

# Program of Medical Informatics Faculty of Computers and Information Mansoura University

#### **Detecting Genetic Variations -II**

(Genomics)

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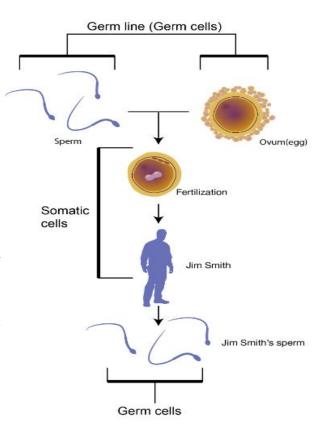


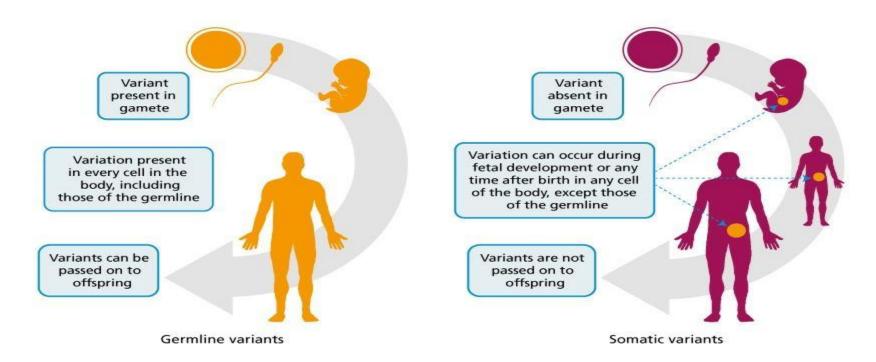




A germ line is the sex cells (eggs and sperm) that are used by sexually reproducing organisms to pass on genes from generation to generation. Egg and sperm cells are called germ cells, in contrast to the other cells of the body that are called somatic cells.

□A gene change in a reproductive cell (egg or sperm) that becomes incorporated into the DNA of every cell in the body of the offspring. A variant (or mutation) contained within the germline can be passed from parent to offspring, and is, therefore, hereditary. Also called germline mutation.





## Genetic Allele

- An allele is a variant form of a gene. Some genes have a variety of different forms, which are located at the same position, or genetic locus, on a chromosome.
- Humans are called diploid organisms because they have two alleles at each genetic locus, with one allele inherited from each parent.

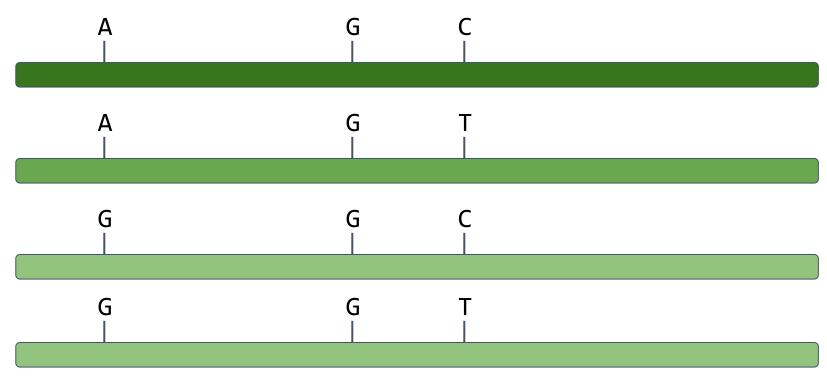
#### Genetic Allele

- Each pair of alleles represents the genotype of a specific gene.
- Genotypes are described as homozygous if there are two identical alleles at a particular locus and as heterozygous if the two alleles differ.
- Alleles contribute to the organism's phenotype, which is the outward appearance of the organism.

#### Genetic Allele

- Some alleles are dominant or recessive. When an organism is heterozygous at a specific locus and carries one dominant and one recessive allele, the organism will express the dominant phenotype.
- A haplotype (haploid genotype) is a group of alleles in an organism that are inherited together from a single parent.
- In addition, the term "haplotype" can also refer to the inheritance of a cluster of single nucleotide polymorphisms (SNPs), which are variations at single positions in the DNA sequence among individuals.

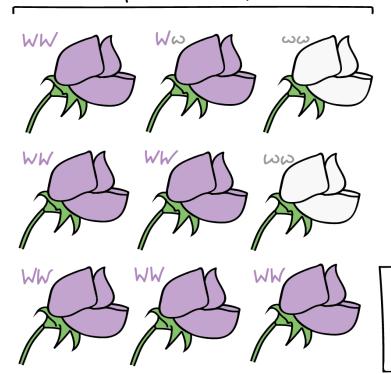
# haplotype

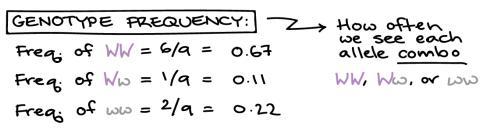


Slides adapted from Aaron Quinlan: https://github.com/quinlan-lab/applied-computational-genomics

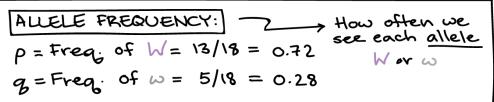
# Genotype vs. Allele frequency

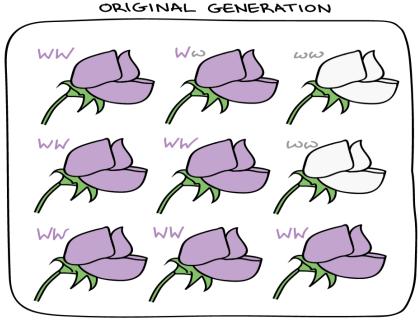
Population of peas





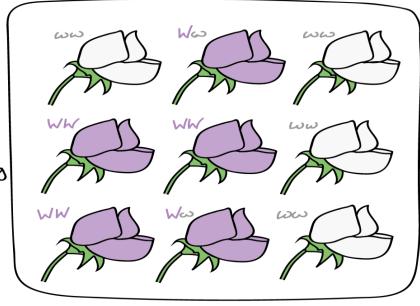
PHENOTYPE FREQUENCY: 
$$2 \rightarrow 100$$
 often we see white Freq. of purple =  $7/9 = 0.78$  vs. purple Freq. of white =  $2/9 = 0.22$ 





p = Frequency of W = 13/18 = 0.72g = Frequency of  $\omega = 5/18 = 0.28$ 

#### NEW GENERATION



p = Frequency of W = 8/18 = 0.44g = Frequency of W = 10/18 = 0.56

Allele frequencies change -> population evolves

old plants die

Their

# Hardy-Weinberg equation

- The equation is an expression of the principle known as Hardy-Weinberg equilibrium, which states that the amount of genetic variation in a population will remain constant from one generation to the next in the absence of disturbing factors.
- To explore the Hardy-Weinberg equation, we can examine a simple genetic locus at which there are two alleles, A and a. The Hardy-Weinberg equation is expressed as:

$$p^2 + 2pq + q^2 = 1$$

# Hardy-Weinberg equation

- p is the frequency of the "A" allele and q is the frequency of the "a" allele in the population. In the equation, p<sup>2</sup> represents the frequency of the homozygous genotype AA, q<sup>2</sup> represents the frequency of the homozygous genotype aa, and 2pq represents the frequency of the heterozygous genotype Aa.
- In addition, the sum of the allele frequencies for all the alleles at the locus must be 1, so p + q = 1. If the p and q allele frequencies are known, then the frequencies of the three genotypes may be calculated using the Hardy-Weinberg equation.

# Hardy-Weinberg Equilibrium

Polymorphic loci that are biallelic (e.g., A and G alleles) have two allele frequencies, <u>p</u> and <u>q</u>.

$$f(A) = p = 4/6 = 0.67$$

$$f(G) = q = 2/6 = 0.33$$

$$p + q = 1$$

# Hardy-Weinberg Equilibrium

In the absence of evolutionary forces such as selection, drift, or bottlenecks, Hardy—Weinberg equilibrium states that allele and genotype frequencies in a population will remain constant from generation to generation. If we know the allele frequencies, p and q, we can predict the genotype frequencies that should be observed (binomial expectation).

$$f(A) = p = 4/6 = 0.67$$
  
 $f(G) = q = 2/6 = 0.33$ 

$$f(AA) = p^2 = (0.67)^2 = 0.4489$$
  
 $f(AG) = 2pq = 2(0.67)(0.33) = 0.4422$   
 $f(GG) = q^2 = (0.33)^2 = 0.1089$ 

$$p^2 + 2pq + q^2 = 1$$

# Hardy-Weinberg Equilibrium: expected genotype freqs

$$p = 0.5, q = 0.5$$
  
 $f(AA) = p^2 = (0.5)^2 = 0.25$   
 $f(AG) = 2pq = 2(0.5)(0.5) = 0.5$   
 $f(GG) = q^2 = (0.5)^2 = 0.25$ 

$$p = 0.01, q = 0.99$$

$$f(AA) = p^2 = (0.01)^2 = 0.0001$$

$$f(AG) = 2pq = 2(0.01)(0.99) = 0.0198$$

$$f(GG) = q^2 = (0.99)^2 = 0.9801$$

$$p = 0.1$$
,  $q = 0.9$   
 $f(AA) = p^2 = (0.1)^2 = 0.01$   
 $f(AG) = 2pq = 2(0.1)(0.9) = 0.18$   
 $f(GG) = q^2 = (0.9)^2 = 0.81$ 

$$p = 0.001, q = 0.999$$

$$f(AA) = p^2 = (0.001)^2 = 0.000001$$

$$f(AG) = 2pq = 2(0.001)(0.999) = 0.001998$$

$$f(GG) = q^2 = (0.999)^2 = 0.998001$$

# Hardy-Weinberg Equilibrium

$$p = 0.1$$
,  $q = 0.9$   
 $f(AA) = p^2 = (0.1)^2 = 0.01$   
 $f(AG) = 2pq = 2(0.1)(0.9) = 0.18$   
 $f(GG) = q^2 = (0.9)^2 = 0.81$ 

If we sequenced 100 individuals, how many A/G heterozygotes would we expect? How many A/A homozygotes?

# Given genotype frequencies, calculate allele frequencies in a gene pool!

Alleles = A, a

Genotypes = AA, Aa, aa

Frequency of allele A: f(A) = f(AA) + 1/2 f(Aa)

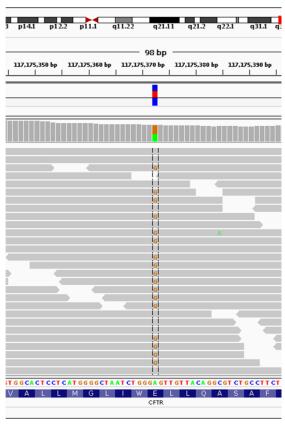
Frequency of allele a: f(a) = f(aa) + 1/2 f(Aa)





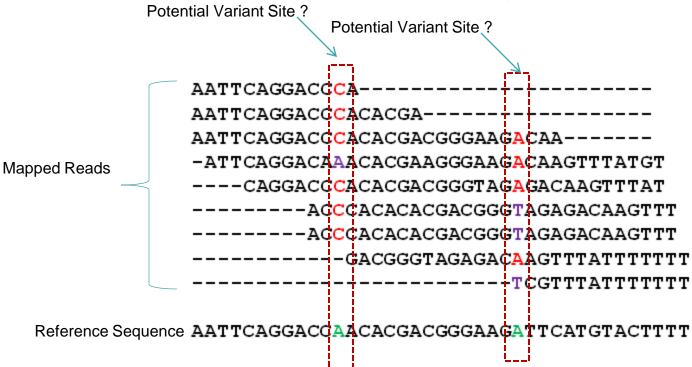


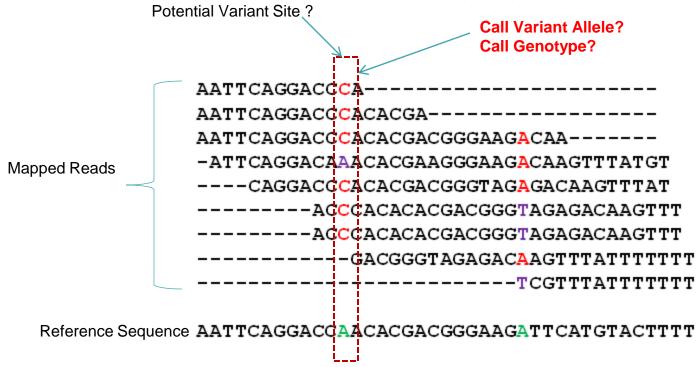
#### What information is needed to decide if a variant exists?



- Depth of coverage at the locus
- Bases observed at the locus
- The base qualities of each allele
- The strand composition
- Mapping qualities
- Proper pairs?
- Expected polymorphism rate

Reference Sequence AATTCAGGACCAACACGACGGGAAGATTCATGTACTTTT

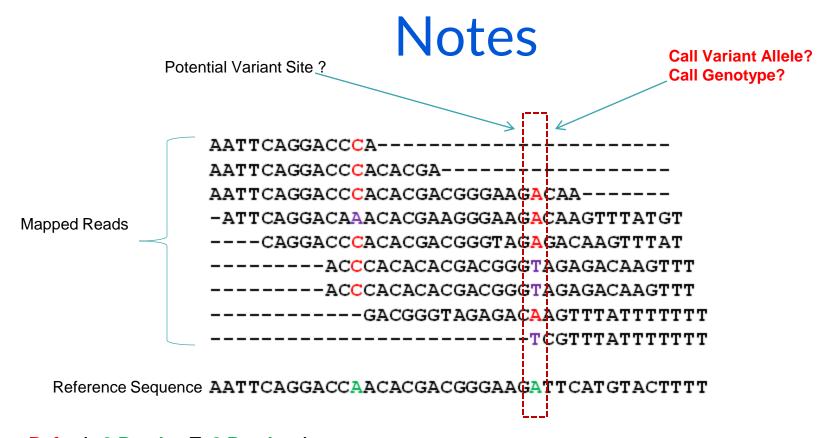




Ref. =A, 6 Reads = C, 1 Reads =A

So:

This potential site has a Variant non reference Allele C and homozygous non reference genotype CC.



Ref. =A, 3 Reads = T, 3 Reads =A
So: This potential site has a Variant reference Allele A and non reference Allele T and heterozygous genotype AT.

Suppose that there are at maximum two alleles a and b at each site and we want to call a variants along the reference sequence.

AATTCAGGACCCA-----AATTCAGGACCCACACGA-----AATTCAGGACCCACACGACGGGAAGACAA--ATTCAGGACAAACACGAAGGGAAGACAAGTTTATGT Mapped Reads ----CAGGACCCACACGACGGTAGAGACAAGTTTAT ----ACCCACACGACGGGTAGAGACAAGTTT -----ACCCACACACGACGGGTAGAGACAAGTTT -----GACGGGTAGAGACAAGTTTATTTTTTT ----TCGTTTATTTTT Reference Sequence AATTCAGGACCAACACGACGGGAAGATTCATGTACTTTT

6

Reference Allele

None-Reference Allele

a 3 3

 $\mathbf{b} \mid \mathbf{0} \mid \mathbf{0}$ 

0

# Pipeline SNP detection

Sequencing Reads Aligner BAM Variant Caller VCF file Variant Filtering

SNP DB

Genotyper, allele calling, are another names for variant calling

# Pipeline SNP detection

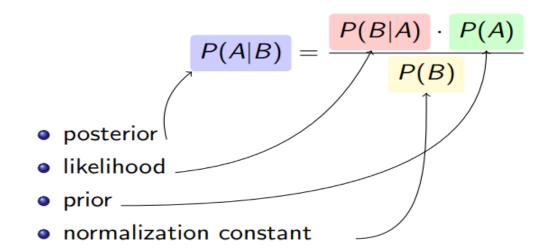
☐ The purpose of the Probabilistic Variant Caller is to identify variants in a sample by using a probabilistic model built from read mapping data.

□This tool can detect variants in data sets from haploid (e.g. Bacteria), diploid (e.g. Human) and polyploid organisms (e.g. Cancer and higher plants).



SNP DB

Bayes' theorem follows from the definition of the conditional probability and relates the conditional probability P(A|B) to P(B|A) for two events A and B such that  $P(B) \neq 0$ :



Example 1. There are three types of coins which have different probabilities of landing heads when tossed.

- Type A coins are fair, with probability 0.5 of heads
- Type B coins are bent and have probability 0.6 of heads
- Type C coins are bent and have probability 0.9 of heads

Suppose I have a drawer containing 5 coins: 2 of type A, 2 of type B, and 1 of type C. I reach into the drawer and pick a coin at random. Without showing you the coin I flip it once and get heads. What is the probability it is type A? Type B? Type C?

☐ Let:

A: Event that the chosen coin was type A.

B: Event that the chosen coin was type B.

C: Event that the chosen coin was type C.

D: Event that the toss was head.

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Prior Information before doing the experiment which is fixed and not changed if we change the experiment. P(A) = 2/5, P(B) = 2/5, P(C) = 1/5.

Example 1. There are three types of coins which have different probabilities of landing heads when tossed.

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Data or Observations during doing the experiment and can be changed if the experiment is changed.

Example 1. There are three types of coins which have different probabilities of landing heads when tossed.

- Type A coins are fair, with probability 0.5 of heads
- Type B coins are bent and have probability 0.6 of heads
- ullet Type C coins are bent and have probability 0.9 of heads

Likelihood which describes
how likely that our observation
or data is true given some
hypotheses.
P(D|A)

Suppose I have a drawer containing 5 coins: 2 of type A, 2 of type B, and 1 of type C. I reach into the drawer and pick a coin at random. Without showing you the coin I flip it once and get heads. What is the probability it is type A? Type B? Type C?

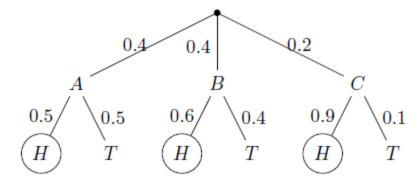
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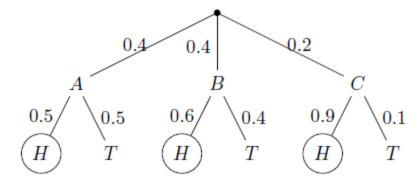
Posterior probability which describes what is the probability that a coin of type A, B, C given that the toss was head which our observations. P(A|D)

First we organize the probabilities into a tree:



Probability tree for choosing and tossing a coin.

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Probability tree for choosing and tossing a coin.

Bayes' theorem says, e.g.  $P(A|\mathcal{D}) = \frac{P(\mathcal{D}|A)P(A)}{P(\mathcal{D})}$ . The denominator  $P(\mathcal{D})$  is computed using the law of total probability:

$$P(\mathcal{D}) = P(\mathcal{D}|A)P(A) + P(\mathcal{D}|B)P(B) + P(\mathcal{D}|C)P(C) = 0.5 \cdot 0.4 + 0.6 \cdot 0.4 + 0.9 \cdot 0.2 = 0.62.$$

Now each of the three posterior probabilities can be computed:

$$P(A|\mathcal{D}) = \frac{P(\mathcal{D}|A)P(A)}{P(\mathcal{D})} = \frac{0.5 \cdot 0.4}{0.62} = \frac{0.2}{0.62}$$

$$P(B|\mathcal{D}) = \frac{P(\mathcal{D}|B)P(B)}{P(\mathcal{D})} = \frac{0.6 \cdot 0.4}{0.62} = \frac{0.24}{0.62}$$

$$P(C|\mathcal{D}) = \frac{P(\mathcal{D}|C)P(C)}{P(\mathcal{D})} = \frac{0.9 \cdot 0.2}{0.62} = \frac{0.18}{0.62}$$

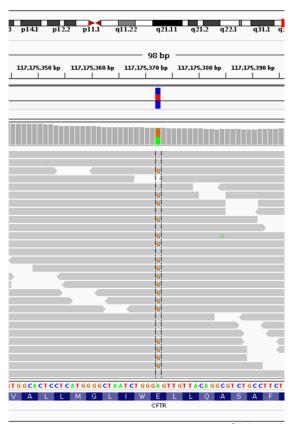
### Bayes theorem

Notice that the total probability  $P(\mathcal{D})$  is the same in each of the denominators and that it is the sum of the three numerators. We can organize all of this very neatly in a Bayesian update table:

		Bayes		
hypothesis	prior	likelihood	numerator	posterior
$\mathcal{H}$	$P(\mathcal{H})$	$P(\mathcal{D} \mathcal{H})$	$P(\mathcal{D} \mathcal{H})P(\mathcal{H})$	$P(\mathcal{H} \mathcal{D})$
A	0.4	0.5	0.2	0.3226
B	0.4	0.6	0.24	0.3871
C	0.2	0.9	0.18	0.2903
total	1		0.62	1 1

If the question is changed to decide the type of a coin, then you will compute probabilities of all types and choose the type corresponding to a maximum probability.

## Bayesian SNP calling



$$P(SNP|Data) = P(Data|SNP) * P(SNP)$$
  
 $P(Data)$ 

☐ At each locus along the reference genome, one SNP site is called if there are a sufficient number of high-quality nucleotides to indicate a difference between the reference genome and the sample genome.

## Bayesian SNP calling

- ☐ Expected Polymorphism Rate
- □ Expected Allele frequency at a potential site (i.e. 0.5) and then use the Hardy–Weinberg equilibrium (HWE).
- ☐ 1 polymorphic in 700 bp human, 1 in 120 for drosophila.
- □ Use dbSNP prior probabilities. For example, if a G/T polymorphism is reported in dbSNP, the prior probabilities are set to be 0.454 for each of the genotypes GG and TT, 0.0909 for GT and less than 10<sup>-4</sup> for all other genotypes.
- ☐ If allele frequencies are known, genotype probabilities can then be calculated using the Hardy–Weinberg equilibrium (HWE) assumption or other assumptions that relate allele frequencies to genotype frequencies.
- ☐ Transition Ti is more frequent than Transversion Tv.
- □One can assign a polymorphic rate P<sub>polymorphic</sub> to {AC,AG,AT,CG,CT,GT}, and (1- P<sub>polymorphic</sub>)/4 to another non-polymorphic permutations {AA,CC,TT,GG}.

### **Expected Prior Polymorphism rate**

Journal List > Genome Res > v.19(6); 2009 Jun > PMC2694485



Genome Res. 2009 Jun; 19(6): 1124-1132.

doi: 10.1101/gr.088013.108

PMCID: PMC2694485

PMID: 19420381

#### SNP detection for massively parallel whole-genome resequencing

Ruiqiang Li, 1,2,3 Yingrui Li, 1,3 Xiaodong Fang, 1 Huanming Yang, 1 Jian Wang, 1 Karsten Kristiansen, 1,2 and Jun Wang 1,2,4

► Author information ► Article notes ► Copyright and License information Disclaimer

Callolla

### **Expected Prior Polymorphism rate**

### Prior probability of genotypes

- Example: Assuming
  - heterozygous SNP rate 0.001, homozygous SNP rate 0.0005
  - Reference allele: G
  - Transition/transversion ratio 2

	Α	С	G	Т
A C G T	$3.33 \times 10^{-4}$	$1.11 \times 10^{-7} \\ 8.33 \times 10^{-5}$	$6.67 \times 10^{-4} \\ 1.67 \times 10^{-4} \\ 0.9985$	$1.11 \times 10^{-7}$ $2.78 \times 10^{-8}$ $1.67 \times 10^{-4}$ $8.33 \times 10^{-5}$

### **Expected Prior Polymorphism rate**

### Prior probability of genotypes

Other information that can be used in setting priors:

- Use dbSNP prior probability
- Use different polymorphism rate for different genomic regions
- Consider different Ti/Tv rate for exonic regions

An example of prior probability for a dbSNP G/T site used in Li et al (2009)

	Α	С	G	Т
Α	4.55*10 <sup>-7</sup>	9.11*10-8	9.1*10-5	9.1*10-5
С		4.55*10 <sup>-7</sup>	9.1*10-5	9.1*10-5
G			.454	.0909
Т				.454

**Table 35.1:** Site Types for a diploid organism with example probabilities.

Site Type	Prior probability
A/A	0.2475
A/C	0.001
A/G	0.001
A/T	0.001
T/C	0.001
T/G	0.001
T/T	0.2475
G/C	0.001
C/C	0.2475
G/G	0.2475
G/-	0.001
A/-	0.001
C/-	0.001
T/-	0.001

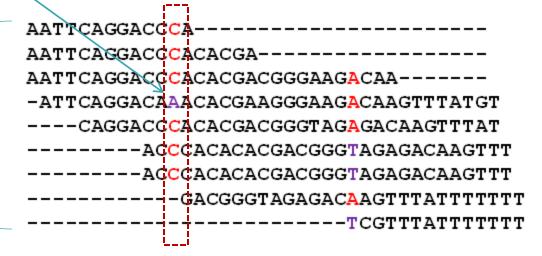
# Bayesian SNP calling

- ☐ Reads that cover a variant calling position with a minimum mapping quality threshold.
- ☐ Quality score of the bases that cover the variant calling position.
- ☐ Reads count that cover a particular variant position.

May be rare variant and not covered by the most of reads.

## Notes

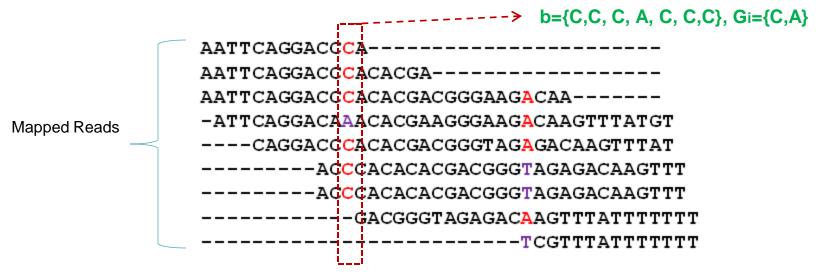
Mapped Reads



Reference Sequence AATTCAGGACCAACACGACGGGAAGATTCATGTACTTTT

Various Genotypes available at this locus { CC, AC, AA}

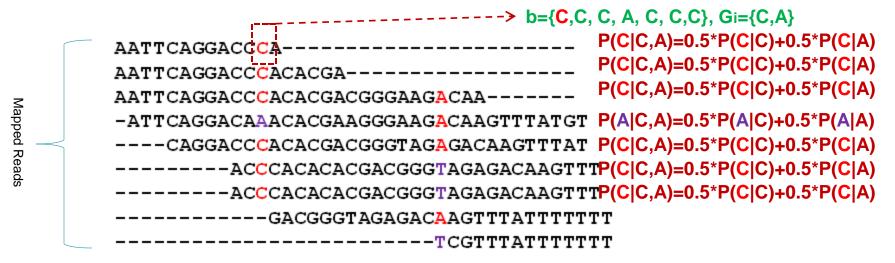
All Genotypes available at this locus { AA, AC, AG, AT, CC, CT, CG, GG, GT, TT}



Reference Sequence AATTCAGGACCAACACGACGGGAAGATTCATGTACTTTT

$$p(D|G_i) = \prod_b p(b|G_i)$$

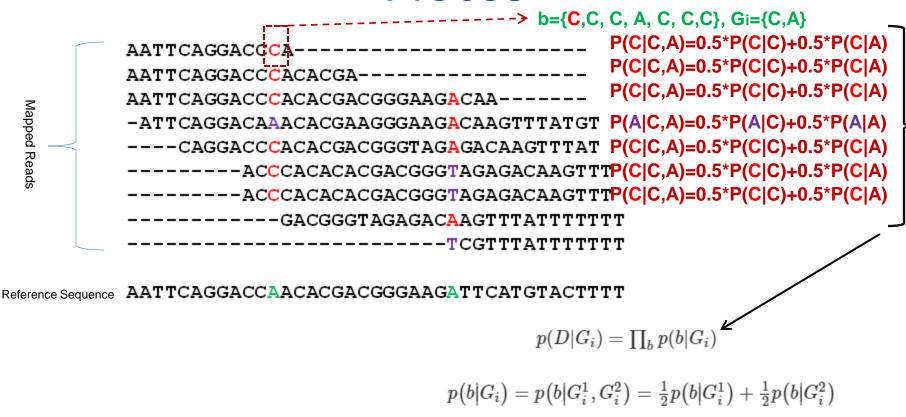
$$p(b|G_i) = p(b|G_i^1, G_i^2) = \frac{1}{2}p(b|G_i^1) + \frac{1}{2}p(b|G_i^2)$$

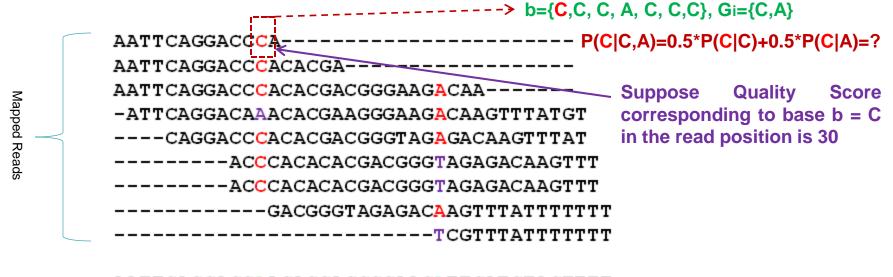


Reference Sequence AATTCAGGACCAACACGACGGGAAGATTCATGTACTTTT

$$p(D|G_i) = \prod_b p(b|G_i)$$

$$p(b|G_i) = p(b|G_i^1, G_i^2) = \frac{1}{2}p(b|G_i^1) + \frac{1}{2}p(b|G_i^2)$$





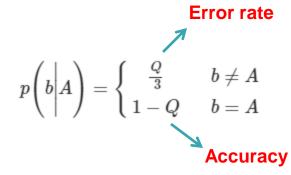
Reference Sequence

AATTCAGGACCAACACGACGGGAAGATTCATGTACTTTT

$$pigg(bigg|Aigg) = \left\{egin{array}{cc} rac{Q}{3} & b 
eq A \ 1-Q & b = A \end{array}
ight.$$

#### Suppose Quality Score corresponding to base b in the read position C is 30

Phred Quality Score	Error	Accuracy (1 - Error)
10	1/10 = 10%	90%
20	1/100 = 196	99%
30	1/1000 = 0.196	99.9%
40	1/10000 = 0.01%	99.99%
50	1/100000 = 0.00196	99.999%
60	1/1000000 = 0.000196	99.9999%



$$P(C|C,A)=0.5*P(C|C)+0.5*P(C|A)=?$$

Q=30, error rate = 0.001/3=0.00033 Q=30, accuracy=0.999

P(C|C)=0.999 P(C|A)=0.00033

Note: another model just use 0.001 for any bases that are different.

#### Suppose Quality Score corresponding to base b in the read position C is 30

Phred Quality Score	Error	Accuracy (1 - Error)
10	1/10 = 1096	90%
20	1/100 = 196	99%
30	1/1000 = 0.1%	99.9%
40	1/10000 = 0.01%	99.99%
50	1/100000 = 0.001%	99.999%
60	1/1000000 = 0.000196	99.9999%

$$pigg(bigg|Aigg) = egin{cases} rac{Q}{3} & b 
eq A \ 1-Q & b = A \end{cases}$$
 Accuracy

#### P(C|C,A)=0.5\*P(C|C)+0.5\*P(C|A)=?

Note: You can incorporate different information in the model such as Mapping quality of the reads, allele frequency, polymorphic rate, reads count, etc.



Reference Sequence AATTCAGGACCAACACGACGGGAAGATTCATGTACTTTT

All Genotypes available at this locus { AA, AC, AG, AT, CC, CT, CG, GG, GT, TT}

P(AC| data) = .99 P(AA| data) = .00000001 P(TT| data) = .0000000001 P(CT|data) = 10<sup>-75</sup> etc

Choose genotype with highest probability





This site is a heterozygous not as we stated before as homozygous to a non reference allele



### **Bioinformatics**

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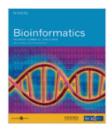
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Volume 34, Issue 12 15 June 2018

**Article Contents** 

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# Progressive approach for SNP calling and haplotype assembly using single molecular sequencing data •

Fei Guo, Dan Wang, Lusheng Wang 🔀

Bioinformatics, Volume 34, Issue 12, 15 June 2018, Pages 2012-2018,

https://doi.org/10.1093/bioinformatics/bty059

Published: 19 February 2018 Article history ▼







Abstract

### PolyBayes: the first statistically rigorous variant detection tool.

letter ≈ © 1999 Nature America Inc. · http://genetics.nature.com

A general approach to single-nucleotide polymorphism discovery

Gabor T. Marth<sup>1</sup>, Ian Korf<sup>1</sup>, Mark D. Yandell<sup>1</sup>, Raymond T. Yeh<sup>1</sup>, Zhijie Gu<sup>2</sup>, Hamideh Zakeri<sup>2</sup>, Nathan O. Stitziel<sup>1</sup>, LaDeana Hillier<sup>1</sup>, Pui-Yan Kwok<sup>2</sup> & Warren R. Gish<sup>1</sup>

This Bayesian statistical framework has been adopted by other modern SNP/INDEL callers such as FreeBayes, GATK, and samtools

## FreeBayes

### Haplotype-based variant detection from short-read sequencing

Erik Garrison and Gabor Marth

July 24, 2012

#### Abstract

The direct detection of haplotypes from short-read DNA sequencing data requires changes to existing small-variant detection methods. Here, we develop a Bayesian statistical framework which is capable of modeling multiallelic loci in sets of individuals with non-uniform copy number. We then describe our implementation of this framework in a haplotype-based variant detector, FreeBayes.

https://arxiv.org/pdf/1207.3907.pdf https://github.com/ekg/freebayes

## **GATK:** Genome Analysis Toolkit

**NATURE GENETICS | TECHNICAL REPORT** 





日本語要約

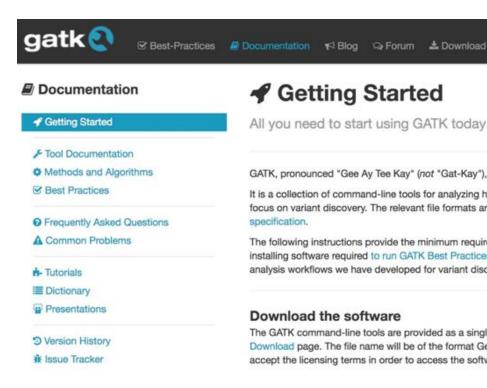
### A framework for variation discovery and genotyping using next-generation DNA sequencing data

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## **GATK:** Genome Analysis Toolkit



### Getting Started

All you need to start using GATK today

GATK, pronounced "Gee Ay Tee Kay" (not "Gat-Kay"), stands for GenomeAnalysisToolkit.

It is a collection of command-line tools for analyzing high-throughput sequencing (HTS) data in formats such as SAM/BAM/CRAM and VCF, with a focus on variant discovery. The relevant file formats are defined in the hts-specs repository; see especially the SAM specification and the VCF specification.

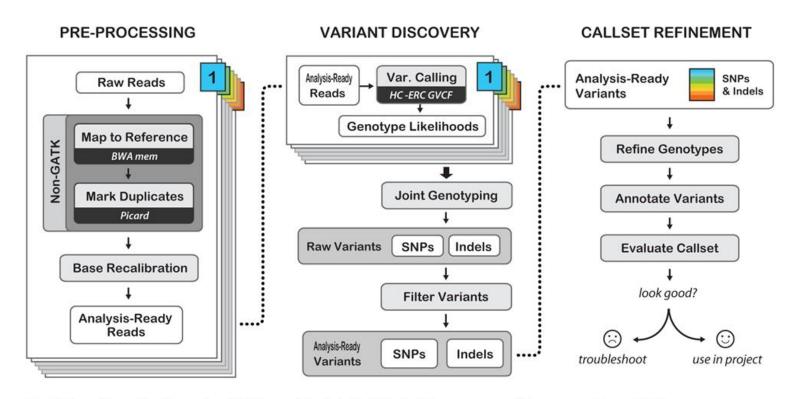
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The following instructions provide the minimum requirements for getting started with GATK. Additional instructions are provided elsewhere for installing software required to run GATK Best Practices workflows and to attend a hands-on GATK workshop. For information about the complete analysis workflows we have developed for variant discovery, see the Best Practices documentation.

#### Download the software

The GATK command-line tools are provided as a single executable jar file. You can download a bzipped package containing the jar file from the Download page. The file name will be of the format GenomeAnalysisTK-x.y-z.tar.bz2. You will need to register for a free account on the forum and accept the licensing terms in order to access the software download.

### **GATK** workflow



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