

# Recherche de variants génomiques en oncologie clinique

Avec des diapos, données & scripts R de:  
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# Technologies de recherche de variants

- Sanger
  - Toujours utilisé en consultation de génétique
- SNP arrays
  - ~1M SNP (recherche GWAS ou « 23&Me »)
- Panel de gènes
  - Une série d'exons d'intérêt (gènes de cancer= 100kb)
- WES (Exome)
  - Tous les exons du génome (30 Mb)
- WGS
  - Whole genome (3 Gb)

NGS

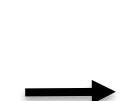
# En génétique constitutionnelle

At hospital



Blood sample

Sequence  
gene panel



Look for  
specific  
alteration  
(BRCA)

Research



Genotype  
or  
Sequence



Compare  
disease and  
healthy  
cohorts

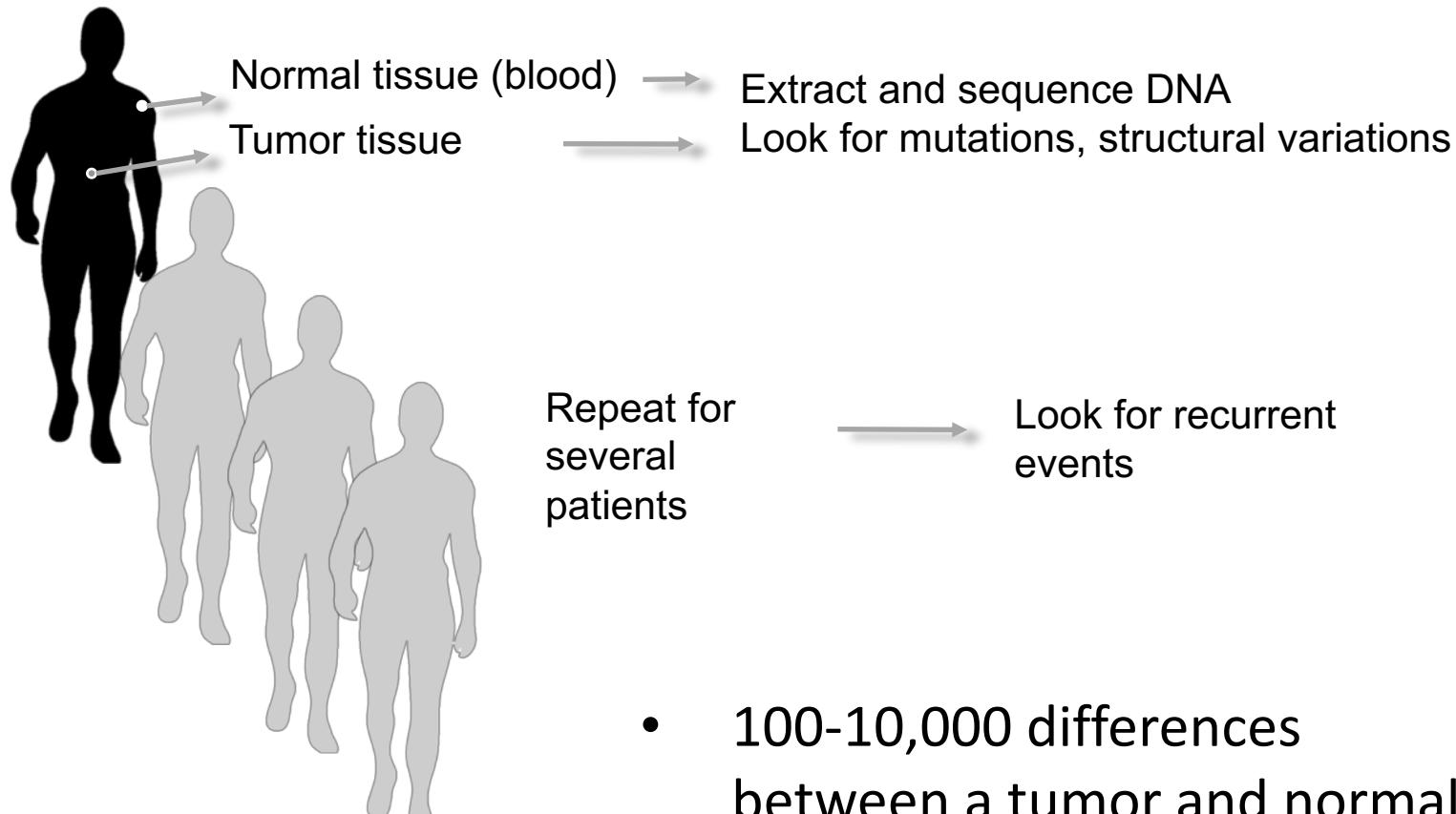
GWAS studies, 1000 Genome Project...

# NGS dans le diagnostic de génétique familiale

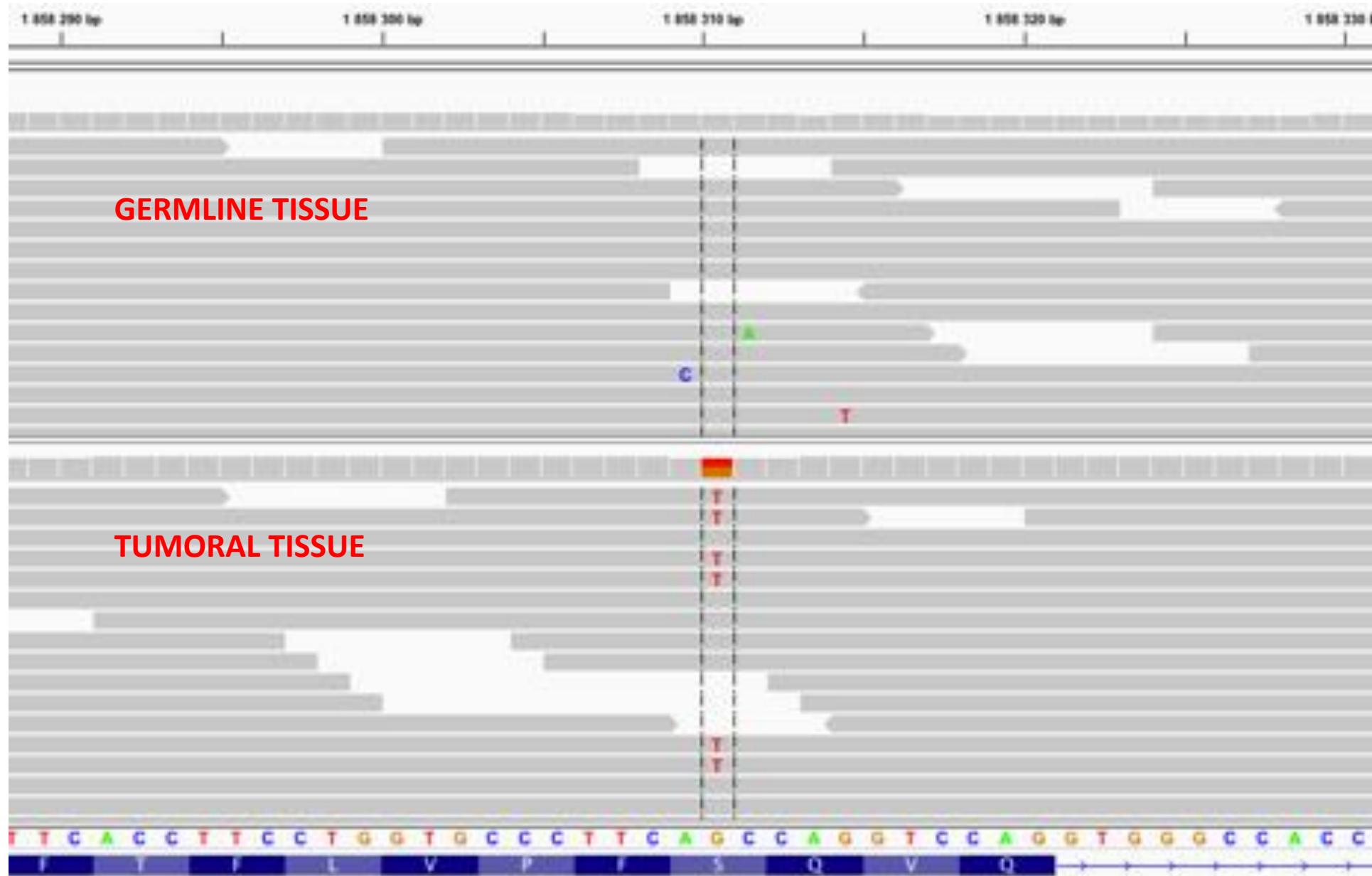
- BRCA1/2 (breast/ovary cancer)
- XPC, XPV.. (melanoma)
- ERCC1 (colorectal cancer)

# En génétique somatique

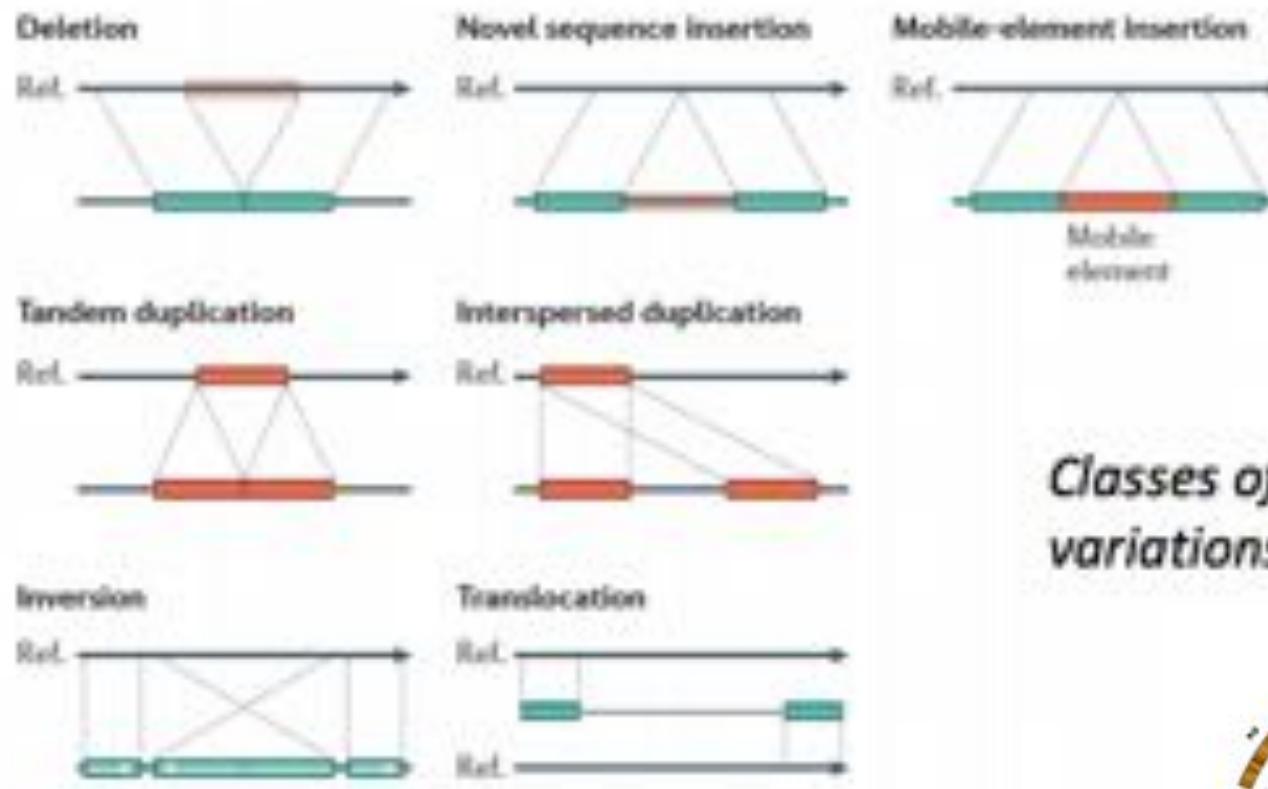
## Finding somatic mutations in the tumor genome



# Les mutations somatiques

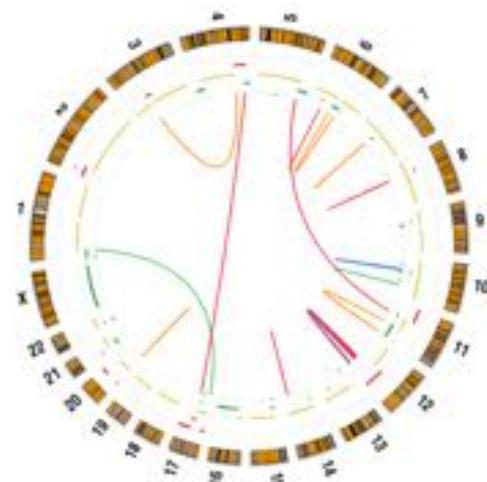


# Variants structuraux

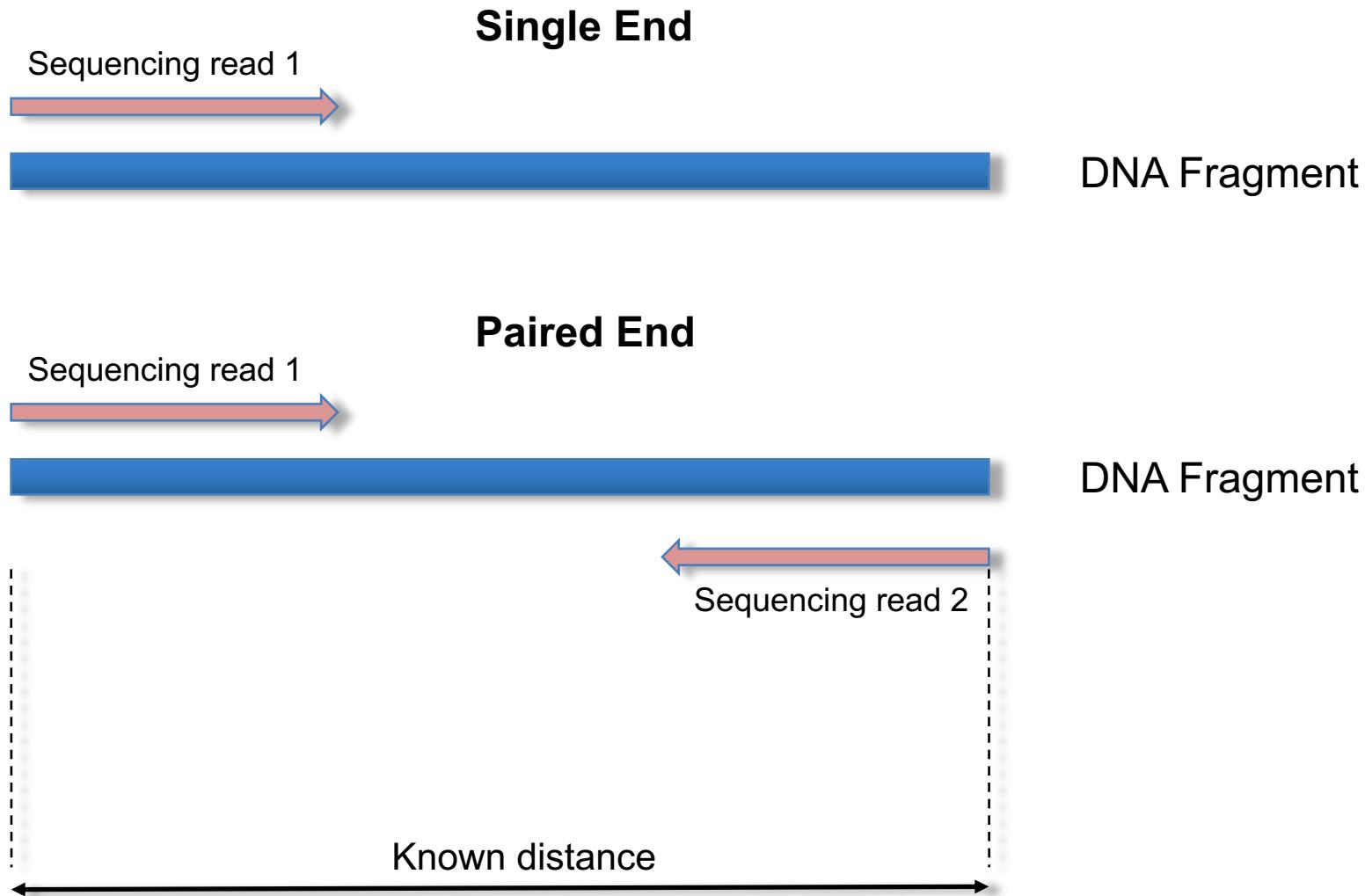


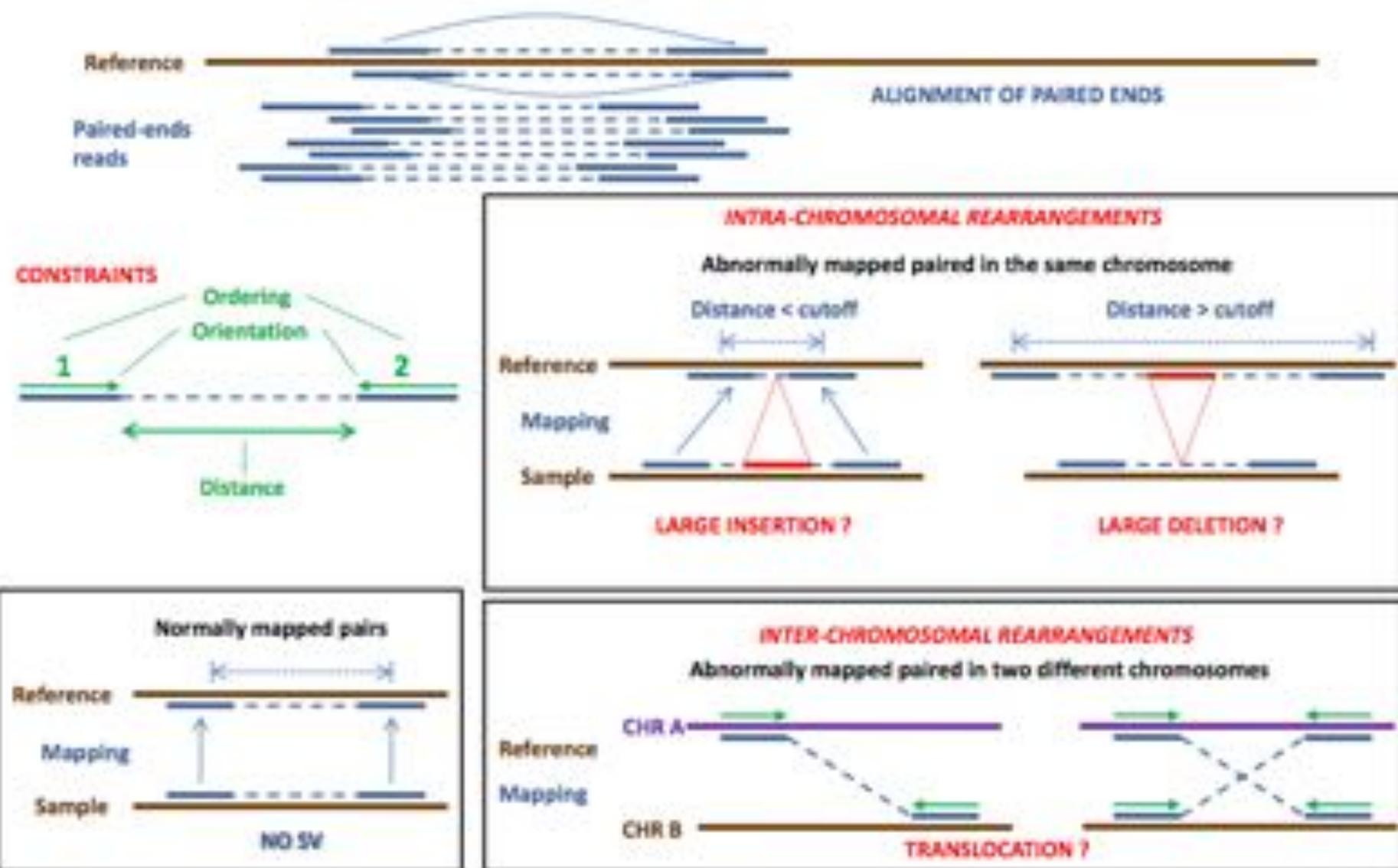
*Classes of structural variations*

Alkan et al 2012

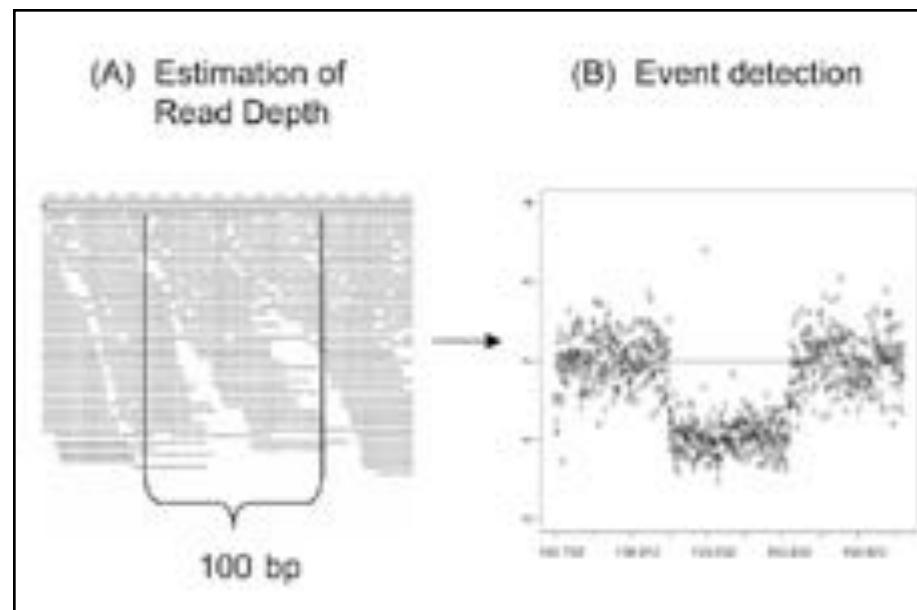
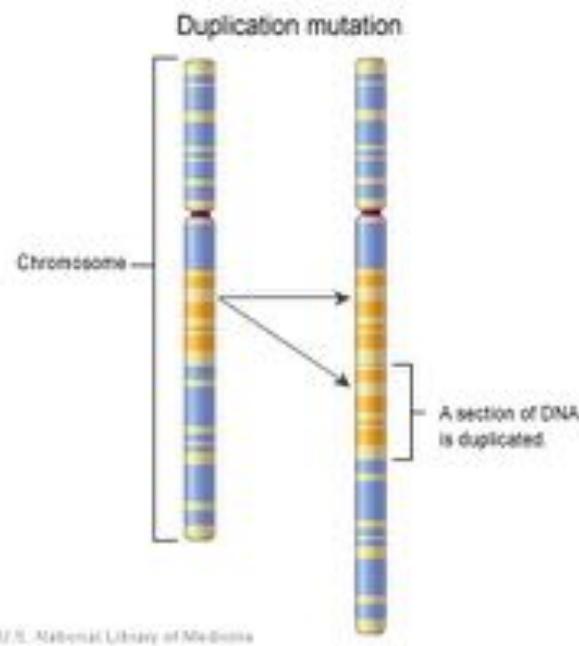


# Intérêt du séquençage paired-end: Résolution des repeats et des variants structuraux





# Recherche de CNV (copy number variations)



- Attempts to infer variations in copy number from the **local read depth**.
- A strong GC% debiasing is required

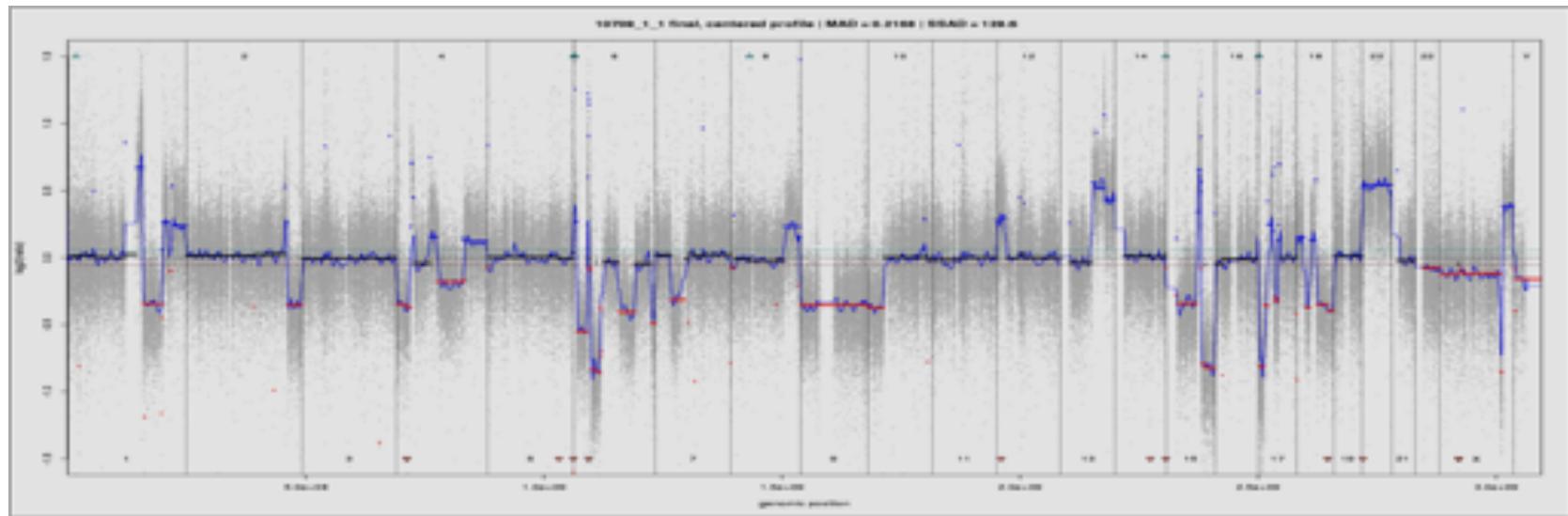
Voir cours Bastien Job

*Yoon, 2009*

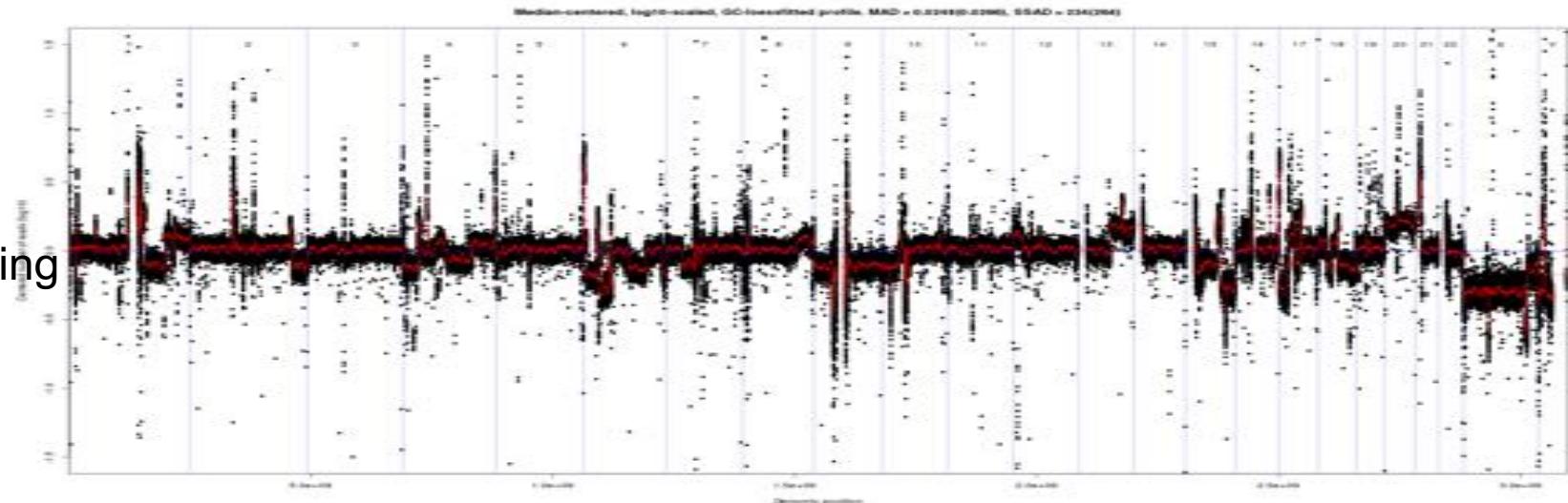
# NGS vs CGH

CGH

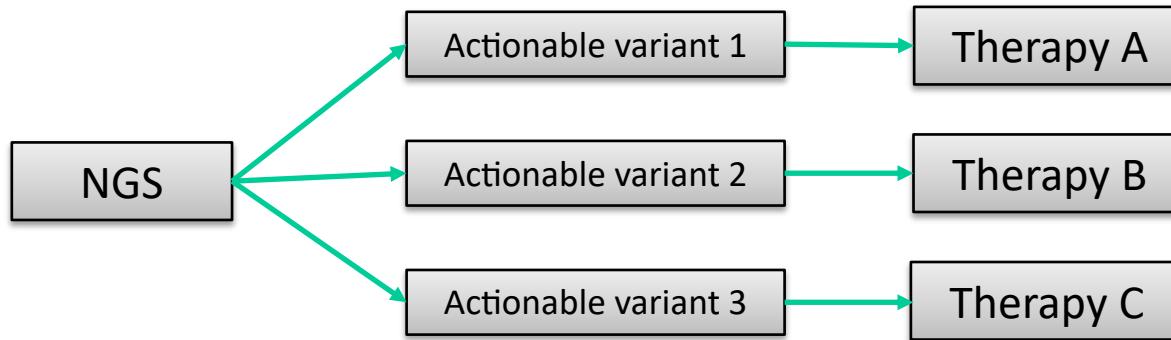
comparative  
genome  
hybridization



Sequencing

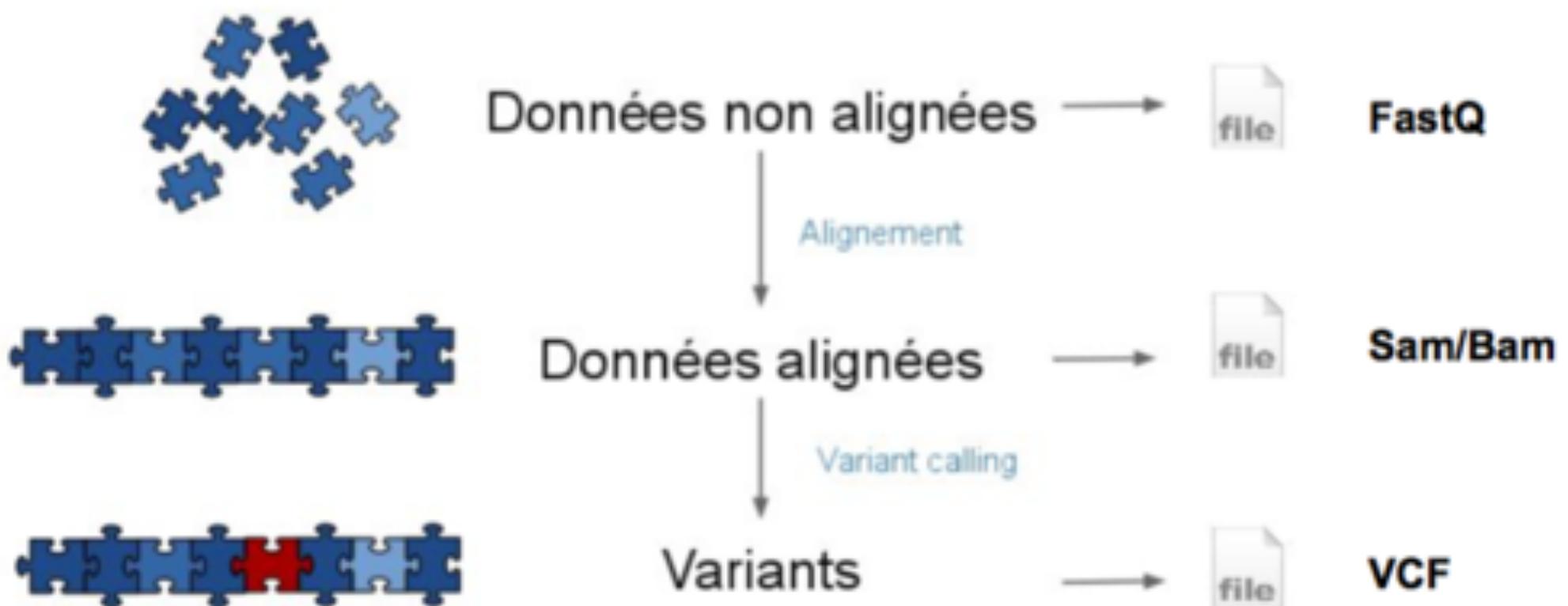


# NGS for precision medicine

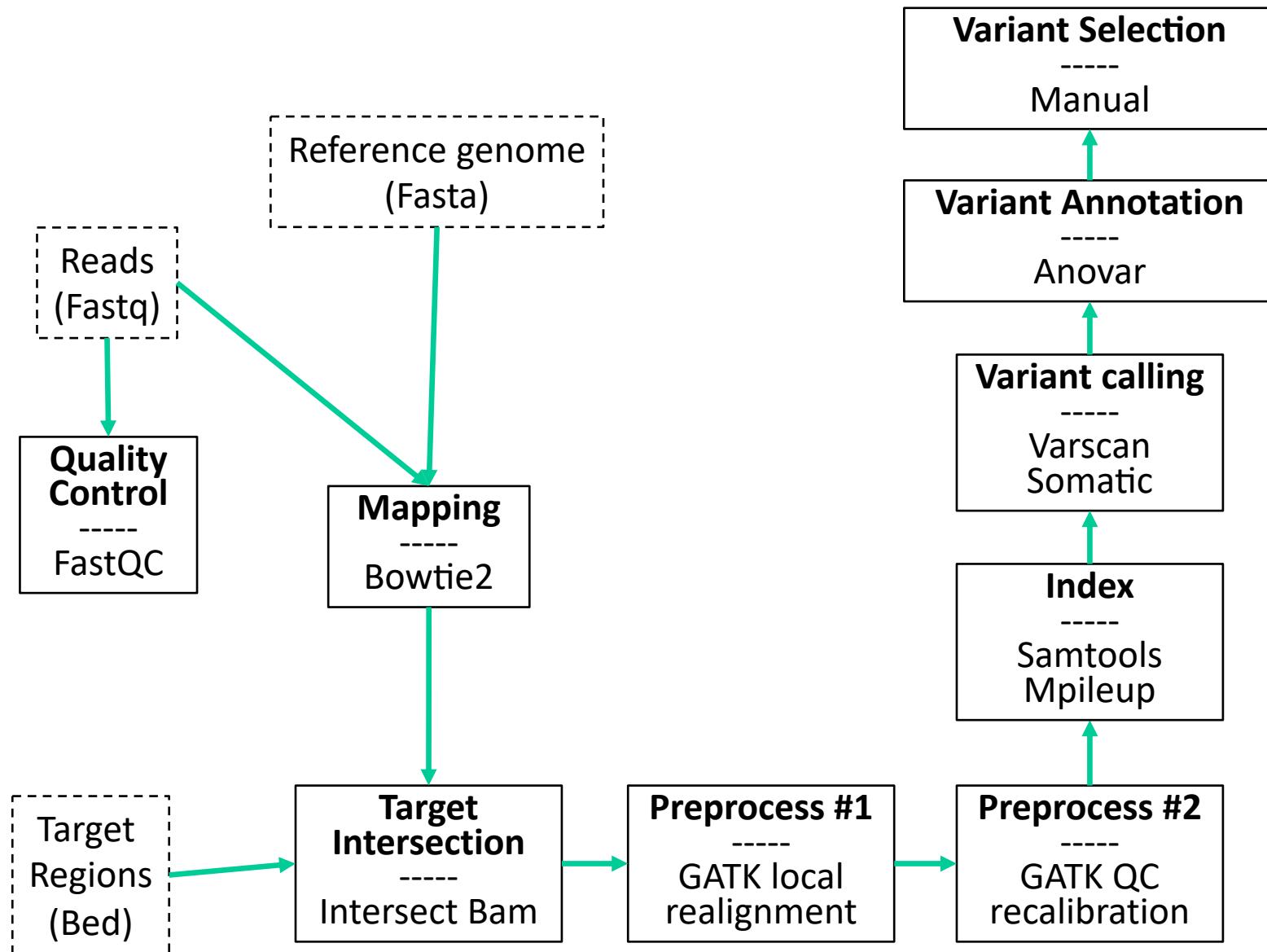


- Clinical trials: MOSCATO (GR), SAFIR (GR), SHIVA (Curie), ...
- Ipilimumab (anti-CTLA4), Nivolumab (anti-PD1), Trastuzumab (anti-HER2), Cetuximab (anti-EGFR)

# Un pipeline « Variants »

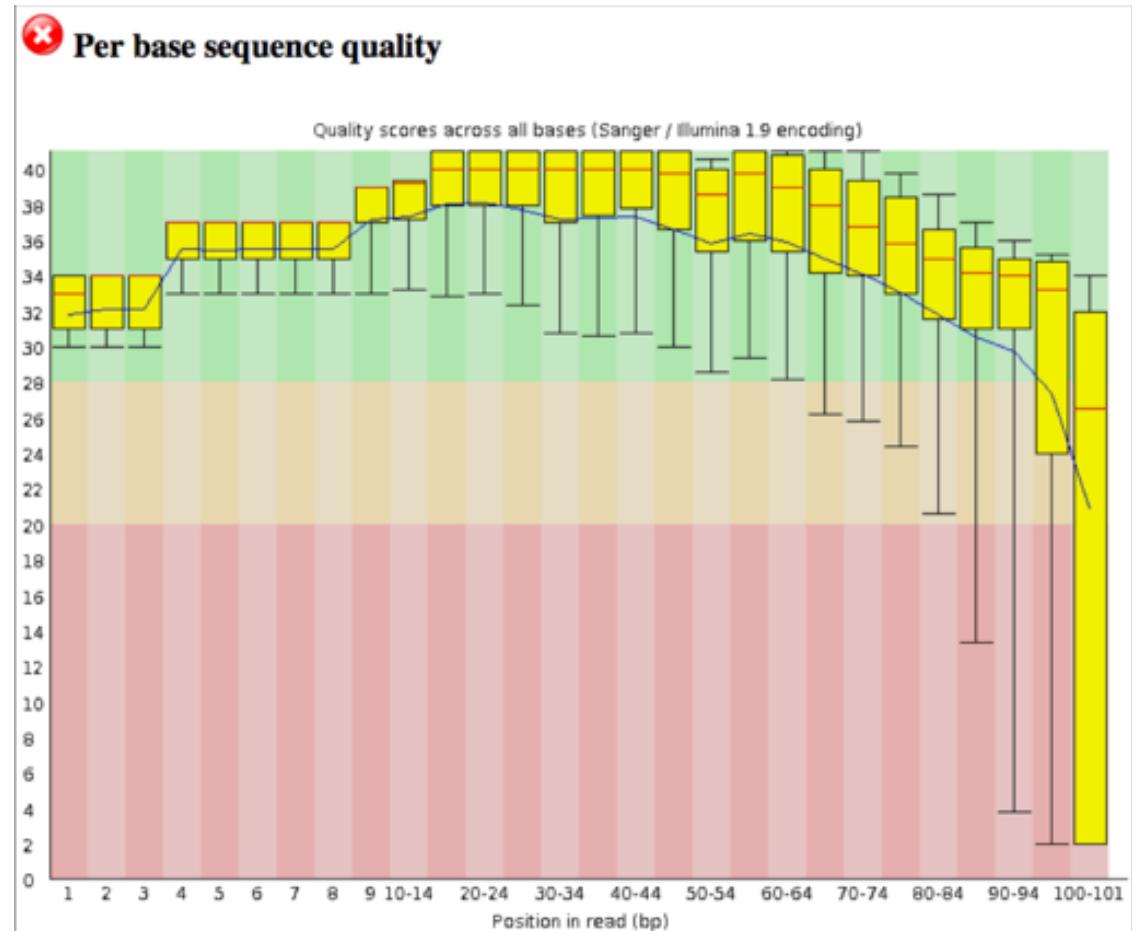


# Un vrai pipeline « variants »



# FastQC Metrics

- Look at the different metrics for both reads
- **Problem:** the per base sequence quality of the Read2 are quite low towards the end



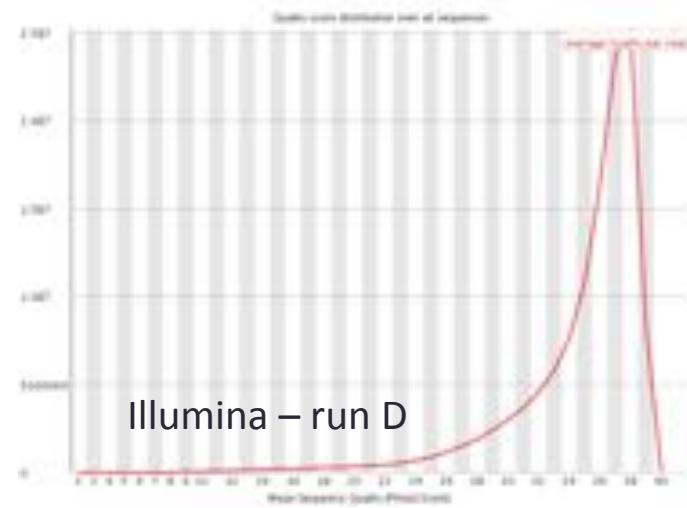
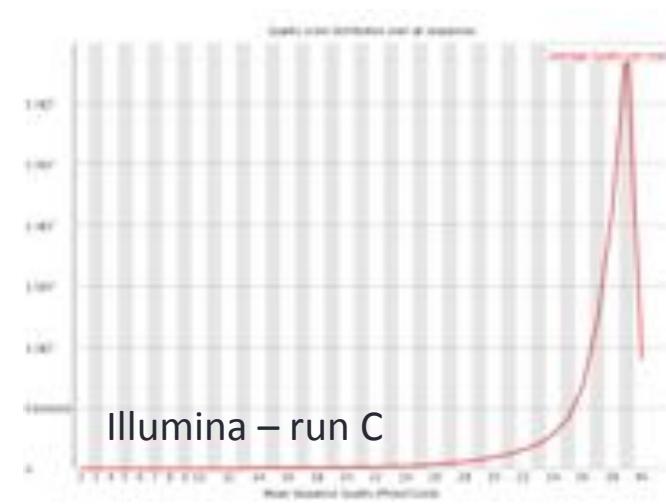
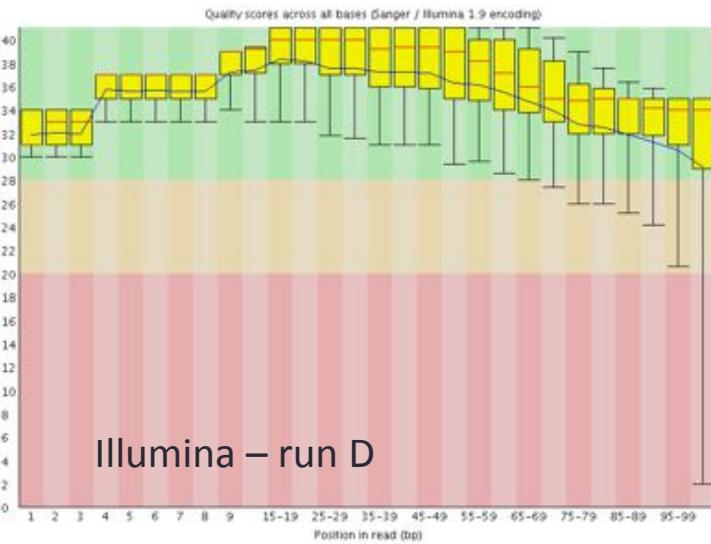
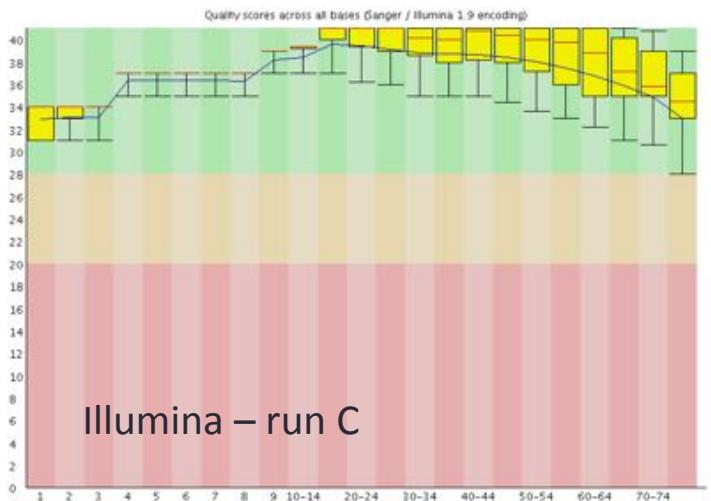
## Solution:

Trim the 25bp from  
the 3' end  
of the reads

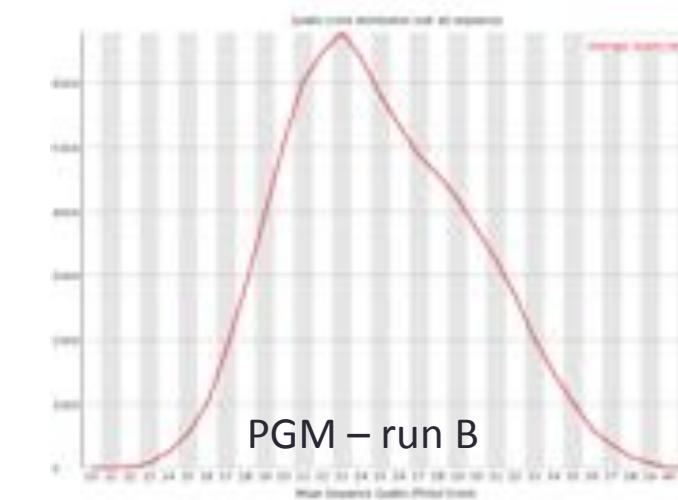
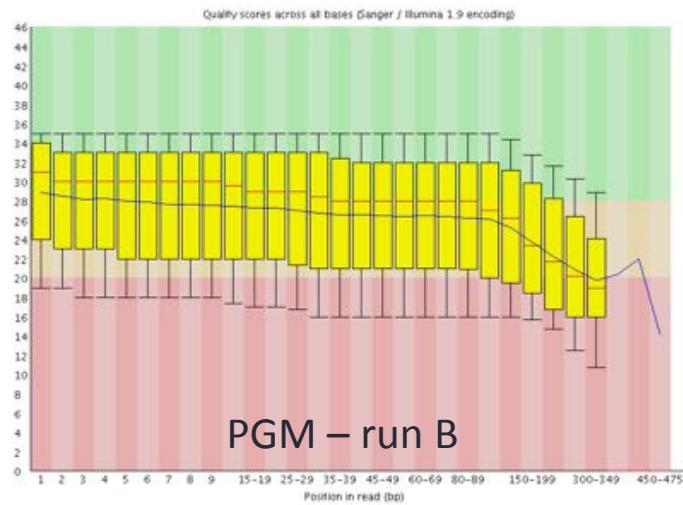
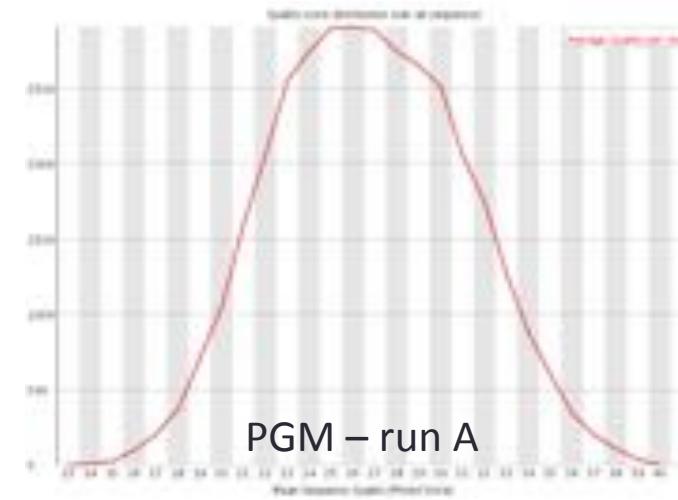
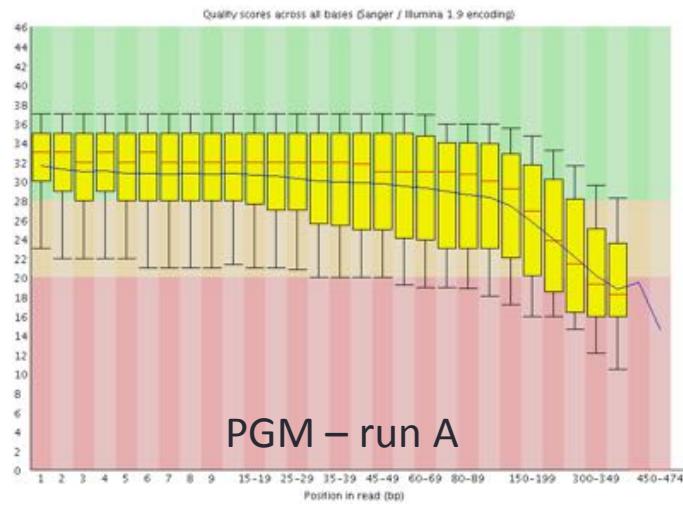
➤ Higher confidence in the  
sequenced information

(FastqTrimmer)

# Illumina

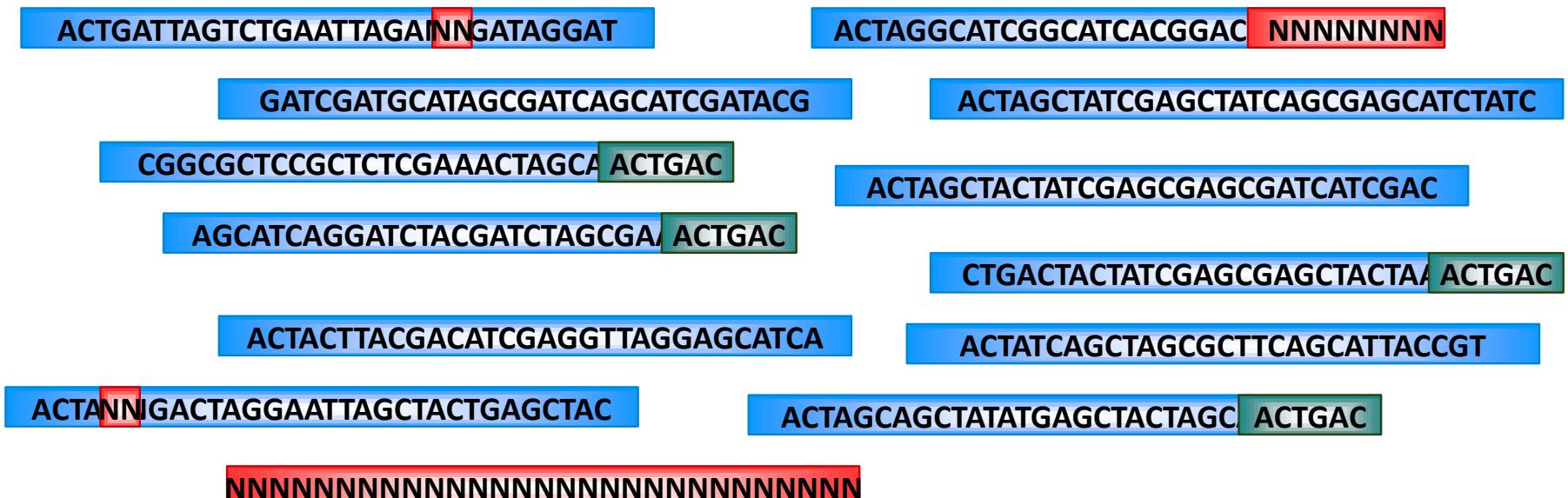


# PGM

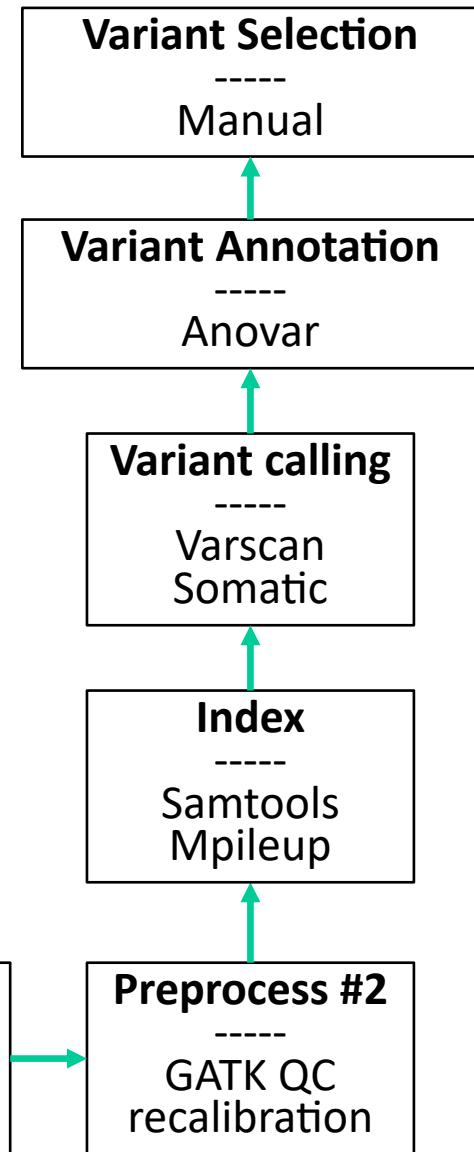
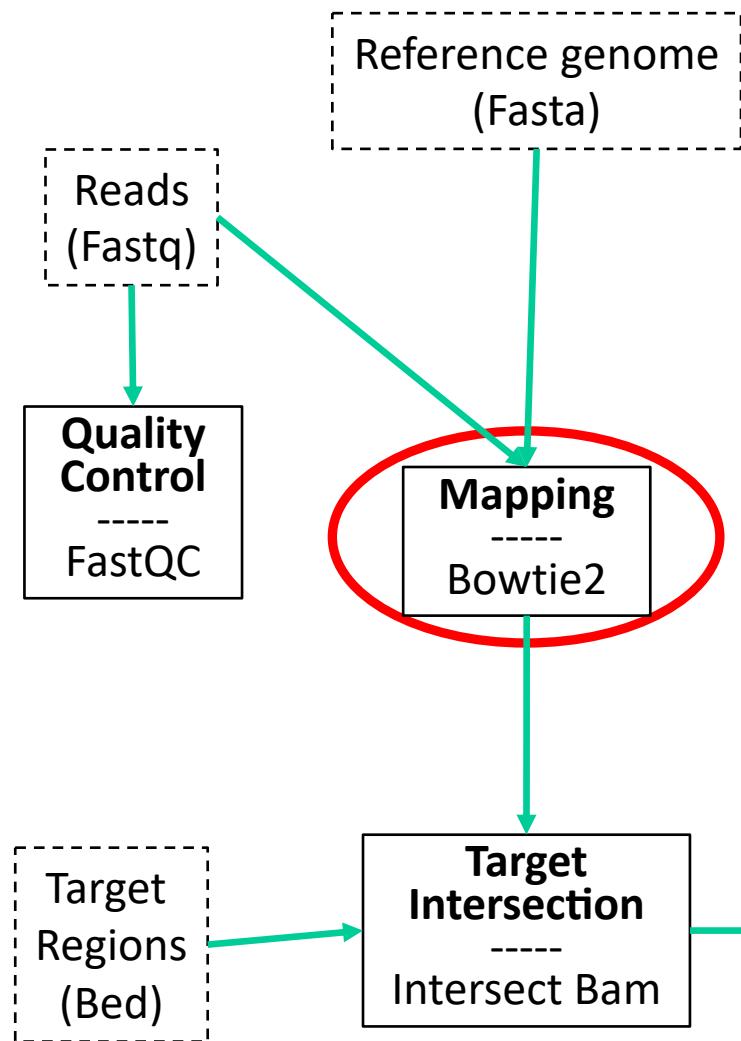


# Trimming and discarding low quality reads...

A first Quality Control of raw reads is mandatory and can be established according to the application ('N', adapter sequences, barcode, contamination, etc.)



Processed reads: blue parts are to  
be kept, green and red parts to be  
removed



# Alignment des reads sur le génome de référence



# Algorithme en $O(mn)$

ACGTTACCGAATCGATCAAGTCGA  
TAC



OK pour 1 read:  $O(3.10^9 \times 100)$   
Mais pour  $1^8$  reads???

« supercalifragilis·ticexpialidocious »

←   →

Préfixe                                      Suffixe

“GOOGOL”

## Suffix array

Tableau trié de tous les suffixes  
d'une chaîne de caractères

0 GOOGOL\$		6 \$	
1 OOGOL\$		3 GOL\$	
2 OGOL\$		0 GOOGOL\$	
3 GOL\$	→	5 L\$	→ (6,3,0,5,2,4,1)
4 OL\$		2 OGOL\$	
5 L\$		4 OL\$	
6 \$		1 OOGOL\$	

Propriété: toutes les occurrences d'une même chaîne  
sont regroupées.

# Suffix arrays

Exemple: trouver la chaîne **GO**

0 GOOGOL\$		6 \$
1 OOGOL\$		3 <b>G</b> O L\$
2 OGOL\$		0 <b>GO</b> OGOL\$
3 GOL\$	→	5 L\$
4 OL\$		2 OGOL\$
5 L\$		4 OL\$
6 \$		1 OOGOL\$

# Most popular aligners for variant analysis

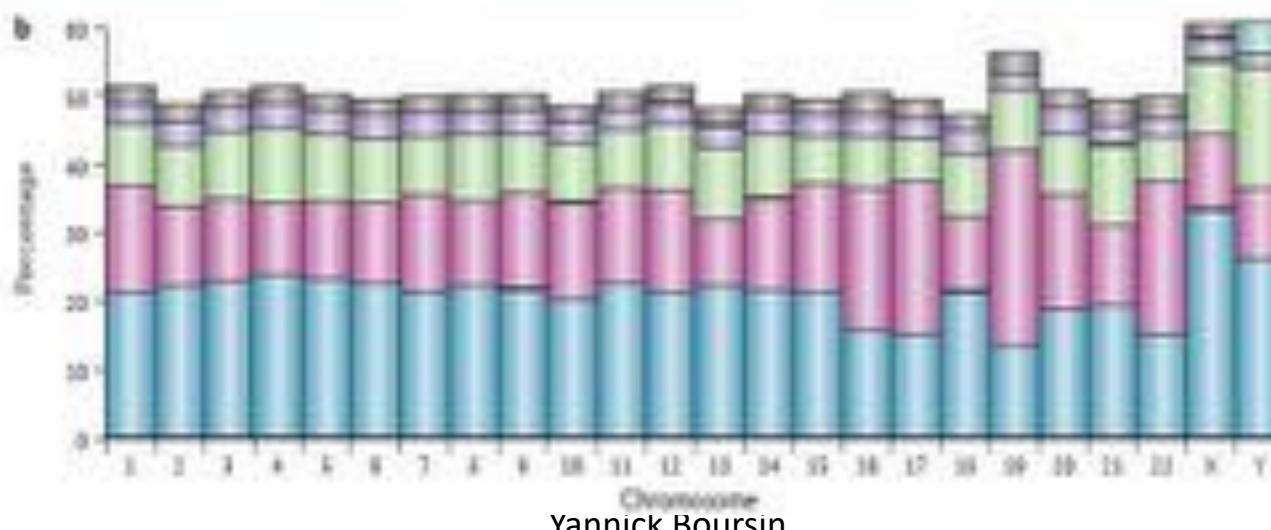
(support mismatched, gapped, paired-end alignment)

- BWA
  - Li H. and Durbin R. (2009)
- Bowtie2
  - Langmead B, Salzberg S (2012)

# Alignment key parameters - Repeats

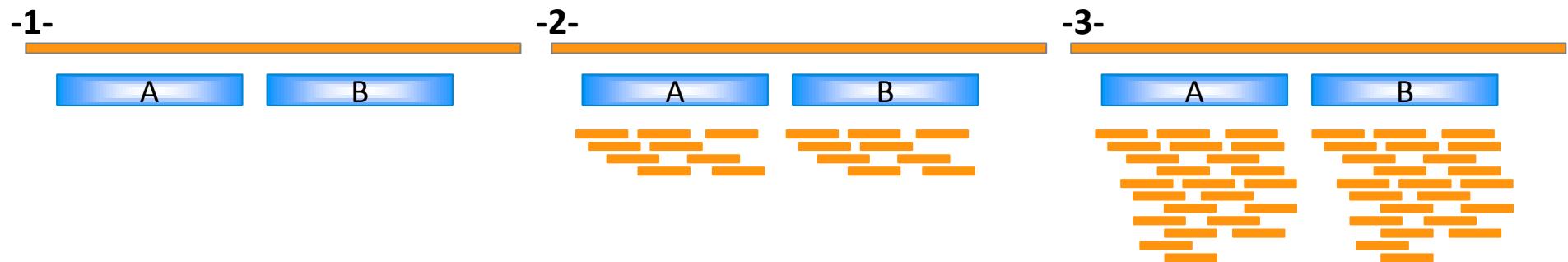
Approximately **50%** of the human genome is comprised of repeats

Repeat class	Repeat type	Number (bp/2L)	Ctg.	Length (bp)
Microsatellite, minisatellite or satellite	Tandem	425,518	1%	2-100
LINE	Interspersed	1,747,375	11%	100-1000
Alu	Interspersed	463,778	2%	200-2000
LTR retrotransposon	Interspersed	718,125	2%	300-5,000
LINE	Interspersed	1,508,345	21%	500-10,000
rDNA (18S, 28S, 5.8S and 25S)	Tandem	601	0.01%	1,000-41,000
Segmental duplications and other elements	Tandem or interspersed	2,779	0.2%	1,000-100,000



# Alignment key parameters – Repeats – 3 strategies

- 1- Report only unique alignment
- 2- Report best alignments and randomly assign reads across equally good loci
- 3- Report all (best) alignments



Treangen T.J. and Salzberg S.L. 2012. Nature review Genetics 13, 36-46

# Alignment key parameters – Using single or paired-end reads ?

The type of sequencing (i.e. single or paired-end reads) is often driven by the application.

**Exemple :** Finding large indels, genomic rearrangements, ...

However, in most cases, the pair information can improve mapping specificity

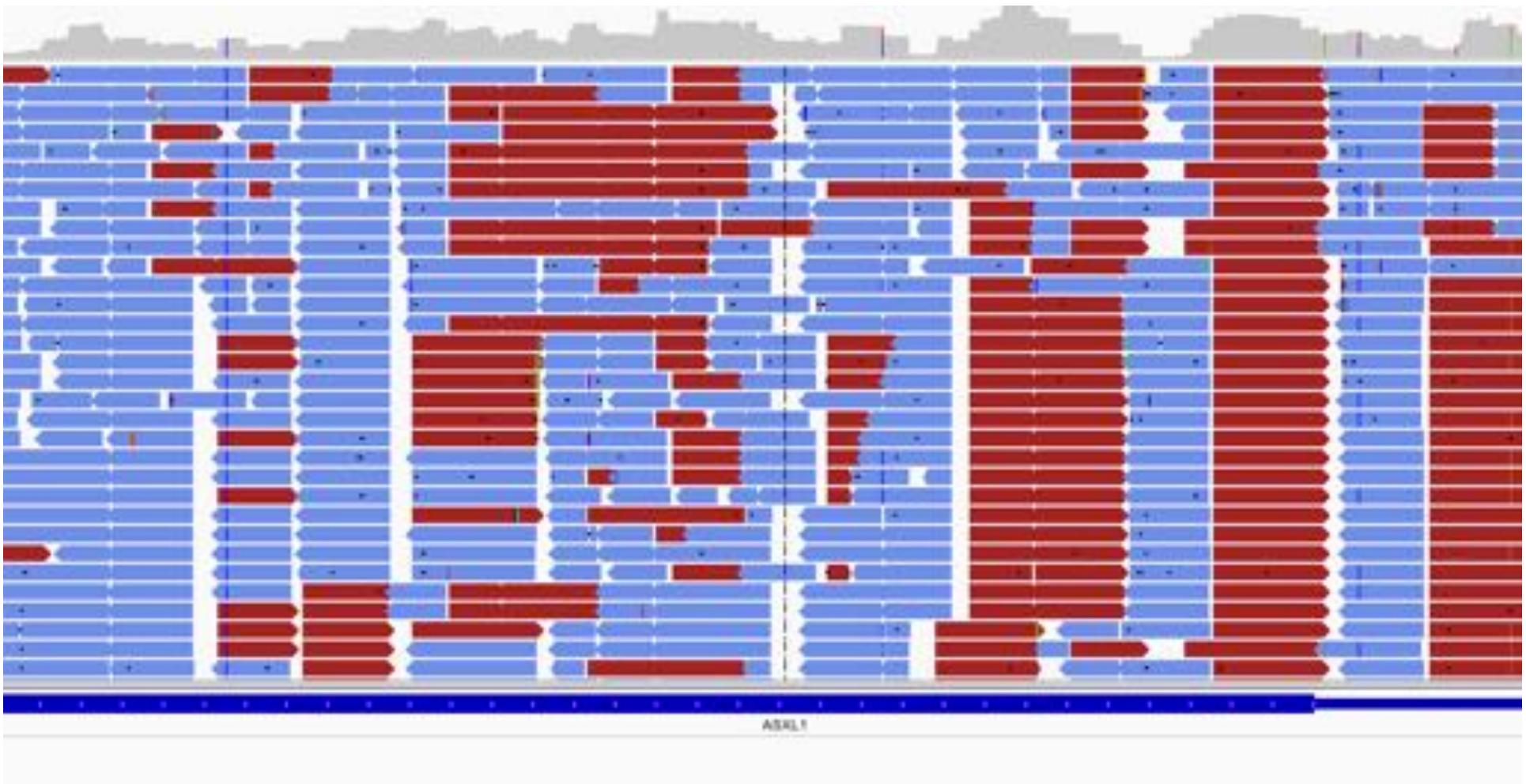
- Single-end alignment – repeated sequence



- Paired-end alignment – unique sequence



# Reads alignés: le format BAM/SAM



# Format BAM/SAM

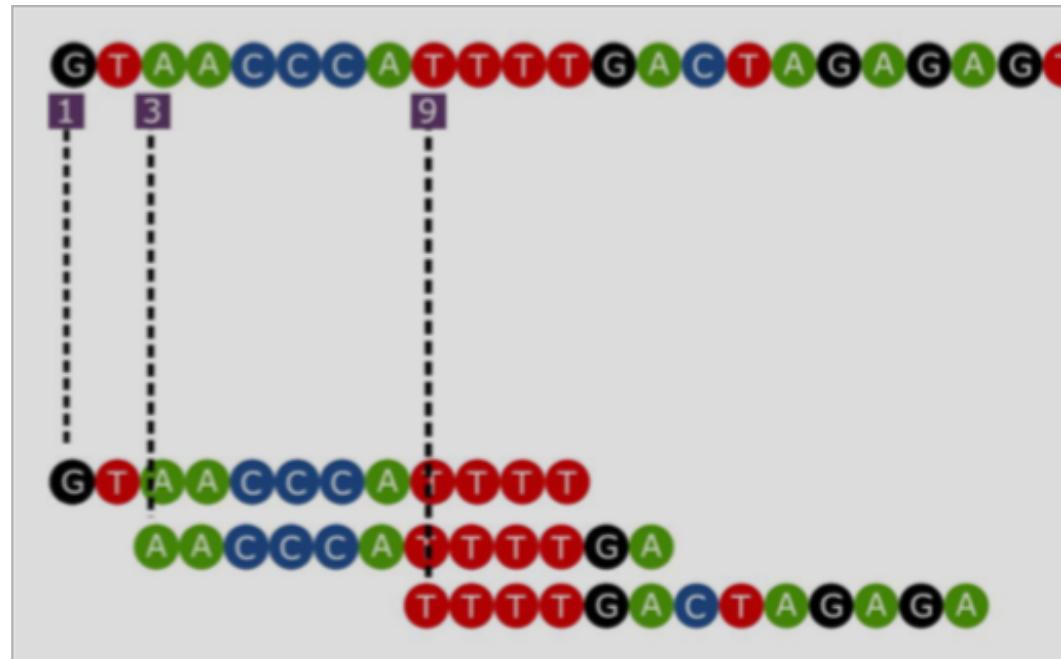
Contient les séquences alignées sur le génome

Dans l'idéal :

chr7 1324324 ACGTGCCTTCGCGT

chr8 1424324 GCGTGATGCGTAAG

chr8 1724354 GTATGTTATATGTA



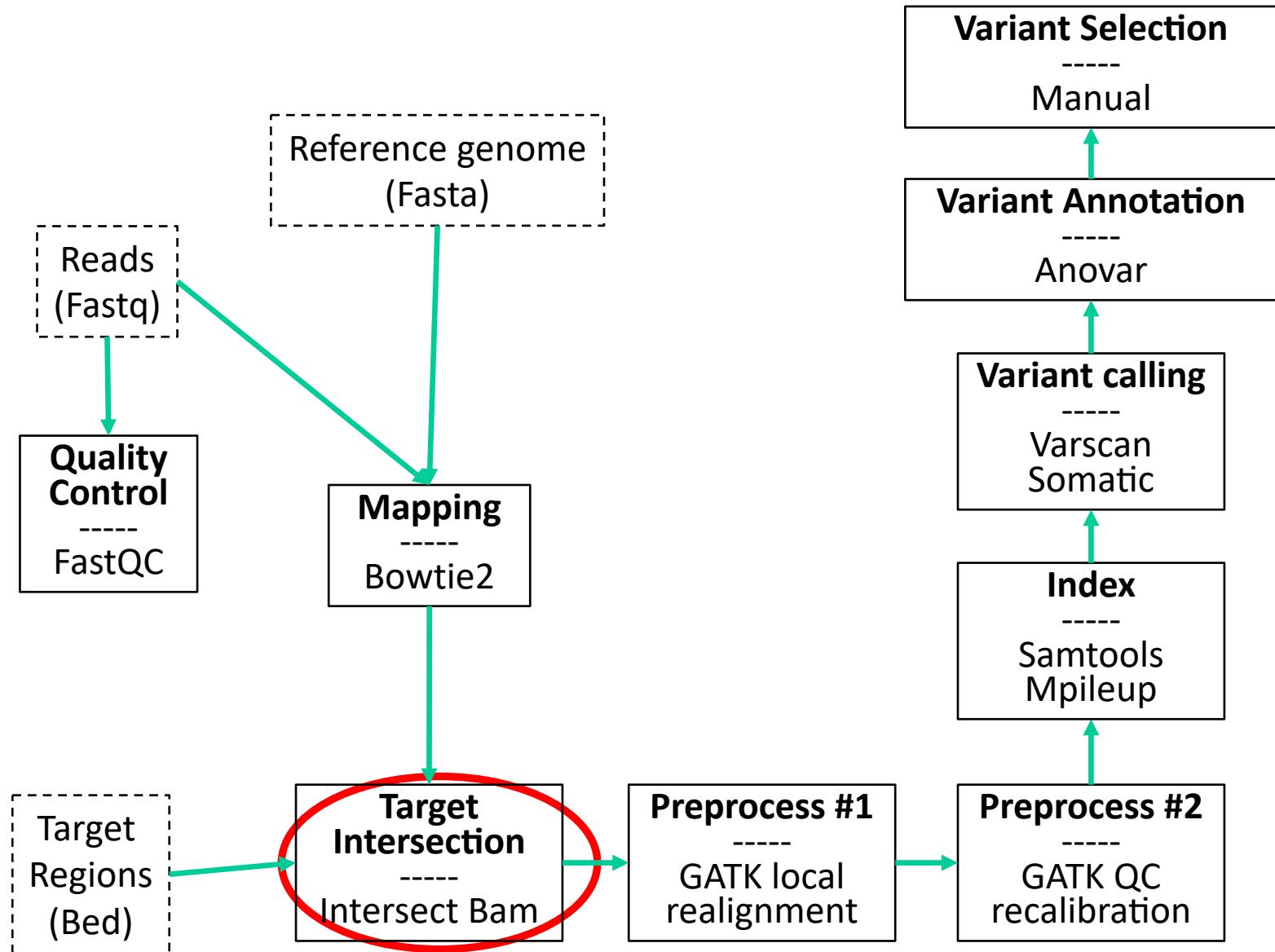
# Format SAM/BAM

- Dans la réalité:

```
@RG  ID:group1  SM:1425_CD34  PL:ILLUMINA  LB:lib1 PU:unit1
@PG  ID:bwa  PN:bwa  VN:0.7.12-r1039 CL:bwa mem -M -t 2 -A 2 -E 1 -R @RG\tID:group1\tSM:1425_CD34\tPL:ILLUMINA\tLB:lib1\tPU:unit1 /root/myd
ERR166338.13782800  83  chr13  32890449  60  101M  =  32890343  -207  GGGACTGAATTAGAACAAATTTCAGCGCTT
ERR166338.13782800  163  chr13  32890343  60  75M  =  32890449  207  CACTAGCCACGTTCGAGTGCTTAATGTGGCTAGTGGC
ERR166338.26716588  99  chr13  32890406  60  101M  =  32890553  222  AATGTTCCCACCTCACAGTAAGCTGTTACCGTTCCAG
ERR166338.26716588  147  chr13  32890553  60  75M  =  32890406  -222  TTGCAGACTTACCAAGCATTGGAGGAATATCGTAA
ERR166338.27259961  99  chr13  32890496  60  101M  =  32890558  137  ACCTCAGTCACATAATAAGGAATGCATCCCTGTGTAAG
ERR166338.27259961  147  chr13  32890558  60  75M  =  32890496  -137  GACTTATTTACCAAGCATTGGAGGAATATCGTAGGTAA
ERR166338.63037998  99  chr13  32890496  60  101M  =  32890558  137  ACCTCAGTCACATAATAAGGAATGCATCCCTGTGTAAG
ERR166338.63037998  147  chr13  32890558  60  75M  =  32890496  -137  GACTTATTTACCAAGCATTGGAGGAATATCGTAGGTAA
```

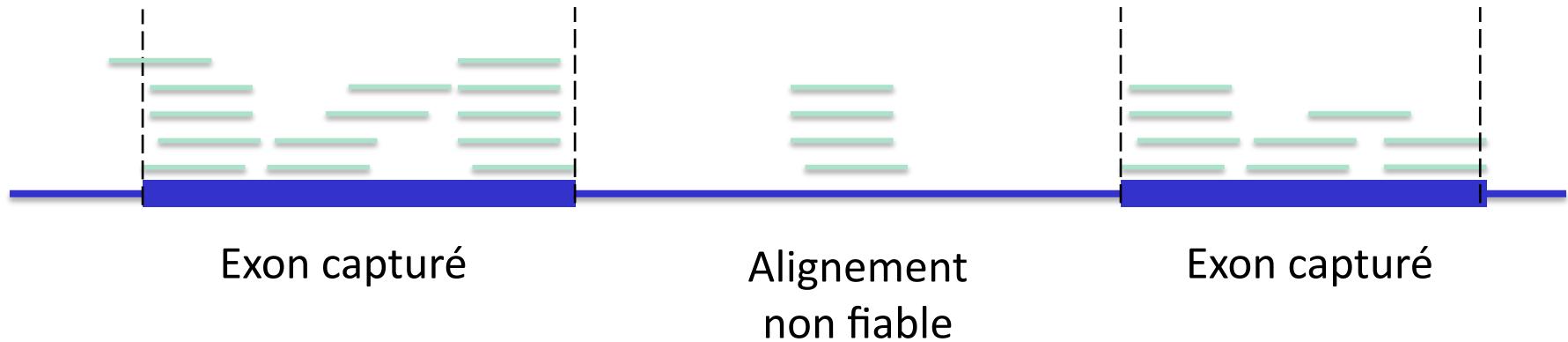
Diagram illustrating the structure of a SAM/BAM header line:

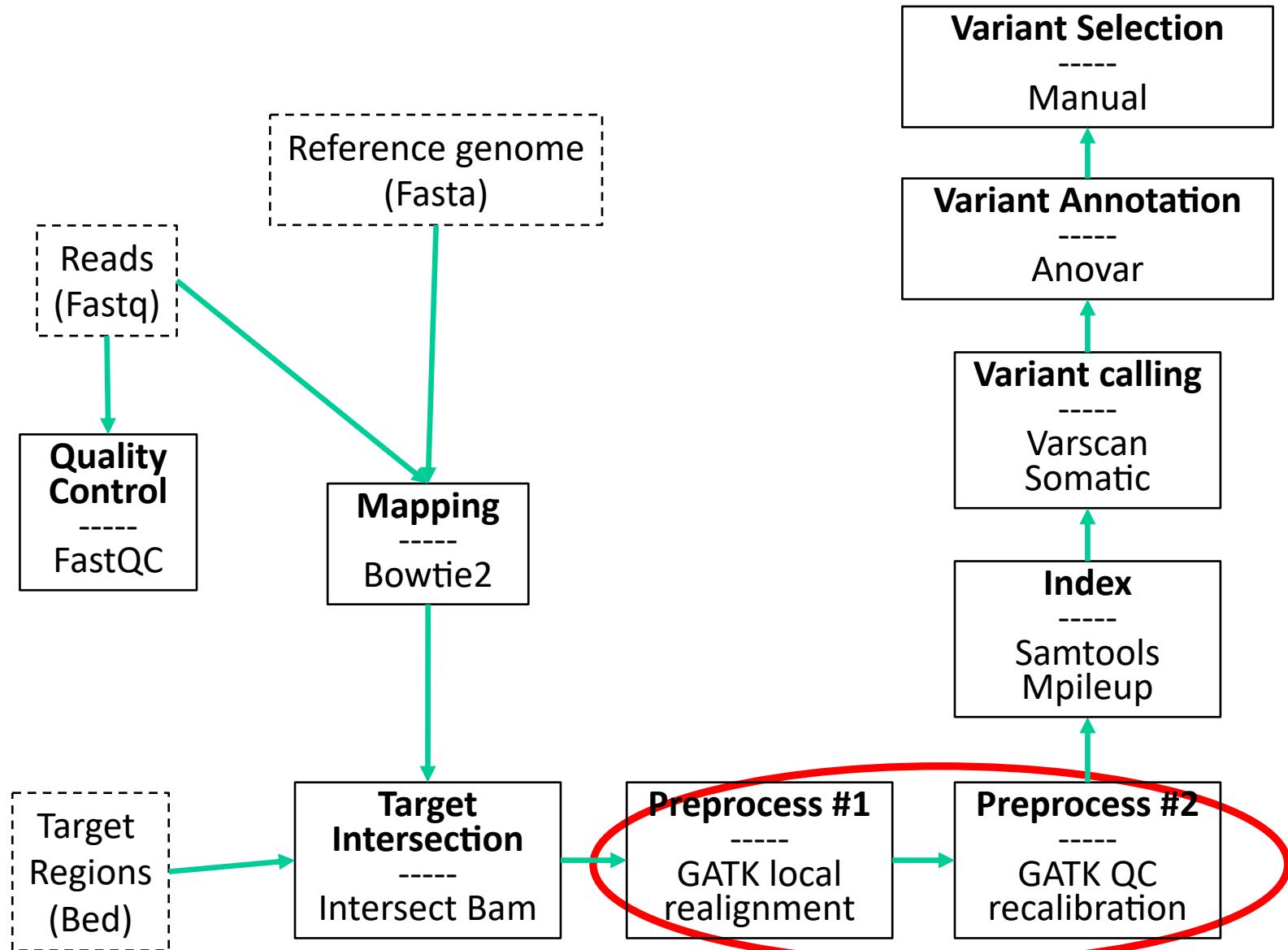
- read ID
- flag
- position
- CIGAR
- mapping qual.
- mate info



# Target intersection

- Comparer l'alignement obtenu à la liste des positions visées par le protocole de capture





# Why realign around indels ?

- Small Insertion/deletion (Indels) in reads (especially near the ends) can trick the mappers into wrong alignments
  - Alignment scoring – cheaper to introduce multiple Single Nucleotide Variants (SNVs) than an indel: induce a lot of false positive SNVs
- ➔ artifactual mismatches
- **Realignment around indels helps improve the downstream processing steps**

# Wrong alignment near indels

Genome

CTACGAAGTAAAAAAAAGAGAGAGTTACT

CTACGAAGT - -AAAAAAAAGAGAGAGTTACT

CTACGAAGTAAAAAAAAGAGAGAG**TTACT**

*Cost for 2 indels < 4 mismatches*

CTACGAAGT - -AAAAAAAAGAGAGA

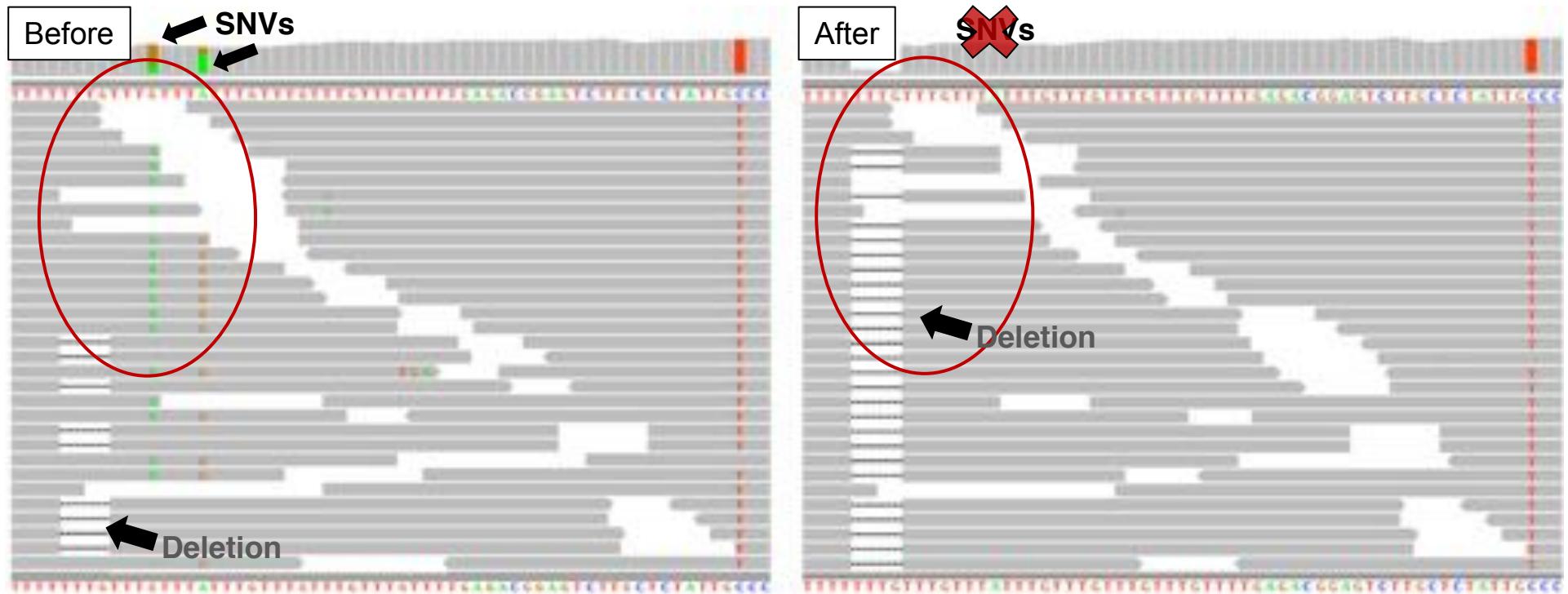
CTACGAAGTAAAAAAAAG**GAGAGA**

*Cost for 2 indels > 1 mismatch*

Read 1: 2 deletions

Read 2: 2 deletions

# Local realignment around indels



# Local realignment around indels



# Local realignment around indels



# Indel realignment in 2 steps

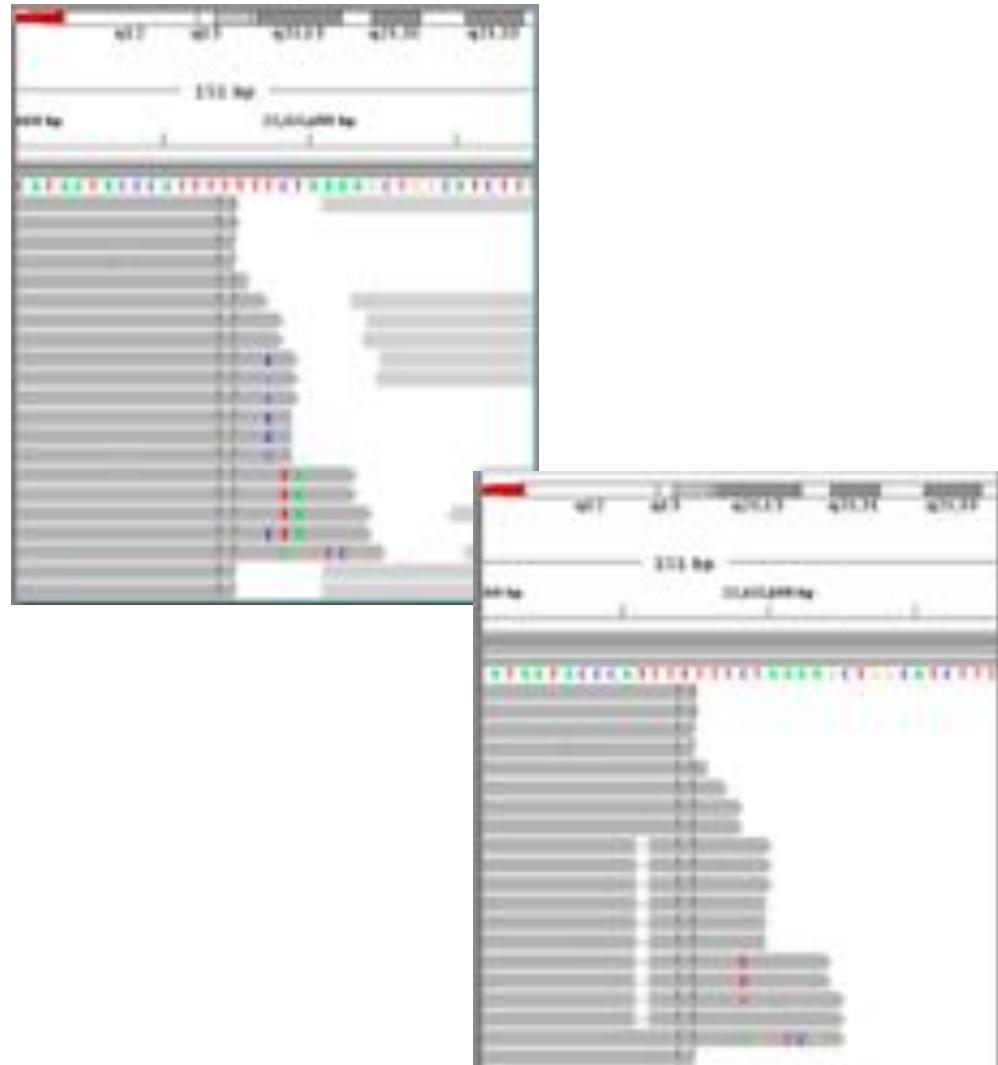
1. Identify what regions need to be realigned

- RealignerTargetCreator + known sites

Intervals  
↓

2. Perform the actual realignment (BAM output)

- IndelRealigner

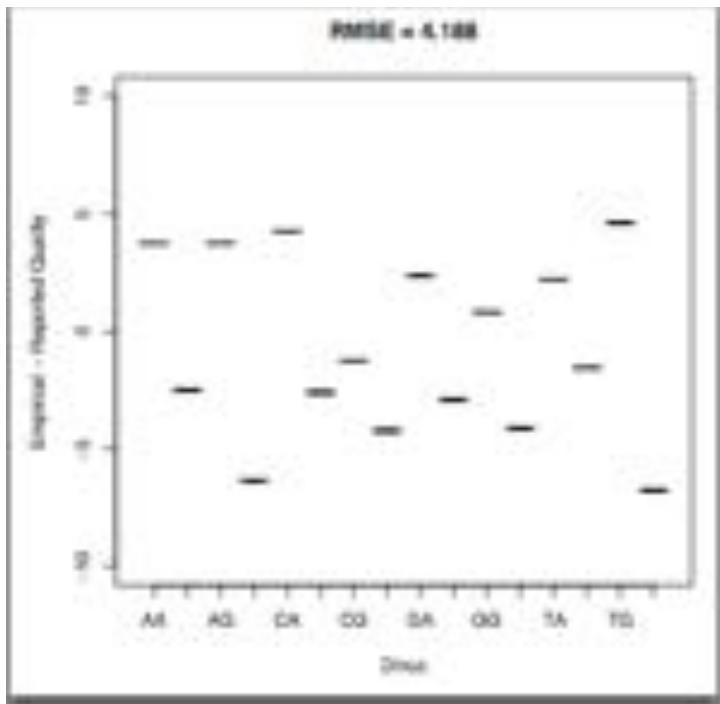


# Types of realignment targets

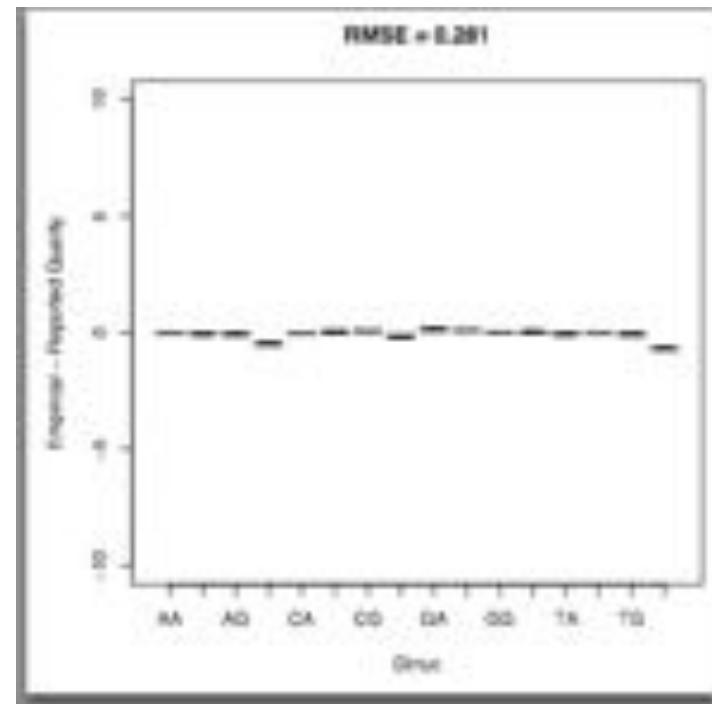
1. Indels seen in original alignments (in CIGAR, indicated by I for Insertion or D for Deletion)
2. Sites where evidences suggest a hidden indel (SNV abundance)
3. Known sites:
  - Common polymorphisms: dbSNP, 1000Genomes

# The quality scores issued by sequencers are biased

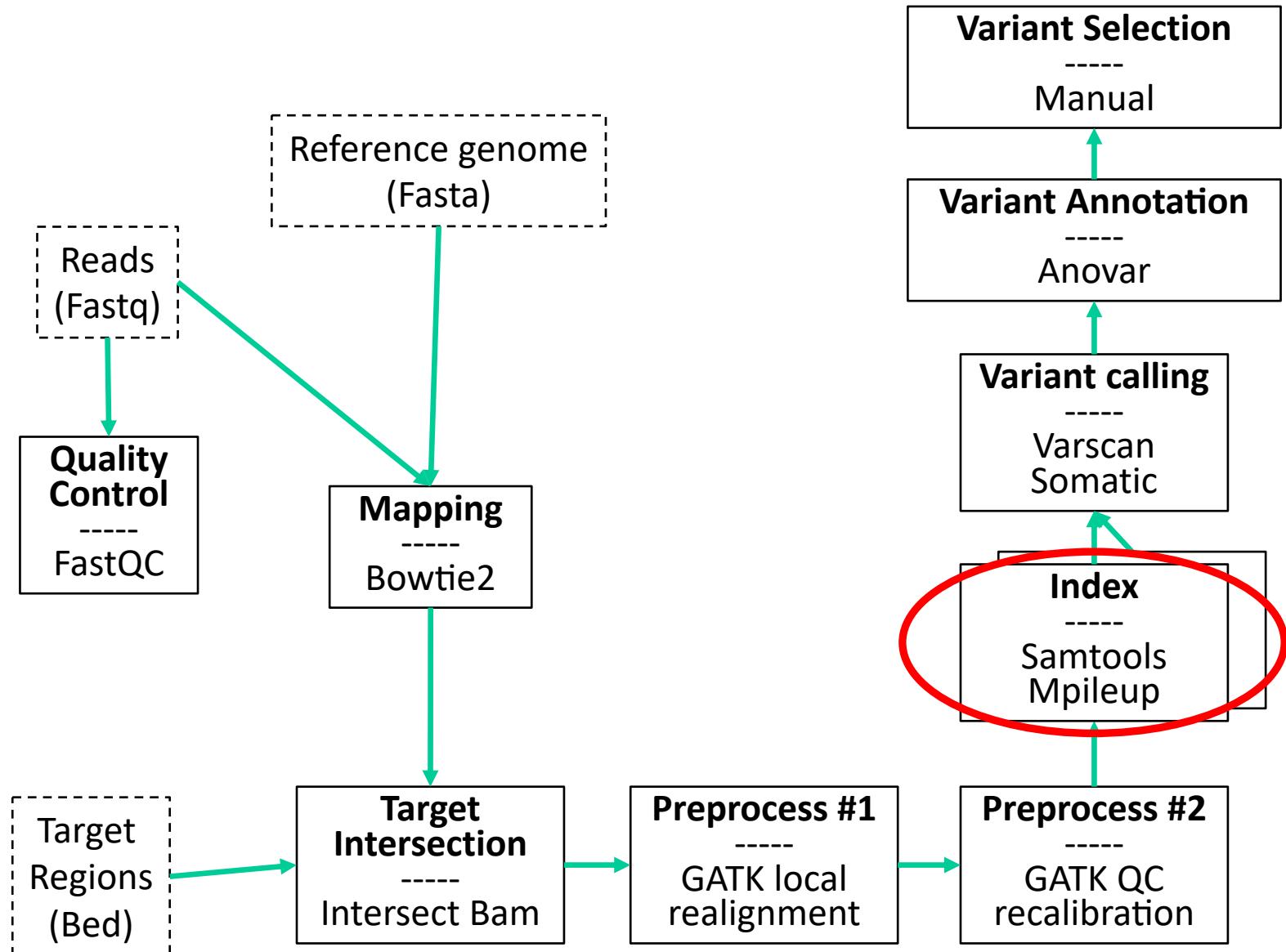
- Quality scores are critical for all downstream analysis
- Systematic biases are a major contributor to bad calls
- Example of sequence context bias in the reported qualities:



before



after



# PileUp:

# Pourquoi un nouveau format de fichier?

Rappel BAM:

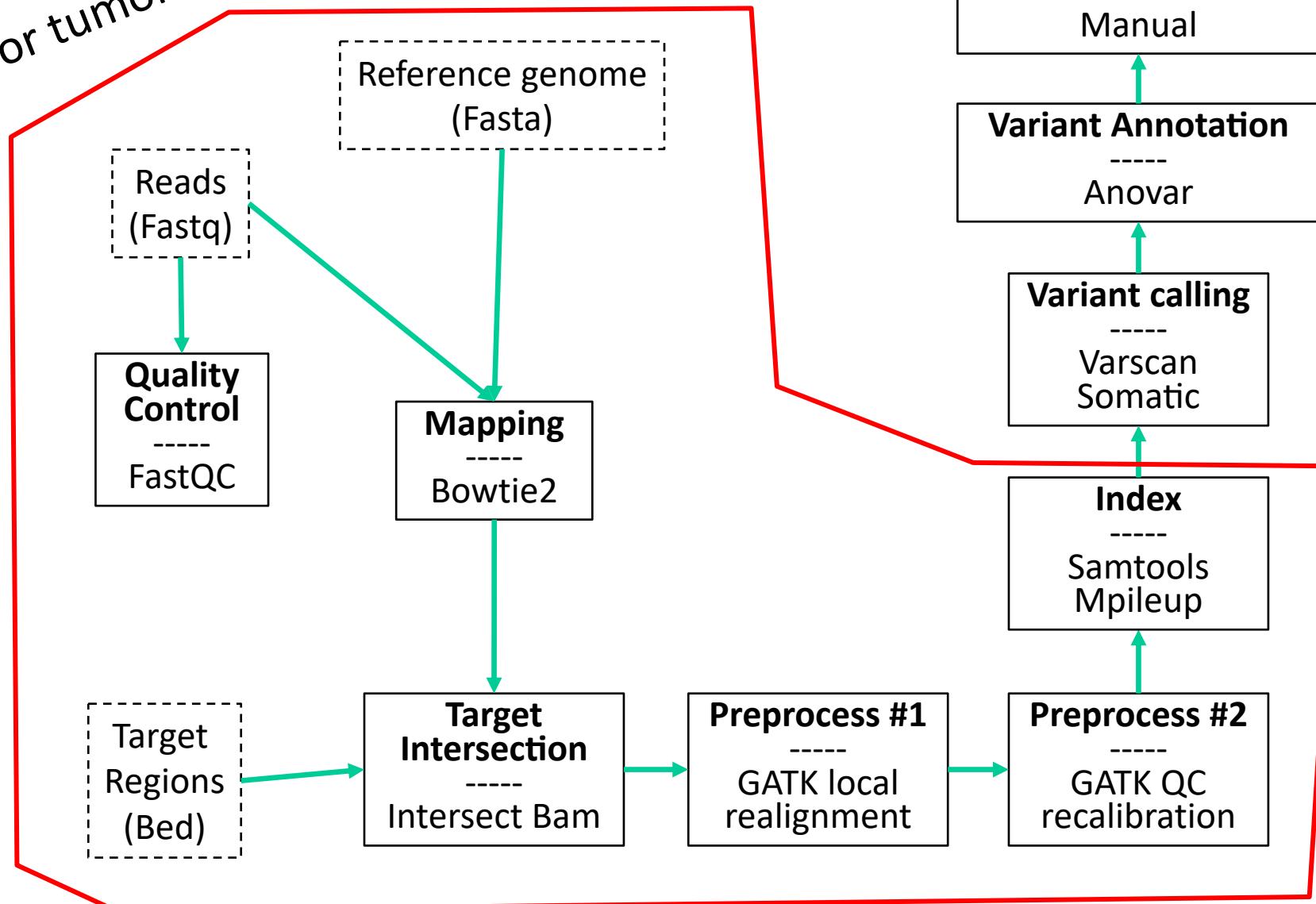
```
@RG  ID:group1  SM:1425_CD34  PL:ILLUMINA  LB:lib1 PU:unit1
@PG  ID:bwa  PN:bwa  VN:0.7.12-r1039 CL:bwa mem -M -t 2 -A 2 -E 1 -R @RG\tID:group1\tSM:1425_CD34\tPL:ILLUMINA\tLB:lib1\tPU:unit1 /root/myd
ERR166338.13782800  83  chr13  32890449  60  101M  =  32890343  -207  GGGACTGAATTAGAACAAATTTCAGCGCTT
ERR166338.13782800  163  chr13  32890343  60  75M  =  32890449  207  CACTAGCCACGTTCGAGTGCTTAATGTGGCTAGTGGC
ERR166338.26716588  99  chr13  32890406  60  101M  =  32890553  222  AATGTTCCCACCTCACAGTAAGCTGTTACCGTTCCAG
ERR166338.26716588  147  chr13  32890553  60  75M  =  32890406  -222  TTGCAGACTTACCAAGCATTGGAGGAATATCGTAA
ERR166338.27259961  99  chr13  32890496  60  101M  =  32890558  137  ACCTCAGTCACATAATAAGGAATGCATCCCTGTGTAAG
ERR166338.27259961  147  chr13  32890558  60  75M  =  32890496  -137  GACTTACCAAGCATTGGAGGAATATCGTAGGTAA
ERR166338.63037998  99  chr13  32890496  60  101M  =  32890558  137  ACCTCAGTCACATAATAAGGAATGCATCCCTGTGTAAG
ERR166338.63037998  147  chr13  32890558  60  75M  =  32890496  -137  GACTTACCAAGCATTGGAGGAATATCGTAGGTAA
```

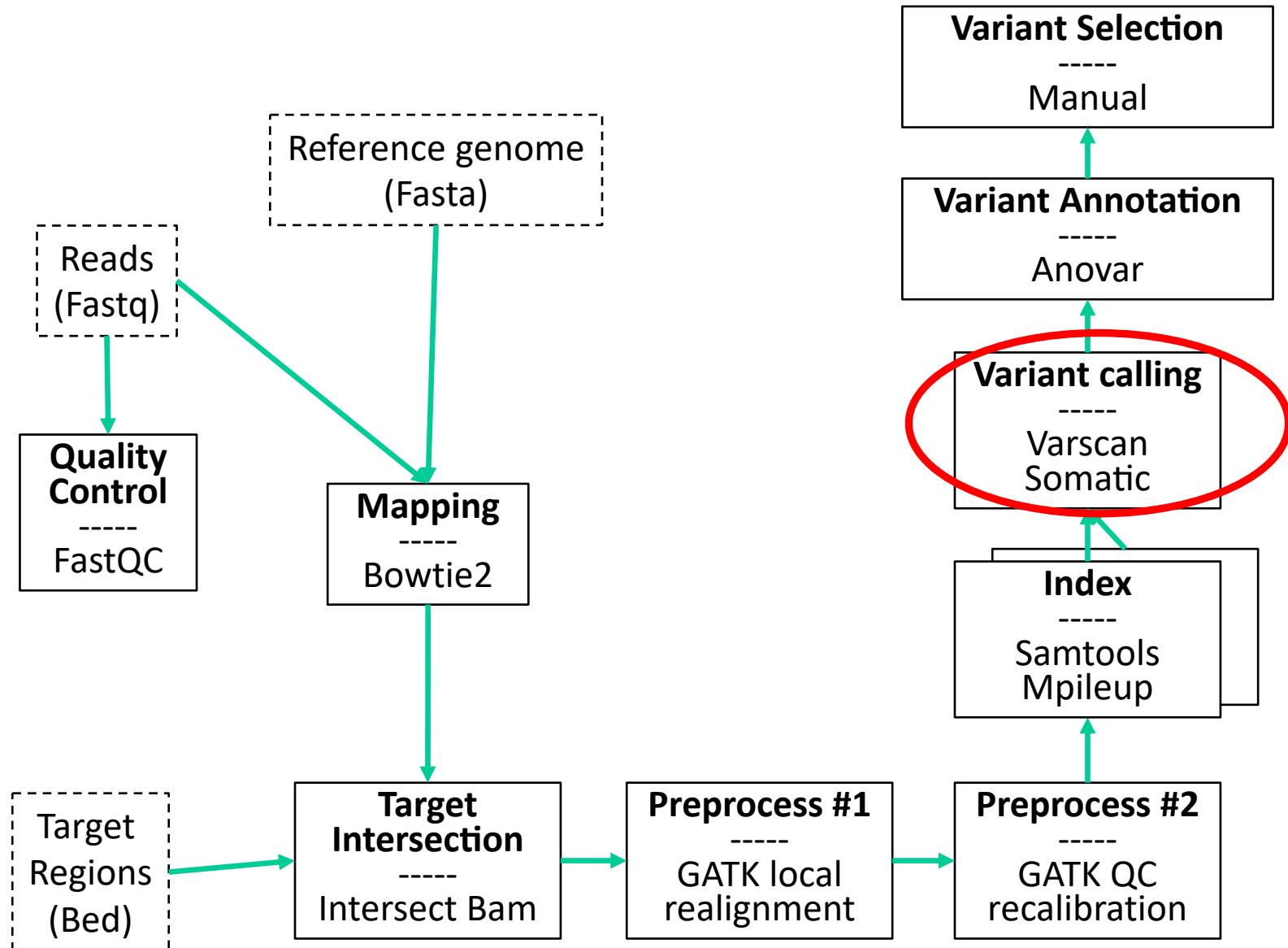
# Pileup format

# Pileup format

Describes base-pair information at each position

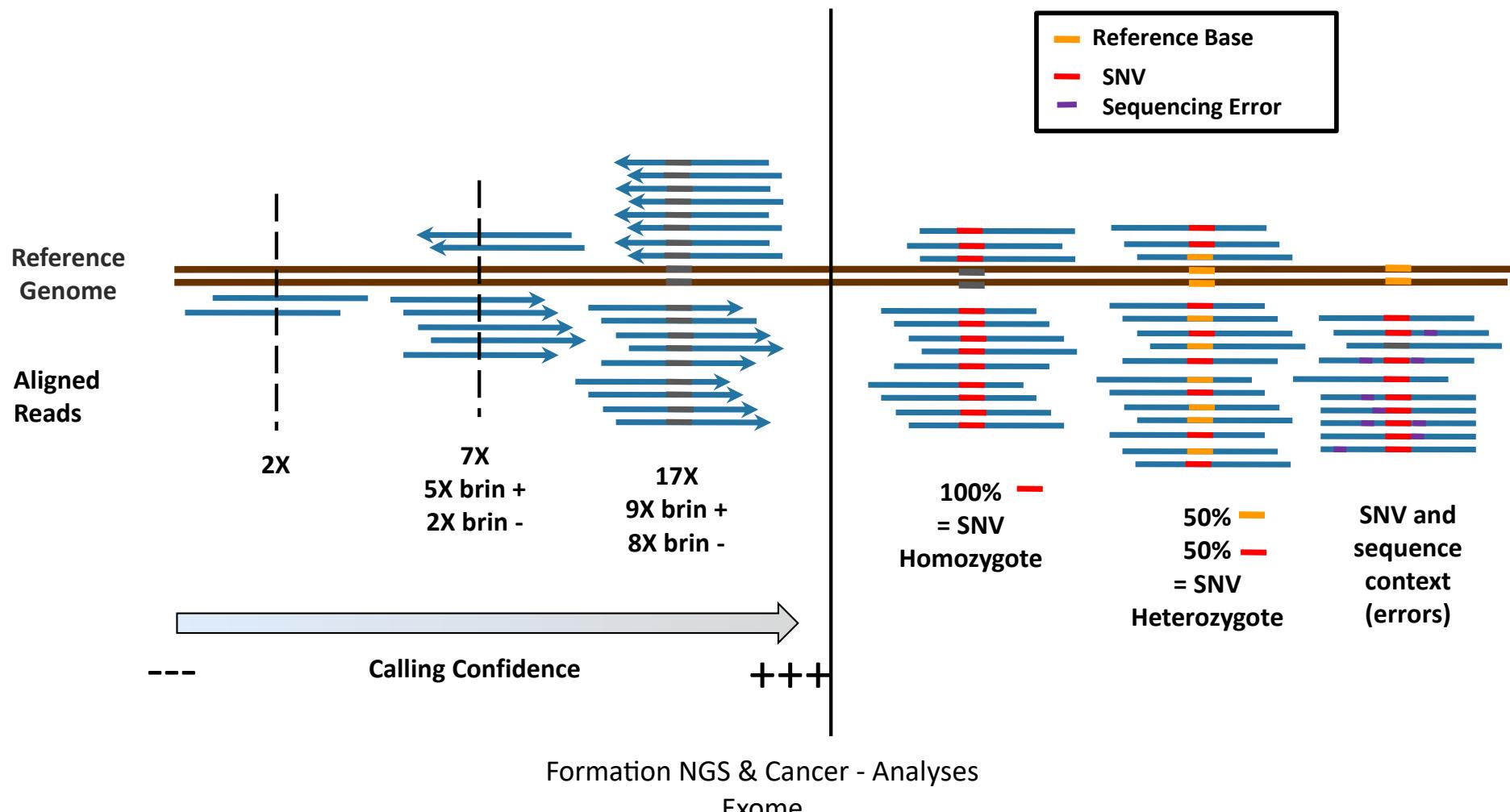
X2 for tumor & normal





# Variant calling criteria

Depth of Coverage = number of reads supporting one position  
ex: 1X, 5X, 100X... >1000X



# Factors to consider for Variant Calling

- Calling a SNV:
  - Base call qualities of each supporting base (base quality)
  - Proximity to small indels, or homopolymer run
  - Mapping qualities of the reads supporting the SNP
  - Sequencing **depth**: >=30x for constit ; >=100 for tumor
  - SNVs position within the reads: Higher error rate at the reads ends
  - Look at strand bias (SNVs supported by only one strand are more likely to be artifactual)
  - **Allelic frequency**: Tumor cellularity will reduce the % of an heterozygous variant
- Calling an indel:
  - Higher stringency (and Sanger validation often needed)

# VarScan2

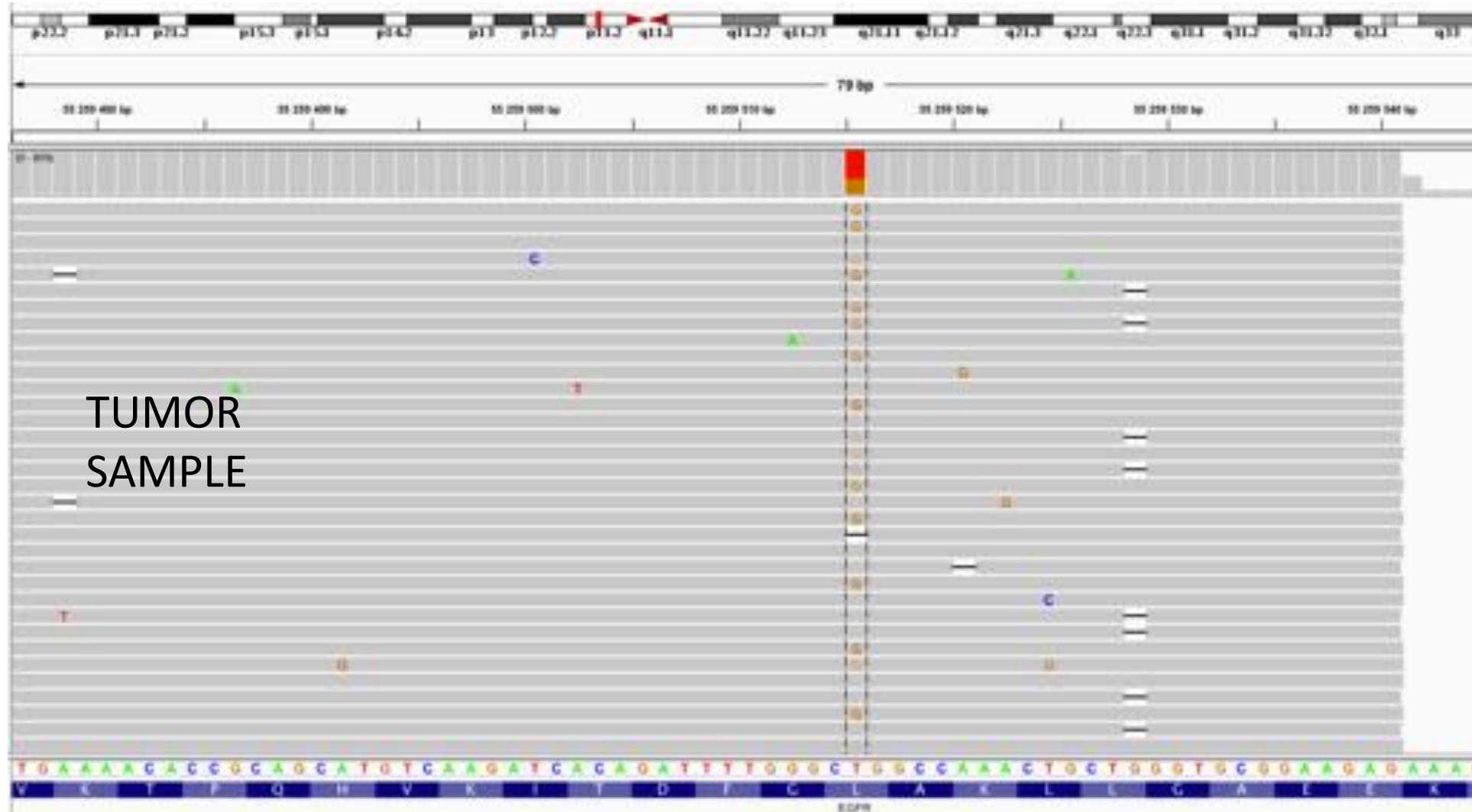
- Mutation caller written in **Java** (no installation required) working with **Pileup files** of Targeted, Exome, and Whole-Genome sequencing data (DNAseq or RNAseq)
- **Multi-platforms:** Illumina, SOLiD, Life/PGM, Roche/454
- Detection of different kinds of Germline SNVs/Indels (classical mode):
  - Variants in individual samples
  - Multi-sample variants **shared or private** in multi-sample datasets
- VarScan specificity is to be able to work with **Tumor/Normal pairs (somatic mode):**
  - Somatic and germline mutation, LOH events in tumor-normal pairs
  - Somatic copy number alterations (CNAs) in tumor-normal exome data

# VarScan2 Performance

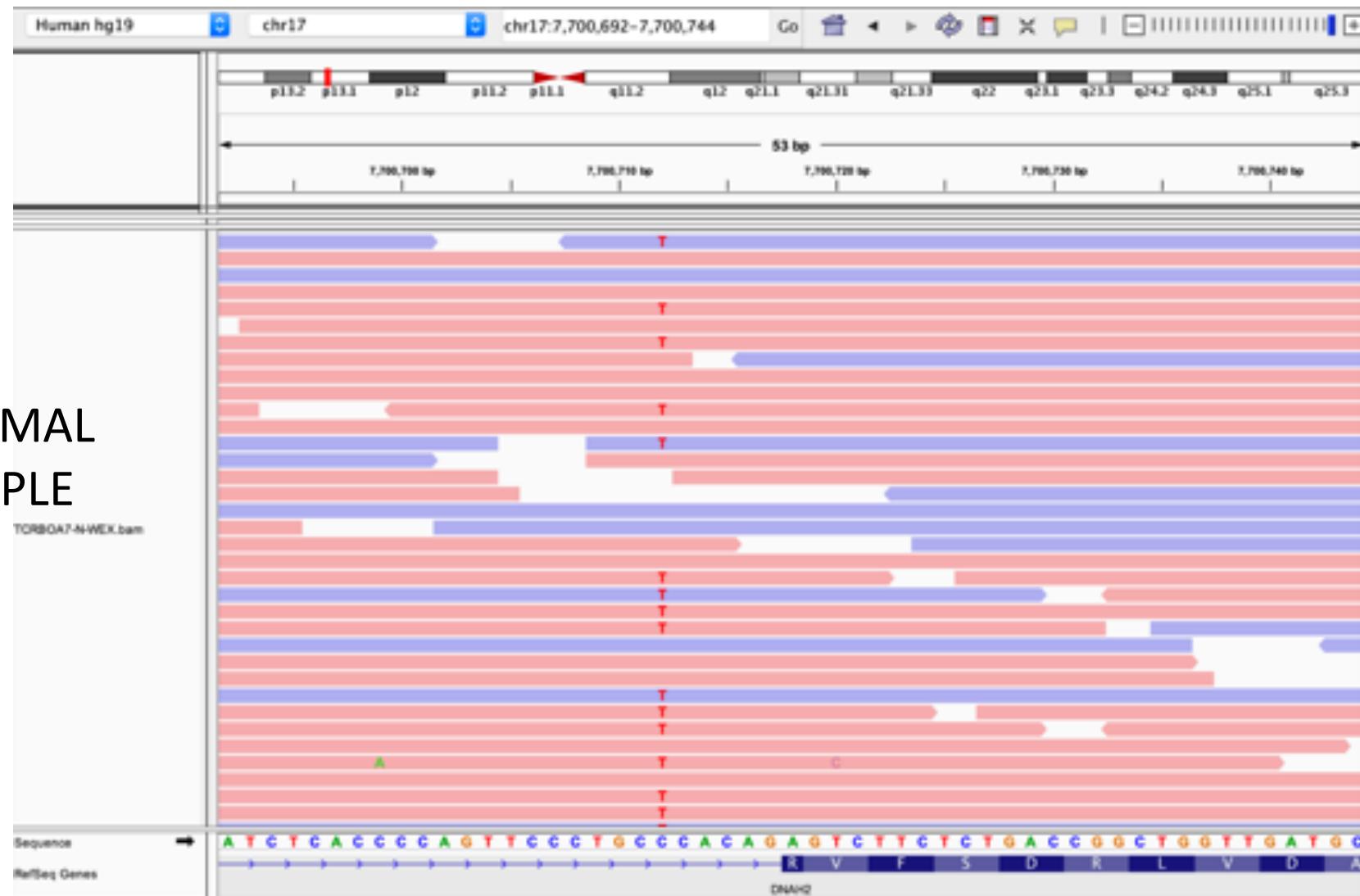
- VarScan uses a robust **heuristic/statistic** approach to call variants that meet desired thresholds for read depth, base quality, variant allele frequency, and statistical significance
- Stead *et al.* (2013) compared 3 different **somatic callers** : MuTect, Strelka, VarScan2
  - **VarScan2 performed best** overall with sequencing depths of 100x, 250x, 500x and 1000x required to accurately identify variants present at 10%, 5%, 2.5% and 1% respectively
- Other widely used tool: **GATK**

# Somatic calling

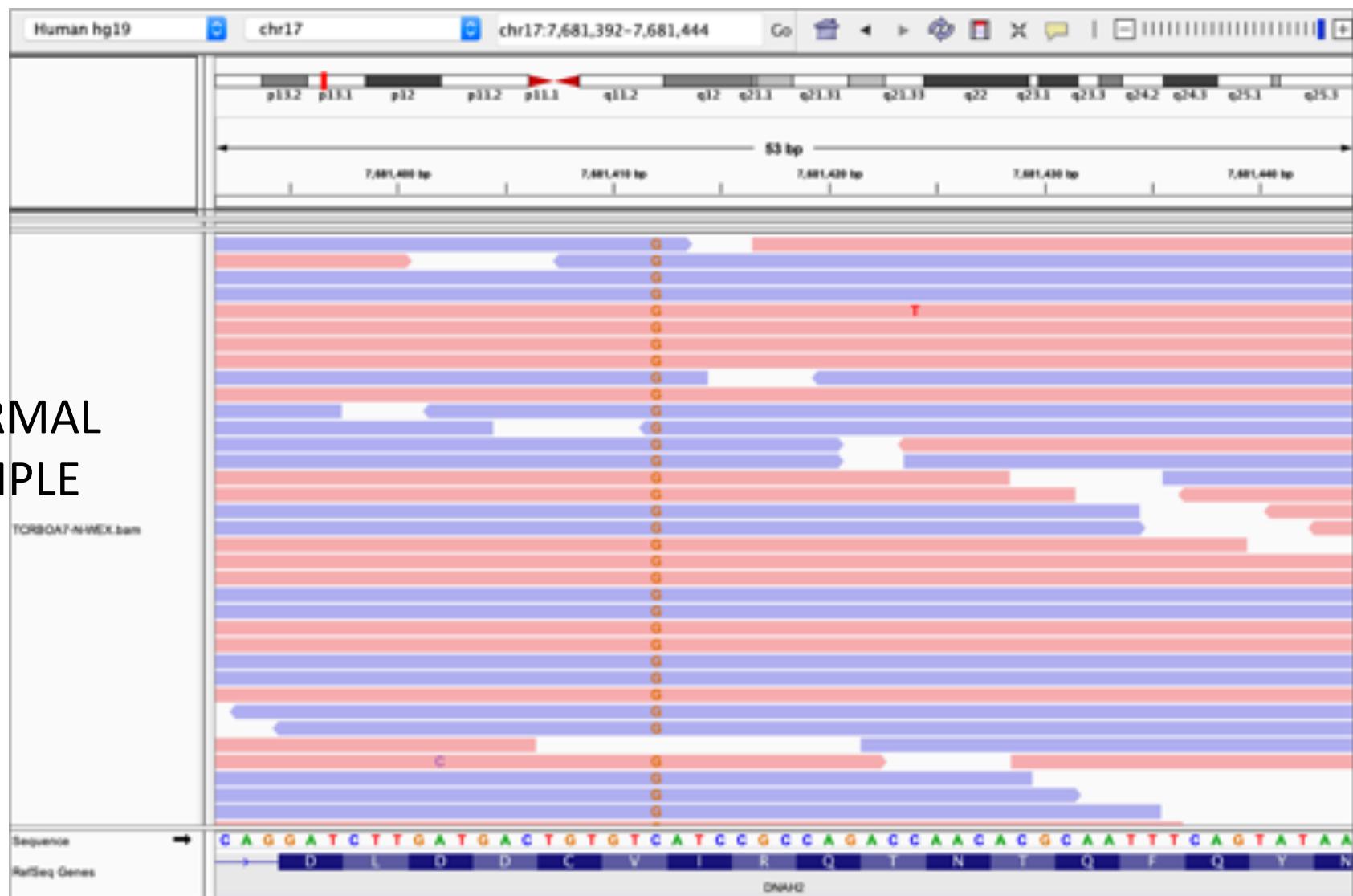
# Un variant tumoral: polymorphisme ou mutation?



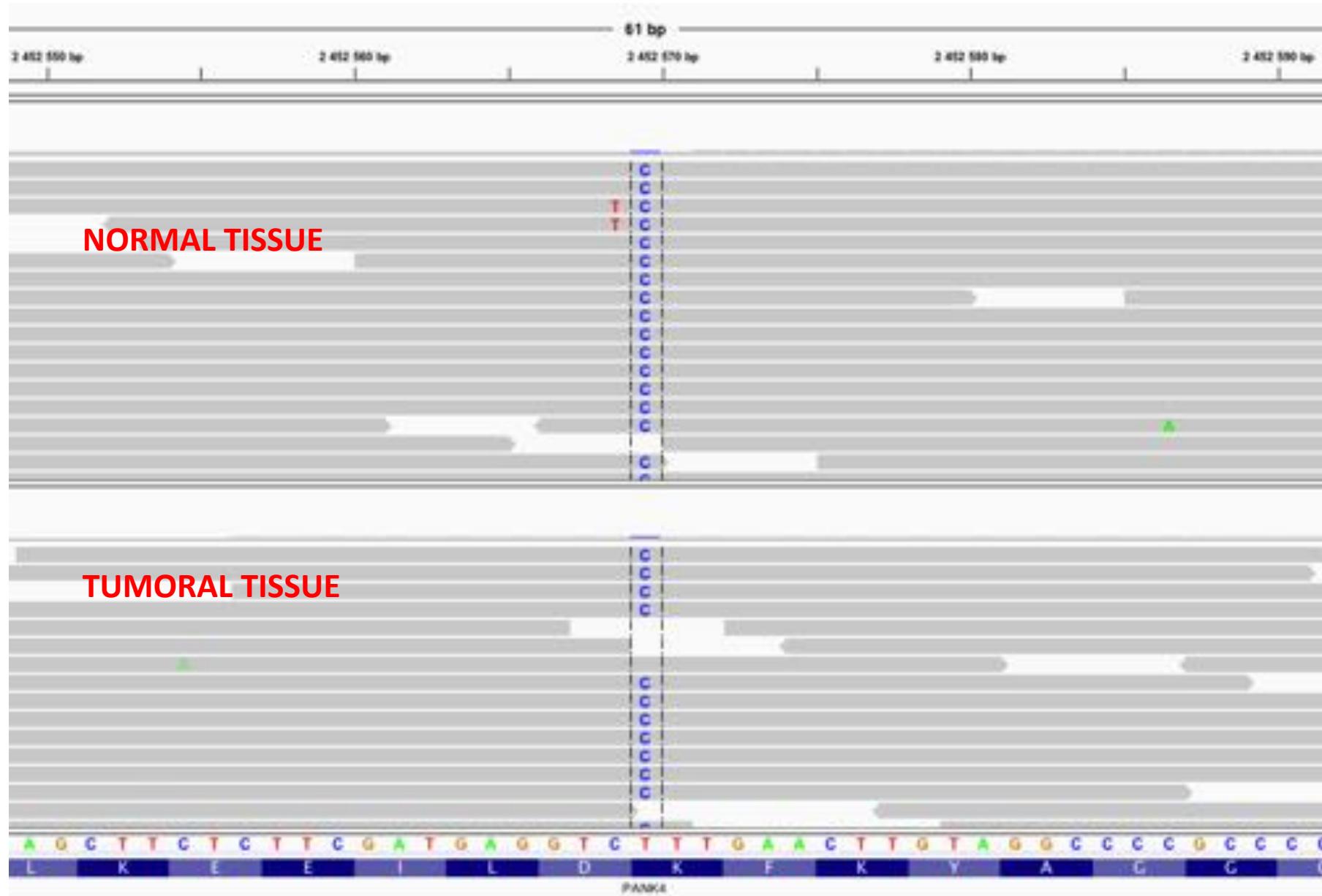
# Un polymorphisme (SNP) hétérozygote



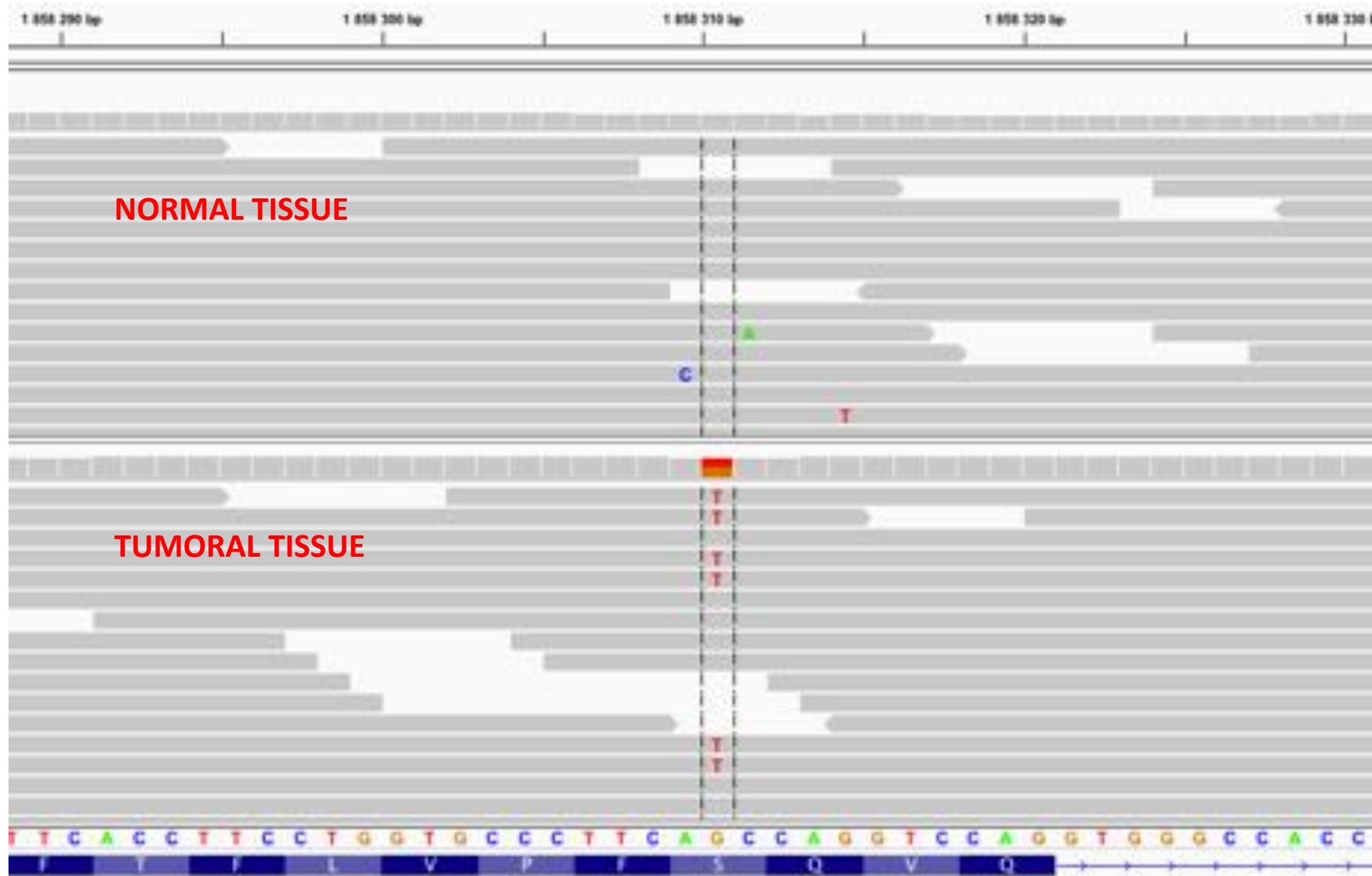
# Un polymorphisme (SNP) homozygote



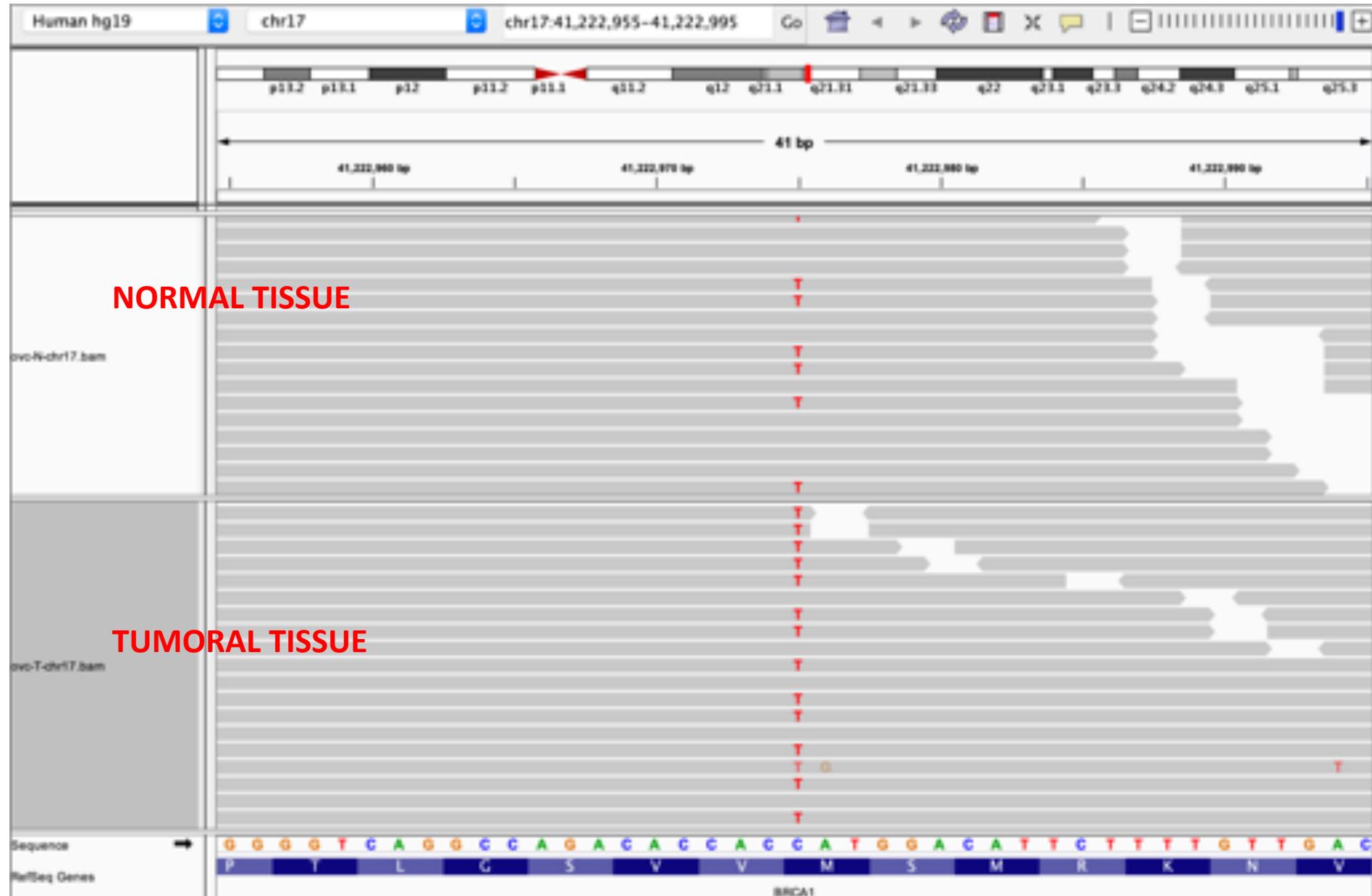
# Un polymorphisme vu dans N et T



# Une mutation somatique



# Une LOH (loss of heterozygosity)



# Varscan's Somatic P-value

## Variant Calling and Comparison

At every position where both normal and tumor have sufficient coverage, a comparison is made. First, normal and tumor are called independently using the germline consensus calling functionality. Then, their genotypes are compared by the following algorithm:

Calculate significance of allele frequency difference by Fisher's Exact Test

**If difference is significant (p-value < threshold):**

If normal matches reference

==> Call Somatic

Else If normal is heterozygous

==> Call LOH

Else normal and tumor are variant, but different

==> Call IndelFilter or Unknown

**If difference is not significant:**

==> Call Germline

The diagram illustrates the logic for determining if a variant is somatic or LOH based on genotype counts in normal (N) and tumor (T) samples.

**Alleles**

	Ref	Var
N	8	0
T	6	7

**Somatic Call:** A red arrow points from the table to the text "Somatic".

	4	4
N	4	4
T	8	1

**LOH Call:** A red arrow points from the table to the text "LOH".

# Format VCF

```

##fileformat=VCFv4.2
##fileDate=20090805
##source=myImputationProgramV3.1
##reference=file:///seq/references/1000GenomesPilot-NCBI36.fasta
##contig=<ID=20,length=62435964,assembly=B36,md5=f126cdf8a6e0c7f379d618ff66beb2da,species="Homo sapiens",taxonomy=x>
##phasing=partial
##INFO<ID=NS,Number=1,Type=Integer,Description="Number of Samples With Data">
##INFO<ID=DP,Number=1,Type=Integer,Description="Total Depth">
##INFO<ID=AF,Number=A,Type=Float,Description="Allele Frequency">
##INFO<ID=AA,Number=1,Type=String,Description="Ancestral Allele">
##INFO<ID=DB,Number=0,Type=Flag,Description="dbSNP membership, build 129">
##INFO<ID=H2,Number=0,Type=Flag,Description="HapMap2 membership">
##FILTER<ID=q10,Description="Quality below 10">
##FILTER<ID=s50,Description="Less than 50% of samples have data">
##FORMAT<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
##FORMAT<ID=DP,Number=1,Type=Integer,Description="Read Depth">
##FORMAT<ID=HQ,Number=2,Type=Integer,Description="Haplotype Quality">
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT NA00001 NA00002 NA00003
20 14370 rs6064257 G A 29 PASS NS=3;DP=14;AF=0.5;DB;H2 GT:GQ:DP:HQ 0|0:48:1:51,51 1|0:48:8:51,51 1/1:43:5:...
20 17330 . T A 3 q10 NS=3;DP=11;AF=0.017 GT:GQ:DP:HQ 0|0:49:3:58,50 0|1:1:3:5:65,3 0/0:41:3
20 1110696 rs6040355 A G,T 67 PASS NS=2;DP=10;AF=0.333,0.667;AA=T;DB GT:GQ:DP:HQ 1|2:21:6:23,27 2|1:2:0:18,2 2/2:36:4
20 1230237 . T . 47 PASS NS=3;DP=13;AA=T GT:GQ:DP:HQ 0|0:54:7:56,60 0|0:48:4:51,51 0/0:61:2
20 1234567 microsat1 GTC G,GTCT 50 PASS NS=3;DP=9;AA=G GT:GQ:DP 0/1:35:4 0/2:17:2 1/1:40:3

```

mandatory

Optional header: meta-data about available annotation

mandatory

samples

deletion

Insertion  
(2 events here)

NS: number of samples with data  
DP: combined depth  
AF: allelic fraction  
AA: ancestral allele

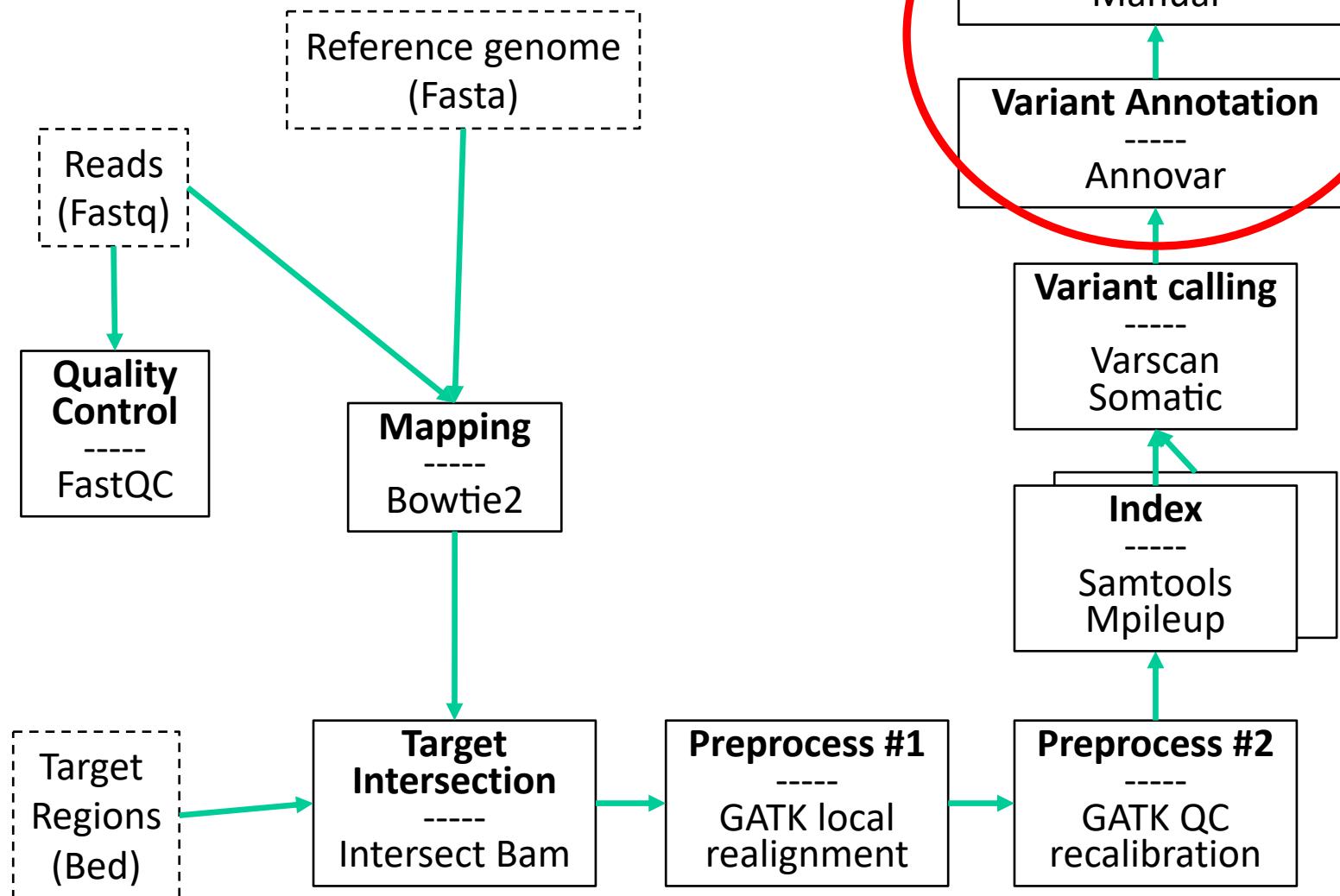
GT: genotype (0=ref, 1=alt)  
GQ: genotype quality  
DP: read depth

# Alternative: VarScan Tabulated Format

Chrom	Position	Ref	Cons	Reads1	Reads2	VarFreq	Strands 1	Strands 2	Qual1	Qual2	Pvalue	Map Qual1	Map Qual2	R1 +	R1 -	R2 +	Rs2 -	Alt
chr12	113348849	C	Y	31	30	49.18%	2	2	27	27	0.98	1	1	19	12	25	5	T
chr12	113354329	G	R	72	2	2.70%	2	2	31	26	0.98	1	1	48	24	1	1	A
chr12	113357193	G	A	2	72	97.30%	1	2	28	24	0.98	1	1	2	0	45	27	A
chr12	113357209	G	A	0	77	100%	0	2	0	29	0.98	0	1	0	0	51	26	A

**Cons** : Consensus Genotype of Variant Called (IUPAC code):

M -> A or C	Y -> C or T	D -> A or G or T	W -> A or T	V -> A or C or G
R -> A or G	K -> G or T	B -> C or G or T	S -> C or G	H -> A or C or T



# Different types of SNVs

- SNVs and short indels are the most frequent events:
  - Intergenic
  - Intronic
  - *cis*-regulatory
  - splice sites
  - frameshift or not
  - synonymous or not
  - benign or damaging etc...
- Example of SNV one want to pinpoint:
  - non-synonymous + highly deleterious + somatically acquired

# Resources dedicated to human genetic variation

- dbSNP and 1000-genomes
  - Population-scale DNA polymorphisms
- COSMIC
  - Catalogue Of Somatic Mutations In Cancer
- Non synonymous SNVs predictions
  - SIFT, Polyphen2 (damaging impact)... PhyloP, GERP++ (conservation)

# Annovar

« Annovar » annotates SNVs and Indels

Takes Multi sample VCF (Tumor &+normal samples)

- RefGene: Gene & Function & AminoAcid Change (HGVS format:  
c.A155G ; p.Lys45Arg)
- 1000g2012apr\_all: Minor Allele Frequency for all ethnies
- ESP6500: Exome Sequencing Project
- Ljb\_all : predictions (**SIFT**, **Polyphen2**, LRT, MutationTaster, PhyloP, GERP++)

❖ Tabulated file

Chr	Start	End	Ref	Alt	Func.refGene	Gene.refGene	ExonicFunc.refGene	AAChange.refGene	1000g2012apr_all	snp137	cosmic68	esp6500_all	LJB_PhylоП	LJB_PhylоП_Pre
chr1	160251792	160251792	A	G	intronic		PEX19		NA		NA			
chr1	167082869	167082869	G	A	intronic		DUSP27		NA		NA			
chr1	167095163	167095163	G	C	exonic		DUSP27		nonsynonymous SNV	DUSP27:NM_001080426:exon5:c.G795C:p.E265D				
chr1	167095881	167095881	G	A	exonic		DUSP27		nonsynonymous SNV	DUSP27:NM_001080426:exon5:c.G1513A:p.A505T				
chr1	167097739	167097739	C	A	exonic		DUSP27		nonsynonymous SNV	DUSP27:NM_001080426:exon5:c.C3371A:p.T1124N				
chr1	214803969	214803969	G	C	exonic		CENPF		nonsynonymous SNV	CENPF:NM_016343:exon9:c.G1287C:p.K429N				

Formation NGS & Cancer - Analyses  
Exome

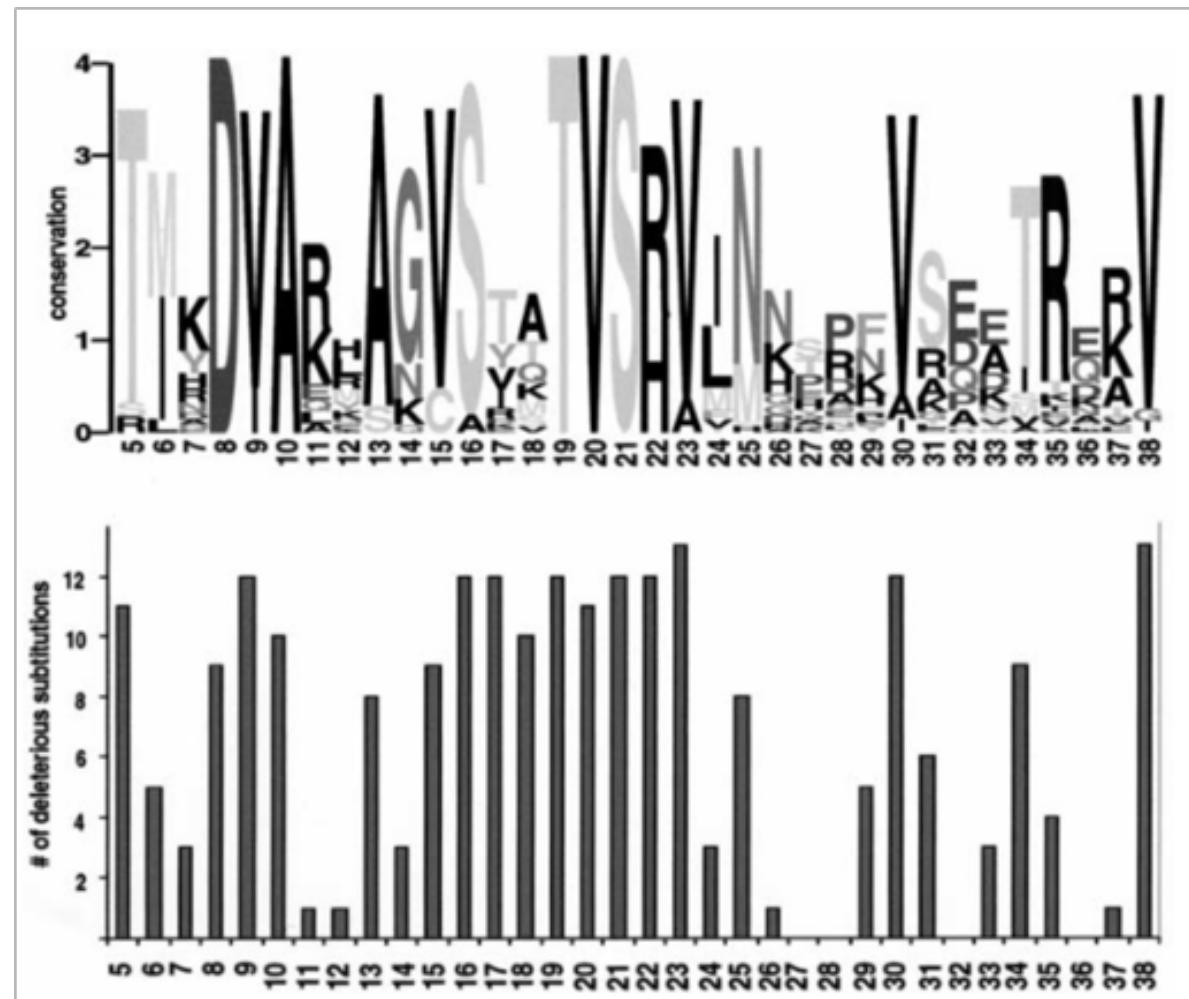


# Sorting Intolerant From Tolerant

Ng & Henikoff,  
Genome Res. 2001

Utilise la  
conservation des  
domaines protéiques  
comme indication du  
caractère délétère  
d'une substitution

Classe en  
**D**(eleterious),  
**T**(olerant),  
. (unknown)

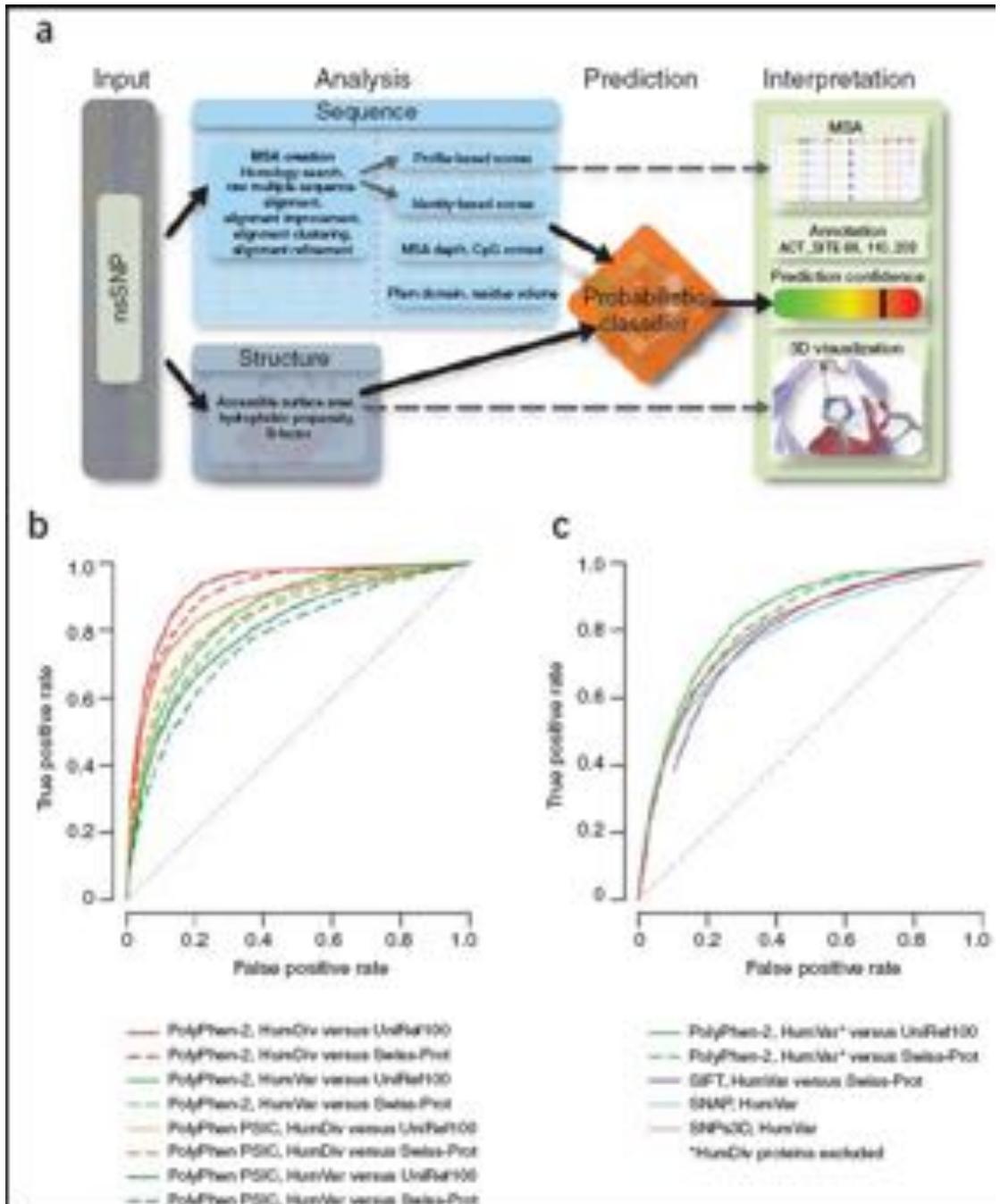


# PolyPhen2

Adzhubei et al. *Nature Methods* 2010.

**Probabilistic classifier:**  
Estimates the probability  
of the missense  
mutation being  
damaging based on a  
combination of  
seq+struct properties.

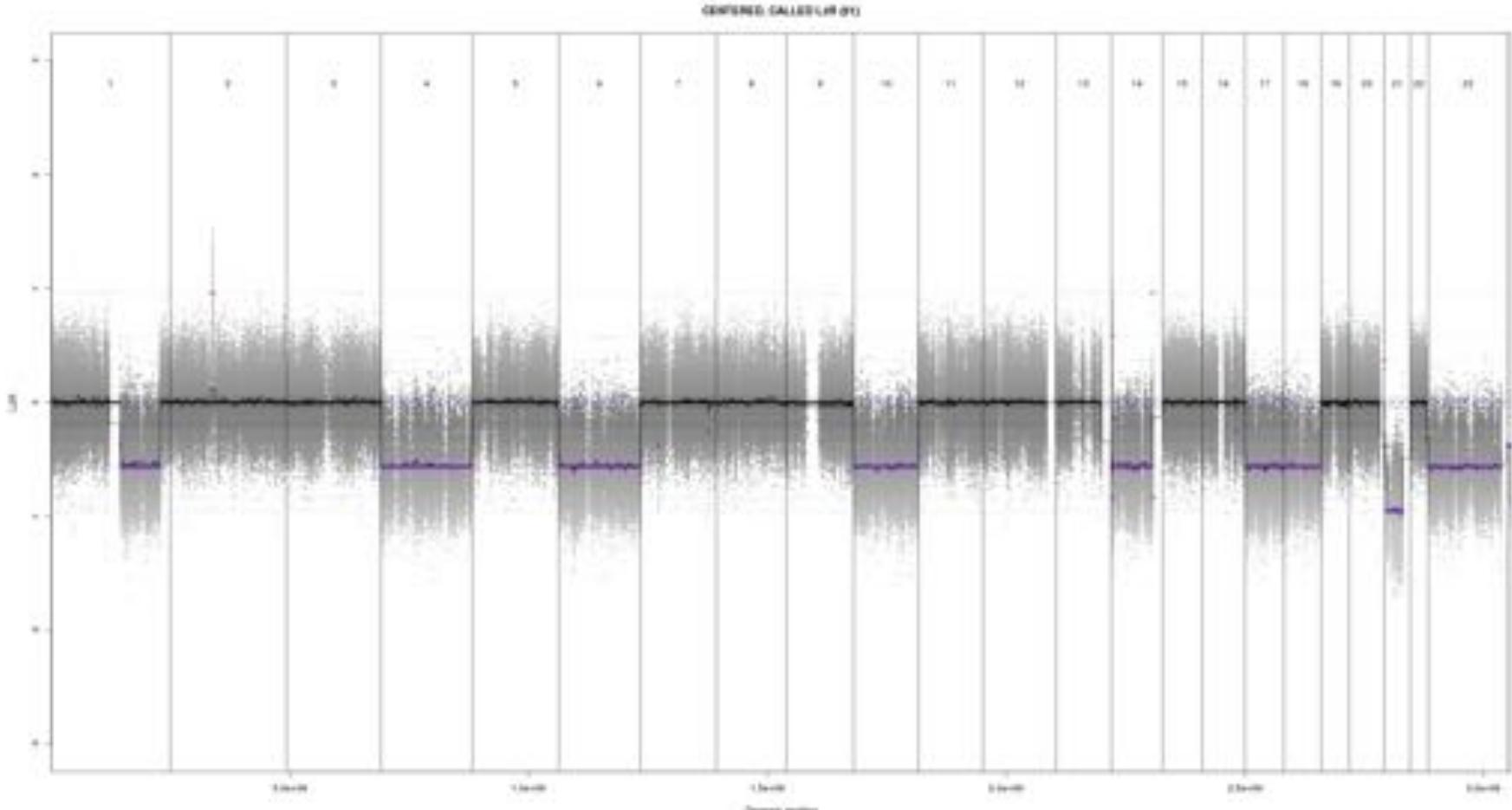
Classe en: **Benign**,  
**Possibly damaging**, or  
probably **Damaging**



# Coverage & Allelic Frequencies For CNV detection

# Detection of copy-number variations

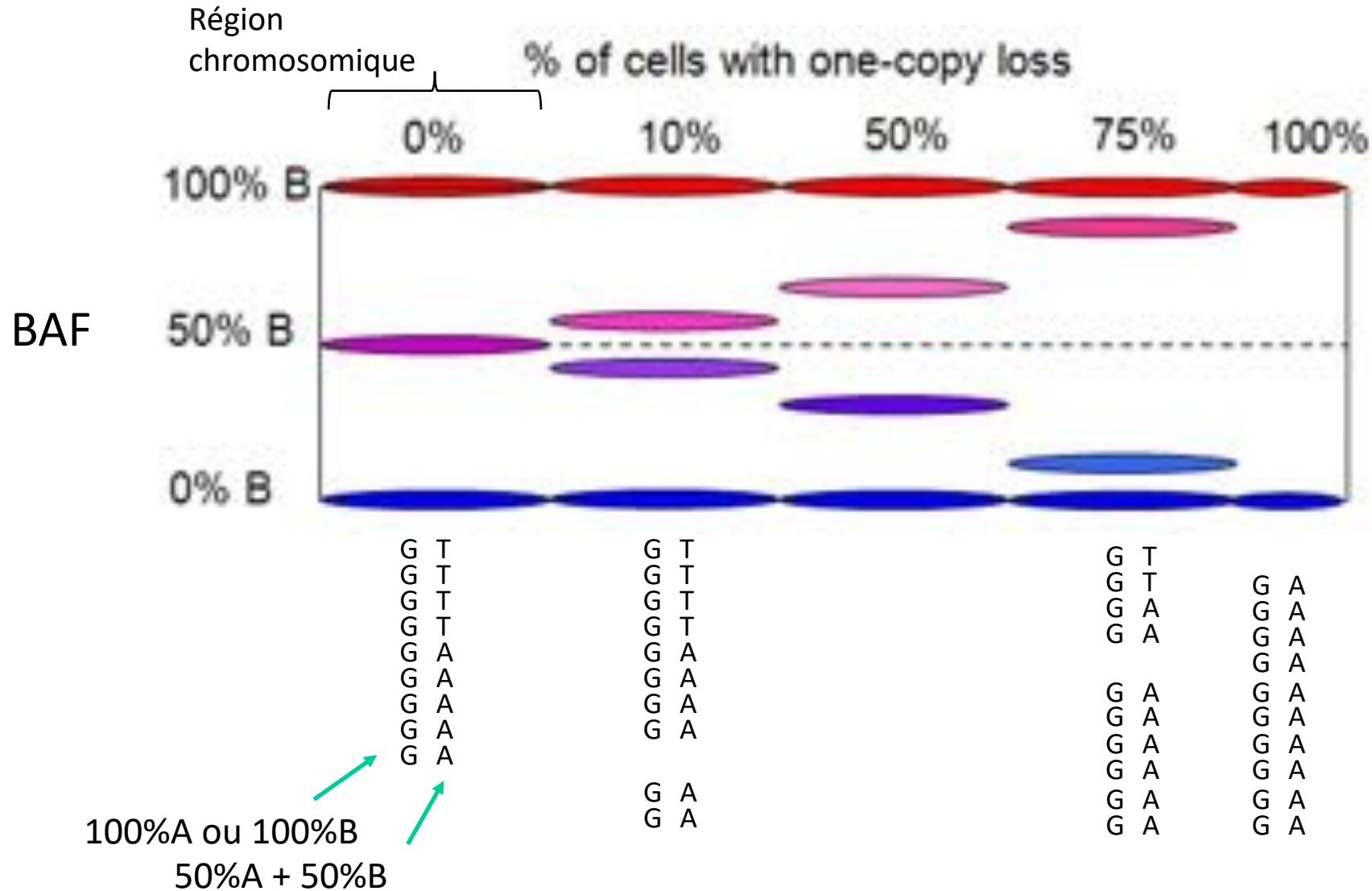
Are there any copy-number alteration (gain or loss of chromosomal regions, amplifications ...) that could explain tumorigenesis ?



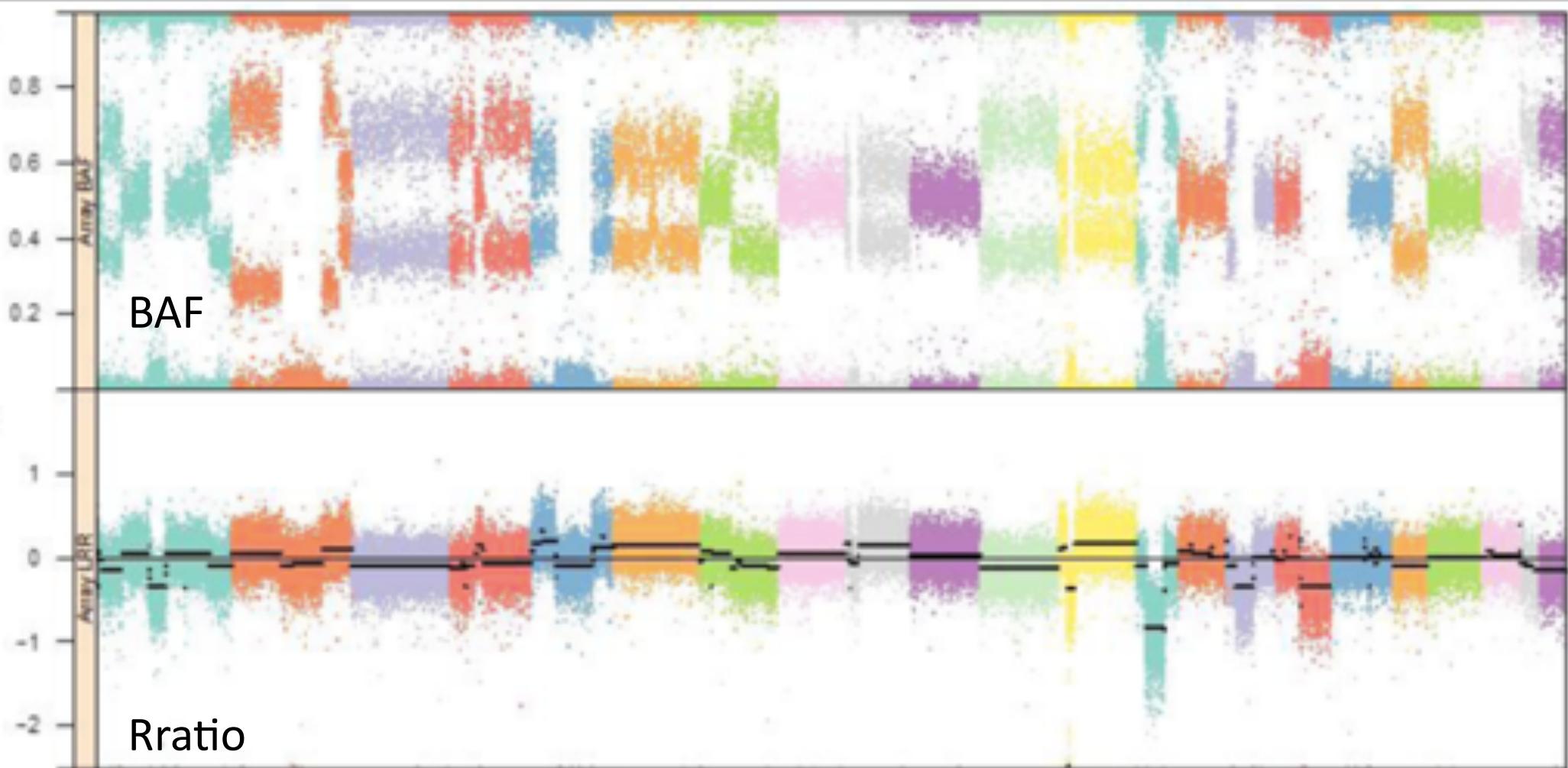
# La fraction allélique

- Vocabulaire:
  - Germline/population: Allelic frequency, MAF (minor allele frequency). Par ex. dans données 1000Genomes.
  - Somatic: Allelic fraction (mais souvent on utilise VAF or BAF: variant allele frequency)
- Où trouver l'info?
  - Colonne info#AF dans VCF

# Cellularité et Fréquence Allelique



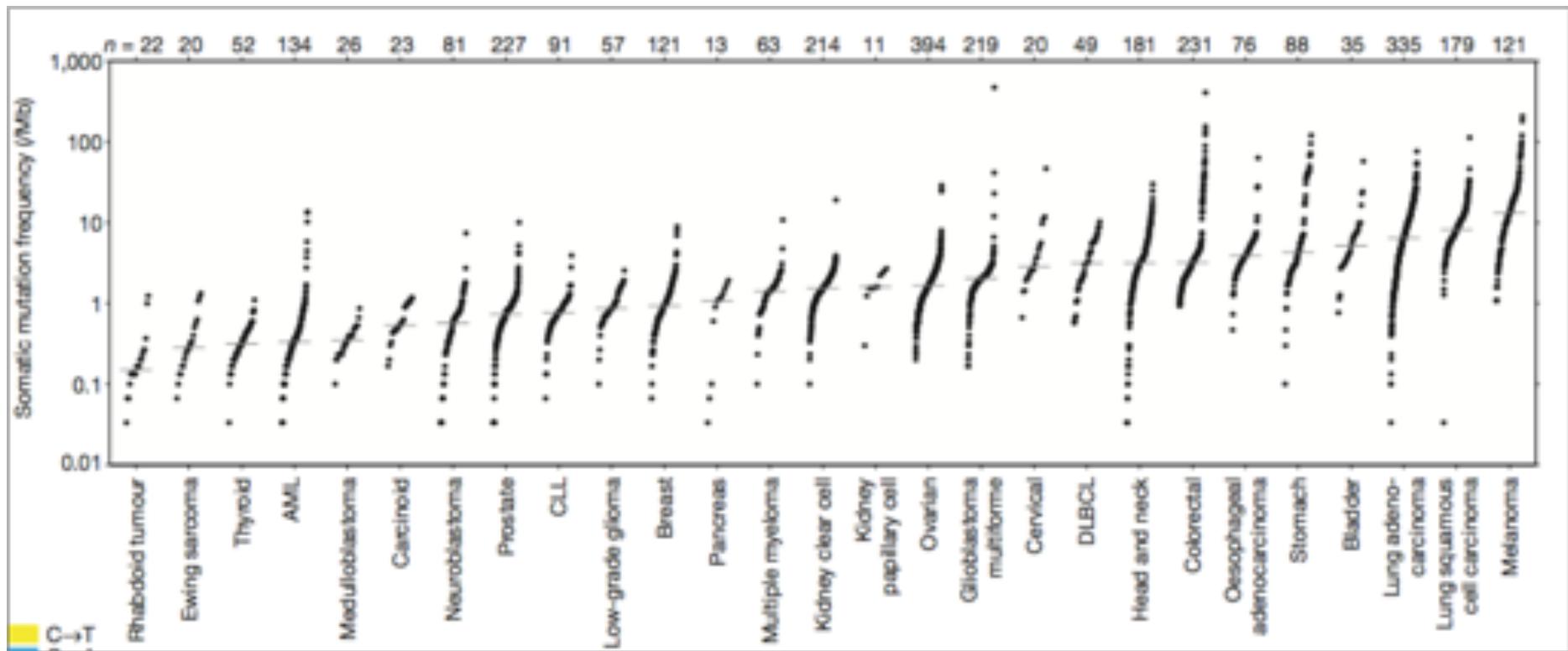
# Segmentation et fréquence allélique



R ratio=utilisé en CGH, =couverture en NGS

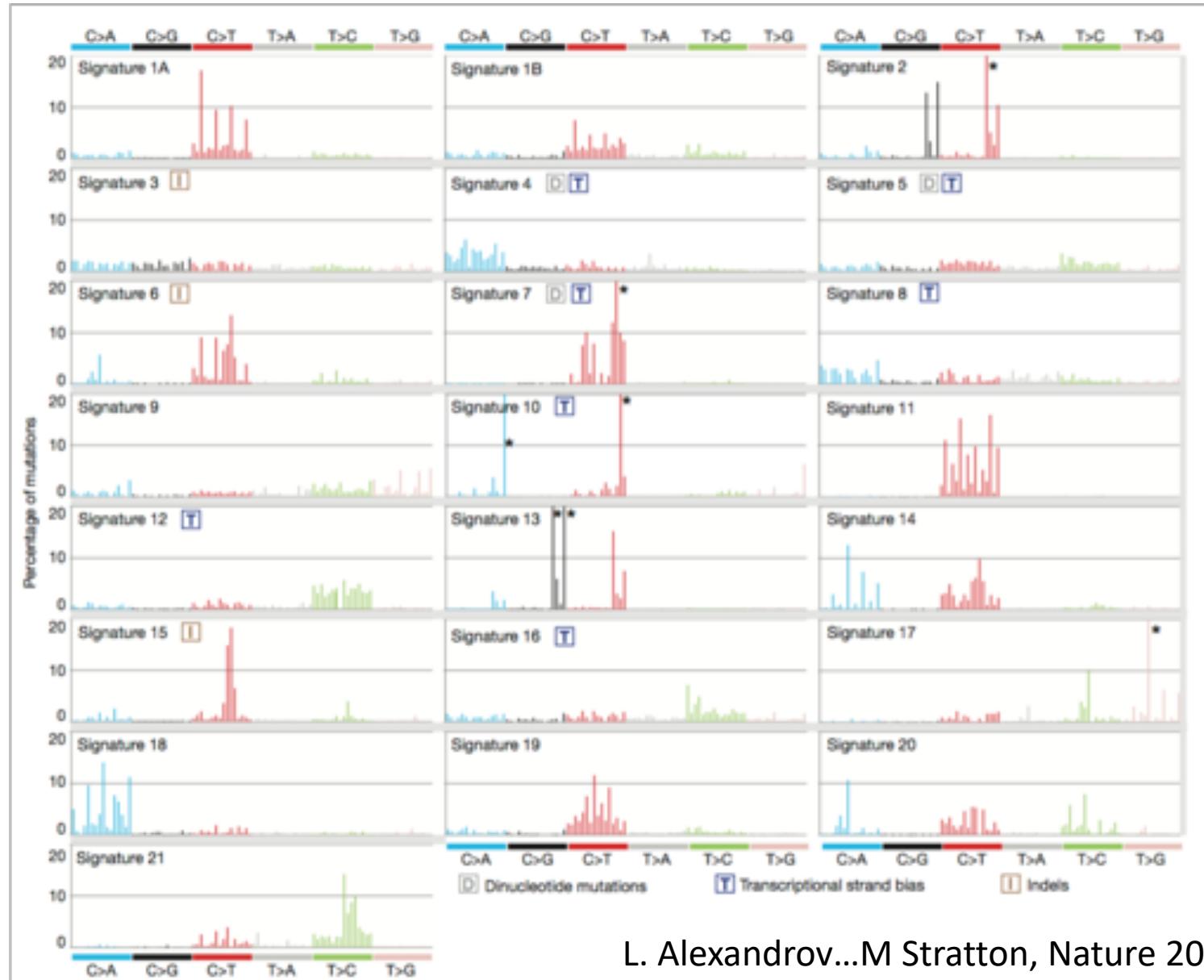
Scott et al. Gene 2014

# 2013: premières études pan-cancer WGS



Alexandrov...M Stratton, Nature 2013.  
Lawrence et al. Nature 2013.

# Les Signatures Mutationnelles



# Signatures et origine des tumeurs



# Annexes

- Formats de fichiers NGS

# Format fasta

\*.fa , \*.fasta

```
>identifiant1 commentaire libre
CAGCATCGATCGTCGGCGATGCATGCGGATGCTAGCTGATCACGATGC
CGCATGCTAGTCAGGCAGGGATATTATTAGCAGGTATCGGATGA
CAGCATTACGGCGGGAGTGCTATTATTATGAGCGGCGAT
>identifiant2 commentaire libre
CAGGCAGGTTCTTATTATATCGGCGGGCGGAGGCAGGCGATGCATC
CAGTGCAGTACGCTAGTCAGCGATGCATTATGACTGACTCAGTTT
CCCGCTAGCTATGCTATGCTATTGATCGATTGAGCTGATCTGGC
CAGCTATGCTTAGTA
```

# Format fastq

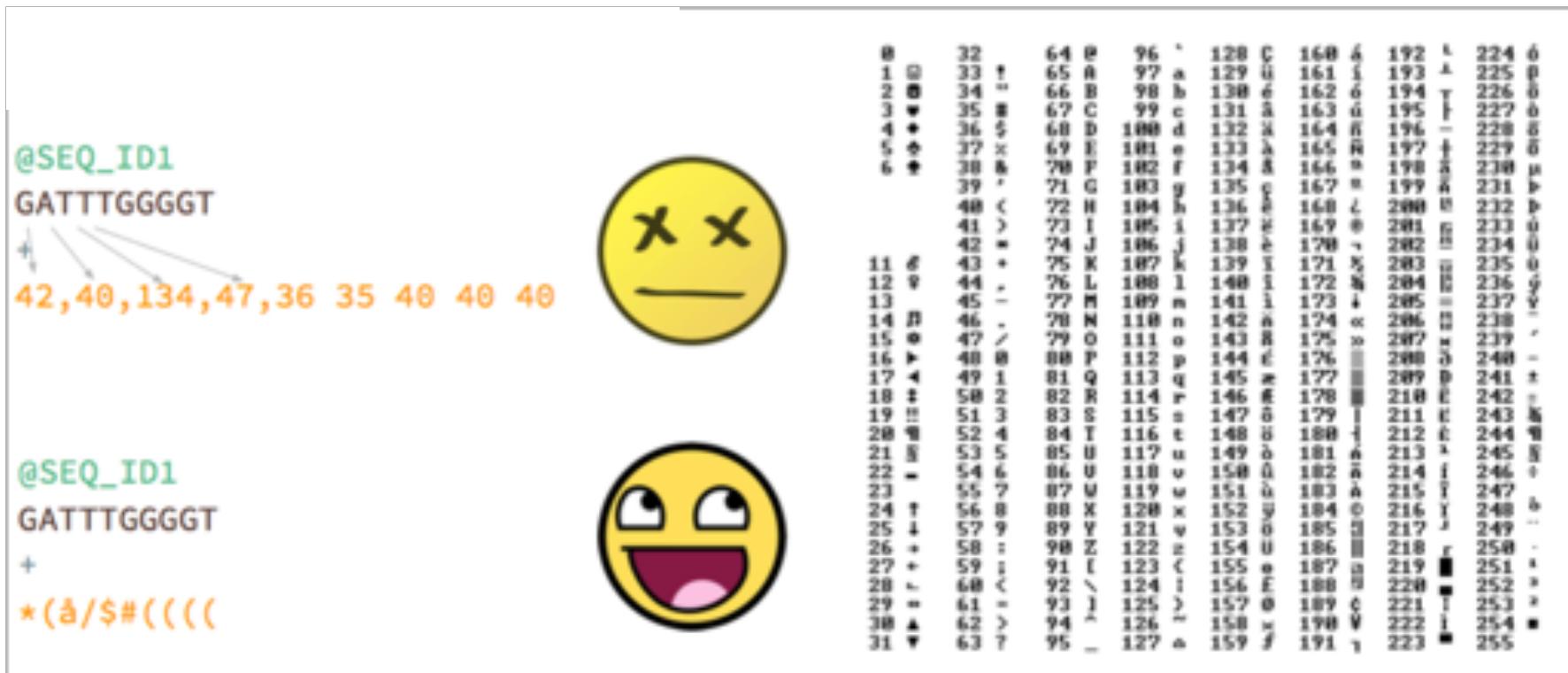
Descriptif du read (position sur la piste de séquençage, taille,...)

```
@SRR001666.1 071112_SLXA-EAS1_s_7:5:1:817:345 length=36
GGGTGATGGCCGCTGCCGATGGCGTCAAATCCCACC
+SRR001666.1 071112_SLXA-EAS1_s_7:5:1:817:345 length=36
IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII9IG9IC
```

Qualité (probabilité que la base soit correcte) encodé par code ASCII

# Qualité dans le format fastq

$$\text{Qualité} = -10 \log_{10}(P_{\text{erreur}})$$



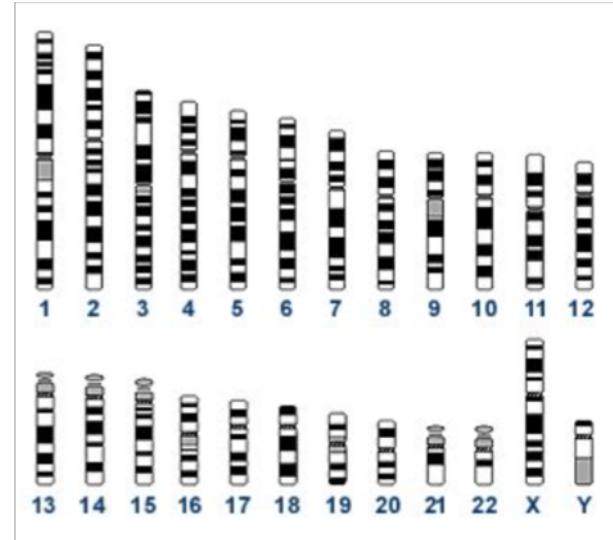
# Fichiers de régions

Coordonnées génomiques indiquant une région du génome

<chromosome>:<start>-<end>  
chr7:117465784-117715971

Formats de régions

- BED
- GTF/GFF: annotation de features dans le génome
- SAM/BAM: alignement de reads de séquence sur le génome
- VCF: variant calling file



# Format bed

obligatoire		<i>name</i>	<i>score</i>	<i>strand</i>	<i>Thick start</i>	<i>Thick end</i>	<i>color</i>	
chr7	127471196	127472363	Pos1	0	+	127471196	127472363	255,0,0
chr7	127472363	127473530	Pos2	0	+	127472363	127473530	255,0,0
chr7	127473530	127474697	Pos3	0	+	127473530	127474697	0,255,0
chr7	127474697	127475864	Pos4	0	+	127474697	127475864	255,0,255

## Attention

Le premier nucléotide est numéroté 0.

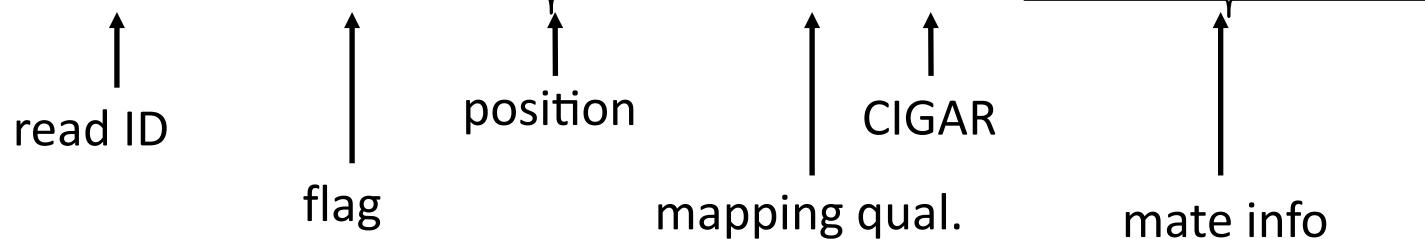
end - start = taille de la séquence



# Format SAM/BAM

Rappel BAM:

```
@RG  ID:group1  SM:1425_CD34  PL:ILLUMINA  LB:lib1 PU:unit1
@PG  ID:bwa  PN:bwa  VN:0.7.12-r1039 CL:bwa mem -M -t 2 -A 2 -E 1 -R @RG\tID:group1\tSM:1425_CD34\tPL:ILLUMINA\tLB:lib1\tPU:unit1 /root/myd
ERR166338.13782800  83  chr13  32890449  60  101M  =  32890343  -207  GGGACTGAATTAGAACAAATTTCAGCGCTT
ERR166338.13782800  163  chr13  32890343  60  75M  =  32890449  207  CACTAGCCACGTTCGAGTGCTTAATGTGGCTAGTGGC
ERR166338.26716588  99  chr13  32890406  60  101M  =  32890553  222  AATGTTCCCACCTCACAGTAAGCTGTTACCGTTCCAG
ERR166338.26716588  147  chr13  32890553  60  75M  =  32890406  -222  TTGCAGACTTACCAAGCATTGGAGGAATATCGTAA
ERR166338.27259961  99  chr13  32890496  60  101M  =  32890558  137  ACCTCAGTCACATAATAAGGAATGCATCCCTGTGTAAG
ERR166338.27259961  147  chr13  32890558  60  75M  =  32890496  -137  GACTTATTTACCAAGCATTGGAGGAATATCGTAGGTAA
ERR166338.63037998  99  chr13  32890496  60  101M  =  32890558  137  ACCTCAGTCACATAATAAGGAATGCATCCCTGTGTAAG
ERR166338.63037998  147  chr13  32890558  60  75M  =  32890496  -137  GACTTATTTACCAAGCATTGGAGGAATATCGTAGGTAA
```



# Le champ CIGAR

Example:

52M36890N45M3S

*REF : chr20*



## All Cigar operations

Op	BAM	Description
M	0	alignment match (can be a sequence match or mismatch)
I	1	insertion to the reference
D	2	deletion from the reference
N	3	skipped region from the reference
S	4	soft clipping (clipped sequences present in SEQ)
H	5	hard clipping (clipped sequences NOT present in SEQ)
P	6	padding (silent deletion from padded reference)
=	7	sequence match
X	8	sequence mismatch

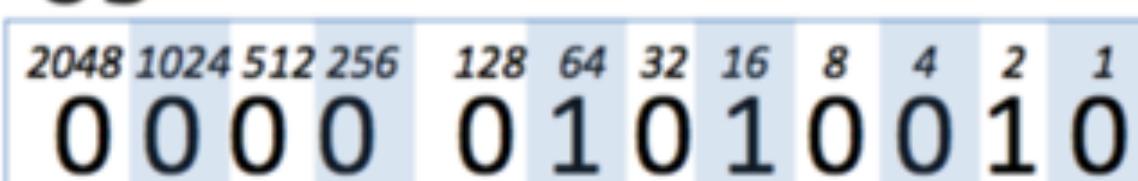
# Les Flags SAM

## Example:

- Decimal Flag Value

83

- Binary Flag Value



- To each bit corresponds a meaning

Bit	Description
1	0x1 template having multiple segments in sequencing
2	0x2 each segment properly aligned according to the aligner
4	0x4 segment unmapped
8	0x8 next segment in the template unmapped
16	0x10 SEQ being reverse complemented
32	0x20 SEQ of the next segment in the template being reverse complemented
64	0x40 the first segment in the template
128	0x80 the last segment in the template
256	0x100 secondary alignment
512	0x200 not passing filters, such as platform/vendor quality controls
1024	0x400 PCR or optical duplicate
2048	0x800 supplementary alignment

# Format VCF

```

##fileformat=VCFv4.2
##fileDate=20090805
##source=myImputationProgramV3.1
##reference=file:///seq/references/1000GenomesPilot-NCBI36.fasta
##contig=<ID=20,length=62435964,assembly=B36,md5=f126cdf8a6e0c7f379d618ff66beb2da,species="Homo sapiens",taxonomy_id=9606>
##phasing=partial
##INFO<ID=NS,Number=1,Type=Integer,Description="Number of Samples With Data">
##INFO<ID=DP,Number=1,Type=Integer,Description="Total Depth">
##INFO<ID=AF,Number=1,Type=Float,Description="Allele Frequency">
##INFO<ID=AA,Number=1,Type=String,Description="Ancestral Allele">
##INFO<ID=DB,Number=0,Type=Flag,Description="dbSNP membership, build 129">
##INFO<ID=H2,Number=0,Type=Flag,Description="HapMap2 membership">
##FILTER<ID=q10,Description="Quality below 10">
##FILTER<ID=s50,Description="Less than 50% of samples have data">
##FORMAT<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
##FORMAT<ID=DP,Number=1,Type=Integer,Description="Read Depth">
##FORMAT<ID=HQ,Number=2,Type=Integer,Description="Haplotype Quality">
#CHROM POS ID REF ALT QUAL FILTER INFO
20 14370 rs6064257 G A 29 PASS NS=3;DP=14;AF=0.5;DB;H2
20 17330 . T A 3 q10 NS=3;DP=11;AF=0.017
20 1110696 rs6040355 A G,T 67 PASS NS=2;DP=10;AF=0.333,0.667;AA=T;DB
20 1230237 . T . 47 PASS NS=3;DP=13;AA=T
20 1234567 microsat1 GTC G,GTCT 50 PASS NS=3;DP=9;AA=G

```

mandatory

Optional header: meta-data about available annotation

mandatory

samples

	FORMAT	NA00001	NA00002	NA00003
	GT:GQ:DP:HQ	0 0:48:1:51,51	1 0:48:8:51,51	1/1:43:5:...
	GT:GQ:DP:HQ	0 0:49:3:58,50	0 1:3:5:65,3	0/0:41:3
	GT:GQ:DP:HQ	1 2:21:6:23,27	2 1:2:0:18,2	2/2:36:4
	GT:GQ:DP:HQ	0 0:54:7:56,60	0 0:48:4:51,51	0/0:61:2
	GT:GQ:DP	0/1:35:4	0/2:17:2	1/1:40:3

deletion

Insertion  
(2 events here)

NS: number of samples with data  
DP: combined depth  
AF: allelic fraction  
AA: ancestral allele

GT: genotype (0=ref, 1=alt)  
GQ: genotype quality  
DP: read depth

# VarScan Tabulated Format

Chrom	Position	Ref	Cons	Reads1	Reads2	VarFreq	Strands 1	Strands 2	Qual1	Qual2	Pvalue	Map Qual1	Map Qual2	R1 +	R1 -	R2 +	Rs2 -	Alt
chr12	113348849	C	Y	31	30	49.18%	2	2	27	27	0.98	1	1	19	12	25	5	T
chr12	113354329	G	R	72	2	2.70%	2	2	31	26	0.98	1	1	48	24	1	1	A
chr12	113357193	G	A	2	72	97.30%	1	2	28	24	0.98	1	1	2	0	45	27	A
chr12	113357209	G	A	0	77	100%	0	2	0	29	0.98	0	1	0	0	51	26	A

**Cons** : Consensus Genotype of Variant Called (IUPAC code):

M -> A or C	Y -> C or T	D -> A or G or T	W -> A or T	V -> A or C or G
R -> A or G	K -> G or T	B -> C or G or T	S -> C or G	H -> A or C or T

# MAF format

Mutation Annotation Format (MAF) is a tab-delimited text file with aggregated mutation information from [VCF Files](#) and are generated on a project-level.

Column	Description
1 - Hugo_Symbol	<a href="#">HUGO</a> symbol for the gene (HUGO symbols are always in all caps). "Unknown" is used for regions that do not correspond to a gene
2 - Entrez_Gene_Id	<a href="#">Entrez gene</a> ID (an integer). "0" is used for regions that do not correspond to a gene region or Ensembl ID
3 - Center	One or more genome sequencing center reporting the variant
4 - NCBI_Build	The reference genome used for the alignment (GRCh38)
5 - Chromosome	The affected chromosome (chr1)
6 - Start_Position	Lowest numeric position of the reported variant on the genomic reference sequence. Mutation start coordinate
7 - End_Position	Highest numeric genomic position of the reported variant on the genomic reference sequence. Mutation end coordinate
8 - Strand	Genomic strand of the reported allele. Currently, all variants will report the positive strand: '+'
9 - Variant_Classification	Translational effect of variant allele
10 - Variant_Type	Type of mutation. TNP (tri-nucleotide polymorphism) is analogous to DNP (di-nucleotide polymorphism) but for three consecutive nucleotides. ONP (oligo-nucleotide polymorphism) is analogous to TNP but for consecutive runs of four or more (SNP, DNP, TNP, ONP, INS, DEL, or Consolidated)
11 - Reference_Allele	The plus strand reference allele at this position. Includes the deleted sequence for a deletion or "-" for an insertion
12 - Tumor_Seq_Allele1	Primary data genotype for tumor sequencing (discovery) allele 1. A "-" symbol for a deletion represents a variant. A "-" symbol for an insertion represents wild-type allele. Novel inserted sequence for insertion does not include flanking reference bases
13 - Tumor_Seq_Allele2	Tumor sequencing (discovery) allele 2
14 - dbSNP_RS	The rs-IDs from the <a href="#">dbSNP</a> database, "novel" if not found in any database used, or null if there is no dbSNP record, but it is found in other databases
15 - dbSNP_Val_Status	The dbSNP validation status is reported as a semicolon-separated list of statuses. The union of all rs-IDs is taken when there are multiple
16 - Tumor_Sample_Barcode	Aliquot barcode for the tumor sample
17 - Matched_Norm_Sample_Barcode	Aliquot barcode for the matched normal sample
18 - Match_Norm_Seq_Allele1	Primary data genotype. Matched normal sequencing allele 1. A "-" symbol for a deletion represents a variant. A "-" symbol for an insertion represents wild-type allele. Novel inserted sequence for insertion does not include flanking reference bases (cleared in somatic MAF)
19 - Match_Norm_Seq_Allele2	Matched normal sequencing allele 2
20 - Tumor_Validation_Allele1	Secondary data from orthogonal technology. Tumor genotyping (validation) for allele 1. A "-" symbol for a deletion represents a variant. A "-" symbol for an insertion represents wild-type allele. Novel inserted sequence for insertion does not include flanking reference bases
21 - Tumor_Validation_Allele2	Secondary data from orthogonal technology. Tumor genotyping (validation) for allele 2
22 - Match_Norm_Validation_Allele1	Secondary data from orthogonal technology. Matched normal genotyping (validation) for allele 1. A "-" symbol for a deletion represents a variant. A "-" symbol for an insertion represents wild-type allele. Novel inserted sequence for insertion does not include flanking reference bases (cleared in somatic MAF)