

# MolAICal

A drug design software combined by artificial intelligence and  
classical programming

## Manual



**MolAICal**

**V1.0**

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# 1. General information of MolAICal

## 1.1. Introduction

Computer-aided drug design (CADD) employs the computational strategies to screen, develop, modify and analyze drugs and similar biologically active compounds<sup>1</sup>. With the development of technology, artificial intelligence methods such as deep learning, etc are permeated into the CADD research fields. The CADD and deep learning have been widely used in *de novo* drug design, bioactivity prediction, biological image analysis and synthesis prediction successfully<sup>2</sup>. The CADD combined with deep learning can save capital and improve the speed of drug discovery.

Here, the MolAICal soft package is introduced which can be used for drug design by deep learning model, classical algorithms such as genetic algorithm, etc. The detail functions of MolAICal contain as below:

- 1 The *de novo* drug design: MolAICal can perform fragment growth on the pocket of the receptor by using the assigned molecular library or the molecular library generated from deep learning models.
- 2 The drug virtual screening: MolAICal combined with Autodock Vina can perform drug virtual screening based on the molecular database generated from deep learning models.
- 3 The useful tools for drug discovery
  - 1) Channel radii measure for channel receptors such as ion channel proteins.
  - 2) Potential of mean force for conformational change measurement of receptor-induced by ligands, etc.
  - 3) Quantitative structure-activity relationship (QSAR) for bioactivity prediction of ligands
  - 4) Lipinski's rule of five<sup>3</sup> for the calculation of drug-like properties.
  - 5) Pan-assay interference compounds (PAINS)<sup>4</sup> filteretc

MolAICal is freely for education, academic and other non-profit purposes. If you want to use MolAICal for any commercial purpose, you should contact with licensors (Email: molaical@yeah.net).

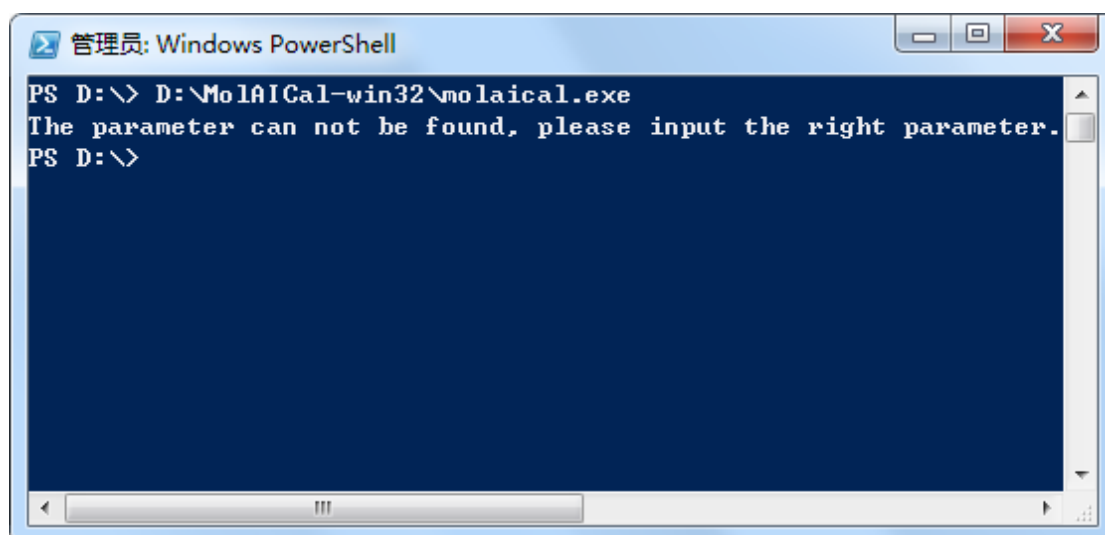
## 1.2. Installation

### Download

MolAICal: <https://molaical.github.io>

### For Window:

MolAICal need not be installed, just decompress the MolAICal file. MolAICal is a command-line tool. You can use the absolute path of MolAICal on DOS or Powershell of Windows. For example: if MolAICal is placed on the D: disk, then you can run as blow:



```
管理员: Windows PowerShell
PS D:\> D:\MolAICal-win32\molaical.exe
The parameter can not be found, please input the right parameter.
PS D:\>
```

**For Linux:**

1. tar -xvzf MolAICal-linux64-xxx.tar.gz

Notice: Please replace the corrected characters in MolAICal-linux64-xxx.tar.gz according to your downloaded MolAICal.

2. cd MolAICal-linux64-xxx

3. chmod +x install.sh

4. ./install.sh

**Notice:** due to the system compatibility problem in Linux operating system, the “AIGenMols” of MolAICal may not run in your system, you can package your trained model by PyInstaller (see the first example in “Specialized topics of MolAICal” of MolAICal tutorials), or choose win64 version of MolAICal to generate your desired number of molecules because the “AIGenMols” of MolAICal have good compatibility in Windows operating system. The memory of 32-bit MolAICal (win32) is limited up to ~2GB. The memory of 64-bit MolAICal (win64, linux64) is unlimited. We recommend 64-bit version of MolAICal.

More detail tutorials, please go to MolAICal website: <https://molaical.github.io>

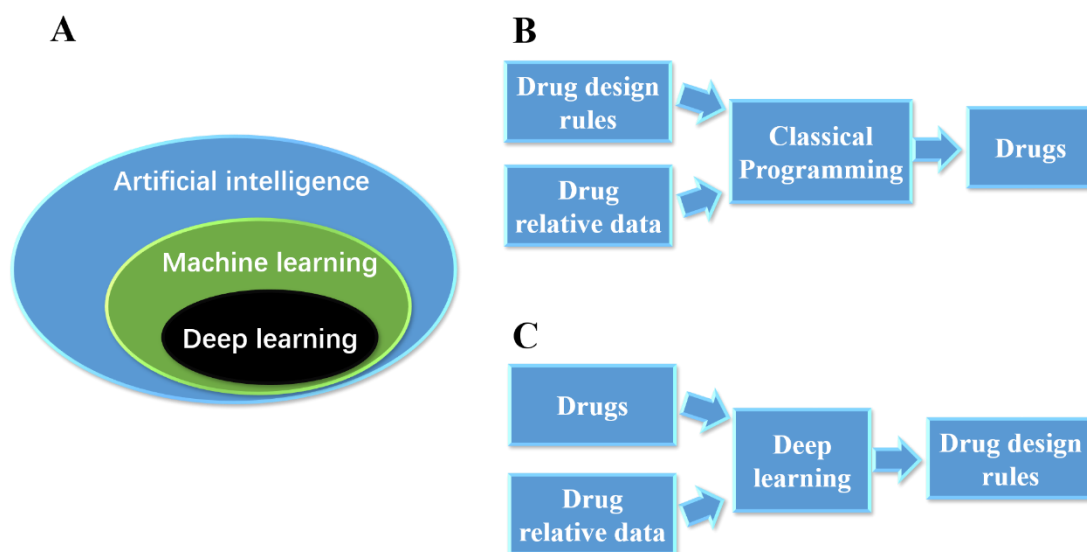
## 2. *De novo* drug design by MolAICal and DL

### 2.1. Theory

The rational drug design is the purpose to find new medications based on the knowledge of the biological targets such as protein, DNA, RNA, etc. Computer-aided drug design is an approach rational drug design to discover, design, optimize, and analyze ligands and similar biologically active drugs in recent years. The fragment-based drug discovery (FBDD) is a *de novo* drug design method to find the lead compounds. The fragments are small organic chips that are with small size

and low molecular weight. The small chemical fragments may bind to the biological target weakly, it can produce a lead with a higher affinity via growing fragments based on the appointed previous anchor fragments in the receptor pocket.

The deep learning which employs the multiple layers to progressively get higher-level features from the raw input data belongs to machine learning and artificial intelligence (see Figure 2.1A). Deep learning has been successfully used in drug discovery<sup>2</sup> and medicine<sup>5</sup>. It can train the molecular generative model which can be used to generate 1D sequence or 2D structural molecules by generative adversarial network (GAN)<sup>6,7</sup>. Traditionally, classical programming produces new drugs through inputting drug design rules and drug relative data such as molecular weight, logP, etc (see Figure 2.1B). The input drug data and drug design algorithm play an important role in the new drug discovery. For the deep learning method, the target drugs and drug relative data are given. The deep learning is responsible for finding the drug design rules via training the deep neural network (see Figure 2.1C). Although the deep learning can trained drug design rules for new similar properties drugs generation, it still need the effective way to generative 3D structural ligands in the 3D receptor pocket. The classical programming and deep learning have their own merit on drug discovery. Here, MolAICal combined the merits of *de novo* drug design and deep learning model for drug design. The molecular generative model which train from the fragments of FDA approved drugs by using GAN, can produce the fragments for ligands growth in the receptor pocket. Besides, MolAICal can perform classical *de novo* drugs which is based on the given small fragments.



**Figure 2.1** The deep learning and algorithm for drug design

## 2.2. Parameters in configure file

These parameters on this part are contained in the configure file which is the input file for *de novo* drug design. The examples of input configure files are shown in Appendix 1 and 2.

➤ **centerPoints** < center position of box >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** The center coordinates of the appointed box.

➤ **boxLengthXYZ** < box length >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** It is box lengths which are sequentially lengths along X, Y, Z axis.

➤ **receptorPDB** < path of receptor >

**Acceptable Values:** String

**Default Value:** null

**Description:** The path of PDB format receptor in your operation system.

➤ **growMethod** < grow method >

**Acceptable Values:** fixFrag, randomFrag

**Default Value:** null

**Description:** The ‘fixFrag’ means the coordinates of the initial molecular fragment are fixed, and this molecular fragment may be extracted from the ligand in the crystal receptor-ligand complex. The “randomFrag” means the positions of the initial fragment are not fixed, and the initial fragment should be assigned as SMILES format. The initial position should also be appointed. If this part is on, it must set the “startAtomPosition” and “startSmiFrag”.

➤ **startFragFile** < path of initial fragment >

**Acceptable Values:** String

**Default Value:** null

**Description:** The path of the initial fragment in your operation system. The initial fragment is Sybyl mol2 format.

➤ **startSmiFrag** < Initial SMILES fragment >

**Acceptable Values:** String

**Default Value:** null

**Description:** If the growMethod is “randomFrag”, this parameter should be set. The value is SMILES such as C

➤ **startAtomPosition** < path of chosen atom file in receptor >

**Acceptable Values:** String

**Default Value:** null

**Description:** If the growMethod is “randomFrag”, this parameter should be set. To set the position of initial SMILES fragment, it should appoint the atom of receptor as the coordinate reference. The appointed atom can be extracted from receptor structure by graphical tools such as pymol, vmd, UCSF Chimera, etc. The atom file must be standard PDB format.

➤ **numCycleGen** < number of grow cycle >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** It is the number of grow cycle. Each cycle grow is based on the previous cycle.

➤ **selTopMols** < number of parent >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** This part is designed by genetic algorithm. The “selTopMols” represents the number of parents.

➤ **selTopRootPercent** < percentage of parent selection >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** It is the percentage of parent selection. Multiply “selTopMols” by “selTopRootPercent” is the selected number.

➤ **selElitePercent** < percentage of elitist selection >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** It is the percentage of elitist selection.

➤ **maximumPOP** < maximum of populations >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** It is the maximum of population.

➤ **libStyle** < Selection way of fragments library >

**Acceptable Values:** mol2, SMILES, AIFrag

**Default Value:** null

**Description:** MolAICal supports three ways for ligands growth. The “mol2” means MolAICal uses its own mol2 format fragments library. The “SMILES” means MolAICal uses its own SMILES sequence library. The “AIFrag” means MolAICal call deep learning model named “AIGenMols” for generation new fragments.

➤ **genAIWay** < selected model for fragment generation >

**Acceptable Values:** Fdafrag

**Default Value:** null

**Description:** If you set “libStyle” to AIFrag. It should set the value of “genAIWay” which can be set to “Fdafrag”.

➤ **genAINumber** < number of generated fragments >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** If you set “libStyle” to AIFrag. It should set the value of “genAINumber”. It can be



an arbitrary integer value. For instance, if you set it to 81, it means deep learning model named “AIGenMols” will generate 81 different fragments.

➤ **perturbeSearch** < whether perform perturbation searching >

**Acceptable Values:** on or off

**Default Value:** null

**Description:** If “on”, the procedure will grow fragment by perturbation searching around bonds. If “off”, the procedure will grow fragment along the hydrogen atom direction.

➤ **genAlgorithm** < searching algorithm for fragment generation anchoring on one point >

**Acceptable Values:** random, fibonacci

**Default Value:** null

**Description:** The “random” means the fragments chosen random points for perturbation growth. The “fibonacci” means the fragments choose the spherical Fibonacci points for perturbation growth.

➤ **ranGenOptNum** < the number times of fragment generation surrounded on anchoring point >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** It represents the number of fragment generation surrounded on anchoring points. The big number will expend more computational resources, but the ligand grow will be more precise.

➤ **atomMutation** < mutation on one atom >

**Acceptable Values:** String

**Default Value:** null

**Description:** The value is “on” or “off”. The “on” means atoms “C”, “N” and “O” will be mutated during the process of ligand grow, and vice versa.

➤ **mutationPercent** < percentage of mutation ratio >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** If the “atomMutation” is set, the “mutationPercent” should be appointed a value.

➤ **randomCrossMutation** < crossover and mutation >

**Acceptable Values:** String

**Default Value:** null

**Description:** This parameter is involved in crossover and mutation. It can be value of “on” or “off”. “On” means the crossover and mutation are carried out during the process of ligand growth, and vice versa. MolAICal regards the ligand as the chromosome. The fragments connected by rotated bonds regards as “A”, “T”, “G”, “C”, etc.

➤ **randomCrossRatio** < ratio for cross operation >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** If the “randomCrossMutation” is set, the “randomCrossRatio” should be appointed. It

is the ratio of crossover operation in the generated population. This mutation unit is one fragment.

➤ **randomMutationRatio** < ratio of mutation >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** If the “randomCrossMutation” is set, the “randomMutationRatio” should be appointed. It is the ratio of mutation. This mutation unit is one fragment.

➤ **randomCrossCompareNum** < number of selected molecules for crossover and mutation >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** If the “randomCrossMutation” is set, the “randomCrossCompareNum” should be appointed. It represents the number of selected molecules from the left molecular population. For example, if the value is 2, it means the current ligand will crossover and mutate with 2 ligands of the left random remainder populations.

➤ **randomCrossRange** < range of crossover and mutation >

**Acceptable Values:** Discrete integer

**Default Value:** null

**Description:** If the “randomCrossMutation” is set, the “randomCrossRange” should be appointed. It corresponds to “numCycleGen”. The discrete integer corresponds to the cycle of “numCycleGen” which represents this cycle performs crossover and mutation.

➤ **writeResultParallel** < parallel writing mode >

**Acceptable Values:** String

**Default Value:** null

**Description:** If “on”, it means the temporal file is saved as the parallel mode, and vice versa.

➤ **unwantedFrag** < filtering unwanted fragments >

**Acceptable Values:** String

**Default Value:** null

**Description:** If “on”, it will filter the generated molecules contained unwanted fragments, and vice versa.

➤ **PAINS** < filtering Pan-assay interference compounds >

**Acceptable Values:** String

**Default Value:** null

**Description:** If “on”, it will filter the generated molecules contained Pan-assay interference compounds (PAINS) fragments, and vice versa.

➤ **growFilter** < filtering wrong position molecules >

**Acceptable Values:** String

**Default Value:** null

**Description:** If “on”, it will filter the generated molecules which are overlap and out of box

fragments, and vice versa.

➤ **growFilterRatio** < cutoff of overlap molecule >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** This ratio is used to filter overlap molecules during the storing process of the temporal file.

➤ **resultsFilterRatio** < cutoff of overlap molecule >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** This ratio is used to filter overlap molecules during the storing process of final results ligands.

➤ **syntheticAccessibility** < synthetic accessibility >

**Acceptable Values:** String

**Default Value:** null

**Description:** If “on”, it will filter the unreachable criterion ligands according to the set value of synthetic accessibility, and vice versa.

➤ **synAccessibilityValue** < cutoff of synthetic accessibility >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** If the “syntheticAccessibility” is set, the “synAccessibilityValue” should be appointed. Synthetic accessibility (SA) is a cutoff ranging from 0 to 100 in which value 100 is maximal synthetic accessibility, it means this molecule is most easily synthesizable.

➤ **MolsimilarPercent** < filtering the similar ligands >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** This parameter is used to filter similar ligands and add new novel ligands into the parents’ populations according to the cutoff.

➤ **RulesOfFive** < Lipinski's rule of five >

**Acceptable Values:** String

**Default Value:** null

**Description:** If “on”, it will filter the unreachable criterion ligands according to the Lipinski's rule of five, and vice versa. The Lipinski's rule of five is changed with the passage of time<sup>8</sup>, please set suitable value according to the research reports or your customization.

➤ **xlogPvalue** < XLogP value >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** It is the XLogP value. If the “RulesOfFive” is set, the “xlogPvalue” should be

appointed.

➤ **acceptors** < number of hydrogen bond acceptors >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** It is the number of hydrogen bond acceptors. If the “RulesOfFive” is set, the “acceptors” should be appointed.

➤ **donors** < number of hydrogen bond donors >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** It is the number of hydrogen bond donors. If the “RulesOfFive” is set, the “donors” should be appointed.

➤ **mwvalue** < molecular weight >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** It is the molecular weight. If the “RulesOfFive” is set, the “mwvalue” should be appointed.

➤ **rotatablebonds** < number of rotatable bonds >

**Acceptable Values:** integer

**Default Value:** null

**Description:** It is the number of rotatable bonds. If the “RulesOfFive” is set, the “rotatablebonds” should be appointed.

➤ **pcClusterNumber** < number of clusters >

**Acceptable Values:** integer

**Default Value:** null

**Description:** Performing appointed number of clusters on the final generated ligands by K-means algorithm.

➤ **adjustOutSimPer** < fingerprint similarity >

**Acceptable Values:** decimal

**Default Value:** 0.999999

**Description:** This parameter filters similar ligands according to the values of fingerprint similarity when performing the cluster on the final generated ligands. The 0.999999 means it filters the same molecule approximatively.

➤ **selMolMaxWeight** < maximum molecular weight >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** This parameter filters the final generated ligands which is greater than maximum molecular weight.

➤ **selMolMinWeight** < minimum molecular weight >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** This parameter filters the final generated ligands which are smaller than minimum molecular weight.

➤ **selBindingScore** < cutoff of binding score >

**Acceptable Values:** decimal

**Default Value:** null

**Description:** This parameter filters the final generated ligands which have higher binding score than the set binding score.

➤ **obabelPath** < path of obabel >

**Acceptable Values:** String

**Default Value:** path of MolAICal

**Description:** It represents the path of obabel of Open Babel. The default value is the path of MolAICal, so you can omit it if you have the full package of MolAICal.

➤ **pcClusterdFile** < path of cluster file >

**Acceptable Values:** String

**Default Value:** current directory

**Description:** It represents the path of cluster file named "PC.dat". The default value is the current directory, so you can omit it.

➤ **moloutputdir** < storing directory of final ligands >

**Acceptable Values:** String

**Default Value:** current directory

**Description:** It represents the storing directory of the final ligands. The default value is the current directory named "results", so you can omit it.

➤ **coreNum** < number of cores for parallel computing >

**Acceptable Values:** Integer

**Default Value:** null

**Description:** It represents the number of cores for parallel computing. You can set it to an arbitrary value, but MolAICal can use the limited core. For example, MolAICal can use maximally ~15 cores for loading 67 molecular fragments at one job.

## 2.3. The commands for fragments grow

The command parameters of MolAICal for *de novo* drug design are as below:

**-denovo:** means perform job of *de novo* drug design. Value is "grow".

- i: input path of configure file.
- g: whether calculate grids, value is “on” or “off”, default value is “on”.
- l: whether perform fragment grow, value is “on” or “off”, default value is “on”.
- o: whether output results, value is “on” or “off”, default value is “on”.

Here, the example commands for fragment grow are listed:

1) If you want to run full process of fragment growth, you can use the command:

`molaical.exe -denovo grow -i InputParFile.dat`

2) This command omits the grids generation, only contains fragments grow and output results.

`molaical.exe -denovo grow -g off -i InputParFile.dat`

3) This command is output results only.

`molaical.exe -denovo grow -g off -l off -i InputParFile.dat`

4) This command is no results output.

`molaical.exe -denovo grow -g off -l off -o off -i InputParFile.dat`

5) This command generates grids only

`molaical.exe -denovo grow -g on -l off -o off -i InputParFile.dat`

## 3. Virtual screening by deep learning model

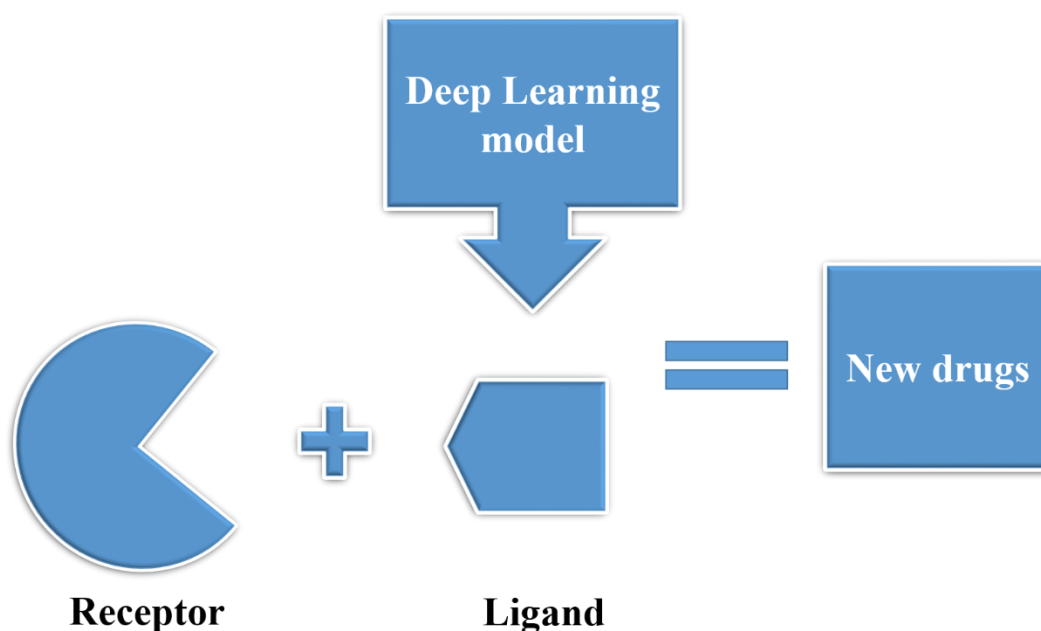
### 3.1. Drug virtual screening by MolAICal, DL model and Autodock Vina

#### 3.1.1. Theory

Virtual screening (VS) which is different from *de novo* drug design, is a computational method used in drug discovery to find potential small molecules that most likely bind to the drug target, typically protein receptor or DNA, etc. The VS contains ligand-based and structure-based virtual screening<sup>9</sup>. Especially, the structure-based virtual screening involves molecular docking which searches good binding poses of the ligands into the protein target followed by applying a scoring function to estimate the affinity ability between ligands and receptors. Molecular docking is likely the theory of the Lock and Key Model (see Figure 3.1.1). The Autodock Vina<sup>10</sup> is an excellent open source molecular docking software that has been studied and applied to the fields of drug discovery. The score function of Autodock Vina is also studied and optimized in used to estimate the binding ability between the ligands and receptors. The Vinardo score<sup>11</sup> is a variant of score function of Autodock

Vina which can improve the prediction of binding ability between ligands and receptors.

The deep learning which employs the multiple layers to progressively get higher-level features from the raw input data belongs to machine learning and artificial intelligence. Deep learning has been successfully used in drug discovery<sup>2</sup> and medicine<sup>5</sup>. It can train the molecular generative model which can be used to generate 1D sequence or 2D structural molecules by generative adversarial network (GAN)<sup>6,7</sup>. However, rational drug design needs to study the 3D structural ligand in the receptor pocket. The *de novo* drug design provides a way to design 3D drugs in the receptor pocket by deep learning model and classical programming algorithm such as genetic algorithm, etc. Besides, the molecular docking supplies another way to take advantage of the deep learning model and classical programming. The strategies for virtual screening is that MolAICal invokes the deep learning model to generate the appointed number of drugs, and then call Autodock Vina to perform the molecular docking to evaluate the binding ability between ligand and receptor (see Figure 3.1.1). Besides, MolAICal also can utilize the appointed small molecular database for drug virtual screening.



**Figure 3.1.1** Virtual screening based on molecular docking and deep learning model

### 3.1.2. Introduction of parameters of Vina configure file

This part introduces the parameters of drug virtual screening by MolAICal, deep learning (DL) model (Note: only 64bit MolAICal can perform the AI and molecular docking completely. 32bit MolAICal can only perform molecular docking. Please run this process on 64 bit operating system.). Before you run this drug virtual screening, please prepare the necessary Vina configure files named "conf.txt". The pdbqt format files of protein must be together with "conf.txt". The content "conf.txt" is like as below:

```
receptor = protein.pdbqt
```

```
cpu = 1  
center_x = -30.011  
center_y = 1.665  
center_z = -36.581
```

```
size_x = 30  
size_y = 30  
size_z = 30
```

```
num_modes = 3
```

Where the “receptor” represents protein file, “cpu” means number of core for running, the “center\_x”, “center\_y”, “center\_z” are the x, y, z coordinates of box center you set, the “size\_x”, “size\_y”, “size\_z” are lengths along x, y, z axis of box you set, the “num\_modes” is the number of generated conformation by Autodock Vina.

### 3.1.3. Command introduction for AI virtual screening

The command parameters of MolAICal for AI drug virtual screening are as below:

- dock:** value is AI, which means performing drug virtual screening by AI.
- s:** selection of deep learning model. Values are “ZINCMol”, “FDAMol”, “FDAFrag”
- n:** number of generated molecules by deep learning model.
- nf:** number of generated molecules in one folder. It avoids very huge of molecules in one folder.
- p:** path of obabel.exe. The default value is in the same directory of MolAICal. So you can omit it if you have full MolAICal soft package.
- nc:** number of CPU cores for running drug virtual screening
- g:** switch AI molecular generations. “on” means generation. “off” means no generation.
- b:** switch molecular format conversion. “on” means conversion. “off” means no conversion.
- v:** switch virtual screening (VS) based on docking. “on” means VS. “off” means no VS.
- m:** molecular format for VS, etc.

Here, the example commands for AI drug virtual screenings are listed:

1) The simple command, the omitted parameters use the default values.  
molaical.exe -dock AI -s FDAMol -n 30 -nf 10 -nc 3

2) Run full drug virtual screening process by MolAICal, deep learning model, and Autodock Vina.  
molaical.exe -dock AI -s FDAMol -n 30 -nf 10 -nc 3 -g on -b on -v on

3) Only run molecular artificial intelligence (AI) generation.



molaical.exe -dock AI -s FDMol -n 30 -nf 10 -nc 3 -g on -b off -v off

4) Only run conversion of molecular formats only

molaical.exe -dock AI -s FDMol -n 30 -nf 10 -nc 3 -g off -b on -v off

5) Only run Vina docking. So you can customize your own AI SMILES file, but this file must be named "tmpGen.dat".

molaical.exe -dock AI -s FDMol -n 30 -nf 10 -nc 3 -g off -b off -v on

6) Command for virtual screening by calling deep learning mode

molaical.exe -dock AI -s FDMol -n 30 -p "C:/Program Files (x86)/OpenBabel-2.4.1/obabel.exe" -nf 10 -nc 3 -g on -b on -v on

7) Command contained full parameters.

molaical.exe -dock AI -s ZINCMol -n 30 -p "C:/Program Files (x86)/OpenBabel-2.4.1/obabel.exe" -nf 10 -nc 3 -g on -b on -v on -m pdbqt

8) This command have no drug virtual screening task.

molaical.exe -dock AI -s FDMol -n 30 -p "C:/Program Files (x86)/OpenBabel-2.4.1/obabel.exe" -nf 10 -nc 3 -g on -b on -v off -m pdbqt

9) Run AI generation and conversion of molecular formats, so you can convert the results to the formats you want.

molaical.exe -dock AI -s FDMol -n 30 -nf 10 -nc 3 -g on -b on -v off -m mol2

### 3.1.4. Command introduction for virtual screening

This command is for virtual screening (VS) based on the known database simply via Autodock Vina. The command parameters of MolaICal for drug virtual screening are as below:

**-dock:** value is vs, means performing drug virtual screening.

**-i:** input directory which contains the molecules files.

**-k:** the files contain the keyword for VS. The default value is ".mol2"

**-p:** path of obabel.exe. The default value is in the same directory of MolaICal. So you can omit it if you have full MolaICal soft package.

**-nc:** number of CPU cores for running drug virtual screening

**-b:** switch molecular format conversion. "on" means conversion. "off" means no conversion.

**-v:** switch virtual screening (VS) based on docking. "on" means VS. "off" means no VS.

**-m:** files with input molecular format for VS in the appointed directory.

Before run VS process, you should split files into directories, for instance:

molaical.exe -tool split -w split -n 1 -v true -i "D:\aa.mol2" -o "D:\split"

Here, the example commands for drug virtual screenings are listed:

1) The simple command, the omitted parameters use the default values.

```
molaical.exe -dock vs -i D:\split
```

2) The simple command, the docked molecular format and running CPU cores are appointed.

```
molaical.exe -dock vs -i D:\split -k .mol2 -nc 3
```

3) Run full drug virtual screening process.

```
molaical.exe -dock vs -i D:\split -k .mol2 -p "C:\Program Files\OpenBabel64\obabel.exe" -b on -v on -m mol2 -nc 3
```

4) Run drug virtual screening process without molecular format conversion.

```
molaical.exe -dock vs -i D:\split -k .mol2 -p "C:\Program Files\OpenBabel64\obabel.exe" -b off -v on -m mol2 -nc 3
```

## 3.2. Commands for MM/GBSA using NAMD

Molecular mechanics generalized Born surface area (MM/GBSA)<sup>12,13</sup> is a popular way to estimate the binding free energy between two molecules such as ligand-protein, protein and protein, etc. Here, MM/GBSA is calculated by MolAICal on basis of MD log file of NAMD software. The MM/GBSA is estimated by the equation as below:

$$\Delta G_{\text{bind}} = \Delta H - T\Delta S \approx \Delta E_{\text{MM}} + \Delta G_{\text{solv}} - T\Delta S$$

$$\Delta E_{\text{MM}} = \Delta E_{\text{internal}} + \Delta E_{\text{ele}} + \Delta E_{\text{vdw}}$$

$$\Delta G_{\text{solv}} = \Delta G_{\text{GB}} + \Delta G_{\text{SA}}$$

Where  $\Delta E_{\text{MM}}$ ,  $\Delta G_{\text{solv}}$ , and  $T\Delta S$  represent the gas phase MM energy, solvation free energy (sum of polar contribution  $\Delta G_{\text{GB}}$  and nonpolar contribution  $\Delta G_{\text{SA}}$ ) and conformational entropy, respectively.  $\Delta E_{\text{MM}}$  contains van der Waals energy  $\Delta E_{\text{vdw}}$ , electrostatic  $\Delta E_{\text{ele}}$  and  $\Delta E_{\text{internal}}$  of bond, angle, and dihedral energies. If there are no binding-induced structural changes in the process of molecular dynamics (MD) simulations, the entropy calculation can be omitted. Currently, the MolAICal can evaluate the binding free energy without entropy calculation based on the MD results log of NAMD. The variables in the above equations correspond to Figure 3.2.1.

```

Info: Reading timestep from file.
Info: Updating unit cell from timestep.
Warning: Cell basis vectors should be specified before reading trajectory.
Info: REMOVING COM VELOCITY 0.0946993 0.165474 -0.0546543
TCL: Setting parameter firstTimestep to 0
TCL: Running for 0 steps
Warning: Energy evaluation is expensive. Increase outputEnergies to improve performance.
ETITLE:    TS      BOND      ANGLE      DIHED      IMPRP      ELECT      VDW      BOUNDARY
ENERGY:    0      859.9728    2393.1243    1018.0991    149.1137    -8636.4042    -961.0351    0.0000

Info: Reading timestep from file.
Info: Updating unit cell from timestep.
Warning: Cell basis vectors should be specified before reading trajectory.
Info: REMOVING COM VELOCITY 0.0946993 0.165474 -0.0546543
TCL: Setting parameter firstTimestep to 1
TCL: Running for 0 steps
ENERGY:    1      818.8525    2486.2754    1012.9058    128.6249    -8586.2623    -1031.4833    0.0000

Info: Reading timestep from file.
Info: Updating unit cell from timestep.
Warning: Cell basis vectors should be specified before reading trajectory.
Info: REMOVING COM VELOCITY 0.0946993 0.165474 -0.0546543
TCL: Setting parameter firstTimestep to 2
TCL: Running for 0 steps
ENERGY:    2      840.4857    2434.8220    1058.8810    139.0865    -8577.0594    -1031.3150    0.0000

```

Annotations in the image:  
- **deltaE(internal)** points to the IMPRP column.  
- **deltaE(vdw)** points to the VDW column.  
- **deltaE(ele)+deltaG(sol)** points to the ELECT column.

Figure 3.2.1 The detail information of NAMD running log file

The parameters for calculating binding free energy are as below:

- mmgbsa**: means MM/GBSA calculation
- c**: the MD log of receptor and ligand
- r**: the MD log of receptor
- l**: the MD log of the ligand.

Example command line as below:

```

molaical.exe -mmgbsa -c "D:\workdir\mmgbsa\complex.log" -r "D:\workdir\mmgbsa\protein.log"
-l "D:\workdir\mmgbsa\ligand.log"

```

## 4. Useful tools for drug design

### 4.1. Channel radii measurement

The Metropolis Monte Carlo simulated annealing method<sup>14</sup> is used to measure the channel radii of objects such as nanotube, ion channel protein, etc. The initial can be defined anywhere in the channel, and the explore vector is approximately along the direction of the channel. The calculation of channel radii is realized in MolAICal. The examples of configure files for radii calculation shows in Appendix 3 and 4. The parameters for channel radii calculation are listed as below:

➤ **pdopath** < The path of object with pdb format>

**Acceptable Values:** String

**Default Value:** null

**Description:** It should point out the path of object for radii measurement.

➤ **psfpath** < The path of object with psf format>

**Acceptable Values:** String

**Default Value:** null

**Description:** The path of object with psf format is not necessary if you only perform the simple radii measurement in the channel of object. If you want to measure the radii of object by using CHARMM force field, you should give the path of psf file which corresponds to pdb file.

➤ **cpoint** < The initial point for radii measurement>

**Acceptable Values:** decimal

**Default Value:** null

**Description:** This is the initial point for radii measurement. The values following cpoint are x, y, z coordinates.

➤ **Vector** < The measurement direction>

**Acceptable Values:** decimal

**Default Value:** null

**Description:** This parameter has three values that represent x, y and z directions. For example, 1.00 0.00 0.00 represent measure channel radii along x axis. 0.00 1.00 0.00 represent measure channel radii along y axis. 0.00 0.00 1.00 represent measure channel radii along z axis.

➤ **sample** < The interval of channel radii measurement along appointed direction>

**Acceptable Values:** decimal

**Default Value:** null

**Description:** It is the interval of channel radii measurement along the appointed direction. You can set it according to your measure requirement.

➤ **minprobe** < The minimum explored probe>

**Acceptable Values:** decimal

**Default Value:** Value calculated from the minimum radius

**Description:** It is the minimum explored detector. The MolAICal program will calculate radii of protein, nanotube, etc. by the minimum explored detector.

➤ **conpar** < control constant>

**Acceptable Values:** decimal

**Default Value:** 0.15

**Description:** It is control constant. The detail parameter is shown in equation 4 in this paper<sup>15</sup>. Usually, it uses the default value of 0.15.

➤ **endrad** < cutoff for radii measurement termination>

**Acceptable Values:** decimal

**Default Value:** null

**Description:** It is the cutoff value for radii measurement termination. If the measured radii are larger than the setting cutoff, the program will end.

➤ **selatoms** < select atoms within the range of object>

**Acceptable Values:** decimal

**Default Value:** null

**Description:** For a large system, it needs to calculate a lot of unnecessary atoms. In this case, the selected atoms can save computing time. This value should be larger than the cutoff value of endrad parameter.

➤ **numpt** < number of points in the surface>

**Acceptable Values:** integer

**Default Value:** null

**Description:** The final output contains grid file which can show the grid surface of the channel in the VMD software<sup>16</sup>. This parameter can determine the number of surface points showed in the VMD software.

➤ **surColor** < surface color>

**Acceptable Values:** string

**Default Value:** null

**Description:** It is the surface color. Because the color is shown in the VMD software, the value of surColor can be chosen from the color values of VMD software.

➤ **lineColor** < color of radii measurement routing>

**Acceptable Values:** string

**Default Value:** null

**Description:** The channel radii are measured along the appointed direction. The whole routing shows a curve in the channel. This parameter sets the color of curve. The value of lineColor can be chosen from the color values of VMD software.

➤ **lineWidth** < width of radii measurement routing>

**Acceptable Values:** integer

**Default Value:** null

**Description:** It represents the width of the curve. The “lineColor” is responsible for the color of measured routing, while the “lineWidth” is responsible for the width of measured routing.

➤ **searchNum** < measured number of radii in one plane>

**Acceptable Values:** integer

**Default Value:** null

**Description:** It is a trial number for the radii measurement of the channel in one plane.

➤ **material** < material property of sphere surface >

**Acceptable Values:** integer

**Default Value:** null

**Description:** It represents the material property of the sphere surface which is shown in VMD software. It can be chosen from the material values of VMD software.

The command parameters of MolaICal for channel radii measurement are as below:

**-channel:** means radii measurement function.

**-cpp:** the path of input configure file which contains the parameters for radii calculations

**-fc:** means the calculated way. It contains two ways. One way is simple radii calculation, the other is radii measurement with CHARMM force field which needs PSF file format.

Here, the example commands for channel radii measurement are shown as below:

1) Command for simple radii calculation.

molaical.exe -channel radii -cpp D:/GramicidinA/parameter.dat -fc simple

2) Command for radii calculation with CHARMM force field.

molaical.exe -channel radii -cpp D:/GramicidinA/parameter.dat -fc charmm

3) Command with default parameters. The default calculated way is simple radii calculation.

molaical.exe -channel radii -cpp D:/GramicidinA/parameter.dat

## 4.2. Potential of mean force

The potential of mean force (PMF) can be used to calculate the free energy changes as the function of a coordinate of system. The PMF along the coordinate is computed from the average distribution function (see below equation).

$$\Delta G = -k_B T \ln \rho(x, y)$$

Where T and  $k_B$  is the temperature and Boltzmann constant, respectively. The x and y represent two principal components.

The command parameters of MolaICal for PMF calculation are as below:

**-pmf:** means PMF calculation function

**-i:** path of input data file

**-g:** number of grids for density calculation in a square

**-l:** number of color levels.

**-m:** choose contours from DISLIN. Currently, it contains “conshd” and “contur”

**-b:** is LABELS. It determines which label types will be plotted on an axis. The values can be referred in the DISLIN help

**-x:** is x-axis label

**-y:** is y-axis label

**-t:** is temperature.

**-f:** is the output figure file.

The example commands for PMF are shown as below:

1) Do PMF calculation by using the default parameter

```
molaical.exe -pmf -i D:/pmf/diher.dat
```

2) Do PMF calculation with “conshd” contour

```
molaical.exe -pmf -i D:/pmf/diher.dat -g 20 -l 10 -m conshd -x "x data" -y "y data"
```

3) Do PMF calculation with “contur” contour without numerical label

```
molaical.exe -pmf -i D:/pmf/diher.dat -g 20 -l 10 -m contur -b none -x "x data" -y "y data"
```

4) Do PMF calculation with “contur” contour without numerical label.

```
molaical.exe -pmf -i D:/pmf/diher.dat -g 20 -l 10 -m contur -b float -x "x data" -y "y data" -t 310 -f Figure.tif
```

## 4.3. Molecular properties calculation

### 4.3.1. Quantitative structure-activity relationship (QSAR)

Quantitative structure-activity relationship (QSAR) is a method involved in regression or classification used in the chemical and biological sciences and engineering. Here, the QSAR is embedded into MolAICal software with genetic algorithm. The detail parameters for run QSAR is listed as below:

**-qsar:** means QSAR calculation function

**-i:** is the path of input data file for QSAR calculation

**-im:** is mutation ratio

**-ic:** number selection for fittest calculation

**-ie:** whether choose elitism way

**-ip:** population size

**-iw:** choose the validation. Currently, it contains least squares R2 and cross validation Q2.

**-in:** number of set variables. If it set 3, it will be like:  $y = a*x_1 + b*x_2 + c*x_3$

**-iv:** number of evolution

**-is:** interval steps for repeating population generation.

**-io:** number core for parallel computing.

The example commands for QSAR calculations are as below:

1) QSAR calculation with default parameters

```
molaical.exe -qsar GA -i D:\\QSAR\\all_QSAR_no.txt
```

2) QSAR calculation with full parameters

```
molaical.exe -qsar GA -i D:\\QSAR\\test1.txt -im 0.5 -ic 5 -ie true -ip 200 -iw Q2 -in 3 -iv 150000  
-is 100 -io 3
```

3) QSAR calculation use parallel running way acquiescently

```
molaical.exe -qsar GA -i D:\\QSAR\\test1.txt -im 0.5 -ic 5 -ie true -ip 200 -iw Q2 -in 3 -iv 150000  
-is 100
```

### 4.3.2. Lipinski's rule of five calculation

Lipinski's rule of five also known as the rule of five (RO5) is a rule to estimate drug-like or determine if a chemical compound has pharmacological or biological activity that would be likely orally active drug<sup>3</sup>. The RO5 cutoff values are multiples of five, which is the origin of the rule's name. It contains three or four rules. The MolAICal evaluate drug-like with five rules:

- 1) No more than 5 hydrogen bond donors
- 2) No more than 10 hydrogen bond acceptors
- 3) A molecular mass less than 500 daltons
- 4) An octanol-water partition coefficient (log P) that does not exceed 5
- 5) No more than 10 rotatable bonds**

In the past years, some rules of Lipinski's rule of five have changed according to the statistical research on the properties of FDA approved drugs (see *J Med Chem.* 2019 Feb 28;62(4):1701-1714)<sup>8</sup>. To estimate the drug-like ligands, the author can set suitable parameters according to the studied need. MolAICal supplies an easy way to set rules parameters.

**Notice:** please distinguish the molecular weight and molecular mass. Please see the detailed information about them: [https://en.wikipedia.org/wiki/Molecular\\_mass](https://en.wikipedia.org/wiki/Molecular_mass). The MolAICal outputs the molecular mass of ligands.

**-tool:** appoint tool function. Here, it is “ruleoffive”

**-f:** molecular format

**-n:** molecule file

**-i:** the file which contains molecules list

**-o:** output results file

**-x:** logP

**-a:** number of hydrogen bond acceptors

**-d:** number of hydrogen bond donors

**-m:** molecular mass

**-r:** number of rotatable bonds



The example commands for RO5 calculations are as below:

1) Calculate RO5 with default original rules (see above five rules)

```
molaical.exe -tool ruleoffive -f mol2 -n "D:\zinc_1879871.mol2"
```

2) Calculate RO5 of ligands set. Note: "mol2List.dat" must be in the directory of molecular set.

```
molaical.exe -tool ruleoffive -f mol2list -i "D:\mol2List.dat" -o "D:\result.dat"
```

3) Calculate RO5 with your given values of rules

```
molaical.exe -tool ruleoffive -f mol2list -i "D:\mol2List.dat" -o "D:\result.dat" -x 5.0 -a 10 -d 5 -m 500.0 -r 10
```

### 4.3.3. PAINS calculation

Pan-assay interference compounds (PAINS) are the compounds which often show the false positive results in the biological assay<sup>4</sup>. PAINS tend to react with numerous biological receptors nonspecifically rather than binding the desired target specifically<sup>17</sup>. Some disruptive functional groups are shared by many PAINS. The Common PAINS contains "toxoflavin, isothiazolones, curcumin, hydroxyphenyl hydrazones, phenol-sulfonamides, enones, rhodanines, catechols, and quianones"<sup>17</sup>. The MolAICal software provides an easy way to calculate PAINS:

**-tool:** appoint tool function. Here, it is "pains"

**-f:** molecular format. It contains "SMILES" and "mol2" formats. We recommend "SMILES" format. You can use Open Babel<sup>18</sup> to change it to "SMILES" format.

**-n:** input molecular file or SMILES sequence

**-i:** input file which contains molecules list or sequences

**-o:** output results

The example commands for PAINS calculations are as below:

1) Calculate PAINS with SMILES format

```
molaical.exe -tool pains -f smiles -n "c1ccccc1N=Nc1ccccc1"
```

2) Calculate PAINS with abbreviation SMILES command parameter

```
molaical.exe -tool pains -f smi -n "c1ccccc1N=Nc1ccccc1"
```

3) Calculate PAINS with mol2 format

```
molaical.exe -tool pains -f mol2 -n "D:\ZINC00154323.mol2"
```

4) Calculate PAINS from the file with contains the list of molecular SMILES sequences

```
molaical.exe -tool pains -f smilist -i "D:\painsTest.txt" -o "D:\smiResult.txt"
```

5) Calculate PAINS from the file with contains mol2 list  
 molaical.exe -tool pains -f mol2list -i "D:\list.dat" -o "D:\mol2Result.txt"

#### 4.3.4. Synthetic Accessibility calculation

The synthetic accessibility (SA)<sup>19</sup> is an important aspect of drug design. The prediction of SA can guide the *de novo* drug design and give chemists useful information for compound synthesis. The calculated parameters of SA by MolAICal is shown as below:

**-tool:** appoint tool function. Here, it is "sa"

**-i:** input molecular SMILES sequence or file contained the list of molecular SMILES sequences.

The example commands for SA calculations are as below:

1) Calculate SA with molecular SMILES sequence

molaical.exe -tool sa -i "FC(F)(F)c1cc(ccc1)N5CCN(CCc2nnc3[C@H]4CCCC[C@H]4Cn23)CC5"

2) Calculate SA from the file contained the list of molecular SMILES sequences

molaical.exe -tool sa -i "D:\\SA\\SmilTest.smi"

#### 4.4. Vinardo score calculation

Vinardo score which is based on Vina score function<sup>11</sup>, is trained on basis of PDBbind database<sup>20-22</sup>. The score function of Vina is show as below. The Vinardo score can be trained by combined with the below equations.

$$Gauss_1(d) = e^{-((d-o_1)/s_1)^2} \quad (1)$$

$$Gauss_2(d) = e^{-((d-o_2)/s_2)^2} \quad (2)$$

$$Repulsion(d) = \begin{cases} d^2, & \text{if } d < 0\text{\AA} \\ 0, & \text{if } d \geq 0\text{\AA} \end{cases} \quad (3)$$

$$Hydrophobic(d) = \begin{cases} 1, & \text{if } d < p_1 \\ p_2 - d, & \text{if } p_1 \leq d \leq p_2 \\ 0, & \text{if } d > p_2 \end{cases} \quad (4)$$

$$Hbond(d) = \begin{cases} 1, & \text{if } d < h_1 \\ 1 - \frac{d - h_1}{-h_1}, & \text{if } h_1 \leq d \leq 0\text{\AA} \\ 0, & \text{if } d > 0\text{\AA} \end{cases} \quad (5)$$

### 4.4.1. Batch of Vinardo score calculation

MolAICal retrain the Vinardo score according to the above five equations. And the command parameters are as below:

- score:** score function for evaluation of binding affinity. Here, it is “vinardo”
- i:** input molecular file or file contained molecules’ list
- o:** output result
- g:** the referred grid file for binding affinity calculation
- p:** pick up the computing way for Vinardo score calculations
- n:** number of cores for parallel computing

The example commands for Vinardo score calculations are as below:

1) Calculate Vinardo score without loading all molecules

```
molaical.exe -score vinardo -i D:\workdir\ligandList.dat -o D:\workdir\output.dat -g D:\workdir\MAICGrid.dat
```

2) Calculate Vinardo score by loading all molecules.

```
molaical.exe -score vinardo -i D:\workdir\ligandList.dat -o D:\workdir\output.dat -g D:\workdir\MAICGrid.dat -p memoryWay -n 2
```

### 4.4.2. Vinardo score Decomposition

The MolAICal can calculate the Gauss<sub>1</sub> score, Gauss<sub>2</sub> score, Repulsion score, Hydrophobic score, Hbond score and Vinardo score based on one ligand, respectively (see equations in 3.7). The calculated parameters are below:

- tool:** appoint tool function. Here, it is “vinardo”
- i:** input molecular file
- g:** the referred grid file
- t:** type of score which is gaussScore1, gaussScore2, repulsionScore, hydrophobicScore, hbondScore or vinardoScore.
- n:** number of cores for parallel computing

1) Calculate Vinardo score from one molecule.

```
molaical.exe -tool vinardo -i D:\workdir\1a1e_ligand.mol2 -g D:\workdir\vinardoScore.dat -t vinardoScore
```

2) Calculate Gauss<sub>1</sub> score with 1 core of computer

```
molaical.exe -tool vinardo -i D:\workdir\1a1e_ligand.mol2 -g D:\workdir\gaussScore1.dat -t gaussScore1 -n 1
```

3) Calculate Gauss<sub>2</sub> score

```
molaical.exe -tool vinardo -i D:\workdir\1a1e_ligand.mol2 -g D:\workdir\gaussScore2.dat -t gaussScore2
```

4) Calculate Repulsion score

```
molaical.exe -tool vinardo -i D:\workdir\1a1e_ligand.mol2 -g D:\workdir\repulsionScore.dat -t repulsionScore
```

5) Calculate Hydrophobic score

```
molaical.exe -tool vinardo -i D:\workdir\1a1e_ligand.mol2 -g D:\workdir\hydrophobicScore.dat -t hydrophobicScore
```

6) Calculate Hbond score

```
molaical.exe -tool vinardo -i D:\workdir\1a1e_ligand.mol2 -g D:\workdir\hbondScore.dat -t hbondScore
```

### 4.4.3. Grid file generation for score calculation

The Gauss<sub>1</sub> score, Gauss<sub>2</sub> score, Repulsion score, Hydrophobic score, Hbond score and Vinardo score need the grid files for binding score calculation. MolAICal supplies a module to calculate the relative grid files for binding score calculation. The parameters are below:

**-tool:** appoint tool function. Here, it is “grid”

**-i:** the path of box file which is measured based on receptor

**-p:** the path of receptor file

**-n:** term of Vinardo score which can be gaussScore1, gaussScore2, repulsionScore, hydrophobicScore, hbondScore or vinardoScore

The example commands are as below:

1) Grid file generation of Gauss<sub>1</sub> score

```
molaical.exe -tool grid -i "D:\1a1e\boxPar.dat" -p "D:\1a1e\protein2.pdb" -n gaussScore1
```

2) Grid file generation of Gauss<sub>2</sub> score

```
molaical.exe -tool grid -i "D:\1a1e\boxPar.dat" -p "D:\1a1e\protein2.pdb" -n gaussScore2
```

3) Grid file generation of Repulsion score

```
molaical.exe -tool grid -i "D:\1a1e\boxPar.dat" -p "D:\1a1e\protein2.pdb" -n repulsionscore
```

4) Grid file generation of Hydrophobic score

```
molaical.exe -tool grid -i "D:\1a1e\boxPar.dat" -p "D:\1a1e\protein2.pdb" -n hydrophobicscore
```

5) Grid file generation of Hbond score

```
molaical.exe -tool grid -i "D:\1a1e\boxPar.dat" -p "D:\1a1e\protein2.pdb" -n hbondscore
```

6) Grid file generation of Vinardo score

```
molaical.exe -tool grid -i "D:\1a1e\boxPar.dat" -p "D:\1a1e\protein2.pdb" -n vinardoScore
```

#### 4.4.4. Box generation based on receptor

Grid file generation needs the box parameters which are extracted on the basis of receptor. MolAICal supplies a function to generate box parameters that can display in UCSF Chimera<sup>23</sup>. The parameters are below:

**-tool:** appoint tool function. Here, it is “box”

**-i:** box center

**-l:** box length

**-t:** transparency shown in UCSF Chimera

**-c:** color shown in UCSF Chimera

**-g:** grid space. Default value is 1.0 Å

The example commands are as below:

1) Calculate the box parameter by using the default value. (Note: **the double quotes are necessary for X, Y, Z coordinates. The interval distance between X, Y, Z coordinates should be one space.**)

```
molaical.exe -tool box -i "52.646 7.634 53.411" -l "30.0 30.0 30.0" -o "D:\chimerabox\box.bild"
```

2) Calculate box parameter with appointed value

```
molaical.exe -tool box -i "52.646 7.634 53.411" -l "40.0 30.0 30.0" -t 0.5 -c red -g 1.0 -o "D:\box.bild"
```

3) This command emphasizes the importance of double quotes. The minus sign must be enclosed by double quote.

```
molaical.exe -tool box -i "-30.011 1.665 -36.581" -l "30.0 30.0 30.0" -o "D:\box.bild"
```

#### 4.4.5. Binding free energy from pKd or pKi

To train the Vinardo score, it needs to calculate the binding free energy according to the Ki, Kd, pKd or pKi values from PDBind database<sup>21,24,25</sup>. Here, the MolAICal provides an easy way to calculate binding free energy if the value of Ki, Kd, pKd or pKi is given.

Here, the International System of Units (SI) is introduced (see Table 4.4.5.1). Most laboratory and literatures uses mol/dm<sup>3</sup>, which is the same with mol/L (also named “M”). For example:

$$\text{mol/m}^3 = 10^{-3} \text{ mol/dm}^3 = 10^{-3} \text{ mol/L} = 10^{-3} \text{ M} = 1 \text{ mmol/L} = 1 \text{ mM}.$$

**Table 4.4.5.1** Molar concentration units

Name	Abbreviation	Concentration	Concentration (SI unit)
millimolar	mM	$10^{-3} \text{ mol/L}$	$10^0 \text{ mol/m}^3$
micromolar	$\mu\text{M}$	$10^{-6} \text{ mol/L}$	$10^{-3} \text{ mol/m}^3$
nanomolar	nM	$10^{-9} \text{ mol/L}$	$10^{-6} \text{ mol/m}^3$
picomolar	pM	$10^{-12} \text{ mol/L}$	$10^{-9} \text{ mol/m}^3$
femtomolar	fM	$10^{-15} \text{ mol/L}$	$10^{-12} \text{ mol/m}^3$
attomolar	aM	$10^{-18} \text{ mol/L}$	$10^{-15} \text{ mol/m}^3$
zeptomolar	zM	$10^{-21} \text{ mol/L}$	$10^{-18} \text{ mol/m}^3$
yoctomolar	yM	$10^{-24} \text{ mol/L}$ (6 particles per 10 L)	$10^{-21} \text{ mol/m}^3$

The "millimolar" and "micromolar" refer to mM and  $\mu\text{M}$  ( $10^{-3} \text{ mol/L}$  and  $10^{-6} \text{ mol/L}$ ), respectively. About the detail relative information of molar concentration units, please see the website: [https://en.wikipedia.org/wiki/Molar\\_concentration](https://en.wikipedia.org/wiki/Molar_concentration)

The parameters for binding free energy calculation in MolaICal are shown as below:

**-tool:** appoint tool function. Here, it is "pkdpki"

**-i:** input value

**-t:** type of calculating way. It can choose "pkx" (means pKd or pKi) or "molar" (means Ki or Kd).

**-u:** unit of molar. It can choose values from the second column values of Table 4.4.5.1.

**-k:** temperature. The default temperature is 298.15 K

1) Calculate binding free energy from pkd (pkd =  $-\log K_d$  or pki =  $-\log K_i$ ), use default temperature: 298.15 K

molaical.exe -tool pkdpki -i 11.92 -t pkx

2) Calculate binding free energy from pkd (pkd =  $-\log K_d$  or pki =  $-\log K_i$ ), use appointed temperature.

molaical.exe -tool pkdpki -i 11.92 -t pkx -k 300

3) Calculate from Kd or Ki with concentration. Default is M (mol/L)

molaical.exe -tool pkdpki -i 1.2 -t molar

4) Calculate from Kd or Ki with pm unit

molaical.exe -tool pkdpki -i 1.2 -t molar -u pm

5) Calculate from Kd or Ki with pm unit under 300 K

molaical.exe -tool pkdpki -i 1.2 -t molar -u pm -k 300

## 4.5. Analysis

### 4.5.1. k-means clustering

The k-means clustering is a popular way to cluster analysis in data mining. The results of drug virtual screening show very similar structures and affinity score. Even though the screened drugs have good binding scores, they maybe have very similar structures and functions. It is difficult to pick up the representative molecule for further drug assay. To solve this problem, the k-means algorithm is used to cluster the results of drug design for further activity assay. The parameters for k-means in MolAICal is:

- tool:** appoint tool function. Here, it is "kmeans"
- n:** clustering number
- i:** the file of principal component
- o:** output file of clustering results

The example commands are as below:

1) k-means command for clustering

```
molaical.exe -tool kmeans -n 3 -i "D:\PC.dat" -o "D:\result.dat"
```

### 4.5.2. Similarity search

#### 4.5.2.1. Fingerprint for similarity search

Molecular fingerprints are a way to encode the molecular structure with a series of binary digits (bits) which can be used to compare the similarity between two molecules. The parameters in MolAICal are shown as below:

- tool:** appoint tool function. Here, it is "finger"
- i:** input file of molecules
- o:** calculated output results

The example commands are as below:

1) Fingerprint calculation command

```
molaical.exe -tool finger -i "D:\workdir\listfiles.dat" -o "D:\workdir\result.dat"
```

### 4.5.2.2. 3D structures similarity comparison

It can compare 3D molecular similarity between two ligands on basis of four specific points which contain the centroid of the ligands, two atoms are closest to and farthest from the centroid, and one atom is farthest from the previous atom. The detailed algorithm is introduced by Ballester PJ and Richards WG<sup>26</sup>. The parameters in MolAICal are shown as below:

- tool:** appoint tool function. Here, it is “3Dcompare”
- i:** the first input file, it can be mol2 file or list file contained molecular mol2 name
- s:** the second file, it can be mol2 file or output file.
- f:** file format, it can be “mol2” or “mol2list”. The default value is “mol2”.

The example commands are as below:

1) Calculate two molecular similarity with default value.

```
molaical.exe -tool 3Dcompare -i D:\workdir\MolAICal\example\3Dcompare\ligand.mol2 -s D:\workdir\MolAICal\example\3Dcompare\1_10023_out.mol2
```

2) Calculate two molecular similarity with mol2 format.

```
molaical.exe -tool 3Dcompare -i D:\workdir\MolAICal\example\3Dcompare\ligand.mol2 -s D:\workdir\MolAICal\example\3Dcompare\1_10023_out.mol2 -f mol2
```

3) Calculate molecular similarity from a list file. **Note:** the first line in file is compared by the remainder lines in file. So please put your compared molecular name in the first place of the file.

```
molaical.exe -tool 3Dcompare -i D:\workdir\MolAICal\example\3Dcompare\list.txt -s D:\workdir\MolAICal\example\3Dcompare\result.dat -f mol2list
```

### 4.5.3. Merge and split files

#### 4.5.3.1. Extract data from one column of file

This function services the drug clusters from the file which contains three independent columns. This function can merge two files by column. For example, one file contains affinity scores, the other is contains fingerprints values. This function can merge these two files to one file which can be named as PC.dat for k-means clustering. The parameters in MolAICal are shown as below:

- tool:** appoint tool function. Here, it is “colth”
- i:** path of input file
- o:** path of output file
- n:** column order. It begins with 0. The first column is 0, and so on.

The example commands are as below:



1) Extract data from the second column of file

```
molaical.exe -tool colth -i "D:\mergecol\5ee7-final_result.txt" -o "D:\mergecol\outputfile.dat" -n 1
```

#### 4.5.3.2. Merge column of two files

MolAICal supplies the function of merging column of two files. You can merge more files one by one based on the previous merged file. The parameters in MolAICal are shown as below:

**-tool:** appoint tool function. Here, it is “col”

**-f:** the path of first input file

**-l:** the path of second input file

**-s:** separator for merging column of two files

**-o:** path of output merged file

The example commands are as below:

1) command example for merging two files

```
molaical.exe -tool col -f "D:\name.dat" -l "D:\Fingerprint.dat" -s " " -o "D:\all.dat"
```

#### 4.5.3.3. Merge two files

ZINC database<sup>27</sup> supplies many small molecular sets with mol2 format. Merge and split molecular files with mol2 format are necessary handling for drug design. Here, MolAICal supplies the merge function for dealing with molecular files with mol2 format. The parameters in MolAICal are shown as below:

**-tool:** appoint tool function. Here, it is “merge”

**-m:** appointed keyword. MolAICal will merge the file whose name contains this keyword.

**-i:** path of input directory

**-o:** path of output file

The example commands are as below:

1) Mol2 format files merging command example

```
molaical.exe -tool merge -m mol2 -i "D:\merge" -o "D:\merge\all.mol2"
```

#### 4.5.3.4. Split and number statistics of mol2 file

In some case, it needs to split the molecular file for drug design. For instance, Autodock vina performs drug virtual screening one by one. It needs to divide the molecular set into one molecule. The MolAICal software supplies a function for splitting the molecular set with mol2 format. The parameters in MolAICal are shown as below:

**-tool:** appoint tool function. Here, it is “split”

**-w:** choose functions. It can be “split”, “chunk” or “num”. “split” means file split, “chunk” means to split the mol2 file to small chunk, “num” means to count the molecular number in the entire mol2 file.

**-n:** when choosing parameter “chunk”, it means the appointed number of molecules in one split chunk file. When choosing parameter “num”, it means the interval number for mol2 file splitting

**-v:** whether overlap the existed molecular file.

**-i:** path of input molecular mol2 file

**-o:** path of output directory

The example commands are as below:

1) Split molecular set to single molecule

```
molaical.exe -tool split -w split -n 1 -v true -i "D:\split\all.mol2" -o "D:\split\1"
```

2) Census the number of molecular set

```
molaical.exe -tool split -w num -i "D:\split\all.mol2"
```

3) Splits mol2 file to the small chunk which contains 20 molecules.

```
molaical.exe -tool split -w chunk -n 20 -i "D:\split\all.mol2" -o "D:\split\2"
```

#### 4.5.3.5. Change Mol2 to SMILES format

This function just service for changing Mol2 to SMILES format of molecules in the *de novo* drug design. This function will continue to be developed. The parameters in MolAICal are shown as below:

**-tool:** appoint tool function. Here, it is “mol2tosmi”

**-i:** path of molecular input file with mol2 format

The example commands are as below:

1) command example of mol2 to SMILES

```
molaical.exe -tool mol2tosmi -i "D:/mol2tosmi/zinc_98180786.mol2"
```

#### 4.5.4. Split molecules into fragments

It will get useful information on drug design by splitting appointed molecules into fragments. For example, the fragments which are split from FDA approved drugs, can give effective small chip for *de novo* drug design based on fragments growth. In addition, it can find the important rules for drug discovery via analysis of fragment features of appointed molecule set such as natural compounds,

marine drugs, and so on. Here, MolAICal supplied one way to split one molecule into fragments by rotation bond. The parameters in MolAICal are shown as below:

**-tool:** appoint tool function. Here, it is “fragSplit”

**-i:** path of molecular list file

**-o:** path of output fragments

**-oa:** path of output fragments with adding hydrogen

**-w:** method for dealing with hydrogen. It contains “add”, “del” and “off”. The “add” means the split fragments will add hydrogen and copy it into the “-oa” appointed directory. The “del” means the split fragments will delete hydrogen and copy it into the “-oa” appointed directory. The “off” means the split fragments will not perform any action and copy it into the “-oa” appointed directory. The default value is “off”.

The example commands are as below:

1) Split molecules into fragments and do not perform any hydrogen operation

```
molaical.exe -tool fragSplit -i "D:\\fragment\\list.dat" -o "D:\\fragment\\1" -oa "D:\\fragment\\3"
```

2) Split molecules into fragments and do not perform any hydrogen operation

```
molaical.exe -tool fragSplit -i "D:\\fragment\\list.dat" -o "D:\\fragment\\1" -oa "D:\\fragment\\3" -w off
```

3) Split molecules into fragments with adding hydrogen

```
molaical.exe -tool fragSplit -i "D:\\fragment\\list.dat" -o "D:\\fragment\\1" -oa "D:\\fragment\\3" -w add
```

4) Split molecules into fragments with deleting hydrogen

```
molaical.exe -tool fragSplit -i "D:\\fragment\\list.dat" -o "D:\\fragment\\1" -oa "D:\\fragment\\3" -w del
```

# Appendix

## 1. Configure file of fragments growth using DL model

# This is an example of configure file with deep learning (DL) model.

# “#” is the annotation

```
centerPoints          -30.011  1.665  -36.581
boxLengthXYZ          30.0 30.0 30.0
receptorPDB           /home/bqf/create/example/GCGRNoLigand.pdb
```

# Two ways for defining the frag grow way.

# 1. First way for grow. The growMethod contains fixFrag and randomFrag

```
growMethod            fixFrag
startFragFile          /home/bqf/create/example/startFrag.mol2
```

# 2. Second way for grow. It is suitable for no crystal ligand in the receptor pocket.

```
# growMethod          randomFrag
# startFragFile        D:/workdir/MolAICal/example/grow/genstartFrag.mol2
# startAtomPosition    D:/workdir/MolAICal/example/grow/resPosition.pdb
# startSmiFrag         C
```

# define the population

```
numCycleGen          24
selTopMols            200
selTopRootPercent     0.70
selElitePercent       0.10
maximumPOP            3000
```

# define read library way: mol2, SMILES, AIFrag

```
libStyle              AIFrag
genAIWay              Fdafrag
genAINumber            81
# obabelPath           C:\Program Files\OpenBabel-2.4.1\obabel.exe
```

# Search the conformation algorithm

# Two methods: "random" and "fibonacci"

```
perturbeSearch        on
genAlgorithm           fibonacci
ranGenOptNum           361
```

# define mutation way

atomMutation	off
mutationPercent	0.50
# grow algorithm	
randomCrossMutation	on
randomCrossRatio	0.50
randomMutationRatio	0.50
randomCrossCompareNum	2
randomCrossRange	3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18
# value on or off	
writeResultParallel	on
# filter molecule. Value is on or off.	
unwantedFrag	on
PAINS	on
growFilter	on
growFilterRatio	0.50
# default is 0.50	
resultsFilterRatio	0.50
# rules of five (see: <i>J Med Chem.</i> 2019 Feb 28;62(4):1701-1714.)	
RulesOfFive	on
xlogPvalue	6.0
acceptors	12
donors	6
mwvalue	1000.0
rotatablebonds	14
# SA	
syntheticAccessibility	off
synAccessibilityValue	50
MolsimilarPercent	0.90
# define output results	
# pcClusterdFile	D:/workdir/MolAICal/example/grow/PC.dat
pcClusterNumber	10
adjustOutSimPer	0.999999
# moloutdir	D:/workdir/MolAICal/example/grow/OX2Orexin/results
selMolMaxWeight	1000
selMolMinWeight	100
selBindingScore	-3

```
# define cpu and memory
coreNum                20
```

## 2. Configure file of fragments growth without DL model

```
# This is an example of configure file without deep learning (DL) model.
# “#” is the annotation
```

```
centerPoints            -30.011  1.665  -36.581
boxLengthXYZ            30.0 30.0 30.0
receptorPDB             /home/bqf/create/example/GCGRNoLigand.pdb
```

```
# Two ways for defining the frag grow way.
```

```
# 1. First way for grow. The growMethod contains fixFrag and randomFrag
```

```
growMethod              fixFrag
startFragFile           /home/bqf/create/example/startFrag.mol2
```

```
# 2. Second way for grow. It is suitable for no crystal ligand in the receptor pocket.
```

```
# growMethod            randomFrag
# startFragFile         D:/workdir/MolAICal/example/grow/genstartFrag.mol2
# startAtomPosition     D:/workdir/MolAICal/example/grow/resPosition.pdb
# startSmiFrag          C
```

```
# define the population
```

```
numCycleGen            24
selTopMols              200
selTopRootPercent      0.70
selElitePercent        0.10
maximumPOP             3000
```

```
# define read library way: mol2, SMILES, AIFrag
```

```
libStyle               mol2
genAIWay               Fdafrag
genAINumber            81
```

```
# Search the conformation algorithm
```

```
# Two methods: "random" and "fibonacci"
```

```
perturbeSearch         on
genAlgorithm            fibonacci
ranGenOptNum           361
```

```
# define mutation way
```

```

atomMutation                off
mutationPercent              0.50

# grow algorithm
randomCrossMutation          off
randomCrossRatio              0.50
randomMutationRatio          0.50
randomCrossCompareNum        2
randomCrossRange              3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18
# value on or off
writeResultParallel          on

# filter molecule. Value is on or off.
unwantedFrag                 on
PAINS                        on
growFilter                    on
growFilterRatio               0.50
##default is 0.56
resultsFilterRatio            0.50

# rules of five (see: J Med Chem. 2019 Feb 28;62(4):1701-1714.)
RulesOfFive                   on
xlogPvalue                    6.0
acceptors                     12
donors                        6
mwvalue                       1000.0
rotatablebonds                14

# SA
syntheticAccessibility        off
synAccessibilityValue         50

MolsimilarPercent             0.90

# define output results
pcClusterNumber               10
adjustOutSimPer                0.999999

# filters
selMolMaxWeight               1000
selMolMinWeight                100
selBindingScore                -3

# define cpu and memory

```

### 3. Configure file for simple radii measurement

```
pdspath      D:/GramicidinA/1JNO.pdb
cpoint       0.1625 -0.629 -1.838
vector       0.00 0.00 1.00
sample       0.25
conpar       0.15
endrad       5.0
selatoms     5.1
numpt        300
surColor     blue
lineColor    red
lineWidth    2
searchNum    2000
material     Glass2
```

### 4. Configure file for radii measurement with CHARMM ff

```
pdspath      D:/GramicidinA/1JNO.pdb
psfpath      D:/GramicidinA/1JNO.psf
cpoint       0.1625 -0.629 -1.838
vector       0.00 0.00 1.00
sample       0.25
conpar       0.15
endrad       5.0
selatoms     5.1
numpt        300
surColor     blue
lineColor    red
lineWidth    2
searchNum    2000
material     Glass2
```



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