

# Oligator

## user manual - version 1.0.1

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## Contents

<b>1</b>	<b>Oligosaccharide features</b>	<b>3</b>
1.1	Topology design . . . . .	3
1.2	Structure notation . . . . .	3
1.2.1	Import a structure . . . . .	4
1.2.2	Export a structure . . . . .	4
<b>2</b>	<b>Ose features</b>	<b>6</b>
2.1	Structure description of oses . . . . .	6
2.1.1	View controls . . . . .	6
2.1.2	Cycle forms . . . . .	6
2.1.3	Carbon features . . . . .	7
2.2	Chemical substitutions . . . . .	7
2.2.1	List . . . . .	7
2.2.2	Definition . . . . .	8
<b>3</b>	<b>Theoretical spectrum production</b>	<b>10</b>
3.1	Theoretical m/z . . . . .	10
3.2	Theoretical intensities . . . . .	11
3.3	Export . . . . .	12

## Preamble

Oligator tool is composed of 2 categories of windows. The first one is the drawing space where elements constituting the oligosaccharide are viewed and assembled. Hydroxyl groups and hydrogens bound to monosaccharide carbons are not represented on the drawing space but implicitly considered.

The second one is a series of formular windows to refine the description of the structure and the resulting mass spectrum. The drawing space forms the main window whereas formular windows are displayed as pop-ups.

All computed masses correspond to monoisotopic masses of atoms. Charges are not expressed in the structure definition as the mass spectrum is built by selecting ionization mode of the neutral form of the drawn structure.

# 1 Oligosaccharide features

## 1.1 Topology design

An user can quickly draw the skeleton of an oligosaccharide with few controls. From the menu bar, the "Design" choice is used to define default ose type and default osidic linkage. The add button, figure 1(a), of the drawing bar adds the image corresponding to the default ose type on the drawing space. An user can therefore add multiple units of the same type and then change the default value for a remaining set of units to add.



Figure 1: Drawing bar buttons.

The remove button, figure 1(b), deletes all selected units (see also section 2.1.1) from the topology either linked or not to the polymer. The "del" key press from keyboard has the same effect. The "Clear all" option from the "Design" menu removes all units from the drawing space.

The grid button, figure 1(c), aligns connected units and disposes them into a grid according to their carbon bindings. It has no effect on free units without osidic bond.

The binding and unbinding of oses are managed by right-clicking a selection of units. The bind action connect carbons defined by default (from "Design" menu). This assignment can be modified by double clicking the edge. Available carbons to connect are C#1 (for aldoses) or C#2 (for cetoses) at reducing end of oses and free carbons at other locations. Free carbons are those without chemical substitution and not yet implied in any osidic binding (figure 2). The reducing end of the oligosaccharide is on the right side of the draw space.

## 1.2 Structure notation

From the "Notation" option of the menu bar, user can import and export text files containing the chemical description of an oligosaccharide structure in

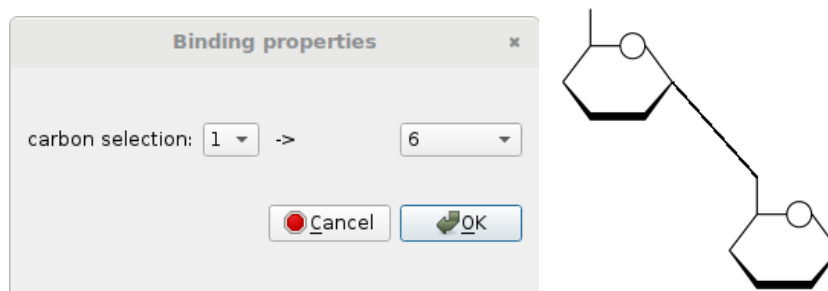


Figure 2: Carbons selection of an osidic binding.

the form of linear notations. The WURCS notation (Web3 unique representation of carbohydrate structures ) is specific to glycans and compliant with the repository GlyTouCan. The SMILES notation (Simplified molecular-input line-entry system) is unspecific to a type of compound and can be used in various chemical structure editors.

### 1.2.1 Import a structure

Oligator reads only the version 2 of the WURCS notation and SMILES from different sources (e.g. PubChem repository, ChemSketch editor). Paste notation from clipboard is unavailable.

When the notation is parsed, the glycan described by both notations is displayed on the drawing space and ose detail is accessible by double-clicking its image. In the presence of a substituent unknown to Oligator, the resulting mass delta is related to a default naming and numbering: ukn#.

The figure 3 displays the GnGnX glycan from both the WURCS and SMILES notations extracted from the PubChem database (compound id : 57339199).

### 1.2.2 Export a structure

Structures can be exported as both notations SMILES and WURCS except when the oligosaccharide carries cyclic substituent or anhydric bond. In that case, only SMILES notation export is possible. Implementation of WURCS notation follows the rules published by Tanaka, Kenichi *et al.*.

Implementation of SMILES follows the OpenSMILES specifications. Nested branches are used during the parsing of the molecule that always starts

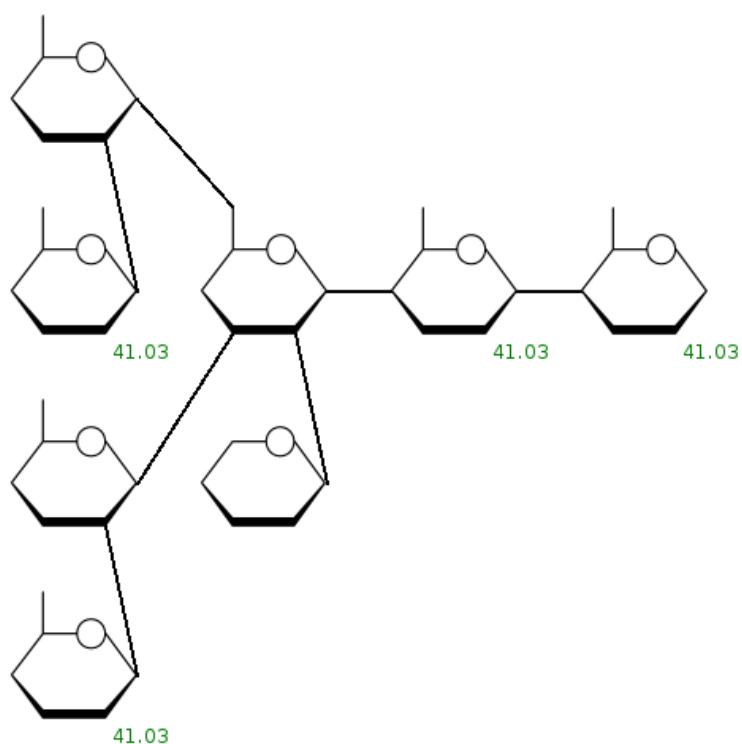


Figure 3: GnGnX compound from WURCS and SMILES notations.

from its reducing end (e.g. "C1(C(C(C(C(O1)CO)O)O)O)O" for hexopyranose). Aromaticity is represented in the Kekule form (C1=CC=CC=C1). The lower-case letters notation is not implemented in the current version of Oligator.

## 2 Ose features

### 2.1 Structure description of oses

Oses are represented through 4 elementary images: hexofuranose, hexopyranose, pentofuranose and pentopyranose.

#### 2.1.1 View controls

Within the draw space, ose controls are accessed through the selection of images. The selection is viewed as dot lines around ose image. There are 3 ways to select oses:

- by clicking on a single image,
- by drawing a rectangular area around a group of images,
- or by pressing the ctrl key + clicking on images one by one.

The selected images can be further dragged and dropped together across the drawing space. Clicking outside any image deselects all oses. Double-clicking on an image triggers the pop-up of ose detail window (figure 4).

Ose description		
Number of carbons :	6	Ring start : 1
		Form : pyrane
Isomers:	Substituents :	
C1	L	hydroxy
C2	D	hydroxy
C3	L	hydroxy
C4	D	hydroxy
C5	D	hydroxy
C6		sulfate
b-dGlc		<input type="checkbox"/> 3,6-anhydro bond
		Cancel OK

Figure 4: Customization of ose structure

#### 2.1.2 Cycle forms

An ose is displayed in either as images derived from the Haworth description with implicate OH groups. As a consequence, the stereo-isomerism of carbons is not displayed (no incidence on molecule mass).

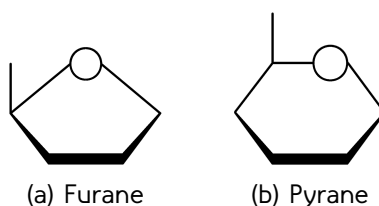


Figure 5: Display of cyclic forms of oses.

Cyclic form can be changed from the detail window (figure 4). By the same way, the total number of carbons constituting the monosaccharide and the carbon starting the cycle can also be changed. A vertical line on the left of the carbon list indicates which ones are in the cycle.

### 2.1.3 Carbon features

For each chiral carbon, the stereochemistry can be assigned according to Fisher projection where D indicates that OH group is on the right of the carbon and L, on its left. For each free carbon, i.e. unrelated to cycle closure or any osidic bond, a substituent can be added. Non-substituted ose has hydroxyl on each carbon.

## 2.2 Chemical substitutions

Numbers at carbon locations indicate chemical substitutions of OH groups resulting in mass gain (green) or loss (red) on the molecule (figure 6).

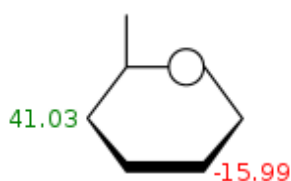


Figure 6: Ose displaying desoxydation of carbon #2 and  $\beta$ -acetylation of carbon #4.

### 2.2.1 List

A substituent list is available from the menu and can be edited by user during the session from the menu option named "Substituents".

This list is a manual extraction of monosaccharidDB with the addition of ferulic acid (found in plants). Anhydro bridge found in algae is a particular ose modification that is presented in Oligator as a check box in ose detail rather than as a substituent of the list.

Displayed as a table where column names are "Name", "Composition", "Mass", "SMILES" and "Linker" (figure 7), the list can be ordered according to one of these columns. The user can add, edit or remove any line of the table.

The substituent list is stored with the Oligator code as a json file in the Ressources directory. Although not recommended, this format enables text edition to store durably additional substituents.

Name	Composition	Mass	SMILES
trifluoroacetyl	C2O2F3	112.985	OC(=O)C(F)(F)F
thio	H1S1	32.980	S
telluro	H1Te1	130.914	Te
sulfate	H1O4S1	96.960	OS(=O)(=O)O
seleno	H1Se1	80.924	Se
s-methyl	C1H3S1	46.996	SC
pyruvate	C3H3O3	87.008	OC(=O)C(=O)O
phospho-choline	C5H15N1O4P1	184.074	OC[N+](C)(C)COP(=O)(O)O
phosphate	H2O4P1	96.969	OP(=O)(O)O
oxydation	O2H1	32.998	(=O)O
nitrat	N1O2	45.993	N(=O)O
n-trimethyl	C3H9N1	59.073	N(C)(C)C
n-trifluoroacetyl	N1C2F3O1H1	112.001	NC(=O)C(F)(F)F
n-sulfate	H2N1O3S1	95.976	NS(=O)(=O)O
n-succinate	C4H5N1O3	115.027	NC(=O)CC(=O)O
n-methyl	C1H4N1	30.034	NC
n-glycolyl	C2H4N1O2	74.024	NC(=O)CO
n-formyl	C1H2N1O1	44.014	NC=O

Figure 7: Substituent list

### 2.2.2 Definition

When adding or editing a substituent, the user has to fill in the SMILES notation beginning by the atom linker to ose carbon (figure 8). In order to help the user to select the atom linker, the graph of atom is displayed according



to the SMILES notation filled. Any atom can be the ose carbon linker according to its valence and is highlighted in green on the graph when binding is possible, in red otherwise. The SMILES notation is reordered according to the user selection. The "refresh view" button rearranges the nodes of the graph and can be used when overlaps occur.

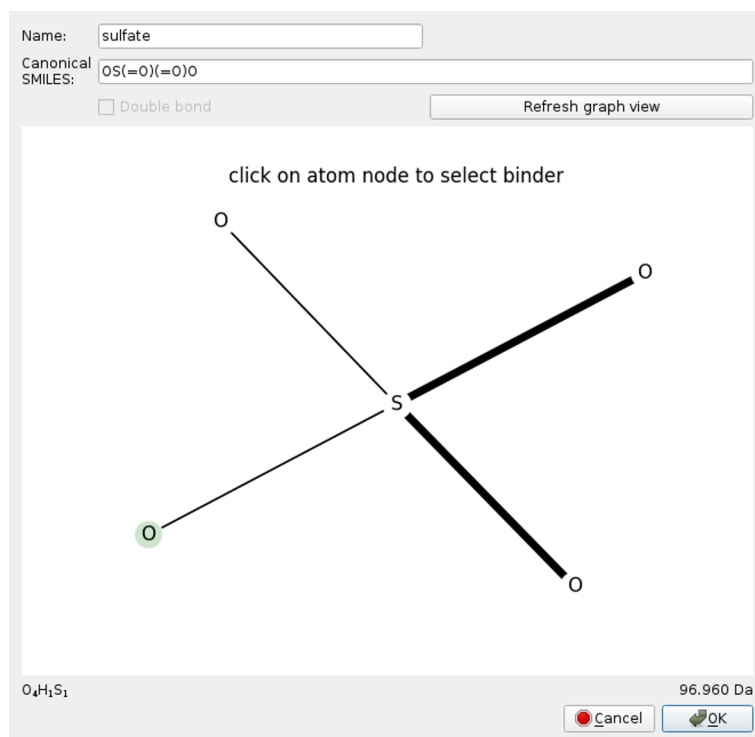


Figure 8: Substituent edition

The chemical name if the substituent is optional and randomly produced when missing. The atom composition is displayed at the left bottom of the window, minus one Hydrogen for ose binding (or 2 for a double bond) and the monoisotopic mass derived from this composition is displayed at the right bottom. The mass displayed on the drawing space of the oligosaccharide corresponds to the difference between this substituant mass and OH mass.

### 3 Theoretical spectrum production

The specification of mass and intensities is made in the "MS/MS spectrum preview" option from the "MS/MS" menu bar (figure 9). By default, the theoretical spectrum is produced from the model of a CID+ experiment. Any changes in the preview window will affect the export of the spectrum.

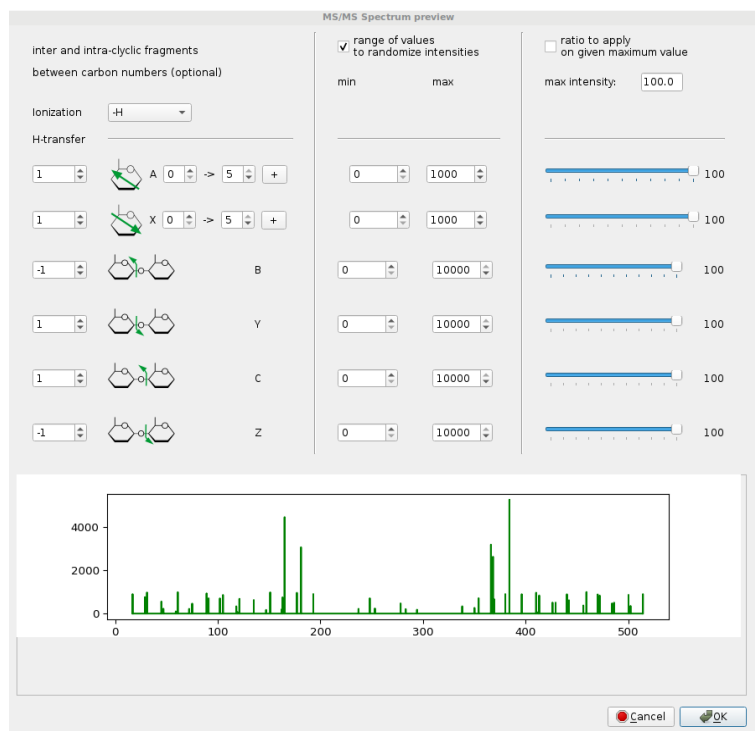


Figure 9: MS/MS spectrum preview.

#### 3.1 Theoretical m/z

The spectrum produced from a theoretical MS/MS fragmentation of oligosaccharide structure relies on the standard nomenclature published by Domon & Costello. Ion types coming from this nomenclature identifies the location of fragmentation between oses (ions Y and Z for the reducing end side, B and C on the other side) or within cycles (ion X for reducing end side, A, for other side). Branching "numbering" with greek letters and quotes is also achieved according to this standard.

Oligator handles singly charged precursor only. User can select +H or -H ionization and alkali cationization (figure 10).

Rearrangements of fragments leading to H-transfers are assigned by default according to the fragmentation pathway described by Domon & Costello. User can adjust the number of H transferred if needed. For these types, parameters can be defined for all fragments occurring between oxygen and carbon#5 or for some specific pairs of carbons ("plus" button). The "– >" symbol means "from carbon i to carbon j", whereas the "-" letter means between carbons i and j.

Figure 10: Parameters influencing the  $m/z$  values of ion types.

Masses are computed according to masses of monoisotopic atoms most frequent in the nature.

### 3.2 Theoretical intensities

Specifying the distribution of intensities is a way to describe the probability of observing ion types. As for example, ions produced by CID experiments at low dissociation energies displays less A and X ions than other types. As for masses, the intensities can be defined for each ion type and each of sub-types coming from the multiple combinations of carbon pairs from internal ose fragmentation.

There are 2 modes to define how intensities are assigned (figure 11):

- randomly between min and max bounds,
- or relatively to a maximum value.

The interface is divided into two main sections. The left section, titled 'range of values to randomize intensities' (checked), contains six rows of input fields for 'min' and 'max' values. The right section, titled 'ratio to apply on given maximum value' (unchecked), contains a 'max intensity' input field set to 100.0 and six horizontal sliders, each with a '100' label at the right end.

Figure 11: Parameters influencing the distribution of intensities.

### 3.3 Export

The exported spectrum is a simple text file including the SMILES notation of the oligosaccharide and ionization mode as headers followed by the list of ions. Each ion line contains its  $m/z$  value, intensity and annotation in a LaTeX syntax. These values are separated by a tabulation.