysics and Biophysical Chemistry* **1989**, *18*, 431–492.
- (8) Straatsma, T. P.; Mccammon, J. A. Computational Alchemy. *Annual Review of Physical Chemistry* **1992**, *43*, 407–435.
- (9) Kollman, P. Free energy calculations: Applications to chemical and biochemical phenomena. *Chemical Reviews* **1993**, *93*, 2395–2417.
- (10) Squire, D. R.; Hoover, W. G. Monte Carlo Simulation of Vacancies in Rare-Gas Crystals. *The Journal of Chemical Physics* **1969**, *50*, 701–706.
- (11) Bennett, C. H. Efficient estimation of free energy differences from Monte Carlo data. *Journal of Computational Physics* **1976**, *22*, 245–268.
- (12) Mruzik, M. R.; Abraham, F. F.; Schreiber, D. E.; Pound, G. M. A Monte Carlo study of ion–water clusters. *The Journal of Chemical Physics* **1976**, *64*, 481–491.
- (13) Postma, J. P. M.; Berendsen, H. J. C.; Haak, J. R. Thermodynamics of cavity formation in water. A molecular dynamics study. *Faraday Symp. Chem. Soc.* **1982**, *17*, 55–67.
- (14) Tembe, B. L.; McCammon, J. Ligand-receptor interactions. *Computers & Chemistry* **1984**, *8*, 281 – 283.
- (15) Jorgensen, W. L.; Ravimohan, C. Monte Carlo simulation of differences in free energies of hydration. *The Journal of Chemical Physics* **1985**, *83*, 3050–3054.
- (16) Gilson, M.; Given, J.; Bush, B.; McCammon, J. The statistical-thermodynamic basis for computation of binding affinities: a critical review. *Biophysical Journal* **1997**, *72*, 1047 – 1069.

- (17) Boresch, S.; Tettinger, F.; Leitgeb, M.; Karplus, M. Absolute Binding Free Energies: A Quantitative Approach for Their Calculation. *The Journal of Physical Chemistry B* **2003**, *107*, 9535–9551.
- (18) Deng, Y.; Roux, B. Computations of Standard Binding Free Energies with Molecular Dynamics Simulations. *The Journal of Physical Chemistry B* **2009**, *113*, 2234–2246.
- (19) Ytreberg, F. M.; Swendsen, R. H.; Zuckerman, D. M. Comparison of free energy methods for molecular systems. *Journal of Chemical Physics* **2006**, *125*, 184114.
- (20) Gapsys, V.; Michielssens, S.; Seeliger, D.; de Groot, B. L. pmx: Automated protein structure and topology generation for alchemical perturbations. *Journal of Computational Chemistry* **2015**, *36*, 348–354.
- (21) Sandberg, R. B.; Banchelli, M.; Guardiani, C.; Menichetti, S.; Caminati, G.; Procacci, P. Efficient Nonequilibrium Method for Binding Free Energy Calculations in Molecular Dynamics Simulations. *Journal of Chemical Theory and Computation* **2015**, *11*, 423–435, PMID: 26580905.
- (22) Duarte Ramos Matos, G.; Kyu, D. Y.; Loeffler, H. H.; Chodera, J. D.; Shirts, M. R.; Mobley, D. L. Approaches for Calculating Solvation Free Energies and Enthalpies Demonstrated with an Update of the FreeSolv Database. *Journal of Chemical & Engineering Data* **0**, *0*, null.
- (23) Jorgensen, W. L.; Buckner, J. K.; Boudon, S.; Tirado-Rives, J. Efficient computation of absolute free energies of binding by computer simulations. Application to the methane dimer in water. *The Journal of chemical physics* **1988**, *89*, 3742–3746.
- (24) Woods, C. J.; Malaisree, M.; Hannongbua, S.; Mulholland, A. J. A water–swap reaction coordinate for the calculation of absolute protein–ligand binding free energies. *The Journal of Chemical Physics* **2011**, *134*, 054114.

- (25) Woods, C. J.; Malaisree, M.; Michel, J.; Long, B.; McIntosh-Smith, S.; Mulholland, A. J. Rapid decomposition and visualisation of protein–ligand binding free energies by residue and by water. *Faraday Discuss.* **2014**, *169*, 477–499.
- (26) Wang, L.; Wu, Y.; Deng, Y.; Kim, B.; Pierce, L.; Krilov, G.; Lupyan, D.; Robinson, S.; Dahlgren, M. K.; Greenwood, J.; Romero, D. L.; Masse, C.; Knight, J. L.; Steinbrecher, T.; Beuming, T.; Damm, W.; Harder, E.; Sherman, W.; Brewer, M.; Wester, R.; Murcko, M.; Frye, L.; Farid, R.; Lin, T.; Mobley, D. L.; Jorgensen, W. L.; Berne, B. J.; Friesner, R. A.; Abel, R. Accurate and Reliable Prediction of Relative Ligand Binding Potency in Prospective Drug Discovery by Way of a Modern Free-Energy Calculation Protocol and Force Field. *Journal of the American Chemical Society* **2015**, *137*, 2695–2703, PMID: 25625324.
- (27) Wang, L.; Deng, Y.; Wu, Y.; Kim, B.; LeBard, D. N.; Wandschneider, D.; Beachy, M.; Friesner, R. A.; Abel, R. Accurate Modeling of Scaffold Hopping Transformations in Drug Discovery. *Journal of Chemical Theory and Computation* **2017**, *13*, 42–54, PMID: 27933808.
- (28) Pearlman, D. A. A Comparison of Alternative Approaches to Free Energy Calculations. *The Journal of Physical Chemistry* **1994**, *98*, 1487–1493.
- (29) Michel, J.; Essex, J. W. Prediction of protein–ligand binding affinity by free energy simulations: assumptions, pitfalls and expectations. *Journal of Computer-Aided Molecular Design* **2010**, *24*, 639–658.
- (30) Boresch, S.; Karplus, M. The Role of Bonded Terms in Free Energy Simulations: 1. Theoretical Analysis. *The Journal of Physical Chemistry A* **1999**, *103*, 103–118.
- (31) Liu, S.; Wang, L.; Mobley, D. L. Is Ring Breaking Feasible in Relative Binding Free Energy Calculations? *Journal of Chemical Information and Modeling* **2015**, *55*, 727–735, PMID: 25835054.

- (32) Shobana, S.; Roux, B.; Andersen, O. S. Free Energy Simulations: Thermodynamic Reversibility and Variability. *The Journal of Physical Chemistry B* **2000**, *104*, 5179–5190.
- (33) Michel, J.; Verdonk, M. L.; Essex, J. W. Protein–Ligand Complexes: Computation of the Relative Free Energy of Different Scaffolds and Binding Modes. *Journal of Chemical Theory and Computation* **2007**, *3*, 1645–1655, PMID: 26627610.
- (34) Rocklin, G. J.; Mobley, D. L.; Dill, K. A. Separated topologies—A method for relative binding free energy calculations using orientational restraints. *The Journal of Chemical Physics* **2013**, *138*, 085104.
- (35) Axelsen, P. H.; Li, D. Improved convergence in dual-topology free energy calculations through use of harmonic restraints. *Journal of Computational Chemistry* **1998**, *19*, 1278–1283.
- (36) Case, D. A.; Cheatham III, T. E.; Darden, T.; Gohlke, H.; Luo, R.; Merz Jr., K. M.; Onufriev, A.; Simmerling, C.; Wang, B.; Woods, R. J. The Amber biomolecular simulation programs. *Journal of Computational Chemistry* **2005**, *26*, 1668–1688.
- (37) Brooks, B. R.; Brooks, C. L.; Mackerell, A. D.; Nilsson, L.; Petrella, R. J.; Roux, B.; Won, Y.; Archontis, G.; Bartels, C.; Boresch, S.; Caffisch, A.; Caves, L.; Cui, Q.; Dinner, A. R.; Feig, M.; Fischer, S.; Gao, J.; Hodoscek, M.; Im, W.; Kuczera, K.; Lazaridis, T.; Ma, J.; Ovchinnikov, V.; Paci, E.; Pastor, R. W.; Post, C. B.; Pu, J. Z.; Schaefer, M.; Tidor, B.; Venable, R. M.; Woodcock, H. L.; Wu, X.; Yang, W.; York, D. M.; Karplus, M. CHARMM: The biomolecular simulation program. *Journal of Computational Chemistry* **2009**, *30*, 1545–1614.
- (38) Abraham, M. J.; Murtola, T.; Schulz, R.; Páll, S.; Smith, J. C.; Hess, B.; Lindahl, E. GROMACS: High performance molecular simulations through multi-level parallelism from laptops to supercomputers. *SoftwareX* **2015**, *1–2*, 19–25.

- (39) Scott, W. R. P.; Hünenberger, P. H.; Tironi, I. G.; Mark, A. E.; Billeter, S. R.; Fennen, J.; Torda, A. E.; Huber, T.; Krüger, P.; van Gunsteren, W. F. The GROMOS Biomolecular Simulation Program Package. *The Journal of Physical Chemistry A* **1999**, *103*, 3596–3607.
- (40) Woods, C.; Mey, A. S.; Calabro, G.; Michel, J. Sire molecular simulations framework. 2016.
- (41) Eastman, P.; Friedrichs, M. S.; Chodera, J. D.; Radmer, R. J.; Bruns, C. M.; Ku, J. P.; Beauchamp, K. A.; Lane, T. J.; Wang, L.-P.; Shukla, D.; Tye, T.; Houston, M.; Stich, T.; Klein, C.; Shirts, M. R.; Pande, V. S. OpenMM 4: A Reusable, Extensible, Hardware Independent Library for High Performance Molecular Simulation. *Journal of Chemical Theory and Computation* **2013**, *9*, 461–469, PMID: 23316124.
- (42) Shirts, M. R.; Klein, C.; Swails, J. M.; Yin, J.; Gilson, M. K.; Mobley, D. L.; Case, D. A.; Zhong, E. D. Lessons learned from comparing molecular dynamics engines on the SAMPL5 dataset. *Journal of Computer-Aided Molecular Design* **2017**, *31*, 147–161.
- (43) Schappals, M.; Mecklenfeld, A.; KrÄger, L.; Botan, V.; KÄster, A.; Stephan, S.; GarcÄa, E. J.; Rutkai, G.; Raabe, G.; Klein, P.; Leonhard, K.; Glass, C. W.; Lenhard, J.; Vrabec, J.; Hasse, H. Round Robin Study: Molecular Simulation of Thermodynamic Properties from Models with Internal Degrees of Freedom. *Journal of Chemical Theory and Computation* **0**, *0*, null, PMID: 28738147.
- (44) Pronk, S.; Larsson, P.; Pouya, I.; Bowman, G. R.; Haque, I. S.; Beauchamp, K.; Hess, B.; Pande, V. S.; Kasson, P. M.; Lindahl, E. Copernicus: A New Paradigm for Parallel Adaptive Molecular Dynamics. Proceedings of 2011 International Conference for High Performance Computing, Networking, Storage and Analysis. New York, NY, USA, 2011; pp 60:1–60:10.
- (45) Sadiq, S. K.; Wright, D.; Watson, S. J.; Zasada, S. J.; Stoica, I.; Coveney, P. V.

- Automated Molecular Simulation Based Binding Affinity Calculator for Ligand-Bound HIV-1 Proteases. *Journal of Chemical Information and Modeling* **2008**, *48*, 1909–1919, PMID: 18710212.
- (46) Lundborg, M.; Lindahl, E. Automatic GROMACS Topology Generation and Comparisons of Force Fields for Solvation Free Energy Calculations. *The Journal of Physical Chemistry B* **2015**, *119*, 810–823, PMID: 25343332.
- (47) Loeffler, H. H.; Michel, J.; Woods, C. FESetup: Automating Setup for Alchemical Free Energy Simulations. *Journal of Chemical Information and Modeling* **2015**, *55*, 2485–2490.
- (48) Balasubramanian, V.; Bethune, I.; Shkurti, A.; Breitmoser, E.; Hruska, E.; Clementi, C.; Laughton, C. A.; Jha, S. ExTASY: Scalable and Flexible Coupling of MD Simulations and Advanced Sampling Techniques. *CoRR* **2016**, *abs/1606.00093*.
- (49) Skyner, R. E.; McDonagh, J. L.; Groom, C. R.; van Mourik, T.; Mitchell, J. B. O. A Review of Methods for the Calculation of Solution Free Energies and the Modelling of Systems in Solution. *Phys. Chem. Chem. Phys.* **2015**, *17*, 6174–6191.
- (50) Mobley, D. L.; Guthrie, J. P. FreeSolv: a database of experimental and calculated hydration free energies, with input files. *Journal of Computer-Aided Molecular Design* **2014**, *28*, 711–720.
- (51) Beutler, T. C.; Mark, A. E.; van Schaik, R. C.; Gerber, P. R.; van Gunsteren, W. F. Avoiding singularities and numerical instabilities in free energy calculations based on molecular simulations. *Chemical Physics Letters* **1994**, *222*, 529–539.
- (52) Zacharias, M.; Straatsma, T. P.; McCammon, J. A. Separation-shifted scaling, a new scaling method for Lennard–Jones interactions in thermodynamic integration. *The Journal of Chemical Physics* **1994**, *100*, 9025–9031.

- (53) Steinbrecher, T.; Mobley, D. L.; Case, D. A. Nonlinear scaling schemes for Lennard-Jones interactions in free energy calculations. *The Journal of Chemical Physics* **2007**, *127*, 214108.
- (54) Cross, A. J. Influence of hamiltonian parameterization on convergence of kirkwood free energy calculations. *Chemical Physics Letters* **1986**, *128*, 198 – 202.
- (55) Phillips, J. C.; Braun, R.; Wang, W.; Gumbart, J.; Tajkhorshid, E.; Villa, E.; Chipot, C.; Skeel, R. D.; Kalé, L.; Schulten, K. Scalable molecular dynamics with NAMD. *Journal of Computational Chemistry* **2005**, *26*, 1781–1802.
- (56) Boresch, S.; Karplus, M. The Role of Bonded Terms in Free Energy Simulations. 2. Calculation of Their Influence on Free Energy Differences of Solvation. *The Journal of Physical Chemistry A* **1999**, *103*, 119–136.
- (57) Rocklin, G. J.; Mobley, D. L.; Dill, K.; Hünenberger, P. H. Calculating the binding free energies of charged species based on explicit-solvent simulations employing lattice-sum methods: an accurate correction scheme for electrostatic finite-size effects. *The Journal of chemical physics* **2013**, *139*, 184103.
- (58) Loeffler, H. H.; Sotriffer, C. A.; Winger, R. H.; Liedl, K. R.; Rode, B. M. Calculation of sequence-dependent free energies of hydration of dipeptides formed by alanine and glycine. *Journal of Computational Chemistry* **2001**, *22*, 846–860.
- (59) Wang, J.; Wolf, R. M.; Caldwell, J. W.; Kollman, P. A.; Case, D. A. Development and testing of a general amber force field. *Journal of Computational Chemistry* **2004**, *25*, 1157–1174.
- (60) Jakalian, A.; Bush, B. L.; Jack, D. B.; Bayly, C. I. Fast, efficient generation of high-quality atomic charges. AM1-BCC model: I. Method. *Journal of Computational Chemistry* **2000**, *21*, 132–146.

- (61) Jakalian, A.; Jack, D. B.; Bayly, C. I. Fast, efficient generation of high-quality atomic charges. AM1-BCC model: II. Parameterization and validation. *Journal of Computational Chemistry* **2002**, *23*, 1623–1641.
- (62) Jorgensen, W. L.; Chandrasekhar, J.; Madura, J. D.; Impey, R. W.; Klein, M. L. Comparison of simple potential functions for simulating liquid water. *The Journal of Chemical Physics* **1983**, *79*, 926–935.
- (63) Shivakumar, D.; Harder, E.; Damm, W.; Friesner, R. A.; Sherman, W. Improving the Prediction of Absolute Solvation Free Energies Using the Next Generation OPLS Force Field. *Journal of Chemical Theory and Computation* **2012**, *8*, 2553–2558, PMID: 26592101.
- (64) Hu, Y.; Sherborne, B.; Lee, T.-S.; Case, D. A.; York, D. M.; Guo, Z. The importance of protonation and tautomerization in relative binding affinity prediction: a comparison of AMBER TI and Schrödinger FEP. *Journal of Computer-Aided Molecular Design* **2016**, *30*, 533–539.
- (65) Steinbrecher, T.; Joung, I.; Case, D. A. Soft-core potentials in thermodynamic integration: comparing one- and two-step transformations. *Journal of Computational Chemistry* **2011**, *32*, 3253–3263.
- (66) Naden, L. N.; Pham, T. T.; Shirts, M. R. Linear basis function approach to efficient alchemical free energy calculations. 1. Removal of uncharged atomic sites. *Journal of Chemical Theory and Computation* **2014**, *10*, 1128–1149.
- (67) Naden, L. N.; Shirts, M. R. Linear basis function approach to efficient alchemical free energy calculations. 2. Inserting and deleting particles with coulombic interactions. *Journal of Chemical Theory and Computation* **2015**, *11*, 2536–2549.
- (68) Barker, J.; Watts, R. Monte Carlo studies of the dielectric properties of water-like models. *Molecular Physics* **1973**, *26*, 789–792.

- (69) Andersen, H. C. Molecular dynamics simulations at constant pressure and/or temperature. *The Journal of Chemical Physics* **1980**, *72*, 2384–2393.
- (70) Bosisio, S.; Mey, A. S. J. S.; Michel, J. Blinded predictions of distribution coefficients in the SAMPL5 challenge. *Journal of Computer-Aided Molecular Design* **2016**, *30*, 1101–1114.
- (71) Zwanzig, R. High-Temperature Equation of State by a Perturbation Method. I. Non-polar Gases. *The Journal of Chemical Physics* **1954**, *22*, 1420–1426.
- (72) Klimovich, P. V.; Shirts, M. R.; Mobley, D. L. Guidelines for the analysis of free energy calculations. *Journal of Computer-Aided Molecular Design* **2015**, *29*, 397–411.
- (73) Klimovich, P. V.; Mobley, D. L. Predicting hydration free energies using all-atom molecular dynamics simulations and multiple starting conformations. *Journal of Computer-Aided Molecular Design* **2010**, *24*, 307–316.
- (74) Lee, T.-S.; Hu, Y.; Sherborne, B.; Guo, Z.; York, D. M. Toward Fast and Accurate Binding Affinity Prediction with pmemdGTI: An Efficient Implementation of GPU-Accelerated Thermodynamic Integration. *Journal of Chemical Theory and Computation* **0**, *0*, null, PMID: 28618232.
- (75) Kaus, J. W.; Pierce, L. T.; Walker, R. C.; McCammon, J. A. Improving the Efficiency of Free Energy Calculations in the Amber Molecular Dynamics Package. *Journal of Chemical Theory and Computation* **2013**, *9*, 4131–4139.
- (76) Pearlman, D. A.; Kollman, P. A. The overlooked bond-stretching contribution in free energy perturbation calculations. *The Journal of chemical physics* **1991**, *94*, 4532–4545.

Graphical TOC Entry

