

Chinese Pharmaceutical Association
Institute of Materia Medica, Chinese Academy of Medical Sciences

Acta Pharmaceutica Sinica B

www.elsevier.com/locate/apsb www.sciencedirect.com



REVIEW

Dissecting the role of AMP-activated protein kinase in human diseases



Jin Li^a, Liping Zhong^b, Fengzhong Wang^{a,*}, Haibo Zhu^{c,d,e,**}

Received 10 October 2016; received in revised form 12 November 2016; accepted 17 November 2016

KEY WORDS

AMPK; Cancer; Type 2 diabetes; Atherosclerosis; Myocardial ischemia; Neurodegenerative disease **Abstract** AMP-activated protein kinase (AMPK), known as a sensor and a master of cellular energy balance, integrates various regulatory signals including anabolic and catabolic metabolic processes. Accompanying the application of genetic methods and a plethora of AMPK agonists, rapid progress has identified AMPK as an attractive therapeutic target for several human diseases, such as cancer, type 2 diabetes, atherosclerosis, myocardial ischemia/reperfusion injury and neurodegenerative disease. The role of AMPK in metabolic and energetic modulation both at the intracellular and whole body levels has been reviewed elsewhere. In the present review, we summarize and update the paradoxical role of AMPK implicated in the diseases mentioned above and put forward the challenge encountered. Thus it will be expected to provide important clues for exploring rational methods of intervention in human diseases.

© 2017 Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Peer review under responsibility of Institute of Materia Medica, Chinese Academy of Medical Sciences and Chinese Pharmaceutical Association.

^aInstitute of Food Science and Technology, Chinese Academy of Agricultural Sciences (CAAS), Beijing 100193, China

^bLife Science College of Tarim University, Xinjiang 843300, China

cState Key Laboratory for Bioactive Substances and Functions of Natural Medicines, Beijing 100050, China

^dBeijing Key Laboratory of New Drug Mechanisms and Pharmacological Evaluation Study, Beijing 100050, China

^eInstitute of Materia Medica, Chinese Academy of Medical Sciences & Peking Union Medical College, Beijing 100050, China

^{*}Corresponding author. Tel./fax: +86 10 62810295.

^{**}Corresponding author at: Institute of Materia Medica, Chinese Academy of Medical Sciences & Peking Union Medical College, Beijing 100050, China. Tel./fax: +86 10 63188106.

E-mail addresses: wangfengzhong@sina.com (Fengzhong Wang), zhuhaibo@imm.ac.cn (Haibo Zhu).

1. Introduction

Since 1973, AMP-activated protein kinase (AMPK) has been first known as an inhibitory factor of 3-hydroxy-3-methyl glutaryl coenzyme A reductase (HMGCR) and acetyl-coenzyme A carboxylase (ACC) in the presence of ATP. The responsible agents were later shown to be kinases and subsequently named as HMGCR kinase and ACC kinase 3, respectively^{1,2}. With the progress of successful purification of this kinase, it was not until 1989 when the name AMPK was finally adopted as it can be allosterically regulated by AMP³. Besides key regulatory enzymes controlling sterol and fatty acid synthesis, two other enzymes that catalyze important steps in lipolysis and glycogen synthesis (hormone-sensitive triglyceride lipase and glycogen synthase) were subsequently reported as substrates for AMPK^{4,5}. The physiological role of AMPK gradually surfaced. In the meantime, the recognition of nucleotides, AMP, ADP and ATP, as allosteric regulators of AMPK activity deepened the understanding of the physiological significance of AMPK³. The current view is that AMPK, the cellular energy sensor, is activated following cellular stress like hypoxia, starvation, glucose deprivation or muscle contraction which can increase the ratio of ADP:ATP or AMP:ATP. In order to restore cellular energy balance, the highly active enzyme integrates hormonal and nutrients signals which will promote catabolism (fatty acid oxidation and glycolysis) and inhibit anabolism (fatty acid, cholesterol, triglycerides and protein, etc.). Apart from these canonical functions, AMPK is confirmed to be involved in cell growth, development, longevity and cell polarity⁶. Additional AMPK activating factors include liver kinase B1 (LKB1), calmodulin-dependent protein kinase kinase β (CaMKK β) and transforming growth factor-β-activated kinase (TAK-1). LKB1 has been proposed to phosphorylate AMPK when the ratio of AMP:ATP is upregulated. However, AMPK can still be excited by CaMKK β during intracellular Ca²⁺ release or by TAK-1 in response to pro-inflammatory cytokines or apoptosis-inducing agent, even when no change in nucleotides are detected 7-10. The detailed information regarding structure, activity regulation and pharmacological agonists of AMPK have recently been reviewed¹¹. In this article, we will focus on the association between AMPK and a variety of human diseases, including cancer, type 2 diabetes, atherosclerosis, myocardial ischemia/ reperfusion injury and neurodegenerative disorder.

2. The role of AMPK in human diseases

2.1. AMPK in cancer

Cancer is fundamentally a disease of tissue growth regulation failure, which can be driven by dysregulation of several cell cycle components. Apart from these alterations, increased catabolic glucose metabolism was observed in proliferating cells. It is a necessity for tumor cells to overcome the significant energy challenge in the initiation of uncontrolled proliferation; otherwise, they die due to energy deficiency. In the mid-1950s, Warburg¹² discovered that tumor cells still survive in the absence of oxygen by glycolysis, and later it was referred as "Warburg effect". The metabolic switch towards the Warburg effect not only supplies the biogenetic source but also confers important metabolic intermediates for cell growth¹³. Presently, this peculiar metabolic shift is universally recognized as a hallmark for tumor cells. Progress in

identification of oncogenes and tumor suppressor genes makes the targeting of this metabolic switch a viable new approach for cancer treatments.

It has been proposed that AMPK is closely related to the regulation of the cell cycle because AMPK activation stimulates phosphorylation and activation of tumor suppressor p53, stabilization of cyclin-dependent kinase inhibitor p27 and reduction of key cell cycle regulators like cyclin A and cyclin B1^{14–16}. Oncogenic BRAF repressing LKB1 and AMPK activity accelerates melanoma cell proliferation¹⁷. On the other hand, AMPK activation blocks the progression of keratinocyte cell cycle via phosphorylation of B-Raf¹⁸. A recent study revealed that CAMKKβ-induced AMPK activation also induces cell cycle arrest19. Apart from the regulatory effect on cell cycle checkpoints, AMPK suppresses the anabolic processes required for rapid cell growth. These processes include mammalian target of rapamycin complex 1 (mTORC1)-dependent protein biosynthesis induced by direct phosphorylation of the tumor suppressor TSC2 and the regulatory-associated protein raptor²⁰, and *de novo* biosynthesis of fatty acid and cholesterol caused by inactivating ACC1, HMGCR and lipogenic transcription factors sterol regulatory element-binding proteins (SREBPs)^{21,22}. More recent genetic researches showed that the growth-suppressive action of AMPK may be mediated by the Hippo pathway 23-25. Furthermore, Faubert et al.26,27 provided evidence to demonstrate that AMPK is a negative controller of the Warburg effect using models with genetic deletion of either LKB1 or AMPKα1. AMPK activation promotes mitochondrial biosynthesis and expression of oxidative enzymes and thus attenuates the glycolytic pathway by inhibiting the transcription factor hypoxia-inducible factor 1α (HIF- 1α). Genetic ablation of AMPKα1 accelerates Myc-induced lymphomagenesis, suggesting that the absence of AMPK may enhance oncogene activity to boost tumorigenesis²⁶. Thus, AMPK can be classified as a metabolic tumor suppressor (Fig. 1).

However, several groups have recently reported that diverse variety of tumor repressors or proto-oncogenes negatively regulate AMPK activity. The absence of the tumor repressor gene folliculin (FLCN) in association with Birt-Hogg-Dube syndrome (BHD) confers tumorigenesis through activation of AMPK^{28,29}. Likewise, AMPK stimulation was found to be indispensable for the proliferation of astrocytic tumor cells or the growth of experimental human breast cancer tumor^{30,31}. Besides, microphthalmiaassociated transcription factor (MITF), a melanoma oncoprotein, is regulated by AMPK to maintain cell viability³². The requirement of AMPK for prostate cancer progression and colon tumor cell survival has also been recently reported^{33,34}. It seems that there are two faces of AMPK in tumorigenesis. In established solid tumors, AMPK activation can provide metabolic adaptive responses to maintain energy supply, although inhibition of AMPK is beneficial in earlier phases of tumor growth. It is worth noting that AMPK α 2 has been reported to selectively suppress Ras-induced mouse embryo fibroblasts (MEFs) transformation and reduce the growth of human mammary epithelial cells (HMECs)^{35,36}. However, two more recent reviews have pointed out that the PRKAA1 gene encoding $\alpha 1$ is often amplified whereas the PRKAA2 gene encoding $\alpha 2$ is more frequently mutated in human cancers^{37,38}. Thus, the two catalytic subunit isoforms may play divergent roles in cancer.

In consideration of the aforementioned role of AMPK in tumorigenesis, pharmacological activation of AMPK may exert beneficial effects on cancer. Indeed, pharmacological activation of AMPK by biguanides or A769662 to $Pten^{+/-}$ mice remarkably inhibits

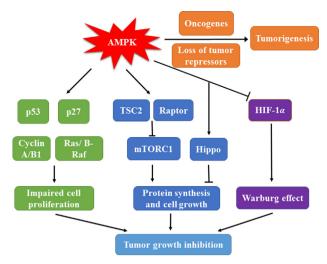


Figure 1 AMP-activated protein kinase (AMPK) plays a double-edged role in cancer. AMPK stimulation promotes activation of tumor repressor p53, increases cyclin-dependent kinase inhibitor p27, decreases cyclin A and cyclin B1, and suppresses ras and B-raf signaling pathway, resulting in cell cycle arrest and impaired cell proliferation. AMPK can induce phosphorylation of TSC2 and raptor to suppress mTOCR1 dependent protein synthesis and activate Hippo pathway to inhibit cell growth. The reduced HIF- 1α transcription activity mediated by AMPK activation leads to the block of Warburg effect and the decreased energy supply, contributing to inhibition of tumor cell growth in further. Thus, AMPK may exert its anticancer activities through multiple approaches mentioned above. However, AMPK may also cooperate with either oncogenes or loss of tumor repressors to promote tumorigenesis.

tumorigenesis³⁹. Subsequently, numerous studies have obtained similar results showing that AMPK agonists can inhibit cancer cell growth and proliferation 40-42. In clinic, a significant reduction in the incidence of cancers in subjects taking metformin compared with other antidiabetic drugs was found by meta-analysis⁴³. However, several groups have recently challenged the role of AMPK in the protective effect of these compounds on tumorigenesis. They pointed out that A769662, widely acknowledged as direct AMPK agonist, may actually accelerate tumor cell proliferation in response to metabolic stress. Additionally, those indirect AMPK agonists prevent tumor cell proliferation and mTOR activity in an AMPK-independent manner 44-46. Biguanides, a group of classic anti-diabetic drugs, was later recognized as potential anti-cancer agents. In fact, they are inhibitors of the mitochondrial respiratory chain and have recently been found to preferentially kill various cancer stem cells, which are dependent on mitochondrial metabolism^{47,48}. In the context of inhibition of proliferation, the upregulation of AMPK activity occurs as an adaptive response to protect cells from the toxicity of biguanides since the mortality increases in cells without LKB1, a critical upstream kinase of AMPK and once known as a tumor repressor⁴⁹. Further, in the mouse model of colon carcinoma or nonsmall cell lung cancer with a defective LKB1/AMPK pathway, the rate of tumor growth declines following treatment with biguanides^{50,51}. Thus, it would be promising to combine biguanides with AMPK inhibitors in the treatment of established solid tumors.

2.2. AMPK in type 2 diabetes

Type 2 diabetes (formerly named noninsulin-dependent diabetes mellitus or adult-onset diabetes) is a metabolic disorder characterized by hyperglycemia and abnormal lipid metabolism in the context of decreased insulin sensitivity of peripheral tissue and inadequate insulin secretion by islet beta cells. It has been agreed that overnutrition, inactivity and consequential obesity are the primary cause of type 2 diabetes in genetically predisposed individuals. Regular exercise and proper dietary are thought to

be first steps to manage the disease. The beneficial effects of exercise may be at least partly mediated by AMPK activation, consistent with a critical role of AMPK in regulating glucose metabolism⁵².

The above mentioned insulin-insensitive peripheral tissues mainly consist of skeletal muscle, liver and fat. Among them, skeletal muscle constitutes around 80% of insulin-stimulated glucose disposal⁵³. It has been reported that AMPK activation by chronic administration of metformin enhances insulinstimulated glucose uptake in mouse soleus muscle⁵⁴. One explanation may be that AMPK activation enhances insulin receptor substrate 1 (IRS-1) Ser789 phosphorylation and subsequent phosphoinositide 3 kinase/protein kinase B (PI3K/PKB) signaling pathway^{55,56}. However, in vivo evidence to support this is lacking, since either AMPK $\beta1\beta2$ muscle knockout mice or mice overexpressing AMPKα2 kinase-dead (KD) in muscle have normal insulin-stimulated glucose transport^{57,58}. The relationship between AMPK and insulin-independent stimulation of glucose uptake has attracted significant interest. AMPK agonists can stimulate glucose uptake in resting skeletal muscle, and genetic techniques confirmed that these effects are indeed mediated by AMPK⁵⁹⁻⁶¹ Moreover, it has also been found that AMPK $\alpha 2\beta 2\gamma 3$ heterotrimer is mainly activated in skeletal muscle⁵². Although there are significant contradictory reports regarding the role of AMPK in exercise-induced glucose transport, results in muscle-specific AMPKβ1/AMPKβ2 double knockout mice have argued convincingly that AMPK is implicated in glucose transport in response to exercise or muscle contraction⁵⁷. When AMPK is activated by exercise or contraction, both Rab GTPase-activating protein TBC1D4 (also known as AS160) and TBC1D1 are phosphorylated and inactivated. Nevertheless, TBC1D1 plays a more pivotal role. The phosphorylated form can recruit scaffolding protein 14-3-3 and allow GLUT4 storage vesicles transport to plasma membrane^{62,63} (Fig. 2).

Apart from impaired glucose uptake in skeletal muscle, the excessive release of glucose into the circulation by liver is another

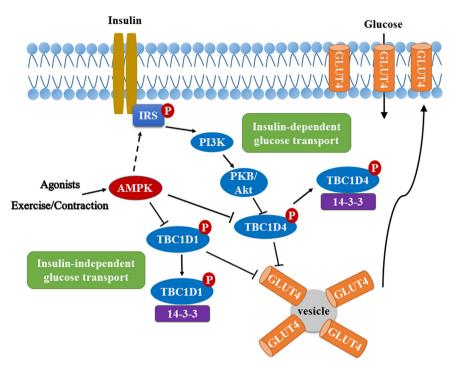


Figure 2 Regulation of glucose uptake in muscle by AMPK. AMPK activation by exercise or muscle contraction or agonists can phosphorylate Rab GTPases TBC1D4 and TBC1D1, increase 14-3-3 binding and subsequent dissociation from GLUT4, which promotes glucose uptake through increasing translocation of GLUT4 to plasma membranes. On the other hand, the insulin-mediated glucose uptake includes insulin receptor and insulin receptor substrate phosphorylation, PI3K/Akt activation and then phosphorylation of TBC1D4. AMPK may participate in the phosphorylation of insulin receptor substrate-1 *in vitro* but there is few *in vivo* evidence to support this.

contributor to hyperglycemia. Dysregulated hepatic glucose production is mainly caused by abnormal gluconeogenesis and elevated plasma glucagon levels. Metformin, a widely used anti-diabetic agent, was first found to inhibit gluconeogenesis through the signaling pathway of LKB1⁶⁴. However, whether AMPK plays a key role in the reduction of hepatic glucose output by biguanides is quite controversial. Foretz et al.65 showed that metformin suppresses gluconeogenesis via a decrease in hepatic energy state independent of LKB1/AMPK pathway by genetic loss-of-function experiments. Subsequently, other groups also found that the inhibition of glucose output by metformin is attributed to the reduced hepatic glucagon signaling and declining mitochondrial glycerophosphate dehydrogenase activity^{66,67}. Conversely, Fullerton et al.⁶⁸ reported that metformin-induced AMPK activation and ACC phosphorylation play crucial roles in lipid-induced insulin resistance. It seems that the acute effect of metformin on hepatic glucose output may be AMPKindependent, whereas the longer-term effects (which are probably more relevant to therapy of humans with metformin) are AMPKdependent. Subsequently, Cao et al.⁶⁹ suggested that a low concentration of metformin inhibited gluconeogenic gene expression via AMPK without increasing the AMP/ATP ratio in primary hepatocytes. Furthermore, the increase in the net phosphorylation of AMPK Thr172 is caused by metformin-mediated increment in formation of the AMPK complex⁷⁰. More recently, a small-molecule AMPK activator 991 was demonstrated to antagonize hepatic glucagon signaling via AMPK-induced cyclic nucleotide phosphodiesterase 4B (PDE4B) activation⁷¹.

Adipose tissue is the main resource for plasma free fatty acids (FFAs) and abundant FFA accumulation in skeletal muscle, liver and adipocytes can drive insulin resistance *via* diacylglycerol (DAG) accumulation and protein kinase C (PKC) activation ^{72,73}. AMPK

activation in adipocytes inhibits lipogenesis due to increased phosphorylation of ACC and decreased expression of lipogenic genes including stearoyl-CoA desaturase 1 (SCD1), fatty acid synthase (FAS) and ACC1, which are under control of transcription factor SREBP-1c^{74,75}. On the other hand, the inactivation of ACC contributes to decreased malonyl-CoA and thus the attenuated inhibition of carnitine palmitoyltransferase 1 (CPT1), a critical enzyme for fatty acid oxidation in mitochondria. Although acute treatment of AICAR in adipocytes was found to attenuate fatty acid oxidation, which is associated with reduced fatty acid uptake⁷⁶, chronic activation of AMPK-stimulated fatty acid oxidation and mitochondrial biogenesis^{74,77}. The role of AMPK in lipolysis is paradoxical. Exercise was reported to promote lipolysis in an AMPK-dependent fashion stimulated by adrenaline in adipocytes⁷⁸. The pro-lipolytic action of AMPK was suggested to be closely associated with increased ATGL phosphorylation⁷⁹. In contrast, mice deficient in AMPK α 1 revealed a phenotype of smaller adipocytes with increased lipolysis. The antilipolytic effect of AMPK was demonstrated to act on hormonesensitive lipase (HSL) by blocking its translocation to the lipid droplet⁸⁰. It seems that acute activation of AMPK suppresses adipose lipolysis and thereby decreasing serum fatty acid concentration, whereas prolonged activation promotes lipolysis⁷⁴. A recent study demonstrated that nicotine acts on adipose tissue to accelerate lipolysis and induce insulin resistance through activating AMPK $\alpha 2^{81}$. However, the overall impact of AMPK on lipolysis still remains controversial.

The fourth contributor to Type 2 diabetes is the decline in numbers of normally functioning islet β cells, which is concurrent with dysregulated glucose-induced insulin secretion. Granot and other groups 82,83 first showed that genetic deletion of LKB1 in pancreatic beta cells dramatically increased insulin secretion in response to glucose and improved glucose tolerance; dramatic

changes in β cell mass and polarity were also seen in vivo. Subsequently, two groups independently reported that LKB1 and AMPK may play different roles in the control of insulin secretion from islet β cells^{84,85}. In these studies, AMPK deficiency both in pancreatic β cell and hypothalamic neurons displayed defective insulin secretion and glucose-intolerance. Recently, Kone and his colleagues⁸⁶ reconfirmed the above results through developing new models without AMPK deletion in the brain. One possible mechanism involves promotion of AMPK-dependent KATP channel trafficking, alleviation of endoplasmic reticulum stress and reduction of lipid accumulation via autophagy stimulation^{87–89}. Thus, AMPK activation seems to have beneficial effect on islet β cells. Indeed, administration of AICAR protects against glucolipotoxicity-induced impaired β -cell function 90 . However, mice with the Arg299Gln γ2-specific mutation develop dysregulated β -cell function and obesity due to sustained activation of AMPK throughout all tissues⁹¹.

Any AMPK activator that crosses the blood-brain barrier would be likely to have adverse effects on food intake because hypothalamic AMPK plays a critical role in the regulation of feeding behavior. This process is suggested closely associated with the expression of orexigenic neuropeptide Y (NPY)/agouti-related protein (AgRP) and anorexigenic proopiomelanocortin- α (POMC)⁹². Either pharmacological activation or expressing constitutively active AMPK in hypothalamus increased food intake, whereas expressing dominant-negative (DN) AMPK in hypothalamus decreased the expression of NPY and AgRP^{93,94}. Claret et al. 95 reported that mice with AMPK α 2 deletion in proopiomelanocortin (POMC)-expressing neurons develop obesity due to increased food intake and decreased energy expenditure. On the contrary, mice with AMPKα2 defective in agouti-related protein (AgRP)-expressing neurons maintain their lean phenotype, suggesting there is a close relationship between AMPK and the activation of these neurons. A recent study pointed out that hypothalamic AMPK regulates neuropeptide expression through induction of autophagy⁹⁶.

2.3. AMPK in atherosclerosis

Atherosclerosis is a slow, progressive disease with accumulated cholesterol, triglyceride, immune cells and fibrin in the intima of coronary and larger arteries, which constitutes plaque. The plaque may gradually plug these arteries causing many other cardiovascular diseases including myocardial infarction and stroke. Solid evidence implicates AMPK in atherosclerosis through modulation of macrophage cholesterol homeostasis, inflammation and vascular dysfunction.

Imbalance of macrophage cholesterol homeostasis is of great importance to atherosclerotic progression. This is mostly because plaque formation requires monocyte infiltration, resulting in generation of vast proinflammatory factors and chemokines, and further development into atherogenic foam cells caused by excessive uptake of modified low-density lipoprotein (LDL) particles. AMPK has been shown to prevent cholesterol accumulation in macrophages through promoting cholesterol efflux to high-density lipoprotein (HDL), causing a marked decrease in atherosclerotic plaque in ApoE^{-/} mice^{97,98}. The beneficial effect of AMPK may be related to the upregulation of ATP-binding cassette sub-family G member 1 (ABCGI) and ATP-binding cassette transporter A1 (ABCAI) gene expression accompanying with increased liver X receptor α (LXR α) expression (Fig. 3). Recently, our results also found out that AMPK predominates in macrophage uptake of cholesterol mediated by oxidized LDL (oxLDL) through downregulation of lectin like oxidized low-density lipoprotein receptor 1 (LOX-1) expression⁹⁹. Similar results were obtained by several other groups with AICAR or berberine 100,101 . The underlying mechanism is probably the decreased

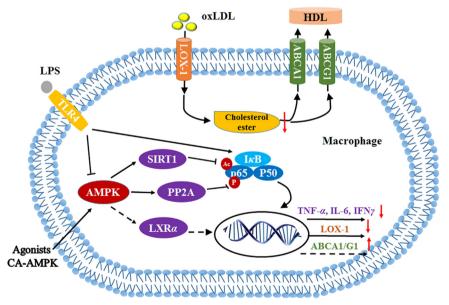


Figure 3 AMPK participates in regulation of macrophage cholesterol homeostasis and lipopolysaccharide (LPS)-stimulated production of inflammation factors. AMPK activation by various agonists (A769662, salicylic acid and AICAR, etc.) or overexpressing constitutively active AMPK upregulates the expression of cholesterol transporter ABCA1 or ABCG1, which may be related to increased expression of nuclear transcription factor LXRα, resulting in accumulated cholesterol efflux to mature HDL. In the meantime, increased PP2A activity stimulated by AMPK causes p65 dephosphorylation and subsequent decreased expression of scavenger receptor LOX-1, resulting in reduced oxLDL uptake and cholesterol accumulation in further. In addition, LPS stimulation can downregulate AMPK activity in macrophage. AMPK activation by pharmacological agonists blocks LPS-stimulated increase of secretary proinflammatory factors by SIRT1-mediated deacetylation of p65.

Ser536 phosphorylation of nuclear transcription factor NF- κ B p65 induced by enhanced protein phosphatase 2A (PP2A) activity ⁹⁹ (Fig. 3). The PP2A B56 γ subunit can be directly phosphorylated at Ser298 and Ser336 by AMPK ¹⁰². Nevertheless, recent studies from Zou's group demonstrated that there was no difference between macrophage-specific AMPK deficiency in $ApoE^{-/-}$ mice and corresponding $ApoE^{-/-}$ mice in western diet—induced atherosclerotic plaque formation and plaque instability, which robustly questions the role of macrophage AMPK in atherosclerosis ^{103,104}.

Simultaneously, AMPK seems to participate in inflammatory cytokine release in macrophages, since reduced AMPK activity was found in lipopolysaccharide (LPS)-stimulated macrophages¹⁰⁵. Accordingly, either activating AMPK in macrophages by pharmacological agonists or constitutive expression of active AMPK attenuates LPS-induced proinflammatory factors, whereas the level of anti-inflammatory cytokine increases, which may be mediated by the reduced acetylation and transcription activity of NF-kB induced by Sirtuin 1 (SIRT1)^{106–108} (Fig. 3), Moreover, AMPK-mediated activation of nucleotide-binding domain and leucine-rich repeat containing protein 3 (NLRP3) inflammasome was demonstrated to be involved in both saturated fatty acid-induced inflammation in macrophages and the anti-inflammatory effect of monounsaturated fatty acid 109-110. Likewise, the NLRP3 inflammasome can also be activated by crystalline cholesterol, an endogenous risk in atherosclerotic progression¹¹¹. However, whether AMPK can regulate this process remains to be addressed.

Vascular dysfunction is acknowledged as an early stage in atherosclerosis and is mainly caused by infiltration of immune cells in the vascular wall, inflammation, oxidative stress, impaired NO bioavailability and endothelial cell apoptosis. All these contributors may be concurrent with a reduction of AMPK activity in aortic endothelium¹¹². Consequently, activating vascular AMPK, then altering all above contributors, is probably an effective way to maintain cardiovascular health. It has been demonstrated that expressing constitutively active AMPK in cultured human aortic endothelial cells inhibit TNFαstimulated leukocyte adhesion associated with reduced monocyte chemotactic protein 1 (MCP-1) secretion¹¹³. AMPK activation by adiponectin reverses palmitate-induced ROS production and following mitogen-activated protein kinase p38-mediated apoptosis in endothelial cells¹¹⁴. Metformin was found to reduce oxidative stress, increase NO bioavailability and restore endothelial function through activation of AMPK/peroxisome proliferator-activated receptor δ (PPAR δ) pathway¹¹⁵. Several other direct or indirect AMPK agonists were also verified to enhance vascular function in succession 116. In addition, activation of AMPK promotes vasorelaxation in an endothelium and NO-independent manner, suggesting a direct effect of AMPK on vascular smooth muscle cells (VSMC) apart from endothelium¹¹⁷. Recently, two papers from Zou's laboratory were published in succession which revealed that specific AMPKa1 knockout in mouse VSMC promoted western diet-induced aortic calcification while specific AMPKα2 deletion in mouse VSMC induced VSMC phenotypic switching and therefore affected atherosclerotic plaque stability 103,10

2.4. AMPK in myocardial ischemia/reperfusion injury

Myocardial ischemia is mainly caused by an interruption in the coronary blood supply, resulting in detrimental effects such as cardiomyocyte death and cardiac dysfunction. Prompt reperfusion restores myocardial blood flow and oxygen supply, but the process of reperfusion brings about myocardial injury as well. The combination

of injury incurred during acute myocardial ischemia and reperfusion following ischemia is therefore named ischemia/reperfusion injury. During ischemia, deficiency of oxygen results in increasing anaerobic metabolism, that is increasing glycolytic pathway instead of oxidative phosphorylation ¹¹⁸. Although reperfusion of the myocardium can temporally maintain cardiomyocyte viability, metabolic alteration happens since glucose oxidation recovers much slower than fatty acid (FA) oxidation ¹¹⁹. These alterations, including increased lactate production, proton generation and decreased intracellular pH, can destroy cardiac efficiency and function ^{120–122}.

It has been suggested that AMPK is activated during ischemia and reperfusion due to the depletion of ATP and the increased activity of AMPK kinases¹²³. AMPK facilitates the delivery of fatty acids *via* increased cardiac lipoprotein lipase (LPL) activity and increased CD36 expression in membrane^{124,125}. The availability of fatty acids accelerates rates of their utilization, which is also promoted by AMPK activation due to decreased ACC activity, thus reducing malonyl-CoA levels. Enhanced fatty acid oxidation during ischemia and reperfusion suppresses glucose oxidative phosphorylation, which is detrimental to the ischemic myocardium¹²². Metabolic and pharmacological activation of AMPK promotes GLUT4 translocation to the sarcolemmal membrane⁶², whereas glucose uptake remains unchanged despite the AMPK activation after ischemia¹²⁶. AMPK can also increase phosphofructokinase-2 phosphorylation, resulting in enhanced glycolysis during ischemia¹²⁷.

It seems that ischemia-induced AMPK activation is harmful to the heart due to the stimulation of fatty acid oxidation or the glycolytic pathway instead of glucose oxidation. Chang et al. 128 suggested that berberine exerted cardioprotective effects by depressing AMPK activity in ischemic areas of rat heart, whereas AMPK was activated in the non-ischemic areas. In fact, AMPK is more like an adaptive response to satisfy the need of ischemic myocardium for ATP and protect the heart from oxygen deprivation. Brief periods of ischemia preconditioning have been confirmed to provide significant protection to the heart from ischemic injury. AMPK is shown to be involved in the setting of ischemic preconditioning and promotes glucose uptake 129. Considerable evidence demonstrates that AMPK stimulation prevents postischemic cardiac dysfunction and cell apoptosis upon reperfusion. For example, when isolated hearts from an AMPK α 2 kinase dead transgenic mice suffered ischemia attack, cardiac function declined, implying that AMPK activation is necessary for the heart to withstand an ischemia insult¹³⁰. Besides, treatment with A769662 prior or during ischemia diminishes the cardiac infarct size and decreases myocardial necrosis in an animal model^{131,132}. It has been suggested that adiponectin prevents the heart from ischemia/reperfusion injury in an AMPK and cyclooxygenase 2 (COX-2) dependent manner¹³³. Recently, some physiologically secreted factors and stress-inducible proteins, such as follistatinlike 1, C1q/TNF related protein 9, antithrombin, omentin and sestrin 2, have also been discovered to benefit the heart from ischemia/reperfusion injury by stimulating cardiac AMPK¹³⁴⁻¹³⁸.

2.5. AMPK in neurodegenerative disorder

Life-threatening, incurable diseases such as Alzheimer's disease (AD), Parkinson's disease (PD) and Huntington's disease (HD) are well-known examples of neurodegenerative disorder, defined as a progressive loss of neuronal structure and function. AD is the most familiar dementia, featuring senile plaques and intracellular neurofibrillary tangles, primarily caused by accumulation of misfolding $A\beta$ peptides and tau hyperphosphorylation in cortical

and hippocampal brain regions ¹³⁹. PD is generally recognized as neurodegenerative motor impairment. PD pathology includes a lack of dopaminergic neurons in the substantia nigra and clumps of α -synuclein, also called Lewy body. One of the main contributors to PD is mitochondrial dysfunction associated with a mutated gene ¹⁴⁰. HD is an age-related disorder involved in both movement and cognizance, induced by CAG triplet repeat expansion in exon 1 of the Htt gene. The major pathological change is the severe decrease of neurons used to synthesize enkephalin and γ -aminobutyric acid (GABA)¹⁴¹.

In mouse models of AD, PD or HD, pathological activation of brain AMPK has been demonstrated 141-143, consistent with the hypothesis that AMPK plays a key role in the development of senile dementia. However, many other reports have shown that AMPK seems to be a double-edged sword, either aggravating or alleviating neurodegeneration under different circumstances.

In AD, it is well established that AMPK is an important modulator of A β generation. Resveratrol prevents A β accumulation through promotion of autophagy induced by AMPK activation and mTOR inhibition¹⁴⁴. Similar results were acquired with another AMPK agonist AICAR¹⁴⁵. Such effects may be associated with changes of neuronal cholesterol and sphingomyelin contents and APP distribution in membrane 139. In addition, Greco et al. 146 identified that AMPK activation is related to the effect of leptin in neuronal cells, including reduction of tau phosphorylation which could lead to neurofibrillary tangles. On the contrary, some studies pointed out that AMPK inhibition by compound C could improve long-term potentiation (LTP) and alleviate impairments induced by amyloid beta¹⁴⁷. Moreover, chronic treatment with metformin was reported to have beneficial effects in females but to enhance memory dysfunction in males, suggesting that the effect of AMPK activation on neuronal cells may be gender-dependent 148

In PD, AMPK activation mitigates dopaminergic dysfunction in *Drosophila* models¹⁴⁹. The neuroprotective effect of ghrelin, a gut hormone, during calorie restriction was mediated by the AMPK signaling pathway¹⁵⁰. On the contrary, inhibition of AMPK by compound C in SH-SY5Y cells supplemented with MPP resulted in neuronal cell death¹⁴³. It was suggested that AMPK cooperates with parkin, an important modulator to maintain mitochondrial homeostasis, which explains why AMPK activation may be beneficial in PD¹⁵¹. However, Kim et al.¹⁵² reported that PARP triggers the degeneration of dopaminergic neurons through activation of AMPK as well, adding the complexity of the roles for AMPK in PD.

It was especially highlighted in HD that overactivation of AMPK by high-dose AICAR in striatal neurons facilitated neuronal loss and formation of Htt aggregates¹⁴¹. The underlying mechanism may be related to AMPK as sensor of oxidative stress to elicit neuronal atrophy¹⁵³. However, Vazquez-Manrique et al.¹⁵⁴ suggested that treatment of HD with metformin may be protective. A potential explanation of the controversial results was that AMPK activation may be beneficial in the onset of HD.

3. Conclusions and perspectives

AMPK has evolved to sense diversified energy and metabolic stress such as produced by hormones, cytokines, growth factors, sheer stress, hypoxia, some xenobiotics, etc. Simultaneously, it may help organisms to survive sudden or chronic stress through regulating a great assay of downstream targets involving glucose, fatty acids, cholesterol and amino acid metabolism,

glucose transport, mitochondrial function, cell growth, etc. The evidence to date indicates that AMPK is implicated in various human diseases. In the early stage, AMPK was suggested as a prime drug target for type 2 diabetes since several agonists displayed dramatic therapeutic potential in this disease. Later, it was exciting to discover the beneficial effect of metformin on cancer. Thus, the role of AMPK in tumorigenesis draws significant attention. In addition, other human diseases like atherosclerosis, myocardial ischemia/reperfusion injury and neurodegenerative disorder are all closely related to metabolism and inflammation processes in which AMPK has been identified to be a critical contributor. However, further studies are required to investigate the underlying molecular mechanisms. Noteworthy, a paradoxical role of AMPK was observed in these diseases. There are four possible reasons. First, indirect AMPK agonists such as AICAR and biguandes were used in most studies to demonstrate the beneficial effects of AMPK activation, even though the direct agonist A769662 may exert its effects in an AMPK-independent way under certain circumstances. Second, it should be noted that LKB1 has at least 13 other substrates other than AMPK¹⁵⁵. When referring to the knockout of LKB1 in mice, some observed phenomena may be mediated by other targets but not AMPK. Third, the widely used AMPK inhibitor, compound C, has also been reported to be highly nonspecific 156. Fourth, AMPK is such a sensitive sensor to various stresses and its multi-subunit structure and complicated activity regulatory mechanism bring complexity to understanding the role of AMPK in these diseases. Therefore, appropriate genetic models and tissue- or isomer-specifically direct AMPK agonists are desperately needed to differentiate the functions of AMPK in these human diseases.

Acknowledgments

This work was supported by grants from National Natural Sciences Foundation of China (NSFC, Grant Nos. 81273514, 91539126 and 81302827) and grants from Innovation Engineering of Chinese Academy of Agricultural Sciences (No. 125161015000150013).

References

- Beg ZH, Allmann DW, Gibson DM. Modulation of 3-hydroxy-3methylglutaryl coenzyme A reductase activity with cAMP and wth protein fractions of rat liver cytosol. *Biochem Biophys Res Commun* 1973:54:1362–9.
- Carlson CA, Kim KH. Regulation of hepatic acetyl coenzyme A carboxylase by phosphorylation and dephosphorylation. *J Biol Chem* 1973;248:378–80.
- Carling D, Clarke PR, Zammit VA, Hardie DG. Purification and characterization of the AMP-activated protein kinase. Copurification of acetyl-CoA carboxylase kinase and 3-hydroxy-3-methylglutaryl-CoA reductase kinase activities. Eur J Biochem 1989;186:129–36.
- Garton AJ, Campbell DG, Carling D, Hardie DG, Colbran RJ, Yeaman SJ. Phosphorylation of bovine hormone-sensitive lipase by the AMP-activated protein kinase. A possible antilipolytic mechanism. *Eur J Biochem* 1989;179:249–54.
- Carling D, Hardie DG. The substrate and sequence specificity of the AMP-activated protein kinase. phosphorylation of glycogen synthase and phosphorylase kinase. *Biochim Biophys Acta* 1989;1012:81–6.
- Dasgupta B, Chhipa RR. Evolving lessons on the complex role of AMPK in normal physiology and cancer. *Trends Pharmacol Sci* 2016;37:192–206.

 Yang Y, Atasoy D, Su HH, Sternson SM. Hunger states switch a flipflop memory circuit via a synaptic AMPK-dependent positive feedback loop. Cell 2011:146:992–1003.

- Srivastava AK, Qin X, Wedhas N, Arnush M, Linkhart TA, Chadwick RB, et al. Tumor necrosis factor-α augments matrix metalloproteinase-9 production in skeletal muscle cells through the activation of transforming growth factor-β-activated kinase 1 (TAK1)-dependent signaling pathway. J Biol Chem 2007;282:35113–24.
- Tamas P, Hawley SA, Clarke RG, Mustard KJ, Green K, Hardie DG, et al. Regulation of the energy sensor AMP-activated protein kinase by antigen receptor and Ca²⁺ in T lymphocytes. *J Exp Med* 2006;203:1665–70.
- Herrero-Martin G, Hoyer-Hansen M, Garcia-Garcia C, Fumarola C, Farkas T, Lopez-Rivas A, et al. TAK1 activates AMPK-dependent cytoprotective autophagy in TRAIL-treated epithelial cells. *EMBO J* 2009;28:677–85.
- Grahame Hardie D. Regulation of AMP-activated protein kinase by natural and synthetic activators. Acta Pharm Sin B 2016;6:1–19.
- Warburg O. On respiratory impairment in cancer cells. Science 1956;124:269–70.
- Levine AJ, Puzio-Kuter AM. The control of the metabolic switch in cancers by oncogenes and tumor suppressor genes. *Science* 2010;330:1340

 –4.
- Jones RG, Plas DR, Kubek S, Buzzai M, Mu J, Xu Y, et al. AMPactivated protein kinase induces a p53-dependent metabolic checkpoint. Mol Cell 2005;18:283–93.
- Liang J, Shao SH, Xu ZX, Hennessy B, Ding Z, Larrea M, et al. The energy sensing LKB1-AMPK pathway regulates p27^{kip1} phosphorylation mediating the decision to enter autophagy or apoptosis. *Nat Cell Biol* 2007;9:218–24.
- Rattan R, Giri S, Singh AK, Singh I. 5-Aminoimidazole-4-carboxamide-1-β-D-ribofuranoside inhibits cancer cell proliferation in vitro and in vivo via AMP-activated protein kinase. J Biol Chem 2005;280:39582–93.
- Zheng B, Jeong JH, Asara JM, Yuan YY, Granter SR, Chin L, et al. Oncogenic B-RAF negatively regulates the tumor suppressor LKB1 to promote melanoma cell proliferation. *Mol Cell* 2009;33:237–47.
- Shen CH, Yuan P, Perez-Lorenzo R, Zhang Y, Lee SX, Ou Y, et al. Phosphorylation of BRAF by AMPK impairs BRAF-KSR1 association and cell proliferation. *Mol Cell* 2013;52:161–72.
- Fogarty S, Ross FA, Vara Ciruelos D, Gray A, Gowans GJ, Hardie DG. AMPK causes cell cycle arrest in LKB1-deficient cells via activation of CAMKK2. Mol Cancer Res 2016;14:683–95.
- Gwinn DM, Shackelford DB, Egan DF, Mihaylova MM, Mery A, Vasquez DS, et al. AMPK phosphorylation of raptor mediates a metabolic checkpoint. *Mol Cell* 2008;30:214–26.
- Scaglia N, Tyekucheva S, Zadra G, Photopoulos C, Loda M. De novo fatty acid synthesis at the mitotic exit is required to complete cellular division. *Cell Cycle* 2014;13:859–68.
- Murtola TJ, Syvala H, Pennanen P, Blauer M, Solakivi T, Ylikomi T, et al. The importance of LDL and cholesterol metabolism for prostate epithelial cell growth. *PLoS One* 2012;7:e39445.
- 23. DeRan M, Yang J, Shen CH, Peters EC, Fitamant J, Chan P, et al. Energy stress regulates hippo-YAP signaling involving AMPK-mediated regulation of angiomotin-like 1 protein. *Cell Rep* 2014;9:495–503.
- 24. Mo JS, Meng Z, Kim YC, Park HW, Hansen CG, Kim S, et al. Cellular energy stress induces AMPK-mediated regulation of YAP and the Hippo pathway. *Nat Cell Biol* 2015;17:500–10.
- Wang W, Xiao ZD, Li X, Aziz KE, Gan B, Johnson RL, et al. AMPK modulates Hippo pathway activity to regulate energy homeostasis. *Nat Cell Biol* 2015;17:490–9.
- **26.** Faubert B, Boily G, Izreig S, Griss T, Samborska B, Dong Z, et al. AMPK is a negative regulator of the Warburg effect and suppresses tumor growth *in vivo*. *Cell Metab* 2013;**17**:113–24.
- Faubert B, Vincent EE, Griss T, Samborska B, Izreig S, Svensson RU, et al. Loss of the tumor suppressor LKB1 promotes metabolic

- reprogramming of cancer cells *via* HIF-1α. *Proc Natl Acad Sci U S A* 2014:**111**:2554–9.
- Yan M, Gingras MC, Dunlop EA, Nouet Y, Dupuy F, Jalali Z, et al. The tumor suppressor folliculin regulates AMPK-dependent metabolic transformation. *J Clin Invest* 2014;124:2640–50.
- Possik E, Jalali Z, Nouet Y, Yan M, Gingras MC, Schmeisser K, et al. Folliculin regulates ampk-dependent autophagy and metabolic stress survival. *PLoS Genet* 2014;10:e1004273.
- Rios M, Foretz M, Viollet B, Prieto A, Fraga M, Costoya JA, et al. AMPK activation by oncogenesis is required to maintain cancer cell proliferation in astrocytic tumors. *Cancer Res* 2013;73:2628–38.
- Laderoute KR, Calaoagan JM, Chao WR, Dinh D, Denko N, Duellman S, et al. 5'-AMP-activated protein kinase (AMPK) supports the growth of aggressive experimental human breast cancer tumors. J Biol Chem 2014;289:22850–64.
- Borgdorff V, Rix U, Winter GE, Gridling M, Muller AC, Breitwieser FP, et al. A chemical biology approach identifies AMPK as a modulator of melanoma oncogene MITF. Oncogene 2014;33:2531–9.
- Tennakoon JB, Shi Y, Han JJ, Tsouko E, White MA, Burns AR, et al. Androgens regulate prostate cancer cell growth *via* an AMPK-PGC-1α-mediated metabolic switch. *Oncogene* 2014;33:5251–61.
- 34. Fisher KW, Das B, Kim HS, Clymer BK, Gehring D, Smith DR, et al. AMPK promotes aberrant PGC1β expression to support human colon tumor cell survival. *Mol Cell Biol* 2015;35:3866–79.
- Fox MM, Phoenix KN, Kopsiaftis SG, Claffey KP. AMP-activated protein kinase α 2 isoform suppression in primary breast cancer alters AMPK growth control and apoptotic signaling. *Genes Cancer* 2013;4:3–14.
- Phoenix KN, Devarakonda CV, Fox MM, Stevens LE, Claffey KP. AMPKα2 suppresses murine embryonic fibroblast transformation and tumorigenesis. *Genes Cancer* 2012;3:51–62.
- Ross FA, MacKintosh C, Hardie DG. AMP-activated protein kinase: a cellular energy sensor that comes in 12 flavours. Febs J 2016;283:2987–3001.
- 38. Monteverde T, Muthalagu N, Port J, Murphy DJ. Evidence of cancer-promoting roles for AMPK and related kinases. *FEBS J* 2015;282:4658–71.
- Huang X, Wullschleger S, Shpiro N, McGuire VA, Sakamoto K, Woods YL, et al. Important role of the LKB1–AMPK pathway in suppressing tumorigenesis in PTEN-deficient mice. *Biochem J* 2008;412:211–21.
- 40. Rosilio C, Lounnas N, Nebout M, Imbert V, Hagenbeek T, Spits H, et al. The metabolic perturbators metformin, phenformin and AICAR interfere with the growth and survival of murine PTEN-deficient T cell lymphomas and human T-ALL/T-LL cancer cells. *Cancer Lett* 2013;336:114–26.
- El-Masry OS, Brown BL, Dobson PR. Effects of activation of AMPK on human breast cancer cell lines with different genetic backgrounds. Oncol Lett 2012;3:224

 –8.
- Buzzai M, Jones RG, Amaravadi RK, Lum JJ, DeBerardinis RJ, Zhao F, et al. Systemic treatment with the antidiabetic drug metformin selectively impairs p53-deficient tumor cell growth. Cancer Res 2007;67:6745–52.
- 43. Wu C, Guo Y, Su Y, Zhang X, Luan H, Zhang X, et al. Cordycepin activates AMP-activated protein kinase (AMPK) via interaction with the γ1 subunit. J Cell Mol Med 2014;18:293–304.
- Vincent EE, Coelho PP, Blagih J, Griss T, Viollet B, Jones RG. Differential effects of AMPK agonists on cell growth and metabolism. *Oncogene* 2015;34:3627–39.
- 45. Nair V, Sreevalsan S, Basha R, Abdelrahim M, Abudayyeh A, Rodrigues Hoffman A, et al. Mechanism of metformin-dependent inhibition of mammalian target of rapamycin (mTOR) and Ras activity in pancreatic cancer: role of specificity protein (Sp) transcription factors. *J Biol Chem* 2014;289:27692–701.
- 46. Liu X, Chhipa RR, Pooya S, Wortman M, Yachyshin S, Chow LM, et al. Discrete mechanisms of mTOR and cell cycle regulation by AMPK agonists independent of AMPK. *Proc Natl Acad Sci U S A* 2014;111:E435–44.

- 47. Janzer A, German NJ, Gonzalez-Herrera KN, Asara JM, Haigis MC, Struhl K. Metformin and phenformin deplete tricarboxylic acid cycle and glycolytic intermediates during cell transformation and NTPs in cancer stem cells. *Proc Natl Acad Sci U S A* 2014;111:10574–9.
- 48. Honjo S, Ajani JA, Scott AW, Chen Q, Skinner HD, Stroehlein J, et al. Metformin sensitizes chemotherapy by targeting cancer stem cells and the mTOR pathway in esophageal cancer. *Int J Oncol* 2014;45:567–74.
- Hemminki A, Markie D, Tomlinson I, Avizienyte E, Roth S, Loukola A, et al. A serine/threonine kinase gene defective in Peutz-Jeghers syndrome. *Nature* 1998:391:184–7.
- Algire C, Amrein L, Bazile M, David S, Zakikhani M, Pollak M. Diet and tumor LKB1 expression interact to determine sensitivity to antineoplastic effects of metformin in vivo. Oncogene 2011;30:1174–82.
- Shackelford DB, Abt E, Gerken L, Vasquez DS, Seki A, Leblanc M, et al. LKB1 inactivation dictates therapeutic response of non-small cell lung cancer to the metabolism drug phenformin. *Cancer Cell* 2013;23:143–58.
- Birk JB, Wojtaszewski JF. Predominant α2/β2/γ3 AMPK activation during exercise in human skeletal muscle. J Physiol 2006;577:1021– 32
- DeFronzo RA, Tripathy D. Skeletal muscle insulin resistance is the primary defect in type 2 diabetes. *Diabetes Care* 2009;32 Suppl 2: \$157-63
- 54. Kristensen JM, Treebak JT, Schjerling P, Goodyear L, Wojtaszewski JF. Two weeks of metformin treatment induces AMPK-dependent enhancement of insulin-stimulated glucose uptake in mouse soleus muscle. Am J Physiol Endocrinol Metab 2014;306:E1099–109.
- 55. Chopra I, Li HF, Wang H, Webster KA. Phosphorylation of the insulin receptor by AMP-activated protein kinase (AMPK) promotes ligand-independent activation of the insulin signalling pathway in rodent muscle. *Diabetologia* 2012;55:783–94.
- Jakobsen SN, Hardie DG, Morrice N, Tornqvist HE. 5'-AMPactivated protein kinase phosphorylates IRS-1 on Ser-789 in mouse C2C12 myotubes in response to 5-aminoimidazole-4-carboxamide riboside. J Biol Chem 2001;276:46912–6.
- 57. O'Neill HM, Maarbjerg SJ, Crane JD, Jeppesen J, Jorgensen SB, Schertzer JD, et al. AMP-activated protein kinase (AMPK) β1β2 muscle null mice reveal an essential role for AMPK in maintaining mitochondrial content and glucose uptake during exercise. *Proc Natl Acad Sci U S A* 2011;108:16092–7.
- 58. Beck Jorgensen S, O'Neill HM, Hewitt K, Kemp BE, Steinberg GR. Reduced AMP-activated protein kinase activity in mouse skeletal muscle does not exacerbate the development of insulin resistance with obesity. *Diabetologia* 2009;52:2395–404.
- Barnes BR, Marklund S, Steiler TL, Walter M, Hjalm G, Amarger V, et al. The 5'-AMP-activated protein kinase γ3 isoform has a key role in carbohydrate and lipid metabolism in glycolytic skeletal muscle. J Biol Chem 2004;279:38441–7.
- 60. Steinberg GR, O'Neill HM, Dzamko NL, Galic S, Naim T, Koopman R, et al. Whole body deletion of AMP-activated protein kinase β2 reduces muscle AMPK activity and exercise capacity. *J Biol Chem* 2010;285:37198–209.
- Brunmair B, Staniek K, Gras F, Scharf N, Althaym A, Clara R, et al. Thiazolidinediones, like metformin, inhibit respiratory complex I: a common mechanism contributing to their antidiabetic actions?. *Diabetes* 2004:53:1052–9.
- Cartee GD. Roles of TBC1D1 and TBC1D4 in insulin- and exercisestimulated glucose transport of skeletal muscle. *Diabetologia* 2015;58:19–30.
- 63. Frosig C, Pehmoller C, Birk JB, Richter EA, Wojtaszewski JF. Exercise-induced TBC1D1 Ser237 phosphorylation and 14-3-3 protein binding capacity in human skeletal muscle. *J Physiol* 2010;588:4539–48.
- 64. Shaw RJ, Lamia KA, Vasquez D, Koo SH, Bardeesy N, Depinho RA, et al. The kinase LKB1 mediates glucose homeostasis in liver and therapeutic effects of metformin. *Science* 2005;310:1642–6.

- **65.** Foretz M, Hebrard S, Leclerc J, Zarrinpashneh E, Soty M, Mithieux G, et al. Metformin inhibits hepatic gluconeogenesis in mice independently of the LKB1/AMPK pathway *via* a decrease in hepatic energy state. *J Clin Invest* 2010;**120**:2355–69.
- Miller RA, Chu Q, Xie J, Foretz M, Viollet B, Birnbaum MJ. Biguanides suppress hepatic glucagon signalling by decreasing production of cyclic AMP. *Nature* 2013;494:256–60.
- Madiraju AK, Erion DM, Rahimi Y, Zhang XM, Braddock DT, Albright RA, et al. Metformin suppresses gluconeogenesis by inhibiting mitochondrial glycerophosphate dehydrogenase. *Nature* 2014:510:542-6
- 68. Fullerton MD, Galic S, Marcinko K, Sikkema S, Pulinilkunnil T, Chen Z-P, et al. Single phosphorylation sites in Acc1 and Acc2 regulate lipid homeostasis and the insulin-sensitizing effects of metformin. *Nat Med* 2013;19:1649–54.
- 69. Cao J, Meng S, Chang E, Beckwith-Fickas K, Xiong L, Cole RN, et al. Low concentrations of metformin suppress glucose production in hepatocytes through AMP-activated protein kinase (AMPK). *J Biol Chem* 2014;289:20435–46.
- 70. Meng S, Cao J, He Q, Xiong L, Chang E, Radovick S, et al. Metformin activates AMP-activated protein kinase by promoting formation of the $\alpha\beta\gamma$ heterotrimeric complex. *J Biol Chem* 2015;**290**:3793–802.
- Johanns M, Lai YC, Hsu MF, Jacobs R, Vertommen D, Van Sande J, et al. AMPK antagonizes hepatic glucagon-stimulated cyclic AMP signalling via phosphorylation-induced activation of cyclic nucleotide phosphodiesterase 4B. Nat Commun 2016;7:10856.
- Rainero E, Cianflone C, Porporato PE, Chianale F, Malacarne V, Bettio V, et al. The diacylglycerol kinase α/atypical PKC/β1 integrin pathway in SDF-1α mammary carcinoma invasiveness. *PLoS One* 2014;9:e97144.
- Turban S, Hajduch E. Protein kinase C isoforms: mediators of reactive lipid metabolites in the development of insulin resistance. FEBS Lett 2011;585:269–74.
- 74. Gaidhu MP, Fediuc S, Anthony NM, So M, Mirpourian M, Perry RL, et al. Prolonged AICAR-induced AMP-kinase activation promotes energy dissipation in white adipocytes: novel mechanisms integrating HSL and ATGL. J Lipid Res 2009;50:704–15.
- Li Y, Xu S, Mihaylova MM, Zheng B, Hou X, Jiang B, et al. AMPK phosphorylates and inhibits SREBP activity to attenuate hepatic steatosis and atherosclerosis in diet-induced insulin-resistant mice. *Cell Metab* 2011;13:376–88.
- Gaidhu MP, Fediuc S, Ceddia RB. 5-Aminoimidazole-4-carboxamide-1-β-D-ribofuranoside-induced AMP-activated protein kinase phosphorylation inhibits basal and insulin-stimulated glucose uptake, lipid synthesis, and fatty acid oxidation in isolated rat adipocytes. *J Biol Chem* 2006;281:25956–64.
- Gaidhu MP, Frontini A, Hung S, Pistor K, Cinti S, Ceddia RB. Chronic AMP-kinase activation with AICAR reduces adiposity by remodeling adipocyte metabolism and increasing leptin sensitivity. J Lipid Res 2011;52:1702–11.
- Koh HJ, Hirshman MF, He H, Li Y, Manabe Y, Balschi JA, et al. Adrenaline is a critical mediator of acute exercise-induced AMPactivated protein kinase activation in adipocytes. *Biochem J* 2007;403:473–81.
- Ahmadian M, Abbott MJ, Tang T, Hudak CS, Kim Y, Bruss M, et al. Desnutrin/ATGL is regulated by AMPK and is required for a brown adipose phenotype. *Cell Metab* 2011;13:739–48.
- Daval M, Diot-Dupuy F, Bazin R, Hainault I, Viollet B, Vaulont S, et al. Anti-lipolytic action of AMP-activated protein kinase in rodent adipocytes. J Biol Chem 2005;280:25250–7.
- 81. Wu Y, Song P, Zhang W, Liu J, Dai X, Liu Z, et al. Activation of AMPKα2 in adipocytes is essential for nicotine-induced insulin resistance in vivo. Nat Med 2015;21:373–82.
- Granot Z, Swisa A, Magenheim J, Stolovich-Rain M, Fujimoto W, Manduchi E, et al. LKB1 regulates pancreatic β cell size, polarity, and function. *Cell Metab* 2009;10:296–308.

83. Fu A, Ng AC, Depatie C, Wijesekara N, He Y, Wang GS, et al. Loss of Lkb1 in adult β cells increases β cell mass and enhances glucose tolerance in mice. *Cell Metab* 2009;**10**:285–95.

- 84. Sun G, Tarasov AI, McGinty JA, French PM, McDonald A, Leclerc I, et al. LKB1 deletion with the RIP2.Cre transgene modifies pancreatic β-cell morphology and enhances insulin secretion in vivo. Am J Physiol Endocrinol Metab 2010;298:E1261–73.
- 85. Sun G, Tarasov AI, McGinty J, McDonald A, da Silva Xavier G, Gorman T, et al. Ablation of AMP-activated protein kinase α1 and α2 from mouse pancreatic β cells and RIP2.Cre neurons suppresses insulin release in vivo. Diabetologia 2010;53:924–36.
- 86. Kone M, Pullen TJ, Sun G, Ibberson M, Martinez-Sanchez A, Sayers S, et al. LKB1 and AMPK differentially regulate pancreatic β-cell identity. FASEB J 2014;28:4972–85.
- 87. Wang L, Khambu B, Zhang H, Yin XM. Autophagy in alcoholic liver disease, self-eating triggered by drinking. *Clin Res Hepatol Gastroenterol* 2015;39 Suppl 1:S2–6.
- Bachar-Wikstrom E, Wikstrom JD, Ariav Y, Tirosh B, Kaiser N, Cerasi E, et al. Stimulation of autophagy improves endoplasmic reticulum stress-induced diabetes. *Diabetes* 2013;62:1227–37.
- 89. Park SH, Ryu SY, Yu WJ, Han YE, Ji YS, Oh K, et al. Leptin promotes K_{ATP} channel trafficking by AMPK signaling in pancreatic β-cells. Proc Natl Acad Sci U S A 2013;110:12673–8.
- Kim JW, You YH, Ham DS, Yang HK, Yoon KH. The paradoxical effects of AMPK on insulin gene expression and glucose-induced insulin secretion. *J Cell Biochem* 2016;117:239–46.
- Yavari A, Stocker CJ, Ghaffari S, Wargent ET, Steeples V, Czibik G, et al. Chronic activation of γ2 AMPK induces obesity and reduces β cell function. *Cell Metab* 2016;23:821–36.
- Varela L, Horvath TL. Leptin and insulin pathways in POMC and AgRP neurons that modulate energy balance and glucose homeostasis. *EMBO Rep* 2012;13:1079–86.
- Andersson U, Filipsson K, Abbott CR, Woods A, Smith K, Bloom SR, et al. AMP-activated protein kinase plays a role in the control of food intake. J Biol Chem 2004;279:12005–8.
- 94. Minokoshi Y, Alquier T, Furukawa N, Kim YB, Lee A, Xue B, et al. AMP-kinase regulates food intake by responding to hormonal and nutrient signals in the hypothalamus. *Nature* 2004;428:569–74.
- Claret M, Smith MA, Batterham RL, Selman C, Choudhury AI, Fryer LG, et al. AMPK is essential for energy homeostasis regulation and glucose sensing by POMC and AgRP neurons. *J Clin Invest* 2007;117:2325–36.
- Oh TS, Cho H, Cho JH, Yu SW, Kim EK. Hypothalamic AMPKinduced autophagy increases food intake by regulating NPY and POMC expression. *Autophagy* 2016:1–17.
- Fullerton MD, Ford RJ, McGregor CP, LeBlond ND, Snider SA, Stypa SA, et al. Salicylate improves macrophage cholesterol homeostasis via activation of AMPK. J Lipid Res 2015;56:1025–33.
- 98. Li D, Wang D, Wang Y, Ling W, Feng X, Xia M. Adenosine monophosphate-activated protein kinase induces cholesterol efflux from macrophage-derived foam cells and alleviates atherosclerosis in apolipoprotein E-deficient mice. J Biol Chem 2010;285:33499–509.
- Chen B, Li J, Zhu H. AMP-activated protein kinase attenuates oxLDL uptake in macrophages through PP2A/NF-κB/LOX-1 pathway. Vasc Pharmacol 2016;85:1–10.
- 100. Namgaladze D, Kemmerer M, von Knethen A, Brune B. AICAR inhibits PPARγ1 during monocyte differentiation to attenuate inflammatory responses to atherogenic lipids. Cardiovasc Res 2013;98:479–87
- 101. Huang Z, Dong F, Li S, Chu M, Zhou H, Lu Z, et al. Berberine-induced inhibition of adipocyte enhancer-binding protein 1 attenuates oxidized low-density lipoprotein accumulation and foam cell formation in phorbol 12-myristate 13-acetate-induced macrophages. Eur J Pharmacol 2012;690:164–9.
- 102. Kim KY, Baek A, Hwang JE, Choi YA, Jeong J, Lee MS, et al. Adiponectin-activated AMPK stimulates dephosphorylation of AKT through protein phosphatase 2A activation. Cancer Res 2009;69:4018–26.

- 103. Cai Z, Ding Y, Zhang M, Lu Q, Wu S, Zhu H, et al. Ablation of adenosine monophosphate-activated protein kinase α1 in vascular smooth muscle cells promotes diet-induced atherosclerotic calcification in vivo. Circ Res 2016;119:422–33.
- 104. Ding Y, Zhang M, Zhang W, Lu Q, Cai Z, Song P, et al. AMP-activated protein kinase alpha 2 deletion induces VSMC phenotypic switching and reduces features of atherosclerotic plaque stability. Circ Res 2016;119:718–30.
- 105. Steinberg GR, Michell BJ, van Denderen BJ, Watt MJ, Carey AL, Fam BC, et al. Tumor necrosis factor α-induced skeletal muscle insulin resistance involves suppression of AMP-kinase signaling. *Cell Metab* 2006;4:465–74.
- 106. Jeong HW, Hsu KC, Lee JW, Ham M, Huh JY, Shin HJ, et al. Berberine suppresses proinflammatory responses through AMPK activation in macrophages. Am J Physiol Endocrinol Metab 2009;296:E955–64.
- 107. Wolf AM, Wolf D, Rumpold H, Enrich B, Tilg H. Adiponectin induces the anti-inflammatory cytokines IL-10 and IL-1RA in human leukocytes. *Biochem Biophys Res Commun* 2004;323:630–5.
- 108. Yang Z, Kahn BB, Shi H, Xue BZ. Macrophage alpha1 AMP-activated protein kinase (α1AMPK) antagonizes fatty acid-induced inflammation through SIRT1. *J Biol Chem* 2010;285:19051–9.
- 109. Wen H, Gris D, Lei Y, Jha S, Zhang L, Huang MT, et al. Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signaling. *Nat Immunol* 2011;12:408–15.
- 110. Finucane OM, Lyons CL, Murphy AM, Reynolds CM, Klinger R, Healy NP, et al. Monounsaturated fatty acid–enriched high-fat diets impede adipose NLRP3 inflammasome-mediated IL-1β secretion and insulin resistance despite obesity. *Diabetes* 2015;64:2116–28.
- 111. Duewell P, Kono H, Rayner KJ, Sirois CM, Vladimer G, Bauernfeind FG, et al. NLRP3 inflammasomes are required for atherogenesis and activated by cholesterol crystals. *Nature* 2010;464:1357–61.
- 112. Weikel KA, Cacicedo JM, Ruderman NB, Ido Y. Glucose and palmitate uncouple AMPK from autophagy in human aortic endothelial cells. Am J Physiol Cell Physiol 2015;308:C249–63.
- 113. Ewart MA, Kennedy S. AMPK and vasculoprotection. *Pharmacol Ther* 2011;131:242–53.
- 114. Kim JE, Song SE, Kim YW, Kim JY, Park SC, Park YK, et al. Adiponectin inhibits palmitate-induced apoptosis through suppression of reactive oxygen species in endothelial cells: involvement of cAMP/protein kinase A and AMP-activated protein kinase. J Endocrinol 2010;207:35–44.
- 115. Cheang WS, Tian XY, Wong WT, Lau CW, Lee SS, Chen ZY, et al. Metformin protects endothelial function in diet-induced obese mice by inhibition of endoplasmic reticulum stress through 5' adenosine monophosphate-activated protein kinase-peroxisome proliferator-activated receptor delta pathway. Arterioscler Thromb Vasc Biol 2014;34:830-6.
- Fullerton MD, Steinberg GR, Schertzer JD. Immunometabolism of AMPK in insulin resistance and atherosclerosis. *Mol Cell Endocrinol* 2013;366:224–34.
- 117. Miyabe M, Ohashi K, Shibata R, Uemura Y, Ogura Y, Yuasa D, et al. Muscle-derived follistatin-like 1 functions to reduce neointimal formation after vascular injury. *Cardiovasc Res* 2014;103:111–20.
- 118. Oliver MF, Opie LH. Effects of glucose and fatty acids on myocardial ischaemia and arrhythmias. *Lancet* 1994;**343**:155–8.
- 119. Lopaschuk GD, Wambolt RB, Barr RL. An imbalance between glycolysis and glucose oxidation is a possible explanation for the detrimental effects of high levels of fatty acids during aerobic reperfusion of ischemic hearts. J Pharmacol Exp Ther 1993;264:135–44.
- Lopaschuk GD. Alterations in fatty acid oxidation during reperfusion of the heart after myocardial ischemia. Am J Cardiol 1997;80:11A–16AA.
- 121. Liu Q, Docherty JC, Rendell JC, Clanachan AS, Lopaschuk GD. High levels of fatty acids delay the recovery of intracellular pH and cardiac efficiency in post-ischemic hearts by inhibiting glucose oxidation. J Am Coll Cardiol 2002;39:718–25.
- 122. Liu B, el Alaoui-Talibi Z, Clanachan AS, Schulz R, Lopaschuk GD. Uncoupling of contractile function from mitochondrial TCA cycle

- activity and MVO2 during reperfusion of ischemic hearts. Am J Physiol 1996;270:H72-80.
- Altarejos JY, Taniguchi M, Clanachan AS, Lopaschuk GD. Myocardial ischemia differentially regulates LKB1 and an alternate 5'-AMP-activated protein kinase kinase. J Biol Chem 2005;280:183–90.
- 124. Pulinilkunnil T, Puthanveetil P, Kim MS, Wang F, Schmitt V, Rodrigues B. Ischemia-reperfusion alters cardiac lipoprotein lipase. *Biochim Biophys Acta* 2010;1801:171–5.
- 125. Habets DD, Coumans WA, Voshol PJ, den Boer MA, Febbraio M, Bonen A, et al. AMPK-mediated increase in myocardial long-chain fatty acid uptake critically depends on sarcolemmal CD36. *Biochem Biophys Res Commun* 2007;355:204–10.
- 126. Omar MA, Fraser H, Clanachan AS. Ischemia-induced activation of AMPK does not increase glucose uptake in glycogen-replete isolated working rat hearts. Am J Physiol Heart Circ Physiol 2008;294:H1266–73.
- 127. Marsin AS, Bertrand L, Rider MH, Deprez J, Beauloye C, Vincent MF, et al. Phosphorylation and activation of heart PFK-2 by AMPK has a role in the stimulation of glycolysis during ischaemia. *Curr Biol* 2000:10:1247–55.
- 128. Chang W, Zhang M, Li J, Meng Z, Xiao D, Wei S, et al. Berberine attenuates ischemia-reperfusion injury via regulation of adenosine-5'-monophosphate kinase activity in both non-ischemic and ischemic areas of the rat heart. Cardiovasc Drugs Ther 2012;26:467–78.
- 129. Nishino Y, Miura T, Miki T, Sakamoto J, Nakamura Y, Ikeda Y, et al. Ischemic preconditioning activates AMPK in a PKC-dependent manner and induces GLUT4 up-regulation in the late phase of cardioprotection. *Cardiovasc Res* 2004;61:610–9.
- 130. Russell 3rd RR, Li J, Coven DL, Pypaert M, Zechner C, Palmeri M, et al. AMP-activated protein kinase mediates ischemic glucose uptake and prevents postischemic cardiac dysfunction, apoptosis, and injury. *J Clin Invest* 2004;114:495–503.
- 131. Kim AS, Miller EJ, Wright TM, Li J, Qi D, Atsina K, et al. A small molecule AMPK activator protects the heart against ischemiareperfusion injury. J Mol Cell Cardiol 2011;51:24–32.
- 132. Paiva MA, Rutter-Locher Z, Goncalves LM, Providencia LA, Davidson SM, Yellon DM, et al. Enhancing AMPK activation during ischemia protects the diabetic heart against reperfusion injury. Am J Physiol Heart Circ Physiol 2011;300:H2123–34.
- 133. Shibata R, Sato K, Pimentel DR, Takemura Y, Kihara S, Ohashi K, et al. Adiponectin protects against myocardial ischemia-reperfusion injury through AMPK- and COX-2-dependent mechanisms. *Nat Med* 2005;11:1096–103.
- 134. Morrison A, Chen L, Wang J, Zhang M, Yang H, Ma Y, et al. Sestrin2 promotes LKB1-mediated AMPK activation in the ischemic heart. FASEB J 2015;29:408–17.
- 135. Ma Y, Wang J, Gao J, Yang H, Wang Y, Manithody C, et al. Antithrombin up-regulates AMP-activated protein kinase signalling during myocardial ischaemia/reperfusion injury. *Thromb Haemost* 2015;113:338–49.
- 136. Kambara T, Shibata R, Ohashi K, Matsuo K, Hiramatsu-Ito M, Enomoto T, et al. C1q/tumor necrosis factor-related protein 9 protects against acute myocardial injury through an adiponectin receptor I-AMPK-dependent mechanism. Mol Cell Biol 2015;35:2173–85.
- 137. Ogura Y, Ouchi N, Ohashi K, Shibata R, Kataoka Y, Kambara T, et al. Therapeutic impact of follistatin-like 1 on myocardial ischemic injury in preclinical models. *Circulation* 2012;126:1728–38.
- 138. Kataoka Y, Shibata R, Ohashi K, Kambara T, Enomoto T, Uemura Y, et al. Omentin prevents myocardial ischemic injury through AMP-activated protein kinase- and Akt-dependent mechanisms. *J Am Coll Cardiol* 2014;63:2722–33.

- Cai Z, Yan LJ, Li K, Quazi SH, Zhao B. Roles of AMP-activated protein kinase in Alzheimer's disease. Neuromolecular Med 2012;14:1–14.
- 140. Corti O, Lesage S, Brice A. What genetics tells us about the causes and mechanisms of Parkinson's disease. *Physiol Rev* 2011;91:1161–218.
- 141. Ju TC, Chen HM, Lin JT, Chang CP, Chang WC, Kang JJ, et al. Nuclear translocation of AMPK-α1 potentiates striatal neurodegeneration in Huntington's disease. J Cell Biol 2011;194:209–27.
- 142. Mairet-Coello G, Courchet J, Pieraut S, Courchet V, Maximov A, Polleux F. The CAMKK2-AMPK kinase pathway mediates the synaptotoxic effects of $A\beta$ oligomers through Tau phosphorylation. *Neuron* 2013;**78**:94–108.
- 143. Choi JS, Park C, Jeong JW. AMP-activated protein kinase is activated in Parkinson's disease models mediated by 1-methyl-4phenyl-1,2,3,6-tetrahydropyridine. *Biochem Biophys Res Commun* 2010;391:147–51.
- 144. Vingtdeux V, Giliberto L, Zhao H, Chandakkar P, Wu Q, Simon JE, et al. AMP-activated protein kinase signaling activation by resveratrol modulates amyloid-β peptide metabolism. J Biol Chem 2010;285:9100–13.
- Won JS, Im YB, Kim J, Singh AK, Singh I. Involvement of AMPactivated-protein-kinase (AMPK) in neuronal amyloidogenesis. *Bio*chem Biophys Res Commun 2010;399:487–91.
- 146. Greco SJ, Sarkar S, Johnston JM, Tezapsidis N. Leptin regulates tau phosphorylation and amyloid through AMPK in neuronal cells. *Biochem Biophys Res Commun* 2009;380:98–104.
- 147. Ma T, Chen Y, Vingtdeux V, Zhao H, Viollet B, Marambaud P, et al. Inhibition of AMP-activated protein kinase signaling alleviates impairments in hippocampal synaptic plasticity induced by amyloid β. J Neurosci 2014;34:12230–8.
- 148. DiTacchio KA, Heinemann SF, Dziewczapolski G. Metformin treatment alters memory function in a mouse model of Alzheimer's disease. J Alzheimers Dis 2015;44:43–8.
- 149. Ng CH, Guan MS, Koh C, Ouyang X, Yu F, Tan EK, et al. AMP kinase activation mitigates dopaminergic dysfunction and mitochondrial abnormalities in Drosophila models of Parkinson's disease. J Neurosci 2012;32:14311–7.
- 150. Bayliss JA, Lemus MB, Stark R, Santos VV, Thompson A, Rees DJ, et al. Ghrelin-AMPK signaling mediates the neuroprotective effects of calorie restriction in Parkinson's disease. *J Neurosci* 2016;36:3049–63.
- 151. Hang L, Thundyil J, Lim KL. Mitochondrial dysfunction and Parkinson disease: a Parkin–AMPK alliance in neuroprotection. Ann N Y Acad Sci 2015;1350:37–47.
- 152. Kim TW, Cho HM, Choi SY, Suguira Y, Hayasaka T, Setou M, et al. ADP-ribose) polymerase 1 and AMP-activated protein kinase mediate progressive dopaminergic neuronal degeneration in a mouse model of Parkinson's disease. *Cell Death Dis* 2013;4:e919.
- 153. Ju TC, Chen HM, Chen YC, Chang CP, Chang C, Chern Y. AMPK-α1 functions downstream of oxidative stress to mediate neuronal atrophy in Huntington's disease. *Biochim Biophys Acta* 2014;1842:1668–80.
- 154. Vazquez-Manrique RP, Farina F, Cambon K, Dolores Sequedo M, Parker AJ, Millan JM, et al. AMPK activation protects from neuronal dysfunction and vulnerability across nematode, cellular and mouse models of Huntington's disease. *Hum Mol Genet* 2016;25:1043–58.
- 155. Lizcano JM, Goransson O, Toth R, Deak M, Morrice NA, Boudeau J, et al. LKB1 is a master kinase that activates 13 kinases of the AMPK subfamily, including MARK/PAR-1. EMBO J 2004;23:833–43.
- 156. Liu X, Chhipa RR, Nakano I, Dasgupta B. The AMPK inhibitor compound C is a potent AMPK-independent antiglioma agent. *Mol Cancer Ther* 2014;13:596–605.