

# Four Column Layout :: CHEAT SHEET

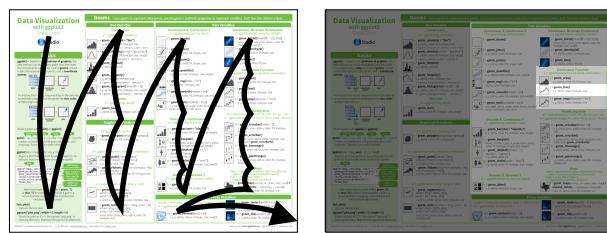
## Basics

**Thank you** for making a new cheatsheet for R! These cheatsheets have an important job:

**Cheatsheets make it easy for R users to look up useful information.**

Remember that the best cheatsheets are **visual**—not written—documents. Whenever possible use visual elements to make it easier for readers to find the information they need.

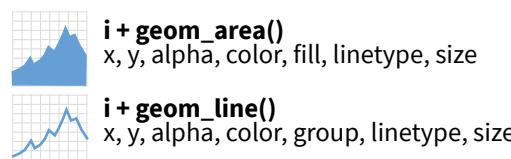
1. Use a **layout** that flows and makes it easy to zero in on specific topics.



2. Use **visualizations** to explain concepts quickly and concisely.



3. Use visual elements to make the sheet **scannable**.



4. Use visual **emphasis** (like color, size, and font weight) to make important information easy to find.

`dplyr::lag()` - Offset elements by 1  
`dplyr::lead()` - Offset elements by -1

## Layout Suggestions

Use headers, colors, and/or backgrounds to **separate or group together sections**.

Section 1      Section 2      Section 3

**Create a visual hierarchy.** Help users navigate the page with titles, subtitles, and subsubtitles

### Title

#### SUBTITLE

#### SUBSUBTITLE

**Fit sections to content.** Try several different layouts.

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Quickly identify content with a **package hexsticker** (if available)



## Copyright

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**YOUR LOGO**  
(optional)

## Useful Elements

### CODE

Where possible, use **code that works** when run.

```
ggplot(mpg, aes(hwy, cty)) +
  geom_point(aes(size = fl)) +
  geom_smooth(method = "lm")
```

Word balloons

can help explain

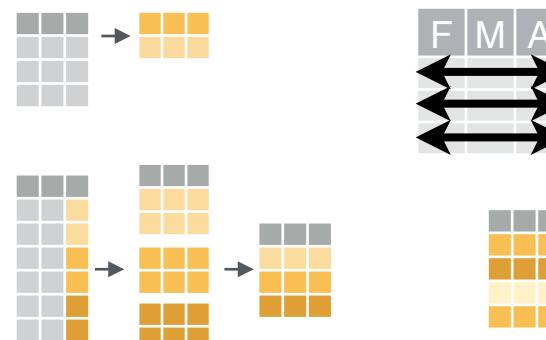
code

### ICONS

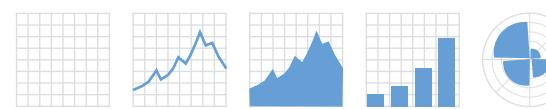
These are just **font awesome** characters



### MOCK TABLES



### MOCK GRAPHS



### TABLES

sub-option	description
citation_package	The LaTeX package to process
code_folding	Let readers to toggle the display of R
colortheme	Beamer color theme to use

## Logistics

### FONTS

This template uses several fonts: **Helvetica Neue, Menlo, Source Sans pro**, which you can acquire for free here, [www.fontsquirrel.com/fonts/source-sans-pro](http://www.fontsquirrel.com/fonts/source-sans-pro), and **Font Awesome**, which you can acquire here, [fortawesome.github.io/Font-Awesome/get-started/](https://fontawesome.github.io/Font-Awesome/get-started/)

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### KEYNOTE

I make my cheatsheets in **Apple Keynote**, and not latex or R Markdown, because presentation software makes it much easier to tweak the visual appearance of a document

### KEYNOTE TIPS

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# Three Column Layout: : CHEAT SHEET



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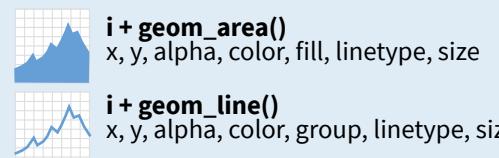
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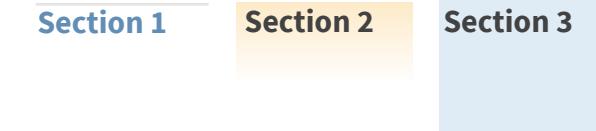
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## Manipulate Variables

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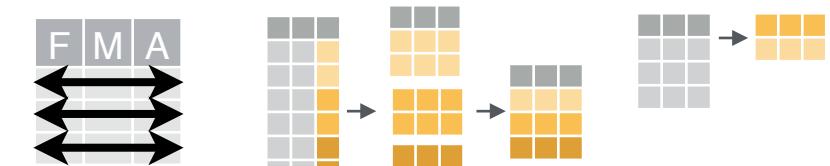
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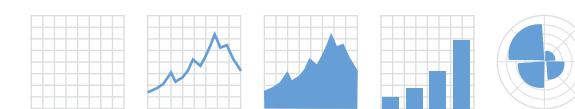


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# Arrow for R :: CHEAT SHEET

## Arrow

Apache Arrow is a multi-language toolbox for accelerated data interchange and processing. It specifies a standardized language-independent column-based memory format for flat and hierarchical data, organized for efficient analytic operations on modern hardware.

The **arrow** R package provides access to the Arrow C++ library from R, and supplies an interface with **dplyr** and other familiar R functions.

## Arrow Data Structures

**Table**: a tabular, column-oriented data structure capable of efficiently storing and processing large amounts of data with expanded column data types.

**Dataset**: similar to **Table** but with the capability to work on larger-than-memory data partitioned across multiple files.

You can convert an existing `data.frame` or `tibble` object into an **arrow Table**, and an **arrow Table** to a `data.frame` or `tibble` to view or work with the data in R:

```
library(arrow)
library(dplyr)

df_table <- arrow_table(df)

df <- as.data.frame(df_table)
df <- as_tibble(df_table)
```

## Read Individual Files

Read a data file from disk:

```
df <- read_parquet("file.parquet")
df <- read_feather("file.feather")
df <- read_csv_arrow("file.csv")
df <- read_json_arrow("file.json")
```

The **arrow** `read_*` functions return a `data.frame`, setting `as_data_frame = FALSE` returns an **arrow Table**.

## Read Multi-file Datasets

**arrow** defines `Dataset` objects for reading and writing very large files or sets of multi-files. The functions `open_dataset()` and `write_dataset()` enable analysis and processing of larger-than-memory data.

Read in multi-files from a directory:

```
open_dataset("folder")
```

Read in multi-files partitioned by year and month within a directory:

```
open_dataset("folder",
            partitioning = c("year", "month"))
```

The file format for `open_dataset()` is controlled by the `format` parameter, which has a default value of "parquet". Other supported formats include:

- "arrow"
- "feather" or "ipc" (aliases for "arrow")
- "csv" and "tsv"
- "text" (generic text-delimited files - use the `delimiter` argument to specify which to use)

Hive style partitioning is also supported, with partitions detected automatically from the file paths:

```
year=2021/month=12/data.parquet
year=2022/month=01/data.parquet
year=2022/month=02/data.parquet
...
```

## Write Multi-file Datasets

Save partitioned data to disk based on columns in the data:

```
write_dataset(df, "data_partitioned",
              partitioning = c("year", "month"))
```

Default partitioning is based on any existing groups in the `tibble` or `data.frame`.

## Write Individual Files

Write an R object `df` to disk:

```
write_parquet(df, "file.parquet")
write_feather(df, "file.feather")
write_csv_arrow(df, "file.csv")
```

To save a compressed file to disk, you specify the compression algorithm with the `compression` argument:

```
write_parquet(df, "file.parquet",
              compression = "gzip")
```

Some file formats write compressed data by default. For more information on the supported compression algorithms see:

```
?write_parquet()
?write_feather()
?write_dataset()
```

## Manipulate Larger-than-Memory Datasets

**arrow** lets you work efficiently with large, multi-file datasets by providing a **dplyr**—and many other R functions—interface to query, manipulate and summarise large datasets *before* pulling data into your R session with **dplyr**'s `collect()`:

```
arrow_table(starwars) %>%
  filter(homeworld == "Tatooine") %>%
  rename(height_cm = height,
        mass_kg = mass) %>%
  mutate(height_in = height_cm / 2.54,
        mass_lbs = mass_kg * 2.2046) %>%
  arrange(desc(birth_year)) %>%
  select(name, height_in, mass_lbs) %>%
  collect()
```

In addition to most single-table **dplyr** verbs, many other function mappings are implemented in **arrow**, including base R, **lubridate**, and **stringr** functions.

# Arrow for R :: CHEAT SHEET



## Manipulate Larger-than-Memory Datasets (cont)

**arrow** supports joins for joining multiple tables:

```
robot <- data.frame(  
  species = c("Human", "Droid", "Ewok"),  
  bot = c(FALSE, TRUE, FALSE)  
)  
  
arrow_table(starwars) %>%  
  select(name, species) %>%  
  left_join(robot) %>%  
  collect()
```

If you use **arrow** with partitioned data, **arrow** will only read from the relevant partitions:

```
year=2021/month=12/data.parquet  
year=2022/month=01/data.parquet  
year=2022/month=02/data.parquet  
...  
  
open_dataset("folder",  
  partitioning = c("year", "month")) %>%  
  filter(year == 2022) %>%  
  group_by(month) %>%  
  summarise(total = sum(amount)) %>%  
  collect()
```

For queries on Table objects, if **arrow** detects an unimplemented function, it will automatically call `collect()` and pull the data into R with a warning message:

```
## Warning: Expression  
unimplemented_function() not supported in  
Arrow;  
## pulling data into R
```

For queries on Dataset objects (which can be larger than memory), if **arrow** detects an unimplemented function, it will raise an error. You will need to explicitly tell **arrow** to `collect()` before the unimplemented function.

## Zero-Copy R and Python Data Sharing

**arrow** provides **reticulate** methods for passing data between R and Python using the Python **pyarrow** library. Install, load, and import **pyarrow** in a virtual environment:

```
library(reticulate)  
  
virtualenv_create("arrow-env")  
install_pyarrow("arrow-env")  
use_virtualenv("arrow-env")  
pa <- import("pyarrow")
```

Use **pyarrow** to create an Arrow array object in an R session:

```
a <- pa$array(c(1, 2, 3))
```

Call a **pyarrow** function from your R session:

```
table1 <- arrow_table(starwars[1:5,])  
table2 <- arrow_table(starwars[11:15,])  
  
pa$concat_tables(tables = list(table1,  
  table2)) %>%  
  collect()
```

## Transport Data with Flight

Connect to Flight RPC server to send and receive data with **arrow**:

```
library(reticulate)  
library(arrow)  
install_pyarrow()  
  
demo <- load_flight_server("flight_server")  
server <- demo$FlightServer(port = 8089)  
server$serve()  
  
client <- flight_connect(port = 8089)  
  
flight_put(client, df, path = "data/df")  
df <- flight_get(client, "data/df")
```

## Cloud Storage Support (S3)

**Arrow** supports reading files and multi-file datasets from cloud storage without having to download them first—`open_dataset()`, `write_dataset()` and **arrow**'s `read_*` and `write_*` functions all accept an S3 Uniform Resource Identifier (URI) as the source or destination file.

Read a file, a multi-file dataset, or partitioned multi-file dataset:

```
df <- read_parquet(  
  "s3://ursa-labs-taxi-data/2019/  
  06/data.parquet")  
  
df <-  
  open_dataset("s3://ursa-labs-taxi-data")  
  
df <- open_dataset(  
  "s3://ursa-labs-taxi-data",  
  partitioning = c("year", "month"))
```

Create an **arrow** `FileSystem` object and pass that to **arrow**'s read and write functions:

```
bucket <- s3_bucket("ursa-labs-taxi-data")  
df <- read_parquet(bucket$path(  
  "2019/06/data.parquet"))
```

Copy data from cloud storage to your computer:

```
copy_files("s3://ursa-labs-taxi-data",  
  "~/nyc-taxi")
```

Detailed instructions for working with S3 cloud storage are available here: [arrow.apache.org/docs/r/articles/fs](https://arrow.apache.org/docs/r/articles/fs)

## Additional Resources

**Arrow** R Cookbook: [arrow.apache.org/cookbook/r/](https://arrow.apache.org/cookbook/r/)

Reference guide to **arrow** in R: [arrow.apache.org/docs/r/](https://arrow.apache.org/docs/r/)

**Arrow** communities: [arrow.apache.org/community/](https://arrow.apache.org/community/)

# Base R Cheat Sheet

## Getting Help

### Accessing the help files

?mean

Get help of a particular function.

help.search('weighted mean')

Search the help files for a word or phrase.

help(package = 'dplyr')

Find help for a package.

### More about an object

str(iris)

Get a summary of an object's structure.

class(iris)

Find the class an object belongs to.

## Using Packages

install.packages('dplyr')

Download and install a package from CRAN.

library(dplyr)

Load the package into the session, making all its functions available to use.

dplyr::select

Use a particular function from a package.

data(iris)

Load a built-in dataset into the environment.

## Working Directory

getwd()

Find the current working directory (where inputs are found and outputs are sent).

setwd('C://file/path')

Change the current working directory.

**Use projects in RStudio to set the working directory to the folder you are working in.**

## Vectors

### Creating Vectors

c(2, 4, 6)	2 4 6	Join elements into a vector
2:6	2 3 4 5 6	An integer sequence
seq(2, 3, by=0.5)	2.0 2.5 3.0	A complex sequence
rep(1:2, times=3)	1 2 1 2 1 2	Repeat a vector
rep(1:2, each=3)	1 1 1 2 2 2	Repeat elements of a vector

### Vector Functions

sort(x)

Return x sorted.

rev(x)

Return x reversed.

table(x)

See counts of values.

unique(x)

See unique values.

### Selecting Vector Elements

#### By Position

x[4]

The fourth element.

x[-4]

All but the fourth.

x[2:4]

Elements two to four.

x[!(2:4)]

All elements except two to four.

x[c(1, 5)]

Elements one and five.

#### By Value

x[x == 10]

Elements which are equal to 10.

x[x < 0]

All elements less than zero.

x[x %in% c(1, 2, 5)]

Elements in the set 1, 2, 5.

### Named Vectors

x['apple']

Element with name 'apple'.

## Programming

### For Loop

```
for (variable in sequence){  
  Do something  
}
```

### Example

```
for (i in 1:4){  
  j <- i + 10  
  print(j)  
}
```

### While Loop

```
while (condition){  
  Do something  
}
```

### Example

```
while (i < 5){  
  print(i)  
  i <- i + 1  
}
```

### Functions

```
function_name <- function(var){  
  Do something  
  return(new_variable)  
}
```

### Example

```
square <- function(x){  
  squared <- x*x  
  return(squared)  
}
```

## Reading and Writing Data

Also see the **readr** package.

Input	Output	Description
df <- read.table('file.txt')	write.table(df, 'file.txt')	Read and write a delimited text file.
df <- read.csv('file.csv')	write.csv(df, 'file.csv')	Read and write a comma separated value file. This is a special case of read.table/write.table.
load('file.RData')	save(df, file = 'file.Rdata')	Read and write an R data file, a file type special for R.

Conditions	a == b	Are equal	a > b	Greater than	a >= b	Greater than or equal to	is.na(a)	Is missing
	a != b	Not equal	a < b	Less than	a <= b	Less than or equal to	is.null(a)	Is null

## Types

Converting between common data types in R. Can always go from a higher value in the table to a lower value.

as.logical	TRUE, FALSE, TRUE	Boolean values (TRUE or FALSE).
as.numeric	1, 0, 1	Integers or floating point numbers.
as.character	'1', '0', '1'	Character strings. Generally preferred to factors.
as.factor	'1', '0', '1', levels: '1', '0'	Character strings with preset levels. Needed for some statistical models.

## Maths Functions

log(x)	Natural log.	sum(x)	Sum.
exp(x)	Exponential.	mean(x)	Mean.
max(x)	Largest element.	median(x)	Median.
min(x)	Smallest element.	quantile(x)	Percentage quantiles.
round(x, n)	Round to n decimal places.	rank(x)	Rank of elements.
signif(x, n)	Round to n significant figures.	var(x)	The variance.
cor(x, y)	Correlation.	sd(x)	The standard deviation.

## Variable Assignment

```
> a <- 'apple'  
> a  
[1] 'apple'
```

## The Environment

ls()	List all variables in the environment.
rm(x)	Remove x from the environment.
rm(list = ls())	Remove all variables from the environment.

You can use the environment panel in RStudio to browse variables in your environment.

## Matrices

`m <- matrix(x, nrow = 3, ncol = 3)`  
Create a matrix from x.

	<code>m[2, ]</code> - Select a row	<code>t(m)</code> Transpose
	<code>m[, 1]</code> - Select a column	<code>m %*% n</code> Matrix Multiplication
	<code>m[2, 3]</code> - Select an element	<code>solve(m, n)</code> Find x in: $m \cdot x = n$

## Lists

`l <- list(x = 1:5, y = c('a', 'b'))`  
A list is a collection of elements which can be of different types.

<code>l[[2]]</code>	<code>l[1]</code>	<code>l\$x</code>	<code>l['y']</code>
Second element of l.	New list with only the first element.	Element named x.	New list with only element named y.

Also see the `dplyr` package.

## Data Frames

`df <- data.frame(x = 1:3, y = c('a', 'b', 'c'))`  
A special case of a list where all elements are the same length.

x	y
1	a
2	b
3	c

## Matrix subsetting

<code>df[, 2]</code>	
<code>df[2, ]</code>	
<code>df[2, 2]</code>	

List subsetting	
<code>df\$x</code>	<code>df[[2]]</code>

<i>Understanding a data frame</i>
<code>View(df)</code>
See the full data frame.

<i>head(df)</i>
See the first 6 rows.

`nrow(df)`  
Number of rows.

`cbind` - Bind columns.

`ncol(df)`  
Number of columns.

`rbind` - Bind rows.

`dim(df)`  
Number of columns and rows.

## Strings

<code>paste(x, y, sep = ' ')</code>	Join multiple vectors together.
<code>paste(x, collapse = ' ')</code>	Join elements of a vector together.
<code>grep(pattern, x)</code>	Find regular expression matches in x.
<code>gsub(pattern, replace, x)</code>	Replace matches in x with a string.
<code>toupper(x)</code>	Convert to uppercase.
<code>tolower(x)</code>	Convert to lowercase.
<code>nchar(x)</code>	Number of characters in a string.

## Factors

<code>factor(x)</code>	
Turn a vector into a factor. Can set the levels of the factor and the order.	Turn a numeric vector into a factor by 'cutting' into sections.

## Statistics

<code>lm(y ~ x, data=df)</code>	Linear model.
<code>glm(y ~ x, data=df)</code>	Generalised linear model.
<code>summary</code>	Get more detailed information out a model.
<code>pairwise.t.test</code>	Perform a t-test for paired data.

## Distributions

	Random Variates	Density Function	Cumulative Distribution	Quantile
Normal	<code>rnorm</code>	<code>dnorm</code>	<code>pnorm</code>	<code>qnorm</code>
Poisson	<code>rpois</code>	<code>dpois</code>	<code>ppois</code>	<code>qpois</code>
Binomial	<code>rbinom</code>	<code>dbinom</code>	<code>pbinom</code>	<code>qbinom</code>
Uniform	<code>runif</code>	<code>dunif</code>	<code>unif</code>	<code>qunif</code>

## Plotting

<code>plot(x)</code>	Values of x in order.
<code>plot(x, y)</code>	Values of x against y.
<code>hist(x)</code>	Histogram of x.

## Dates

See the `lubridate` package.

# Bayesplot :: CHEAT SHEET



```
library("bayesplot")
library("rstanarm")
options(mc.cores = parallel::detectCores())
library("ggplot2")
library("dplyr")
```

## Model Parameters

To showcase bayesplot, we'll fit linear regression using `rstanarm::stan_glm` and use this model throughout.

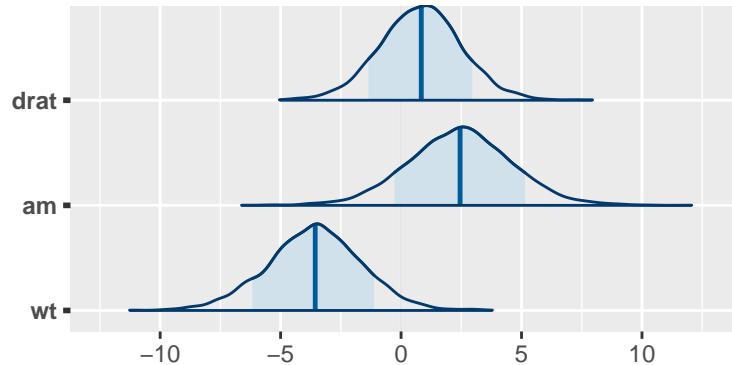
```
model <- stan_glm(mpg ~ ., data=mtcars, chains=4)
posterior <- as.matrix(model)
```

Chances are good you're most interested in the posterior distributions for select parameters.

```
plot_title <- ggtitle("Posterior distributions",
                      "medians and 80% intervals")
mcmc_areas(posterior,
            pars = c("drat", "am", "wt"),
            prob = 0.8) + plot_title
```

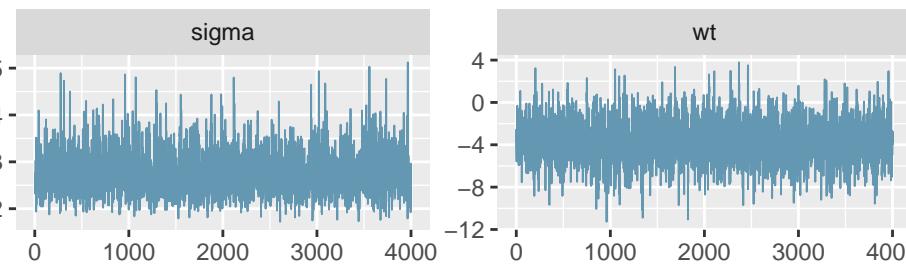
## Posterior distributions

medians and 80% intervals



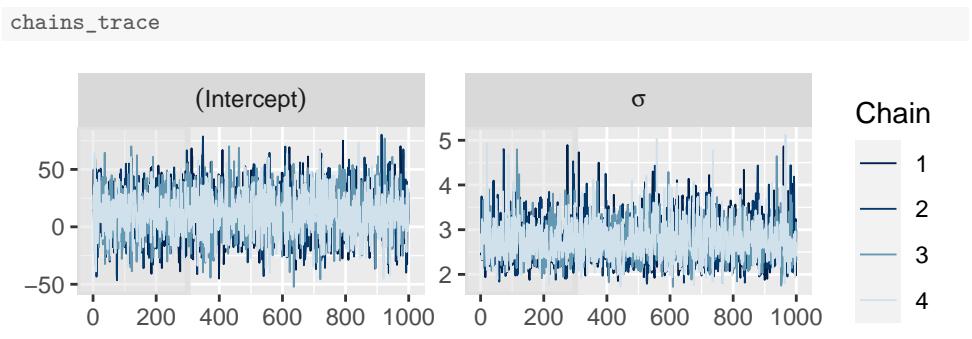
Diagnosing convergence with traceplots is simple.

```
mcmc_trace(posterior, pars=c("sigma", "wt"))
```



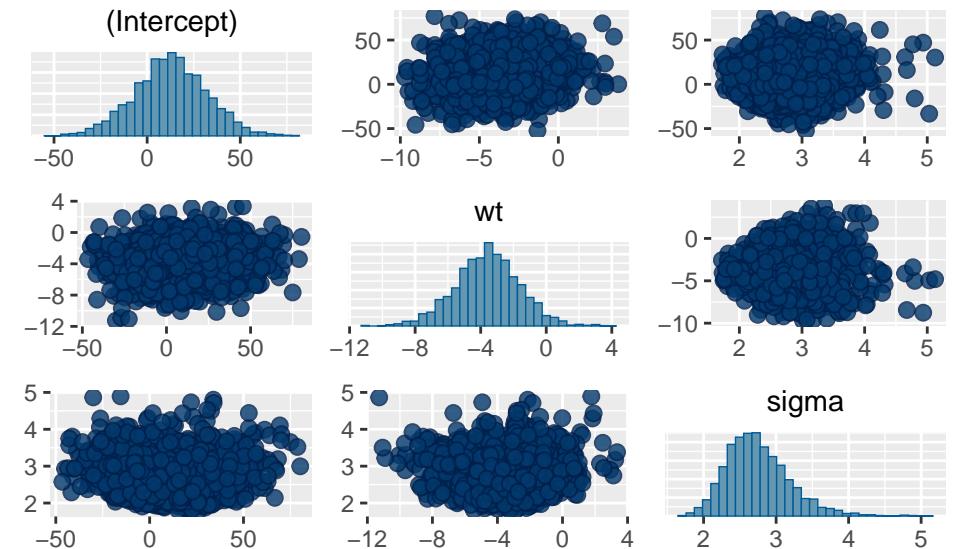
Using `as.array`, you can extract each of the four chain's posterior draws, different from above. This allows you to see each chain's traceplot for selected parameters.

```
posterior_chains <- as.array(model)
fargs <- list(ncol = 2, labeller = label_parsed)
pars <- c("Intercept", "sigma")
chains_trace <- mcmc_trace(posterior_chains, pars = pars,
                           n_warmup = 300, facet_args = fargs)
```



The pairs plot is helpful in determining if you have any highly correlated parameters.

```
posterior_chains %>%
  mcmc_pairs(pars = c("(Intercept)", "wt", "sigma"))
```

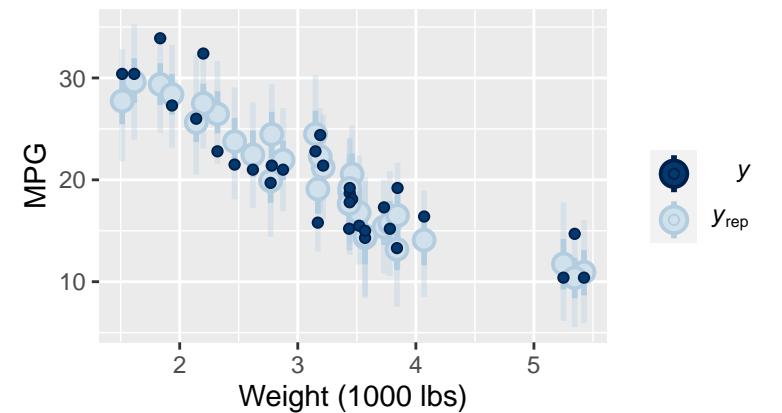


## Posterior Predictive Checks

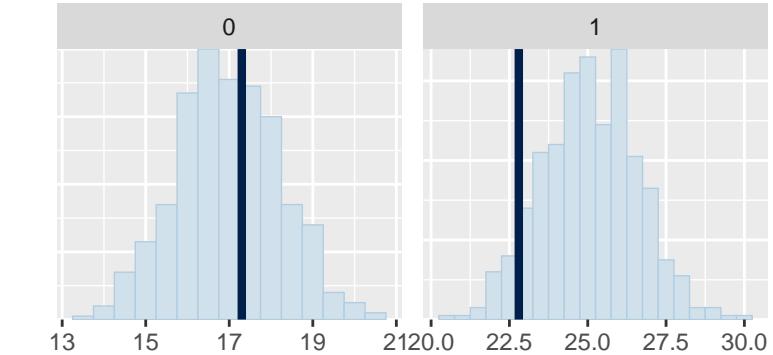
Check how well the model covers your data with draws from the posterior predictive density.

```
ppd <- posterior_predict(model, draws=500)
ppd %>%
  ppc_intervals(y = mtcars$mpg, yrep = ., x = mtcars$wt, prob = 0.5) +
  labs(x = "Weight (1000 lbs)", y = "MPG",
       title = "50% posterior predictive intervals of MPG by weight")
```

## 50% posterior predictive intervals of MPG by



```
ppd %>%
  ppc_stat_grouped(y = mtcars$mpg, group = mtcars$am,
                     stat = "median", binwidth=0.5)
```

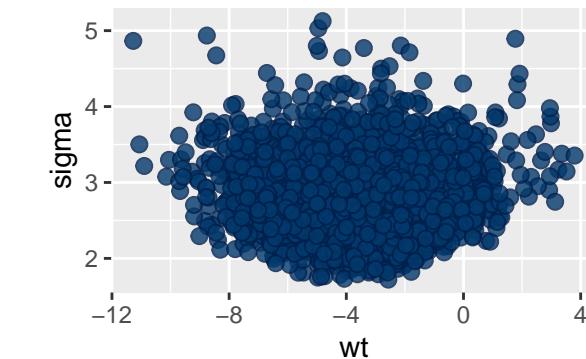


$T = \text{median}$   
 $T(y_{\text{rep}})$   
 $T(y)$

## Diagnostics

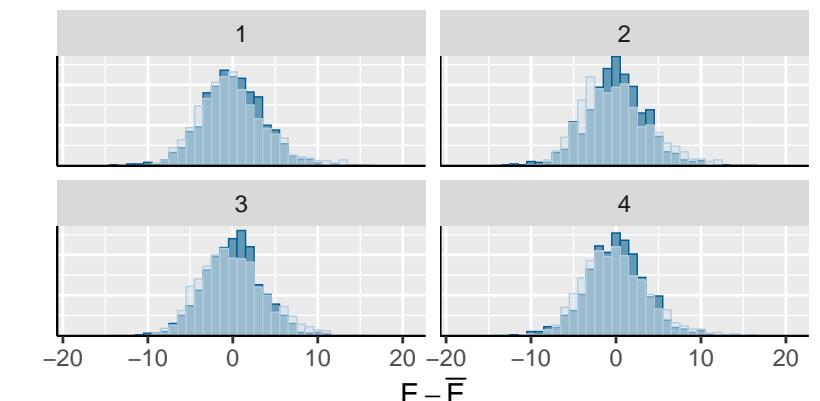
Bayesplot makes it easy to check diagnostics specific to the NUTS sampling method that `rstanarm` uses by default.

```
mcmc_scatter(posterior, pars = c("wt", "sigma"),
              np = nuts_params(model$stanfit))
```



```
np <- nuts_params(model$stanfit)
mcmc_nuts_energy(np, binwidth=1) +
  ggtitle("NUTS Energy Diagnostic")
```

## NUTS Energy Diagnostic



$\pi_E$   
 $\pi_{\Delta E}$

# BCEA :: CHEAT SHEET



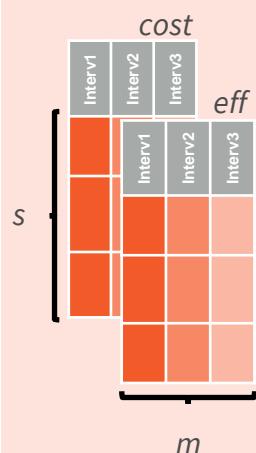
## Introduction

### Bayesian Cost-Effectiveness Analysis in R

Given a random sample of suitable variables of costs (*cost*) and clinical benefits (*eff*) for two or more interventions produces a health economic evaluation. Inputs may be the results of a Bayesian model (possibly based on MCMC) in the form of simulations from the posterior distributions. For *s* sample points compares one of the *m* interventions (*reference*) to the others (*.comparison*).

```
bcea(eff, cost, ref, .comparison, interventions)
```

#### INPUT ARRAY PAIR



#### CONSTITUENT FUNCTIONS

- `compute_U()` : Expected utility for each WTP & intervention
- `compute_Ustar()` : Maximum 'known-distribution' utility for each WTP
- `compute_vi()` : Value of information for each WTP
- `compute_ol()` : Opportunity Loss for each WTP
- `compute_ICER()` : Incremental cost-effectiveness ratio
- `compute_IB()` : Incremental benefit for each WTP
- `compute_CEAC()` : Cost-effectiveness acceptability for each WTP
- `compute_EIB()` : Expected incremental benefit for each WTP
- `compute_kstar()` : WTP break-even value

`bcea()` calculates numerous cost-effectiveness analysis statistics. These can be called directly, using the constituent functions, but would require some pre-processing which is already handled by `bcea()`.

## Value assignment

There are 3 equivalent ways to assign values to analysis parameters.

1. *In Constructor*: When first creating a `bcea` object.

```
he <- bcea(eff, cost, ref, .comparison, ...)
```

2. *Using Setters*: Change directly using replacement functions.

```
setComparison(he) <- comparison
setKmax(he) <- Kmax
setReference(he) <- ref
```

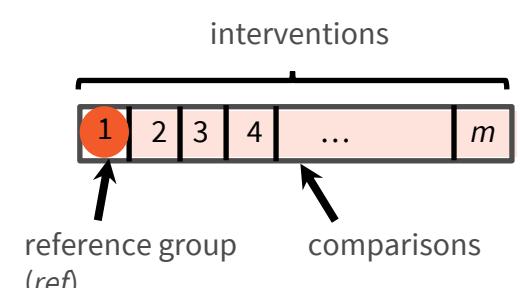
3. *In plotting call*: At the point of making a plot.

```
eib.plot(he, comparison, ...)
ceac.plot(he, comparison, ...)
ceplane.plot(he, comparison, ...)
```

#### SELECTING ANALYSIS INTERVENTIONS

##### Default

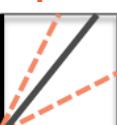
The first columns in (*eff*, *cost*) are the default reference intervention. All other interventions are the comparison interventions unless defined otherwise. E.g. for *m* interventions



## Plot

Standard cost-effectiveness analysis output plots. Base R, ggplot2 and plotly versions of plots are available and can be called directly but require extra default parameters.

#### Expected incremental benefit

  
`eib.plot(he, comparison = NULL, pos = c(1, 0), size = NULL, plot.cri = NULL, graph = c("base", "ggplot2", "plotly"), ...)`  
calls: • `eib_plot_base()`  
• `eib_plot_ggplot()`  
• `eib_plot_plotly()`

#### Expected value of information

  
`evi.plot(he, graph = c("base", "ggplot2", "plotly"), ... )`

#### Cost-effectiveness planes with contours

  
`contour[2](he, comparison = 1, scale = 0.5, nlevels = 4, levels = NULL, pos = c(1, 0), xlim = NULL, ylim = NULL, graph = c("base", "ggplot2"), ...)`

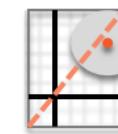
#### Compare optimal scenario to mixed case

  
`plot.mixedAn(x, y.limits = NULL, pos = c(0, 1), graph = c("base", "ggplot2"), ...)`

#### Cost-effectiveness acceptability curve

  
`ceac_plot(he, comparison = NULL, pos = c(1, 0), graph = c("base", "ggplot2", "plotly"), ...)`  
calls: • `ceac_plot_base()`  
• `ceac_plot_ggplot()`  
• `ceac_plot_plotly()`

#### Cost-effectiveness plane

  
`ceplane_plot(he, comparison = NULL, pos = c(1, 0), graph = c("base", "ggplot2", "plotly"), ...)`  
calls: • `ceplane_plot_base()`  
• `ceplane_plot_ggplot()`  
• `ceplane_plot_plotly()`

#### Grid of CE plane, EIB, EVI & CEAC

  
`plot.bcea(x, comparison = NULL, pos = c(1, 0), graph = c("base", "ggplot2", "plotly"), ...)`

#### Expected value of perfect partial information

  
`plot.evppi(x, pos = c(0, 0.8), graph = c("base", "ggplot2"), col = NULL, ...)`

## Summarise data

Summary statistics and formatted tables can be used to interrogate a `bcea()` object.

`summary.bcea(he, ...)`

Prints a table with summary results of the health economic evaluation

`summary.mixedAn(he, ...)`

Prints summary table for results of mixed analysis

`sim.table(he, ...)`

Summary table of simulations from the cost-effectiveness analysis

`make.report(he, ...)`

Constructs the automated report from the output of the BCEA

# caret Package

## Cheat Sheet

### Specifying the Model

Possible syntaxes for specifying the variables in the model:

```
train(y ~ x1 + x2, data = dat, ...)
train(x = predictor_df, y = outcome_vector, ...)
train(recipe_object, data = dat, ...)
```

- `rfe`, `sbf`, `gafs`, and `safs` only have the `x/y` interface.
- The `train` formula method will **always** create dummy variables.
- The `x/y` interface to `train` will not create dummy variables (but the underlying model function might).

**Remember** to:

- Have column names in your data.
- Use factors for a classification outcome (not 0/1 or integers).
- Have valid R names for class levels (not "0"/"1")
- Set the random number seed prior to calling `train` repeatedly to get the same resamples across calls.
- Use the `train` option `na.action = na.pass` if you will be imputing missing data. Also, use this option when predicting new data containing missing values.

To pass options to the underlying model function, you can pass them to `train` via the ellipses:

```
train(y ~ ., data = dat, method = "rf",
      # options to `randomForest`:
      importance = TRUE)
```

### Parallel Processing

The `foreach` package is used to run models in parallel. The `train` code does not change but a "`do`" package must be called first.

```
# on Mac OS or Linux      # on Windows
library(doMC)              library(doParallel)
registerDoMC(cores=4)       cl <- makeCluster(2)
                           registerDoParallel(cl)
```

The function `parallel::detectCores` can help too.

### Preprocessing

Transformations, filters, and other operations can be applied to the *predictors* with the `preProc` option.

```
train(..., preProc = c("method1", "method2"), ...)
```

Methods include:

- `"center"`, `"scale"`, and `"range"` to normalize predictors.
- `"BoxCox"`, `"YeoJohnson"`, or `"expoTrans"` to transform predictors.
- `"knnImpute"`, `"bagImpute"`, or `"medianImpute"` to impute.
- `"corr"`, `"nzv"`, `"zv"`, and `"conditionalX"` to filter.
- `"pca"`, `"ica"`, or `"spatialSign"` to transform groups.

`train` determines the order of operations; the order that the methods are declared does not matter.

The `recipes` package has a more extensive list of preprocessing operations.

### Adding Options

Many `train` options can be specified using the `trainControl` function:

```
train(y ~ ., data = dat, method = "cubist",
      trControl = trainControl(<options>))
```

### Resampling Options

`trainControl` is used to choose a resampling method:

```
trainControl(method = <method>, <options>)
```

Methods and options are:

- `"cv"` for K-fold cross-validation (`number` sets the # folds).
- `"repeatedcv"` for repeated cross-validation (`repeats` for # repeats).
- `"boot"` for bootstrap (`number` sets the iterations).
- `"LGOCV"` for leave-group-out (`number` and `p` are options).
- `"L0O"` for leave-one-out cross-validation.
- `"oob"` for out-of-bag resampling (only for some models).
- `"timeslice"` for time-series data (options are `initialWindow`, `horizon`, `fixedWindow`, and `skip`).

### Performance Metrics

To choose how to summarize a model, the `trainControl` function is used again.

```
trainControl(summaryFunction = <R function>,
             classProbs = <logical>)
```

Custom R functions can be used but `caret` includes several: `defaultSummary` (for accuracy, RMSE, etc), `twoClassSummary` (for ROC curves), and `prSummary` (for information retrieval). For the last two functions, the option `classProbs` must be set to `TRUE`.

### Grid Search

To let `train` determine the values of the tuning parameter(s), the `tuneLength` option controls how many values `per tuning` parameter to evaluate.

Alternatively, specific values of the tuning parameters can be declared using the `tuneGrid` argument:

```
grid <- expand.grid(alpha = c(0.1, 0.5, 0.9),
                      lambda = c(0.001, 0.01))
```

```
train(x = x, y = y, method = "glmnet",
      preProc = c("center", "scale"),
      tuneGrid = grid)
```

### Random Search

For tuning, `train` can also generate random tuning parameter combinations over a wide range. `tuneLength` controls the total number of combinations to evaluate. To use random search:

```
trainControl(search = "random")
```

### Subsampling

With a large class imbalance, `train` can subsample the data to balance the classes them prior to model fitting.

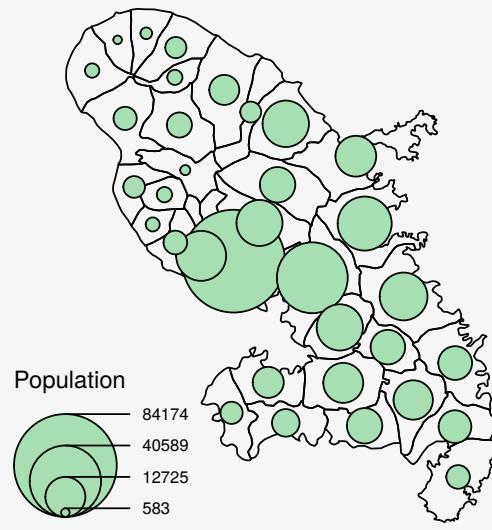
```
trainControl(sampling = "down")
```

Other values are `"up"`, `"smote"`, or `"rose"`. The latter two may require additional package installs.

# Thematic maps with cartography :: CHEAT SHEET

Use cartography with spatial objects from sf or sp packages to create thematic maps.

```
library(cartography)
library(sf)
mtq <- st_read("martinique.shp")
plot(st_geometry(mtq))
propSymbolsLayer(x = mtq, var = "P13_POP",
  legend.title.txt = "Population",
  col = "#a7dfb4")
```



## Classification

Available methods are: quantile, equal, q6, fisher-jenks, mean-sd, sd, geometric progression...

```
bks1 <- getBreaks(v = var, nclass = 6,
  method = "quantile")
bks2 <- getBreaks(v = var, nclass = 6,
  method = "fisher-jenks")
pal <- carto.pal("green.pal", 3, "wine.pal", 3)
hist(var, breaks = bks1, col = pal)
```



```
hist(var, breaks = bks2, col = pal)
```



## Symbology

In most functions the x argument should be an sf object. sp objects are handled through spdf and df arguments.



Choropleth  
choroLayer(x = mtq, var = "myvar",  
method = "quantile", nclass = 8)



Typology  
typoLayer(x = mtq, var = "myvar")



Proportional Symbols  
propSymbolsLayer(x = mtq, var = "myvar",  
inches = 0.1, symbols = "circle")



Colorized Proportional Symbols (relative data)  
propSymbolsChoroLayer(x = mtq, var = "myvar",  
var2 = "myvar2")



Colorized Proportional Symbols (qualitative data)  
propSymbolsTypoLayer(x = mtq, var = "myvar",  
var2 = "myvar2")



Double Proportional Symbols  
propTrianglesLayer(x = mtq, var1 = "myvar",  
var2 = "myvar2")



OpenStreetMap Basemap (see rosm package)  
tiles <- getTiles(x = mtq, type = "osm")  
tilesLayer(tiles)



Isopleth (see SpatialPosition package)  
smoothLayer(x = mtq, var = "myvar",  
typefc = "exponential", span = 500,  
beta = 2)



Discontinuities  
discLayer(x = mtq.borders, df = mtq\_df,  
var = "myvar", threshold = 0.5)



Flows  
propLinkLayer(x = mtq\_link, df = mtq\_df,  
var = "fij")



Dot Density  
dotDensityLayer(x = mtq, var = "myvar")



Labels  
labelLayer(x = mtq, txt = "myvar",  
halo = TRUE, overlap = FALSE)

## Transformations

Polygons to Grid

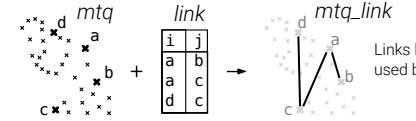
```
mtq_grid <- getGridLayer(x = mtq, cellsize = 3.6e+07,
  type = "hexagonal", var = "myvar")
```



Grids layers can be used by  
choroLayer() or propSymbolsLayer().

Points to Links

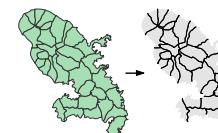
```
mtq_link <- getLinkLayer(x = mtq, df = link)
```



Links layers can be  
used by \*LinkLayer().

Polygons to Borders

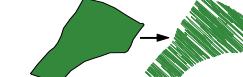
```
mtq_border <- getBorders(x = mtq)
```



Borders layers can be used by  
discLayer() function

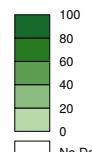
Polygons to Pencil Lines

```
mtq_pen <- getPencilLayer(x = mtq)
```



## Legends

legendChoro()



```
legendChoro(pos = "topleft",
  title.txt = "legendChoro()",  
breaks = c(0,20,40,60,80,100),  
col = carto.pal("green.pal", 6),  
nodata = TRUE, nodata.txt = "No Data")
```

legendTypo()



```
legendTypo(title.txt = "legendTypo()",  
col = c("peru", "skyblue", "gray77"),  
categ = c("type 1", "type 2", "type 3"),  
nodata = FALSE)
```

legendCirclesSymbols()

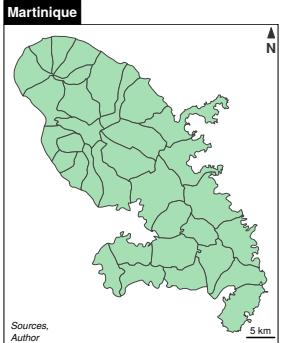


```
legendCirclesSymbols(var = c(10,100),  
title.txt = "legendCirclesSymbols()",  
col = "#a7dfb4ff", inches = 0.3)
```

See also legendSquaresSymbols(), legendBarsSymbols(),  
legendGradLines(), legendPropLines() and legendPropTriangles().

## Map Layout

North Arrow:  
north(pos = "topright")



Scale Bar:  
barscale(size = 5)

Full Layout:

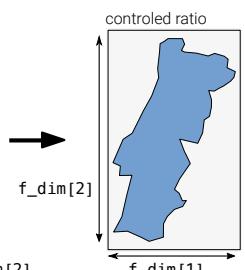
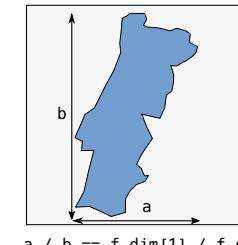
```
layoutLayer(
  title = "Martinique",
  tabtitle = TRUE,
  frame = TRUE,
  author = "Author",
  sources = "Sources",
  north = TRUE,
  scale = 5)
```

Figure Dimensions

Get figure dimensions based on the dimension ratio of a spatial object,  
figure margins and output resolution.

```
f_dim <- getFigDim(x = sf_obj, width = 500,
  mar = c(0,0,0,0))
png("fig.png", width = 500, height = f_dim[2])
par(mar = c(0,0,0,0))
plot(sf_obj, col = "#729fcf")
dev.off()
```

default



## Color Palettes

```
carto.pal(pal1 = "blue.pal", n1 = 5,
  pal2 = sand.pal, n2 = 3)
```



display.carto.all(n = 8)



# Advanced and Fast Data Transformation with `collapse` :: CHEAT SHEET



## Introduction

`collapse` is a C/C++ based package supporting advanced (grouped, weighted, time series, panel data and recursive) statistical operations in R, with very efficient low-level vectorizations across both groups and columns.

It also offers a flexible, class-agnostic, approach to data transformation in R: handling matrix and data frame based objects in a uniform, attribute preserving, way, and ensuring seamless compatibility with base R, `dplyr` / (grouped) `tibble`, `data.table`, `xts/zoo`, `sf`, and `plm` classes for panel data.

`collapse` provides full control to the user for statistical programming - with several ways to reach the same outcome and rich optimization possibilities. It is globally configurable using `setCollapse()` which includes algorithm defaults, multithreading, and the exported namespace (see below).

Calling `help("collapse-documentation")` brings up a detailed documentation, which is also available [online](#). See also the [fastverse](#) package/project for a recommended set of complimentary packages and easy package management.

## Row/Column Arithmetic (by Reference)

Column-wise sweeping out of vectors/matrices/DFs/lists

```
%cr%, %c+%, %c-%, %c*%, %c/% e.g. Z = X %c% rowSums(X)
```

Row-wise sweeping vectors from vectors/matrices/DFs/lists

```
%rr%, %r+%, %r-%, %r*%, %r/% e.g. Z = X %r% colSums(X)
```

Standard (column-wise) math by reference (returns invisibly)

```
%+=%, %-=%, %*=%, %=% e.g. X %-=% rowSums(X)
```

Same thing, also supports row-wise operations by reference

```
setup(X, "/", rowSums(X))
setup(X, "/", colSums(X), rowwise = TRUE)
```

## Transform Data by (Grouped) Replacing or Sweeping out Statistics (by Reference)

A generalisation of rowwise operations, that also supports sweeping by groups e.g. aggregate statistics

```
TRA(x, STATS, FUN = "-", g = NULL, set = FALSE)
setTRA(x, STATS, FUN = "-", g = NULL)
```

x vector, matrix, or (grouped) data frame / list

STATS statistics matching (columns of) x (i.e. aggregated vector, matrix or data frame / list)

FUN integer/string indicating transformation to perform:

Int.	String	Description
0	"replace.NA"	replace missing values in x
1	"replace.fill"	replace data and missing values in x
2	"replace"	replace data but preserve missing values in x
3	"+"	subtract: x - STATS(g)
4	"+"	x - STATS(g) + mean(STATS, w = GRP%)
5	"%"	divide: x / STATS(g)
6	"%"	compute percentages: x * 100/STATS(g)
7	"%"	add: x + STATS(g)
8	"%"	multiply: x * STATS(g)
9	"%"	modulus: x %% STATS(g)
10	"%"	subtract modulus: x - x %% STATS(g)

g [optional] (list of) vectors / factors or GRP() object

set TRUE transforms x by reference. `setTRA` is equivalent to `invisible(TRA(..., set = TRUE))`

## Fast Statistical Functions

Fast functions to perform column-wise grouped and weighted computations on matrix-like objects

```
fmean, fmedian, fmode, fsum, fprod, fsd, fvar,
fmin, fmax, fnth, ffirst, flast, fnobs, fndistinct
```

## Syntax

```
FUN(x, g = NULL, [w = NULL], TRA = NULL,
[na.rm = TRUE], use.g.names = TRUE,
[drop = TRUE], [nthreads = 1L])
```

x vector, matrix, or (grouped) data frame / list

g [optional] (list of) vectors / factors or GRP() object

w [optional] vector of (frequency) weights

TRA [optional] operation to transform data with computed statistics (see FUN argument to TRA() and Examples)

drop drop matrix / data frame dimensions. default TRUE

## Examples

```
fmean(AirPassengers) # Vector
## [1] 280.2986
fmean(AirPassengers, w = cycle(AirPassengers)) # Weighted mean
## [1] 284.3397
fmean(EuStockMarkets) # Matrix
##   DAX    SMI    CAC    FTSE
## 2530.657 3376.224 2227.828 3565.643
fmean(airquality) # Data Frame (use drop = FALSE to keep frame)
##   Ozone Solar.R Wind Temp Month Day
## 42.129310 185.931507 9.957516 77.882253 6.993464 15.803922
fmean(iris[1:4], g = iris$Species) # Grouped
##             Sepal.Length Sepal.Width Petal.Length Petal.Width
## setosa      5.006     3.428     1.462     0.246
## versicolor  5.936     2.770     4.260     1.326
## virginica   6.588     2.974     5.552     2.026
X = iris[1:4]; g = iris$Species; w <- abs(rnorm(nrow(X)))
fmean(X, g, w) # Grouped and weighted (random weights)
##             Sepal.Length Sepal.Width Petal.Length Petal.Width
## setosa      4.974946  3.376431  1.467120  0.2613994
## versicolor  6.020234  2.827597  4.310551  1.3258578
## virginica   6.542129  2.947751  5.463577  2.0390838
## Transformations: here centering data on the weighted group median
TRA(X, fmedian(X, g, w, "-", g)) |> head(2)
##   Sepal.Length Sepal.Width Petal.Length Petal.Width
## 1          0.1        0.1       -0.1        0
## 2         -0.1       -0.4       -0.1        0
fmedian(X, g, w, TRA = "-") |> head(2) # Same thing: more compact
##   Sepal.Length Sepal.Width Petal.Length Petal.Width
## 1          0.1        0.1       -0.1        0
## 2         -0.1       -0.4       -0.1        0
fmedian(X, g, w, "-", set = TRUE) # Modify in-place (same as setTRA())
```

## Other Statistical Functions

Fast (weighted) sample quantiles, range, and distances

```
fquantile(x, probs, w, o, na.rm = TRUE, type = 7)
```

```
frange(x, na.rm = TRUE)
```

```
fdist(x, v, method = "euclidean", nthreads = 1)
```

## Basic Computing with R Functions

Apply R functions to rows or columns (by groups)

```
dapply(x, FUN, ..., MARGIN = 2) - column/row apply
```

```
BY(x, g, FUN, ...) - split-apply-combine computing
```

## Grouping and Ordering

Optimized functions for grouping, ordering, unique values, matching, splitting, and dealing with factors

GRP() - create a grouping object (class 'GRP'): pass to g arg.

```
g <- GRP(iris, ~ Species) # or GRP(iris$Species) or GRP(iris[["Species"]])
fndistinct(iris[1:4], g) # Computation without grouping overhead
```

	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width
## setosa	15	16	9	6
## versicolor	21	14	19	9
## virginica	21	13	20	12

fgroup\_by() - attach 'GRP' object to data: a class-agnostic grouped frame supporting fast computations

```
mtcars |> fgroup_by(cyl, vs, am) |> ss(1:2)
##          mpg cyl disp hp drat wt qsec vs am gear carb
## Mazda RX4 21   6 160 110 3.9 2.620 16.46 0 1 4 4
## Mazda RX4 Wag 21   6 160 110 3.9 2.875 17.02 0 1 4 4
##
## Grouped by: cyl, vs, am [7 | 5 | 3 (3.8) 1-12]
# Group Stats: [N. groups | mean (sd) min-max of group sizes]
# Fast Functions also have a grouped_df method: here wt-weighted medians
mtcars |> fgroup_by(cyl, vs, am) |> fmedian(wt) |> head(2)
##   cyl vs am sum.wt mpg disp hp drat qsec gear carb
## 1   4   0   1 21.0  6.0 120.0 3.9 4.05 18.00 3.0 1.5 4
## 2   4   1   0 28.8  6.9 162.0 4.9 3.21 18.40 3.0 1.0 4
## 2   4   1   0 8.05 22.8 140.8 95.37 3.0 20.01 4 4
```

GRPN(), fcountr[v]() , fgroup\_vars(), fungroup() - get group count, grouping columns, and ungroup data

qF(), qG() - quick as.factor, and vector grouping object of class 'qG': a factor-light without levels attribute

group() - (multivariate) group id ('qG') in appearance order

groupid() - run-length-type group id ('qG')

seqid() - group-id from integer-sequences ('qG')

radixorder[v]() - (multivariate) radix-based ordering

finteraction() - fast factor interactions (or return 'qG')

fdroplevels() - fast removal of unused factor levels

f[n]unique(), fduplicated() - fast unique values / rows

fmatch(), %![!][i]in% - fast matching of values / rows

gsplit() - fast splitting vector based on 'GRP' objects

greorder() - efficiently reorder y = unlist(gsplit(x, g)) such that identical(greorder(y, g), x)

`collapse` optimizes grouping using both factors / 'qG' objects and 'GRP' objects. 'GRP' objects contain most information and are thus most efficient for complex computations.

```
X <- iris[1:4]; v <- as.character(iris$Species)
f <- qF(v, na, exclude = FALSE) # adds 'na.included' class: no NA checks
## > group(v) # 'qG' object: first appearance order, with 'na.included'
microbenchmark(fmode(X, v), fmode(X, f), fmode(X, gv), fmode(X, g))
```

	Unit: microseconds
## fmode(X, v)	expr min lq mean median uq max neval
## fmode(X, f)	11.890 12.9150 15.17697 13.3455 13.7350 162.073 100
## fmode(X, gv)	9.225 9.8195 11.33035 10.0860 10.4550 92.947 100
## fmode(X, g)	8.569 9.3480 10.73667 9.6555 10.1065 73.021 100

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## fmode(X, v) 11.890 12.9150 15.17697 13.3455 13.7350 162.073 100

## fmode(X, f) 9.225 9.8195 11.33035

## Advanced Transformations

Common transformations (in econometrics)

Scaling, Centering and Averaging

```
fscale(x, g = NULL, w = NULL, na.rm = TRUE,
       mean = 0, sd = 1, ...)
fwithin(x, g = NULL, w = NULL, na.rm = TRUE,
        mean = 0, theta = 1, ...)
fbetween(x, g = NULL, w = NULL, na.rm = TRUE,
          fill = FALSE, ...)
```

Higher-Dimensional Centering/Avg. and Linear Prediction  
`fhdwithin(x, f1, w = NULL, na.rm = TRUE,`  
`fill = FALSE, lm.method = "qr", ...)`  
`fdbetween() - same arguments as fhdwithin()`

Statistical Operators (function shorthands with extra features)  
`STD(), W(), B(), HDW(), HDB()`

## Examples

```
# Grouped scaling
iris > fgroup_by(Species) |> fscale() |> head(2)
## Species Sepal.Length Sepal.Width Petal.Length Petal.Width
## 1 setosa   0.2666745  0.1899414 -0.3570112 -0.4364923
## 2 setosa   -0.3007180 -1.1290958 -0.3570112 -0.4364923
STD(iris, ~Species, stub = FALSE) # Same thing + faster
# Grouped and weighted scaling. Operators support formulas and keep ids
STD(mtcars, mpg + carb ~ cyl, w = "wt") |> head(2)
##      cyl   wt STD.mpg STD.carb
## Mazda RX4  6 2.620 0.9691687 0.386125
## Mazda RX4 Wag 6 2.875 0.9691687 0.386125
# Much shorter than fsubset(mpg > fmean(mpg, cyl, TRA = "replace"))
mtcars > fsubset(mpg > B(mpg, cyl)) |> head(2)
##      mpg cyl disp hp drat wt qsec vs am gear carb
## Mazda RX4  21  6 160 110  3.9 2.620 16.46  0  1  4   4
## Mazda RX4 Wag 21  6 160 110  3.9 2.875 17.02  0  1  4   4
# Regression with cyl fixed effects - a la Mundlak (1978)
lm(mpg ~ carb + B(carb, cyl), data = mtcars) |> coef()
## (Intercept) carb B(carb, cyl)
## 34.829652 -0.465511 -4.775032
# Fast grouped (vs) bivariate regression slopes: mpg ~ carb
mtcars > fgroup_by(carb) |> fmutate(dm_carb = W(carb)) |>
  summarise(beta = fsum(mpg, dm_carb) %~% fsum(dm_carb)^2)
##    vs      beta
## 1  0 -0.5557241
## 2  1 -2.0706468
# Residuals from regressing on 'Petal' vars and 'Species' FE
fhdwithin(iris[1:2], iris[3:5]) |> head(2)
## Sepal.Length Sepal.Width
## 1  0.14989286  0.1102684
## 2 -0.05010714 -0.3897316
# Detrending with country-level cubic polynomials
HDW(wlddev, PCGDP + LIFEEX + POP ~ iso3c * poly(year, 3)) |> head(2)
## HDW.PCGDP HDW.LIFEEX HDW.POP
## 1 9.963947 0.023669531 314373.06
## 2 14.044984 0.006742957 -32402.75
# Note: HD centering/prediction and polynomials requires package 'fjtest'
```

## Linear Models

Fast (barebones) linear model fitting with 6 different solvers  
`flm(y, X, w = NULL, add.icpt = FALSE, method = "lm")`

Fast  $R^2$ -based F-test of exclusion restrictions for lm's (with FE)  
`fFtest(y, exc, X = NULL, w = NULL, full.df = TRUE)`

Both functions also have formula interfaces:

```
flm(cbind(mpg, disp) ~ hp + carb, weights = wt, mtcars)
##      mpg      disp
## (Intercept) 28.48041839 42.155002
## hp         -0.06834996  2.101036
## carb        0.33207257 -38.183910
# Test the exclusion of cyl-dummies and hp.
fFtest(mpg ~ qB(cyl) + hp | carb + qF(am), weights = wt, mtcars)
##          R-Sq. DF1 DF2 F-Stat. P-Value
## Full Model 0.812 5 26 22.479 0.000
## Restricted Model 0.674 2 29 30.041 0.000
## Exclusion Rest. 0.138 3 26 6.351 0.002
```

## Time Series and Panel Series

Fast and flexible indexed series and data frames: a modern upgrade of plm's 'pseries' and 'pdata.frame'

Turn DF into an 'indexed\_frame' using id and/or time vars  
`data_ix = findex_by(data, id1, ..., time)`

`data_ix$indexed_series - columns are 'indexed_series'`  
`index_df = findex(data_ix) - retrieve 'index_df': DF of ids`  
`index_df = with(data_ix, findex(indexed_series)) - can fetch 'index_df' from 'indexed_series' in any caller environment`

`data = unindex(data_ix) - unindex (also 'indexed.series')`  
`reindex(data, index = index_df) - reindex / new pointers`  
`'indexed_series' can be 1-or-2D atomic objects. Vectors / time series / matrices can also be indexed directly using:`  
`reindex(vec/mat, index = vec/index_df)`  
`is_irregular() - irregularity in any index[ed] obj. or time vec`

### Example: Indexing Panel Data

```
wldi <- wlddev |> findex_by(iso3c, year) # Balanced: 216 countries
fsubset(wldi, 1:12, iso3c, year, PCGDP:POP)
## iso3c year PCGDP LIFEEX GINI ODA POP
## 1 AFG 1960 NA 32.446 116769997 8996973
## 2 AFG 1961 NA 32.962 232080002 9169410
##
## Indexed by: iso3c [1] | year [2 (61)]
## Index stats: [N..ids] / [N.. periods (tot.N.. periods: (max-min)/GCD)]
LIFEEXI = wldi$LIFEEX # Indexed series
str(LIFEEXI, strict.width = "cut")
## 'indexed_series' num [1:13176] 32.4 33 33.5 34 34.5 ...
## - attr(*, "index_df")=Classes 'index_df', 'pindex' and 'data.frame'...
## .. $ iso3c: Factor w/ 216 levels "ABW","AFG","AGO",... 2 2 2 2 2 2 ...
## .. $ year: Ord.factor w/ 61 levels "1960"<"1961"<...: 1 2 3 4 5 6 ...
LIFEEXI[1:7] # Subsetting indexed series
## #> [1] 32.446 32.962 33.471 33.971 34.463 34.948 35.430
##
## Indexed by: iso3c [1] | year [7 (61)]
c(is_irregular(LIFEEXI), is_irregular(LIFEEXI[-5])) # Is irregular?
## #> [1] FALSE TRUE
```

Note: 'indexed\_series' and frames are supported via existing 'pseries'/pdata.frame' methods for time series/panel functions.

Fast functions to perform time-based computations on (irregular) time series and (unbalanced) panel data

Lags/Leads, Differences, Growth Rates and Cumulative Sums  
`flag(x, n = 1, g = NULL, t = NULL, fill = NA, ...)`  
`fdiff(x, n = 1, diff = 1, g = NULL, t = NULL,`  
 `fill = NA, log = FALSE, rho = 1, ...)`  
`fgrowth(x, n = 1, diff = 1, g = NULL, t = NULL, fill = NA, logdiff = FALSE, scale = 100, power = 1, ...)`  
`fcumsum(x, g = NULL, o = NULL, na.rm = TRUE, fill = FALSE, check.o = TRUE, ...)`

Statistical Operators: L(), F(), D(), Dlog(), G()

### Example: Computing Growth Rates

```
# Ad-hoc use: note that G() supports formulas which fgrowth() doesn't
fgrowth(AirPassengers) |> head()
## #> [1] NA 5.351743 11.864407 -2.272727 -6.201550 11.570248
G(wlddev, c(1, 10), by = PCGDP ~ iso3c, t = ~ year) |> ss(11:12)
## iso3c year G1.PCGDP L10G1.PCGDP
## 1 AFG 1970 NA NA
## 2 AFG 1971 NA NA
wlddev |> fgroup_by(iso3c) |> fselect(iso3c, year, PCGDP, LIFEEX) |>
  fmutate(PCGDP_growth = fgrowth(PCGDP, t = year)) |> head(2)
## iso3c year PCGDP LIFEEX PCGDP_growth
## 1 AFG 1960 NA 32.446 NA
## 2 AFG 1961 NA 32.962 NA
settransform(wlddev, PCGDP_growth = G(PCGDP, g = iso3c, t = year))
# Note: can omit t -> requires consecutive observations and groups
# Usage with indexed series / frames:
```

## Recode and Replace Values

recode\_num(), recode\_char() - recode numeric / character values (+ regex recoding) in matrix-like objects

replace\_[NA|Inf|outliers]() - replace special values  
`pad() - add (missing) observations / rows i.e. expand objects`

## (Memory) Efficient Programming

Functions for (memory) efficient R programming

```
any|all[v|NA], which[v|NA], %[!]=%, copyv, setv, alloc
missing_cases, na.[insert|rm|omit], vlengths, vtypes,
vcgdc, fnlevels, fn[row|col], fdim, seq_[row|col], vec
fsubset(wlddev, year %=% 2010) # 2x faster fsubset(wlddev, year == 2010)
attach(mtcars) # Efficient sub-assignment by reference, various options...
setv(am, 0, vs); setv(am, 1:10, vs); setv(am, 1:10, vs[10:20])
```

## Small (Helper) Functions

Functions for (meta-)programming and attributes

```
.c, massign, %=%, vlabels[<-], setLabels, vclasses,
namlab, [add|rm]_stub, all_identical, all_obj_equal,
all_funcs, set[Dim]Row[Col]names, unattrAttrib, setAttrib,
copyAttrib, copyMostAttrib, is_categorical, is_date
.c(vari, var2, var3) # Non-standard concatenation
## [1] "var1" "var2" "var3"
.c(values, vectors) %=% eigen(cov(mtcars)) # Multiple Assignment
# Variable labels: vlabels[<-], (set|relabel) etc. namlab() shows summary
namlab(wlddev[c(2, 9)], N = TRUE, Mdist = TRUE, class = TRUE)
## Variable Class N Mdist Label
## 1 iso3c factor 13176 216 Country Code
## 2 PCGDP numeric 9470 9470 GDP per capita (constant 2010 US$)
```

## API Extensions and Global Options

Shorthands for frequently used functions

```
fselect -> slt, fsubset -> sbt, fmutate -> mtt,
[f/set]transform[v] -> [set]tfm[v], fsmarimise ->
smr, across -> acr, fgroup_by -> gby, finteraction
-> itn, findex_by -> iby, findex -> ix, frename ->
rnm, get_vars -> gv, num_vars -> nv, add_vars -> av
```

Namespace masking and other global options

Use `set_colollapse(mask = c(...))` with a vector of functions starting with f, to export versions without f, masking base R and/or dplyr. A few keywords exist to mask multiple functions, see `help("collapse-options")`. There are also many other global defaults and optimizations that can be controlled with `set.colollapse(...)`. Retrieve options using `get.colollapse()`.

```
# Masking all (f-)functions and changing some defaults (optimizing)
library(collapse)
set_colollapse(mask = "all", na.rm = FALSE, sort = FALSE, nthreads = 4)
# The following is now 100% collapse code and executed without regard for
# missing values, using unsorted grouping and 4 threads (where applicable)
wlddev |>
  subset(year >= 1990 & is.finite(GINI)) |>
  group_by(year) |>
  summarise(n = n(), across(PCGDP:GINI, mean, w = POP))

with(mtcars, table(cyl, vs, am))
sum(mtcars)
diff(EuStockMarkets)
droplevels(wlddev)
mean(nv(iris), g = iris$Species)
scale(nv(GGD10S), g = GGD10S$Variable)
unique(GGD10S, cols = c("Variable", "Country"))
range(wlddev$Date)

wlddev |>
  index_by(iso3c, year) |>
  mutate(PCGDP_lag = lag(PCGDP),
         PCGDP_diff = PCGDP - PCGDP_lag,
         PCGDP_growth = growth(PCGDP)) |> unindex()
```

# Data Transformation with data.table :: CHEAT SHEET



## Basics

data.table is an extremely fast and memory efficient package for transforming data in R. It works by converting R's native data frame objects into data.tables with new and enhanced functionality. The basics of working with data.tables are:

**dt[i, j, by]**

Take data.table **dt**,  
subset rows using **i**  
and manipulate columns with **j**,  
grouped according to **by**.

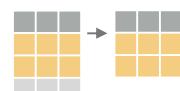
data.tables are also data frames – functions that work with data frames therefore also work with data.tables.

## Create a data.table

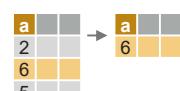
**data.table(a = c(1, 2), b = c("a", "b"))** – create a data.table from scratch. Analogous to `data.frame()`.

**setDT(df)\* or as.data.table(df)** – convert a data frame or a list to a data.table.

## Subset rows using i



**dt[1:2, ]** – subset rows based on row numbers.



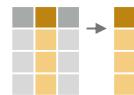
**dt[a > 5, ]** – subset rows based on values in one or more columns.

### LOGICAL OPERATORS TO USE IN i

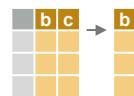
<	<=	is.na()	%in%		%like%
>	>=	!is.na()	!	&	%between%

## Manipulate columns with j

### EXTRACT



**dt[, c(2)]** – extract columns by number. Prefix column numbers with “-” to drop.



**dt[, -(b, c)]** – extract columns by name.

### SUMMARIZE



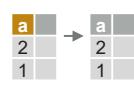
**dt[, .(x = sum(a))]** – create a data.table with new columns based on the summarized values of rows.

Summary functions like `mean()`, `median()`, `min()`, `max()`, etc. can be used to summarize rows.

### COMPUTE COLUMNS\*



**dt[, c := 1 + 2]** – compute a column based on an expression.



**dt[a == 1, c := 1 + 2]** – compute a column based on an expression but only for a subset of rows.



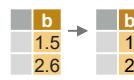
**dt[, `:=` (c = 1, d = 2)]** – compute multiple columns based on separate expressions.

### DELETE COLUMN



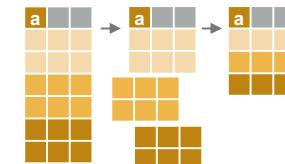
**dt[, c := NULL]** – delete a column.

### CONVERT COLUMN TYPE



**dt[, b := as.integer(b)]** – convert the type of a column using `as.integer()`, `as.numeric()`, `as.character()`, `as.Date()`, etc..

## Group according to by



**dt[, j, by = .(a)]** – group rows by values in specified columns.

**dt[, j, keyby = .(a)]** – group and simultaneously sort rows by values in specified columns.

### COMMON GROUPED OPERATIONS

**dt[, .(c = sum(b)), by = a]** – summarize rows within groups.

**dt[, c := sum(b), by = a]** – create a new column and compute rows within groups.

**dt[, .SD[1], by = a]** – extract first row of groups.

**dt[, .SD[N], by = a]** – extract last row of groups.

## Chaining

**dt[...][...]** – perform a sequence of data.table operations by chaining multiple “[]”.

## Functions for data.tables

### REORDER



**setorder(dt, a, -b)** – reorder a data.table according to specified columns. Prefix column names with “-” for descending order.

### \* SET FUNCTIONS AND :=

data.table's functions prefixed with “set” and the operator “:=” work without “-<” to alter data without making copies in memory. E.g., the more efficient “`setDT(df)`” is analogous to “`df <- as.data.table(df)`”.



## Apply function to cols.

### APPLY A FUNCTION TO MULTIPLE COLUMNS

<b>a b</b>	$\rightarrow$	<b>a b</b>
1 2		1 2
2 2		2 2

`dt[, lapply(.SD, mean), .SDcols = c("a", "b")]` – apply a function – e.g. `mean()`, `as.character()`, `which.max()` – to columns specified in `.SDcols` with `lapply()` and the `.SD` symbol. Also works with groups.

<b>a</b>	$\rightarrow$	<b>a m</b>
1		1 2
2		2 2
3		3 2

`cols <- c("a")`  
`dt[, paste0(cols, "_m") := lapply(.SD, mean), .SDcols = cols]` – apply a function to specified columns and assign the result with suffixed variable names to the original data.

## Reshape a data.table

### RESHAPE TO WIDE FORMAT

<b>id y a b</b>	$\rightarrow$	<b>id a x a z b x b z</b>	<b>dcast(dt,</b>
A x 1 3		A 1 2 3 4	id ~ y,
A x 1 3		B 1 2 3 4	value.var = c("a", "b"))
B z 2 4			
B z 2 4			

Reshape a data.table from long to wide format.

`dt` A data.table.  
`id ~ y` Formula with a LHS: ID columns containing IDs for multiple entries. And a RHS: columns with values to spread in column headers.  
`value.var` Columns containing values to fill into cells.

### RESHAPE TO LONG FORMAT

<b>id a x a z b x b z</b>	$\rightarrow$	<b>id y a b</b>	<b>melt(dt,</b>
A 1 2 3 4		A x 1 3	measure.vars = measure (
B 1 2 3 4		B x 1 3	value.name, y, sep = "_"))
		A z 2 4	
		B z 2 4	

Reshape a data.table from wide to long format.

`dt` A data.table.  
`measure.vars` Columns containing values to fill into cells, often using `measure()` or patterns `(.)`.  
`id.vars` Character vector of ID column names (optional).  
`variable.name,`  
`value.name` Names for output columns (optional).

`measure(out_name1, out_name2, sep = "_", pattern = "[ab]_(.*_)")`  
`sep`(separator) or `pattern` (regular expression) are used to specify columns to melt, and to parse input column names.  
`out_name1, out_name2`: names for output columns (creates single value column), or `value.name` (creates a value column for each unique part of the melted column name).

## Sequential rows

### ROW IDS

<b>a b</b>	$\rightarrow$	<b>a b c</b>
1 a		1 a 1
2 a		2 a 2
3 b		3 b 1

`dt[, c := 1:N, by = b]` – within groups, compute a column with sequential row IDs.

### LAG & LEAD

<b>a b</b>	$\rightarrow$	<b>a b c</b>
1 a		1 a NA
2 a		2 a 1
3 b		3 b NA
4 b		4 b 3
5 b		5 b 4

`dt[, c := shift(a, 1), by = b]` – within groups, duplicate a column with rows lagged by specified amount.

`dt[, c := shift(a, 1, type = "lead"), by = b]` – within groups, duplicate a column with rows leading by specified amount.

## read & write files

### IMPORT

`fread("file.csv")` – read data from a flat file such as `.csv` or `.tsv` into R.

`fread("file.csv", select = c("a", "b"))` – read specified columns from a flat file into R.

### EXPORT

`fwrite(dt, "file.csv")` – write data to a flat file from R.



# Data import with the tidyverse :: CHEATSHEET

## Read Tabular Data with readr

```
read_*(file, col_names = TRUE, col_types = NULL, col_select = NULL, id = NULL, locale, n_max = Inf,
skip = 0, na = c("", "NA"), guess_max = min(1000, n_max), show_col_types = TRUE) See ?read_delim
```

A B C 1 2 3 4 5 NA	→	A   B   C 1 2 3 4 NA
		<b>read_delim("file.txt", delim = " ")</b> Read files with any delimiter. If no delimiter is specified, it will automatically guess. To make file.txt, run: write_file("A B C\n1 2 3\n4 5 NA", file = "file.txt")

A,B,C 1,2,3 4,5,NA	→	A   B   C 1 2 3 4 NA
		<b>read_csv("file.csv")</b> Read a comma delimited file with period decimal marks. write_file("A,B,C\n1,2,3\n4,5,NA", file = "file.csv")

A;B;C 1,5;2;3 4,5;5;NA	→	A   B   C 1.5 2 3 4.5 NA
		<b>read_csv2("file2.csv")</b> Read semicolon delimited files with comma decimal marks. write_file("A;B;C\n1,5;2;3\n4,5;5;NA", file = "file2.csv")

A B C 1 2 3 4 5 NA	→	A   B   C 1 2 3 4 NA
		<b>read_tsv("file.tsv")</b> Read a tab delimited file. Also <b>read_table()</b> . <b>read_fwf("file.tsv", fwf_widths(c(2, 2, NA)))</b> Read a fixed width file. write_file("A\tB\tC\n1\t2\t3\n4\t5\tNA", file = "file.tsv")

### USEFUL READ ARGUMENTS

**No header**  
read\_csv("file.csv", col\_names = FALSE)

**Provide header**  
read\_csv("file.csv", col\_names = c("x", "y", "z"))

**Read multiple files into a single table**  
read\_csv(c("f1.csv", "f2.csv", "f3.csv"), id = "origin\_file")

**Skip lines**  
read\_csv("file.csv", skip = 1)

**Read a subset of lines**  
read\_csv("file.csv", n\_max = 1)

**Read values as missing**  
read\_csv("file.csv", na = c("1"))

**Specify decimal marks**  
read\_delim("file2.csv", locale = locale(decimal\_mark = ","))

One of the first steps of a project is to import outside data into R. Data is often stored in tabular formats, like csv files or spreadsheets.



The front page of this sheet shows how to import and save text files into R using **readr**.



The back page shows how to import spreadsheet data from Excel files using **readxl** or Google Sheets using **googlesheets4**.

### OTHER TYPES OF DATA

Try one of the following packages to import other types of files:

- **haven** - SPSS, Stata, and SAS files
- **DBI** - databases
- **jsonlite** - json
- **xml2** - XML
- **httr** - Web APIs
- **rvest** - HTML (Web Scraping)
- **readr::read\_lines()** - text data

## Column Specification with readr

Column specifications define what data type each column of a file will be imported as. By default **readr** will generate a column spec when a file is read and output a summary.

**spec(x)** Extract the full column specification for the given imported data frame.

```
spec(x)
# cols(
#   age = col_integer(),
#   edu = col_character(),
#   earn = col_double()
# )
```

age is an integer  
edu is a character  
earn is a double (numeric)

### COLUMN TYPES

Each column type has a function and corresponding string abbreviation.

- **col\_logical()** - "l"
- **col\_integer()** - "i"
- **col\_double()** - "d"
- **col\_number()** - "n"
- **col\_character()** - "c"
- **col\_factor(levels, ordered = FALSE)** - "f"
- **col\_datetime(format = "")** - "T"
- **col\_date(format = "")** - "D"
- **col\_time(format = "")** - "t"
- **col\_skip() - "-"**, "\_"
- **col\_guess() - "?"**

### USEFUL COLUMN ARGUMENTS

#### Hide col spec message

read\_\*(file, show\_col\_types = FALSE)

#### Select columns to import

Use names, position, or selection helpers.  
read\_\*(file, col\_select = c(age, earn))

#### Guess column types

To guess a column type, **read\_\***() looks at the first 1000 rows of data. Increase with **guess\_max**.  
read\_\*(file, guess\_max = Inf)

### DEFINE COLUMN SPECIFICATION

#### Set a default type

```
read_csv(
  file,
  col_type = list(.default = col_double())
)
```

#### Use column type or string abbreviation

```
read_csv(
  file,
  col_type = list(x = col_double(), y = "l", z = "_")
)
```

#### Use a single string of abbreviations

```
# col types: skip, guess, integer, logical, character
read_csv(
  file,
  col_type = "_?ilc"
)
```

## Save Data with readr

```
write_*(x, file, na = "NA", append, col_names, quote, escape, eol, num_threads, progress)
```

A   B   C 1 2 3 4 5 NA	→	A,B,C 1,2,3 4,5,NA
		<b>write_delim(x, file, delim = " ")</b> Write files with any delimiter. <b>write_csv(x, file)</b> Write a comma delimited file. <b>write_csv2(x, file)</b> Write a semicolon delimited file. <b>write_tsv(x, file)</b> Write a tab delimited file.



# Import Spreadsheets with readxl

## READ EXCEL FILES

	A	B	C	D	E
1	x1	x2	x3	x4	x5
2	x		z	8	
3	y	7		9	10

```
read_excel(path, sheet = NULL, range = NULL)  
Read a .xls or .xlsx file based on the file extension.  
See front page for more read arguments. Also  
read_xls() and read_xlsx().  
read_excel("excel_file.xlsx")
```

## READ SHEETS

A	B	C	D	E
s1	s2	s3		

s1	s2	s3
----	----	----

A	B	C	D	E
A	B	C	D	E
A	B	C	D	E

- To **read multiple sheets**:
1. Get a vector of sheet names from the file path.
  2. Set the vector names to be the sheet names.
  3. Use purrr::map() and purrr::list\_rbind() to read multiple files into one data frame.

```
path <- "your_file_path.xlsx"  
path >  
  excel_sheets() |>  
  set_names() |>  
  map(read_excel, path = path) |>  
  list_rbind()
```

## OTHER USEFUL EXCEL PACKAGES

For functions to write data to Excel files, see:

- **openxlsx**
- **writexl**

For working with non-tabular Excel data, see:

- **tidyxl**



# with googlesheets4

## READ SHEETS

	A	B	C	D	E
1	x1	x2	x3	x4	x5
2	x		z	8	
3	y	7		9	10

```
read_sheet(ss, sheet = NULL, range = NULL)  
Read a sheet from a URL, a Sheet ID, or a dribble  
from the googledrive package. See front page for  
more read arguments. Same as range_read().
```

## SHEETS METADATA

**URLs** are in the form:  
<https://docs.google.com/spreadsheets/d/>  
**SPREADSHEET\_ID**/edit#gid=**SHEET\_ID**

**gs4\_get(ss)** Get spreadsheet meta data.

**gs4\_find(...)** Get data on all spreadsheet files.

**sheet\_properties(ss)** Get a tibble of properties  
for each worksheet. Also **sheet\_names()**.

## WRITE SHEETS

1	x	4
2	y	5
3	z	6

→

A	B	C
1	1	x
2	2	y
3	3	z

s1

**write\_sheet(data, ss = NULL, sheet = NULL)**  
Write a data frame into a new or existing Sheet.

**gs4\_create(name, ..., sheets = NULL)** Create a new Sheet with a vector of names, a data frame, or a (named) list of data frames.

**sheet\_append(ss, data, sheet = 1)** Add rows to the end of a worksheet.

1	A	B	C
2			

→

A	B	C
1	x1	x2
2	y	5
3	z	6
4		

s1

**gs4\_update(ss, sheet, data)**  
Update a sheet with a data frame.

**sheet\_replace(ss, sheet, data)**  
Replace a sheet with a data frame.

**sheet\_delete(ss, sheet)**  
Delete a sheet.

I	n	c	D	L
TRUE	2	hello	1947-01-08	hello
FALSE	3.45	world	1956-10-21	1

- skip - "\_" or "-"
- guess - "?"
- logical - "l"
- integer - "i"
- double - "d"
- numeric - "n"
- date - "D"
- datetime - "T"
- character - "c"
- list-column - "L"
- cell - "C" Returns list of raw cell data.

Use list for columns that include multiple data types. See **tidyr** and **purrr** for list-column data.

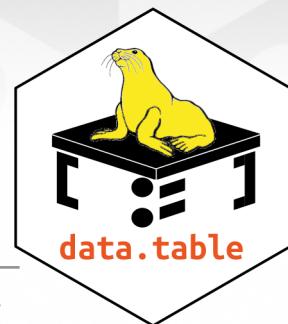
## FILE LEVEL OPERATIONS

**googlesheets4** also offers ways to modify other aspects of Sheets (e.g. freeze rows, set column width, manage (work)sheets). Go to [googlesheets4.tidyverse.org](https://googlesheets4.tidyverse.org) to read more.

For whole-file operations (e.g. renaming, sharing, placing within a folder), see the tidyverse package **googledrive** at [googledrive.tidyverse.org](https://googledrive.tidyverse.org).



# Data Transformation with data.table :: CHEAT SHEET



## Basics

data.table is an extremely fast and memory efficient package for transforming data in R. It works by converting R's native data frame objects into data.tables with new and enhanced functionality. The basics of working with data.tables are:

**dt[i, j, by]**

Take data.table **dt**,  
subset rows using **i**  
and manipulate columns with **j**,  
grouped according to **by**.

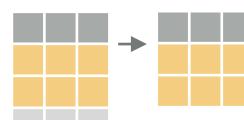
data.tables are also data frames – functions that work with data frames therefore also work with data.tables.

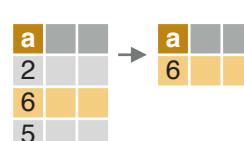
## Create a data.table

**data.table(a = c(1, 2), b = c("a", "b"))** – create a data.table from scratch. Analogous to `data.frame()`.

**setDT(df)\* or as.data.table(df)** – convert a data frame or a list to a data.table.

## Subset rows using **i**

 **dt[1:2, ]** – subset rows based on row numbers.

 **dt[a > 5, ]** – subset rows based on values in one or more columns.

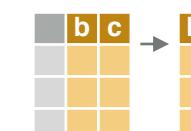
### LOGICAL OPERATORS TO USE IN **i**

<	<=	is.na()	%in%		%like%
>	>=	!is.na()	!	&	%between%

## Manipulate columns with **j**

### EXTRACT

 **dt[, c(2)]** – extract columns by number. Prefix column numbers with “-” to drop.

 **dt[, .(b, c)]** – extract columns by name.

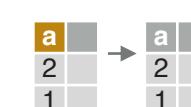
### SUMMARIZE

 **dt[, .(x = sum(a))]** – create a data.table with new columns based on the summarized values of rows.

Summary functions like `mean()`, `median()`, `min()`, `max()`, etc. can be used to summarize rows.

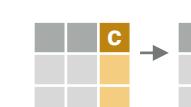
### COMPUTE COLUMNS\*

 **dt[, c := 1 + 2]** – compute a column based on an expression.

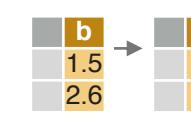
 **dt[a == 1, c := 1 + 2]** – compute a column based on an expression but only for a subset of rows.

 **dt[, `:=` (c = 1, d = 2)]** – compute multiple columns based on separate expressions.

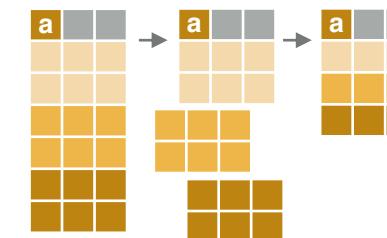
### DELETE COLUMN

 **dt[, c := NULL]** – delete a column.

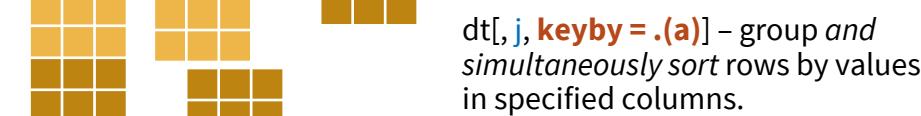
### CONVERT COLUMN TYPE

 **dt[, b := as.integer(b)]** – convert the type of a column using `as.integer()`, `as.numeric()`, `as.character()`, `as.Date()`, etc..

## Group according to **by**



**dt[, j, by = .(a)]** – group rows by values in specified columns.



**dt[, j, keyby = .(a)]** – group and simultaneously sort rows by values in specified columns.

### COMMON GROUPED OPERATIONS

**dt[, .(c = sum(b)), by = a]** – summarize rows within groups.

**dt[, c := sum(b), by = a]** – create a new column and compute rows within groups.

**dt[, .SD[1], by = a]** – extract first row of groups.

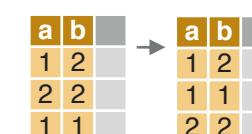
**dt[, .SD[N], by = a]** – extract last row of groups.

## Chaining

**dt[...][...]** – perform a sequence of data.table operations by chaining multiple “[]”.

## Functions for data.tables

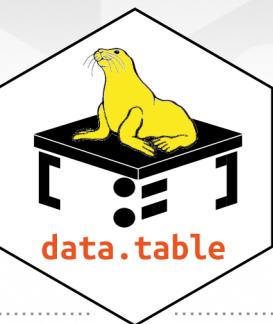
### REORDER



**setorder(dt, a, -b)** – reorder a data.table according to specified columns. Prefix column names with “-” for descending order.

### \* SET FUNCTIONS AND :=

data.table's functions prefixed with “set” and the operator “:=” work without “<-” to alter data without making copies in memory. E.g., the more efficient “`setDT(df)`” is analogous to “`df <- as.data.table(df)`”.



## UNIQUE ROWS

a	b
1	2
2	2
1	2

`unique(dt, by = c("a", "b"))` – extract unique rows based on columns specified in “by”. Leave out “by” to use all columns.

`uniqueN(dt, by = c("a", "b"))` – count the number of unique rows based on columns specified in “by”.

## RENAME COLUMNS

a	b
x	y

`setnames(dt, c("a", "b"), c("x", "y"))` – rename columns.

## SET KEYS

`setkey(dt, a, b)` – set keys to enable fast repeated lookup in specified columns using “`dt[.(value), ]`” or for merging without specifying merging columns using “`dt_a[dt_b]`”.

## Combine data.tables

### JOIN

a	b
1	c
2	a
3	b

x	y
3	b
2	c
1	a

a	b	x
3	b	3
2	c	2
1	a	1

`dt_a[dt_b, on = .(b = y)]` – join data.tables on rows with equal values.

a	b	c
1	c	7
2	a	5
3	b	6

x	y	z
3	b	4
2	c	5
1	a	8

a	b	c	x
3	b	4	3
1	c	5	2
2	a	8	NA

`dt_a[dt_b, on = .(b = y, c > z)]` – join data.tables on rows with equal and unequal values.

### ROLLING JOIN

a	id	date
1	A	01-01-2010
2	A	01-01-2012
3	A	01-01-2014
1	B	01-01-2010
2	B	01-01-2012

b	id	date
1	A	01-01-2013
1	B	01-01-2013

a	id	date	b
2	A	01-01-2013	1
1	B	01-01-2013	1

`dt_a[dt_b, on = .(id = id, date = date), roll = TRUE]` – join data.tables on matching rows in id columns but only keep the most recent preceding match with the left data.table according to date columns. “`roll = -Inf`” reverses direction.

## BIND

a	b

a	b

a	b

a	b

`rbind(dt_a, dt_b)` – combine rows of two data.tables.

a	b

x	y

a	b	x	y

`cbind(dt_a, dt_b)` – combine columns of two data.tables.

## Apply function to cols.

### APPLY A FUNCTION TO MULTIPLE COLUMNS

a	b
1	4
2	5
3	6

`dt[, lapply(.SD, mean), .SDcols = c("a", "b")]` – apply a function – e.g. `mean()`, `as.character()`, `which.max()` – to columns specified in `.SDcols` with `lapply()` and the `.SD` symbol. Also works with groups.

a	b
1	1
2	2
3	3

`cols <- c("a")`  
`dt[, paste0(cols, "_m") := lapply(.SD, mean), .SDcols = cols]` – apply a function to specified columns and assign the result with suffixed variable names to the original data.

## RESHAPE TO WIDE FORMAT

id	y	a	b
A	x	1	3
A	z	2	4
B	x	1	3
B	z	2	4

`dcast(dt,`  
`id ~ y,`  
`value.var = c("a", "b"))`

Reshape a data.table from long to wide format.

`dt` A data.table.  
`id ~ y` Formula with a LHS: ID columns containing IDs for multiple entries. And a RHS: columns with values to spread in column headers.  
`value.var` Columns containing values to fill into cells.

## RESHAPE TO LONG FORMAT

id	a	x	a	z	b	x	b	z
A	1	2	3	4	A	1	1	3
B	1	2	3	4	B	1	1	3

`melt(dt,`  
`id.vars = c("id"),`  
`measure.vars = patterns("^a", "^b"),`  
`variable.name = "y",`  
`value.name = c("a", "b"))`

Reshape a data.table from wide to long format.

`dt` A data.table.  
`id.vars` ID columns with IDs for multiple entries.  
`measure.vars` Columns containing values to fill into cells (often in pattern form).  
`variable.name, value.name` Names of new columns for variables and values derived from old headers.

a	b
1	a
2	a
3	b
4	b
5	b

`dt[, c := shift(a, 1), by = b]` – within groups, duplicate a column with rows lagged by specified amount.  
`dt[, c := shift(a, 1, type = "lead"), by = b]` – within groups, duplicate a column with rows leading by specified amount.

## read & write files

### IMPORT

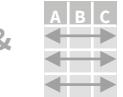
# Data transformation with dplyr :: CHEATSHEET



dplyr functions work with pipes and expect **tidy data**. In tidy data:



Each **variable** is in its own **column**



Each **observation**, or **case**, is in its own **row**

## pipes

$x |> f(y)$  becomes  $f(x, y)$

## Summarize Cases

Apply **summary functions** to columns to create a new table of summary statistics. Summary functions take vectors as input and return one value (see back).



**summarize(.data, ...)**  
Compute table of summaries.  
`mtcars |> summarize(avg = mean(mpg))`

**count(.data, ..., wt = NULL, sort = FALSE, name = NULL)** Count number of rows in each group defined by the variables in ... Also **tally()**, **add\_count()**, **add\_tally()**.  
`mtcars |> count(cyl)`

## Group Cases

Use **group\_by(.data, ..., .add = FALSE, .drop = TRUE)** to create a "grouped" copy of a table grouped by columns in ... dplyr functions will manipulate each "group" separately and combine the results.

`mtcars |> group_by(cyl) |> summarize(avg = mean(mpg))`

Use **rowwise(.data, ...)** to group data into individual rows. dplyr functions will compute results for each row. Also apply functions to list-columns. See tidyverse cheat sheet for list-column workflow.

`starwars |> rowwise() |> mutate(film_count = length(films))`

**ungroup(x, ...)** Returns ungrouped copy of table.  
`g_mtcars <- mtcars |> group_by(cyl)  
ungroup(g_mtcars)`

## Manipulate Cases

### EXTRACT CASES

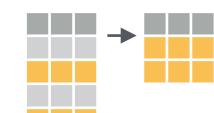
Row functions return a subset of rows as a new table.



**filter(.data, ..., .preserve = FALSE)** Extract rows that meet logical criteria.  
`mtcars |> filter(mpg > 20)`



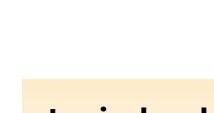
**distinct(.data, ..., .keep\_all = FALSE)** Remove rows with duplicate values.  
`mtcars |> distinct(gear)`



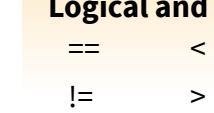
**slice(.data, ..., .preserve = FALSE)** Select rows by position.  
`mtcars |> slice(10:15)`



**slice\_sample(.data, ..., n, prop, weight\_by = NULL, replace = FALSE)** Randomly select rows. Use n to select a number of rows and prop to select a fraction of rows.  
`mtcars |> slice_sample(n = 5, replace = TRUE)`



**slice\_min(.data, order\_by, ..., n, prop, with\_ties = TRUE)** and **slice\_max()** Select rows with the lowest and highest values.  
`mtcars |> slice_min(mpg, prop = 0.25)`



**slice\_head(.data, ..., n, prop)** and **slice\_tail()**  
Select the first or last rows.  
`mtcars |> slice_head(n = 5)`

### Logical and boolean operators to use with filter()

<code>==</code>	<code>&lt;</code>	<code>&lt;=</code>	<code>is.na()</code>	<code>%in%</code>	<code> </code>	<code>xor()</code>
<code>!=</code>	<code>&gt;</code>	<code>&gt;=</code>	<code>!is.na()</code>	<code>!</code>	<code>&amp;</code>	

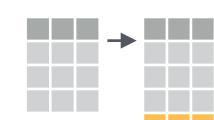
See [?base::Logic](#) and [?Comparison](#) for help.

### ARRANGE CASES



**arrange(.data, ..., .by\_group = FALSE)** Order rows by values of a column or columns (low to high), use with **desc()** to order from high to low.  
`mtcars |> arrange(mpg)  
mtcars |> arrange(desc(mpg))`

### ADD CASES



**add\_row(.data, ..., .before = NULL, .after = NULL)**  
Add one or more rows to a table.  
`cars |> add_row(speed = 1, dist = 1)`

## Manipulate Variables

### EXTRACT VARIABLES

Column functions return a set of columns as a new vector or table.



**pull(.data, var = -1, name = NULL, ...)** Extract column values as a vector, by name or index.  
`mtcars |> pull(wt)`



**select(.data, ...)** Extract columns as a table.  
`mtcars |> select(mpg, wt)`



**relocate(.data, ..., .before = NULL, .after = NULL)** Move columns to new position.  
`mtcars |> relocate(mpg, cyl, .after = last_col())`

### Use these helpers with select() and across()

e.g. `mtcars |> select(mpg:cyl)`

**contains(match)**

**ends\_with(match)**

**starts\_with(match)**

**num\_range(prefix, range)**

**all\_of(x)/any\_of(x, ..., vars)**

! e.g., mpg:cyl

**matches(match)**

### MANIPULATE MULTIPLE VARIABLES AT ONCE

`df <- tibble(x_1 = c(1, 2), x_2 = c(3, 4), y = c(4, 5))`



**across(.cols, .funs, ..., .names = NULL)** Summarize or mutate multiple columns in the same way.  
`df |> summarize(across(everything()), mean))`



**c\_across(.cols)** Compute across columns in row-wise data.  
`df |> rowwise() |> mutate(x_total = sum(c_across(1:2)))`

### MAKE NEW VARIABLES

Apply **vectorized functions** to columns. Vectorized functions take vectors as input and return vectors of the same length as output (see back).

**mutate(.data, ..., .keep = "all", .before = NULL, .after = NULL)** Compute new column(s). Also **add\_column()**.  
`mtcars |> mutate(gpm = 1 / mpg)  
mtcars |> mutate(gpm = 1 / mpg, .keep = "none")`

**rename(.data, ...)** Rename columns. Use **rename\_with()** to rename with a function.  
`mtcars |> rename(miles_per_gallon = mpg)`



# Vectorized Functions

## TO USE WITH MUTATE ()

**mutate()** applies vectorized functions to columns to create new columns. Vectorized functions take vectors as input and return vectors of the same length as output.

vectorized function →

## OFFSET

dplyr::lag() - offset elements by 1  
dplyr::lead() - offset elements by -1

## CUMULATIVE AGGREGATE

dplyr::cumall() - cumulative all()  
dplyr::cumany() - cumulative any()  
cummax() - cumulative max()  
dplyr::cummean() - cumulative mean()  
cummin() - cumulative min()  
cumprod() - cumulative prod()  
cumsum() - cumulative sum()

## RANKING

dplyr::cume\_dist() - proportion of all values <=  
dplyr::dense\_rank() - rank w ties = min, no gaps  
dplyr::min\_rank() - rank with ties = min  
dplyr::ntile() - bins into n bins  
dplyr::percent\_rank() - min\_rank scaled to [0,1]  
dplyr::row\_number() - rank with ties = "first"

## MATH

+, -, \*, /, ^, %/%, %% - arithmetic ops  
log(), log2(), log10() - logs  
<, <=, >, >=, !=, == - logical comparisons  
dplyr::between() - x >= left & x <= right  
dplyr::near() - safe == for floating point numbers

## MISCELLANEOUS

dplyr::case\_when() - multi-case if\_else()  
starwars |>  
  mutate(type = case\_when(  
    height > 200 | mass > 200 ~ "large",  
    species == "Droid" ~ "robot",  
    TRUE ~ "other"))  
dplyr::coalesce() - first non-NA values by element across a set of vectors  
dplyr::if\_else() - element-wise if() + else()  
dplyr::na\_if() - replace specific values with NA  
pmax() - element-wise max()  
pmin() - element-wise min()

# Summary Functions

## TO USE WITH SUMMARIZE ()

**summarize()** applies summary functions to columns to create a new table. Summary functions take vectors as input and return single values as output.

summary function →

## COUNT

dplyr::n() - number of values/rows  
dplyr::n\_distinct() - # of uniques  
sum(!is.na()) - # of non-NAs

## POSITION

mean() - mean, also mean(!is.na())  
median() - median

## LOGICAL

mean() - proportion of TRUEs  
sum() - # of TRUEs

## ORDER

dplyr::first() - first value  
dplyr::last() - last value  
dplyr::nth() - value in nth location of vector

## RANK

quantile() - nth quantile  
min() - minimum value  
max() - maximum value

## SPREAD

IQR() - Inter-Quartile Range  
mad() - median absolute deviation  
sd() - standard deviation  
var() - variance

# Row Names

Tidy data does not use rownames, which store a variable outside of the columns. To work with the rownames, first move them into a column.

A B → C A B tibble::rownames\_to\_column()  
1 a t → 1 a Move row names into col.  
2 b u → 2 b a <- mtcars |>  
3 c v → 3 c rownames\_to\_column(var = "C")

A B C → A B tibble::column\_to\_rownames()  
1 a t → 1 a Move col into row names.  
2 b u → 2 b a |> column\_to\_rownames(var = "C")  
3 c v → 3 c

Also tibble::has\_rownames() and tibble::remove\_rownames().

# Combine Tables

## COMBINE VARIABLES

X	y
A B C	E F G
a t 1	a t 3
b u 2	b u 2
c v 3	d w 1

**bind\_cols(..., .name\_repair)** Returns tables placed side by side as a single table. Column lengths must be equal. Columns will NOT be matched by id (to do that look at Relational Data below), so be sure to check that both tables are ordered the way you want before binding.

## RELATIONAL DATA

Use a "Mutating Join" to join one table to columns from another, matching values with the rows that they correspond to. Each join retains a different combination of values from the tables.

A B C D	left_join(x, y, by = NULL, copy = FALSE, suffix = c(".x", ".y"), ..., keep = FALSE, na_matches = "na")
a t 1 3	Join matching values from y to x.
b u 2 2	
c v 3 NA	

A B C D	right_join(x, y, by = NULL, copy = FALSE, suffix = c(".x", ".y"), ..., keep = FALSE, na_matches = "na")
a t 1 3	Join matching values from x to y.
b u 2 2	
d w NA 1	

A B C D	inner_join(x, y, by = NULL, copy = FALSE, suffix = c(".x", ".y"), ..., keep = FALSE, na_matches = "na")
a t 1 3	Join data. Retain only rows with matches.
b u 2 2	

A B C D	full_join(x, y, by = NULL, copy = FALSE, suffix = c(".x", ".y"), ..., keep = FALSE, na_matches = "na")
a t 1 3	Join data. Retain all values, all rows.
b u 2 2	
c v 3 NA 1	

## COLUMN MATCHING FOR JOINS

A B x C B y D	Use <b>by = c("col1", "col2", ...)</b> to specify one or more common columns to match on.
a t 1 t 3	left_join(x, y, by = "A")
b u 2 u 2	
c v 3 NA NA	

A x B x C A y B y	Use a named vector, <b>by = c("col1" = "col2")</b> , to match on columns that have different names in each table.
a t 1 d w	left_join(x, y, by = c("C" = "D"))
b u 2 b u	
c v 3 a t	

A1 B1 C A2 B2	Use <b>suffix</b> to specify the suffix to give to unmatched columns that have the same name in both tables.
a t 1 d w	left_join(x, y, by = c("C" = "D"), suffix = c("1", "2"))
b u 2 b u	
c v 3 a t	

## COMBINE CASES

X	y
A B C	A B C
a t 1	a t 1
b u 2	b u 2
c v 3	d w 4

**bind\_rows(..., id = NULL)** Returns tables one on top of the other as a single table. Set **.id** to a column name to add a column of the original table names (as pictured).

Use a "Filtering Join" to filter one table against the rows of another.

X	y
A B C	A B C
a t 1	a t 3
b u 2	b u 2
c v 3	d w 1

**semi\_join(x, y, by = NULL, copy = FALSE, ..., na\_matches = "na")** Return rows of x that have a match in y. Use to see what will be included in a join.

**anti\_join(x, y, by = NULL, copy = FALSE, ..., na\_matches = "na")** Return rows of x that do not have a match in y. Use to see what will not be included in a join.

Use a "Nest Join" to inner join one table to another into a nested data frame.

A B C	y
a t 1	<tibble [1x2]>
b u 2	<tibble [1x2]>
c v 3	<tibble [1x2]>

## SET OPERATIONS

**intersect(x, y, ...)** Rows that appear in both x and y.



**setdiff(x, y, ...)** Rows that appear in x but not y.



**union(x, y, ...)** Rows that appear in x or y, duplicates removed). **union\_all()** retains duplicates.



Use **setequal()** to test whether two data sets contain the exact same rows (in any order).





# DeclareDesign:: CHEAT SHEET

## Model

What is your model of the world, including how outcomes respond to interventions in the world?

## Population

Define the size of the population, hierarchical structure (if any), and background variables.

Simple dataset with no background variables

```
pop <- declare_population(N = 100)
pop()
```

Simple dataset with background variables

```
declare_population(N = 100,
                  X = rnorm(N))
```

Two-level dataset

```
declare_population(
  schools =
    add_level(N = 10,
              funding = rnorm(N)),
  students =
    add_level(N = 100,
              scores = rnorm(N))
)
```

## Outcomes

### Outcomes that depend on a treatment (Z)

Using a formula

```
declare_potential_outcomes(
  Y ~ .5 * Z + rnorm(N))
```

As separate variables

```
declare_potential_outcomes(
  Y_Z_0 = rnorm(N),
  Y_Z_1 = Y_Z_0 + .5)
```

### Outcomes that do not depend on treatment

```
declare_potential_outcomes(
  Y = rnorm(N))
```

## Inquiry

What is the research question you want to answer?

Causal inquiries

```
declare_estimand(
  ATE = mean(Y_Z_1 - Y_Z_0))
```

Descriptive inquiries

```
declare_estimand(
  Y_median = median(Y))
```

Conditional estimands

```
declare_estimand(
  LATE = mean(Y_Z_1 - Y_Z_0),
  subset = complier == TRUE)
```

## Data Strategy

How will you generate data to answer your inquiry?

### Sampling

```
declare_sampling(n = 100)
```

```
declare_sampling(
  strata_n = 20,
  strata = urban_area)
```

### Treatment assignment

```
declare_assignment(m = 100)
```

```
declare_assignment(
  clusters = villages,
  m = 10)
```

## Answer Strategy

How will you generate an answer to your inquiry?

OLS with robust standard errors

```
declare_estimator(
  Y ~ Z, model = lm_robust)
```

2SLS instrumental variables regression with robust SEs

```
declare_estimator(
  Y ~ D | Z, model = iv_robust)
```

Difference-in-means

```
declare_estimator(
  Y ~ Z,
  model = difference_in_means)
```

**DeclareDesign** is a software implementation of the MIDA framework, according to which research designs have a **Model** of the world, an **Inquiry** about that model, a **Data strategy** that generates information about the world, and an **Answer** strategy that uses data to make a guess about the **Inquiry**. Declared designs can be “diagnosed” to calculate the properties of the design such as power and bias using Monte Carlo simulation.

All `declare_*` functions return *functions*. Most functions take a `data.frame` and return a `data.frame`.

## Design Declaration

Put together all the steps into a declared design using the `+` operator

```
design <-
  declare_population(N = 200, X = rnorm(N)) +
  declare_potential_outcomes(Y ~ .5 * Z + X) +
  declare_estimand(ATE = mean(Y_Z_1 - Y_Z_0)) +
  declare_sampling(n = 100) +
  declare_assignment(m = 50) +
  declare_estimator(Y ~ Z, model = lm_robust)
```

```
draw_data(design)
draw_estimates(design)
get_estimates(design, data = real_data)
draw_estimands(design)
run_design(design)
summary(design)
compare_designs(design_1, design_2)
```

## Design Diagnosis

Diagnose the properties of your design

```
diagnosis <- diagnose_design(
  design, sims = 100, bootstrap_sims = 100)
```

```
summary(diagnosis)
get_diagnosands(diagnosis)
get_simulations(diagnosis)
```

Custom diagnosands

```
diagnose_design(
  design,
  diagnosands = declare_diagnosands(
    sig_pos = mean(p.value < .05 & estimate > 0)))
```

# Create, query and manipulate distributions with distr6 :: CHEAT SHEET



## Introduction

distr6 is an object-oriented interface for probability distributions. Including distributions as objects, statistical properties of distributions, composite modelling and decorators for numerical imputation. As well as this cheat sheet, see:

- [GitHub](#) for an issue tracker and latest development branch
- [CRAN](#) for package meta-data
- The distr6 [website](#) for more complete tutorials.

## R6 Classes

<b>Distribution</b>	The parent class to most distr6 classes.	<code>Distribution</code>
<b>SDistribution</b>	Class given to all probability distributions implemented in distr6.	<code>Distribution</code> ↑ <code>SDistribution</code>
<b>Kernel</b>	Class given to all kernel-like probability distributions.	<code>Distribution</code> ↑ <code>Kernel</code>
<b>Decorator</b>	Used to add or impute methods to a Distribution.	
<b>Wrapper</b>	Create composite distributions by adapting class properties	
<b>ParameterSet</b>	Class used to add parameters to a distribution.	

## R6 Basics

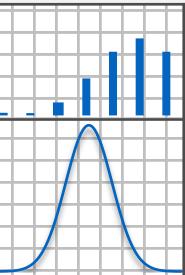
\$ All methods are called using dollar-sign notation	<code>N &lt;- Normal\$new() N\$mean() N\$pdf(2)</code>
<b>clone</b> Objects are copied using the clone method	<code>N1 &lt;- Normal\$new() N2 &lt;- N1\$clone()</code>
<b>Method chaining</b> Call one method after another	<code>Normal\$new()\$pdf(2)</code>

## Construct a Distribution

Each distribution has a default parameterisation, and all common parameterisations are available.

```
Binomial$new()
Binomial$new(size=5, prob=0.6)
Binomial$new(size=5, qprob=0.4)
```

```
Normal$new()
Normal$new(mean=0, sd=1)
Normal$new(mean=0, var=1)
Normal$new(mean=0, prec=1)
```



You can list all the implemented probability distributions and kernels

```
listDistributions()
listKernels()
```

## S3 and Piping

distr6 uses '[R6toS3](#)' so every R6 method has an S3 dispatch available.

<code>N &lt;- Normal\$new()</code>	<code>N\$mean()</code> ➔ <code>mean(N)</code>
	<code>N\$getParameterValue("mean")</code> ➔ <code>getParameterValue(N, "mean")</code>
	<code>N\$pdf(1:5)</code> ➔ <code>pdf(N, 1:5)</code>

Use the 'magrittr' package for method chaining and piping (%>%).

<code>&gt; N &lt;- Normal\$new()</code>	<code>&gt; N\$setParameterValue(sd=2)\$getParameterValue("var")</code>
	↓ library(magrittr)
<code>&gt; N &lt;- Normal\$new()</code>	<code>&gt; N %&gt;% setParameterValue(sd=2) %&gt;% getParameterValue("var")</code>

## Multivariate Distributions

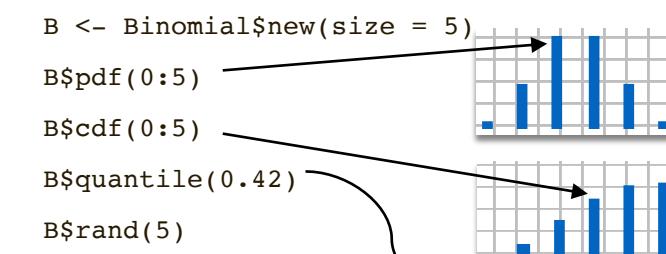
Multivariate distributions are handled just like univariate distributions, except the pdf/cdf functions take multiple arguments, as do cf and mgf where available.

<code>&gt; MN &lt;- MultivariateNormal\$new(mean = c(0,0,0), cov = c(3,-1,-1,-1,1,0,-1,0,1))</code>	
	<code>&gt; MN &lt;- MultivariateNormal\$new(mean = c(0,0,0), prec = c(3,-1,-1,-1,1,0,-1,0,1))</code>
	<code>&gt; MN\$pdf(1, 2, 3)</code>
	<code>&gt; MN\$cdf(1, 1, 1)</code>
	Once again vectorization is available
	<code>&gt; MN\$pdf(1:2, 2:3, 1:2)</code>
	<code>&gt; MN\$cdf(c(0.45, 0.65), c(0.12, 0.99), c(0, 1))</code>

## Statistical Methods

```
N <- Normal$new()
N$mean()
N$variance()
N$skewness()
N$kurtosis()
N$entropy()
```

Use ?SDistribution, ?Normal (or any other distribution) to see available methods.

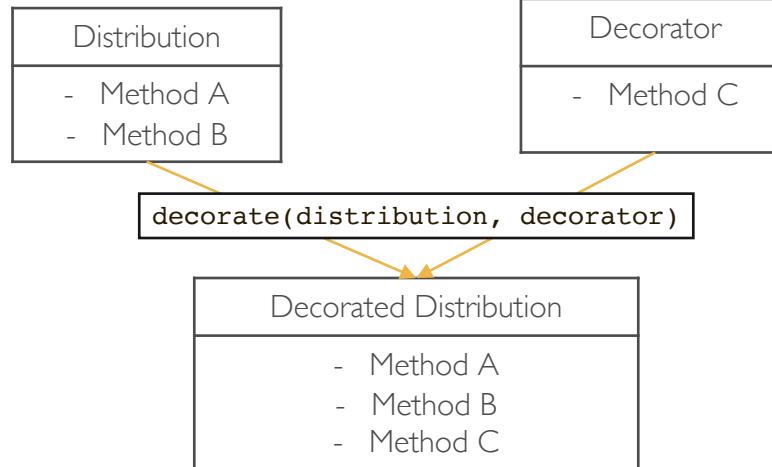


# Create, query and manipulate distributions with distr6 :: CHEAT SHEET



## Decorators

Decorators are a design pattern (Gamma et al., 1994) used to add methods to objects.



## Available Decorators

**CoreStatistics** Imputes common numeric statistical results, adds generalised expectation and moments function.

**ExoticStatistics** Adds methods for survival analysis and statistical modelling.

**FunctionImputation** Uses numerical methods to impute missing pdf/cdf/quantile/rand functions

Remember to decorate first before using a method from a decorator

```

> N <- Normal$new()
> N$survival(1)
Error: attempt to apply non-function
> decorate(N, ExoticStatistics)
> N$survival(1)
[1] 0.1586553
  
```

S3 methods will now work too

```

> N <- Normal$new(decorators = ExoticStatistics)
> pdfPNorm(N, 3, -1, 1)
[1] 0.4383636
  
```

Use listing to see which decorators are currently implemented.

```
listDecorators()
```

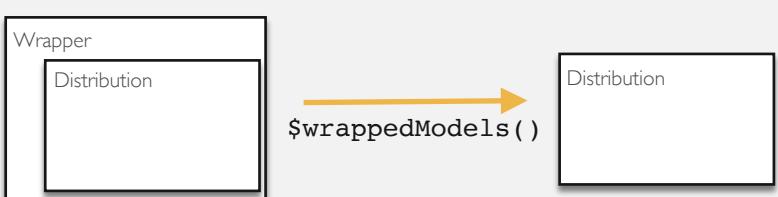
## Wrappers

Wrappers are based on the **Adapter** design pattern (Gamma et al., 1994) and are used to change the interface of an object.



## Available Wrappers

ProductDistribution	VectorDistribution
Product of two or more distributions.	Vectorizes two or more distributions.
Convolution	Addition (or subtraction) of two distributions
HuberizedDistribution	MixtureDistribution
Huberizes a distribution between limits.	Weighted mixture of two or more distributions
TruncatedDistribution	
	Truncates a distribution between limits.



```

> TruncatedDistribution$new(Normal$new(),
  lower = -1, upper = 1)
> MixtureDistribution$new(list(Binomial$new(),
  Normal$new()), weights = c(0.4, 0.6))
> ProductDistribution$new(list(Exponential$new(),
  Normal$new()))$pdf(1,1)
  
```

Use listing to see which wrappers are currently implemented.

```
listWrappers()
```

## Custom Distributions

Custom distributions can be created using `Distribution$new`, this is not the same as implementing a new `SDistribution`!

```

pdf <-
function(x1) return(1/(self$getParameterValue("upper") - self$getParameterValue("lower")))
  
```

The `self` argument tells the object to call the method on itself.

All `pdf/cdf` methods in `distr6` use '`x1,x2,...`' as their arguments

```

cdf <- function(x1) return((x1 -
self$getParameterValue("lower")) /
(self$getParameterValue("upper") -
self$getParameterValue("lower")))
  
```

`ParameterSet` is the class used for `distr6` parameters.

```

ps <- ParameterSet$new(id = list("lower", "upper"),
value = c(1,10), support =
list(Reals$new(), Reals$new()), settable =
list(TRUE, TRUE))
  
```

The argument `support` is of type `SetInterval`. See `listSpecialSets()`

Unique distribution name and one-word `short_name` (`ID`)

```

dist <- Distribution$new(name = "Uniform",
short_name = "unif", type = Reals$new(), support =
Interval$new(1, 10), symmetric = TRUE, pdf = pdf,
cdf = cdf, parameters = ps, description = "Custom
uniform distribution", decorators = CoreStatistics)
  
```

Distribution type and support is of type `SetInterval`.

`CoreStatistics` decorator is optionally used to impute numeric results.

`log` and `lower.tail` arguments are added automatically

```

> dist$pdf(1, log = TRUE)
[1] -2.197225
> dist$cdf(2, lower.tail = FALSE)
[1] 0.8888889
> decorate(dist, FunctionImputation)
> dist$mean()
[1] 5.5
> dist$quantile(0.42)
[1] 4.78
  
```

impute missing `quantile` and `rand` methods

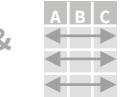
# Data transformation with dplyr :: CHEATSHEET



dplyr functions work with pipes and expect **tidy data**. In tidy data:



Each **variable** is in its own **column**



Each **observation**, or **case**, is in its own **row**

## pipes

$x |> f(y)$  becomes  $f(x, y)$

## Summarize Cases

Apply **summary functions** to columns to create a new table of summary statistics. Summary functions take vectors as input and return one value (see back).



→ **summarize(.data, ...)**  
Compute table of summaries.  
mtcars |> summarize(avg = mean(mpg))

→ **count(.data, ..., wt = NULL, sort = FALSE, name = NULL)** Count number of rows in each group defined by the variables in ... Also **tally()**, **add\_count()**, **add\_tally()**.  
mtcars |> count(cyl)

## Group Cases

Use **group\_by(.data, ..., .add = FALSE, .drop = TRUE)** to create a "grouped" copy of a table grouped by columns in ... dplyr functions will manipulate each "group" separately and combine the results.

→ → mtcars |>  
group\_by(cyl) |>  
summarize(avg = mean(mpg))

Use **rowwise(.data, ...)** to group data into individual rows. dplyr functions will compute results for each row. Also apply functions to list-columns. See tidyverse cheat sheet for list-column workflow.

→ → starwars |>  
rowwise() |>  
mutate(film\_count = length(films))

**ungroup(x, ...)** Returns ungrouped copy of table.  
g\_mtcars <- mtcars |> group\_by(cyl)  
ungroup(g\_mtcars)

## Manipulate Cases

### EXTRACT CASES

Row functions return a subset of rows as a new table.



**filter(.data, ..., .preserve = FALSE)** Extract rows that meet logical criteria.  
mtcars |> filter(mpg > 20)



**distinct(.data, ..., .keep\_all = FALSE)** Remove rows with duplicate values.  
mtcars |> distinct(gear)



**slice(.data, ..., .preserve = FALSE)** Select rows by position.  
mtcars |> slice(10:15)



**slice\_sample(.data, ..., n, prop, weight\_by = NULL, replace = FALSE)** Randomly select rows. Use n to select a number of rows and prop to select a fraction of rows.  
mtcars |> slice\_sample(n = 5, replace = TRUE)



**slice\_min(.data, order\_by, ..., n, prop, with\_ties = TRUE)** and **slice\_max()** Select rows with the lowest and highest values.  
mtcars |> slice\_min(mpg, prop = 0.25)



**slice\_head(.data, ..., n, prop)** and **slice\_tail()**  
Select the first or last rows.  
mtcars |> slice\_head(n = 5)

### Logical and boolean operators to use with filter()

<code>==</code>	<code>&lt;</code>	<code>&lt;=</code>	<code>is.na()</code>	<code>%in%</code>	<code> </code>	<code>xor()</code>
<code>!=</code>	<code>&gt;</code>	<code>&gt;=</code>	<code>!is.na()</code>	<code>!</code>	<code>&amp;</code>	

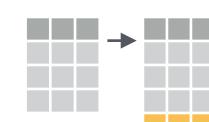
See [?base::Logic](#) and [?Comparison](#) for help.

### ARRANGE CASES



**arrange(.data, ..., .by\_group = FALSE)** Order rows by values of a column or columns (low to high), use with **desc()** to order from high to low.  
mtcars |> arrange(mpg)  
mtcars |> arrange(desc(mpg))

### ADD CASES



**add\_row(.data, ..., .before = NULL, .after = NULL)**  
Add one or more rows to a table.  
cars |> add\_row(speed = 1, dist = 1)

## Manipulate Variables

### EXTRACT VARIABLES

Column functions return a set of columns as a new vector or table.



**pull(.data, var = -1, name = NULL, ...)** Extract column values as a vector, by name or index.  
mtcars |> pull(wt)



**select(.data, ...)** Extract columns as a table.  
mtcars |> select(mpg, wt)



**relocate(.data, ..., .before = NULL, .after = NULL)**  
Move columns to new position.  
mtcars |> relocate(mpg, cyl, .after = last\_col())

### Use these helpers with select() and across()

e.g. mtcars |> select(mpg:cyl)

**contains(match)**

**ends\_with(match)**

**starts\_with(match)**

**num\_range(prefix, range)**

**all\_of(x)/any\_of(x, ..., vars)**

**matches(match)**

; e.g., mpg:cyl

!, e.g., !gear

**everything()**

### MANIPULATE MULTIPLE VARIABLES AT ONCE

df <- tibble(x\_1 = c(1, 2), x\_2 = c(3, 4), y = c(4, 5))



**across(.cols, .funs, ..., .names = NULL)** Summarize or mutate multiple columns in the same way.  
df |> summarize(across(everything(), mean))



**c\_across(.cols)** Compute across columns in row-wise data.  
df |>  
rowwise() |>  
mutate(x\_total = sum(c\_across(1:2)))

### MAKE NEW VARIABLES

Apply **vectorized functions** to columns. Vectorized functions take vectors as input and return vectors of the same length as output (see back).



**mutate(.data, ..., .keep = "all", .before = NULL, .after = NULL)** Compute new column(s). Also **add\_column()**.  
mtcars |> mutate(gpm = 1 / mpg)  
mtcars |> mutate(gpm = 1 / mpg, .keep = "none")



**rename(.data, ...)** Rename columns. Use **rename\_with()** to rename with a function.  
mtcars |> rename(miles\_per\_gallon = mpg)



# Vectorized Functions

## TO USE WITH MUTATE ()

**mutate()** applies vectorized functions to columns to create new columns. Vectorized functions take vectors as input and return vectors of the same length as output.

vectorized function →

## OFFSET

dplyr::lag() - offset elements by 1  
dplyr::lead() - offset elements by -1

## CUMULATIVE AGGREGATE

dplyr::cumall() - cumulative all()  
dplyr::cumany() - cumulative any()  
cummax() - cumulative max()  
dplyr::cummean() - cumulative mean()  
cummin() - cumulative min()  
cumprod() - cumulative prod()  
cumsum() - cumulative sum()

## RANKING

dplyr::cume\_dist() - proportion of all values <=  
dplyr::dense\_rank() - rank w ties = min, no gaps  
dplyr::min\_rank() - rank with ties = min  
dplyr::ntile() - bins into n bins  
dplyr::percent\_rank() - min\_rank scaled to [0,1]  
dplyr::row\_number() - rank with ties = "first"

## MATH

+, -, \*, /, ^, %/%, %% - arithmetic ops  
log(), log2(), log10() - logs  
<, <=, >, >=, !=, == - logical comparisons  
dplyr::between() - x >= left & x <= right  
dplyr::near() - safe == for floating point numbers

## MISCELLANEOUS

dplyr::case\_when() - multi-case if\_else()  
starwars |>  
  mutate(type = case\_when(  
    height > 200 | mass > 200 ~ "large",  
    species == "Droid" ~ "robot",  
    TRUE ~ "other"))  
dplyr::coalesce() - first non-NA values by element across a set of vectors  
dplyr::if\_else() - element-wise if() + else()  
dplyr::na\_if() - replace specific values with NA  
pmax() - element-wise max()  
pmin() - element-wise min()

# Summary Functions

## TO USE WITH SUMMARIZE ()

**summarize()** applies summary functions to columns to create a new table. Summary functions take vectors as input and return single values as output.

summary function →

## COUNT

dplyr::n() - number of values/rows  
dplyr::n\_distinct() - # of uniques  
sum(!is.na()) - # of non-NAs

## POSITION

mean() - mean, also mean(!is.na())  
median() - median

## LOGICAL

mean() - proportion of TRUEs  
sum() - # of TRUEs

## ORDER

dplyr::first() - first value  
dplyr::last() - last value  
dplyr::nth() - value in nth location of vector

## RANK

quantile() - nth quantile  
min() - minimum value  
max() - maximum value

## SPREAD

IQR() - Inter-Quartile Range  
mad() - median absolute deviation  
sd() - standard deviation  
var() - variance

# Row Names

Tidy data does not use rownames, which store a variable outside of the columns. To work with the rownames, first move them into a column.

A B → C A B tibble::rownames\_to\_column()  
1 a t → 1 a Move row names into col.  
2 b u → 2 b a <- mtcars |>  
3 c v → 3 c rownames\_to\_column(var = "C")

A B C → A B tibble::column\_to\_rownames()  
1 a t → 1 a Move col into row names.  
2 b u → 2 b a |> column\_to\_rownames(var = "C")  
3 c v → 3 c

Also tibble::has\_rownames() and tibble::remove\_rownames().

# Combine Tables

## COMBINE VARIABLES

X	y
A B C	E F G
a t 1	a t 3
b u 2	b u 2
c v 3	d w 1

**bind\_cols(..., .name\_repair)** Returns tables placed side by side as a single table. Column lengths must be equal. Columns will NOT be matched by id (to do that look at Relational Data below), so be sure to check that both tables are ordered the way you want before binding.

## RELATIONAL DATA

Use a "Mutating Join" to join one table to columns from another, matching values with the rows that they correspond to. Each join retains a different combination of values from the tables.

A B C D	left_join(x, y, by = NULL, copy = FALSE, suffix = c(".x", ".y"), ..., keep = FALSE, na_matches = "na")
a t 1 3	Join matching values from y to x.
b u 2 2	
c v 3 NA	

A B C D	right_join(x, y, by = NULL, copy = FALSE, suffix = c(".x", ".y"), ..., keep = FALSE, na_matches = "na")
a t 1 3	Join matching values from x to y.
b u 2 2	
d w NA 1	

A B C D	inner_join(x, y, by = NULL, copy = FALSE, suffix = c(".x", ".y"), ..., keep = FALSE, na_matches = "na")
a t 1 3	Join data. Retain only rows with matches.
b u 2 2	

A B C D	full_join(x, y, by = NULL, copy = FALSE, suffix = c(".x", ".y"), ..., keep = FALSE, na_matches = "na")
a t 1 3	Join data. Retain all values, all rows.
b u 2 2	
c v 3 NA 1	

## COLUMN MATCHING FOR JOINS

A B x C B y D	Use <b>by = c("col1", "col2", ...)</b> to specify one or more common columns to match on.
a t 1 t 3	left_join(x, y, by = "A")
b u 2 u 2	
c v 3 NA NA	

A x B x C A y B y	Use a named vector, <b>by = c("col1" = "col2")</b> , to match on columns that have different names in each table.
a t 1 d w	left_join(x, y, by = c("C" = "D"))
b u 2 b u	
c v 3 a t	

A1 B1 C A2 B2	Use <b>suffix</b> to specify the suffix to give to unmatched columns that have the same name in both tables.
a t 1 d w	left_join(x, y, by = c("C" = "D"), suffix = c("1", "2"))
b u 2 b u	
c v 3 a t	

## COMBINE CASES

X	y
A B C	A B C
a t 1	a t 1
b u 2	b u 2
c v 3	c v 3

**bind\_rows(..., id = NULL)**

Returns tables one on top of the other as a single table. Set `.id` to a column name to add a column of the original table names (as pictured).

X	y
A B C	A B C
a t 1	a t 1
b u 2	b u 2
c v 3	c v 3

**semi\_join(x, y, by = NULL, copy = FALSE, ..., na\_matches = "na")**

Return rows of x that have a match in y. Use to see what will be included in a join.

**anti\_join(x, y, by = NULL, copy = FALSE, ..., na\_matches = "na")**

Return rows of x that do not have a match in y. Use to see what will not be included in a join.

Use a "Nest Join" to inner join one table to another into a nested data frame.

A B C	y
a t 1	<tibble [1x2]>
b u 2	<tibble [1x2]>
c v 3	<tibble [1x2]>

## SET OPERATIONS

**intersect(x, y, ...)**

Rows that appear in both x and y.



**setdiff(x, y, ...)**

Rows that appear in x but not y.



**union(x, y, ...)**

Rows that appear in x or y, duplicates removed). **union\_all()** retains duplicates.



Use **setequal()** to test whether two data sets contain the exact same rows (in any order).



# DRomics :: CHEAT SHEET

Written by the authors of the DRomics package - Updated in january 2023  
See <https://lbbi.univ-lyon1.fr/fr/dromics>  
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## Format of data

Data can be imported from a **.txt file** containing one row per item after a first row giving the doses or concentrations for each sample, with the first column corresponding to the identifier of each item.

Alternatively an R object of class **data.frame** can be directly given as input, corresponding to the output of `read.table(file, header = FALSE)` on a file described as above.

`formatdata4DRomics()` can be used to help formatting such an R object.

	Identifiers of items (contigs, probes, metabolites, ...)			
RefSeq	0	0	0.22	0.2
NM_144958	2072	2506	2519	2111
NR_102758	0	0	0	0
NM_172405	198	265	250	241
NM_029777	18	29	25	16
NM_0011301	0	0	0	0
NM_0011623	3	1	2	0
NM_008117	0	0	0	0
NM_0011682	61	65	79	81
NM_0010910	7	10	9	0
NR_002862	139	172	165	159
NR_033520	318	407	425	43

Tested doses or concentrations

Signal (counts of reads, continuous signal in log2, ...)

## Workflow for analysis of data

See below the functions with their main arguments (see help pages for their complete description).

### Step 1: import, check and pretreatment

```
microarraydata(file, norm.method = c("cyclicloess", "quantile", "scale",
"none"))
RNaseqdata(file, transfo.method = c("rlog", "vst"))
continuousomicdata(file)
continuousanchoringdata(file)
```

### Step 2: selection of significantly responsive items

```
itemselect(omicdata, select.method = c("quadratic", "linear", "ANOVA"),
FDR = 0.05)
```

### Step 3: dose-response modelling for responsive items

```
drcfit(itemselect, information.criterion = c("AICC", "BIC", "AIC"))
```

### Step 4: Computation of benchmark doses

```
bmdcalc(f, z = 1, x = 10)
```

### Step 5: Bootstrap to compute BMD confidence intervals

```
bmdboot(r, niter = 1000, conf.level = 0.95)
```

## Typical script for the workflow

```
o <- RNaseq(datafilename)
s <- itemselect(o)
f <- drcfit(s)
r <- bmdcalc(f)
b <- bmdboot(r)
b$res
```

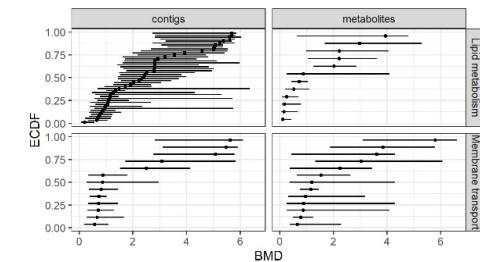
Each function of this workflow returns a S3 class object that can be printed and plotted using `print()` and `plot()` functions.  
Targetted items can be explored whatever they are or not in the selection using `targetplot(items, f)`.

## Other functions to help the interpretation of results within a multi-level approach using a unique biological annotation

Functions taking as a first argument `extendedres`, a data frame with the main workflow results, optionally gathering results obtained at different experimental (different molecular levels, different time points, different pre-exposure histories, ...) extended with additional columns coding for the biological annotation of items and optionally for the experimental level. Some lines of the workflow results can be replicated for items having more than one annotation. See help pages for a complete description of argument of those functions.

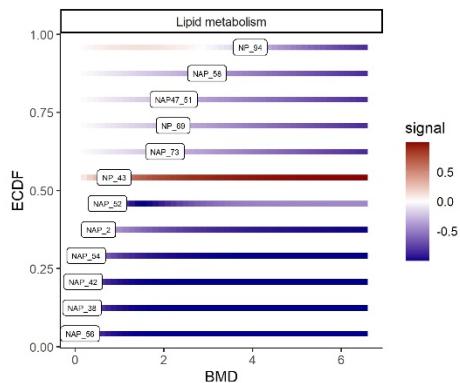
## BMD plot

```
bmdplot(extendedres, add.CI, facetby,
facetby2, shapeby, colorby,
add.label, BMD_log_transfo)
```



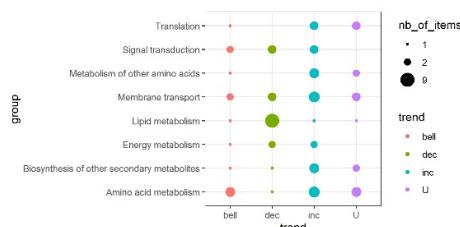
## BMD plot with gradient

```
bmdplotwithgradient(extendedres, xmin,
xmax, scaling, facetby, facetby2,
shapeby, line.size, add.label,
BMD_log_transfo)
```



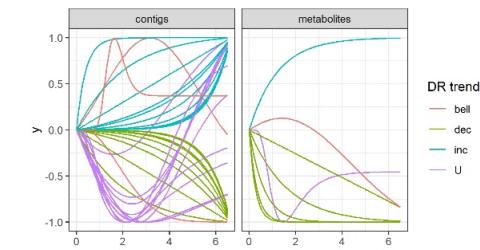
## Trend plot

```
trendplot(extendedres, group, facetby)
```



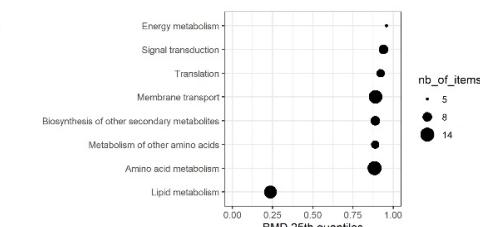
## Dose-response curves plot

```
curvesplot(extendedres, xmin, xmax,
scaling, facetby, facetby2, colorby,
line.size, dose_log_transfo = FALSE)
```



## Sensitivity plot

```
sensitivityplot(extendedres, group,
colorby, BMDsummary =
c("first.quartile", "median",
"median.and.IQR"), BMD_log_transfo)
```



# estimatr :: CHEAT SHEET

## OLS with lm\_robust()

lm\_robust() is lm() with robust SEs. HC2 is the default.

```
lm_robust(mpg ~ hp, data = mtcars)
lm_robust(mpg ~ hp, se_type = "HC1",
          data = mtcars)
lm_robust(mpg ~ hp, se_type = "classical",
          data = mtcars)
```

Indicate clusters to get clustered SEs. CR2 is the default.

```
lm_robust(mpg ~ hp, clusters = carb,
          data = mtcars)
lm_robust(mpg ~ hp, clusters = carb,
          se_type = "stata", data = mtcars)
```

Fixed effects two ways:

```
# FEs as "dummies"
lm_robust(mpg ~ hp + as.factor(am),
          data = mtcars)

# "Absorbing" FEs (substantially faster)
lm_robust(mpg ~ hp,
          fixed_effects = ~ am,
          data = mtcars)
```

post-estimation commands:

```
fit <- lm_robust(mpg ~ hp, data = mtcars)
summary(fit)
print(fit)
tidy(fit)
vcov(fit)
confint(fit)
nobs(fit)
predict(fit, newdata = mtcars)
```

estimatr is part of the DeclareDesign suite of packages for designing, implementing, and analyzing social science research designs.

## 2SLS with iv\_robust()

iv\_robust() is AER:::ivreg() with robust SEs.

```
iv_robust(mpg ~ hp | am, data = mtcars)
iv_robust(mpg ~ hp | am,
          clusters = carb, data = mtcars)
```

## Two-group estimators

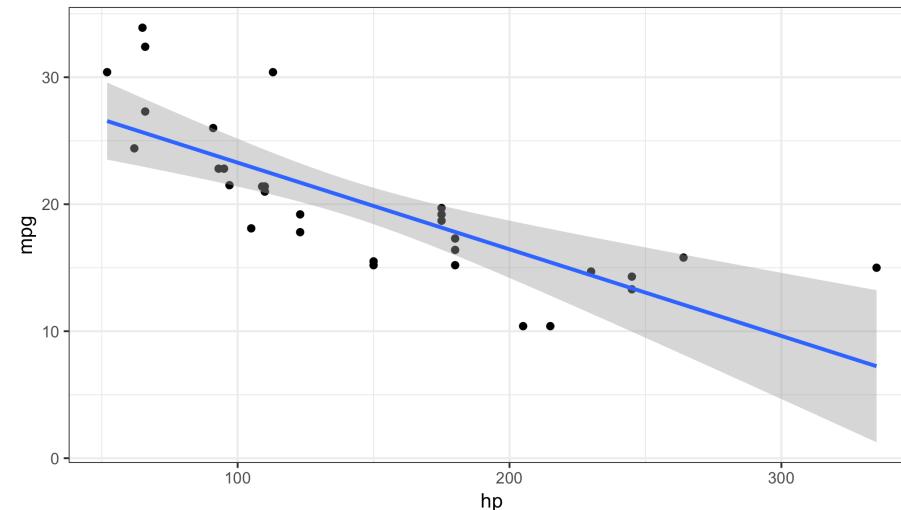
difference\_in\_means() and horvitz\_thompson()  
compare two groups

```
difference_in_means(mpg ~ am, data = mtcars)
horvitz_thompson(mpg ~ am, data = mtcars)
```

## ggplot2 integration

Use robust variance estimates for drawing confidence intervals:

```
library(ggplot2)
ggplot(mtcars, aes(mpg, hp)) +
  geom_point() +
  stat_smooth(method = "lm_robust") +
  theme_bw()
```



## Multiple models

Same outcome, different subsets:

```
library(tidyverse)
mtcars %>%
  split(.cyl) %>%
  map(~lm_robust(mpg ~ hp, data = .)) %>%
  map(tidy) %>%
  bind_rows(.id = "cyl")
```

Different outcomes, same subset:

```
c("mpg", "disp") %>%
  map(~formula(paste0(., " ~ hp"))) %>%
  map(~lm_robust(., data = mtcars)) %>%
  map(tidy) %>%
  bind_rows
```

## Extras

```
# Lin (2013) covariate adjustment
lm_lin(mpg ~ am, covariates = ~ hp,
        data = mtcars)
```

```
# regression tables with texreg
fit <- lm_robust(mpg ~ hp, data = mtcars)
texreg::texreg(fit, include.ci = FALSE)
```

## estimatr-to-Stata dictionary

### estimatr

```
lm_robust(y ~ z,
           data = dat)
```

### Stata

```
reg y z, vce(hc2)
```

```
lm_robust(y ~ z,
           clusters = cl,
           se_type = "stata",
           data = dat)
```

```
reg y z, vce(cluster cl)
```

```
lm_robust(mpg ~ hp,
           fixed_effects = ~ am,
           se_type = "stata",
           data = mtcars)
```

```
areg mpg hp, absorb(am)
vce(robust)
```

```
iv_robust(mpg ~ hp | am,
           se_type = "HC1",
           data = mtcars)
```

```
ivregress 2sls mpg (hp =
am), vce(robust) small
```

# Access Eurostat data with eurostat::cheat sheet

## Search and download

Data in the Eurostat database is stored in tables. Each table has an identifier, a short table\_code, and a description (e.g. tps00199 - Total fertility rate).

Key eurostat functions allow to find the table\_code, download the eurostat table and polish labels in the table.

### Find the table code

The **search\_eurostat(pattern,...)** function scans the directory of Eurostat tables and returns codes and descriptions of tables that match pattern.

```
library("eurostat")
query <- search_eurostat(pattern = "fertility rate",
                         type = "table", fixed = FALSE)
query[,1:2]
## title                           code
## <chr>                            <chr>
## Total fertility rate by NUTS 2 region tgs00100
## Total fertility rate           tps00199
## Total fertility rate by NUTS 2 region tgs00100
```

### Download the table

The **get\_eurostat(id, time\_format = "date", filters = "none", type = "code", cache = TRUE,...)** function downloads the requested table from the Eurostat bulk download facility or from The Eurostat Web Services JSON API (if filters are defined). Downloaded data is cached (if cache=TRUE). Additional arguments define how to read the time column (time\_format) and if table dimensions shall be kept as codes or converted to labels (type).

```
ct <- c("AT", "BE", "BG", "CH", "CY", "CZ", "DE", "DK", "EE", "EL", "ES",
       "FI", "FR", "HR", "HU", "IE", "IS", "IT", "LI", "LT", "LU", "LV",
       "MT", "NL", "NO", "PL", "PT", "RO", "SE", "SI", "SK", "UK")
dat <- get_eurostat(id="tps00199", time_format="num",
                    filters = list(geo = ct))
dat[1:2,]
## indic_de geo   time values
## TOTFERRT AT    2006  1.41
## TOTFERRT AT    2007  1.38
```

### Add labels

The **label\_eurostat(x, lang = "en",...)** gets definitions for Eurostat codes and replace them with labels in given language ("en", "fr" or "de")

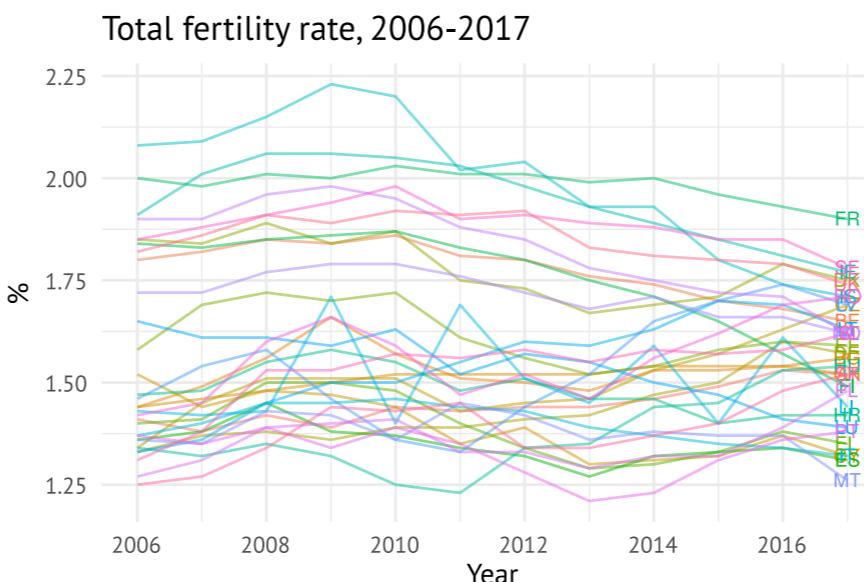
```
dat <- label_eurostat(dat)
dat[1:3,]
## indic_de      geo   time values
## <fct>        <fct> <dbl> <dbl>
## Total fertility rate Andorra 2006  1.24
## Total fertility rate Albania 2006  1.67
## Total fertility rate Armenia 2006  1.34
```



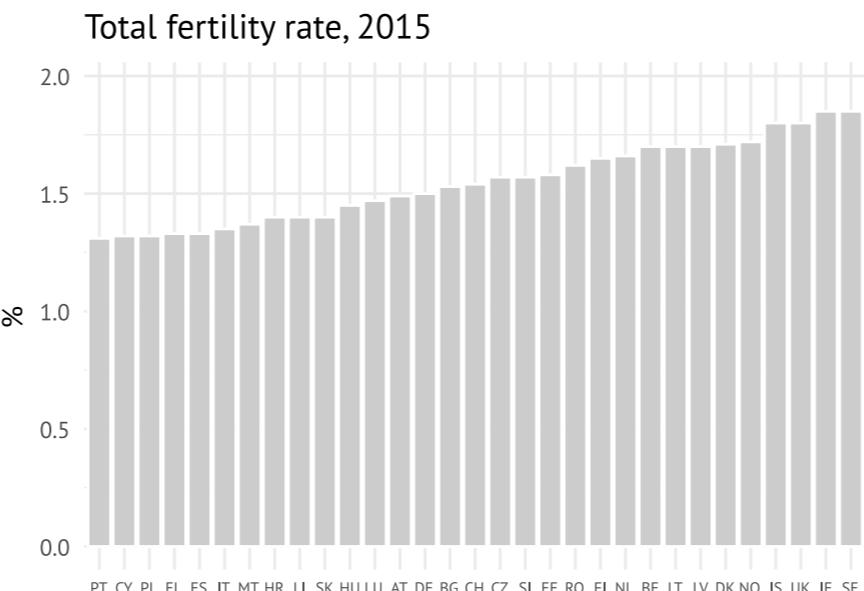
## eurostat and plots

The **get\_eurostat()** function returns tibbles in the long format. Packages dplyr and tidyr are well suited to transform these objects. The **ggplot2**-package is well suited to plot these objects.

```
dat <- get_eurostat(id="tps00199", filters = list(geo = ct))
library(ggplot2)
library(dplyr)
ggplot(dat,
       aes(x = time, y = values, color = geo, label = geo)) +
  geom_line(alpha = .5) +
  geom_text(data = dat %>% group_by(geo) %>%
              filter(time == max(time)),
            size = 2.6) +
  theme(legend.position = "none") +
  labs(title = "Total fertility rate, 2006-2017",
       x = "Year", y = "%")
```



```
dat_2015 <- dat %>%
  filter(time == "2015-01-01")
ggplot(dat_2015, aes(x = reorder(geo, values), y = values)) +
  geom_col(color = "white", fill = "grey80") +
  theme(axis.text.x = element_text(size = 6)) +
  labs(title = "Total fertility rate, 2015",
       y = "%", x = NULL)
```



## eurostat and maps

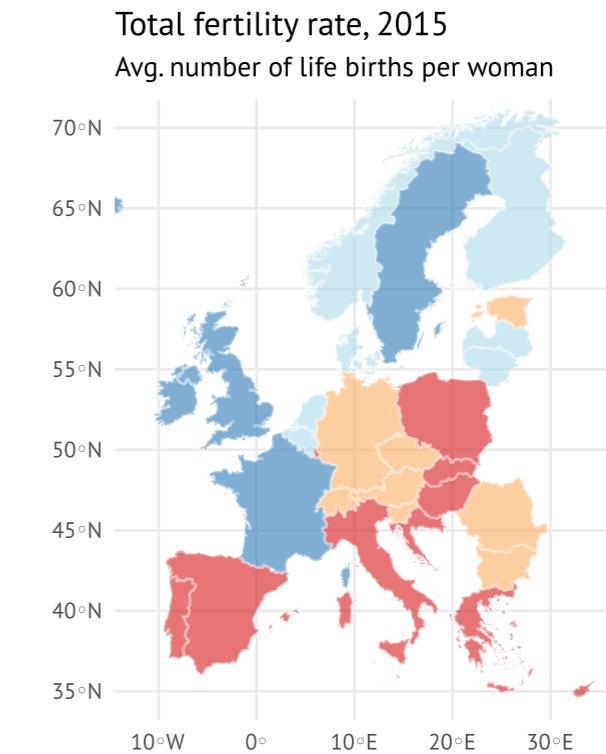
There are two function to work with geospatial data from GISCO. The **get\_eurostat\_geospatial()** returns spatial data as sf-object. Object can me merged with data.frames using **dplyr::\*\_join()**-functions. The **cut\_to\_classes()** is a wrapper for **cut()** - function and is used for categorizing data for maps with tidy labels.

```
mapdata <- get_eurostat_geospatial(nuts_level = 0) %>%
  right_join(dat_2015) %>%
  mutate(cat = cut_to_classes(values, n=4, decimals=1))
head(select(mapdata, geo, values, cat), 3)
## #> #> geo values      cat      geometry
## #> #> AT    1.49 1.5 ~< 1.6 MULTIPOLYGON (((15.54245 48...
```

### Plot a Map

The **sf-object** returned are ready to be plotted with **ggplot::geom\_sf()**-function.

```
ggplot(mapdata, aes(fill = cat)) +
  scale_fill_brewer(palette = "RdYlBu") +
  geom_sf(color = alpha("white", 1/3), alpha = .6) +
  xlim(c(-12, 44)) + ylim(c(35, 70)) +
  labs(title = "Total fertility rate, 2015",
       subtitle = "Avg. number of life births per woman",
       fill = "%")
```



This onepager presents the eurostat package 2014-2019  
 Leo Lahti, Janne Huovari, Markus Kainu, Przemyslaw Biecek  
 package version 3.3.55 URL: <https://github.com/rOpenGov/eurostat>

Retrieval and Analysis of Eurostat Open Data with the eurostat Package.  
 Leo Lahti, Janne Huovari, Markus Kainu, and Przemysław Biecek.  
 The R Journal, 9(1):385–392, 2017.

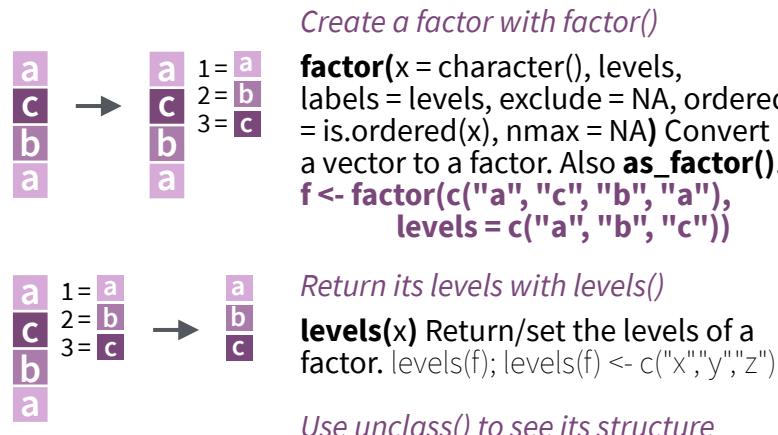


# Factors withforcats :: CHEATSHEET

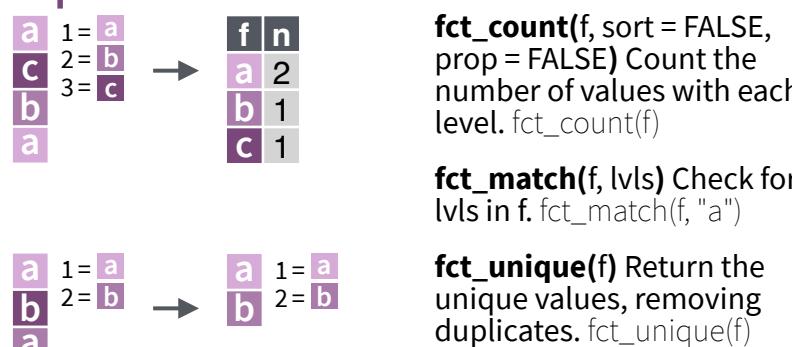
The **forcats** package provides tools for working with factors, which are R's data structure for categorical data.

## Factors

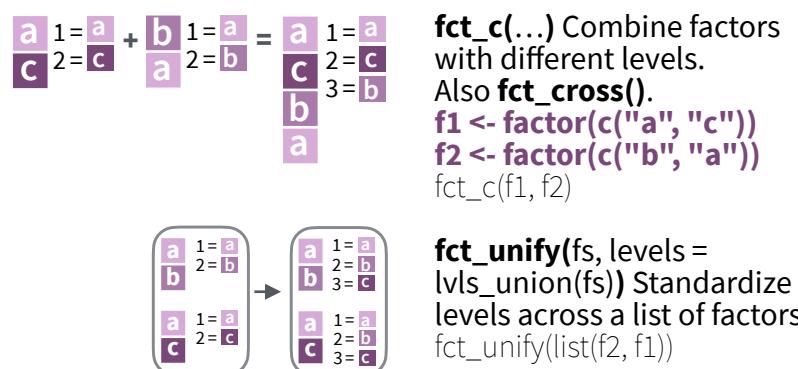
R represents categorical data with factors. A **factor** is an integer vector with a **levels** attribute that stores a set of mappings between integers and categorical values. When you view a factor, R displays not the integers, but the levels associated with them.



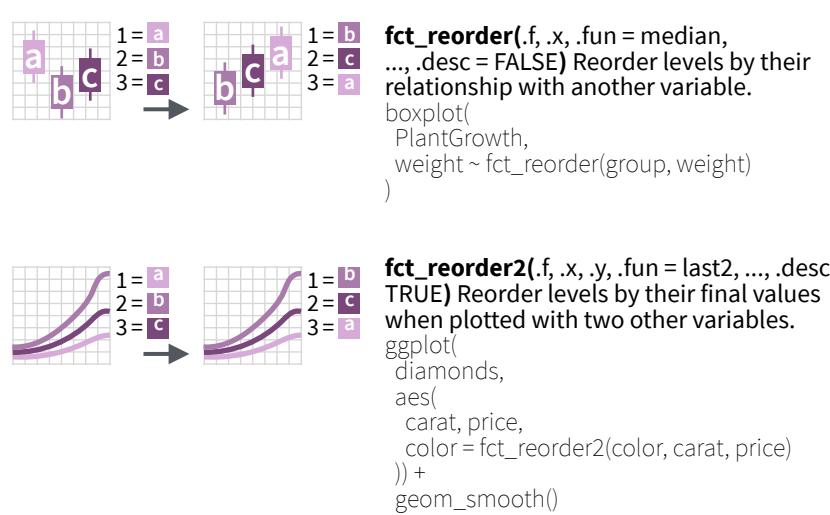
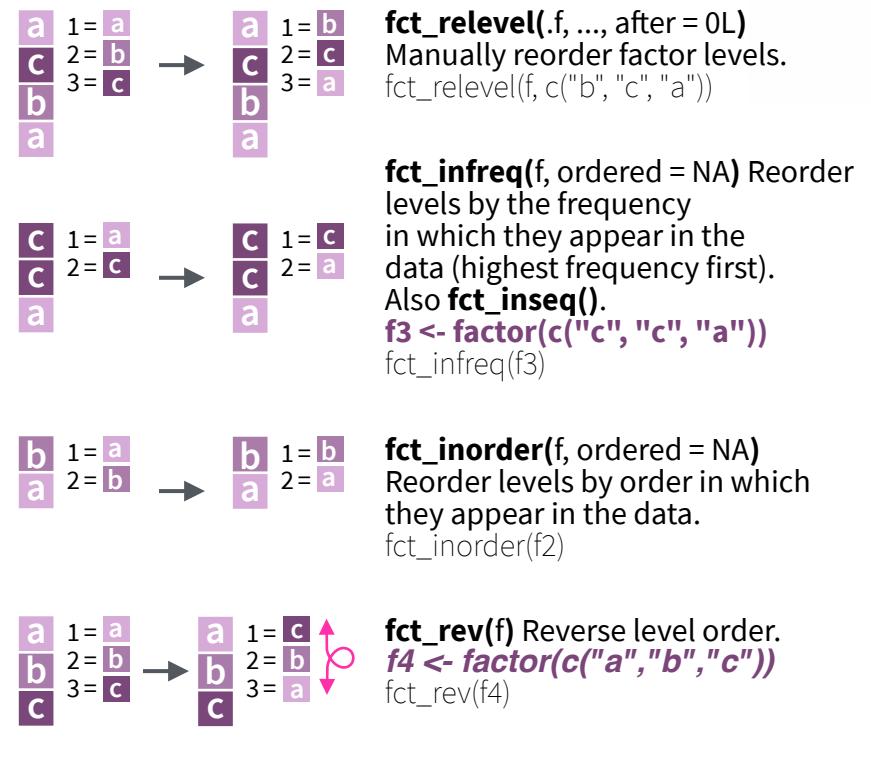
## Inspect Factors



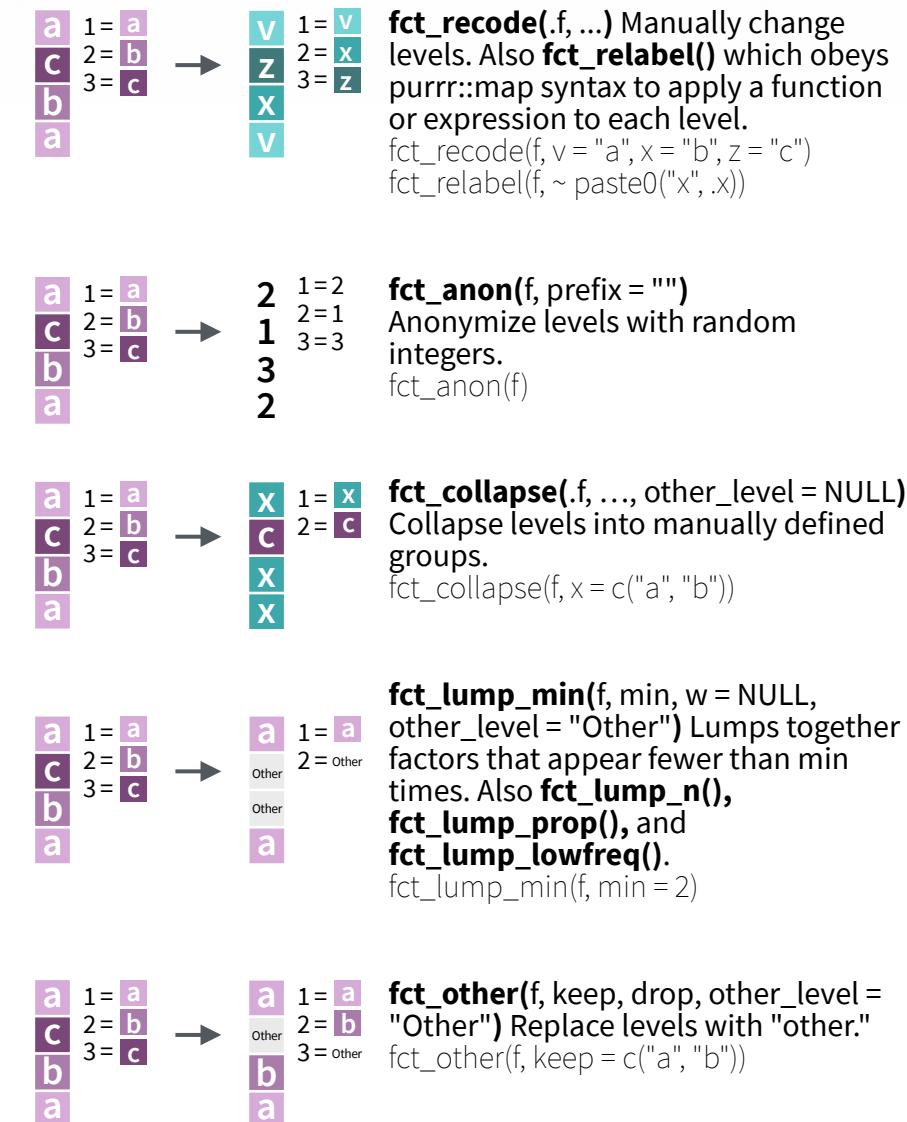
## Combine Factors



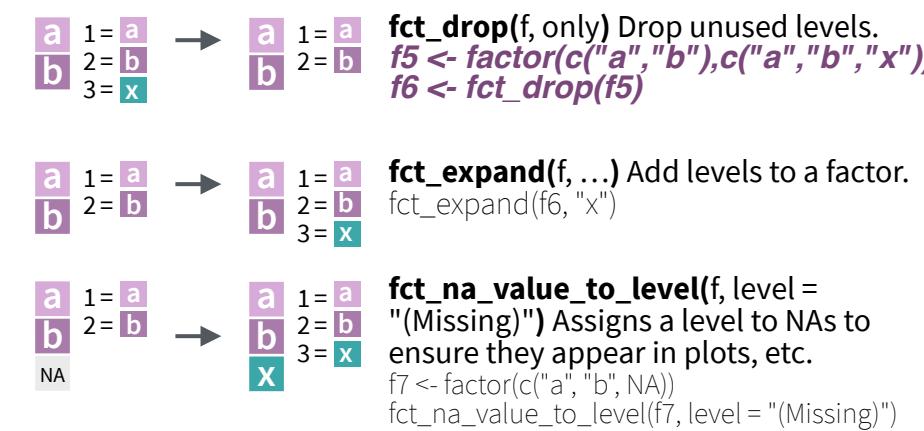
## Change the order of levels



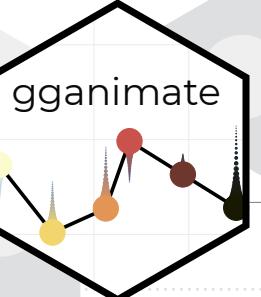
## Change the value of levels



## Add or drop levels



# Animate ggplots with gganimate :: CHEAT SHEET



## Core Concepts

gganimate builds on ggplot2's grammar of graphics to provide functions for animation. You add them to plots created with `ggplot()` the same way you add a geom.

### Main Function Groups

- `transition_*`(): What variable controls change and how?
- `view_*`(): Should the axes change with the data?
- `enter/exit_*`(): How does new data get added the plot? How does old data leave?
- `shadow_*`(): Should previous data be "remembered" and shown with current data?
- `ease_aes()`: How do you want to handle the pace of change between transition values?

**Note:** you only need a `transition_*`() or `view_*`() to make an animation. The other function groups enable you to add features or alter gganimate's default settings .

## Starting Plots

```
library(tidyverse)
library(gganimate)

a <- ggplot(diamonds,
            aes(carat, price)) +
  geom_point()

b <- ggplot(txhousing,
            aes(month, sales)) +
  geom_col()

c <- ggplot(economics,
            aes(date, psavert)) +
  geom_line()
```

## transition\_\*

### transition\_states()

```
a + transition_states(color, transition_length = 3, state_length = 1)
```

We're cycling between values of `color`, ...

... and spending **3** times as long going to the next cut as we do pausing there.

### transition\_time()

```
b + transition_time(year, range = c(2002L, 2006L))
```

We're cycling through each `year` of the data...

...from **2002** to **2006** (range is optional; default is the whole time frame). Unlike `transition_states()`, `transition_time()` treats the data as continuous and so the transition length is based on the actual values. Using **2002L** instead of **2002** because the underlying data is an integer.

### transition\_reveal()

```
c + transition_reveal(date)
```

We're adding each `date` of the data on top of 'old' data

### transition\_filters()

```
a + transition_filter(transition_length = 3,
                      filter_length = 1,
                      cut == "Ideal",
                      Deep = depth >= 60)
```

`transition_length` and `filter_length` work the same as `transition/state_length()` in `transition_states()`...

... but now we're cycling between these two filtering conditions. **Names** are optional, but can be useful (see "Label variables" on next page).

### Other transitions

- `transition_manual()`: Similar to `transition_states()`, but without intermediate states.
- `transition_layers()`: Add layers (geoms) one at time.
- `transition_components()`: Transition elements independently from each other.
- `transition_events()`: Each element's duration can be controlled individually.

## Baseline Animation

```
anim_a <- a + transition_states(color, transition_length = 3, state_length = 1)
```

## view\_\*

### view\_follow()

```
anim_a +
  view_follow(fixed_x = TRUE,
              fixed_y = c(2500, NA))
```

x-axis shows **full range**, y shows **[2500, as much is needed for that frame]**. Default is for both axis to vary as needed.

### view\_step()

```
anim_a +
  view_step(pause_length = 2,
            step_length = 1,
            nstep = 7)
```

We're spending **twice** as long moving between views as staying at them...

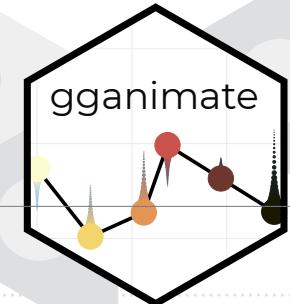
... and we're cycling between **seven** views. Seven is the number of steps in the transition, so the view is changing when the points are static, and visa versa. Views are determined by what data is in the current frame.

### view\_zoom()

`view_zoom()` works similarly to `view_step()`, except it changes the view by zooming and panning.

**Note:** both `view_step()` and `view_zoom()` have `view_*_manual()` versions for setting views directly instead of inferring it from frame data.

# Animate ggplots with gganimate :: CHEAT SHEET



## enter/exit\_\*

Every enter\_\*() function has a corresponding exit\_\*() function, and visa versa.

### enter/exit\_fade()

```
anim_a + enter_fade()
```

When new points need to be added, they will start transparent and become opaque.

### enter\_grow()/exit\_shrink()

```
anim_a + exit_shrink()
```

When extra points need to be removed, they will shrink in size before disappearing.

### enter/exit\_fly()

```
anim_a + enter_fly(x_loc = 0,  
y_loc = 0)
```

When new points need to be added, they will fly in from (0, 0).

### enter/exit\_drift()

```
anim_a + exit_drift(x_mod = 3, y_mod = -2)
```

When extra points need to be removed, They drift 3 units to the right and down 2 units before disappearing.

### enter/exit\_recolour() (or enter/exit\_recolor())

```
anim_a + enter_recolour(color = "red")
```

When new points need to be added, they start as red before transitioning to their correct color.

**Note:** enter/exit\_\*() functions can be combined so that you can have old data fade away and shrink to nothing by adding exit\_fade() and exit\_shrink() to the plot.

## shadow\_\*

### shadow\_wake()

```
anim_a + shadow_wake(wake_length = 0.05)
```

Points have a wake of points with the data from the last 5% of frames.

### shadow\_trail()

```
anim_a + shadow_trail(distance = 0.05)
```

Animation will keep the points from 5% of the frames, spaced as evenly as possible.

### shadow\_mark()

```
anim_a + shadow_mark(color = "red")
```

Animation will keep past states plotted in red (but not the intermediate frames).

## ease\_aes()

ease\_aes() allows you to set an easing function to control the rate of change between transition states. See ?ease\_aes for the full list.

Compare:

```
anim_a
```

```
anim_a + ease_aes("cubic-in") # Change easing of all aesthetics
```

```
anim_a + ease_aes(x = "elastic-in") # Only change `x` (others remain "linear")
```

## Saving animations

```
animation_to_save <- anim_a + exit_shrink()  
anim_save("first_saved_animation.gif", animation = animation_to_save)
```

Since the animation argument uses your last rendered animation by default, this also works:

```
anim_a + exit_shrink()  
anim_save("second_saved_animation.gif")
```

anim\_save() uses gifski to render the animation as a .gif file by default. You can use the renderer argument for other output types including video files (av\_renderer() or ffmpeg\_renderer()) or spritesheets (sprite\_renderer()):

```
# requires you to have the av package installed  
anim_save("third_saved_animation.mp4",  
renderer = av_renderer())
```

## Label variables

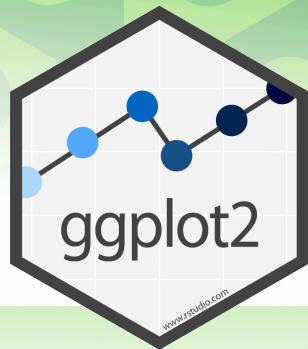
gganimate's transition\_\*() functions create label variables you can pass to (sub)titles and other labels with the glue package. For example, transition\_states() has next\_state, which is the name of the state the animation is transitioning towards. Label variables are different between transitions, and details are included in the documentation of each.

```
anim_a + labs(subtitle = "Moving to {next_state}")
```

We're using the **next\_state** label variable to tell the viewer where we're going.

Label variable	Description	Transitions
transitioning	TRUE if the current frame is an transition frame, FALSE otherwise	states, layers, filter
previous_state/layer	Last shown state/layer	states, layers
next_state/layer	State/layer that will been shown next	states, layers
closest_state/layer	State/layer that current frame is closest to (if between states/layers, either next or closest).	states, layers
previous/closest/_filter/_expression	Similar to their state/layer analogs. *_filter variables return the name of the filter, *_expression variables return the condition.	filter
frame_time	Time of current frame	time, components, events
frame_along	Current frame's value for the dimension we're transitioning over	reveal
nlayers	Number of layers (total, not just currently shown)	layer

# Data visualization with ggplot2 :: CHEAT SHEET



## Basics

ggplot2 is based on the **grammar of graphics**, the idea that you can build every graph from the same components: a **data** set, a **coordinate system**, and **geoms**—visual marks that represent data points.



To display values, map variables in the data to visual properties of the geom (**aesthetics**) like **size**, **color**, and **x** and **y** locations.



Complete the template below to build a graph.

```
ggplot (data = <DATA>) +
  <GEOM_FUNCTION>(mapping = aes(<MAPPINGS>),
  stat = <STAT>, position = <POSITION>) +
  <COORDINATE_FUNCTION> +
  <FACET_FUNCTION> +
  <SCALE_FUNCTION> +
  <THEME_FUNCTION>
```

required  
Not required, sensible defaults supplied

`ggplot(data = mpg, aes(x = cty, y = hwy))` Begins a plot that you finish by adding layers to. Add one geom function per layer.

`last_plot()` Returns the last plot.

`ggsave("plot.png", width = 5, height = 5)` Saves last plot as 5' x 5' file named "plot.png" in working directory. Matches file type to file extension.

## Aes Common aesthetic values.

**color** and **fill** - string ("red", "#RRGGBB")

**linetype** - integer or string (0 = "blank", 1 = "solid", 2 = "dashed", 3 = "dotted", 4 = "dotdash", 5 = "longdash", 6 = "twodash")

**lineend** - string ("round", "butt", or "square")

**linejoin** - string ("round", "mitre", or "bevel")

**size** - integer (line width in mm)

**shape** - integer/shape name or a single character ("a")  


## Geoms

Use a geom function to represent data points, use the geom's aesthetic properties to represent variables.  
Each function returns a layer.

### GRAPHICAL PRIMITIVES

```
a <- ggplot(economics, aes(date, unemploy))
b <- ggplot(seals, aes(x = long, y = lat))
```

- a + geom\_blank()** and **a + expand\_limits()**  
Ensure limits include values across all plots.
- b + geom\_curve(aes(yend = lat + 1, xend = long + 1, curvature = 1))** - x, yend, alpha, angle, curvature, linetype, size
- a + geom\_path(lineend = "butt", linejoin = "round", linemitre = 1)** - x, y, alpha, color, group, linetype, size
- a + geom\_polygon(aes(alpha = 50))** - x, y, alpha, color, fill, group, subgroup, linetype, size
- b + geom\_rect(aes(xmin = long, ymin = lat, xmax = long + 1, ymax = lat + 1))** - xmax, xmin, ymax, ymin, alpha, color, fill, linetype, size
- a + geom\_ribbon(aes(ymin = unemploy - 900, ymax = unemploy + 900))** - x, ymax, ymin, alpha, color, fill, group, linetype, size

### LINE SEGMENTS

common aesthetics: x, y, alpha, color, linetype, size

- b + geom\_abline(aes(intercept = 0, slope = 1))**
- b + geom\_hline(aes(yintercept = lat))**
- b + geom\_vline(aes(xintercept = long))**
- b + geom\_segment(aes(yend = lat + 1, xend = long + 1))**
- b + geom\_spoke(aes(angle = 1:1155, radius = 1))**

### ONE VARIABLE continuous

- ```
c <- ggplot(mpg, aes(hwy)); c2 <- ggplot(mpg)
```
- c + geom\_area(stat = "bin")** - x, y, alpha, color, fill, linetype, size
  - c + geom\_density(kernel = "gaussian")** - x, y, alpha, color, fill, group, linetype, size, weight
  - c + geom\_dotplot()** - x, y, alpha, color, fill
  - c + geom\_freqpoly()** - x, y, alpha, color, group, linetype, size
  - c + geom\_histogram(binwidth = 5)** - x, y, alpha, color, fill, linetype, size, weight
  - c2 + geom\_qq(aes(sample = hwy))** - x, y, alpha, color, fill, linetype, size, weight

### discrete

```
d <- ggplot(mpg, aes(f1))
```

- d + geom\_bar()** - x, alpha, color, fill, linetype, size, weight

### TWO VARIABLES both continuous

```
e <- ggplot(mpg, aes(cty, hwy))
```

- e + geom\_label(aes(label = cty), nudge\_x = 1, nudge\_y = 1)** - x, y, label, alpha, angle, color, family, fontface, hjust, lineheight, size, vjust
- e + geom\_point()** - x, y, alpha, color, fill, shape, size, stroke
- e + geom\_quantile()** - x, y, alpha, color, group, linetype, size, weight
- e + geom\_rug(sides = "bl")** - x, y, alpha, color, linetype, size
- e + geom\_smooth(method = lm)** - x, y, alpha, color, fill, group, linetype, size, weight
- e + geom\_text(aes(label = cty), nudge\_x = 1, nudge\_y = 1)** - x, y, label, alpha, angle, color, family, fontface, hjust, lineheight, size, vjust

### one discrete, one continuous

```
f <- ggplot(mpg, aes(class, hwy))
```

- f + geom\_col()** - x, y, alpha, color, fill, group, linetype, size
- f + geom\_boxplot()** - x, y, lower, middle, upper, ymax, ymin, alpha, color, fill, group, linetype, shape, size, weight
- f + geom\_dotplot(binaxis = "y", stackdir = "center")** - x, y, alpha, color, fill, group
- f + geom\_violin(scale = "area")** - x, y, alpha, color, fill, group, linetype, size, weight

### both discrete

```
g <- ggplot(diamonds, aes(cut, color))
```

- g + geom\_count()** - x, y, alpha, color, fill, shape, size, stroke
- e + geom\_jitter(height = 2, width = 2)** - x, y, alpha, color, fill, shape, size

### THREE VARIABLES

```
seals$z <- with(seals, sqrt(delta_long^2 + delta_lat^2)); l <- ggplot(seals, aes(long, lat))
```

- l + geom\_contour(aes(z = z))** - x, y, z, alpha, color, group, linetype, size, weight
- l + geom\_contour\_filled(aes(fill = z))** - x, y, alpha, color, fill, group, linetype, size, subgroup
- l + geom\_raster(aes(fill = z), hjust = 0.5, vjust = 0.5, interpolate = FALSE)** - x, y, alpha, fill
- l + geom\_tile(aes(fill = z))** - x, y, alpha, color, fill, linetype, size, width

### continuous bivariate distribution

```
h <- ggplot(diamonds, aes(carat, price))
```

- h + geom\_bin2d(binwidth = c(0.25, 500))** - x, y, alpha, color, fill, linetype, size, weight
- h + geom\_density\_2d()** - x, y, alpha, color, group, linetype, size
- h + geom\_hex()** - x, y, alpha, color, fill, size

### continuous function

```
i <- ggplot(economics, aes(date, unemploy))
```

- i + geom\_area()** - x, y, alpha, color, fill, linetype, size
- i + geom\_line()** - x, y, alpha, color, group, linetype, size
- i + geom\_step(direction = "hv")** - x, y, alpha, color, group, linetype, size

### visualizing error

```
df <- data.frame(grp = c("A", "B"), fit = 4:5, se = 1:2)
j <- ggplot(df, aes(grp, fit, ymin = fit - se, ymax = fit + se))
```

- j + geom\_crossbar(fatten = 2)** - x, y, ymax, ymin, alpha, color, fill, group, linetype, size
- j + geom\_errorbar()** - x, ymax, ymin, alpha, color, group, linetype, size, width  
Also **geom\_errorbarh()**.
- j + geom\_linerange()** - x, ymin, ymax, alpha, color, group, linetype, size
- j + geom\_pointrange()** - x, y, ymin, ymax, alpha, color, fill, group, linetype, shape, size

### maps

```
data <- data.frame(murder = USArrests$Murder, state = tolower(rownames(USArrests)))
```

```
map <- map_data("state")
```

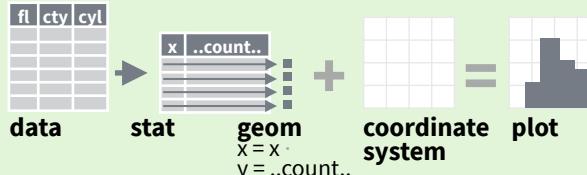
```
k <- ggplot(data, aes(fill = murder))
```

- k + geom\_map(aes(map\_id = state), map = map) + expand\_limits(x = map\$long, y = map\$lat)**  
map\_id, alpha, color, fill, linetype, size

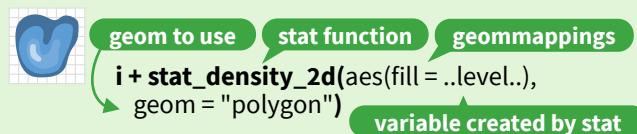
# Stats

An alternative way to build a layer.

A stat builds new variables to plot (e.g., count, prop).



Visualize a stat by changing the default stat of a geom function, `geom_bar(stat="count")` or by using a stat function, `stat_count(geom="bar")`, which calls a default geom to make a layer (equivalent to a geom function). Use `..name..` syntax to map stat variables to aesthetics.



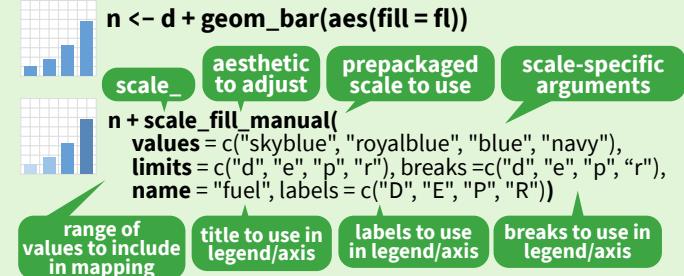
```

c + stat_bin(binwidth = 1, boundary = 10)
x, y | ..count.., ..ncount.., ..density.., ..ndensity..
c + stat_count(width = 1) x, y | ..count.., ..prop..
c + stat_density(adjust = 1, kernel = "gaussian")
x, y | ..count.., ..density.., ..scaled..
e + stat_bin_2d(bins = 30, drop = T)
x, y, fill | ..count.., ..density..
e + stat_bin_hex(bins = 30) x, y, fill | ..count.., ..density..
e + stat_density_2d(contour = TRUE, n = 100)
x, y, color, size | ..level..
e + stat_ellipse(level = 0.95, segments = 51, type = "t")
l + stat_contour(aes(z = z)) x, y, z, order | ..level..
l + stat_summary_hex(aes(z = z), bins = 30, fun = max)
x, y, z, fill | ..value..
l + stat_summary_2d(aes(z = z), bins = 30, fun = mean)
x, y, z, fill | ..value..
f + stat_boxplot(coef = 1.5)
x, y | ..lower.., ..middle.., ..upper.., ..width.., ..ymin.., ..ymax..
f + stat_ydensity(kernel = "gaussian", scale = "area") x, y
| ..density.., ..scaled.., ..count.., ..n.., ..violinwidth.., ..width..
e + stat_ecdf(n = 40) x, y | ..x.., ..y..
e + stat_quantile(quantiles = c(0.1, 0.9),
formula = y ~ log(x), method = "rq") x, y | ..quantile..
e + stat_smooth(method = "lm", formula = y ~ x, se = T,
level = 0.95) x, y | ..se.., ..x.., ..y.., ..ymin.., ..ymax..
ggplot() + xlim(-5, 5) + stat_function(fun = dnorm,
n = 20, geom = "point") x | ..x.., ..y..
ggplot() + stat_qq(aes(sample = 1:100))
x, y, sample | ..sample.., ..theoretical..
e + stat_sum() x, y, size | ..n.., ..prop..
e + stat_summary(fun.data = "mean_cl_boot")
h + stat_summary_bin(fun = "mean", geom = "bar")
e + stat_identity()
e + stat_unique()
  
```

# Scales

Override defaults with `scales` package.

**Scales** map data values to the visual values of an aesthetic. To change a mapping, add a new scale.



## GENERAL PURPOSE SCALES

Use with most aesthetics

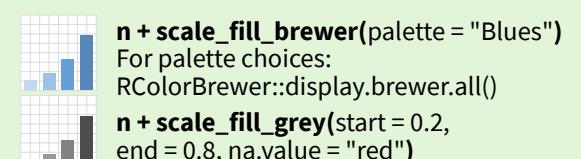
`scale_*_continuous()` - Map cont' values to visual ones.  
`scale_*_discrete()` - Map discrete values to visual ones.  
`scale_*_binned()` - Map continuous values to discrete bins.  
`scale_*_identity()` - Use data values as visual ones.  
`scale_*_manual(values = c())` - Map discrete values to manually chosen visual ones.  
`scale_*_date(date_labels = "%m/%d")`,  
`date_breaks = "2 weeks"` - Treat data values as dates.  
`scale_*_datetime()` - Treat data values as date times.  
 Same as `scale_*_date()`. See `?strptime` for label formats.

## X & Y LOCATION SCALES

Use with x or y aesthetics (x shown here)

`scale_x_log10()` - Plot x on log10 scale.  
`scale_x_reverse()` - Reverse the direction of the x axis.  
`scale_x_sqrt()` - Plot x on square root scale.

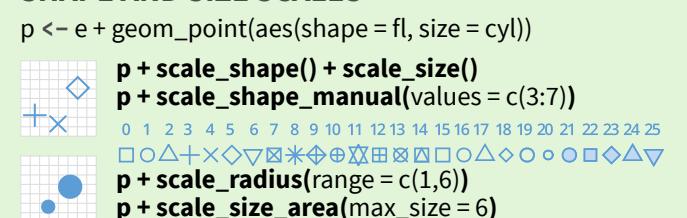
## COLOR AND FILL SCALES (DISCRETE)



## COLOR AND FILL SCALES (CONTINUOUS)



## SHAPE AND SIZE SCALES



# Coordinate Systems

`r <- d + geom_bar()`

`r + coord_cartesian(xlim = c(0, 5))` - xlim, ylim  
The default cartesian coordinate system.

`r + coord_fixed(ratio = 1/2)`  
ratio, xlim, ylim - Cartesian coordinates with fixed aspect ratio between x and y units.

`ggplot(mpg, aes(y = fl)) + geom_bar()`  
Flip cartesian coordinates by switching x and y aesthetic mappings.

`r + coord_polar(theta = "x", direction=1)`  
theta, start, direction - Polar coordinates.

`r + coord_trans(y = "sqrt")` - x, y, xlim, ylim  
Transformed cartesian coordinates. Set xtrans and ytrans to the name of a window function.

`pi + coord_quickmap()`  
`pi + coord_map(projection = "ortho", orientation = c(41, -74, 0))` - projection, xlim, ylim  
Map projections from the mapproj package (mercator (default), azequalarea, lagrange, etc.).

## Position Adjustments

Position adjustments determine how to arrange geoms that would otherwise occupy the same space.

`s <- ggplot(mpg, aes(fl, fill = drv))`

`s + geom_bar(position = "dodge")`  
Arrange elements side by side.

`s + geom_bar(position = "fill")`  
Stack elements on top of one another, normalize height.

`e + geom_point(position = "jitter")`  
Add random noise to X and Y position of each element to avoid overplotting.

`e + geom_label(position = "nudge")`  
Nudge labels away from points.

`s + geom_bar(position = "stack")`  
Stack elements on top of one another.

Each position adjustment can be recast as a function with manual `width` and `height` arguments:  
`s + geom_bar(position = position_dodge(width = 1))`

## Themes

`r + theme_bw()`  
White background with grid lines.

`r + theme_gray()`  
Grey background (default theme).

`r + theme_dark()`  
Dark for contrast.

`r + theme_classic()`

`r + theme_light()`

`r + theme_linedraw()`

`r + theme_minimal()`  
Minimal theme.

`r + theme_void()`  
Empty theme.

`r + theme()` Customize aspects of the theme such as axis, legend, panel, and facet properties.

`r + ggtitle("Title") + theme(plot.title.position = "plot")`  
`r + theme(panel.background = element_rect(fill = "blue"))`

# Faceting

Facets divide a plot into subplots based on the values of one or more discrete variables.

`t <- ggplot(mpg, aes(cty, hwy)) + geom_point()`

`t + facet_grid(cols = vars(fl))`  
Facet into columns based on fl.

`t + facet_grid(rows = vars(year))`  
Facet into rows based on year.

`t + facet_grid(rows = vars(year), cols = vars(fl))`  
Facet into both rows and columns.

`t + facet_wrap(vars(fl))`  
Wrap facets into a rectangular layout.

Set `scales` to let axis limits vary across facets.

`t + facet_grid(rows = vars(drv), cols = vars(fl), scales = "free")`

x and y axis limits adjust to individual facets:  
`"free_x"` - x axis limits adjust  
`"free_y"` - y axis limits adjust

Set `labeler` to adjust facet label:

`t + facet_grid(cols = vars(fl), labeler = label_both)`

`fl: c fl: d fl: e fl: p fl: r`

`t + facet_grid(rows = vars(fl), labeler = label_bquote(alpha ^ .(fl)))`

`alpha^c alpha^d alpha^e alpha^p alpha^r`

# Labels and Legends

Use `labs()` to label the elements of your plot.

`t + labs(x = "New x axis label", y = "New y axis label", title = "Add a title above the plot", subtitle = "Add a subtitle below title", caption = "Add a caption below plot", alt = "Add alt text to the plot", <AES> = "New <AES> legend title")`

`t + annotate(geom = "text", x = 8, y = 9, label = "A")`  
Places a geom with manually selected aesthetics.

`p + guides(x = guide_axis(n.dodge = 2))` Avoid crowded or overlapping labels with `guide_axis(n.dodge` or `angle`).

`n + guides(fill = "none")` Set legend type for each aesthetic: colorbar, legend, or none (no legend).

`n + theme(legend.position = "bottom")`  
Place legend at "bottom", "top", "left", or "right".

`n + scale_fill_discrete(name = "Title", labels = c("A", "B", "C", "D", "E"))`  
Set legend title and labels with a scale function.

# Zooming

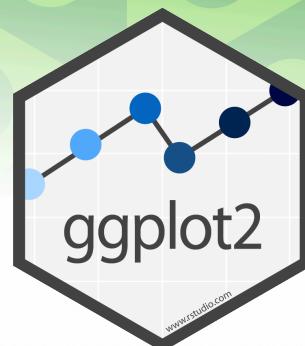
`Without clipping` (preferred):

`t + coord_cartesian(xlim = c(0, 100), ylim = c(10, 20))`

`With clipping` (removes unseen data points):

`t + xlim(0, 100) + ylim(10, 20)`

`t + scale_x_continuous(limits = c(0, 100)) + scale_y_continuous(limits = c(0, 100))`







# Using Git and GitHub with RStudio: : CHEATSHEET



**Version control** control, also known as **source control**, is the practice of tracking and managing changes to software code.

Version control systems are software tools that help software teams manage changes to source code over time.

Git is an **open-source** software for version control, originally developed in 2005 by Linus Torvalds, the creator of the Linux operating system kernel.

**Git** is a version control tool to track the changes in the source code of a project.

**GitHub** is the most popular hosting service for collaborating on code using Git.

## Requirements

1. R and RStudio installed
2. Git installed
3. Register a free GitHub account



## Check that Git is installed

In the Terminal of RStudio, enter `which git` to request the path to your Git executable:

```
which git
## /usr/bin/git
```

and `git --version` to see its version:

```
git --version
## git version 2.34.1
```

## Introduce yourself to Git

Open a shell from RStudio *Tools > Shell* and type each line separately by substituting your name and the email associated with your GitHub account:

```
git config --global user.name 'Jane Doe'
git config --global user.email
'jane@example.com'
```

## Github Glossary

This [glossary](#) introduces common Git and GitHub terminology.

## Basics

|                                                    |                                                                                                                               |
|----------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------|
| <code>git init &lt;directory&gt;</code>            | Create empty Git repository in specified directory.                                                                           |
| <code>git clone &lt;repository&gt;</code>          | Clone a repository located at <code>&lt;repository&gt;</code> on your local machine.                                          |
| <code>git config user.name &lt;username&gt;</code> | Define author name to be used for all commits in current repository.                                                          |
| <code>git add &lt;directory&gt;</code>             | Stage all changes in <code>&lt;directory&gt;</code> for the next commit.                                                      |
| <code>git commit -m &lt;"message"&gt;</code>       | Commit the staged snapshot, but instead of launching a text editor, use <code>&lt;"message"&gt;</code> as the commit message. |
| <code>git status</code>                            | List which files are staged, unstaged, and untracked.                                                                         |
| <code>git log</code>                               | Display the entire commit history using the default format.                                                                   |
| <code>git diff</code>                              | Show unstaged changes between your index and working directory.                                                               |

## Remote Repositories

|                                                      |                                                                                                                                                                            |
|------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <code>git remote add &lt;name&gt; &lt;url&gt;</code> | Create a new connection to a remote repository. After adding a remote, you can use <code>&lt;name&gt;</code> as a shortcut for <code>&lt;url&gt;</code> in other commands. |
| <code>git fetch &lt;remote&gt; &lt;branch&gt;</code> | Fetches a specific <code>&lt;branch&gt;</code> , from the repository. Leave off <code>&lt;branch&gt;</code> to fetch all remote refs.                                      |
| <code>git pull &lt;remote&gt;</code>                 | Fetch the specified remote's copy of current branch and <b>immediately</b> merge it into the local copy.                                                                   |
| <code>git push &lt;remote&gt; &lt;branch&gt;</code>  | Push the branch to <code>&lt;remote&gt;</code> , along with necessary commits and objects. Creates named branch in the remote repository if it doesn't exist.              |

## Undoing Changes

|                                        |                                                                                                                                                         |
|----------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------|
| <code>git revert &lt;commit&gt;</code> | Create new commit that undoes all of the changes made in <code>&lt;commit&gt;</code> , then apply it to the current branch.                             |
| <code>git reset &lt;file&gt;</code>    | Remove <code>&lt;file&gt;</code> from the staging area but leave the working directory unchanged. This unstages a file without overwriting any changes. |
| <code>git clean -n</code>              | Shows which files would be removed from working directory. Use the <code>-f</code> flag in place of the <code>-n</code> flag to execute the clean.      |

## Rewriting Git History

|                                      |                                                                                                                                                               |
|--------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <code>git commit --amend</code>      | Replace the last commit with the staged changes and last commit combined. Use with nothing staged to edit the last commit's message.                          |
| <code>git rebase &lt;base&gt;</code> | Rebase the current branch onto <code>&lt;base&gt;</code> . <code>&lt;base&gt;</code> can be a commit ID, branch name, a tag, or a relative reference to HEAD. |
| <code>git reflog</code>              | Show a log of changes to the local repository's HEAD. Add <code>--relative-date</code> flag to show date info or <code>--all</code> to show all refs.         |

## Git Branches

|                                             |                                                                                                                                                      |
|---------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------|
| <code>git branch</code>                     | List all of the branches in your repo. Add a <code>&lt;branch&gt;</code> argument to create a new branch with the name <code>&lt;branch&gt;</code> . |
| <code>git checkout -b &lt;branch&gt;</code> | Create and check out a new named <code>&lt;branch&gt;</code> . Drop the <code>-b</code> flag to checkout an existing branch.                         |
| <code>git merge &lt;branch&gt;</code>       | Merge <code>&lt;branch&gt;</code> into the current branch.                                                                                           |

# golem :: A Framework for Building Robust Shiny Apps

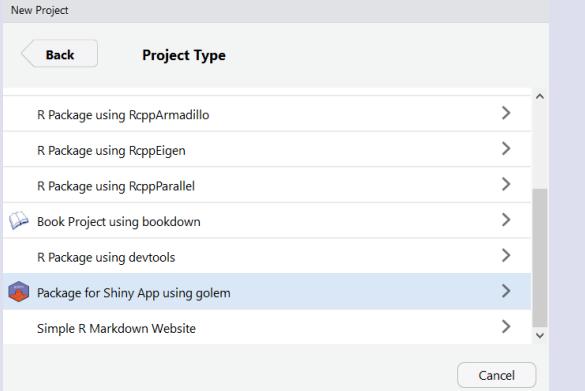
Create, maintain & deploy a packaged Shiny Application



## 1. Create a golem

With RStudio:

File ➔ New Project ➔ New Directory ➔ Shiny App using golem



Using the command line:

```
golem::create_golem(path = "~/appdemo")  
Creates a golem at '~/appdemo'.
```

## 2. Set up your golem with dev/01\_start.R

```
golem::fill_desc(pkg_name = "appdemo", ...)
```

Fills the package DESCRIPTION with the author information, the application title & description, links...

```
golem::set_golem_options()  
Sets {golem} global options.
```

```
golem::use_recommended_tests()  
Creates a test template for your app.
```

```
golem::use_recommended_deps()  
Adds {shiny}, {DT}, {attempt}, {glue}, {htmltools}, and  
{golem} as dependencies.
```

```
golem::use_favicon(path = "path/to/favicon.ico")  
Changes the default favicon.
```

```
golem::use_utils_ui()  
Creates 'R/golem_utils_ui.R', with UI-related helper functions.
```

```
golem::use_utils_server()  
Creates 'R/golem_utils_server.R', with server-related helper functions.
```

## 3. Day-to-day dev with golem

### A. Look at your golem

- Launch your app with `dev/run_dev.R`:

```
options(golem.app.prod = FALSE)  
Sets the prod or dev mode. (see ?golem::app_dev)  
golem::detach_all_attached()  
Detaches all loaded packages and cleans your environment.  
golem::document_and_reload()  
Documents and reloads your package.  
appdemo::run_app()  
Launches your application.
```

### B. Customise your golem with dev/02\_dev.R

- Edit R/app\_ui.R & R/app\_server.R

'R/app\_ui.R' & 'R/app\_server.R' hold the UI and server logic of your app. You can edit them directly, or add elements created with golem (e.g. modules).

- Add shiny modules

```
golem::add_module(name = "example")  
Creates 'R/mod_example.R', with mod_example_ui and  
mod_example_server functions inside.
```

- Add external files

```
golem::add_js_file("script")  
Creates 'inst/app/www/script.js'.
```

```
golem::add_js_handler("script")  
Creates 'inst/app/www/script.js' with a skeleton for shiny  
custom handlers.
```

```
golem::add_css_file("custom")  
Creates 'inst/app/www/custom.css'.
```

- Use golem built-in JavaScript functions

```
golem::activate_js()  
Activates the built-in JavaScript functions. To be inserted in the UI.
```

```
golem::invoke_js("jsfunction", ns("ref_ui"))  
Invokes from the server any JS function: built-in golem JS  
functions or custom ones created with add_js_handler()
```

## 4. Exhibit your golem

### Locally

```
remotes::install_local()
```

Installs your golem locally like any other package.

### To Rstudio products

```
golem::add_rstudioconnect_file()
```

Creates an app.R file, ready to be deployed to RStudio Connect.

```
golem::add_shinyappsio_file()
```

Creates an app.R file, ready to be deployed to shinyapps.io.

```
golem::add_shinyserver_file()
```

Creates an app.R file, ready to be deployed to Shiny Server.

### With Docker

```
golem::add_dockerfile()
```

Creates a Dockerfile that can launch your app.

```
golem::add_dockerfile_shinyproxy()
```

Creates a Dockerfile for ShinyProxy.

```
golem::add_dockerfile_heroku()
```

Creates a Dockerfile for Heroku.

## Tips and tricks

```
golem::print_dev("text")
```

Prints text in your console if `golem::app_dev()` is TRUE.

```
golem::make_dev(function)
```

Makes `function` depend on `golem::app_dev()` being TRUE.

```
golem::browser_button()
```

Creates a backdoor to your app  
(see ?golem::browser\_button).

- How to make a `run_dev` script for a specific module:

```
golem::detach_all_attached()  
golem::document_and_reload()
```

```
ui <- mod_example_ui("my_module")  
server <- function(input, output, session){  
  callModule(mod_example_server, "my_module", session)  
}  
shinyApp(ui, server)
```

Keep in mind that a golem is a package. Everything you know about package development works with your packaged Shiny App created with {golem}!  
(documentation, tests, CI & CD, ...)

# gt summary

Publication-ready analytical  
and summary tables with R  
Cheat Sheet

## Core Table Functions

### tbl\_summary()

Calculates descriptive stats for continuous, categorical, and dichotomous variables.

### tbl\_regression()

Turns a regression model object into a customized, formatted table.

### tbl\_survfit()

Turns a survfit object into a customized table with time-to-event estimates.



## tbl\_summary() using tidyverse syntax to summarize specific columns of a dataset with flexible customization options (See vignette!)

### Basic code

```
trial %>% select(trt, age, grade, response) %>% tbl_summary()
```

### Basic table

| Characteristic         | N = 200 <sup>1</sup> |
|------------------------|----------------------|
| Chemotherapy Treatment |                      |
| Drug A                 | 98 (49%)             |
| Drug B                 | 102 (51%)            |
| Age, yrs               | 47 (38, 57)          |
| Unknown                | 11                   |
| Grade                  |                      |
| I                      | 68 (34%)             |
| II                     | 68 (34%)             |
| III                    | 64 (32%)             |
| Tumor Response         | 61 (32%)             |
| Unknown                | 7                    |

<sup>1</sup>n (%); Median (IQR)

### tbl\_svysummary() for survey objects

Same functionality as `tbl_summary()`, but takes a survey object as input, and accounts for survey weights and design.

More info at:  
[http://www.danielsjoberg.com/gtsummary/reference/tbl\\_svysummary.html](http://www.danielsjoberg.com/gtsummary/reference/tbl_svysummary.html)

For more info on customization arguments and options, visit [http://www.danielsjoberg.com/gtsummary/reference/tbl\\_summary.html](http://www.danielsjoberg.com/gtsummary/reference/tbl_summary.html)

### Argument Input

### Customization options

#### Effect on table

|              |                                                                            |                                                                                                 |
|--------------|----------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------|
| by           | Column to crosstabulate by                                                 | Summary statistics will be calculated separately for each level of the variable                 |
| label        | Formula list of variable labels                                            | Changes text of variable name in table                                                          |
| statistic    | Formula list of summary statistic type for each variable                   | Changes summary statistics displayed for specified variables in table                           |
| digits       | Formula list of number of decimal places to display                        | Changes number of rounded decimal places in table for specified continuous variables            |
| type         | Formula list specifying variable types                                     | Changes variable type for specified variables, affecting which summary statistics are displayed |
| value        | Formula list of value to display for dichotomous variables                 | Changes the value displayed for dichotomous type variables                                      |
| missing      | "no", "ifany", "always"                                                    | Changes whether missing observations are reported                                               |
| missing_text | String to display for count of missing observations                        | Changes the name of the missing data level for appropriate variables                            |
| sort         | Formula list of type of sorting to perform ("frequency" or "alphanumeric") | Changes the type of sorting for categorical variables                                           |
| percent      | "column", "row", or "cell"                                                 | Changes how percentage statistics are calculated and displayed                                  |

### Customized code

```
trial %>%  
  select(trt, age, grade, response) %>%  
  tbl_summary(  
    by = trt,  
    label = list(age ~ "Age (years)",  
                grade ~ "Tumor grade"),  
    percent = "row"  
    digits = list(age ~ 2),  
    statistic = list(age ~ "{mean} ({sd})",  
                    response ~ "{n}/{N} ({p}%)"),  
    type = list(response ~ "categorical"),  
    missing = "always",  
    missing_text = "Missing",  
    )
```

### Customized table

| Characteristic | Drug A, N = 98 <sup>1</sup> | Drug B, N = 102 <sup>1</sup> |
|----------------|-----------------------------|------------------------------|
| Age (years)    | 47.01 (14.71)               | 47.45 (14.01)                |
| Missing        | 7                           | 4                            |
| Tumor grade    |                             |                              |
| I              | 35 (51%)                    | 33 (49%)                     |
| II             | 32 (47%)                    | 36 (53%)                     |
| III            | 31 (48%)                    | 33 (52%)                     |
| Missing        | 0                           | 0                            |
| Tumor Response |                             |                              |
| 0              | 67/132 (51%)                | 65/132 (49%)                 |
| 1              | 28/61 (46%)                 | 33/61 (54%)                  |
| Missing        | 3                           | 4                            |

<sup>1</sup>Mean (SD); n (%) ; n/N (%)

### Extended code

```
trial %>%  
  select(trt, age,  
         response) %>%  
  tbl_summary(  
    by = trt,  
    missing= "no"  
    ) %>%  
  add_n() %>%  
  add_overall() %>%  
  add_p()
```

### Extended table

| Characteristic | N   | Overall, N = 200 <sup>1</sup> | Drug A, N = 98 <sup>1</sup> | Drug B, N = 102 <sup>1</sup> | p-value <sup>2</sup> |
|----------------|-----|-------------------------------|-----------------------------|------------------------------|----------------------|
| Age, yrs       | 189 | 47 (38, 57)                   | 46 (37, 59)                 | 48 (39, 56)                  | 0.7                  |
| Tumor Response | 193 | 61 (32%)                      | 28 (29%)                    | 33 (34%)                     | 0.5                  |

### add\_n()

Adds a column with the total number of non-missing observations

### add\_overall()

Adds a column with overall summary statistics

See also:  
`add_q()`  
`bold_p()`  
`bold_labels()`  
`add_stat()`

**add\_p()** Adds column of p-values generated by testing for differences between groups. Takes arguments below.

| Argument   | Default                                                                                                                                               | Input                                                                                                                                                                     | Effect on table                                              |
|------------|-------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------|
| test       | Continuous: "Kruskal test";<br>Categorical, expected cell counts ≥5: "chisq.test.no.correct";<br>Categorical, expected cell counts < 5: "fisher.test" | Formula list specifying statistical test to perform for each variable: "t.test", "aov", "wilcox.test", "kruskal.test", "chisq.test" and "lme4"; custom tests possible too | Changes p-value in table based on specified statistical test |
| pvalue_fun | <code>style_pvalue()</code>                                                                                                                           | Function to round and format p-values                                                                                                                                     | Changes format of p-values in table                          |

For more info, visit <http://www.danielsjoberg.com/gtsummary/reference/index.html>

## tbl\_regression() Present regression model object in publication-ready table

### cox model: basic code

```
library(survival)
cox1 = coxph(Surv(ttdeath, death) ~ age + marker,
             data = trial)
tbl_regression(cox1, exponentiate = TRUE)
```

### cox model: basic table

| Characteristic      | HR <sup>1</sup> | 95% CI <sup>1</sup> | p-value |
|---------------------|-----------------|---------------------|---------|
| Age, yrs            | 1.01            | 0.99, 1.02          | 0.4     |
| Marker Level, ng/mL | 0.96            | 0.76, 1.21          | 0.7     |

<sup>1</sup> HR = Hazard Ratio, CI = Confidence Interval

### glm model: basic code

```
m1 <- glm(response ~ age + stage, data = trial,
           family = binomial)
tbl_regression(m1, exponentiate = TRUE)
```

### glm model: custom code using helper functions

```
m1 %>%
  tbl_regression(exponentiate = TRUE) %>%
  add_global_p() %>%
  bold_p(t = 0.10) %>%
  bold_labels() %>%
  italicize_levels()
```

### tbl\_uvregression() displays multiple univariate regression models at once

#### cox univariate models: code

```
library(survival)
tbl_uvregression(
  trial %>% select(ttdeath, death, age,
                     grade, response),
  method = coxph,
  y = Surv(ttdeath, death),
  exponentiate = TRUE,
  label = list(grade ~ "Tumor grade"),
  ) %>%
  add_global_p()
  add_n(location = "level") %>%
  add_nevent(location = "level")
```

#### cox univariate models: table

| Characteristic | N   | Event N | HR <sup>1</sup> | 95% CI <sup>1</sup> | p-value |
|----------------|-----|---------|-----------------|---------------------|---------|
| Age, yrs       | 189 | 103     | 1.01            | 0.99, 1.02          | 0.3     |
| Tumor grade    |     |         |                 |                     | 0.075   |
| I              | 68  | 33      | —               | —                   |         |
| II             | 68  | 36      | 1.28            | 0.80, 2.05          |         |
| III            | 64  | 43      | 1.69            | 1.07, 2.66          |         |
| Tumor Response | 193 | 107     | 0.50            | 0.31, 0.78          | 0.001   |

<sup>1</sup> HR = Hazard Ratio, CI = Confidence Interval

Requires “method” parameter specifying model type. Can estimate univariate regression models holding either outcome (“y”) or covariate (“x”) constant, or both (see “formula” parameter). For more info about “formula” and other parameters, see:

[https://www.danielsjoberg.com/gtsummary/reference/tbl\\_uvregression.html](https://www.danielsjoberg.com/gtsummary/reference/tbl_uvregression.html)

## tbl\_survfit() Present survfit object with custom estimates in publication-ready table

### library(survival)

```
tbl_survfit(
  list(
    survfit(Surv(ttdeath, death) ~ 1, trial),
    survfit(Surv(ttdeath, death) ~ trt, trial)
  ),
  times = c(12, 24),
  label_header = "***{time} Month***"
)
```

### Options for defining model:

1. x=explicit survfit model (or list of models) from dataframe
2. x=dataframe and designate y=Surv object and include=covariates in model

### Characteristic

### 12 Month

### 24 Month

|                        |                |                |
|------------------------|----------------|----------------|
| Overall                | 88% (84%, 93%) | 44% (38%, 51%) |
| Chemotherapy Treatment |                |                |
| Drug A                 | 91% (85%, 97%) | 47% (38%, 58%) |
| Drug B                 | 86% (80%, 93%) | 41% (33%, 52%) |

### library(survival)

```
tbl_survfit(
  trial,
  y = Surv(ttdeath, death),
  include = c(trt, grade),
  probs = 0.5,
  label_header = "***Median Survival***"
) %>% add_p()
```

| Characteristic         | Median Survival | p-value <sup>1</sup> |
|------------------------|-----------------|----------------------|
| Chemotherapy Treatment | 24 (21, —)      | 0.2                  |
| Drug A                 | 24 (21, —)      |                      |
| Drug B                 | 21 (18, —)      |                      |
| Grade                  |                 | 0.072                |
| I                      | — (22, —)       |                      |
| II                     | 22 (18, —)      |                      |
| III                    | 20 (18, 23)     |                      |

<sup>1</sup> Log-rank test

### tbl\_merge(), tbl\_stack() combine tables by row or column

```
t1 = tbl_survfit(
  list(survfit(Surv(ttdeath, death) ~ trt +
               grade, trial),
       times = c(12, 24),
       label_header = "***{time} Month***"
  )
)
```

```
t2 = tbl_survfit(
  trial,
  y = Surv(ttdeath, death),
  include = c(trt, grade),
  probs = 0.5,
  label_header = "***Median Survival***"
) %>% add_p()
```

`tbl_merge(list(t1,t2), tab_spacer = FALSE)`

| Characteristic         | 12 Month        | 24 Month       | Median Survival | p-value <sup>1</sup> |
|------------------------|-----------------|----------------|-----------------|----------------------|
| Chemotherapy Treatment |                 |                |                 | 0.2                  |
| Drug A                 | 91% (85%, 97%)  | 47% (38%, 58%) | 24 (21, —)      |                      |
| Drug B                 | 86% (80%, 93%)  | 41% (33%, 52%) | 21 (18, —)      |                      |
| Grade                  |                 |                |                 | 0.072                |
| I                      | 97% (93%, 100%) | 51% (41%, 65%) | — (22, —)       |                      |
| II                     | 82% (74%, 92%)  | 47% (37%, 61%) | 22 (18, —)      |                      |
| III                    | 86% (78%, 95%)  | 33% (23%, 47%) | 20 (18, 23)     |                      |

<sup>1</sup> Log-rank test

tbl\_merge combines columns, tbl\_stack() combines rows. For more info, see <https://www.danielsjoberg.com/gtsummary/reference/>



# GWAS Catalog access with gwasrapidd

## Introduction

The **GWAS Catalog** is a service provided by the EMBL-EBI and NHGRI that offers a manually curated and freely available database of published genome-wide association studies (GWAS).

The GWAS Catalog data provided by the **RESTful API** is organized around four core entities:

- **studies**
- **associations**
- **variants**
- **traits**

## Get GWAS Catalog Entities

**gwasrapidd** facilitates the access to the Catalog via the RESTful API, allowing you to programmatically retrieve data directly into R. Each of the four entities is mapped to an S4 object of a class of the same name.

| GWAS CATALOG     | RETRIEVAL FUNCTIONS                                  | S4 CLASSES                          |
|------------------|------------------------------------------------------|-------------------------------------|
|                  | get_studies()                                        | <b>S</b> studies                    |
|                  | get_associations()                                   | <b>A</b> associations               |
|                  | get_variants()                                       | <b>V</b> variants                   |
|                  | get_traits()                                         | <b>T</b> traits                     |
| Search by        | Example                                              |                                     |
| study_id         | "CCST000858"                                         | <b>S</b> <b>A</b> <b>V</b> <b>T</b> |
| association_id   | "24300113"                                           |                                     |
| variant_id       | "rs12752552"                                         |                                     |
| efo_id           | "EFO_0005543"                                        |                                     |
| pubmed_id        | "21626137"                                           |                                     |
| user_requested   | TRUE                                                 |                                     |
| full_pvalue_set  | FALSE                                                |                                     |
| efo_uri          | "http://www.ebi.ac.uk/efo/EFO_0004761"               |                                     |
| genomic_range    | list(chromosome = "22", start = 1L, end = 15473564L) |                                     |
| gene_name        | "BRCA1"                                              |                                     |
| efo_trait        | "lung adenocarcinoma"                                |                                     |
| reported_trait   | "Breast cancer"                                      |                                     |
| cytogenetic_band | "1p36.33"                                            |                                     |

## S4 Representation of GWAS Catalog Entities

### S4 class studies

The **studies** object consists of eight slots, each a table (tibble). Each study is an observation (row) in the studies table — main table. All tables have the column `study_id` as primary key.

For details about the studies S4 class: `class?studies`.

| studies                                | genotyping_techs                     | countries_of_recruitment        |
|----------------------------------------|--------------------------------------|---------------------------------|
| • <code>study_id</code>                | • <code>study_id</code>              | • <code>study_id</code>         |
| • <code>reported_trait</code>          | • genotyping technology              | • <code>ancestry_id</code>      |
| • <code>initial_sample_size</code>     |                                      | • <code>country_name</code>     |
| • <code>replication_sample_size</code> | • <code>study_id</code>              | • <code>major_area</code>       |
| • <code>gxe</code>                     | • manufacturer                       | • <code>region</code>           |
| • <code>gxg</code>                     |                                      |                                 |
| • <code>snp_count</code>               | • <code>study_id</code>              | • <code>study_id</code>         |
| • <code>qualifier</code>               | • <code>ancestry_id</code>           | • <code>ancestry_id</code>      |
| • <code>imputed</code>                 | • <code>type</code>                  | • <code>country_name</code>     |
| • <code>pooled</code>                  | • <code>number_of_individuals</code> | • <code>major_area</code>       |
| • <code>study_design_comment</code>    |                                      | • <code>region</code>           |
| • <code>full_pvalue_set</code>         | • <code>study_id</code>              |                                 |
| • <code>user_requested</code>          | • <code>ancestry_id</code>           | • <code>study_id</code>         |
|                                        | • <code>ancestral_group</code>       | • <code>pubmed_id</code>        |
|                                        |                                      | • <code>publication_date</code> |
|                                        |                                      | • publication                   |
|                                        |                                      | • title                         |
|                                        |                                      | • author_fullname               |
|                                        |                                      | • author_orcid                  |

### S4 class associations

The **associations** object consists of six slots, each a table (tibble). Each association is an observation (row) in the associations table — main table. All tables have the column `association_id` as primary key.

For details about the associations S4 class: `class?associations`.

| associations                          | loci                               | genes                         |
|---------------------------------------|------------------------------------|-------------------------------|
| • <code>association_id</code>         | • <code>association_id</code>      | • <code>association_id</code> |
| • <code>pvalue</code>                 | • <code>locus_id</code>            | • <code>locus_id</code>       |
| • <code>pvalue_description</code>     | • <code>haplotype.snp_count</code> | • <code>gene_name</code>      |
| • <code>pvalue_mantissa</code>        | • <code>description</code>         |                               |
| • <code>pvalue_exponent</code>        |                                    | • <code>ensembl_id</code>     |
| • <code>multiple.snp.haplotype</code> | • <code>association_id</code>      | • <code>variant_id</code>     |
| • <code>snp_interaction</code>        | • <code>locus_id</code>            | • <code>gene_name</code>      |
| • <code>snp_type</code>               | • <code>variant_id</code>          | • <code>ensembl_id</code>     |
| • <code>standard_error</code>         | • <code>risk_allele</code>         | • <code>entrez_id</code>      |
| • <code>range</code>                  | • <code>risk_frequency</code>      | • <code>association_id</code> |
| • <code>or_per_copy_number</code>     | • <code>genome_wide</code>         | • <code>locus_id</code>       |
| • <code>beta_number</code>            | • <code>limited_list</code>        | • <code>gene_name</code>      |
| • <code>beta_unit</code>              |                                    | • <code>entrez_id</code>      |
| • <code>beta_direction</code>         |                                    |                               |
| • <code>beta_description</code>       |                                    |                               |
| • <code>last_mapping_date</code>      |                                    |                               |
| • <code>last_update_date</code>       |                                    |                               |

# h2o:: CHEAT SHEET

## Dataset Operations

### DATA IMPORT / EXPORT

**h2o.uploadFile:** Upload a file into H2O from a client-side path, and parse it.

**h2o.downloadCSV:** Download a H2O dataset to a client-side CSV file.

**h2o.importFile:** Import a file into H2O from a server-side path, and parse it.

**h2o.exportFile:** Export an H2O Data Frame to a server-side file.

**h2o.parseRaw:** Parse a raw data file.

### NATIVE R TO H2O COERCION

**as.h2o:** Convert a R object to an H2O object

### H2O TO NATIVE R COERCION

**as.data.frame:** Check if an object is a data frame, and coerce it if possible.

### DATA GENERATION

**h2o.createFrame:** Creates a data frame in H2O with real-valued, categorical, integer, and binary columns specified by the user, with optional randomization.

**h2o.runif:** Produce a vector of random uniform numbers.

**h2o.interaction:** Create interaction terms between categorical features of an H2O Frame.

**h2o.target\_encode\_apply:** Target encoding map to an H2O Data Frame, which can improve performance of supervised learning models for high cardinality categorical columns.

### DATA SAMPLING / SPLITTING

**h2o.splitFrame:** Split an existing H2O dataset according to user-specified ratios.

### MISSING DATA HANDLING

**h2o.impute:** Impute a column of data using the mean, median, or mode.

**h2o.insertMissingValues:** Replaces a user-specified fraction of entries in an H2O dataset with missing values.

**h2o.na.omit:** Remove Rows With NAs.

## General Operations

### SUBSCRIPTING

Subscripting example to pull (/push) pieces from (/to) a H2O Parsed Data object.

|                  |              |          |                  |
|------------------|--------------|----------|------------------|
| x[j] ## column J | x[i]         | <- value | Value Assignment |
| x[i, j]          | x[i, j, ...] | <- value |                  |
| x[[i]]           | x[[i]]       | <- value |                  |
| x\$name          | x\$i         | <- value |                  |

### Selection

### Value Assignment

### SUBSETTING

**h2o.head, h2o.tail:** Object's Start or End.

### DATA ATTRIBUTES

**h2o.names:** Return column names for an H2O Frame. Also: **h2o.colnames**

**names<-:** Set the row or column names of a H2O Frame. Also: **colnames<-**

**h2o.dim:** Retrieve object dimensions.

**h2o.length:** Length of vector, list or factor.

**h2o.nrow:** Number of H2O Frame rows.

**h2o.ncol:** Number of H2O Frame columns.

**h2o.anyFactor:** Check if an H2O Frame object has any categorical data columns.

**is.factor, is.character, is.numeric:** Check Column's Data Type.

### DATA TYPE COERCION:

**h2o.asfactor, as.factor:** Factor.

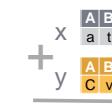
**h2o.as\_date, as.Date:** Date.

**h2o.ascharacter, as.character:** Character.

**h2o.asnumeric, as.numeric:** Numeric.

### BASIC DATA MANIPULATION

**c:** Combine Values into a Vector or List.

 **h2o.cbind; h2o.rbind:** Combine a sequence of H2O datasets by column (cbind) or rows (rbind).

 **h2o.merge:** Merges 2 H2OFrames.

 **h2o.arrange:** Sorts an H2OFrame by columns.

### ELEMENT INDEX SELECTION

**h2o.which:** True Condition's Row Numbers

### CONDITIONAL VALUE SELECTION

**h2o.ifelse:** Apply conditional statements to numeric vectors in an H2O Frame.

## Math Operations

### (math) vectorized function

### MATH

**h2o.abs:** Compute the absolute value of x.

**h2o.sqrt:** Principal Square Root of x,  $\sqrt{x}$ .

**h2o.ceiling:** Take a single numeric argument x and return a numeric vector containing the smallest integers not less than the corresponding elements of x.

**h2o.floor:** Take a single numeric argument x and return a numeric vector containing the largest integers not greater than the corresponding elements of x.

**h2o.trunc:** Take a single numeric argument x and return a numeric vector containing the integers formed by truncating the values in x toward 0.

**h2o.log:** Compute natural logarithms. See also: **h2o.log10, h2o.log2, h2o.log1p**

**h2o.exp:** Compute the exponential function

**h2o.cos, h2o.cosh, h2o.acos, h2o.sin, h2o.tan, h2o.tanh, Math:** ?groupGeneric

**sign:** Return a vector with the signs of the corresponding elements of x (the sign of a real number is 1, 0, or -1 if the number is positive, zero, or negative, respectively).

**&& (Vectorized AND), || (Vectorized OR), !x, %in%, Ops: +, -, \*, /, ^, %%, %/%, ==, !=, <, <=, >=, >, &, |, !**

### CUMULATIVE

**h2o.cummax:** Vector of the cumulative maxima of the elements of the argument.

**h2o.cummin:** Vector of the cumulative minima of the elements of the argument.

**h2o.cumprod:** Vector of the cumulative products of the elements of the argument.

**h2o.cumsum:** Vector of the cumulative sums of the elements of the argument.

### PRECISION

**h2o.round:** Round values to the specified number of decimal places. The default is 0.

**h2o.signif:** Round values to the specified number of significant digits.

## Group By Summaries

### (group by) summary function

**nrow:** Count the number of rows.

**max:** All input argument's Maximum.

**min:** All input argument's Minimum.

**sum:** All argument values Sum.

**mean:** (Trimmed) arithmetic mean.

**sd:** Calculate the standard deviation of a column of continuous real valued data.

**var:** Compute the variance of x.

## Generic Summaries

### NON-GROUP\_BY SUMMARIES

**h2o.median:** Calculate the median of x.

**h2o.range:** Input argument's Min/Max Vector

**h2o.cor:** Correlation Matrix of H2O Frames.

**h2o.quantile:** Obtain and display quantiles for an H2O Frame Column.

 **h2o.hist:** Compute a histogram over a numeric H2O Frame Column.

**h2o.prod:** Product of all arguments values.

**h2o.any:** Given a set of logical vectors, determine if at least one of the values is true.

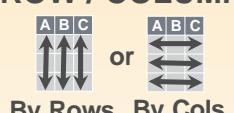
**h2o.all:** Given a set of logical vectors, determine if all of the values are true.

### NON-GROUP\_BY SUMMARIES: GENERIC

**h2o.summary:** Produce result summaries of the results of various model fitting functions.

## Aggregations

### ROW / COLUMN AGGREGATION



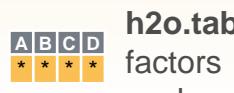
**apply:** Apply a function over an H2O parsed data object (an array) margins.

### GROUP BY AGGREGATION



**h2o.group\_by:** Apply an aggregate function to each group of an H2O dataset.

### TABULATION



**h2o.table:** Use the cross-classifying factors to build a table of counts at each combination of factor levels.

# h2o::CHEAT SHEET



## Data Modeling

### MODEL TRAINING: SUPERVISED LEARNING

**h2o.deeplearning:** Perform Deep Learning Neural Networks on an H2OFrame.

**h2o.gbm:** Build Gradient Boosted Regression Trees or Classification Trees.

**h2o.glm:** Fit a Generalized Linear Model, specified by a response variable, a set of predictors, and the error distribution.

**h2o.naiveBayes:** Compute Naive Bayes classification probabilities on an H2O Frame.

**h2o.randomForest:** Perform Random Forest Classification on an H2O Frame.

**h2o.xgboost:** Build an Extreme Gradient Boosted Model using the XGBoost backend.

**h2o.stackedEnsemble:** Build a stacked ensemble (aka. Super Learner) using the specified H2O base learning algorithms.

**h2o.automl:** Automates the Supervised Machine Learning Model Training Process: Automatically Trains and Cross-validates a set of Models, and trains a Stacked Ensemble.

### MODEL TRAINING: UNSUPERVISED LEARNING

**h2o.prcomp:** Perform Principal Components Analysis on the given H2O Frame.

**h2o.kmeans:** Perform k-means Clustering on the given H2O Frame.

**h2o.anomaly:** Detect anomalies in a H2O Frame using a H2O Deep Learning Model with Auto-Encoding.

**h2o.deepfeatures:** Extract the non-linear features from a H2O Frame using a H2O Deep Learning Model.

**h2o.glrn:** Builds a Generalized Low Rank Decomposition of an H2O Frame.

**h2o.svd:** Singular value decomposition of an H2O Frame using the power method.

**h2o.word2vec:** Trains a word2vec model on a String column of an H2O data frame.

### SURVIVAL MODELS: TIME-TO-EVENT

**h2o.coxph:** Trains a Cox Proportional Hazards Model (CoxPH) on an H2O Frame.

### GRID SEARCH

**h2o.grid:** Efficient method to build multiple models with different hyperparameters.

**h2o.getGrid:** Get a grid object from H2O distributed K/V store.

### MODEL SCORING

**h2o.predict:** Obtain predictions from various fitted H2O model objects.

**h2o.scoreHistory:** Get Model Score History.

### MODEL METRICS

**h2o.make\_metrics:** Given predicted values (target for regression, class-1 probabilities, or binomial or per-class probabilities for multinomial), compute a model metrics object.

### GENERAL MODEL HELPER

**h2o.performance:** Evaluate the predictive performance of a Supervised Learning Regression or Classification Model via various metrics. Set **xval = TRUE** for retrieving the training cross-validation metrics.

### REGRESSION MODEL HELPER

**h2o.mse:** Display the mean squared error calculated from "Predicted Responses" and "Actual (Reference) Responses". Set **xval = TRUE** for retrieving the cross-validation MSE.

### CLASSIFICATION MODEL HELPERS

**h2o.accuracy:** Get Model Accuracy metric.

**h2o.auc:** Retrieve the AUC (area under ROC curve). Set **xval = TRUE** for retrieving the cross-validation AUC.

**h2o.confusionMatrix:** Display prediction errors for classification data ("Predicted" vs "Reference : Real Values").

**h2o.hit\_ratio\_table:** Retrieve the Hit Ratios. Set **xval = TRUE** for retrieving the cross-validation Hit Ratio.

### CLUSTERING MODEL HELPER

**h2o.betweenss:** Get the between cluster Sum of Squares.

**h2o.centers:** Retrieve the Model Centers.

### PREDICTOR VARIABLE IMPORTANCE

**h2o.varimp:** Retrieve the variable importance

**h2o.varimp\_plot:** Plot Variable Importances.

## Data Munging

### GENERAL COLUMN MANIPULATION

**is.na:** Display missing elements.

### FACTOR LEVEL MANIPULATIONS

**h2o.levels:** Display a list of the unique values found in a categorical data column.

**h2o.relevel:** Reorders levels of an H2O factor, similarly to standard R's `relevel`.

**h2o.setLevels:** Set Levels of H2O Factor.

### NUMERIC COLUMN MANIPULATIONS

**h2o.cut:** Convert H2O Numeric Data to Factor by breaking it into Intervals.

### CHARACTER COLUMN MANIPULATIONS

**h2o.strsplit:** "String Split": Splits the given factor column on the input split.

**h2o.tolower:** Convert the characters of a character vector to lower case.

**h2o.toupper:** Convert the characters of a character vector to upper case.

**h2o.trim:** "Trim spaces": Remove leading and trailing white space.

**h2o.gsub:** Match a pattern & replace **all** instances (occurrences) of the matched pattern with the replacement string globally.

**h2o.sub:** Match a pattern & replace the **first** instance (occurrence) of the matched pattern with the replacement string.

### DATE MANIPULATIONS

**h2o.month:** Convert Milliseconds to Months in H2O Datasets (Scale: 0 to 11).

**h2o.year:** Convert Milliseconds to Years in H2O Datasets, indexed starting from 1900.

**h2o.day:** Convert Milliseconds to Day of Month in H2O Datasets (Scale: 1 to 31).

**h2o.hour:** Convert Milliseconds to Hour of Day in H2O Datasets (Scale: 0 to 23).

**h2o.dayOfWeek:** Convert Milliseconds to Day of Week in a H2OFrame (Scale: 0 to 6)

### MATRIX OPERATIONS

**%\*%:** Multiply two conformable matrices.

**t:** Returns the transpose of an H2OFrame.

## Cluster Operations

### H2O KEY VALUE STORE ACCESS

**h2o.assign:** Assign H2O hex.keys to R objects.

**h2o.getFrame:** Get H2O dataset Reference.

**h2o.getModel:** Get H2O model reference.

**h2o.ls:** Display a list of object keys in the running instance of H2O.

**h2o.rm:** Remove specified H2O Objects from the H2O server, but not from the R environment.

**h2o.removeAll:** Remove All H2O Objects from the H2O server, but not from the R environment.

### H2O MODEL IMPORT / EXPORT

**h2o.loadModel:** Load H2OModel from disk.

**h2o.saveModel:** Save H2OModel object to disk.

**h2o.download\_pojo:** Download the Scoring POJO (Plain Old Java Object) of an H2O Model.

**h2o.download\_mojo:** Download the model in MOJO format.

### H2O CLUSTER CONNECTION

**h2o.init:** Connect to a running H2O instance using all CPUs on the host.

**h2o.shutdown:** Shut down the specified H2O instance. All data on the server will be lost!

### H2O CLUSTER INFORMATION

**h2o.clusterInfo:** Display the name, version, uptime, total nodes, total memory, total cores and health of a cluster running H2O.

**h2o.clusterStatus:** Retrieve information on the status of the cluster running H2O.

### H2O LOGGING

**h2o.clearLog:** Clear all H2O R command and error response logs from the local disk.

**h2o.downloadAllLogs:** Download all H2O log files to the local disk.

**h2o.logAndEcho:** Write a message to the H2O Java log file and echo it back.

**h2o.openLog:** Open existing logs of H2O R POST commands and error responses on disk.

**h2o.getLogPath:** Get the file path for the H2O R command and error response logs.

**h2o.startLogging:** Begin logging H2O R POST commands and error responses.

**h2o.stopLogging:** Stop logging H2O R POST commands and error responses.

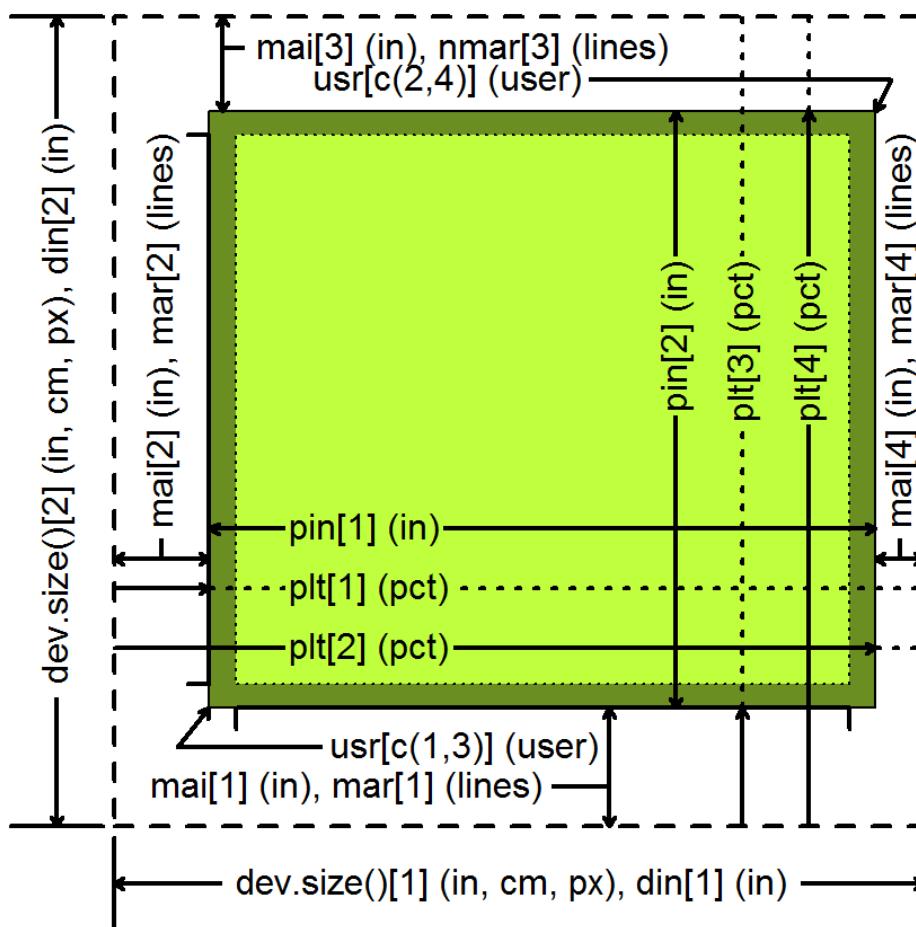


# How Big is Your Graph?

## An R Cheat Sheet

### Introduction

All functions that open a device for graphics will have **height** and **width** arguments to control the size of the graph and a **pointsize** argument to control the relative font size. In **knitr**, you control the size of the graph with the chunk options, **fig.width** and **fig.height**. This sheet will help you with calculating the size of the graph and various parts of the graph within R.



### Your graphics device

**dev.size()** (width, height)  
**par("din")** (r.o.) (width, height) in inches

Both the **dev.size** function and the **din** argument of **par** will tell you the size of the graphics device. The **dev.size** function will report the size in

1. inches (**units="in"**), the default
2. centimeters (**units="cm"**)
3. pixels (**units="px"**)

Like several other **par** arguments, **din** is read only (r.o.) meaning that you can ask its current value (**par("din")**) but you cannot change it (**par(din=c(5,7))** will fail).

### Your plot margins

**par("mai")** (bottom, left, top, right) in inches  
**par("mar")** (bottom, left, top, right) in lines

Margins provide you space for your axes, axis labels, and titles.

A "line" is the amount of vertical space needed for a line of text.

If your graph has no axes or titles, you can remove the margins (and maximize the plotting region) with

```
par(mar=rep(0,4))
```

### Your plotting region

**par("pin")** (width, height) in inches  
**par("plt")** (left, right, bottom, top) in pct

The **pin** argument **par** gives you the size of the plotting region (the size of the device minus the size of the margins) in inches.

The **plt** argument **par** gives you the percentage of the device from the left/bottom edge up to the left edge of the plotting region, the right edge, the bottom edge, and the top edge. The first and third values are equivalent to the percentage of space devoted to the left and bottom margins. Subtract the second and fourth values from 1 to get the percentage of space devoted to the right and top margins.

### Your x-y coordinates

**par("usr")** (xmin, ymin, xmax, ymax)

Your x-y coordinates are the values you use when plotting your data. This normally is not the same as the values you specified with the **xlim** and **ylim** arguments in **plot**. By default, R adds an extra 4% to the plotting range (see the dark green region on the figure) so that points right up on the edges of your plot do not get partially clipped. You can override this by setting **xaxs="i"** and/or the **yaxs="i"** in **par**.

Run **par("usr")** to find the minimum X value, the maximum X value, the minimum Y value, and the maximum Y value. If you assign new values to **usr**, you will update the x-y coordinates to the new values.

### Getting a square graph

**par("pty")**

You can produce a square graph manually by setting the width and height to the same value and setting the margins so that the sum of the top and bottom margins equal the sum of the left and right margins. But a much easier way is to specify **pty="s"**, which adjusts the margins so that the size of the plotting region is always square, even if you resize the graphics window.

### Converting units

For many applications, you need to be able to translate user coordinates to pixels or inches. There are some cryptic shortcuts, but the simplest way is to get the range in user coordinates and measure the proportion of the graphics device devoted to the plotting region.

```
user.range <- par("usr")[c(2,4)] - par("usr")[c(1,3)]
```

```
region.pct <- par("plt")[c(2,4)] - par("plt")[c(1,3)]
```

```
region.px <- dev.size(units="px") * region.pct
```

```
px.per.xy <- region.px / user.range
```

To convert a horizontal or distance from the x-coordinate value to pixels, multiply by **px.per.xy[1]**. To convert a vertical distance, multiply by **region.px.per.xy[2]**. To convert a diagonal distance, you need to invoke Pythagoras.

```
a.px <- x.dist*px.per.xy[1]
b.px <- y.dist*px.per.xy[2]
c.px <- sqrt(a.px^2+b.px^2)
```

To rotate a string to match the slope of a line segment, you need to convert the distances to pixels, calculate the arctangent, and convert from radians to degrees.

```
segments(x0, y0, x1, y1)
delta.x <- (x1 - x0) * px.per.xy[1]
delta.y <- (y1 - y0) * px.per.xy[2]
angle.radians <- atan2(delta.y, delta.x)
angle.degrees <- angle.radians * 180 / pi
text(x1, y1, "TEXT", srt=angle.degrees)
```

## Panels

`par("fig")` (width, height) in pct  
`par("fin")` (width, height) in inches

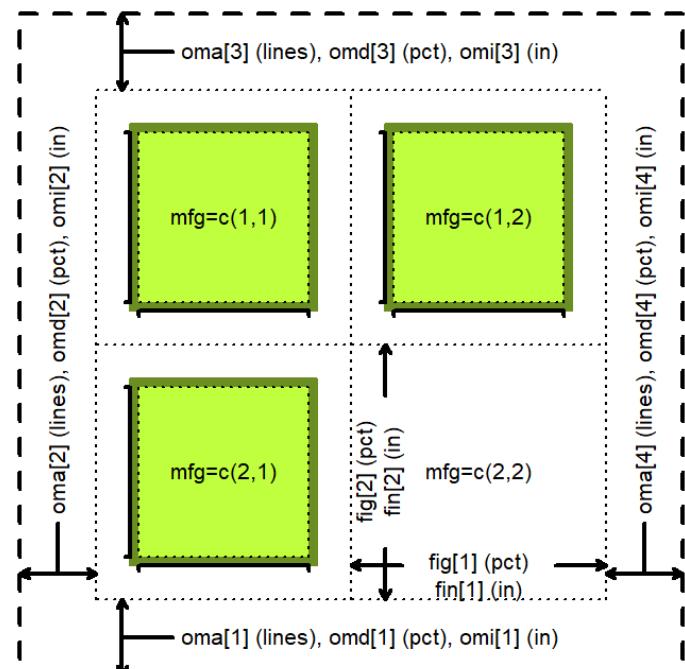
If you display multiple plots within a single graphics window (e.g., with the `mfrow` or `mfcol` arguments of `par` or with the `layout` function), then the `fig` and `fin` arguments will tell you the size of the current subplot window in percent or inches, respectively.

`par("oma")` (bottom, left, top, right) in lines  
`par("omd")` (bottom, left, top, right) in pct  
`par("omi")` (bottom, left, top, right) in inches

Each subplot will have margins specified by `mai` or `mar`, but no outer margin around the entire set of plots, unless you specify them using `oma`, `omd`, or `omi`. You can place text in the outer margins using the `mtext` function with the argument `outer=TRUE`.

`par("mfg")` (r, c) or (r, c, maxr, maxc)

The `mfg` argument of `par` will allow you to jump to a subplot in a particular row and column. If you query with `par("mfg")`, you will get the current row and column followed by the maximum row and column.



## Character and string sizes

### `strheight()`

The `strheight` functions will tell you the height of a specified string in inches (`units="inches"`), x-y user coordinates (`units="user"`) or as a percentage of the graphics device (`units="figure"`).

For a single line of text, `strheight` will give you the height of the letter "M". If you have a string with one or more linebreaks ("n"), the `strheight` function will measure the height of the letter "M" plus the height of one or more additional lines. The height of a line is dependent on the line spacing, set by the `lheight` argument of `par`. The default line height (`lheight=1`), corresponding to single spaced lines, produces a line height roughly 1.5 times the height of "M".

### `strwidth()`

The `strwidth` function will produce different widths to individual characters, representing the proportional spacing used by most fonts ("W" using much more space than an "i"). For the width of a string, the `strwidth` function will sum up the lengths of the individual characters in the string.

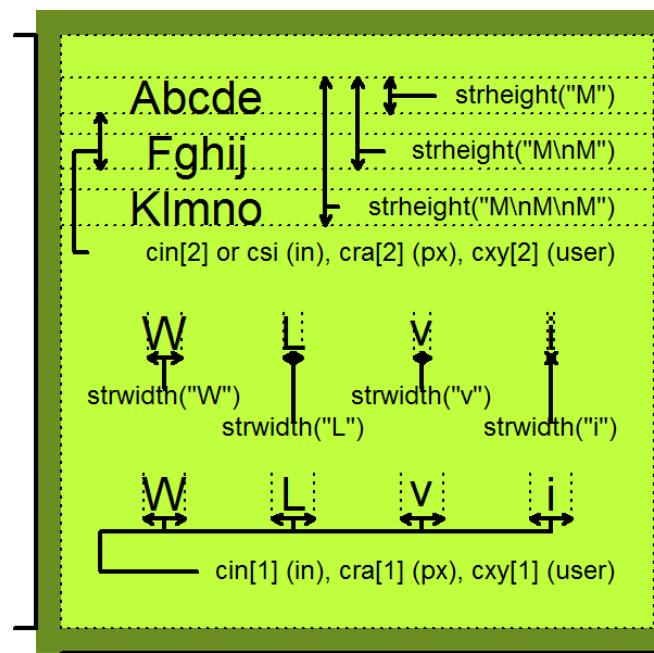
`par("cin")` (r.o.) (width, height) in inches  
`par("csi")` (r.o.) height in inches  
`par("cra")` (r.o.) (width, height) in pixels  
`par("cxy")` (r.o.) (width, height) in xy coordinates

The single value returned by the `csi` argument of `par` gives you the height of a line of text in inches. The second of the two values returned by `cin`, `cra`, and `cxy` gives you the height of a line, in inches, pixels, or xy (user) coordinates.

The first of the two values returned by the `cin`, `cra`, and `cxy` arguments to `par` gives you the approximate width of a single character, in inches, pixels, or xy (user) coordinates. The width, very slightly smaller than the actual width of the letter "W", is a rough estimate at best and ignores the variable width of individual letters.

These values are useful, however, in providing fast ratios of the relative sizes of the differing units of measure

`px.per.in <- par("cra") / par("cin")`  
`px.per.xy <- par("cra") / par("cxy")`  
`xy.per.in <- par("cxy") / par("cin")`



## If your fonts are too big or too small

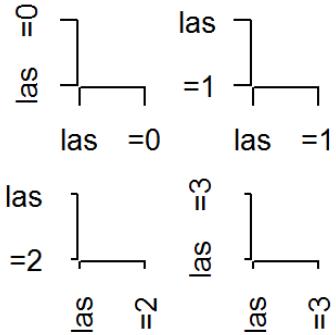
Fixing this takes a bit of trial and error.

- Specify a larger/smaller value for the `pointsize` argument when you open your graphics device.
- Try opening your graphics device with different values for `height` and `width`. Fonts that look too big might be better proportioned in a larger graphics window.
- Use the `cex` argument to increase or decrease the relative size of your fonts.

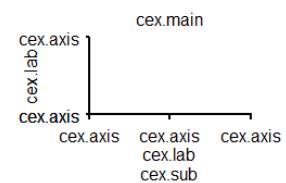
## If your axes don't fit

There are several possible solutions.

- You can assign wider margins using the `mar` or `mai` argument in `par`.
- You can change the orientation of the axis labels with `las`. Choose among
  - `las=0` both axis labels parallel
  - `las=1` both axis labels horizontal
  - `las=2` both axis labels perpendicular
  - `las=3` both axis labels vertical.



- change the relative size of the font
  - `cex.axis` for the tick mark labels.
  - `cex.lab` for `xlab` and `ylab`.
  - `cex.main` for the main title
  - `cex.sub` for the subtitle.





# Time Series Imputation with imputeTS: : CHEAT SHEET

## Mission

Missing Data is nearly everywhere. Also in time series, especially in sensor recordings missing data is common.

*imputeTS helps you with your missing data problems.*

## Features

The package provides easy to use functions in these areas:

### 1. Imputation Functions

Several algorithms for replacing NAs with reasonable values (imputation).



### 2. Missing Data Visualizations

Plots for analysis of the distribution of NAs, patterns and imputation performance.



### 3. Stats and Datasets

Functions for printing missing data stats and benchmarking datasets.



## Scope

imputeTS specializes on univariate time series that are:



- numeric
- equally-spaced

## Visualizations

There are multiple plots provided for analysing the missing data before and after imputation. All plotting functions start with `ggplot_na_plotname`.

### Function

### Description

|                                     |                                        |
|-------------------------------------|----------------------------------------|
| <code>ggplot_na_distribution</code> | Getting a first overview of NAs        |
| <code>ggplot_na_intervals</code>    | Insights about NAs in specific periods |
| <code>ggplot_na_gapsizes</code>     | Insights about occurring NA gapsizes   |
| <code>ggplot_na_imputations</code>  | Evaluating imputation quality          |

## Imputation

The package offers multiple missing data replacement (imputation) functions, which are really easy to use.

```
na_interpolation(x, option = "spline")
```

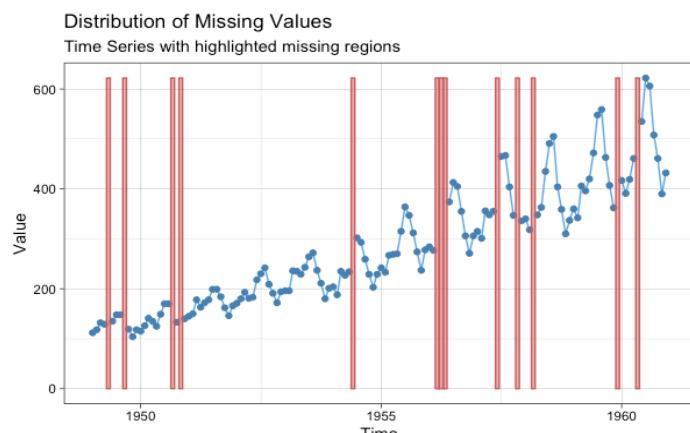
Imputation Function    Your input time series    Additional Parameters

### List of available Algorithms

| Function                      | Description                               |
|-------------------------------|-------------------------------------------|
| <code>na_interpolation</code> | Imputation by Interpolation               |
| <code>na_kalman</code>        | Imputation by Kalman Smoothing            |
| <code>na_locf</code>          | Last Observation Carried Forward          |
| <code>na_ma</code>            | Imputation by Moving Average              |
| <code>na_mean</code>          | Imputation by Mean Value                  |
| <code>na_random</code>        | Imputation by Random Sample               |
| <code>na_remove</code>        | Remove Missing Values                     |
| <code>na_replace</code>       | Replace Missing Values by a Defined Value |
| <code>na_seadec</code>        | Seasonally Decomposed Imputation          |
| <code>na_seasplit</code>      | Seasonally Splitted Imputation            |

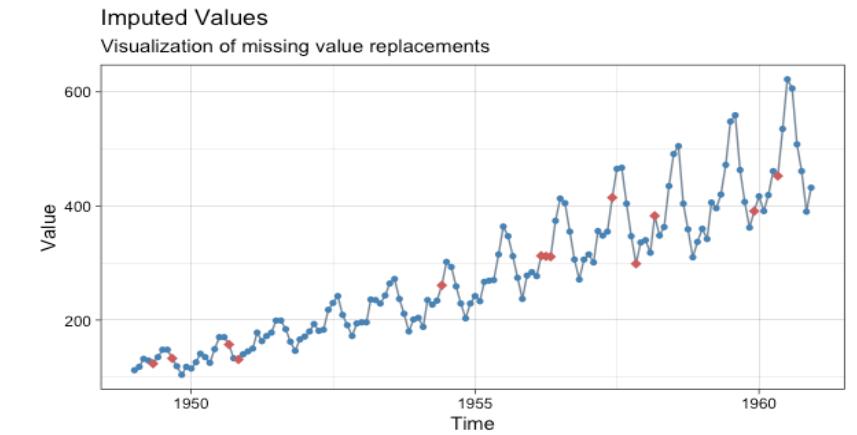
## Missing Data Overview Plots

The ‘distribution’, ‘intervals’ and ‘gapsize’ plots can be used on new datasets to gain insights about missing data patterns and distribution.



## Imputation Analysis Plots

Imputation results can be visualized with the ‘imputations’ plot. Here first `na_kalman` is performed and then the results are plotted.



```
imp <- na_kalman(tsAirgap)
ggplot_na_imputations(tsAirgap, imp)
```

imputation    visualization

## Workflows

The functions also work well in tidy style pipe workflows. Here an example of first using imputation and later forecasting and plotting.

```
library("forecast")
tsAirgap %>% na_interpolation() %>%
  ets() %>% forecast(h=36) %>%
  autoplot()
```

can be put in pipe workflows

a 36 step forecast is created and plotted

## Datasets

The package includes three datasets for imputation experiments.

### Function

### Description

|                        |                                                                                  |
|------------------------|----------------------------------------------------------------------------------|
| <code>tsAirgap</code>  | Monthly totals of international airline passengers.<br>144 Observations / 13 NAs |
| <code>tsNH4</code>     | NH4 concentration in a wastewater system.<br>3552 observations / 883 NAs         |
| <code>tsHeating</code> | A heating systems supply temperature.<br>606837 observations / 57391 NAs         |

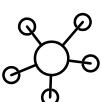
## CITATION

You can cite `imputeTS` the following:

Moritz, Steffen and Bartz-Beielstein, Thomas.

"imputeTS: Time Series Missing Value Imputation in R."

R Journal 9.1 (2017). doi: 10.32614/RJ-2017-009.



# Audit sampling with jfa:: : CHEAT SHEET



## Basics

jfa is an R package that facilitates statistical planning, selection, and evaluation of audit samples.

The package provides five functions that allow users to easily apply Bayesian or classical probability theory in the standard audit sampling workflow.

## Installation

Installing the package can be done via:  
`install.packages('jfa')`

Loading the package can be done via:  
`library(jfa)`

## Example

The blue code blocks next to the function descriptions provide a working example of the intended workflow.

The data for this example can be loaded via:  
`data('BuildIt')`



## Construct a prior probability distribution (optional)

`jfa::auditPrior()`

This function creates a prior distribution for the misstatement in the population based on audit evidence specified via the `method` argument. The prior distribution can be used as input for the `prior` argument in other functions to perform Bayesian inference.

- `likelihood`: Specifies the family of the prior probability distribution.

```
auditPrior(method = 'default',
           likelihood = 'poisson', ...)
```

## Calculate the minimum sample size

`jfa::planning()`

Given a performance materiality or a minimum precision, this function calculates the minimum sample size to achieve these objectives based on the binomial, Poisson, or hypergeometric `likelihood`. A `prior` can be specified to perform Bayesian planning.

- `expected`: A fraction or an integer specifying the expected errors in the sample.

```
planning(materiality = 0.05,
          expected = 0.01,
          likelihood = 'poisson',
          conf.level = 0.95,
          prior = FALSE, ...)
```

## Select the required items from the population

`jfa::selection()`

This function takes a data frame and performs sampling according to one of three popular `method`'s: random sampling, cell sampling, or fixed interval sampling. Sampling is done in combination with one of two sampling `units`: items (rows) or monetary units.

```
selection(data = BuildIt,
           size = 93,
           units = 'items',
           method = 'interval', ...)
```

## Evaluate the misstatement in the population

`jfa::evaluation()`

This function takes a data sample (using `data`, `values`, and `values.audit`) or summary statistics from a sample (using `x` and `n`) and performs statistical evaluation on the misstatement in the population according to the specified `method`. A `prior` can be specified to perform Bayesian evaluation.

- `prior`: An object returned by `auditPrior()` that specifies the prior distribution.

```
evaluation(materiality = 0.05,
            method = 'poisson',
            alternative = 'less',
            conf.level = 0.95,
            x = 0,
            n = 93,
            prior = FALSE, ...)
```

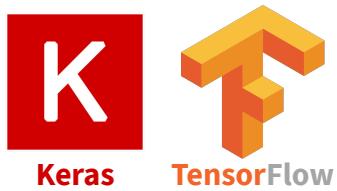
## Create a report of the statistical results

`jfa::report()`

This function takes an object of class `jfaEvaluation` as returned by `evaluation()` and automatically generates a report containing the statistical results and their interpretation.

```
report(object = evaluationResult,
        file = 'report.html', ...)
```

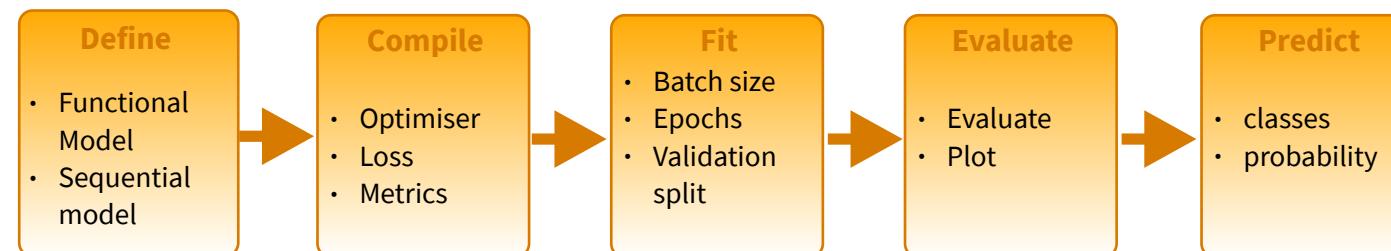
# Deep Learning with Keras3 :: CHEATSHEET



## Intro

[Keras](#) is a high-level neural networks API developed with a focus on enabling fast experimentation. It supports multiple back-ends, including TensorFlow, Jax and Torch.

Backends like TensorFlow are lower level mathematical libraries for building deep neural network architectures. The keras3 R package makes it easy to use Keras with any backend in R.



<https://keras.posit.co>

<https://www.manning.com/books/deep-learning-with-r-second-edition>

The “Hello, World!”  
of deep learning

## Working with Keras Models

### DEFINE A MODEL

#### Functional API: `keras_input()` and `keras_model()`

Define a Functional Model with inputs and outputs.  
`inputs <- keras_input(<input-shape>)`  
`outputs <- inputs |>`  
`layer_dense() |> layer....`  
`model <- keras_model(inputs, outputs)`

#### Sequential API: `keras_model_sequential()`

Define a Sequential Model composed of a linear stack of layers

```
model <-  
  keras_model_sequential(<input-shape>) |>  
  layer_dense() |> layer....
```

#### Subclassing API: `Model()`

Subclass the base Model class

### COMPILE A MODEL

#### `compile()`

Configure a Keras model for training

### FIT A MODEL

#### `fit()`

Train a Keras model for a fixed number of epochs (iterations)

Customize training:

- Provide callbacks to `fit()`:
- Define a custom `Callback()`.
- Call `train_on_batch()` in a custom training loop.
- Subclass `Model()` and implement a custom `train_step` method.
- Write a fully custom training loop. Update weights with `model$optimizer$apply(gradients, weights)`

### INSPECT A MODEL

`print(model)` Print a summary of a Keras model

```
plot(model, show_shapes = FALSE, show_dtype =  
      FALSE, show_layer_names = FALSE, ...)  
Plot a Keras model
```

### EVALUATE A MODEL

`evaluate(object, x = NULL, y = NULL, batch_size = NULL)` Evaluate a Keras model

### PREDICT

`predict()` Generate predictions from a Keras model

`predict_on_batch()` Returns predictions for a single batch of samples.

### SAVE/LOAD A MODEL

`save_model(); load_model()`  
Save/Load models using the ".keras" file format.

`save_model_weights(); load_model_weights()`  
Save/load model weights to/from ".h5" files.

`save_model_config(); load_model_config()`  
Save/load model architecture to/from a ".json" file.

### Deploy

Export just the forward pass of the trained model for inference serving.

`export_savedmodel(model, "my-saved-model/1")`  
Save a TF SavedModel for inference.

`rsconnect::deployTFModel("my-saved-model")`

Deploy a TF SavedModel to Connect for inference.

### CORE LAYERS



`layer_dense()` Add a densely-connected NN layer to an output



`layer_einsum_dense()` Add a dense layer with arbitrary dimensionality



`layer_activation()` Apply an activation function to an output



`layer_dropout()` Applies Dropout to the input



`layer_reshape()` Reshapes an output to a certain shape



`layer_permute()` Permute the dimensions of an input according to a given pattern



`layer_repeat_vector()` Repeats the input n times



`layer_lambda(object, f)` Wraps arbitrary expression as a layer



`layer_activity_regularization()`  
Layer that applies an update to the cost function based input activity



`layer_masking()` Masks a sequence by using a mask value to skip timesteps



`layer_flatten()` Flattens an input

## INSTALLATION

The keras3 R package uses the Python Keras library. You can install all the prerequisites directly from R. See `?keras3::install_keras` for details and options.

```
library(keras3)  
reticulate::install_python()  
install_keras()
```

This installs the required libraries in virtual environment named '**r-keras**'.

It will automatically detect if a GPU is available.

## TRAINING AN IMAGE RECOGNIZER ON MNIST DATA

5041

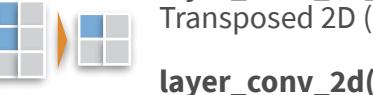
```
# input layer: use MNIST images  
mnist <- dataset_mnist()  
x_train <- mnist$train$x; y_train <- mnist$train$y  
x_test <- mnist$test$x; y_test <- mnist$test$y  
  
# reshape and rescale  
x_train <- array_reshape(x_train, c(nrow(x_train), 784))  
x_test <- array_reshape(x_test, c(nrow(x_test), 784))  
x_train <- x_train / 255; x_test <- x_test / 255  
  
y_train <- to_categorical(y_train, 10)  
y_test <- to_categorical(y_test, 10)  
  
# defining the model and layers  
model <-  
  keras_model_sequential(input_shape = c(28, 28, 1))  
model |>  
  layer_conv_2d(filters = 32, kernel_size = c(3, 3),  
    activation = "relu") |>  
  layer_max_pooling_2d(pool_size = c(2, 2)) |>  
  layer_conv_2d(filters = 64, kernel_size = c(3, 3),  
    activation = "relu") |>  
  layer_max_pooling_2d(pool_size = c(2, 2)) |>  
  layer_flatten() |>  
  layer_dropout(rate = 0.5) |>  
  layer_dense(units = num_classes,  
    activation = "softmax")  
  
# View the model summary  
summary(model)  
plot(model)  
  
# compile (define loss and optimizer)  
model |> compile(  
  loss = 'categorical_crossentropy',  
  optimizer = optimizer_rmsprop(),  
  metrics = c('accuracy'))  
)  
  
# train (fit)  
model |> fit(  
  x_train, y_train,  
  epochs = 30, batch_size = 128,  
  validation_split = 0.2  
)  
model |> evaluate(x_test, y_test)  
model |> predict(x_test)  
  
# save the full model  
save_model(model, "mnist-classifier.keras")  
  
# deploy for serving inference.  
dir.create("serving-mnist-classifier")  
export_savedmodel(model, "serving-mnist-classifier/1")  
rsconnect::deployTFModel("serving-mnist-classifier")
```

## More layers

### CONVOLUTIONAL LAYERS



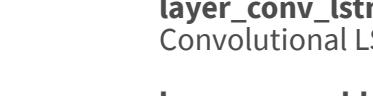
`layer_conv_1d()` 1D, e.g. temporal convolution



`layer_conv_2d_transpose()` Transposed 2D (deconvolution)



`layer_conv_2d()` 2D, e.g. spatial convolution over images

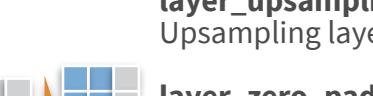


`layer_conv_3d_transpose()` Transposed 3D (deconvolution)

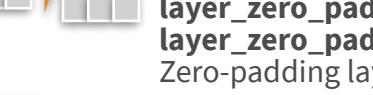
`layer_conv_3d()` 3D, e.g. spatial convolution over volumes



`layer_conv_lstm_2d()` Convolutional LSTM



`layer_separable_conv_2d()` Depthwise separable 2D



`layer_upsampling_1d()`  
`layer_upsampling_2d()`  
`layer_upsampling_3d()` Upsampling layer

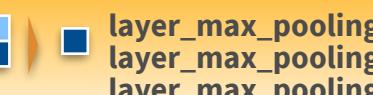


`layer_zero_padding_1d()`  
`layer_zero_padding_2d()`  
`layer_zero_padding_3d()` Zero-padding layer



`layer_cropping_1d()`  
`layer_cropping_2d()`  
`layer_cropping_3d()` Cropping layer

### POOLING LAYERS



`layer_max_pooling_1d()`  
`layer_max_pooling_2d()`  
`layer_max_pooling_3d()` Maximum pooling for 1D to 3D



`layer_average_pooling_1d()`  
`layer_average_pooling_2d()`  
`layer_average_pooling_3d()` Average pooling for 1D to 3D



`layer_global_max_pooling_1d()`  
`layer_global_max_pooling_2d()`  
`layer_global_max_pooling_3d()` Global maximum pooling



`layer_global_average_pooling_1d()`  
`layer_global_average_pooling_2d()`  
`layer_global_average_pooling_3d()` Global average pooling

## Preprocessing

### IMAGE PREPROCESSING

#### *Load Images*

`image_dataset_from_directory()`  
Create a TF Dataset from image files in a directory.

`image_load()`, `image_from_array()`,  
`image_to_array()`, `image_array_save()`

Work with PIL Image instances

#### *Transform Images*

`op_image_crop()`  
`op_image_extract_patches()`  
`op_image_pad()`  
`op_image_resize()`  
`op_image_affine_transform()`  
`op_image_map_coordinates()`  
`op_image_rgb_to_grayscale()`  
Operations that transform image tensors in deterministic ways.

#### `image_smart_resize()`

Resize images without aspect ratio distortion.

#### *Image Layers*

Builtin image preprocessing layers. Note, any image operation function can also be used as a layer in a Model, or used in `layer_lambda()`.

#### *Image Preprocessing Layers*

`layer_resizing()`  
`layer_rescaling()`  
`layer_center_crop()`

#### *Image Augmentation Layers*

Preprocessing layers that randomly augment image inputs during training.

`layer_random_crop()`  
`layer_random_flip()`  
`layer_random_translation()`  
`layer_random_rotation()`  
`layer_random_zoom()`  
`layer_random_contrast()`  
`layer_random_brightness()`

### SEQUENCE PREPROCESSING

#### `timeseries_dataset_from_array()`

Generate a TF Dataset of sliding windows over a timeseries provided as array.

#### `audio_dataset_from_directory()`

Generate a TF Dataset from audio files.

#### `pad_sequences()`

Pad sequences to the same length

## Preprocessing

### TEXT PREPROCESSING

#### `text_dataset_from_directory()`

Generate a TF Dataset from text files in a directory.

`layer_text_vectorization()`,  
`get_vocabulary()`, `set_vocabulary()`

Map text to integer sequences.

### NUMERICAL FEATURES PREPROCESSING

#### `layer_normalization()`

Normalizes continuous features.

#### `layer_discretization()`

Buckets continuous features by ranges.

### CATEGORICAL FEATURES PREPROCESSING

#### `layer_category_encoding()`

Encode integer features.

#### `layer_hashing()`

Hash and bin categorical features.

#### `layer_hashed_crossing()`

Cross features using the "hashing trick".

#### `layer_string_lookup()`

Map strings to (possibly encoded) indices.

#### `layer_integer_lookup()`

Map integers to (possibly encoded) indices.

### TABULAR DATA

One-stop utility for preprocessing and encoding structured data. Define a feature space from a list of table columns (features).

`feature_space <- layer_feature_space(features = list(<features>))`

Adapt the feature space to a dataset

`adapt(feature_space, dataset)`

Use the adapted `feature_space` preprocessing layer as a layer in a Keras Model, or in the data input pipeline with `tfdatasets::dataset_map()`

Available features:

`feature_float()`  
`feature_float_rescaled()`  
`feature_float_normalized()`  
`feature_float_discretized()`

`feature_integer_categorical()`  
`feature_integer_hashed()`

`feature_string_categorical()`  
`feature_string_hashed()`

`feature_cross()`  
`feature_custom()`

## Pre-trained models

Keras applications are deep learning models that are made available with pre-trained weights. These models can be used for prediction, feature extraction, and fine-tuning.

`application_mobilenet_v3_large()`  
`application_mobilenet_v3_small()`

MobileNetV3 Model, pre-trained on ImageNet

`application_efficientnet_v2s()`  
`application_efficientnet_v2m()`  
`application_efficientnet_v2l()`

EfficientNetV2 Model, pre-trained on ImageNet

`application_inception_resnet_v2()`  
`application_inception_v3()`

Inception-ResNet v2 and v3 model, with weights trained on ImageNet

`application_vgg16(); application_vgg19()`

VGG16 and VGG19 models

`application_resnet50()` ResNet50 model

`application_nasnet_large()`  
`application_nasnet_mobile()`

NASNet model architecture

### IMAGENET

`ImageNet` is a large database of images with labels, extensively used for deep learning

`application_preprocess_inputs()`  
`application_decode_predictions()`

Preprocesses a tensor encoding a batch of images for an application, and decodes predictions from an application

## Callbacks

A callback is a set of functions to be applied at given stages of the training procedure. You can use callbacks to get a view on internal states and statistics of the model during training.

`callback_early_stopping()` Stop training when a monitored quantity has stopped improving

`callback_learning_rate_scheduler()` Learning rate scheduler

`callback_tensorboard()` TensorBoard basic visualizations



# Labelled data with labelled :: CHEAT SHEET

The **labelled** package provides a set of functions and methods to handle and to manipulate labelled data, as imported with **haven** package.

## Basics

Labelled data is a common data structure in other statistical environment such as Stata, SAS or SPSS.

It consists of a set of additional attributes for numeric and character vectors (including columns of a data frame).

There are 3 types of attributes:

1. **Variable labels** (a short description of a variable)
2. **Value labels** (labels associated to specific values)
3. **Missing values**:
  - User-defined missing values (SPSS style)
  - Tagged NA (Stata and SAS style)

## Variable labels

### MANIPULATING A VECTOR

- var\_label(x) or var\_label(df\$v1)**  
Get the variable label associated to a vector x
- var\_label(x) <- "variable description"**  
Add/modify a variable label to x
- var\_label(x) <- NULL**  
Remove the variable label associated to x

### MANIPULATING A DATA.FRAME

- var\_label(df)**  
List all variable labels associated with columns of df
- var\_label(df) <- list(v1 = "variable 1", v2 = "variable 2")**  
Update variable labels of some columns of df
- df %>% set\_variable\_labels(v1 = "variable 1", v2 = "variable 2", v3 = NULL)**  
Update variable labels using dplyr syntax
- df %>% look\_for()**  
Return a data frame with all variable names and labels
- df %>% look\_for("s")**  
Search variables containing "s" in their name or label
- df %>% look\_for(details = TRUE)**  
Return additional details on each variable

## Value labels

When value labels are attached to a numeric or character vector, the vector's class becomes **haven\_labelled**. A major difference with a factor is that values of the vector are not changed and it is not mandatory to attach a label to each value.

### MANIPULATING A VECTOR

- val\_label(x, value) or val\_label(df\$v1, value)**  
Get the label attached to a specific value of a vector
- val\_label(x, value) <- "label"**  
Set/Update the label attached to a specific value
- val\_label(x, value) <- NULL**  
Remove the label attached to a specific value
- val\_labels(x)**  
Get all value labels attached to a vector
- val\_labels(x) <- c(no = 0, yes = 1, maybe = 9)**  
Set/Update all value labels attached to a vector
- val\_labels(x) <- NULL**  
Remove all value labels attached to a vector
- labelled(c("F", "F", "M"), c(Female = "F", Male = "M"))**  
Create a labelled vector
- sort\_val\_labels(x, according\_to = "values")**  
Sort value labels according to values (or labels)
- drop\_unused\_value\_labels(x)**  
Remove value labels not observed in the data

### MANIPULATING A DATA.FRAME

- df %>% set\_value\_labels(v1 = c(Yes = 1, No = 2), v2 = c(Male = "M", Female = "F"))**  
Define value labels of several variables
- df %>% add\_value\_labels(v1 = c(Unknown = 9))**  
Add specific value labels to a variable (other already defined value labels remains unchanged)
- df %>% remove\_value\_labels(v1 = 9)**  
Remove specific value labels to a variable
- df %>% set\_value\_labels(v1 = NULL)**  
Remove all value labels attached to a variable
- df %>% drop\_unused\_value\_labels()**  
Remove value labels not observed in the data

## Missing values

### USER-DEFINED MISSING VALUES (SPSS STYLE)

Used to indicate that some values should be considered as missing. However, they will not be treated as NA as long as they are not converted to proper NA.

When missing values are attached to a numeric or character vector, the vector's class becomes **haven\_labelled\_spss**.

When importing a SPSS file, use the option *user\_na = TRUE* to keep defined missing values (otherwise, they will be converted to NA).

### na\_values(x)

Get individual missing values attached to a vector

### na\_values(x) <- c(8, 9, 10)

df %>% set\_na\_values(v1 = c(8, 9, 10))  
Set/Update individual missing values (NULL to remove)

### na\_range(x)

Get a range of missing values attached to a vector

### na\_range(x) <- c(8, 10)

df %>% set\_na\_range(v1 = c(8, 10))  
Set/Update a range of missing values (NULL to remove)

### user\_na\_to\_na(x) or df %>% user\_na\_to\_na()

Convert user-defined missing values to NA

### is\_na(x)

TRUE if NA or if a user-defined missing value

### TAGGED NAs (STATA & SAS STYLE)

"Tagged" missing values work exactly like regular R missing values except that they store one additional byte of information: a tag, which is usually a letter ("a" to "z").

x <- c(1:5, tagged\_na("a"), tagged\_na("z"), NA)

tagged\_na("a") generates a NA with a tag

### is.na(x)

Tagged NAs work identically to regular NAs

### is\_tagged\_na(x)

Test if it is a tagged NA

### na\_tag(x)

Display the tags associated to tagged NAs

### format\_tagged\_na(x)

Convert x to a character vector showing the tagged NAs



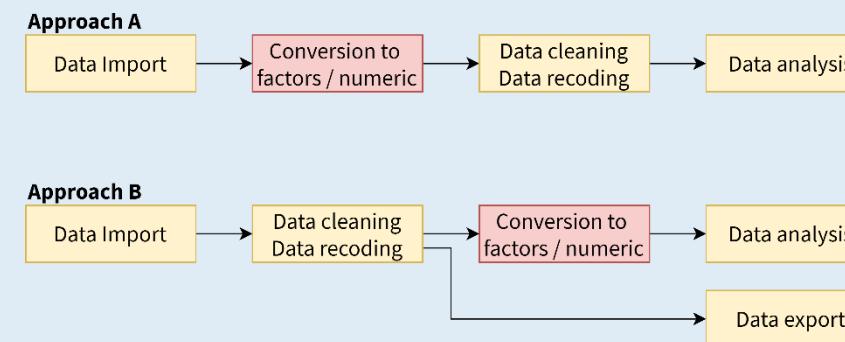
# When using labelled data?

`haven_labelled` and `haven_labelled_spss` classes introduced in **haven** package allow to add metadata (variable labels, value labels and SPSS-style missing values) to vectors / data frame columns and to properly import these metadata from SAS, Stata or SPSS.

Functions and methods provided by **labelled** package are designed for easy manipulation of such labelled data.

It should be noted that **value labels** doesn't imply that your vectors should be considered as categorical or continuous. Therefore, value labels are not intended to be used for data analysis. For example, before performing modeling or plotting, you should convert vectors with value labels into factors or into classic numeric/character vectors.

Two main approaches could be considered:



In **approach A**, labelled vectors are converted into factors or into numeric/character vectors just after data import, using **unlabelled()**, **to\_factor()** or **unclass()**. Then, data cleaning, data recoding and analysis are performed using classic R vector types.

In **approach B**, labelled vectors are kept for data cleaning and recoding, allowing to preserve original coding, in particular if data should be reexported after that step. Functions provided by **labelled** will be useful for managing value labels.

However, as in approach A, labelled vectors will have to be converted into classic factors or numeric vectors before data analysis as this is the way categorical and continuous variables should be coded for analysis.

# Conversion

## OF LABELLED VECTORS

If all value labels and user-defined missing values are removed from a labelled vector, the `haven_labelled` class will be removed and the vector will be transformed into a basic numeric or character vector. **Values of the vector will remain unchanged.**

### `remove_val_labels(x)`

Remove value labels attached to a vector.

### `remove_user_na(x)`

Remove user-defined (`na_values` and `na_range`) from a vector

### `unclass(x)`

Remove the `haven_labelled` class. Therefore, the vector will be considered as a classical numeric or character vector. Value labels and user-defined missing values will still be visible as attributes attached to the vector.

When converting a labelled vector into a factor or a character vector of the value labels, be aware that **original values of the vector will be converted.**

### `to_character(x)`

Convert into a character vector replacing values by their corresponding value label

### `to_factor(x)`

Convert into a character vector replacing values by their corresponding value label

### `to_factor(x, levels = "prefixed")`

Value labels will be prefixed with their original value

### `to_factor(x, strict = TRUE)`

Convert into a factor only if all observed values have a value label

### `df %>% to_factor()`

Convert all labelled vectors and only labelled vectors into factors

### `df %>% to_factor(labelled_only = FALSE)`

Convert all columns (including non labelled vectors) into factors

### `unlabelled(x)`

#### `df %>% unlabelled()`

Labelled vectors will be converted into factors only if all observed values have a value label. Otherwise, they will be unclassed. Similar to `df %>% to_factor(labelled_only = T, strict = T, unclass = T)`

### `to_factor(x, drop_unused_labels = TRUE)`

#### `df %>% unlabelled(drop_unused_labels = TRUE)`

Unused value labels will be dropped before conversion into factors

## INTO LABELLED VECTORS

### `to_labelled(f)`

Convert a factor into a numeric labelled vector. Note that `to_labelled(to_factor(x))` and `x` will not be identical (original coding will be lost).

### `to_labelled(df)`

If `df` was imported with the **foreign** package or if it is a data set created with **memisc** package, meta data (variable labels, value labels and user-defined missing values) will be converted into **labelled** format.

If any value label or user-defined missing value is added to a numeric or a character vector, it will be automatically converted into a labelled vector.

**Values of the vector will remain unchanged.**

## Miscellaneous

### `nolabel_to_na(x)` or `df %>% nolabel_to_na()`

For labelled vectors, values without a value label will be converted into NA

### `val_labels_to_na(x)` or `df %>% val_labels_to_na()`

For labelled vectors, values with a value label will be converted into NA

### `df2 %>% copy_labels_from(df1)`

Copy variable labels, values labels and user-defined missing values from `df1` to `df2` based on shared columns names. Useful when attributes are lost after some data manipulation.

### `recode(x, `2` = 1, `3` = 2)`

Apply `dplyr::recode()` to a labelled vector. Attached value labels will remain unchanged.

### `recode(x, `2` = 1, .combine_value_labels = TRUE)`

This option will combine value labels of original values merged together to produce new value labels. It is recommended to check that the result is appropriate.

### `update_labelled(x)` or `df %>% update_labelled()`

If `x/df` was imported/created using an older version of **haven** or **labelled**, you may encounter some unexpected results. `update_labelled()` will update all labelled vectors to be consistent with the current implementation.

# Leaflet Cheat Sheet



an open-source JavaScript library for mobile-friendly interactive maps

## Quick Start

### Installation

Use `install.packages("leaflet")` to install the package or directly from Github `devtools::install_github("rstudio/leaflet")`.

### First Map

```
m <- leaflet() %>%
  addTiles() %>%
  addMarkers(lng = 174.768, lat = -36.852, popup = "The birthplace of R")
# add a single point layer
```



## Map Widget

### Initialization

|                                                             |                                      |
|-------------------------------------------------------------|--------------------------------------|
| <code>m &lt;- leaflet(options = leafletOptions(...))</code> | Initial geographic center of the map |
| <code>center</code>                                         | Initial map zoom level               |
| <code>zoom</code>                                           | Minimum zoom level of the map        |
| <code>minZoom</code>                                        | Maximum zoom level of the map        |
| <code>maxZoom</code>                                        |                                      |

### Map Methods

```
m %>% setView(lng, lat, zoom, options = list())
# Set the view of the map (center and zoom level)
m %>% fitBounds(lng1, lat1, lng2, lat2)
# Fit the view into the rectangle [lng1, lat1] - [lng2, lat2]
m %>% clearBounds()
# Clear the bound, automatically determine from the map elements
```

### Data Object

Both `leaflet()` and the `map` layers have an optional data parameter that is designed to receive spatial data with the following formats:

#### Base R

The arguments of all layers take normal R objects:

```
df <- data.frame(lat = ..., lng = ...)
```

```
leaflet(df) %>% addTiles() %>% addCircles()
```

library(sp) Useful functions:

SpatialPoints, SpatialLines, SpatialPolygons, ...

library(maps) Build a map of states with colors:

```
mapStates <- map("state", fill = TRUE, plot = FALSE)
```

```
leaflet(mapStates) %>% addTiles() %>%
```

```
addPolygons(fillColor = topo.colors(10, alpha = NULL), stroke = FALSE)
```

## Markers

Use markers to call out points, express locations with latitude/longitude coordinates, appear as icons or as circles.

Data come from vectors or assigned data frame, or `sp` package objects.

### Icon Markers

Regular Icons: default and simple

```
addMarkers(lng, lat, popup, label) add basic icon markers
```

```
makeIcon(Icons(iconUrl, iconWidth, iconHeight, iconAnchorX, iconAnchorY,
  shadowUrl, shadowWidth, shadowHeight, ...)) customize marker icons
```

```
iconList() create a list of icons
```

Awesome Icons: customizable with colors and icons

```
addAwesomeMarkers, makeAwesomeIcon, awesomeIcons, awesomeIconList
```

Marker Clusters: option of `addMarkers()`

```
clusterOptions = markerClusterOptions()
```

```
freezeAtZoom Freeze the cluster at assigned zoom level
```

### Circle Markers

```
addCircleMarkers(color, radius, stroke, opacity, ...)
```

Customize their color, radius, stroke, opacity

## Popups and Labels

`addPopups(lng, lat, ...content..., options)` Add standalone popups

```
options = popupOptions(closeButton=FALSE)
```

`addMarkers(..., popup, ...)` Show popups with markers or shapes

`addMarkers(..., label, labelOptions...)` Show labels with markers or shapes

```
labelOptions = labelOptions(noHide, textOnly, textSize, direction, style)
```

`addLabelOnlyMarkers()` Add labels without markers

## Lines and Shapes

### Polygons and Polylines

`addPolygons(color, weight=1, smoothFactor=0.5, opacity=1.0, fillOpacity=0.5,`  
`fillColor= ~colorQuantile("YlOrRd", ALAND)(ALAND), highlightOptions, ... )`

`highlightOptions(color, weight=2, bringToFront=TRUE)` highlight shapes

Use `rmapshaper::ms_simplify` to simplify complex shapes

`Circles addCircles(lng, lat, weight=1, radius, ...)`

`Rectangles addRectangles(lng1, lat1, lng2, lat2, fillColor="transparent", ... )`

## Basemaps

### Default Tiles

`addTiles()`

### Third-Party Tiles

`providers$Stamen.Toner, CartoDB.Positron, Esri.NatGeoWorldMap`

`addProviderTiles()`

Default Tiles  
Use `addTiles()` to add a custom map tile URL template, use `addWMSTiles()` to add WMS (Web Map Service) tiles

## GeoJSON and TopoJSON

There are two options to use the GeoJSON/TopoJSON data:

- \* To read into `sp` objects with the `geojsonio` or `rgdal` package:  
`geojsonio::geojson_read(..., what="sp") rgdal::readOGR(..., "OGRGeoJSON")`

- \* Or to use the `addGeoJSON()` and `addTopoJSON()` functions:  
`addTopoJSON/addGeoJSON(... weight, color, fill, opacity, fillOpacity...)`

Styles can also be tuned separately with a `style: {}` object.

Other packages including `RJSONIO` and `jsonlite` can help fast parse or generate the data needed.

## Shiny Integration

To integrate a Leaflet map into an app:

- \* In the UI, call `leafletOutput("name")`

- \* On the server side, assign a `renderLeaflet(...)` call to the output

- \* Inside the `renderLeaflet` expression, return a Leaflet map object

### Modification

To modify an existing map or add incremental changes to the map, you can use `leafletProxy()`. This should be performed in an observer on the server side.

Other useful functions to edit your map:

`fitBounds(o, 0, 11, 11)` similar to `setView`

fit the view to within these bounds

`addCircles(1:10, 1:10, layerId = LETTERS[1:10])`

create circles with layerIds of "A", "B", "C"...

`removeShape(c("B", "F"))` remove some of the circles

`clearShapes()` clear all circles (and other shapes)

### Inputs/Events

#### Object Events

Object event names generally use this pattern:

`inputs$MAPID_OBJCATEGORY_EVENTNAME`.

Triger an event changes the value of the Shiny input at this variable.

Valid values for `OBJCATEGORY` are `marker`, `shape`, `geojson` and `topojson`.

Valid values for `EVENTNAME` are `click`, `mouseover` and `mouseout`.

All of these events are set to either `NULL` if the event has never happened, or a `list()` that includes:

- \* `lat` The latitude of the object, if available; otherwise, the mouse cursor

- \* `lng` The longitude of the object, if available; otherwise, the mouse cursor

- \* `id` The layerId, if any

GeoJSON events also include additional properties:

- \* `featureId` The feature ID, if any

- \* `properties` The feature properties

#### Map Events

`inputs$MAPID_click`

when the map background or basemap is clicked  
value -- a list with lat and lng

`inputs$MAPID_bounds`

provide the lat/long bounds of the visible map area  
value -- a list with north, east, south and west

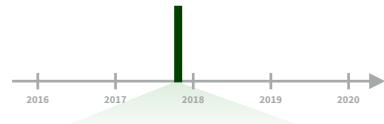
`inputs$MAPID_zoom`

an integer indicates the zoom level

# Dates and times with lubridate :: CHEATSHEET



## Date-times



2017-11-28 12:00:00

2017-11-28 12:00:00

A **date-time** is a point on the timeline, stored as the number of seconds since 1970-01-01 00:00:00 UTC

```
dt <- as_datetime(1511870400)
## "2017-11-28 12:00:00 UTC"
```

### PARSE DATE-TIMES (Convert strings or numbers to date-times)

1. Identify the order of the year (**y**), month (**m**), day (**d**), hour (**h**), minute (**m**) and second (**s**) elements in your data.
2. Use the function below whose name replicates the order. Each accepts a tz argument to set the time zone, e.g. ymd(x, tz = "UTC").

2017-11-28T14:02:00

ymd\_hms(), ymd\_hm(), ymd\_h().  
ymd\_hms("2017-11-28T14:02:00")

2017-22-12 10:00:00

ydm\_hms(), ydm\_hm(), ydm\_h().  
ydm\_hms("2017-22-12 10:00:00")

11/28/2017 1:02:03

mdy\_hms(), mdy\_hm(), mdy\_h().  
mdy\_hms("11/28/2017 1:02:03")

1 Jan 2017 23:59:59

dmy\_hms(), dmy\_hm(), dmy\_h().  
dmy\_hms("1 Jan 2017 23:59:59")

20170131

ymd(), ydm(). ymd(20170131)

July 4th, 2000

mdy(), myd(). mdy("July 4th, 2000")

4th of July '99

dmy(), dym(). dmy("4th of July '99")

2001: Q3

yq() Q for quarter. yq("2001: Q3")

07-2020

my(), ym(). my("07-2020")

2:01

hms::hms() Also lubridate::hms(), hm() and ms(), which return periods.\* hms::hms(seconds = 0, minutes = 1, hours = 2)

2017.5

date\_decimal(decimal, tz = "UTC")  
date\_decimal(2017.5)

now(zone = "") Current time in tz (defaults to system tz). now()

today(zone = "") Current date in a tz (defaults to system tz). today()

fast.strptime() Faster strftime.

fast.strptime("9/1/01", "%y/%m/%d")

parse\_date\_time() Easier strftime.

parse\_date\_time("09-01-01", "ymd")



2017-11-28

A **date** is a day stored as the number of days since 1970-01-01

```
d <- as_date(17498)
## "2017-11-28"
```

### GET AND SET COMPONENTS

Use an accessor function to get a component. Assign into an accessor function to change a component in place.

2018-01-31 11:59:59

**date(x)** Date component. date(dt)

**year(x)** Year. year(dt)  
**isoyear(x)** The ISO 8601 year.  
**epiyear(x)** Epidemiological year.

2018-01-31 11:59:59

**month(x, label, abbr)** Month. month(dt)

2018-01-31 11:59:59

**day(x)** Day of month. day(dt)  
**wday(x, label, abbr)** Day of week.  
**qday(x)** Day of quarter.

2018-01-31 11:59:59

**hour(x)** Hour. hour(dt)

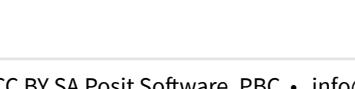
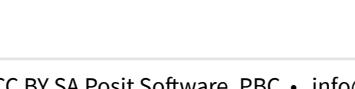
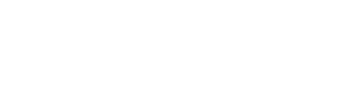
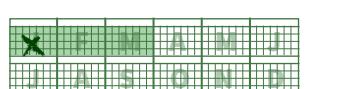
2018-01-31 11:59:59

**minute(x)** Minutes. minute(dt)

2018-01-31 11:59:59

**second(x)** Seconds. second(dt)

2018-01-31 11:59:59 UTC

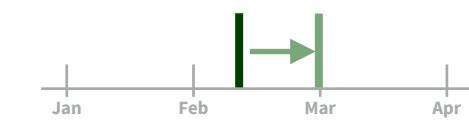
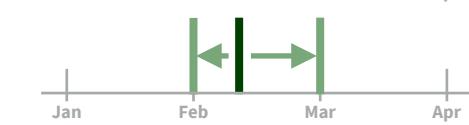
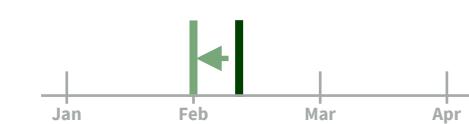


12:00:00

An hms is a **time** stored as the number of seconds since 00:00:00

```
t <- hms::as_hms(85)
## 00:01:25
```

## Round Date-times



Valid units are second, minute, hour, day, week, month, bimonth, quarter, season, halfyear and year.

**rollback(dates, roll\_to\_first = FALSE, preserve\_hms = TRUE)** Roll back to last day of previous month. Also **rollforward()**. rollback(dt)

## Stamp Date-times

**stamp()** Derive a template from an example string and return a new function that will apply the template to date-times. Also **stamp\_date()** and **stamp\_time()**.

1. Derive a template, create a function  
sf <- stamp("Created Sunday, Jan 17, 1999 3:34")

2. Apply the template to dates  
sf(ymd("2010-04-05"))
## [1] "Created Monday, Apr 05, 2010 00:00"

**Tip:** use a date with day > 12

## Time Zones

R recognizes ~600 time zones. Each encodes the time zone, Daylight Savings Time, and historical calendar variations for an area. R assigns one time zone per vector.

Use the **UTC** time zone to avoid Daylight Savings.

**OlsonNames()** Returns a list of valid time zone names. OlsonNames()

**Sys.timezone()** Gets current time zone.



**with\_tz(time, tzzone = "")** Get the same date-time in a new time zone (a new clock time). Also **local\_time(dt, tz, units)**. with\_tz(dt, "US/Pacific")

**force\_tz(time, tzzone = "")** Get the same clock time in a new time zone (a new date-time). Also **force\_tzs()**. force\_tz(dt, "US/Pacific")



# Math with Date-times

Math with date-times relies on the **timeline**, which behaves inconsistently. Consider how the timeline behaves during:

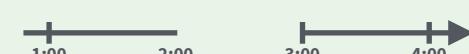
A normal day

```
nor <- ymd_hms("2018-01-01 01:30:00", tz="US/Eastern")
```



The start of daylight savings (spring forward)

```
gap <- ymd_hms("2018-03-11 01:30:00", tz="US/Eastern")
```



The end of daylight savings (fall back)

```
lap <- ymd_hms("2018-11-04 00:30:00", tz="US/Eastern")
```



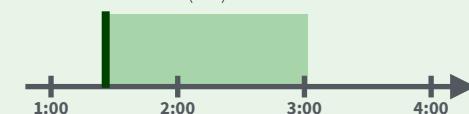
Leap years and leap seconds

```
leap <- ymd("2019-03-01")
```

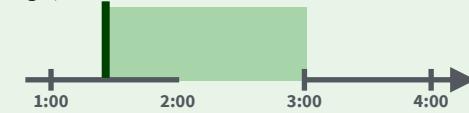


**Periods** track changes in clock times, which ignore time line irregularities.

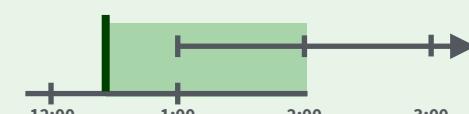
nor + minutes(90)



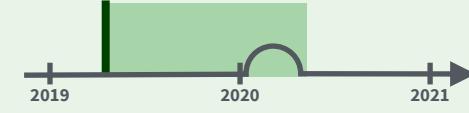
gap + minutes(90)



lap + minutes(90)

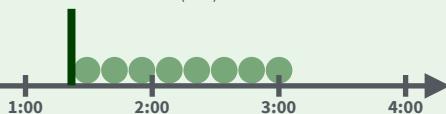


leap + years(1)



**Durations** track the passage of physical time, which deviates from clock time when irregularities occur.

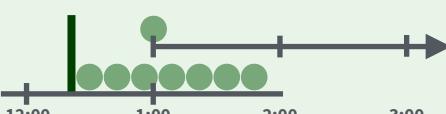
nor + dminutes(90)



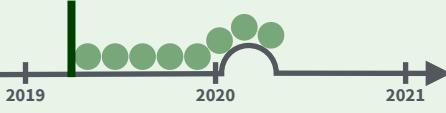
gap + dminutes(90)



lap + dminutes(90)



leap + dyears(1)



**Intervals** represent specific intervals of the timeline, bounded by start and end date-times.

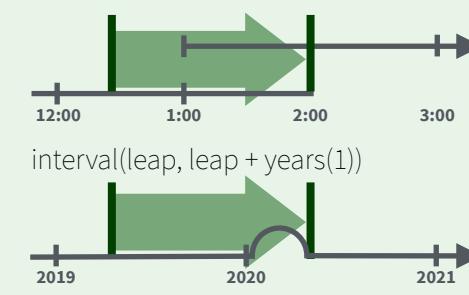
interval(nor, nor + minutes(90))



interval(gap, gap + minutes(90))



interval(lap, lap + minutes(90))



Not all years are 365 days due to **leap days**.

Not all minutes are 60 seconds due to **leap seconds**.

It is possible to create an imaginary date by adding **months**, e.g. February 31st

```
jan31 <- ymd(20180131)
```

```
jan31 + months(1)
```

```
## NA
```

%m+% and %m-% will roll imaginary dates to the last day of the previous month.

```
jan31 %m+% months(1)
```

```
## "2018-02-28"
```

**add\_with\_rollback**(e1, e2, roll\_to\_first = TRUE) will roll imaginary dates to the first day of the new month.

```
add_with_rollback(jan31, months(1), roll_to_first = TRUE)
```

```
## "2018-03-01"
```

## PERIODS

Add or subtract periods to model events that happen at specific clock times, like the NYSE opening bell.

Make a period with the name of a time unit **pluralized**, e.g.

```
p <- months(3) + days(12)
```

```
p
```

```
"3m 12d 0H 0M 0S"
```

Number of months Number of days etc.

**years(x = 1)** x years.

**months(x)** x months.

**weeks(x = 1)** x weeks.

**days(x = 1)** x days.

**hours(x = 1)** x hours.

**minutes(x = 1)** x minutes.

**seconds(x = 1)** x seconds.

**milliseconds(x = 1)** x milliseconds.

**microseconds(x = 1)** x microseconds.

**nanoseconds(x = 1)** x nanoseconds.

**picoseconds(x = 1)** x picoseconds.

**period(num = NULL, units = "second", ...)**

An automation friendly period constructor.  
period(5, unit = "years")

**as.period(x, unit)** Coerce a timespan to a period, optionally in the specified units. Also **is.period()**. as.period(p)

**period\_to\_seconds(x)** Convert a period to the "standard" number of seconds implied by the period. Also **seconds\_to\_period()**. period\_to\_seconds(p)

## DURATIONS

Add or subtract durations to model physical processes, like battery life. Durations are stored as seconds, the only time unit with a consistent length.

**Diftimes** are a class of durations found in base R.

Make a duration with the name of a period prefixed with a **d**, e.g.

```
dd <- ddays(14)
```

```
dd
```

```
"1209600s (~2 weeks)"
```

Exact length in seconds Equivalent in common units

**dyears(x = 1)** 31536000x seconds.

**dmonths(x = 1)** 2629800x seconds.

**dweeks(x = 1)** 604800x seconds.

**ddays(x = 1)** 86400x seconds.

**dhours(x = 1)** 3600x seconds.

**dminutes(x = 1)** 60x seconds.

**dseconds(x = 1)** x seconds.

**dmilliseconds(x = 1)** x × 10<sup>-3</sup> seconds.

**dmicroseconds(x = 1)** x × 10<sup>-6</sup> seconds.

**dnanoseconds(x = 1)** x × 10<sup>-9</sup> seconds.

**dpicoseconds(x = 1)** x × 10<sup>-12</sup> seconds.

**duration(num = NULL, units = "second", ...)**

An automation friendly duration constructor. duration(5, unit = "years")

**as.duration(x, ...)** Coerce a timespan to a duration. Also **is.duration()**, **is.difftime()**. as.duration(i)

**make\_difftime(x)** Make difftime with the specified number of units. make\_difftime(99999)

## INTERVALS

Divide an interval by a duration to determine its physical length, divide an interval by a period to determine its implied length in clock time.

Make an interval with **interval()** or %--%, e.g.

```
i <- interval(ymd("2017-01-01"), d)
```

```
## 2017-01-01 UTC--2017-11-28 UTC
```

```
j <- d %--% ymd("2017-12-31")
```

```
## 2017-11-28 UTC--2017-12-31 UTC
```

- %within%** b Does interval or date-time a fall within interval b? now() %within% i
- int\_start(int)** Access/set the start date-time of an interval. Also **int\_end()**. int\_start(i) <- now(); int\_start(i)
- int\_aligns(int1, int2)** Do two intervals share a boundary? Also **int\_overlaps()**. int\_aligns(i, j)
- int\_diff(times)** Make the intervals that occur between the date-times in a vector. v <- c(dt, dt + 100, dt + 1000); int\_diff(v)
- int\_flip(int)** Reverse the direction of an interval. Also **int\_standardize()**. int\_flip(i)
- int\_length(int)** Length in seconds. int\_length(i)
- int\_shift(int, by)** Shifts an interval up or down the timeline by a timespan. int\_shift(i, days(-1))
- as.interval(x, start, ...)** Coerce a timespan to an interval with the start date-time. Also **is.interval()**. as.interval(days(1), start = now())

# Machine Learning Modelling in R :: CHEAT SHEET

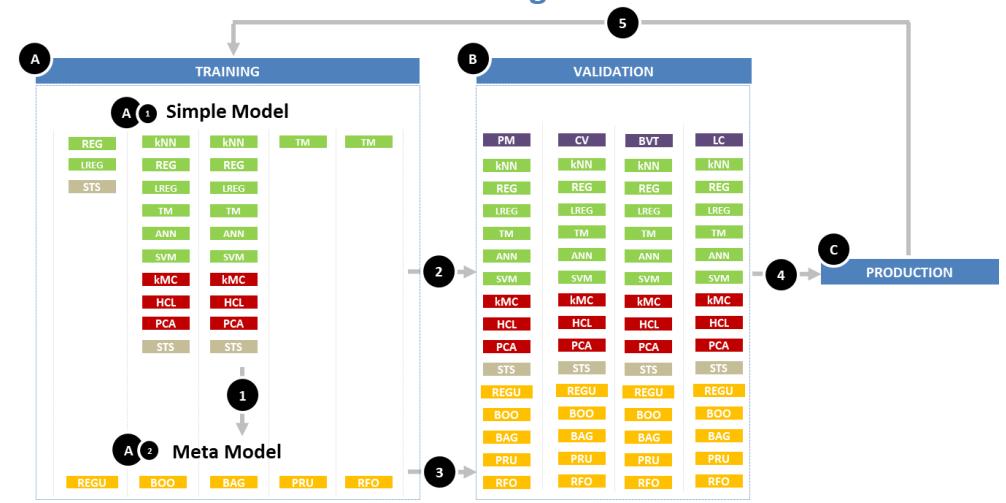
## Supervised & Unsupervised Learning

| ALGORITHM                           | DESCRIPTION                                                                                                                                                                                                                                                                 | R PACKAGE::FUNCTION                                                                | SAMPLE CODE                                                                                                                                                                                                                                                                         |
|-------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| NBC<br>Naïve Bayes classifier       | A classification technique based on Bayes' Theorem with an assumption of independence among predictors. In simple terms, a Naïve Bayes classifier assumes that the presence or absence of a particular feature in a class is unrelated to the presence of any other feature | e1071::naiveBayes                                                                  | naiveBayes(class ~ ., data = x)                                                                                                                                                                                                                                                     |
| kNN<br>k-Nearest Neighbours         | A non-parametric method used for classification and regression. In both cases, the input consists of the k closest training examples in the feature space. The output depends on whether k-NN is used for classification or regression                                      | class::knn                                                                         | knn(train, test, cl, k = 1, l = 0, prob = FALSE, use.all = TRUE)                                                                                                                                                                                                                    |
| LRG<br>Linear Regression            | Model the linear relationship between a scalar dependent variable Y and one or more explanatory variables (or independent variables) denoted X                                                                                                                              | stats::lm                                                                          | lm(dist ~ speed, data=cars)                                                                                                                                                                                                                                                         |
| LRC<br>Logistic Regression          | Used to predict a binary outcome (1 / 0, Yes / No, True / False) given a set of independent variables.                                                                                                                                                                      | stats::glm                                                                         | glm(Y ~ ., family = binomial (link = 'logit'), data = X)                                                                                                                                                                                                                            |
| TM<br>Tree-Based Models             | The idea is to consecutively divide (branch) the training data into smaller and smaller features until an assignment criterion with respect to the target variable into a "data bucket" (leaf) is reached                                                                   | rpart::rpart                                                                       | rpart(Kyphosis ~ Age + Number + Start, data = kyphosis)                                                                                                                                                                                                                             |
| ANN<br>Artificial Neural Network    | Neural networks are built from units called perceptrons. Perceptrons have one or more inputs, an activation function and an output. An ANN model is built up by combining perceptrons in structured layers.                                                                 | neuralnet::neuralnet                                                               | neuralnet(f,data=train_hidden=(5,3),linear.output=T)                                                                                                                                                                                                                                |
| SVM<br>Support Vector Machine       | A data classification method that separates data using hyperplanes                                                                                                                                                                                                          | e1071::svm                                                                         | svm(formula, data = NULL, ..., subset, na.action = na.omit, scale = TRUE)                                                                                                                                                                                                           |
| PCA<br>Principal Component Analysis | A procedure that uses an orthogonal transformation to convert a set of observations of possibly correlated variables into a set of values of linearly uncorrelated variables called principal components.                                                                   | stats::prcomp<br>stats::princomp<br>FactoMineR::PCA<br>ade4::dudi.pca<br>amap::acp | stats::prcomp(formula, data = NULL, subset, na.action, ...) stats::princomp(formula, data = NULL, subset, na.action, ...) FactoMineR::PCA(decatlon, quanti.sup = 11:12, quali.sup = 13) ade4::dudi.pca(deugStab, center = deugCent, scale = FALSE, scan = FALSE) amap::acp(lubisch) |
| HAC<br>k-Mean Clustering            | Aims at partitioning n observations into k clusters in which each observation belongs to the cluster with the nearest mean                                                                                                                                                  | stats::kmeans                                                                      | kmeans(k, centers, iter.max = 10, nstart = 1, algorithm = c("Hartigan-Wong", "Lloyd", "Forgy", "MacQueen"), trace = FALSE)                                                                                                                                                          |
| HCL<br>Hierarchical Clustering      | An approach which builds a hierarchy from the bottom-up, and doesn't require the number of clusters to be specified beforehand.                                                                                                                                             | stats::hclust                                                                      | hclust(d, method = "complete", members = NULL)                                                                                                                                                                                                                                      |

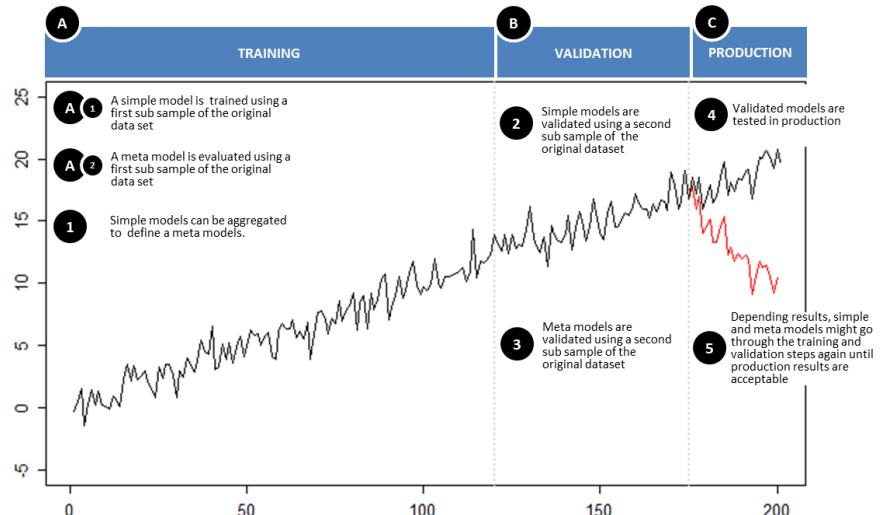
## Meta-Algorithm, Time Series & Model Validation

| ALGORITHM                                       | DESCRIPTION                                                                                                                                                                                                                                                           | R PACKAGE::FUNCTION                               | SAMPLE CODE                                                                                                                                                                                                                              |
|-------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| REGU<br>Regularisation L1 (Lasso)<br>L2 (Ridge) | Regularisation adds a penalty on the different parameters of a model to reduce the freedom of the model. Hence, the model will be less likely to fit the noise of the training data and will improve the generalization abilities of the model                        | glmnet::glmnet                                    | L1 : glmnet(myMatrixA, myMatrixB, family = "gaussian", alpha = 1)<br>L2 : glmnet(myMatrixA, myMatrixB, family = "gaussian", alpha = 0)                                                                                                   |
| BOO<br>Boosting                                 | A process of iteratively refining, e.g. by reweighting, of estimated regression and classification functions (though it has primarily been applied to the latter), in order to improve predictive ability.                                                            | gbm::gbm                                          | gbmboost(Y ~ ., data = curr1[trnidxs,])                                                                                                                                                                                                  |
| BAG<br>Bagging                                  | Bagging is a way to increase the power of a predictive statistical model by taking multiple random samples (with replacement) of the training data set, and using each of them to construct a separate model and separate predictions for the original test set       | randomForest::randomForest                        | foreach : d <- data.frame(x=1:10, y=rnorm(10)) s <- foreach(d=i %in% d, by=row, .combine=rbind, .id=i, .d = i) ipred : bagging(formula, data, subset, na.action=na.rpart, \dots)                                                         |
| PRU<br>Pruning                                  | Pruning is a technique that reduces the size of decision tree by removing sections of the tree that provide little power to classify instances. Pruning reduces the complexity of the final classifier and hence improves predictive accuracy by reducing overfitting | rpart::rpart                                      | prune(x, cp = 0.1)                                                                                                                                                                                                                       |
| RFO<br>Random Forest                            | An ensemble learning method for classification, regression and other tasks, that operate by constructing a multitude of decision trees at training time and outputting the class that is the mode of the classes (classification) or mean prediction (regression)     | randomForest::randomForest                        | randomForest(X ~ ., data = Y, subset = mySub)                                                                                                                                                                                            |
| STS<br>Time Series                              | Random sampling of observations for training and testing a model can be an issue when faced with a times dimension. Random sampling may either destroy serial correlation properties in the data which we would like to exploit                                       | stats::xts<br>forecast::spectral<br>spectral::TTR | Auto-correlation: acf(x, lag.max = NULL, type = c("correlation", "covariance", "partial")) Spectral Analysis: spec.pgram(..., spans = NULL) Seasonal Decomposition of Time Series : stl(x, s.window = 7, t.window = 50, t.jump = 1) .... |
| PM<br>Performance metrics                       | Depends on the problem:<br>• Regression: squared errors, outliers, error rate...<br>• Classification: Accuracy, precision, recall, F-score...                                                                                                                         | ROCR::ROC                                         | Regression:stats::outlierTest, stats::qqPlot ...<br>Classification:ROCR::ROC                                                                                                                                                             |
| JVT<br>Bias-Variance Tradeoff                   | • Simple models with few parameters are easier to compute but may lead to poorer fits ( <b>high bias</b> ).<br>• Complex models may provide more accurate fits but may over-fit the data ( <b>high variance</b> )                                                     | caret::confusionMatrix                            | Tailored to the analysis                                                                                                                                                                                                                 |
| CV<br>Cross validation                          | Cross validation compares the test performances of different model realisations with different sets or values of parameters                                                                                                                                           | caret::createDataPartition<br>caret::createFolds  | Tailored to the analysis                                                                                                                                                                                                                 |
| LC<br>Learning Curves                           | Learning curves plot a model's training and test errors, or the chosen performance metric, depending on the training set size                                                                                                                                         | caret::learning_curve_dat                         | createDataPartition(classes, p = 0.8, list = FALSE)<br>learning_curve_dat(dat, outcome = NULL, proportion = (1:10)/10, test_prop = 0, verbose = TRUE, ...)                                                                               |

## Standard Modelling Workflow



## Time Series View

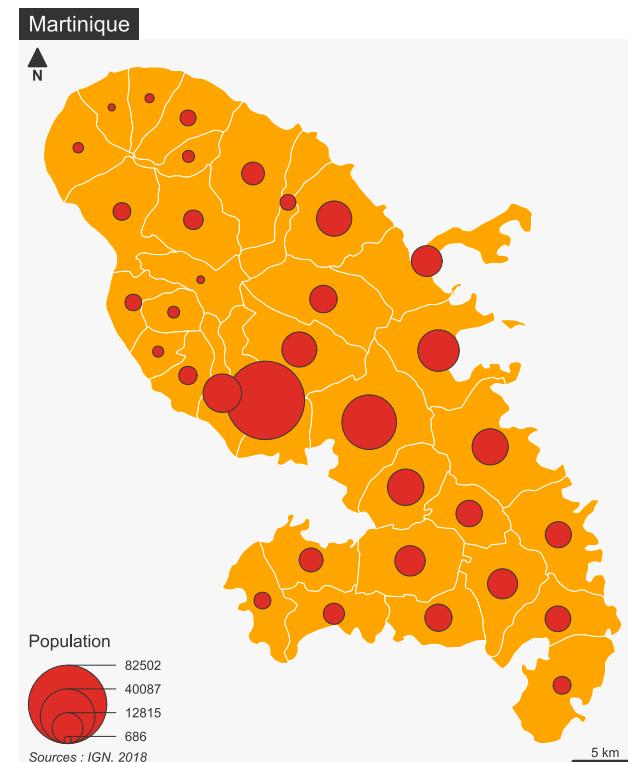


# Thematic maps with mapsf :: CHEAT SHEET

Create and integrate thematic maps in your workflow.

## Base map

```
Import library  
library(mapsf)  
  
Import the sample data set  
mtq <- mf_get_mtq()  
  
Initiate a base map centered on a specific extent  
mf_map(x = mtq, col = "orange",  
       border = "white")  
  
Plot symbology  
mf_map(x = mtq, type = "prop", var = "POP",  
       leg_title = "Population", add = TRUE)  
  
Complete layout (credits, title, north, arrow, scale bar)  
mf_layout(title = "Martinique",  
          credits = "Sources: IGN, 2018")
```



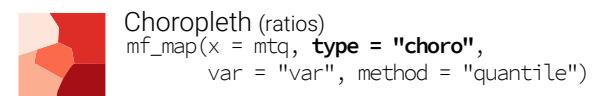
## Colors

mapsf can use color palettes from `hcl.colors()`.  
`mf_get_pal()` is useful to create well-balanced asymmetric diverging palettes

```
mf_get_pal(n = c(7, 2), pal = c("Burg", "Mint"))
```

## Symbology

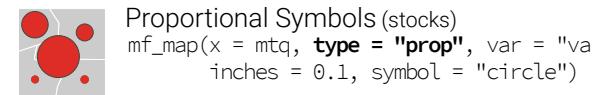
The x argument should be an sf object.  
Input geometries can be polygons, lines or points.



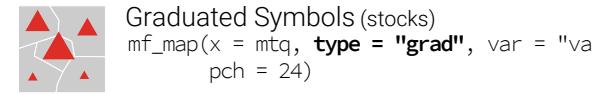
```
Choropleth (ratios)  
mf_map(x = mtq, type = "choro",  
       var = "var", method = "quantile")
```



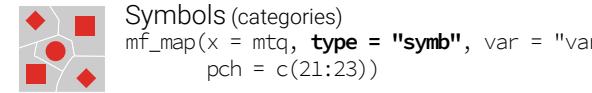
```
Typology (categories)  
mf_map(x = mtq, type = "typo", var = "var")
```



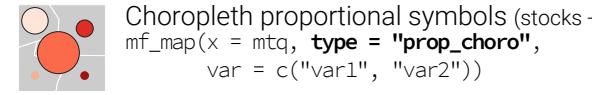
```
Proportional Symbols (stocks)  
mf_map(x = mtq, type = "prop", var = "var",  
       inches = 0.1, symbol = "circle")
```



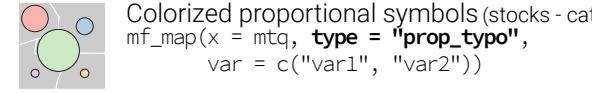
```
Graduated Symbols (stocks)  
mf_map(x = mtq, type = "grad", var = "var",  
       pch = 24)
```



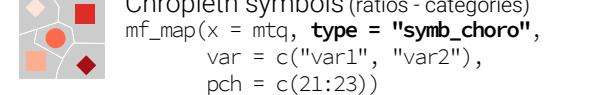
```
Symbols (categories)  
mf_map(x = mtq, type = "symb", var = "var",  
       pch = c(21:23))
```



```
Choropleth proportional symbols (stocks - ratios)  
mf_map(x = mtq, type = "prop_choro",  
       var = c("var1", "var2"))
```



```
Colorized proportional symbols (stocks - categor.)  
mf_map(x = mtq, type = "prop_typo",  
       var = c("var1", "var2"))
```



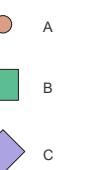
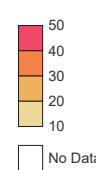
```
Choropleth symbols (ratios - categories)  
mf_map(x = mtq, type = "symb_choro",  
       var = c("var1", "var2"),  
       pch = c(21:23))
```



```
Raster  
mf_raster(x = raster)
```

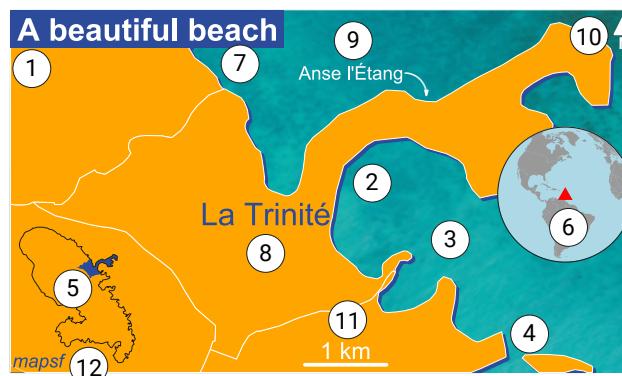
## Legends

Default legends are plotted along maps.  
Customization parameters are available with  
`mf_legend()`



## Map Layout

Along with cartographic functions, other functions are dedicated to customize the layout design.



① Set a map theme (figure margins, colors, title options...)  
`mf_theme(bg = "white", tab = TRUE,  
 mar = c(0,0,0,0), pos = "left")`

Builtin themes are available : default, ink, dark, agolalight, candy, darkula, iceberg, green, nevermind, jsk or barcelona.

② Init a map centered on a specific area  
`mf_init(x = mtq[30, ])`  
`mf_map(x = mtq, col = "orange",  
 border = "white", add = TRUE)`

③ Import external image for background  
`mf_background(filename = "img/sea.jpg")`

④ Create a shadow effect  
`mf_shadow(...)`

⑤ Create a custom inset  
`mf_inset_on(x = mtq, pos = "bottomleft")`  
`mf_map(...)`  
`mf_inset_off()`

⑥ Create a world inset  
`mf_inset_on(x = "worldmap", pos = "right")`  
`mf_worldmap(mtq)`  
`mf_inset_off()`

⑦ Plot title  
`mf_title("A beautiful beach")`

⑧ Plot labels  
`mf_label(...)`

⑨ Plot annotation (in specific locations)  
`mf_annotation(...)`

⑩ North arrow  
`mf_arrow(...)`

⑪ Scale (in km)  
`mf_scale(...)`

⑫ Credits  
`mf_credits(...)`



## Export Maps

`mf_export()` exports maps in PNG or SVG formats.

The exported map width/height ratio will match the one of a spatial object.

Additionally, `mf_export()` can be used to set a theme, to extend the map space on one or several side of the figure, or to center a map on a specific area.

Simple export (PNG)  
`mf_export(x = mtq, width = 500,  
 filename = "my_export.png")`  
`mf_map(x = mtq, add = TRUE)`  
`dev.off()`

Export with a theme (SVG)  
`mf_export(x = mtq, width = 5, export = "svg",  
 filename = "my_export.svg",  
 theme = "nevermind")`  
`mf_map(x = mtq, add = TRUE)`  
`dev.off()`

Extra space on the figure (bottom, left, top, right)  
`mf_export(x = mtq, width = 500,  
 filename = "my_export.png",  
 expandBB = c(0,0,0,0))`  
`mf_map(x = mtq, add = TRUE)`  
`dev.off()`



Export a map centered on a specific area  
`mf_export(x = mtq[30, ], height = 600,  
 filename = "my_export.png")`  
`mf_map(x = mtq, add = TRUE)`  
`dev.off()`

## Further documentation

Vignettes on mapsf website: [riatelab.github.io/maps/](http://riatelab.github.io/maps/)

- > Get started
- > How to Use Themes
- > How to Export Maps
- > How to Create Inset Maps
- > How to Create Faceted Maps
- > How to Use a Custom Font Family

# Prediction Performance with: : metrIca

ShinyApp



## Basics

**metrIca** is a compilation of more than 80 functions designed to quantitatively and visually evaluate the prediction performance of regression (continuous) and classification (categorical) point-forecast models (e.g., APSIM, DSSAT, DNDC, Supervised Machine Learning).

## Using the functions

There are two basic arguments common to all **metrIca** functions: (i) **obs** (O; observed, a.k.a. actual, measured, truth, target, label), and (ii) **pred** (P; predicted, a.k.a. simulated, fitted, modeled, estimate) values.

Optional arguments include data that allows to call an existing data frame containing both observed and predicted vectors, and tidy, which controls the type of output as a list (tidy = FALSE) or as a data.frame (tidy = TRUE).

## Installation

```
install.packages("metrIca")
```

You can install the development version from [GitHub](#) with:

```
#install.packages("devtools")
devtools::install_github("adriancorrendo/metrIca")
```

## Native datasets

The **metrIca** package comes with four example datasets of continuous variables (regression) from the APSIM software:

- **Wheat**: 137 data-points of wheat grain N
- **Barley**: 69 data-points of barley grain number
- **Sorghum**: 36 data-points of sorghum grain number
- **Chickpea**: 39 data-points of chickpea aboveground dry mass

In addition, **metrIca** also provides two native examples for categorical variables (classification):

- **land\_cover**: binary dataset of land cover using satellite images. Values: 1=vegetation, 0=other type of land cover.
- **maize\_phenology**: data set of maize (*Zea mays* L.) phenology (16 crop development stages).

Check the metrics documentation to find all the performance metrics and their details: [metrIca](#)

## Regression

```
R2(data = wheat, obs = obs, pred=pred, tidy = TRUE)
#> R2
#> 1 0.8455538
```

```
RMSE(data = wheat, obs = obs, pred = pred)
#> $RMSE
#> [1] 1.666441
```

```
KGE(data = wheat, obs = obs, pred = pred)
#> $KGE
#> [1] 0.9106471
```

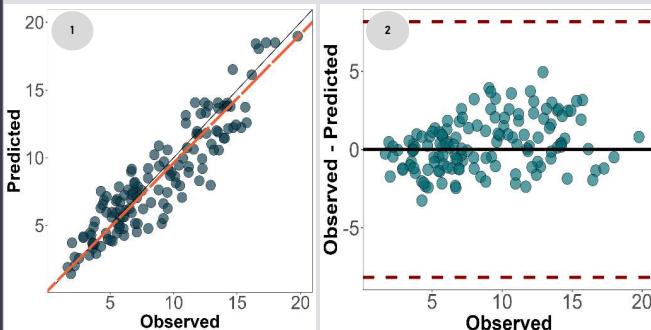
Users can also calculate **all (default)** or a selected list of metrics at once using the function **metrics\_summary()**:

```
sel_r_metrics <- c("R2", "MBE", "RMSE", "RSR", "NSE",
"KGE", "CCC")
```

```
metrics_summary(data = wheat,
                 obs = obs,
                 pred = pred,
                 type = "regression",
                 metrics_list = sel_r_metrics)
```

## Plots

1. `scatter_plot(data = wheat, obs = obs, pred = pred)`
2. `bland_altman_plot(data = wheat, obs = obs, pred = pred)`



## Classification

```
accuracy(data=maize_phenology, obs=actual, pred=predicted)
#> $accuracy
#> [1] 0.8834951
```

```
precision(data=maize_phenology, obs=actual, pred=predicted)
#> $precision
#> [1] 0.8335108
```

```
recall(data = maize_phenology, obs=actual, pred=predicted)
#> $recall
#> [1] 0.8405168
```

For classification, users can also apply the **metrics\_summary()** function to obtain multiple metrics at once:

```
sel_c_metrics <- c("accuracy", "precision", "recall",
"fscore")
```

```
metrics_summary(data = landcover,
                obs = actual, pred = predicted,
                type = "classification",
                metrics_list = sel_c_metrics,
                pos_level = 1)
```

## Confusion matrix

```
confusion_matrix(data = .,
                  obs = labels, pred = predictions,
                  plot = TRUE,
                  unit="count")
```

### Binomial case

Performance metrics:

accuracy = 0.43; precision = 0.43; recall = 0.43.

Observed

|           |     | 0   | 1  |
|-----------|-----|-----|----|
| Predicted | 0   | 151 | 6  |
|           | 1   | 6   | 82 |
| count     | 157 | 88  |    |

### Multinomial case

Performance metrics:

accuracy = 0.43; precision = 0.43; recall = 0.43.

Observed

|           |       |    | Blue | Green | Red |
|-----------|-------|----|------|-------|-----|
| Predicted | Blue  | 9  | 6    | 12    |     |
|           | Green | 13 | 16   | 12    |     |
| count     | 18    | 15 | 12   |       |     |

# Machine Learning with R



## Introduction

**mlr** offers a unified interface for the basic building blocks of machine learning: tasks, learners, hyperparameters, etc.

**Tasks** contain a description of a task (classification, regression, clustering, etc.) and a data set.

**Learners** specify a machine learning algorithm (GLM, SVM, xgboost, etc.) and its parameters.

**Hyperparameters** are learner settings that can be specified directly or tuned. A **parameter set** lists the possible hyperparameters for a given learner.

**Wrapped Models** are learners that have been trained on a task and can be used to make predictions.

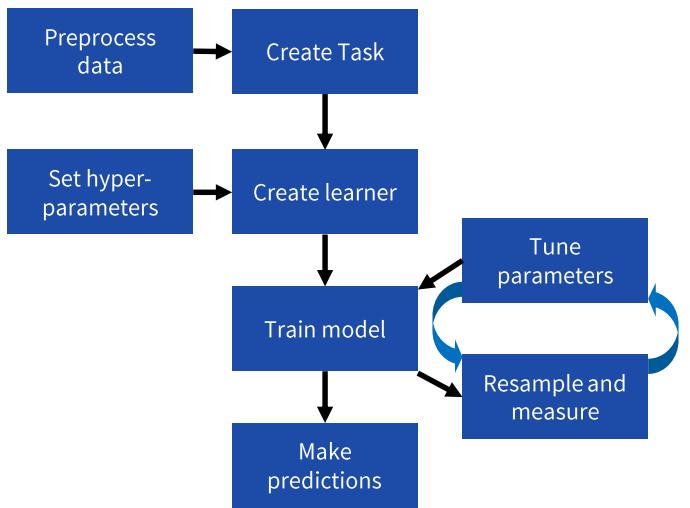
**Predictions** are the results of applying a model to either new data or the original training data.

**Measures** control how learner performance is evaluated, e.g. RMSE, LogLoss, AUC, etc.

**Resampling** estimates generalization performance by separating training data from test data. Common strategies include holdout and cross-validation.

Links: [Tutorial](#) | [CRAN](#) | [Github](#)

## mlr workflow



## Setup

### Preprocessing data

`createDummyFeatures(obj=, target=, method=, cols=)`  
Creates (0,1) flags for each non-numeric variable excluding `target`. Can be applied to entire dataset or only specific `cols`

`normalizeFeatures(obj=, target=, method=, cols=, range=, on.constant=)`  
Normalizes numerical features according to specified `method`:

- "center" (subtract mean)
- "scale" (divide by std. deviation)
- "standardize" (center and scale)
- "range" (linear scale to given range, default `range=c(0,1)`)

`mergeSmallFactorLevels(task=, cols=, min.perc=)`  
Combine infrequent factor levels into a single merged level

`summarizeColumns(obj=)` where `obj` is a data.frame or task.  
Provides type, NA, and distributional data about each column

See also `capLargeValues` `dropFeatures` `removeConstantFeatures` `summarizeLevels`

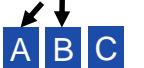
### Creating a task



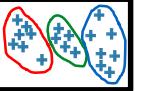
`makeClassifTask(data=, target=)`  
Classification of a target variable, with optional positive class `positive`



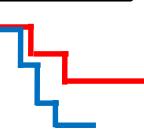
`makeRegrTask(data=, target=)`  
Regression on a target variable



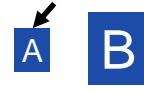
`makeMultilabelTask(data=, target=)`  
Classification where the target can belong to more than one class per observation



`makeClusterTask(data=)`  
Unsupervised clustering on a data set



`makeSurvTask(data=, target= c("time", "event"))`  
Survival analysis with a survival time column and an event column



`makeCostSensTask(data=, costs=)`  
Cost-sensitive classification where each observation-cost pair has a specified cost

Other arguments that can be passed to a `task`:

- `weights`= Weighting vector to apply to observations
- `blocking`= Factor vector where each level indicates a block of observations that will not be split up in resampling

### Making a learner

`makeLearner(cl=, predict.type=, ..., par.vals=)`  
Choose an algorithm class to perform the task and determine what that algorithm will predict

- `cl`=name of algorithm, e.g. `"classif.xgboost"` `"regr.randomForest"` `"cluster.kmeans"`
  - `predict.type="response"` returns a prediction type that matches the source data; `"prob"` returns a predicted probability for classification problems only; `"se"` returns the standard error of the prediction for regression problems only. Only certain learners can return `"prob"` and `"se"`
  - `par.vals`= takes a list of hyperparameters and passes them to the learner; parameters can also be passed directly (...)
- You can make multiple learners at once with `makeLearners()`

mlr has integrated over 170 different learning algorithms

- Full list: `View(listLearners())` shows all learners
- Available learners for a task: `View(listLearners(task))`
- Filtered list: `View(listLearners("classif", properties=c("prob", "factors")))` shows all classification learners `"classif"` which can predict probabilities `"prob"` and handle factor inputs `"factors"`
- See also `getLearnerProperties()`

## Training & Testing

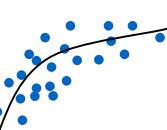
### Setting hyperparameters

`setHyperPars(learner=, ...)`  
Set the hyperparameters (settings) for each learner, if you don't want to use the defaults. You can also specify hyperparameters in the `makeLearner()` call

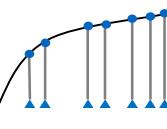


`getParamSet(learner=)`  
Show the possible universe of parameters for your learner; can take a learner directly, or a text string such as `"classif.qda"`

### Train a model and predict



`train(learner=, task=)`  
Train a model (`WrappedModel`) by applying a learner to a task. By default, the model will train on all observations. The underlying model can be extracted with `getLearnerModel()`



`predict(object=, task=, newdata=)`  
Use a trained model to make predictions on a task or dataset. The resulting `pred` object can be viewed with `View(pred)` or accessed by `as.data.frame(pred)`

### Measuring performance



`performance(pred=, measures=)`  
Calculate performance of predictions according to one or more of several measures (use `listMeasures()` for full list):

- `classif` `acc` `auc` `bac` `ber` `brier[,scaled]` `f1` `fdr` `fnr` `fpr` `gmean` `multiclass[,au1]` `aunp` `aunu` `brier` `npv` `ppv` `qsr` `ssr` `tn` `tnr` `tp` `tpr` `wkappa`
- `regr` `rsq` `expvar` `kendalltau` `mae` `mape` `medae` `medse` `mse` `msle` `rae` `rmse` `rmsle` `rrse` `rsq` `sae` `spearmanrho` `sse`
- `cluster` `db` `dunn` `G1` `G2` `silhouette`
- `multilabel` `multilabel[,f1]` `subset01` `.tpr` `.ppv` `.acc` `.hamloss`
- `costsens` `mcp` `meancosts`
- `surv` `cindex`
- `other` `featperc` `timeboth` `timelpredict` `timetrain`

For detailed performance data on classification tasks, use:

- `calculateConfusionMatrix(pred=)`
- `calculateROCMeasures(pred=)`

### Resampling a learner

`makeResampleDesc(method=, ..., stratify=)`

`method` must be one of the following:

- "CV" (cross-validation, for number of folds use `iters=`)
  - "LOO" (leave-one-out cross-validation, for folds use `iters=`)
  - "RepCV" (repeated cross-validation, for number of repetitions use `reps=`, for folds use `folds=`)
  - "Subsample" (aka Monte-Carlo cross-validation, for iterations use `iters=`, for train % use `split=`)
  - "Bootstrap" (out-of-bag bootstrap, uses `iters=`)
  - "Holdout" (for train % use `split=`)
- `stratify` keeps target proportions consistent across samples.

`makeResampleInstance(desc=, task=)` can reduce noise by ensuring the resampling is done identically every time.

`resample(learner=, task=, resampling=, measures=)`  
Train and test model according to specified resampling strategy.

mlr includes several pre-specified resample descriptions: `cv2` (2-fold cross-validation), `cv3`, `cv5`, `cv10`, `hout` (holdout with split 2/3 for training, 1/3 for testing). Convenience functions also exist to `resample()` with a specific strategy: `crossval()`, `repCV()`, `holdout()`, `subsample()`, `bootstrap00B()`, `bootstrapB632()`, `bootstrapB632plus()`

## Refining Performance

### Tuning hyperparameters

Set search space using `makeParamSet(make<type>Param())`

- `makeNumericParam(id=, lower=, upper=, trafo=)`
  - `makeIntegerParam(id=, lower=, upper=, trafo=)`
  - `makeIntegerVectorParam(id=, len=, lower=, upper=, trafo=)`
  - `makeDiscreteParam(id=, values=c(...))` (can also be used to test discrete values of numeric or integer parameters)
- `trafo` transforms the parameter output using a specified function, e.g. `lower=-2, upper=2, trafo=function(x) 10^x` would test values between 0.01 and 100, scaled exponentially
- Other acceptable parameter types include `Logical` `LogicalVector` `CharacterVector` `DiscreteVector`

Set a search algorithm with `makeTuneControl<type>()`

- `Grid(resolution=10)` Grid of all possible points
- `Random(maxit=100)` Randomly sample search space
- `MBO(budget=)` Use Bayesian model-based optimization
- `Irace(n.instances=)` Iterated racing process
- Other types: `CMAES`, `Design`, `GenSA`

Tune using `tuneParams(learner=, task=, resampling=, measures=, par.set=, control=)`

## Quickstart

### Prepare data for training and testing

```

library(mlbench)
data(Soybean)
soy = createDummyFeatures(Soybean, target="Class")
tsk = makeClassifTask(data=soy, target="Class")
ho = makeResampleInstance("Holdout", tsk)
tsk.train = subsetTask(tsk, ho$train.ind[1])
tsk.test = subsetTask(tsk, ho$test.ind[1])

```

Convert the factor inputs in the Soybean dataset into (0,1) dummy features which can be used by the XGboost algorithm. Create a task to predict the "Class" column. Create a train set with 2/3 of data and a test set with the remaining 1/3 (default).

### Create learner and evaluate performance

```

lrn = makeLearner("classif.xgboost", nrounds=10)
cv = makeResampleDesc("CV", iters=5)
res = resample(lrn, tsk.train, cv, acc)

```

Create an XGboost learner which will build 10 trees. Then test performance using 5-fold cross-validation. Accuracy should be between 0.90-0.92.

### Tune hyperparameters and retrain model

```

ps = makeParamSet(makeNumericParam("eta", 0, 1),
                  makeNumericParam("lambda", 0, 200),
                  makeIntegerParam("max_depth", 1, 20))
tc = makeTuneControlMBO(budget=100)
tr = tuneParams(lrn, tsk.train, cv5, acc, ps, tc)
lrn = setHyperPars(lrn, par.vals=tr$x)

```

Tune hyperparameters `eta`, `lambda`, and `max_depth` by defining a search space and using Model Based Optimization (MBO) to control the search. Then perform 100 rounds of 5-fold cross-validation, improving accuracy to ~0.93. Update the XGboost learner with the tuned hyperparameters.

```

mdl = train(lrn, tsk.train)
prd = predict(mdl, tsk.test)
calculateConfusionMatrix(prd)
mdl = train(lrn, tsk)

```

Train the model on the train set and make predictions on the test set. Show performance as a confusion matrix. Finally, re-train model on the full set to use on new data. You are now ready to go out into the real world and make 93% accurate predictions!

Legend for functions (not all parameters shown):

`function(required_parameters, optional_parameters=)`

# Configuration

mlr's default settings can be changed using `configureMlr()`:

- `show.info` Whether to show verbose output by default when training, tuning, resampling, etc. (`TRUE`)
- `on.learner.error` How to handle a learner error. `"stop"` halts execution, `"warn"` returns NAs and displays a warning, `"quiet"` returns NAs with no warning (`"stop"`)
- `on.learner.warning` How to handle a learner warning. `"warn"` displays a warning, `"quiet"` suppresses it (`"warn"`)
- `on.par.without.desc` How to handle a parameter with no description. `"stop"`, `"warn"`, `"quiet"` (`"stop"`)
- `on.par.out.of.bounds` How to handle a parameter with an out-of-bounds value. `"stop"`, `"warn"`, `"quiet"` (`"stop"`)
- `on.measure.not.applicable` How to handle a measure not applicable to a learner. `"stop"`, `"warn"`, `"quiet"` (`"stop"`)
- `show.learner.output` Whether to show learner output to the console during training (`TRUE`)
- `on.error.dump` Whether to create an error dump for crashed learners if `on.learner.error` is not set to `"stop"` (`TRUE`)

Use `getMlrOptions()` to see current settings

# Parallelization

mlr works with the `parallelMap` package to take advantage of multicore and cluster computing for faster operations. mlr automatically detects which operations are able to run in parallel.

To begin parallel operation use:

- ```
parallelStart(mode=, cpus=, level=)
```
- `mode` determines how the parallelization is performed:
    - `"local"` no parallelization applied, simply uses `mapply`
    - `"multicore"` multicore execution on a single machine, uses `parallel::mclapply`. Not available in Windows.
    - `"socket"` multicore execution in socket mode
    - `"mpi"` Snow MPI cluster on one or multiple machines using `parallel::makeCluster` and `parallel::clusterMap`
    - `"BatchJobs"` Batch queuing HPC clusters using `BatchJobs::batchMap`
  - `cpus` determines how many logical cores will be used
  - `level` controls parallelization: `"mlr.benchmark"`, `"mlr.resample"`, `"mlr.selectFeatures"`, `"mlr.tuneParams"`, `"mlr.ensemble"`

To end parallelization, use `parallelStop()`

# Imputation

`impute(obj=, target=, cols=, dummy.cols=, dummy.type=)`  
Applies specified logic to data frame or task containing NAs and returns an imputation description which can be used on new data

- `obj`=data frame or task on which to perform imputation
- `target`=specify target variable which will not be imputed
- `cols`=column names and logic for imputation\*
- `dummy.cols`=column names to create a NA (T/F) column\*
- `dummy.type`=set to `"numeric"` to use (0,1) instead of (T/F)

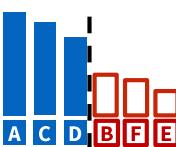
\*Can also use `classes` and `dummy.classes` in place of `cols`

Imputation logic is passed to `cols` or `classes` via a list, e.g.: `cols=list(V1=imputeMean())` where `V1` is the column to which to apply the imputation, and `imputeMean()` is the imputation method. Available imputation methods include:  
`imputeConst(const=)` `imputeMedian()` `imputeMode()` `imputeMin(multiplier=)` `imputeMax(multiplier=)` `imputeNormal(mean=, sd=)` `imputeHist(breaks=, use.mids=)` `imputeLearner(learner=, features=)` `impute` returns a list containing the imputed dataset or task as well as an imputation description that can be used to reapply the same imputation to new data using `reimpute`

`reimpute(obj=, desc=)` Imputes missing values on a task or dataset (`obj`) using a description (`desc`) created by `impute`

# Feature Extraction

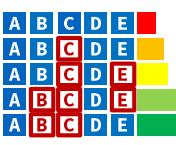
## Feature filtering



`filterFeatures(task=, method=, perc=, abs=, threshold=)`  
Uses a learner-agnostic feature evaluation method to rank feature importance, then includes only features in the top n percent (`perc=`), top n (`abs=`), or which meet a set performance threshold (`threshold=`).

Outputs a task with features that failed the test omitted. `method` defaults to `"randomForestSRC.rfsrc"`, but can be set to:  
`"anova.test"` `"carscore"` `"cforest.importance"`  
`"chi.squared"` `"gain.ratio"` `"information.gain"`  
`"kruskal.test"` `"linear.correlation"` `"mrmr"` `"oneR"`  
`"permutation.importance"` `"randomForest.importance"`  
`"randomForestSRC.rfsrc"` `"randomForestSRC.var.select"`  
`"rank.correlation"` `"relief"`  
`"symmetrical.uncertainty"` `"univariate.model.score"`  
`"variance"`

## Feature selection



`selectFeatures(learner=, task=, resampling=, measures=, control=)`  
Uses a feature selection algorithm (`control`) to resample and build a model repeatedly using different feature sets each time in order to find the best set.

Available controls include:

- `makeFeatSelControlExhaustive(max.features=)` Try every combination of features up to optional `max.features`
- `makeFeatSelControlRandom(maxit=, prob=, max.features=)` Randomly sample features with probability `prob` (default 0.5) until `maxit` (default 100) iterations; return the best one found
- `makeFeatSelControlSequential(method=, maxit=, max.features=, alpha=, beta=)` Perform an iterative search using a `method` from the following: `"sfs"` forward search, `"sbs"` backward search, `"sfbs"` floating forward search, `"sfbs"` floating backward search. `alpha` indicates minimum improvement required to add a feature; `beta` indicates minimum required to remove a feature
- `makeFeatSelControlGA(maxit=, max.features=, mu=, lambda=, crossover.rate=, mutation.rate=)` Genetic algorithm trains on random feature vectors, then uses crossover on the best performers to produce 'offspring', repeated over generations. `mu` is size of parent population, `lambda` is size of children population, `crossover.rate` is probability of choosing a bit from first parent, `mutation.rate` is probability of flipping a bit (on or off)

`selectFeatures` returns a `FeatSelResult` object which contains optimal features and an optimization path. To apply feature selection result (`fsr`) to your task (`tsk`), use:  
`tsk = subsetTask(tsk, features=fsr$x)`

# Benchmarking

`benchmark(learners=, tasks=, resamplings=, measures=)`  
Allows easy comparison of multiple learners on a single task, a single learner on multiple tasks, or multiple learners on multiple tasks. Returns a benchmark result object.

Benchmark results can be accessed with a variety of functions beginning with `getBMR<object>.AggrPerformance`  
`FeatSelResults` `FilteredFeatures` `LearnerIds`  
`LeanerShortNames` `Learners` `MeasureIds` `Measures`  
`Models` `Performances` `Predictions` `TaskDescs` `TaskIds`  
`TuneResults`

mlr contains several toy tasks which are useful for benchmarking:  
`agri.task` `bc.task` `bh.task` `costiris.task` `iris.task`  
`lung.task` `mtcars.task` `pid.task` `sonar.task`  
`wpbc.task` `yeast.task`

# Visualization

## Performance

`generateThreshVsPerfData(obj=, measures=)` Measure performance at different probability cutoffs to determine optimal decision threshold for binary classification problems

- `plotThreshVsPerf(obj=)` Plot visual representation of threshold curve(s) from `ThreshVsPerfData`
- `plotROCCurves(obj=)` Plot receiver operating characteristic (ROC) curve from `ThreshVsPerfData`. Must set `measures=list(fpr, tpr)`

## Residuals

- `plotResiduals(obj=)` Plots residuals for `Prediction` or `BenchmarkResult`

## Learning curve

`generateLearningCurveData(learners=, task=, resampling=, percs=, measures=)` Measure performance of learner(s) trained on different percentages of task data

- `plotLearningCurve(obj=)` Plot curve showing learner performance vs. proportion of data used, uses `LearningCurveData`

## Feature importance

`generateFilterValuesData(task=, method=)` Get feature importance rankings using specified filter method

- `plotFilterValues(obj=)` Plot bar chart of feature importance based on filter method using `FilterValuesData`

## Hyperparameter tuning

`generateHyperParsEffectData(tune.result=)` Get the impact of different hyperparameter settings on model performance

- `plotHyperParsEffect(hyperpars.effec.t.data=, x=, y=, z=)` Create a plot showing hyperparameter impact on performance using `HyperParsEffectData`

See also:

- `plotOptPath(op=)` Display details of optimization process. Takes `<obj>$opt.path`, where `<obj>` is an object of class `tuneResult` or `featSelResult`
- `plotTuneMultiCritResult(res=)` Show pareto front for results of tuning to multiple performance measures

## Partial dependence

`generatePartialDependenceData(obj=, input=)` Get partial dependence of model (`obj`) prediction over each feature of data (`input`)

- `plotPartialDependence(obj=)` Plots partial dependence of model using `PartialDependenceData`

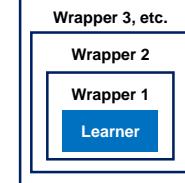
## Benchmarking

- `plotBMRBoxplots(bmr=)` Distribution of performances
- `plotBMRSummary(bmr=)` Scatterplot of avg. performances
- `plotBMRanksAsBarChart(bmr=)` Rank learners in bar plot

## Other

- `generateCritDifferencesData(bmr=, measure=, p.value=, test=)` Perform critical-differences test using either the Bonferroni-Dunn ("bd") or "Nemenyi" test
- `plotCritDifferences(obj=)`
- `generateCalibrationData(obj=)` Evaluate calibration of probability predictions vs. true incidence
- `plotCalibration(obj=)`

# Wrappers



**Wrappers** fuse a learner with additional functionality. mlr treats a learner with wrappers as a single learner, and hyperparameters of wrappers can be tuned jointly with underlying model parameters. Models trained with wrappers will apply them to new data.

## Preprocessing and imputation

`makeDummyFeaturesWrapper(learner=)`  
`makeImputeWrapper(learner=, classes=, cols=)`  
`makePreprocWrapper(learner=, train=, predict=)`  
`makePreprocWrapperCaret(learner=, ...)`  
`makeRemoveConstantFeaturesWrapper(learner=)`

## Class imbalance

`makeOverBaggingWrapper(learner=)`  
`makeSMOTEWrapper(learner=)`  
`makeUndersampleWrapper(learner=)`  
`makeWeightedClassesWrapper(learner=)`

## Cost-sensitive learning

`makeCostSensClassifWrapper(learner=)`  
`makeCostSensRegrWrapper(learner=)`  
`makeCostSensWeightedPairsWrapper(learner=)`

## Multilabel classification

`makeMultilabelBinaryRelevanceWrapper(learner=)`  
`makeMultilabelClassifierChainsWrapper(learner=)`  
`makeMultilabelDBRWrapper(learner=)`  
`makeMultilabelNestedStackingWrapper(learner=)`  
`makeMultilabelStackingWrapper(learner=)`

## Other

`makeBaggingWrapper(learner=)`  
`makeConstantClassWrapper(learner=)`  
`makeDownsampleWrapper(learner=, dw.perc=)`  
`makeFeatSelWrapper(learner=, resampling=, control=)`  
`makeFilterWrapper(learner=, fw.perc=, fw.abs=, fw.threshold=)`  
`makeMultiClassWrapper(learner=)`  
`makeTuneWrapper(learner=, resampling=, par.set=, control=)`

## Nested Resampling

mlr supports **nested resampling** for complex operations such as tuning and feature selection through wrappers. In order to get a good estimate of generalization performance and avoid data leakage, both an outer (for tuning/feature selection) and an inner (for the base model) resampling process are advised.

- Outer resampling can be specified in `resample` or `benchmark`
- Inner resampling can be specified in `makeTuneWrapper`, `makeFeatSelWrapper`, etc.

## Ensembles

`makeStackedLearner(base.learners=, super.learner=, method=)` Combines multiple learners to create an ensemble

- `base.learners`=learners to use for initial predictions
- `super.learner`=learner to use for final prediction
- `method`=how to combine base learner predictions:
  - `"average"` simple average of all base learners
  - `"stack.nocv", "stack.cv"` train super learner on results of base learners, with or without cross-validation
  - `"hill.climb"` search for optimal weighted average
  - `"compress"` with a neural network for faster performance

# Intro stats with mosaic

(lattice version)

## Essential R syntax

Names in R are *case sensitive*

Function and arguments

`rflip(10)`

Optional arguments

`rflip(10, prob = 0.8)`

Assignment

`x <- rflip(10, prob = 0.8)`

Getting help on any function

`help(mean)`

## Loading packages

`library(mosaic)`

## Arithmetic operations

`+ - * /` basic operations

`^` exponentiation

`( )` grouping

`sqrt(x)` square root

`abs(x)` absolute value

`log10(x)` logarithm, base 10

`log(x)` natural logarithm, base  $e$

`exp(x)` exponential function  $e^x$

`factorial(k)`  $k! = k(k-1) \dots 1$

## Logical operators

`==` is equal to (note double equal sign)

`!=` is not equal to

`<` is less than

`<=` is less than or equal to

`>` is greater than

`>=` is greater than or equal to

`&` **A & B** is TRUE if both **A** and **B** are TRUE

`|` **A | B** is TRUE if one or both of **A** and **B** are TRUE

`%in%` includes; for example

`"C" %in% c("A", "B")` is FALSE

## Formula interface

Use for graphics, statistics, inference, and modeling operations.

`goal(y ~ x, data = mydata)`  
Read as “Calculate **goal** for **y** using **mydata** “broken down by” **x**, or “modeled by” **x**.

`mean(age ~ sex, data = HELPrc)`

For graphics:

`goal(y ~ x | z, groups = w, data = mydata)`

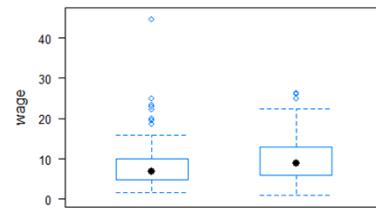
**y** : y-axis variable (*optional*)

**x** : x-axis variable (*required*)

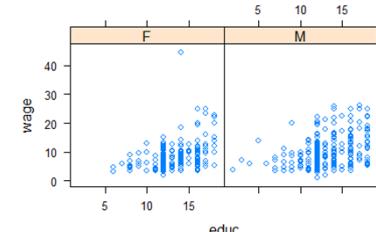
**z** : panel-by variable (*optional*)

**w** : color-by variable (*optional*)

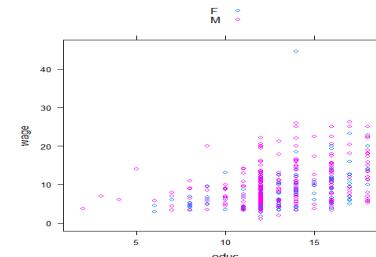
`bwplot(wage ~ sex, data = CPS85)`



`xyplot(wage ~ educ | sex, data = CPS85)`



`xyplot(wage ~ educ, groups = sex, data = CPS85, auto.key = TRUE)`



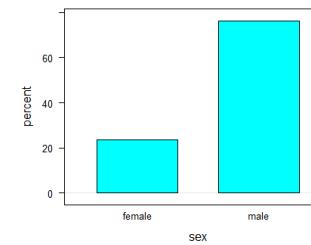
## One categorical variable

Counts by category

`tally(~ sex, data = HELPrc)`

Percentages by category

`tally(~ sex, format = "percent", data = HELPrc)`  
`bargraph(~ sex, type = "percent", data = HELPrc)`



Tests and confidence intervals

Exact test

`result1 <- binom.test(~ (homeless == "homeless"), data = HELPrc)`

Approximate test (large samples)

`result2 <- prop.test(~ (homeless == "homeless"), data = HELPrc)`

Extract confidence intervals and p-values

`confint(result1)`  
`pval(result2)`

## Examining data

Print short summary of all variables

`inspect(HELPrc)`

Number of rows and columns

`dim(HELPrc)`

`nrow(HELPrc)`

`ncol(HELPrc)`

Print first rows or last rows

`head(KidsFeet)`  
`tail(KidsFeet, 10)`

Names of variables

`names(HELPrc)`

## One quantitative variable

Make output more readable

`options(digits = 3)`

Compute summary statistics

`mean(~ cesd, data = HELPrc)`

Other summary statistics work similarly

`median()` `iqr()` `max()` `min()`

`fivenum()` `sd()` `var()` `sum()`

Table of summary statistics

`favstats(~ cesd, data = HELPrc)`

Summary statistics by group

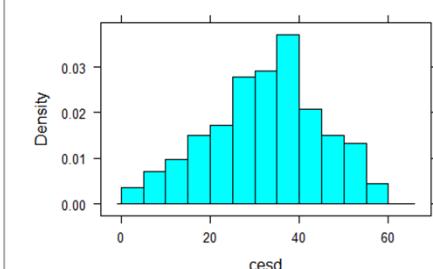
`favstats(cesd ~ sex, data = HELPrc)`

Quantiles

`quantile(~ cesd, data = HELPrc, prob = c(0.25, 0.5, 0.8))`

Histogram

`histogram(~ cesd, width = 5, center = 2.5, data = HELPrc)`



Normal probability plot

`qqmath(~ cesd, dist = "qnorm", data = HELPrc)`

Density plot

`densityplot(~ cesd, data = HELPrc)`

Dot plot

`dotPlot(~ cesd, data = HELPrc)`

One-sample t-test

`result <- t.test(~ cesd, mu = 34, data = HELPrc)`

Extract confidence intervals and p-values

`confint(result)`  
`pval(result)`

## Two categorical variables

Contingency table with margins

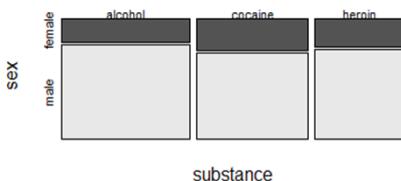
```
tally(~ substance + sex,  
      margins = TRUE,  
      data = HELPrc)
```

Percentages by column

```
tally(~ sex | substance,  
      format = "percent",  
      data = HELPrc)
```

Mosaic plot

```
mosaicplot(~ substance + sex,  
           color = TRUE, data = HELPrc)
```



Chi-square test

```
xchisq.test(~ substance + sex,  
            data = HELPrc,  
            correct = FALSE)
```

## Distributions

Normal distribution function

```
pnorm(13, mean = 10, sd = 2)
```

Normal distribution function with graph

```
xpnorm(1.645, mean = 0, sd = 1)
```

Normal distribution quantiles

```
qnorm(0.95) # mean = 0, sd = 1
```

Normal distribution quantiles with graph

```
xqnorm(0.85, mean = 10, sd = 2)
```

Binomial density function ("size" means  $n$ )

```
dbinom(5, size = 8, prob = 0.65)
```

Binomial distribution function

```
pbinom(5, size = 8, prob = 0.65)
```

Central portion of distribution

```
cdist("norm", 0.95)
```

```
cdist("t", c(0.90, 0.99), df = 5)
```

Plotting distributions

```
plotDist("binom", size = 8,  
        prob = 0.65, xlim = c(-1, 9))
```

```
plotDist("norm", mean = 10,  
        sd = 2)
```

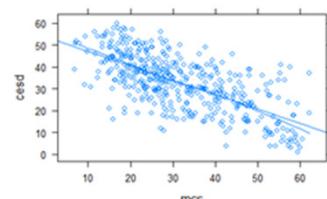
## Two quantitative variables

Correlation coefficient

```
cor(cesd ~ mcs, data = HELPrc)
```

Scatterplot with regression line and smooth

```
xyplot(cesd ~ mcs,  
       type = c("p", "r", "smooth"),  
       data = HELPrc)
```



Simple linear regression

```
cesdmodel <- lm(cesd ~ mcs,  
                  data = HELPrc)
```

```
msummary(cesdmodel)
```

Prediction

```
lmfunction <- makeFun(cesdmodel)  
lmfunction(mcs = 35)
```

Extract useful quantities

```
anova(cesdmodel)
```

```
coef(cesdmodel)
```

```
confint(cesdmodel)
```

```
rsquared(cesdmodel)
```

Diagnostics; plot residuals

```
histogram(~resid(cesdmodel),  
          density = TRUE)
```

```
qqmath(~resid(cesdmodel))
```

Diagnostics; plot residuals vs. fitted

```
xyplot(resid(cesdmodel) ~  
       fitted(cesdmodel),  
       type = c("p", "smooth", "r"))
```

## Categorical response, quantitative predictor

Logistic regression

```
logit_mod <-  
  glm(homeless ~ age + female,  
       family = binomial, data = HELPrc)
```

```
msummary(logitmod)
```

Odds ratios and confidence intervals

```
exp(coef(logit_mod))
```

```
exp(confint(logit_mod))
```

## Data management

From dplyr package

Drop or reorder variables

```
select()
```

Create new variables from existing ones

```
mutate()
```

Retain specific rows from data

```
filter()
```

Sort data rows

```
arrange()
```

Compute summary statistics by group

```
group_by()
```

```
summarize()
```

Merge data tables

```
left_join()
```

```
inner_join()
```

## Importing data

Import file from computer or URL

```
MustangPrice <-  
  read.file("C:/MustangPrice.csv")  
# NOTE: R uses forward slashes!  
Dome <-  
  read.file("http://www.mosaic-  
web.org/go/datasets/Dome.csv")
```

## Randomization and simulation

Fix random number sequence

```
set.seed(42)
```

Tossing coins

```
rflip(10) # default prob is 0.5
```

Do something repeatedly

```
do(5) * rflip(10, prob = 0.75)
```

Draw a simple random sample

```
sample(LETTERS, 10)
```

```
deal(Cards, 5) # poker hand
```

Resample with replacement

```
Small <- sample(KidsFeet, 10)
```

```
resample(�Small)
```

Random permutation (shuffling)

```
shuffle(Cards)
```

Random values from distributions

```
rbinom(5, size = 10, prob = 0.7)
```

```
rnorm(5, mean = 10, sd = 2)
```

## Quantitative response, categorical predictor

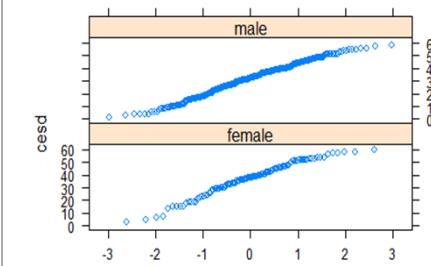
Two-level predictor: two-sample  $t$  test

Numeric summaries

```
favstats(~cesd | sex,  
         data = HELPrc)
```

Comparative normal probability plot

```
qqmath(~cesd | sex, data = HELPrc,  
      layout = c(1, 2)) # also bwplot
```



Dotplot for smaller samples

```
xyplot(sex ~ length, alpha = 0.6,  
      cex = 1.4, data = KidsFeet)
```

Two-sample  $t$ -test and confidence interval

```
result <- t.test(cesd ~ sex,  
                 var.equal = FALSE, data = HELPrc)
```

```
confint(result)
```

More than two levels: Analysis of variance

Numeric summaries

```
favstats(cesd ~ substance,  
         data = HELPrc)
```

Graphic summaries

```
bwplot(cesd ~ substance, pch = "|",  
       data = HELPrc)
```

Fitt and summarize model

```
modsubstance <- lm(cesd ~ substance,  
                     data = HELPrc)
```

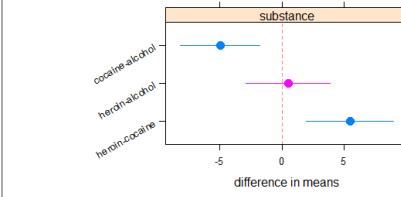
```
anova(modsubstance)
```

Which differences are significant?

```
pairwise <- TukeyHSD(modsubstance)
```

```
mplot(pairwise)
```

95% family-wise confidence level



# nardl Package

An R package to estimate the nonlinear cointegrating autoregressive distributed lag model

## Specifying the Model

Possible syntaxes for specifying the variables in the model:

- nardl with fixed p and q lags**

```
nardl(fod~inf,p,q,data=fod,ic="aic",maxlags = FALSE,graph = FALSE,case=3)
```

- Auto selected lags (maxlags=TRUE)**

```
nardl(food~inf,data=fod,ic="aic",maxlags = TRUE,graph = FALSE,case=3)
```

**The formula:**

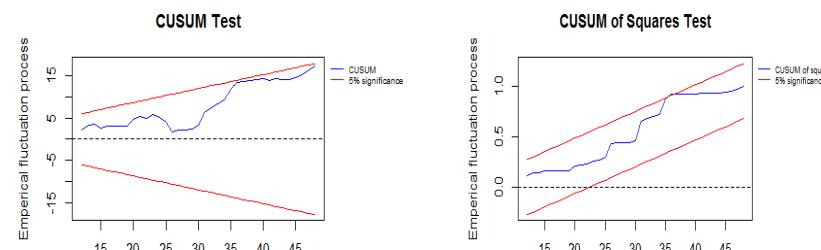
- $y \sim x | z_1 + z_2 \dots$
- $y$  the dependent variable
- $x$  the decomposed variable (this package version can't assume more than one decomposed variable)
- $z_1 + z_2 + \dots$  independent variables

- Data** is the dataframe
- p** number of lags of the dependent variable
- q** number of lags of the independent variables
- ic** : c("aic", "bic", "ll", "R2") criteria model selection
- maxlags** if **TRUE** auto lags selection
- case** case number 3 for (unrestricted intercept, no trend) and 5 (unrestricted intercept, unrestricted trend), 1 2 and 4 not supported

## Cusum and CusumQ plot

Cusum and CusumQ plot (graph=TRUE)

```
nardl(food~inf,data=fod,ic="aic",maxlags = TRUE,graph = TRUE,case=3)
```



## Cointegration bounds test

```
pssbounds(obs, fstat, tstat = NULL, case, k)
```

**pssbounds specification include:**

- Case** case number 3 for (unrestricted intercept, no trend) and 5 (unrestricted intercept, unrestricted trend), 1 2 and 4 not supported
- fstat** represent the value of the F-statistic
- obs** represent the number of observation
- k** number of regressors appearing in lag levels

**Example:**

```
reg<-nardl(food~inf,fod,ic="aic",maxlags = TRUE,graph = TRUE,case=3)
pssbounds(case=reg$case,fstat=reg$fstat,obs=reg$obs,k=reg$k)
```

## LM test for serial correlation

LM test for serial correlation

```
bp2(object, nlags, fill = NULL, type = c("F", "Chi2"))
```

**Methods and options are:**

- object** fitted lm model
- nlags** positive integer number of lags
- fill** starting values for the lagged residuals in the auxiliary regression. By default 0.
- type** Fisher or Chisquare statistics

**Example :**

```
reg<-nardl(food~inf,fod,ic="aic",maxlags = TRUE,graph = TRUE,case=3)
```

```
bp2(reg$fit,reg$np,fill=0,type="F")
```

## Lagrange multiplier test

Lagrange multiplier test for conditional heteroscedasticity of Engle (1982), as described by Tsay (2005, pp. 101-102)

```
ArchTest(x, lags = 12, demean = FALSE)
```

**Methods and options are:**

- x** numeric vector
- lags** positive integer number of lags.
- demean** logical: If TRUE, remove the mean before computing the test statistic.

**Example :**

```
reg<-nardl(food~inf,fod,ic="aic",maxlags = TRUE,graph = TRUE,case=3)
x<-reg$selresidu
nlag<-reg$np
ArchTest(x, lags=nlag)
```

## Dynamic multipliers plot

Dynamic multiplier plot

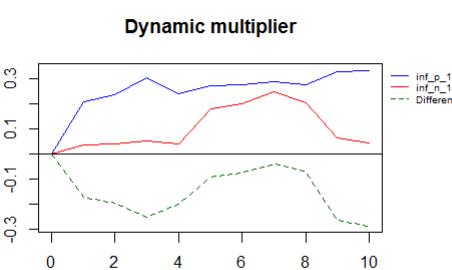
```
plotmplier(model, np, k, h)
```

**Methods and options are:**

- model** the fitted model
- np** the selected number of lags
- k** number of decomposed independent variables
- h** is the horizon over which multipliers will be computed

**Example**

```
reg<-nardl(food~inf,p=4,q=4,fod,ic="aic",maxlags = FALSE,graph = TRUE,case=3)
plotmplier(reg,reg$np,1,10)
```



## pssbounds

**pssbound** function display the necessary critical values to conduct the Pesaran, Shin and Smith 2001 bounds test for cointegration. See <http://andyphilips.github.io/pssbounds/>.

```
pssbounds(obs, fstat, tstat = NULL, case, k)
```

**Methods and options are:**

- obs** number of observations
- fstat** value of the F-statistic
- tstat** value of the t-statistic
- case** case number
- k** number of regressors appearing in lag levels

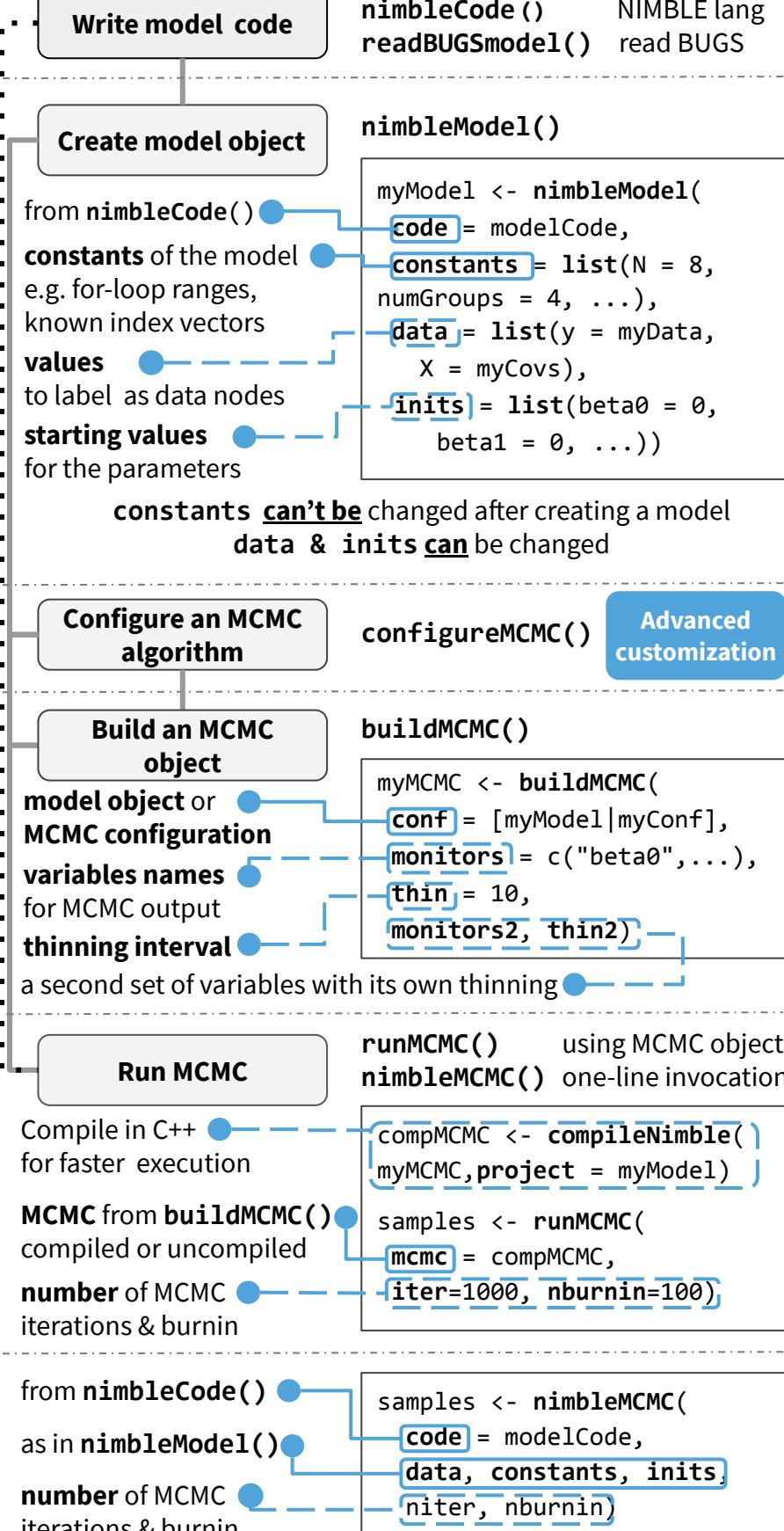
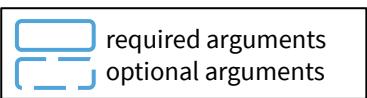
**Example**

```
reg<-nardl(food~inf,fod,ic="aic",maxlags = TRUE,graph = TRUE,case=3)
pssbounds(case=reg$case,fstat=reg$fstat,obs=reg$obs,k=reg$k)
# F-stat concludes I(1) and cointegrating, t-stat concludes I(0).
```

# nimble models: : CHEAT SHEET



## NIMBLE workflow



## Writing model code

Split code over multiple lines to help people read it.

**Use named arguments**  
for non-default parameterization  
e.g. `beta0` and `beta1` follow equivalent distributions  
(default is precision, `tau`).

**Link functions**  
can be declared on the left-hand side.

**Order of declaration**  
**does not matter**  
`alpha[iGroup]` can be declared after being used in other declarations.

```

modelCode <- nimbleCode({
  beta0 ~ dnorm(0, sd = 1000)
  beta1 ~ dnorm(0, 1E-6)
  sdGroups ~ dunif(0, 100)
  fixed_effects[1:N] <- beta0 + beta1 * X[1:N]
  for(i in 1:N) {
    log(eta[i]) <- fixed_effects[i] +
      alpha[groupID[i]]
    y[i] ~ dpois(eta[i])
  }
  for(iGroup in 1:numGroups) {
    alpha[iGroup] ~ dnorm(0, sd = sdGroups)
  }
})
  
```

**Vectorized declarations**  
create vector nodes. This means `fixed_effects[1:N]` will be a single node. One vector node vs. multiple scalar nodes give different model graphs, so use with care.

**Provide explicit index ranges**  
or use empty brackets `( )` and provide the `dimensions` argument to `nimbleModel()`.

**Nested indexing** is a good way to implement experimental groups or factor levels. If groups are known from the design, include them in `constants`.

## Using models

**Models can be compiled.**  
`cModel <- compileNimble(myModel)`  
In methods below, "model" can be `cModel` or `myModel`.

**Models can access and change variables.**  
`model$beta0 <- 5`  
`model[["beta0"]] <- 5`

**Models can simulate or calculate log-probabilities.**

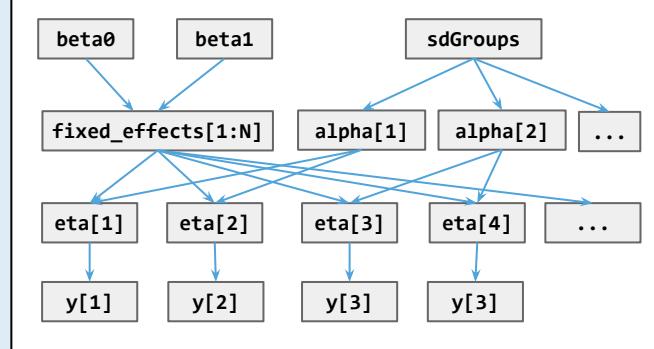
`model$calculate(nodes)`  
returns sum of log probability densities.

`model$calculateDiff(nodes)`  
returns difference in sum of log probability densities between current and previous node values.

`model$getLogProb(nodes)`  
returns sum of most recently calculated log probability densities.

`model$simulate(nodes,`  
`includeData = FALSE)`  
simulates into stochastic nodes.  
`includeData = FALSE` protects data.

**Models are graphs**



**Models know about nodes, variables and relationships.**

`model$getNodeNames()`  
returns node names  
e.g. "eta[1]", "eta[2]", ...

`model$getVarNames()`  
returns variable names  
e.g. "eta"

`model$expandNodeNames(nodes)`  
e.g. "y" is expanded to "y[1]", "y[2]", ...

`model$getDependencies(nodes, ...)`  
returns nodes that depend on input nodes.

**Uncompiled models can be debugged, updated, and copied.**

**Flag nodes as data and set inits**

`myModel$setData("y")`  
`myModel$setInits(inits)`

**Debug model errors**

`myModel$check()`  
check for missing/invalid values.

`myModel$initializeInfo()`  
which nodes are not fully initialized?

`myModel$checkBasics()`  
check for size/dimension mismatches and NA.

**Make a copy**

`myModel$newModel(replicate = TRUE)`

**Models know properties of nodes.**

`model$getDimension(node)`  
`model$getDistribution(nodes)`  
`model$isDeterm(nodes)`  
`model$isStoch(nodes)`  
`model$isNullData(nodes)`  
`model$Discrete(nodes)`  
`model$Multivariate(nodes)`  
`model$Binary(nodes)`  
`model$EndNode(nodes)`  
`model$Truncated(nodes)`

# nimble distributions and functions: : CHEAT SHEET



## Declarations

**STOCHASTIC**  
 $x \sim ddist(args)$

**DETERMINISTIC**  
 $z \leftarrow fn(args)$

**TRUNCATED STOCHASTIC**  
 $x \sim T(ddist(args), min, max)$

**CENSORED STOCHASTIC**  
 $seg \sim dinterval(t, c[1:nSegments])$   
 $t \sim ddist(args)$

**CONSTRAINT**  
 $one \sim dconstraint(condition)$

## Deterministic Functions

### SCALAR or COMPONENT-WISE

**Logical:** |, &, !, >, >=, <, <=, !=,  
 ==, equals, step

**Arithmetic:** +, -, \*, /, ^, pow(x, y)  
 %%, exp, log, sqrt, abs, cube

**Trigonometric:** sin, cos, tan, asin,  
 acos, atan, asinh, acosh, atanh

**Links:** logit, probit, cloglog  
*(links can also be used on left-hand side of a declaration)*

**Inverse links:** ilogit/expit,  
 iprobit/phi, icloglog

**Rounding:** ceiling, floor, round,  
 trunc

**Specials:** lgamma/loggam, besselK,  
 log1p, lfactorial, logfact

**Distributions:** d, p, q, r forms of available  
 distributions can be used as deterministic  
 functions.

### VECTOR and/or MATRIX

**Returning scalar:** inprod, logdet, sum,  
 mean, sd, prod, min, max

**Returning vector:** pmin, pmax,  
 eigen(x)\$values, svd(x)\$d

**Returning matrix:** inverse, chol, %\*%,  
 t, solve, forwardsolve,  
 backsolve, eigen(x)\$vectors,  
 svd(x)\$u, svd(x)\$v

**Write your own!**

See Ch 12 of  
User Manual

NIMBLE allows you to write **new distributions and functions** using nimbleFunction().

## Univariate Distributions

### Continuous



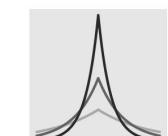
#### BETA

$y \sim dbeta([shape1, shape2 | mean, sd])$   
 $shape1 = mean^2 * (1 - mean) / sd^2 - mean$   
 $shape2 = mean * (1 - mean)^2 / sd^2 + mean - 1$



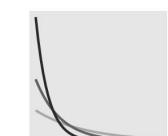
#### CHI-SQUARE

$y \sim dchisq(df)$



#### DOUBLE EXPONENTIAL (LAPLACE)

$y \sim ddexp(location, [scale|rate|var])$   
 $scale = 1/rate$   
 $scale = sqrt(var/2)$



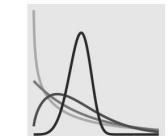
#### EXPONENTIAL

$y \sim dexp([rate|scale])$   
 $rate = 1/scale$



#### FLAT (improper)

$y \sim dflat()$



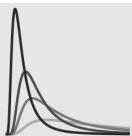
#### GAMMA

$y \sim dgamma([shape, [rate|scale] | [mean, sd]])$   
 $scale = 1/rate$   
 $shape = mean^2 / sd^2$   
 $scale = sd^2 / mean$



#### HALF FLAT (improper)

$y \sim dhalfflat()$



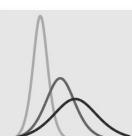
#### INVERSE GAMMA

$y \sim dinvgamma(shape, [rate|scale])$   
 $rate = 1/scale$



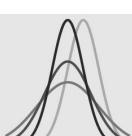
#### LOGISTIC

$y \sim dlogis(location, [rate|scale])$   
 $scale = 1/rate$



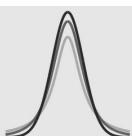
#### LOG-NORMAL

$y \sim dlnorm(meanlog, [taulog|sdlog|varlog])$   
 $sdlog = 1/sqrt(taulog)$   
 $sdlog = sqrt(varlog)$



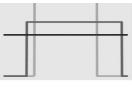
#### NORMAL

$y \sim dnorm(mean, [tau|sd|var])$   
 $sd = 1/sqrt(tau)$   
 $sd = sqrt(var)$



#### STUDENT T

$y \sim dt(mu, [tau|sigma|sigma2], df)$   
 $sigma = 1/sqrt(tau)$   
 $sigma = sqrt(sigma2)$



#### UNIFORM

$y \sim dunif(min, max)$



#### WEIBULL

$y \sim dweib(shape, [lambda|scale|rate])$   
 $scale = lambda^{-1/shape}$   
 $scale = 1/rate$

## DISTRIBUTION NAME

$y \sim ddist([default|alternative])$   
 canonical = fn(provided)

Lifted nodes are inserted when non-canonical parameters are used. Default parameters are not necessarily canonical.

## Discrete



#### BERNOULLI

$y \sim dbern(prob)$



#### BINOMIAL

$y \sim dbinom(prob, size)$



#### CATEGORICAL

$y \sim dcat(prob)$



#### NEGATIVE BINOMIAL

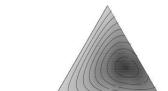
$y \sim dnegbin(prob, size)$



#### POISSON

$y \sim dpois(lambda)$

## Multivariate distributions



#### DIRICHLET

$y[] \sim ddirch(alpha[])$



#### MULTINOMIAL

$y[] \sim dmulti(prob[], size)$



#### MULTIVARIATE NORMAL

$y[] \sim dmnorm(mean[], [prec[,] | cov[,] | cholesky[,], prec_param])$



#### MULTIVARIATE STUDENT T

$y[] \sim dmvt(mu[], [prec[,] | scale[,] | cholesky[,], df, prec_param])$



cholesky = chol(prec) : prec\_param=1  
 cholesky = chol(cov) : prec\_param=0 for dmnorm  
 cholesky = chol(scale) : prec\_param=0 for dmvt  
 cholesky is chol(prec) when prec\_param=1,  
 chol(R) when prec\_param=0  
 cholesky = chol(prec) : prec\_param=1  
 cholesky = chol(cov) : prec\_param=0 for dmnorm  
 cholesky = chol(scale) : prec\_param=0 for dmvt  
 cholesky is chol(prec) when prec\_param=1,  
 chol(cov)|chol(scale) when prec\_param=0



#### WISHART

$y[,] \sim dwish([R[,] | S[,] | cholesky[,], df, scale_param])$



#### INVERSE WISHART

$y[,] \sim dinvwish([S[,] | R[,] | cholesky[,], df, scale_param])$

cholesky = chol(R): scale\_param=0  
 cholesky = chol(S): scale\_param=1  
 cholesky is chol(S) when scale\_param=1,  
 chol(R) when scale\_param=0



#### CONDITIONAL AUTOREGRESSIVE intrinsic (improper)

$y[] \sim dcar_normal(adj[], weights[], num[], tau, c, zero_mean)$



#### proper

See Ch 9 of User Manual  
 $y[] \sim dcar_proper(mu[], C[], adj[], num[], M[], tau, gamma)$

## Bayesian nonparametric distributions



#### CHINESE RESTAURANT PROCESS

$y[] \sim dCRP(conc, size)$   
 conc = concentration parameter



#### STICK BREAKING PROCESS

$y[] \sim stick_breaking(z[])$   
 $z = \text{vector of breaking points}$

# oSCR :: CHEAT SHEET



The oSCR package, pronounced “Oscar”, provides a set of functions for working with Spatial Capture Recapture (SCR) models.

## Getting the package

Package hosted on [GitHub](#)

```
library (devtools)
install_github("jaroyle/oSCR")
library(oSCR)
```

## Workflow

- Every model you run on oSCR has the following 4 basic steps.
- Modeled after [unmarked](#) workflow

### 1. Format the sampling data

- One file for each one:
- Spatial encounter histories
  - Detector information

### 2. Define and format the State Space

- Size and resolution of the state space
- Spatial covariates for density

### 3. Analyze the data - model fitting

- Likelihood based: use AIC to do model selection
- No need to use other packages, oSCR has helper functions to do the model selection.

### 4. Post processing model output for inference:

- This means that now that you have your parameters all you have to do is interpret your results!

## Modelling framework

### A. Single-session models

- Repeated sample occasions on a single population of individuals using a single array of traps.

### B. Multi-session models

- Data grouped in strata or groups which are independent in space or time.

### C. Explicit sex-structured models

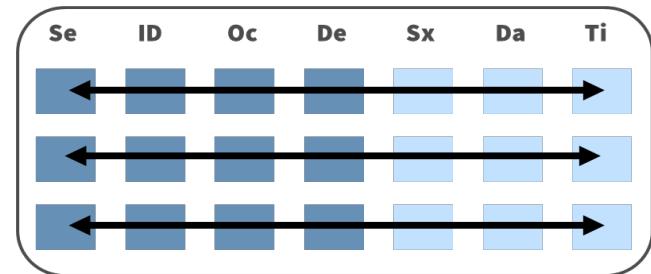
### D. Multi-session sex-structured models

## 1. Format sampling data

Before starting to use oSCR you need to format the datafiles in a scrFrame which consists of two basic spreadsheets: **edf** and **tdf**.

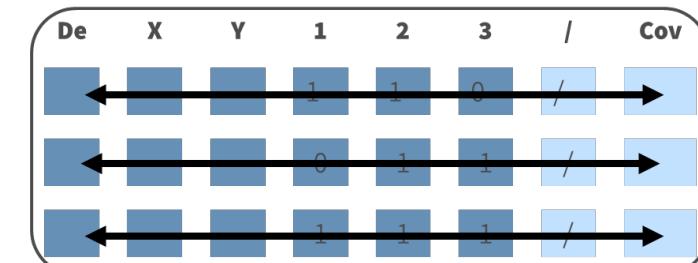
### 1.1 edf: encounter data file.

- Single **data frame**.
- Each row has individual detection events.
- Dark blue = required; light blue = optional.
- Columns contain capture information:
  - Session (Se)
  - Individual ID (ID)
  - Occasion (Oc)
  - Detector\* (De)
  - Sex (Sx)
  - Date (Da)
  - Time (Ti)



### 1.2 tdf: trap deployment data file.

- A **list** with information for each session (tdf1, tdf2, ...).
- Each row is a trap.
- Columns contain trap information
  - Detector\* (De)
  - X (required, UTM)
  - Y (required, UTM)
  - Binary trap operation data for malfunctions, rotations (required if problems were found;
  - 1, 2, 3, ... n)
  - Separator (e.g., /)
  - Trap level covariates (different column per covariate)



\*Notice that both edf and tdf have the same **Detector (De)** column that **MUST** match (same name, class, relational database).

**1.3 data2oscr()**: is a function that links **edf** and **tdf** files via the detector\* names. Creates **scrFrame**.

```
data <- data2oscr(
  edf,      # encounter data file
  tdf,      # list containing trap deployment file
  sess.col*, # session col number or name in edf
  id.col*, # individual ID col # or name in edf
  occ.col, # occasion col number or name in edf
  trap.col*, # detector col number or name in edf
  sex.col*, # sex col number or name in edf
  sex.nancode, # character for unknown sex in edf
  K,        # number of occasions
  ntraps,   # number of traps
  trapcov.names, # vector of un-numbered cov
  names
  tdf.sep) # separator (e.g., "/")
```

\* `which(colnames(edf) %in% "name of column in edf")`

### 1.4 Summary functions for scrFrame :

- scrFrame contains information from the **edf** and **tdf** via detector names.

```
sf<-data$scrFrame
```

**sf\$caphist** Array of individual-by-trap-by-occasion (n x J x K). Binary or counts.

**sf\$traps** Data frame containing at least trap ID and coordinates of traps. Best with UTM.

**sf\$indcovs** Sex data (0 female, 1 male) or any bivariate covariate. NAs allowed.

**sf\$trapCovs** List of session specific trap covariates. Row per trap, and column per covariate.

**sf\$sigCovs** A data frame of covariates that affect space use ( $\sigma$ ,  $\sigma$ ).

**sf\$trapOperation** A list of session specific information on trap operational data.

**sf\$occasions** A vector of number of occasions per session .

**sf\$mmdm** Mean maximum distance moved pooled across sessions.  $\frac{1}{2} mmdm \sim \sigma$

**sf\$mdm** Maximum distance moved pooled across sessions.

**\$telemetry** Telemetry object for fitting resource selection models.

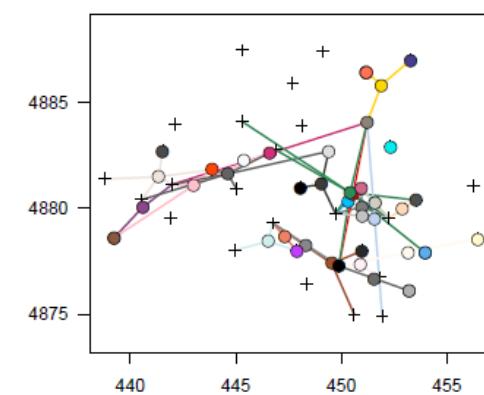
## 1.5 Summary of scrFrame

**sf**

S1	
n individuals	47
n traps	38
n occasions	8
S1	
avg caps	3.21
avg spatial caps	2.02
mmdm	4.65

## 1.6 Spatial captures per session

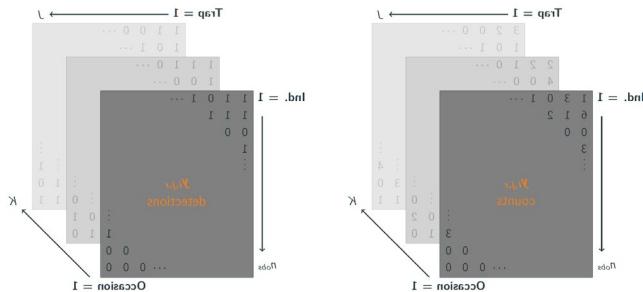
`plot(sf)` #y and x are UTM



# oSCR :: CHEAT SHEET



## 1.4.1 Navigating the scrFrame



### Capture history

- Session 1, all individuals, all traps, occasion 3  
`sf$caphist[[1]][ , , 3]`
- Session 1, individual 4, all traps, all occasions  
`sf$caphist[[1]][4, , ]`

### Traps

- Session 1 trap coordinates  
`sf$traps[[1]]`

### Trap covariates

- Trap covariate df session 1 occasion 4  
`sf$trapCovs[[1]][[4]]`

### Trap operation

- Session 1 trap trap operation matrix  
`sf$trapOperation [[1]]`

### Covariates that affect sigma ( $\sigma$ )

- These covariates are NOT session specific. This is a sessions=rows dataframe  
`sf$ sigCovs[[1]]`

### Vectors and single numbers

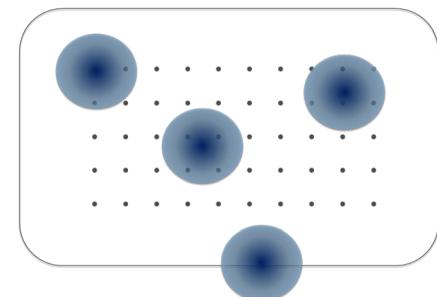
```
sf$ occasions
sf$mmdm
sf$mdm
```

## Datasets available

```
> data(package = "oSCR")
> data(ocelot)
> data("beardata")
> data("nybears")
> data("peromyscus")
> data("mink")
```

## 2. Create the State Space

The **State Space (S)** is the core element of SCR models. It defines where individuals can live and should represent activity centers of all sampled individuals.



### ssDF: the State Space Data Frame

- List with spatially explicit information from each session.
- At least include the coordinates (X, Y) of the discrete state space (UTM).
- Can include spatial covariates for a continuous state space to study variation in Density.
- Non habitat can be removed by removing unwanted coordinates (e.g., parking lot).

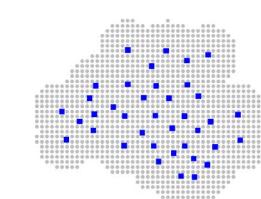


### 2.1. make.ssDF():

- Remember that  $\frac{1}{2} mmdm \sim \sigma$
  - Extracts covariates and removes non habitat
- ```
ss <- make.ssDF(scrFrame,
                  buffer, #~3 to 4σ around traps
                  res) # ≤ σ
```

### 2.2. Plot the state space

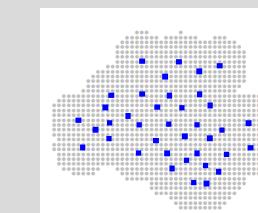
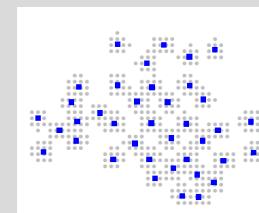
- Plot state space  
`plot(ss)`
- Plot state space & traps  
`plot(ss, sf)`



### Vary the buffer and/or resolution

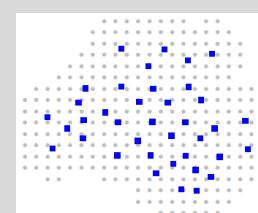
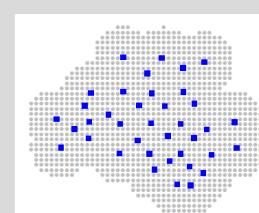
- ↗ Varying buffer, fixed resolution

```
make.ssDF(sf,
           buffer = 1,
           res = 0.5)
```



- ↖ Fixed buffer, varying resolution

```
make.ssDF(sf,
           buffer = 3,
           res = 0.1)
```



## 3. Fit the model

### 3.1. Single-session model: Fit the model with oSCR.fit():

```
sf <- data$scrFrame
mod <- oSCR.fit(model,
                  scrFrame, #sf
                  ssDF, ...)
```

- See pg. 3 for null model and multi-session models.

**model** is a list with 3 basic formulations:

```
list(D ~ 1, p0 ~ 1, sig ~ 1)
```

| Variation in... |                              |
|-----------------|------------------------------|
|                 |                              |
| <b>D</b>        | pixel density                |
| <b>p0</b>       | baseline encounter prob/rate |
| <b>sig</b>      | sigma ( $\sigma$ )           |

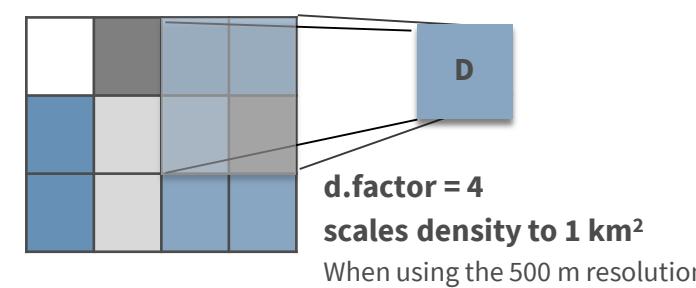
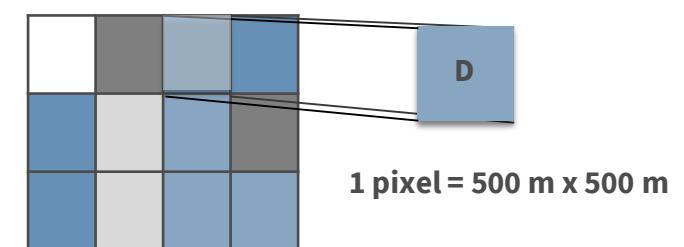
### 3.2. Backtransform to the real scale

```
get.real(model,
          newdata,
          d.factor,
          type)
```

|                 |                                                               |
|-----------------|---------------------------------------------------------------|
| <b>model</b>    | fitted model                                                  |
| <b>newdata</b>  | Optional new data object for predictions                      |
| <b>d.factor</b> | optional scale the estimates to a different resolution        |
| <b>type</b>     | density ("dens"), detection probability ("det"), sigma("sig") |

|               |                                                                             |
|---------------|-----------------------------------------------------------------------------|
| <b>"dens"</b> | Sex-specific estimates of density, and the density estimates are per pixel. |
| <b>"det"</b>  | Estimate of detection at distance from activity center = 0.                 |
| <b>"sig"</b>  | Estimates of the spatial scale of detection.                                |

### d.factor



# oSCR :: CHEAT SHEET



Page 3 describes the specific functions and workflow for the null model and multi-session model in the oSCR package.

## Model specifics

### Null model ( $SCR_0$ )

- The null model assumes homogeneous density which means all pixels have the same expected density.
- For additional arguments see `?oSCR.fit()`

```
mod1 <- oSCR.fit(list(D ~ 1,
p0 ~ 1, sig ~ 1),
scrFrame, #scrFrame object
ssDF, #ssDF object
... ) #other arguments
mod1 #summary
```



- If you included sex as a covariate in the scrFrame:
- Sex ratio psi() will be included in the summary
  - Can compare AIC with and without sex effects

### Multi-session model

Are your data organized in multi-sessions and you want to analyze all of them jointly?



**Spatial sessions:** different study areas (e.g., parks, trapping grids)



**Temporal sessions:** same areas different times (e.g. seasons, years)



Session specific **population size**  $N_g$  (g=group/session)

- Test for differences among sessions using AIC.
- Can share parameters among sessions or not.

- The **multi-session** model follows similar steps as the single session model.
- The **edf** files from multiple sessions may be merged into one data frame prior to `data2oscr`  
`edf <- rbind(edf1, edf2, ...)`
- The **tdf** files must be separate files for each session.

### 1. data2oscr for multi-session scrFrame

```
data <- data2oscr(
  edf, # include session column
  list(tdf1, tdf2, ...), # tdf files
  sess.col*, # session col in edf
  id.col*, # individual ID col in edf
  occ.col, # occasion col in edf
  trap.col*, # detector col in edf
  sex.col*, # sex col in edf
  sex.nancode, # unknown sex in edf
  K, # vector with occasions per session
  ntraps) # vector with traps per session
```

```
sf <- data$sf
```

```
sf # summary info per session (S1, S2..)
```

### 1.2. Summary of multi-session scrFrame

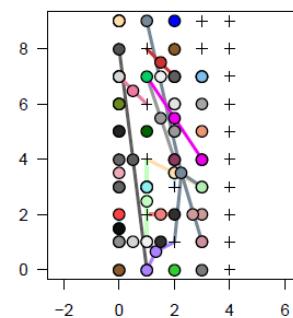
|                  | S1   | S2   | S3   | S4   |
|------------------|------|------|------|------|
| n individuals    | 77   | 60   | 108  | 54   |
| n traps          | 50   | 50   | 50   | 50   |
| n occasions      | 7    | 5    | 6    | 4    |
|                  | S1   | S2   | S3   | S4   |
| avg caps         | 1.91 | 1.47 | 1.71 | 1.37 |
| avg spatial caps | 1.30 | 1.15 | 1.27 | 1.13 |
| mmdm             | 2.57 | 2.32 | 1.76 | 2.84 |
| Pooled MMDM:     | 2.21 |      |      |      |

### 1.3. Plot spatial captures in a multi-session scrFrame

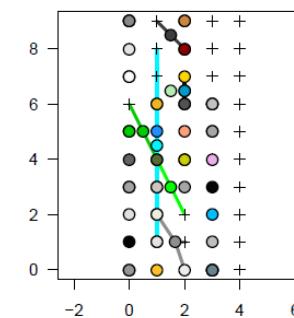
- Use `plot(sf)` to plot a spatial capture per session

```
par(mfrow=c(1,n)) # n = sessions
plot(sf) # plot all sessions
```

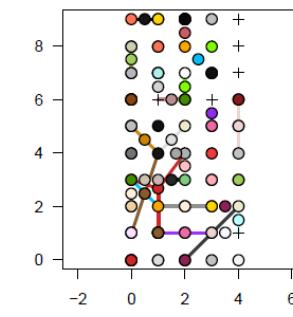
Session 1



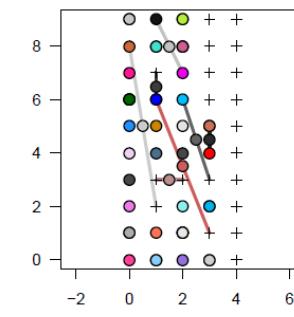
Session 2



Session 3



Session 4



### 2. Make the State Space Data Frame of a multi-session scrFrame

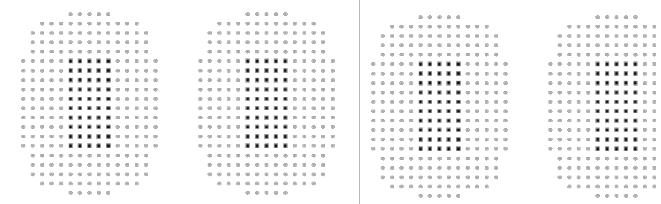
```
ss <- make.ssDF(
  scrFrame, # multi-session
  buffer, #buffer width
  res) #state space resolution
```

- You can vary the buffer and resolution as in the single-session model.

```
?make.ssDF() # Look at the help file for other arguments
```

- Visualize the state space

```
par(mfrow=c(1,n)) # n = sessions
plot.ssDF(ss, # state space
          sf) # traps
```



### 3. Model fitting

- Specify models that consider or not variation among sessions.
  - fixed vs. session specific **D**
  - fixed vs. session specific **p0**
  - fixed vs. session specific **space use ( $\sigma$ )**

| Model     | Algebra                                                  | In <code>oSCR.fit</code>   |
|-----------|----------------------------------------------------------|----------------------------|
| Density   | $\log(D_{(s_i)}) = \beta_0$                              | <code>D ~ 1</code>         |
| Density   | $\log(D_{(s_i)}) = \beta_0 + \beta_{1(g)} Session_g$     | <code>D ~ session</code>   |
| Detection | $\text{logit}(p_0) = \alpha_0$                           | <code>p0 ~ 1</code>        |
| Detection | $\text{logit}(p_0) = \alpha_0 + \alpha_{1(g)} Session_g$ | <code>p0 ~ session</code>  |
| Space use | $\log(\sigma) = \gamma_0$                                | <code>sig ~ 1</code>       |
| Space use | $\log(\sigma) = \gamma_0 + \gamma_{1(g)} Session_g$      | <code>sig ~ session</code> |

- Include all models into a list using `fitList.oSCR()`:
- ```
f1 <- fitList.oSCR(
  mods, # list of fitted models
  rename) # if TRUE models are renamed with sensible names
```
- Compare multiple models
  - `ms <- modSel.oSCR(f1)`
  - Generate an AIC table to compare multiple models
  - `ms$aic`
  - Generate a coefficient table
  - `ms$coef.tab`
  - Generate a model averaged coefficients
  - `ma <- ma.coef(ms) # include a modSel.oSCR object`

### 3.1. Back transform to the real scale

```
top.model <- m3
```

```
pred.df <- data.frame(session =
  factor (c(1, 2, 3, 4, ...)))
```

```
pred.det <- get.real(
  model = top.model, type = "det",
  newdata = pred.df)
```

# Get an Overview with overviewR: : CHEAT SHEET



## Generate Tables

**overview\_tab** generates a data frame that collapses the time condition for each id by taking into account potential gaps in the time frame

id	time	Var1	Var2
A	1990		
A	1991		
A	1992		
B	1990		

```
output_table <-  
  overview_tab(  
    dat = toydata,  
    id = ccode,  
    time = year)
```

**add data frame**  
**define your time and scope variables**

It also works with multiple time arguments (day, month, and year)

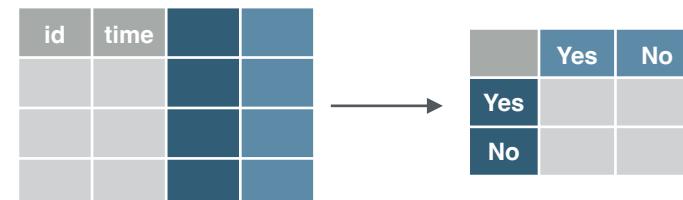
id	year	month	day	Var1	Var2
A	1990	3	1		
A	1990	3	2		
A	1990	3	3		
B	1991	4	10		

```
overview_tab(  
  dat = toydata,  
  id = ccode,  
  time = list(year = toydata$year,  
             month = toydata$month,  
             day = toydata$day),  
  complex_date = TRUE)
```

**define the time variables**  
**set argument complex\_date to**

**overview\_tab** and **overview\_na** can also handle **data.table** objects (all other functions currently only with **data.frames**) to increase the performance on larger data sets

**overview\_crosstab** generates a cross table that divides the data based on two conditions



```
output_crosstab <-  
  overview_crosstab(  
    dat = toydata,  
    cond1 = gdp,  
    cond2 = population,  
    threshold1 = 25000,  
    threshold2 = 27000,  
    id = ccode,  
    time = year  
)
```

**define your conditions with cond1 and cond2**  
**set your thresholds**

⚠ If a data set is used that has multiple observations on the id-time unit, the function automatically aggregates the data set using the mean of condition 1 (**cond1**) and condition 2 (**cond2**).

## Workflows

All **overviewR** functions can be easily integrated in the **tidyverse**.

```
toydata %>%  
  dplyr::filter(year > 1993) %>%  
  overview_na()
```

The example shows how to filter or wrangle data before calling **overview\_na**.

To change the visual appearance of plots, the user can rely on **ggplot2** layering logic.

## Export Results

### Tables

**overview\_latex** generates a LaTeX output (works with both **overview\_tab** and **overview\_crosstab** output)

```
overview_latex(  
  obj = output_table)
```

```
overview_latex(  
  obj = output_crosstab,  
  crosstab = TRUE)
```

**TRUE for cross tables**

The table can be modified with the **title**, **id**, **time**, **cond1**, and **cond2** arguments to replace default names

It also allows to save your output in a .tex file

```
overview_latex(  
  obj = output_table,  
  save_out = TRUE,  
  file_path = "path/output.tex")
```

**define where your output should**

We introduced **overview\_latex** (instead of **overview\_print**) and **file\_path** (instead of the separate **file** and **path** arguments) as of v0.0.12.

The outputs of **overview\_tab** and **overview\_crosstab** are also compatible with other packages and functions such as **xtable**, **flextable**, or **kable** from **knitr**.

To generate a table in RMarkdown with **knitr::kable**:

```
knitr::kable(output_table)
```

### Plots

As the plots are based on ggplot2, plots can be stored with **ggplot2::ggsave**

```
ggplot2::ggsave(  
  output_plot,  
  filename = "FILENAME.png")
```

**add plot object**  
**add filename**

Alternatively, storing the object also works this way:

```
png("FILENAME.png")  
output_plot  
dev.off()
```

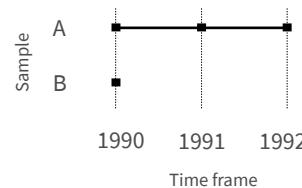
# Get an Overview with overviewR:: CHEAT SHEET



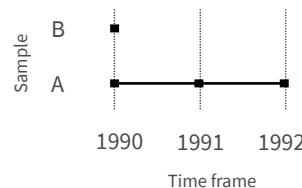
## Generate Plots

### Sample overview

**overview\_plot** illustrates the information that is generated in overview\_tab in a ggplot2 graphic

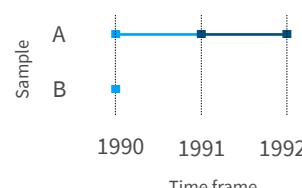


```
overview_plot(  
  dat = toydata,  
  id = ccode,  
  time = year)
```



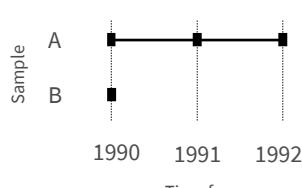
```
overview_plot(  
  dat = toydata,  
  id = ccode,  
  time = year,  
  asc = FALSE)
```

Reverse order of y-axis



```
overview_plot(  
  dat = toydata,  
  id = ccode,  
  time = year,  
  color = before)
```

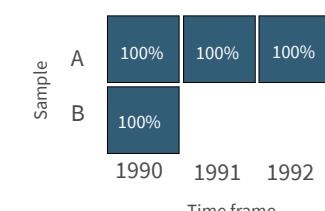
Add variable to identify time periods



```
overview_plot(  
  dat = toydata,  
  id = ccode,  
  time = year,  
  dot_size = 5)
```

change size of dots

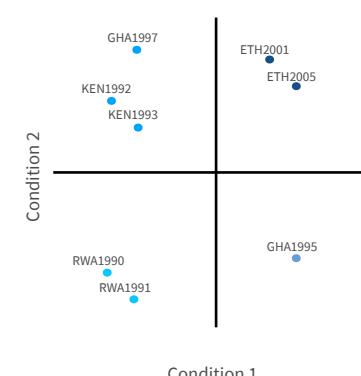
**overview\_heat** is similar to overview\_plot but presents the frequency of data points by id-time-unit in a heat map



```
overview_heat(  
  dat = toydata,  
  id = ccode,  
  time = year,  
  perc = TRUE,  
  exp_total = 12)
```

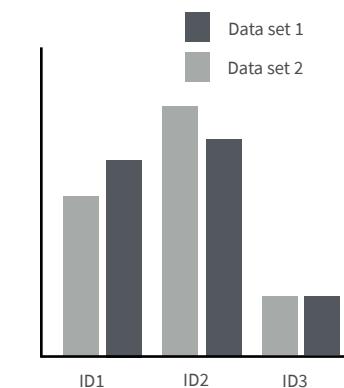
displays percentage  
max observations by id-

**overview\_crossplot** allows to illustrate similar information as presented in a cross table (overview\_crosstab)



```
overview_crossplot(  
  toydata,  
  id = ccode,  
  time = year,  
  cond1 = gdp,  
  cond2 = population,  
  threshold1 = 25000,  
  threshold2 = 27000,  
  color = TRUE,  
  label = TRUE  
)
```

**overview\_overlap** plots the overlap between two data sets



```
overview_overlap(  
  dat1 = toydata,  
  dat2 = toydata2,  
  dat1_id = ccode,  
  dat2_id = ccode,  
  plot_type = "bar"  
)
```

This is the default

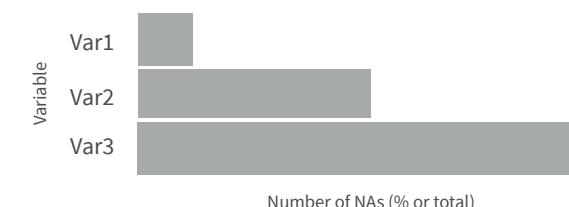


```
overview_overlap(  
  dat1 = toydata,  
  dat2 = toydata2,  
  dat1_id = ccode,  
  dat2_id = ccode,  
  plot_type = "venn"  
)
```

Represents a Venn diagramm

### Missing values (NAs)

**overview\_na** returns a horizontal ggplot2 bar plot that indicates the amount of missing data (NAs) for each variable (column-wise)



Number of NAs (% or total)

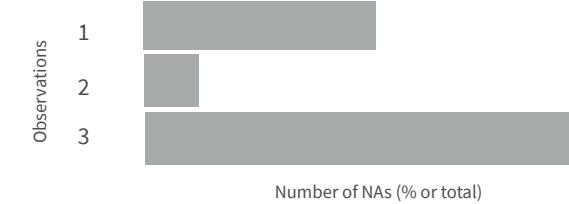
relative distribution

```
overview_na(toydata_with_na)
```

```
overview_na(toydata_with_na,  
           perc = FALSE)
```

FALSE gives total number (instead of percentage)

It also allows you to plot the NAs row-wise. This can be helpful when looking at survey data where each respondent is represented in a row. Using this function then helps to understand the share of unit or item non-responses.



Number of NAs (% or total)

Plot NAs row-wise

```
overview_na(toydata_with_na,  
           row_wise = TRUE)
```

```
overview_na(toydata_with_na,  
           row_wise = TRUE,  
           perc = FALSE)
```

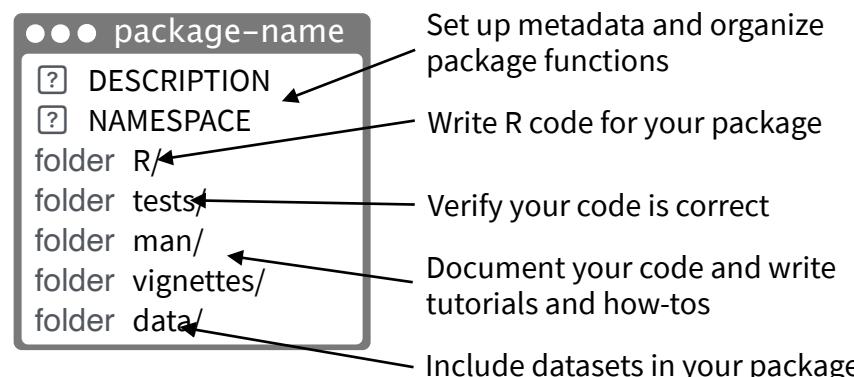
FALSE gives total number (instead of percentage)

# Package Development :: CHEATSHEET



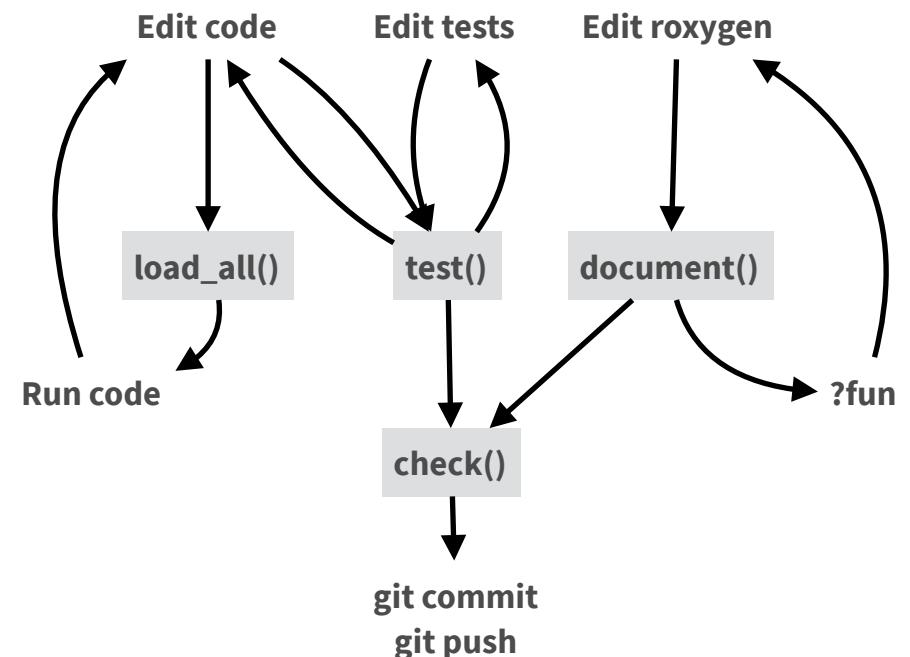
## Package Structure

A package is a convention for organizing files into directories. This cheat sheet shows how to work with the 7 most common parts of an R package:



There are multiple packages useful to package development, including **usethis** which handily automates many of the more repetitive tasks. Install and load **devtools**, which wraps together several of these packages to access everything in one step.

## Workflow



## Getting Started

### Once per machine:

- Get set up with **use\_devtools()** so **devtools** is always loaded in interactive R sessions

```
if (interactive()) {  
  require("devtools", quietly = TRUE)  
  # automatically attaches usethis  
}
```

- create\_github\_token()** — Set up GitHub credentials
- git\_vaccinate()** — Ignores common special files

### Once per package:

- create\_package()** — Create a project with package scaffolding
- use\_git()** — Activate git
- use\_github()** — Connect to GitHub
- use\_github\_action()** — Set up automated package checks

Having problems with git? Get a situation report with **git\_sitrep()**.

- load\_all()** (Ctrl/Cmd + Shift + L) — Load code
- document()** (Ctrl/Cmd + Shift + D) — Rebuild docs and NAMESPACE
- test()** (Ctrl/Cmd + Shift + T) — Run tests
- check()** (Ctrl/Cmd + Shift + E) — Check complete package

## folder R/

All of the R code in your package goes in folder R/. A package with just an R/ directory is still a very useful package.

- Create a new package project with **create\_package("path/to/name")**.
- Create R files with **use\_r("file-name")**.

- Follow the tidyverse style guide at [style.tidyverse.org](https://style.tidyverse.org)
- Click on a function and press **F2** to go to its definition
- Find a function or file with **Ctrl + .**

## DESCRIPTION

The **DESCRIPTION** file describes your work, sets up how your package will work with other packages, and applies a license.

- Pick a license with **use\_mit\_license()**, **use\_gpl3\_license()**, **use\_proprietary\_license()**.
- Add packages that you need with **use\_package()**.

**Import** packages that your package requires to work. R will install them when it installs your package.

**use\_package(x, type = "imports")**

**Suggest** packages that developers of your package need. Users can install or not, as they like.

**use\_package(x, type = "suggests")**

## NAMESPACE

The **NAMESPACE** file helps you make your package self-contained: it won't interfere with other packages, and other packages won't interfere with it.

- Export functions for users by placing **@export** in their roxygen comments.
- Use objects from other packages with **package::object** or **@importFrom package object** (recommended) or **@import package** (use with caution).
- Call **document()** to generate NAMESPACE and **load\_all()** to reload.

### DESCRIPTION

Makes **packages** available

Mandatory

**use\_package()**

### NAMESPACE

Makes **function** available

Optional (can use `::` instead)

**use\_import\_from()**

## folder man/

The documentation will become the help pages in your package.

- Document each function with a roxygen block above its definition in R/. In RStudio, Code > Insert Roxygen Skeleton helps (Ctrl/Cmd + Alt + Shift + R).
- Document each dataset with roxygen block above the name of the dataset in quotes.
- Document the package with `use_package_doc()`.
- Build documentation in folder man/ from Roxygen blocks with `document()`.

## folder vignettes/

- Create a vignette that is included with your package with `use_vignette()`.
- Create an article that only appears on the website with `use_article()`.
- Write the body of your vignettes in R Markdown.

## Websites with pkgdown



- Use GitHub and `use_pkdown_github_pages()` to set up pkdown and configures an automated workflow using GitHub Actions and Pages.
- If you're not using GitHub, call `use_pkdown()` to configure pkdown. Then build locally with `pkdown::build_site()`.

## folder tests/



- Set up test infrastructure with `use_testthat()`.
- Create a test file with `use_test()`.
- Write tests with `test_that()` and `expect_()`.
- Run all tests with `test()` and run tests for current file with `test_active_file()`.
- See coverage of all files with `test_coverage()` and see coverage of current file with `test_coverage_active_file()`.

### ROXYGEN2

The **roxygen2** package lets you write documentation inline in your .R files with shorthand syntax.

- Add roxygen documentation as comments beginning with `#'`.
- Place a roxygen `@` tag (right) after `#'` to supply a specific section of documentation.
- Untagged paragraphs will be used to generate a title, description, and details section (in that order).

```
#' Add together two numbers
#'
#' @param x A number.
#' @param y A number.
#' @returns The sum of `x` and `y`.
#' @export
#' @examples
#' add(1, 1)
add <- function(x, y) {
  x + y
}
```



### COMMON ROXYGEN TAGS

<code>@description</code>	<code>@family</code>	<code>@returns</code>
<code>@examples</code>	<code>@inheritParams</code>	<code>@seealso</code>
<code>@examplesIf</code>	<code>@param</code>	
<code>@export</code>	<code>@rdname</code>	

## README.Rmd + NEWS.md

- Create a README and NEWS markdown files with `use_readme_rmd()` and `use_news_md()`.

### Expect statement

#### `expect_equal()`

Is equal? (within numerical tolerance)

#### `expect_error()`

Throws specified error?

#### `expect_snapshot()`

Output is unchanged?

```
test_that("Math works", {
  expect_equal(1 + 1, 2)
  expect_equal(1 + 2, 3)
  expect_equal(1 + 3, 4)
})
```

## folder data/

- Record how a data set was prepared as an R script and save that script to folder data-raw/ with `use_data_raw()`.
- Save a prepared data object to folder data/ with `use_data()`.

## Package States

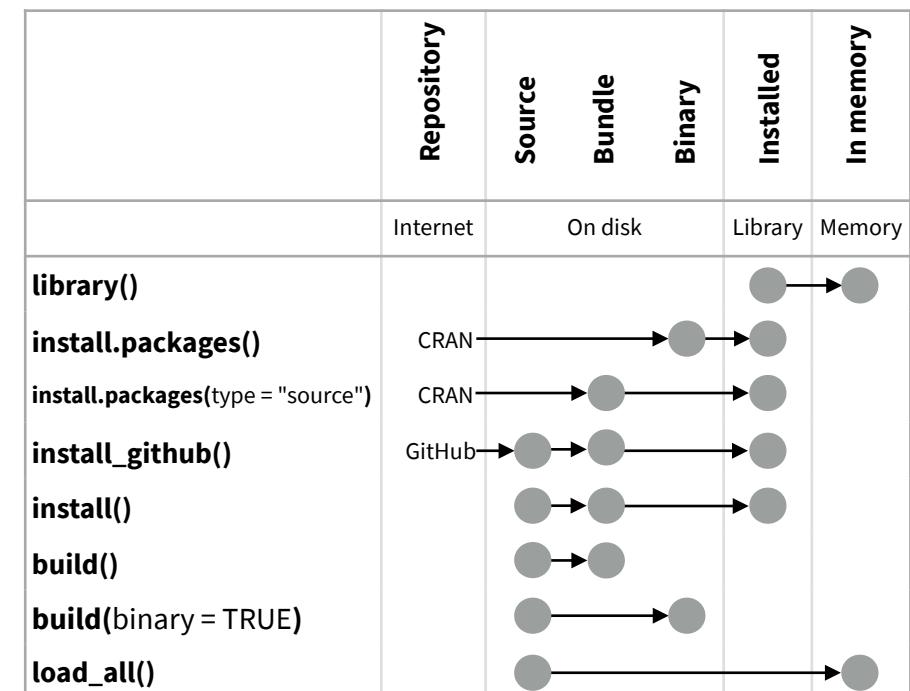
The contents of a package can be stored on disk as a:

- **source** - a directory with sub-directories (as shown in Package structure)
- **bundle** - a single compressed file (.tar.gz)
- **binary** - a single compressed file optimized for a specific OS

Packages exist in those states locally or remotely, e.g. on CRAN or on GitHub.

From those states, a package can be installed into an R library and then loaded into memory for use during an R session.

Use the functions below to move between these states.



Visit [r-pkgs.org](https://r-pkgs.org) to learn much more about writing and publishing packages for R.

# Searching CRAN with packagefinder:: CHEAT SHEET



## CONSOLE

```
findPackage(keywords, mode = "or", case.sensitive = FALSE, always.sensitive = NULL,  
weights = c(2,2,1,2), display = "viewer", results.longdesc = FALSE, limit.results = 15, silent =  
FALSE, index = NULL, advanced.ranking = TRUE, return.df = FALSE, clipboard = FALSE)
```

Most important arguments

keywords

Word or vector of words to search for

mode

Find packages with every keyword ("and") or with any of the keywords ("or")?  
Will be overruled if keywords contain logical operators like keywords = "X and Y"

case.sensitive

Case-sensitive search?

always.sensitive

Vector of words that will always be treated as case-sensitive, e.g. abbreviations

limit.results

How many results to display in console or viewer?

Outputs

display

Score	Name	Short Description	Link
100	xmld	Parse XML	16203
93	XML	Tools for Parsing and Generating XML Within R and S-Plus	16356
92.9	XML	Tools for Parsing and Generating XML Within R and S-Plus	16202
77.6	xmtr	Read, write and work with XML Data	16207
66	XML2R	Easier XML data collection	16204
65.7	xmldpcz	Implementation of the XML Procedure Call Protocol (XMLRPC)	16208
54.8	flatxml	Tools for working with XML Files as R DataFrames	4639

display = "viewer"

display = "console"

display = "browser"

return.df = TRUE Return results as dataframe

clipboard = TRUE Copy results to clipboard

Examples

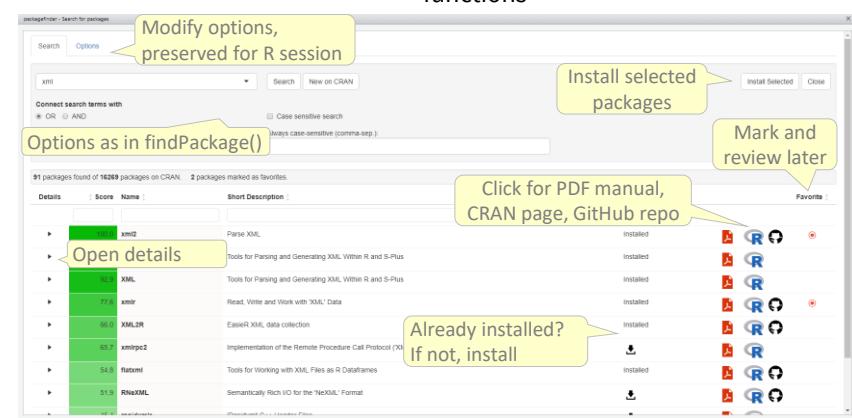
- > findPackage("parameters", mode = "and", always.sensitive = "SEM")
- > findPackage("meta and regression")
- > my.results <- findPackage(c("meta", "regression"), "and", return.df = TRUE)
- > findPackage("xml", display = "browser")

## RSTUDIO ADD-IN



Automatically installed with the package.

Provides a graphical interface to the findPackage() and whatsNew() functions



## ADDITIONAL FUNCTIONS

whatsNew(last.days = 0) Show new packages on CRAN

packageDetails(package) Show details of a CRAN package in the console

lastResults(package = "viewer") Show results of last search again

fp(...) Short hand for findPackage(...)

go(package, where.to = "details") Install CRAN package, show PDF manual, details or package website

# Parallel Computing :: CHEAT SHEET

## Splitting :

### Splitting a code by :

1. Task (different tasks on same data)
2. Data (one task on different data)

### Hardware needs :

CPU (+2 cores)

RAM (shared memory vs distributed memory)

## 2 ideas in parallel computing :

### 1. Map-Reduced Models :

(distributed data; physically on different devices)

- Hadoop
- Spark

### R Packages:

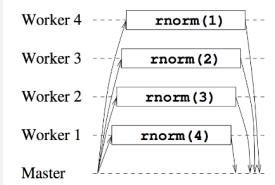
- sparklyr, iotools
- pbdr (programming with big data in R)

### 2. Master - Worker Models :

(M tasks on C cores; usually  $1 < C \ll M$ )

### R Packages:

- snow, snowFT, snowfall
- foreach
- future, future.apply



## Not always parallel computing:

stop/start cluster takes time

overhead (communication time b/w master and workers ; not good for repeatedly sending big data!)

## Sequential vs Parallel:

```
library(microbenchmark)
microbenchmark( FUN1(...), FUN2(...),
times = 10)
```

## parallel.R : core package

```
library(parallel)
ncores <- detectCores(logical=F) # physical cores
cl <- makeCluster(ncores)
clusterApply(cl, x = c(...), fun = FUN) # FUN(x,...)
stopCluster(cl)
```

## Initialization of workers :

```
clusterCall(cl,FUN) # calls FUN on workers
clusterEvalQ(cl, exp) # eval an exp. on workers
## clusterEvalQ(cl, library(foo))
clusterExport(cl, varlist) # varlist on workers
## clusterExport(cl, c("mean")) where mean = 10
```

## Data Chunk on workers :

1. generated on workers  
# clusterApply(cl,x, FUN) e.g FUN(){ rnorm()}
2. generated on master and pass to workers  
# ind <- splitIndices(200, 5)  
# clusterApply(cl, ind, FUN)  
# (-) : not efficient in Big Data : heavy
3. chunk on workers #copy of original Data on all workers  
# clusterExport(cl, M) e.g. M is a matrix  
# clusterApply(cl, x, FUN) FUN contains subset M

## foreach.R : Sequential

```
library(foreach) # by default return a list
foreach(n = rep(5,3), m = 10^(0:2)) %do% FUN(n,m)
foreach(n, .packages = "X") %do% FUN(n)
# FUN needs package X to be run
foreach(n, .export = c("Y")) %do% FUN(n,b=Y)
# FUN needs outside object/function "Y"
foreach(n,.combine = rbind) %do% FUN(n) #row bind
foreach(n,.combine = '+') %do% FUN(n) #rbind + colSum
foreach(n,.combine = c) %do% FUN(n) # vector
foreach(n,.combine = c) %:% when(n > 2) %do% FUN(n)
```

## future.R : asynchronously

```
library(future) (variables run as soon as created)
plan(multicore)
# plans : sequential, cluster, multicore, multiprocess
x <- mean(rnorm(100))
y <- mean(rnorm(100))
```

## future.apply.R : parallel\_apply

```
library(future.apply) (parallel _apply functions)
plan(multicore) # can be other plans
future_apply(n,FUN),future_lapply(...),future_sapply(...)
```

## foreach.R : Parallel

needs backend packages support parallel computing

- doParallel(parallel.R), doFuture (future.R), doSEQ

## doParallel.R : backend of foreach

```
library(doParallel)
cl <- makeCluster(ncores) # ncores = 2,3,...
registerDoParallel(cl) # register the backend
foreach(...) %dopar% FUN(...)
```

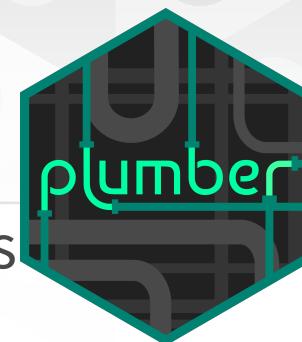
## doFuture.R : backend of foreach

```
library(doFuture)
registerDoFuture()
plan(cluster , workers = 3) # can be other plans
foreach(...) %dopar% FUN(...)
```

## Load Balancing: for uneven task times

```
clusterApplyLB(cl,x,FUN) # not for small task time
clusterApply(cl, x = splitIndices(10,2), FUN)
library(iertools)
foreach(s=isplitVector(1:10, chunks =2))%dopar% FUN
# e.g. FUN = sapply(s,"*",100)
future_sapply(..., future.scheduling = 1)
```

# REST APIs with plumber: : CHEATSHEET



## Introduction to REST APIs

Web APIs use **HTTP** to communicate between **client** and **server**.

### HTTP



HTTP is built around a **request** and a **response**. A **client** makes a request to a **server**, which handles the request and provides a response. Requests and responses are specially formatted text containing details and data about the exchange between client and server.

### REQUEST

**HTTP Method**: curl -v "http://httpbin.org/get"  
#> GET / get HTTP/1.1  
#> Host: httpbin.org  
#> User-Agent: curl/7.55.1  
#> Accept: \*/\*  
#  
# Request Body

**Headers**: Path

**Message body**

### RESPONSE

**HTTP Version**: #< HTTP/1.1 200 OK  
**Status code**: Reason phrase  
#< Connection: keep-alive  
#< Date: Thu, 02 Aug 2018 18:22:22 GMT  
**Headers**:  
#< # Response Body

**Message body**

## Plumber: Build APIs with R

Plumber uses special comments to turn any arbitrary R code into API endpoints. The example below defines a function that takes the **msg** argument and returns it embedded in additional text.

**Plumber comments begin with #\***

```
library(plumber)
#* @apiTitle Plumber Example API
#* Echo back the input
#* @param msg The message to echo
#* @get /echo
function(msg = "") {
  list(
    msg = paste0(
      "The message is: '", msg, "'"))
}
```

**HTTP Method**: /<path> is used to define the location of the endpoint

**@decorators define API characteristics**

## Plumber pipeline

Plumber endpoints contain R code that is executed in response to an HTTP request. Incoming requests pass through a set of mechanisms before a response is returned to the client.

### FILTERS

Filters can forward requests (after potentially mutating them), throw errors, or return a response without forwarding the request. Filters are defined similarly to endpoints using the `@filter [name]` tag. By default, filters apply to all endpoints. Endpoints can opt out of filters using the `@preempt` tag.

### PARSER

parsers determine how Plumber parses the incoming request body. By default Plumber parses the request body as JavaScript Object Notation (JSON). Other parsers, including custom parsers, are identified using the `@parser [parser name]` tag. All registered parsers can be viewed with `registered_parsers()`.

### ENDPOINT

Endpoints define the R code that is executed in response to incoming requests. These endpoints correspond to HTTP methods and respond to incoming requests that match the defined method.

### METHODS

- `@get` - request a resource
- `@post` - send data in body
- `@put` - store / update data
- `@delete` - delete resource
- `@head` - no request body
- `@options` - describe options
- `@patch` - partial changes
- `@use` - use all methods

### SERIALIZER

Serializers determine how Plumber returns results to the client. By default Plumber serializes the R object returned into JavaScript Object Notation (JSON). Other serializers, including custom serializers, are identified using the `@serializer [serializer name]` tag. All registered serializers can be viewed with `registered_serializers()`.

### Identify as filter

### Forward

### Endpoint description

### Parser

### HTTP Method

### library(plumber)

```
#* @filter log
function(req, res) {
  print(req$HTTP_USER_AGENT)
  forward()
}
```

#\* Convert request body to uppercase

#\* @preempt log

#\* @parser json

#\* @post /uppercase

#\* @serializer json

```
function(req, res) {
  toupper(req$body)
}
```

### Filter name

### Opt out of the log

### Endpoint path

### Serializer

## Running Plumber APIs

Plumber APIs can be run programmatically from within an R session.

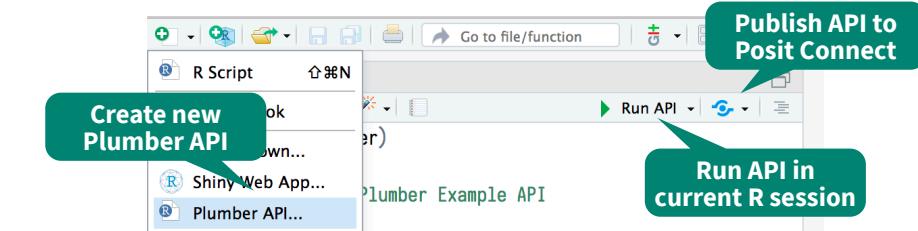
```
library(plumber)
plumb("plumber.R") %>%
  pr_run(port = 5762)
```

Path to API definition

Specify API port

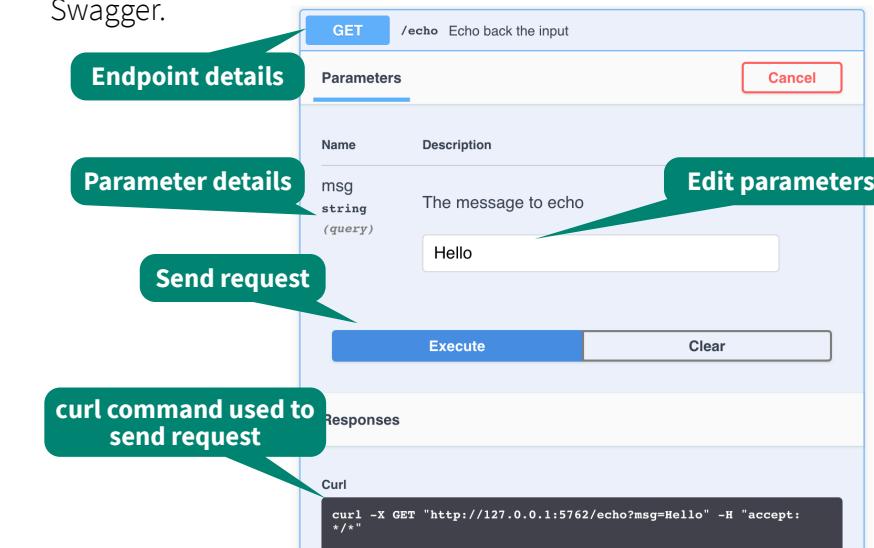
This runs the API on the host machine supported by the current R session.

### IDE INTEGRATION



## Documentation

Plumber APIs automatically generate an OpenAPI specification file. This specification file can be interpreted to generate a dynamic user-interface for the API. The default interface is generated via Swagger.



## Interact with the API

Once the API is running, it can be interacted with using any HTTP client. Note that using `httr` requires using a separate R session from the one serving the API.

```
(resp <- httr::GET("localhost:5762/echo?msg=Hello"))
#> Response [http://localhost:5762/echo?msg=Hello]
#>   Date: 2018-08-07 20:06
#>   Status: 200
#>   Content-Type: application/json
#>   Size: 35 B
httr::content(resp, as = "text")
#> [1] "{ \"msg\": [\"The message is: 'Hello'\"]}"
```

# Programmatic Plumber

## Tidy Plumber

Plumber is exceptionally customizable. In addition to using special comments to create APIs, APIs can be created entirely programmatically. This exposes additional features and functionality. Plumber has a convenient “tidy” interface that allows API routers to be built piece by piece. The following example is part of a standard `plumber.R` file.

```
library(plumber)

#* @plumber
function(pr) {
  pr %>%
    pr_get(path = "/echo",
           handler = function(msg = "") {
             list(msg = paste0(
               "The message is: '", msg, "'"))
           }) %>%
    pr_get(path = "/plot",
           handler = function() {
             rand <- rnorm(100)
             hist(rand)
           },
           serializer = serializer_png()) %>%
    pr_post(path = "/sum",
           handler = function(a, b) {
             as.numeric(a) + as.numeric(b)
           })
}
```

## OpenAPI

Plumber automatically creates an OpenAPI specification file based on Plumber comments. This file can be further modified using `pr_set_api_spec()` with either a function that modifies the existing specification or a path to a `.yaml` or `.json` specification file.

```
library(plumber)

#* @param msg The message to echo
#* @get /echo
function(msg = "") {
  list(
    msg = paste0(
      "The message is: '", msg, "'"))
}

#* @plumber
function(pr) {
  pr %>%
    pr_set_api_spec(function(spec) {
      spec$paths[["/echo"]る$get$summary <-
        "Echo back the input"
      spec
    })
}
```

By default, Swagger is used to interpret the OpenAPI specification file and generate the user interface for the API. Other interpreters can be used to adjust the look and feel of the user interface via `pr_set_docs()`.



# Advanced Plumber

## REQUEST and RESPONSE

Plumber provides access to special `req` and `res` objects that can be passed to Plumber functions. These objects provide access to the request submitted by the client and the response that will be sent to the client. Each object has several components, the most helpful of which are outlined below:

Name	Example	Description
<code>req</code>		
<code>req\$pr</code>	<code>plumber::pr()</code>	The Plumber router processing the request
<code>req\$body</code>	<code>list(a=1)</code>	Typically the same as <code>argsBody</code>
<code>req\$argsBody</code>	<code>list(a=1)</code>	The parsed body output
<code>req\$argsPath</code>	<code>list(c=3)</code>	The values of the path arguments
<code>req\$argsQuery</code>	<code>list(e=5)</code>	The parsed output from <code>req\$QUERY_STRING</code>
<code>req\$cookies</code>	<code>list(cook = "a")</code>	A list of cookies
<code>req\$REQUEST_METHOD</code>	"GET"	The method used for the HTTP request
<code>req\$PATH_INFO</code>	"/"	The path of the incoming HTTP request
<code>req\$HTTP_*</code>	"HTTP_USER_AGENT"	All of the HTTP headers sent with the request
<code>req\$bodyRaw</code>	<code>charToRaw("a=1")</code>	The <code>raw()</code> contents of the request body
<code>res</code>		
<code>res\$headers</code>	<code>list(header = "abc")</code>	HTTP headers to include in the response
<code>res\$setHeader()</code>	<code>setHeader("foo", "bar")</code>	Sets an HTTP header
<code>res\$setCookie()</code>	<code>setCookie("foo", "bar")</code>	Sets an HTTP cookie on the client
<code>res\$removeCookie</code>	<code>removeCookie("foo")</code>	Removes an HTTP cookie
<code>res\$body</code>	"{"a": [1]}"	Serialized output
<code>res\$status</code>	200	The response HTTP status code
<code>res\$toResponse()</code>	<code>toResponse()</code>	A list of status, headers, and body

## ASYNC PLUMBER

Plumber supports asynchronous execution via the `future` R package. This pattern allows Plumber to concurrently process multiple requests.

```
library(plumber)
future::plan("multisession")

#* @get /slow
function() {
  promises::future_promise({
    slow_calc()
  })
}
```



Set the execution plan

Slow calculation

## MOUNTING ROUTERS

Plumber routers can be combined by mounting routers into other routers. This can be beneficial when building routers that involve several different endpoints and you want to break each component out into a separate router. These separate routers can even be separate files loaded using `plumb()`.

```
library(plumber)
route <- pr() %>%
  pr_get("/foo", function() "foo")

#* @plumber
function(pr) {
  pr %>%
    pr_mount("/bar", route)
}
```

Create an initial router

Mount one router into another

In the above example, the final route is `/bar/foo`.

## RUNNING EXAMPLES

Some packages, like the Plumber package itself, may include example Plumber APIs. Available APIs can be viewed using `available_apis()`. These example APIs can be run with `plumb_api()` combined with `pr_run()`.

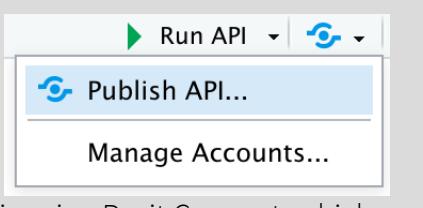
```
library(plumber)
plumb_api(package = "plumber",
          name = "01-append",
          edit = TRUE) %>%
  pr_run()
```

Identify the package name and API name

Optionally open the file for editing

Run the example API

# Deploying Plumber APIs



Once Plumber APIs have been developed, they often need to be deployed somewhere to be useful. Plumber APIs can be deployed in a variety of different ways. One of the easiest way to deploy Plumber APIs is using Posit Connect, which supports push button publishing from the RStudio IDE.



## 5 Publish your Content to Posit Connect

### Supported Content Types on Posit Connect

#### Documents



#### Applications



#### APIs



Used from Tableau workbooks to make real-time requests from Tableau to your Python and R code.

### Methods for Publishing to Posit Connect



Publish content directly from R using the rsconnect R package



Publish content using the CLI within the rsconnect-python package



Publish content with the click of a button using push button deployment



Publish directly from a git repository



### Need Help? Want to learn More?

Questions about sales and licensing → [sales@posit.co](mailto:sales@posit.co)

Technical Issues → [support@posit.co](mailto:support@posit.co)

Other questions? → [info@posit.co](mailto:info@posit.co)

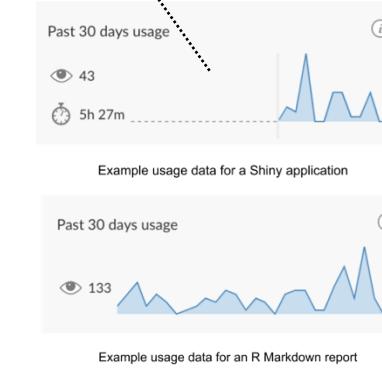
## 6 Share and Control your Content on Posit Connect

Content controls are accessed by clicking the icon

### Info and Content

#### Metadata

- Change content title
- Content information
- Add thumbnail
- Usage data



More usage info here:  
[pos.it/cookbook](#)

### Access Controls

Control who has access to your content

**SHARING**

Anyone - no login required  
 All users - login required  
 Specific users or groups  
 Who can view or change this document  
 Ryan Johnson ryan@posit.co

Share your content with a customized URL

**CONTENT URL**

Path: /public/awesome\_quarto/  
URL: [https://pub.current.posit.team/public/aweso...me\\_quarto/](https://pub.current.posit.team/public/aweso...me_quarto/)

### posit Team

Posit Team docs → <https://docs.posit.co>

Release notes → <https://docs.posit.co/release-notes.html>

### Runtime Settings

Tune and scale **applications** and **APIs** by modifying the number of processes (e.g., Python and R sessions) and connections per process

Min processes: 0 – no minimum, 1,   
Max processes: 3–, 5,   
Max connections per process: 20,

In this above configuration, the content can accommodate up to 100 concurrent connections (20 x 5). There will also be at least one process always running, meaning the content never goes to sleep

### Other Runtime Settings

Easily modify how your content runs by modifying any of the below settings. **Note:** some options may not be available depending on how Posit Connect is configured in your environment

PROCESS EXECUTION	CPU & RAM SETTINGS
Who runs this content on the server	Initial Number of CPUs: 0 – none requested, <input type="button" value="enter override"/> Max Number of CPUs: 0 – no limit, 1.5, <input type="button" value="x"/> Initial RAM Requested (GiB): 0 – none requested, <input type="button" value="enter override"/> Max RAM (GiB): 0 – no limit, 10, <input type="button" value="x"/>
EXECUTION ENVIRONMENT	PYTHON ENVIRONMENT MANAGEMENT
Last time: posit/dev/connect:latest Next time: Automatic (default)	Last time: Unknown Next time: Disabled
GPU SETTINGS	R ENVIRONMENT MANAGEMENT
AMD GPUs Requested: 0 – none requested, 1, <input type="button" value="x"/> Nvidia GPUs Requested: 0 – none requested, <input type="button" value="enter override"/>	Last time: Disabled Next time: Disabled

### Schedule and Emailing

**Documents** can be configured to execute on a schedule

Timezone: (GMT-05:00) America - Lima  
Start date & time: Sep 9, 2024 10 : 54 am pm  
Reset to local time  
Schedule type: Daily  
 Run every 1 day.  
 Run every weekday (Monday to Friday).  
 Publish output after it is generated  
 Send email after update

Schedule type options

- By Minute
- Hourly
- Daily
- Weekly
- Semimonthly
- Monthly
- Yearly

Email configuration options

Owners are always notified unless they opt-out

Send to all collaborators

Send to all viewers

Additional Recipients:

**Tip:** Learn how to send custom and conditional emails: [pos.it/rmd\\_email](#) & [pos.it/qmd\\_email](#)

### Tags

**Tags** make content organization, discovery, and filtering easier across the Connect dashboard

Technology: Deep Learning, Output Types, Email, BackEnds, Dashboards, Scheduled

Projects and Presentations: Calendars, QuickStart, Production Webinar

### Environment Variables

#### Environment Variables

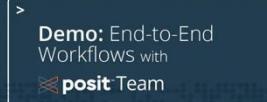
Note: Do not wrap your text in quotation marks; all symbols become part of the value available to your code.

Name:

Value:

Add Variable

Securely pass configuration options to your content as environment variables including **API Keys** and **Passwords**



LAST WEDNESDAY OF EVERY MONTH

# Apply functions with purrr :: CHEATSHEET

## Map Functions



### ONE LIST

**map(.x, .f, ...)** Apply a function to each element of a list or vector, and return a list.  
x <- list(a = 1:10, b = 11:20, c = 21:30)  
l1 <- list(x = c("a", "b"), y = c("c", "d"))  
map(l1, sort, decreasing = TRUE)



**map\_dbl(.x, .f, ...)**  
Return a double vector.  
map\_dbl(x, mean)

**map\_int(.x, .f, ...)**  
Return an integer vector.  
map\_int(x, length)

**map\_chr(.x, .f, ...)**  
Return a character vector.  
map\_chr(l1, paste, collapse = "")

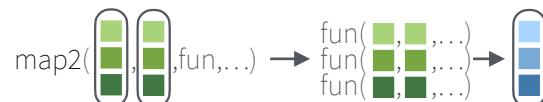
**map\_lgl(.x, .f, ...)**  
Return a logical vector.  
map\_lgl(x, is.integer)

**map\_vec(.x, .f, ...)**  
Return a vector that is of the simplest common type.  
map\_vec(l1, paste, collapse = "")

**walk(.x, .f, ...)** Trigger side effects, return invisibly.  
walk(x, print)

### TWO LISTS

**map2(.x, .y, .f, ...)** Apply a function to pairs of elements from two lists or vectors, return a list.  
y <- list(1, 2, 3); z <- list(4, 5, 6); l2 <- list(x = "a", y = "z")  
map2(x, y, \((x, y) x^\* y)



**map2\_dbl(.x, .y, .f, ...)** Return a double vector.  
map2\_dbl(y, z, ~.x / .y)

**map2\_int(.x, .y, .f, ...)** Return an integer vector.  
map2\_int(y, z, `+`)

**map2\_chr(.x, .y, .f, ...)** Return a character vector.  
map2\_chr(l1, l2, paste, collapse = "", sep = ":")

**map2\_lgl(.x, .y, .f, ...)** Return a logical vector.  
map2\_lgl(l2, l1, `%in%`)

**map2\_vec(.x, .f, ...)** Return a vector that is of the simplest common type.  
map2\_vec(l1, l2, paste, collapse = "", sep = ".")

**walk2(.x, .y, .f, ...)** Trigger side effects, return invisibly.  
walk2(objs, paths, save)

### MANY LISTS

**pmap(.l, .f, ...)** Apply a function to groups of elements from a list of lists or vectors, return a list.  
pmap(  
list(x, y, z),  
function(first, second, third) first \* (second + third)  
)



**pmap\_dbl(.l, .f, ...)** Return a double vector.  
pmap\_dbl(list(y, z), ~.x / .y)

**pmap\_int(.l, .f, ...)** Return an integer vector.  
pmap\_int(list(y, z), `+`)

**pmap\_chr(.l, .f, ...)** Return a character vector.  
pmap\_chr(list(l1, l2), paste, collapse = "", sep = ":")

**pmap\_lgl(.l, .f, ...)** Return a logical vector.  
pmap\_lgl(list(l2, l1), `%in%`)

**pmap\_vec(.l, .f, ...)** Return a vector that is of the simplest common type.  
pmap\_vec(list(l1, l2), paste, collapse = "", sep = ".")

**pwalk(.l, .f, ...)** Trigger side effects, return invisibly.  
pwalk(list(objs, paths), save)

## Function Shortcuts

Use `\(x)` with functions like **map()** that have single arguments.

**map(l, \(\(x) x + 2)**  
becomes  
**map(l, function(x) x + 2)**

Use `\(x, y)` with functions like **map2()** that have two arguments.

**map2(l, p, \(\(x, y) x + y)**  
becomes  
**map2(l, p, function(l, p) l + p)**

Use `\(x, y, z)` etc with functions like **pmap()** that have many arguments.

**pmap(list(x, y, z), \(\(x, y, z) x + y / z)**  
becomes  
**pmap(list(x, y, z), function(x, y, z) x \* (y + z))**

Use `\(x, y)` with functions like **imap()**. `.x` will get the list value and `.y` will get the index, or name if available.

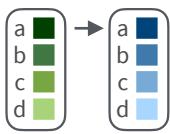
**imap(list("a", "b", "c"), \(\(x, y) paste0(y, ":", x))**  
outputs "index: value" for each item

Use a **string** or an **integer** with any map function to index list elements by name or position. **map(l, "name")** becomes **map(l, function(x) x[["name"]])**

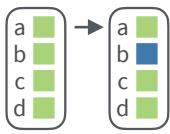


## Vectors

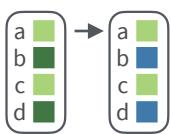
### Modify



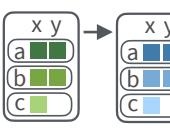
**modify(x, .f, ...)** Apply a function to each element. Also **modify2()**, and **imodify()**.  
modify(x, ~.+ 2)



**modify\_at(x, .at, .f, ...)** Apply a function to selected elements. Also **map\_at()**.  
modify\_at(x, "b", ~.+ 2)



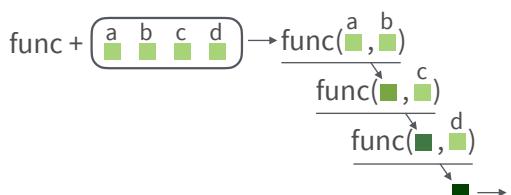
**modify\_if(x, .p, .f, ...)** Apply a function to elements that pass a test. Also **map\_if()**.  
modify\_if(x, is.numeric, ~.+ 2)



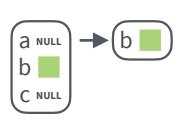
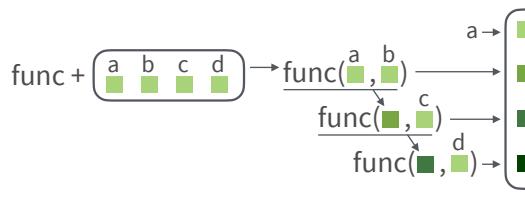
**modify\_depth(x, .depth, .f, ...)** Apply function to each element at a given level of a list. Also **map\_depth()**.  
modify\_depth(x, 1, ~.+ 2)

### Reduce

**reduce(x, .f, ..., .init, .dir = c("forward", "backward"))**  
Apply function recursively to each element of a list or vector. Also **reduce2()**.  
reduce(x, sum)



**accumulate(x, .f, ..., .init)** Reduce a list, but also return intermediate results. Also **accumulate2()**.  
accumulate(x, sum)



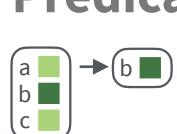
**compact(x, .p = identity)**  
Discard empty elements.  
compact(x)



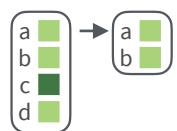
**keep\_at(x, at)**  
Keep/discard elements based by name or position.  
Conversely, **discard\_at()**.  
keep\_at(x, "a")



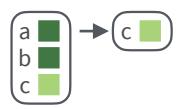
**set\_names(x, nm = x)**  
Set the names of a vector/list directly or with a function.  
set\_names(x, c("p", "q", "r"))  
set\_names(x, tolower)



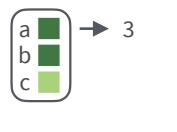
**keep(x, .p, ...)**  
Keep elements that pass a logical test.  
Conversely, **discard()**.  
keep(x, is.numeric)



**head\_while(x, .p, ...)**  
Return head elements until one does not pass.  
Also **tail\_while()**.  
head\_while(x, is.character)



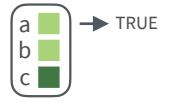
**detect(x, .f, ..., dir = c("forward", "backward"), .right = NULL, .default = NULL)** Find first element to pass.  
detect(x, is.character)



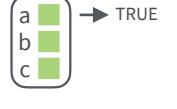
**detect\_index(x, .f, ..., dir = c("forward", "backward"), .right = NULL)** Find index of first element to pass.  
detect\_index(x, is.character)



**every(x, .p, ...)**  
Do all elements pass a test?  
every(x, is.character)



**some(x, .p, ...)**  
Do some elements pass a test?  
some(x, is.character)

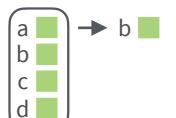


**none(x, .p, ...)**  
Do no elements pass a test?  
none(x, is.character)

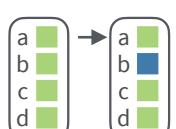


**has\_element(x, y)**  
Does a list contain an element?  
has\_element(x, "foo")

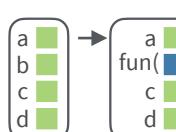
### Pluck



**pluck(x, ..., .default=NULL)**  
Select an element by name or index. Also **attr\_getter()** and **chuck()**.  
pluck(x, "b")  
x |> pluck("b")



**assign\_in(x, where, value)**  
Assign a value to a location using pluck selection.  
assign\_in(x, "b", 5)  
x |> assign\_in("b", 5)



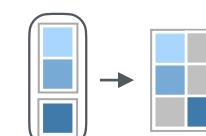
**modify\_in(x, .where, .f)** Apply a function to a value at a selected location.  
modify\_in(x, "b", abs)  
x |> modify\_in("b", abs)

### Concatenate

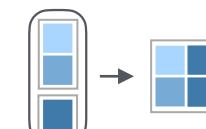
x1 <- list(a = 1, b = 2, c = 3)  
x2 <- list(  
  a = data.frame(x = 1:2),  
  b = data.frame(y = "a")  
)



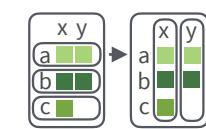
**list\_c(x)** Combines elements into a vector by concatenating them together.  
list\_c(x1)



**list\_rbind(x)** Combines elements into a data frame by row-binding them together.  
list\_rbind(x2)



**list\_cbind(x)** Combines elements into a data frame by column-binding them together.  
list\_cbind(x2)



**list\_transpose(l, .names = NULL)**  
Transposes the index order in a multi-level list.  
list\_transpose(x)

### Reshape



**list\_flatten(x)** Remove a level of indexes from a list.  
list\_flatten(x)



**map()**, **map2()**, or **pmap()** return lists and will create new list-columns.



Suffixed map functions like **map\_int()** return an atomic data type and will simplify list-columns into regular columns.



max	seq
3	<int [3]>
4	<int [4]>
5	<int [5]>

**List-columns** are columns of a data frame where each element is a list or vector instead of an atomic value. Columns can also be lists of data frames. See **tidyverse** for more about nested data and list columns.

### List-Columns

Manipulate list-columns like any other kind of column, using **dplyr** functions like **mutate()**. Because each element is a list, use **map functions** within a column function to manipulate each element.

# quanteda Cheat Sheet

Quantitative Analysis of Textual Data

## General syntax

- **corpus\_\*** manage text collections/metadata
- **tokens\_\*** create/modify tokenized texts
- **dfm\_\*** create/modify doc-feature matrices
- **fcm\_\*** work with co-occurrence matrices
- **textstat\_\*** calculate text-based statistics
- **textmodel\_\*** fit (un-)supervised models
- **textplot\_\*** create text-based visualizations

### Consistent grammar:

- **object()** constructor for the object type
- **object\_verb()** inputs & returns object type

## Extensions

quanteda works well with these companion packages:

- **readtext**: an easy way to read text data
- **spacyr**: NLP using the spaCy library
- **quanteda.corpora**: additional text corpora
- **stopwords**: multilingual stopword lists in R
- **quanteda.[textstats/textmodels/textplots]**: text analysis packages

## Create a corpus from texts (corpus\_\*)

### Read texts (txt, pdf, csv, doc, docx, json, xml)

```
my_texts <- readtext::readtext("~/link/to/path/*")
```

### Construct a corpus from a character vector

```
x <- corpus(data_char_ukimmig2010, text_field = "text")
```

### Explore a corpus

```
summary(data_corpus_inaugural, n = 2)
```

## Corpus consisting of 58 documents, showing 2 documents:

```
##          Text Types Tokens Sentences Year President FirstName Party
## 1 1789-Washington    625   1537      23 1789 Washington  George  none
## 2 1793-Washington     96    147       4 1793 Washington  George  none
```

### Extract or add document-level variables

```
party <- data_corpus_inaugural$Party
x$serial_number <- seq_len(ndoc(x))
docvars(x, "serial_number") <- seq_len(ndoc(x)) # alternative
```

### Bind or subset corpora

```
corpus(x[1:5]) + corpus(x[7:9])
corpus_subset(x, Year > 1990)
```

### Change units of a corpus

```
corpus_reshape(x, to = "sentences")
```

### Segment texts on a pattern match

```
corpus_segment(x, pattern, valuetype, extract_pattern = TRUE)
```

### Take a random sample of corpus texts

```
corpus_sample(x, size = 10, replace = FALSE)
```

## Tokenize a set of texts (tokens\_\*)

### Tokenize texts from a character vector or corpus

```
toks <- tokens("Powerful tool for text analysis.")
```

### Convert sequences into compound tokens

```
myseqs <- phrase(c("text analysis"))
tokens_compound(toks, myseqs)
```

### Select tokens

```
tokens_select(toks, c("powerful", "text"), selection = "keep")
```

### Create a dictionary

```
dict <- dictionary(list(negative = c("bad", "awful", "sad"),
positive = c("good", "wonderful", "happy")))
```

### Apply a dictionary

```
tokens_lookup(toks, dictionary = data_dictionary_LSD2015)
```

### Create ngrams and skipgrams from tokens

```
tokens_ngrams(toks, n = 1:3)
```

```
tokens_skipgrams(toks, n = 2, skip = 0:1)
```

### Convert case of tokens

```
tokens_tolower(toks) tokens_toupper(toks)
```

### Stem tokens

```
tokens_wordstem(toks)
```

`tokens_remove/select/toupper/tolower()` are also available

## Extract features (dfm\_\*)

### Create a document-feature matrix (dfm) from a tokens object

```
dfmat <- dfm(toks)
```

### Select features

```
dfm_select(dfmat, pattern = "recommend*"), selection = "keep")
```

### Randomly sample documents or features

```
dfm_sample(dfmat, what = c("documents", "features"))
```

### Weight or smooth the feature frequencies

```
dfm_weight(dfmat, scheme = "prop")
```

```
dfm_smooth(dfmat, smoothing = 0.5)
```

### Sort or group a dfm

```
dfm_sort(dfmat, margin = c("features", "documents", "both"))
```

```
dfm_group(dfmat, groups = President)
```

### Combine identical dimension elements of a dfm

```
dfm_compress(dfmat, margin = c("both", "documents", "features"))
```

### Create a feature co-occurrence matrix (fcm)

```
x <- fcm(data_corpus_inaugural, context = "window", size = 5)
```

`fcm_compress/remove/select/toupper/tolower()` are also available

## Useful additional functions

### Locate keywords-in-context

```
kwic(tokens(data_corpus_ inaugural), pattern = "america*")  
## Keyword-in-context with 499 matches.  
## [1789-Washington, 1069] hands of the | American | people. Besides  
## [1789-Washington, 1472] to favor the | American | people with opportunities  
## [1793-Washington, 63] people of united | America | . Previous to  
## [1797-Adams, 16] middle course for | America | remained between unlimited
```

### Utility functions

as.character(corpus)	Show texts of a corpus
ndoc(corpus / dfm / tokens)	Count documents/features
nfeat(corpus / dfm / tokens)	Count features
ntoken(corpus / dfm / tokens)	Count tokens
summary(corpus / dfm)	Print summary
head(corpus / dfm)	Return first part
tail(corpus / dfm)	Return last part

## Calculate text statistics (textstat\_\*)

These functions require the **quanteda.textstats** package

### Tabulate feature frequencies from a dfm

```
textstat_frequency(x) topfeatures(x)
```

### Identify and score collocations from a tokenized text

```
toks <- tokens(c("quanteda is a pkg for quant text analysis",  
"quant text analysis is a growing field"))  
textstat_collocations(toks, size = 3, min_count = 2)
```

### Calculate readability of a corpus

```
textstat_readability(x, measure = c("Flesch", "FOG"))
```

### Calculate lexical diversity of a dfm

```
textstat_lexdiv(x, measure = "TTR")
```

### Measure distance or similarity from a dfm

```
textstat_simil(x, "2017-Trump", method = "cosine",  
margin = c("documents", "features"))
```

```
textstat_dist(x, "2017-Trump",  
margin = c("documents", "features"))
```

### Calculate keyness statistics

```
textstat_keyness(x, target = "2017-Trump")
```

## Fit text models based on a dfm (textmodel\_\*)

These functions require the **quanteda.textmodels** package

### Correspondence Analysis (CA)

```
textmodel_ca(x, threads = 2, sparse = TRUE, residual_floor = 0.1)
```

### Naïve Bayes classifier for texts

```
textmodel_nb(x, y = training_labels, distribution = "multinomial")
```

### SVM classifier for texts

```
textmodel_svm(x, y = training_labels)
```

### Wordscores text model

```
refscores <- c(seq(-1.5, 1.5, .75), NA)
```

```
textmodel_wordscores(data_dfm_lbgexample, refscores)
```

### Wordfish Poisson scaling model

```
textmodel_wordfish(dfm(data_corpus_irishbudget2010), dir = c(6,5))
```

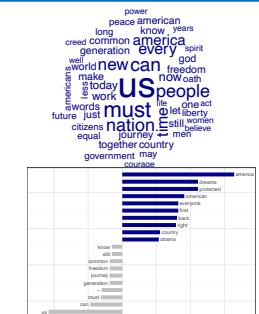
**Textmodel methods:** `predict()`, `coef()`, `summary()`, `print()`

## Plot features or models (textplot\_\*)

These functions require the **quanteda.textplots** package

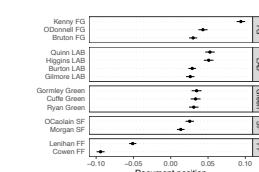
### Plot features as a wordcloud

```
data_corpus_ inaugural |>  
corpus_subset(President == "Obama") |>  
tokens() |>  
tokens_remove(pattern = stopwords("en")) |>  
dfm() |>  
textplot_wordcloud()
```



### Plot word keyness

```
data_corpus_ inaugural |>  
corpus_subset(President %in%  
c("Obama", "Trump")) |>  
tokens() |>  
dfm() |>  
dfm_group(groups = President) |>  
textstat_keyness(target = "Trump") |>  
textplot_keyness()
```



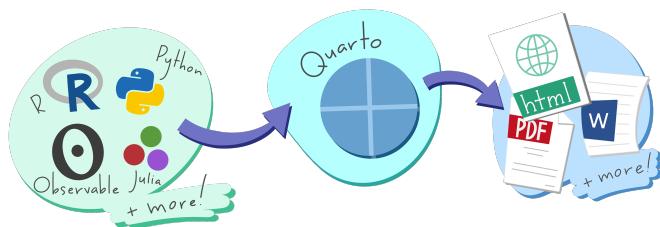
### Plot Wordfish, Wordscores or CA models

```
textplot_scale1d(scaling_model, margin = "documents")
```

## Convert dfm to a non-quanteda format

```
convert(x, to = c("lda", "tm", "stm", "austin", "topicmodels",  
"lsa", "matrix", "data.frame"))
```

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## Author

### SOURCE FILE: hello.qmd

```
---  
title: "Hello, Penguins"  
format: html  
execute:  
  echo: false  
---  
  
## Meet the penguins  
  
The `penguins` data contains size measurements for three species of penguins in the Palmer Archipelago, Antarctica.  
  
The three species of penguins have quite distinct distributions of physical dimensions (@fig-penguins).  
  
```{r}  
#| label: fig-penguins  
#| fig-cap: "Dimensions of penguins across three species."  
#| warning: false  
library(tidyverse, quietly = TRUE)  
library(palmerpenguins)  
penguins %>  
  ggplot(aes(x = flipper_length_mm, y = bill_length_mm)) +  
  geom_point(aes(color = species)) +  
  scale_color_manual(  
    values = c("darkorange", "purple", "cyan4")) +
```

**Set format(s) and options**  
Use YAML Syntax

**## Write with \*\*Markdown\*\***  
RStudio: Help > Markdown Quick Reference  
 Use Visual Editor

**Include code**  
R, Python, Julia, Observable, or any language with a Jupyter kernel

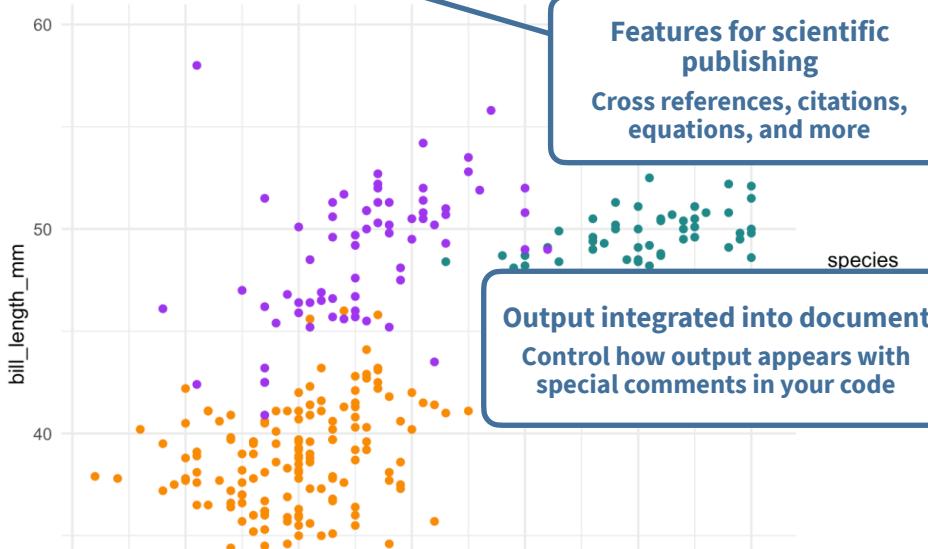
## Render

### RENDERED OUTPUT: hello.html

#### Hello, Penguins

#### Meet the penguins

The three species of penguins have quite distinct distributions of physical dimensions (Figure 1).



### USE A TOOL WITH A RICH EDITING EXPERIENCE

RStudio Visual Studio Code + Quarto extension

Run code cells as you write

Render with a button or keyboard shortcut

Edit Quarto documents with a Visual Editor

### OR ANY TEXT EDITOR

Quarto documents (.qmd) can be edited in any tool that edits text.

Apply formatting in Visual Editor. Saved as Markdown in source.  
Insert elements like code cells, cross references, and more.

Save, then render to preview the document output.

Terminal  
quarto preview hello.qmd

Use Render button   
 Use Preview button

The resulting HTML/PDF/MS Word/etc. document will be created and saved in the same directory as the source .qmd file.

### BEHIND THE SCENES

When you render a document, Quarto:

- Runs the code and embeds results and text into an .md file with: **Knitr**, if any {r} cells or, **Jupyter**, if any other cells.
- Converts the .md file into the output format with Pandoc.

## GET QUARTO

<https://quarto.org/docs/download/>

Or use version **bundled with RStudio**

## GET STARTED

<https://quarto.org/docs/get-started/>

## Publish

### Terminal

quarto publish {venue} hello.qmd

{venue}: quarto-pub, connect, gh-pages, netlify, confluence, posit-cloud

Use Publish button

Quarto Pub

Free publishing service for Quarto content.

posit Cloud

Cloud-hosted, control access to project and output.

posit Connect

Org-hosted, control access, schedule updates.

## Quarto Projects

### CREATE WEBSITES, BOOKS, AND MORE

A directory of Quarto documents + a configuration file (\_quarto.yml)

See examples at <https://quarto.org/docs/gallery/>

Get started from the command line:

### Terminal

quarto create project {type}

{type}: default, website, blog, book, confluence, manuscript

Use File > New Project

Artwork from "Hello, Quarto" keynote by Julia Lowndes and Mine Çetinkaya-Rundel, presented at RStudio Conference 2022. Illustrated by Allison Horst.

# Include Code

## CODE CELLS

Code cells start with `{{language}}` and end with `{{}}`.

Use **Insert Code Chunk/Cell**

```
```{r}
# I label: chunk-id
library(tidyverse)
```
```{python}
# I label: chunk-id
import pandas as pd
```
```

Other languages: {julia}, {ojs}

Add code cell options with **#I** comments.

Cell options control **execution**, figures, tables, layout and more. See them all at: <https://quarto.org/docs/reference/cells>

## EXECUTION OPTIONS

### OPTION DEFAULT EFFECTS

|                |       |                                                                     |
|----------------|-------|---------------------------------------------------------------------|
| <b>echo</b>    | true  | false: hide code<br>fenced: include code cell syntax                |
| <b>eval</b>    | true  | false: don't run code                                               |
| <b>include</b> | true  | false: don't include code or results                                |
| <b>output</b>  | true  | false: don't include results<br>asis: treat results as raw markdown |
| <b>warning</b> | true  | false: don't include warnings in output                             |
| <b>error</b>   | false | true: include error in output and continue with render              |

Set execution options at the **cell level**:

```
```{r}
# I echo: false
```
```{python}
# I echo: false
```
```
```

Or, **globally** in the YAML header with the **execute** option:

```
---
execute:
  echo: false
---
```

**Set options in code cells with **#I** comments and YAML syntax:**  
**key: value**

## INLINE CODE

Use computed values directly in text sections.  
Code is evaluated at render and results appear as text.

**KNITR**      **JUPYTER**      **OUTPUT**  
Value is `r 2 + 2`. Value is `{{python}} 2 + 2`. Value is 4.

# Set Format and Options

## SET FORMAT OPTIONS

```
---  
title: "My Document"  
format:  
  html:  
    code-fold: true  
    toc: true  
---  
Indent options 4 spaces  
Indent format 2 spaces
```

## MULTIPLE FORMATS

```
---  
title: "My Document"  
toc: true  
format:  
  html:  
    code-fold: true  
  pdf: default  
---
```

Top-level options apply to all formats

Common formats: **html, pdf, docx, odt, rtf, gfm, pptx, revealjs, beamer**

Render **all** formats:

Terminal  
quarto render hello.qmd

Render a **specific** format:

Terminal  
quarto render hello.qmd --to pdf

## OPTION

html/revealjs  
pdf/beamer  
docx/pptx

## DESCRIPTION

<b>toc</b>	X X X	Add a table of contents (true or false)
<b>toc-depth</b>	X X X	Lowest level of headings to add to table of contents (e.g. 2, 3)
<b>anchor-sections</b>	X	Show section anchors on mouse hover (true or false)
<b>highlight-style</b>	X X X	Syntax highlighting theme (e.g. arrow, pygments, kate, zenburn)
<b>mainfont, monofont</b>	X X	Font name. HTML: sets CSS font-family; LaTeX: via fonts package
<b>theme</b>	X	Bootswatch theme name (e.g. cosmo, darkly, solar etc.)
<b>css</b>	X	CSS or SCSS file to use to style the document (e.g. "style.css")
<b>reference-doc</b>	X	docx/pptx file containing template styles (e.g. file.docx, file.pptx)
<b>include-in-header</b>	X X	Files of content to include in header of output document, also <b>include-before-body, include-after-body</b>
<b>keep-md</b>	X X X	Keep intermediate markdown (true or false), also <b>keep-ipynb, keep-tex</b>
<b>documentclass</b>	X	LaTeX document class, set document class options with <b>classoption</b>
<b>pdf-engine</b>	X	LaTeX engine to produce PDF output (xelatex, pdflatex, lualatex)
<b>cite-method</b>	X	Method used to format citations (citeproc, natbib, biblatex)
<b>code-fold</b>	X	Let readers toggle the display of R code (false, true, or show)
<b>code-tools</b>	X	Add menu for hiding, showing, and downloading code (true or false)
<b>code-overflow</b>	X	Display of wide code (scroll, or wrap)
<b>fig-align</b>	X X /	Alignment of figures (default, left, right, or center)
<b>fig-width, fig-height</b>	X X X	Default width and height for figures in inches
<b>fig-format</b>	X X X	Format for Matplotlib or R figures (retina, png, jpeg, svg, or pdf)

Visit <https://quarto.org/docs/reference/> to see **all options** by format

Also use in code cells

# Add Content

## FIGURES ?

## MARKDOWN

```
![[CAP]](image.png){#fig-LABEL fig-alt="ALT"}
```

## COMPUTATION

```
```{python}
# I label: fig-LABEL
# I fig-cap: CAP
# I fig-alt: ALT
{{ plot code here }}
```
```

Or {r}

## CROSS REFERENCES

### 1. Add labels

Code cell: add option label: prefix-LABEL  
Markdown: add attribute #prefix-LABEL

### 2. Add references @prefix-LABEL, e.g.

You can see in @fig-scatterplot, that...

| Prefix | Renders  | Prefix | Renders    |
|--------|----------|--------|------------|
| fig-   | Figure 1 | eq-    | Equation 1 |
| tbl-   | Table 1  | sec-   | Section 1  |

## TABLES ?

## MARKDOWN

```
lobject	radius!
ISun | 1 696000  
IEarth | 6371  
:  
: CAPTION {#tbl-LABEL}
```

Use **Insert Table** in the **Visual Editor**

## CITATIONS

1. Add a bibliography **file** to the YAML header:

```
---  
bibliography: references.bib  
---
```

2. Add citations: **[@citation]**, or **@citation**

Use **Insert Citations** dialog in the **Visual Editor**

Build your bibliography file from your Zotero library, DOI, Crossref, DataCite, or PubMed

**COMPUTATION** Output a Markdown table or an HTML table from your code

## KNITR

Use knitr::kable() to produce Markdown:

```
```{r}
# I label: tbl-LABEL
# I tbl-cap: CAPTION
import pandas as pd, tabulate
from IPython.display import Markdown
df = pd.DataFrame({ "A": [1, 2],
                     "B": [1, 2] })
Markdown(df.to_markdown(index=False))
```
```

Also see the R packages: gt, flextable, kableExtra.

**JUPYTER** Add Markdown() to Markdown output:

```
```{python}
# I label: tbl-LABEL
# I tbl-cap: CAPTION
import pandas as pd, tabulate
from IPython.display import Markdown
df = pd.DataFrame({ "A": [1, 2],
                     "B": [1, 2] })
Markdown(df.to_markdown(index=False))
```
```

Instead of **tip** use one of:  
note, caution, warning,  
or important.

note    warning  
 caution    important

## CALLOUTS

... {.callout-tip}  
## Title

Text

...

## SHORTCODES

```
{{< include _file.qmd >}}
{{< embed file.ipynb#id >}}
{{< video video.mp4 >}}
```

# PGS Catalog access with quincunx



## Introduction

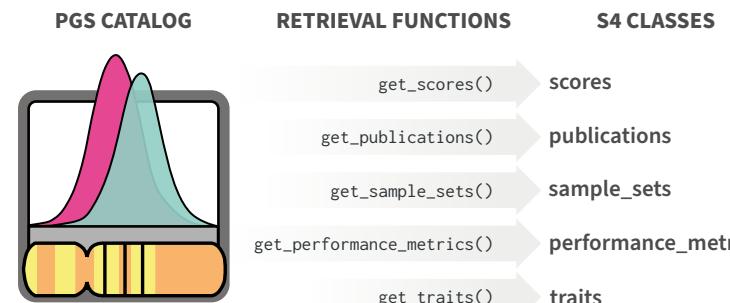
The **PGS Catalog** is a service provided by the EMBL-EBI and University of Cambridge that offers a manually curated and freely available database of published polygenic scores (PGS): <https://www.pgscatalog.org/>.

The PGS Catalog data provided by the **REST API** is organised around five core entities:

**PGS** Polygenic Scores  
**PGP** PGS Publications  
**PSS** PGS Sample Sets  
**PPM** PGS Performance Metrics  
**EFO** EFO traits

# Get PGS Catalog Entities

**quincunx** facilitates the access to the Catalog via the REST API, allowing you to programmatically retrieve data directly into R. Each of the five entities is mapped to an S4 object of a class of the same name.



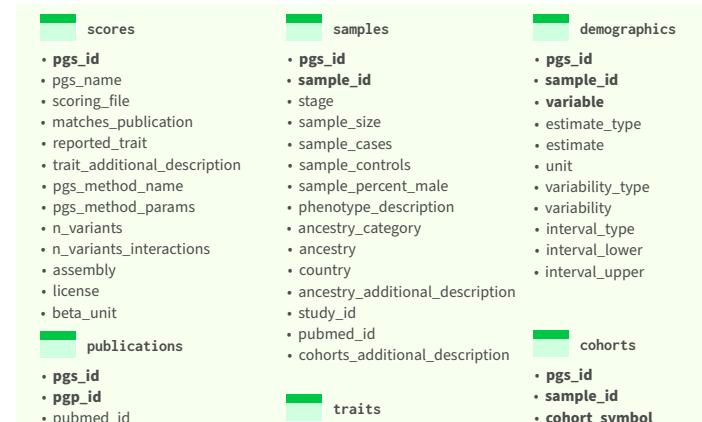
Query criteria for retrieval functions, e.g., PGS can be queried by either pgs\_id, efo\_id or pubmed\_id. These correspond to the criteria exposed by the PGS Catalog REST API: <https://www.pgscatalog.org/rest/>.

| Search by  | Example       | PGS    | PGP    | PSS  | PPM | EFC    |
|------------|---------------|--------|--------|------|-----|--------|
| pgs_id     | "PGS000001"   | Green  | Yellow | Blue | Red |        |
| pgp_id     | "PGP000001"   | Yellow | Yellow |      |     |        |
| pss_id     | "PSS000001"   | Yellow |        | Blue |     |        |
| ppm_id     | "PPM_000001"  | Yellow |        |      | Red |        |
| efo_id     | "EFO_0000249" | Green  |        |      |     | Yellow |
| pubmed_id  | "25855707"    | Green  | Yellow |      |     |        |
| author     | "Mavaddat"    | Yellow | Yellow |      |     |        |
| trait term | "Alzheimer"   | Yellow |        |      |     | Yellow |

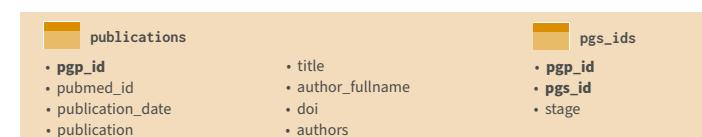
# PGS Catalog Entities in R

PGS Catalog entities are represented as S4 classes in R. Each class represents a relational database of tidy data tables. All objects start with a table with the same name as the class. Combination of variables indicated in bold renders each row unique in each table.

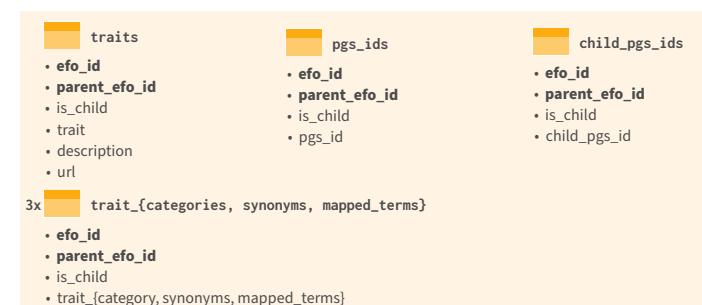
## S4 class scores



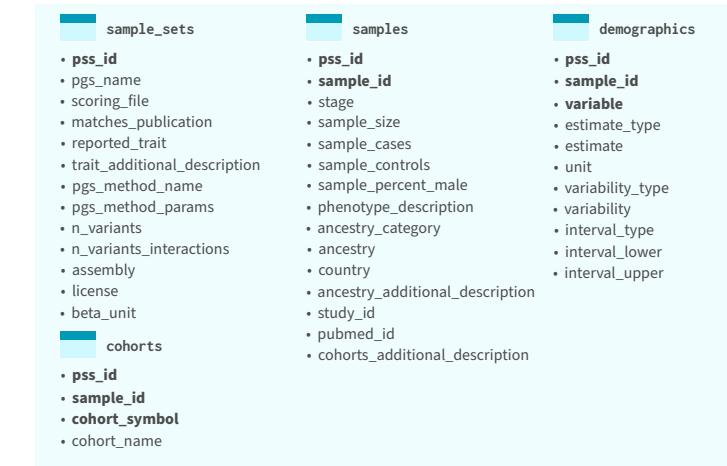
## S4 class publications



## S4 class traits



S4 class sample\_sets



## S4 class performance metrics





## Other S4 Entities

Besides the five PGS Catalog entities, there are three other objects that can be retrieved from the REST API: trait\_categories, cohorts and releases.

### S4 class trait\_categories

|   |                  |
|---|------------------|
|   | trait_categories |
| • | trait_category   |
| • | efo_id           |
| • | trait            |
| • | description      |
| • | url              |

### S4 class cohorts

|   |               |
|---|---------------|
|   | cohorts       |
| • | cohort_symbol |
| • | cohort_name   |

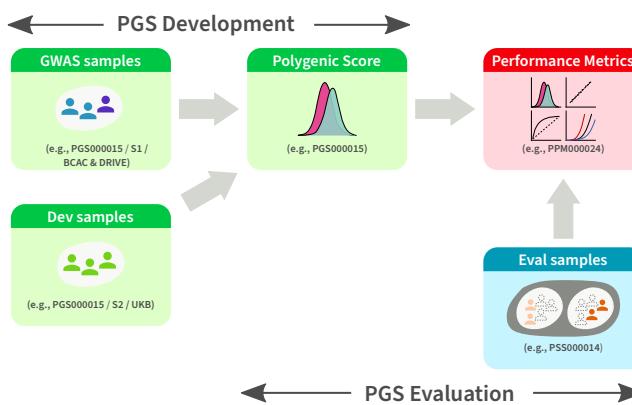
  

|   |               |
|---|---------------|
|   | pgs_ids       |
| • | cohort_symbol |
| • | pgs_id        |
| • | stage         |

### S4 class releases

|    |                           |
|----|---------------------------|
|    | releases                  |
| 3x |                           |
| •  | date                      |
| •  | pgs_ids, ppm_ids, ppg_ids |
| •  | date                      |
| •  | (pgs_id, ppm_id, ppg_id)  |
| •  | n_ppm                     |
| •  | n_pgp                     |
| •  | notes                     |

## PGS Construction Process



Samples and Polygenic Scores (PGS) are annotated according to their utilisation context in the PGS construction process, i.e. the stage variable in quincunx:

- Source of Variant Associations (GWAS): stage="gwas"
- Score Development/Training: stage="dev"
- Development: stage="gwas/dev" ("gwas" and "dev")
- PGS Evaluation: stage="eval"

## Cohorts, Samples and Sample Sets

### Cohorts

A cohort is a group of individuals with a shared characteristic. Cohorts are identified in quincunx by the cohort\_symbol variable.



### Samples

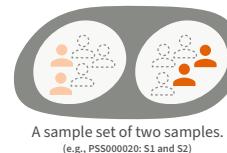
A sample is a group of participants associated with none, one or more catalogued cohorts. The selection from a cohort can be either a subset or its totality. Samples are not identified in PGS Catalog with a global unique identifier, but quincunx assigns a surrogate identifier (sample\_id) to allow relations between tables.

Possible compositions of samples:



### Sample Sets

A sample set is a group of samples used in a polygenic score evaluation. Each sample set is identified in the PGS Catalog by a unique sample set identifier (PSS ID).



A sample set of two samples.  
(e.g., PSS000020: S1 and S2)

## Manipulate Cases of S4 Entities

Get a scores object **s** consisting of two polygenic scores (PGS):

```
s <- get_scores(pgs_id = c('a', 'b'))
```

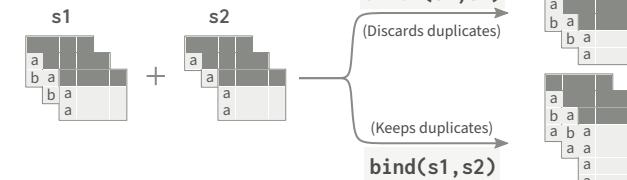
Subset object **s** by either identifier or position using '**[**':



s['a'] # Subset by identifier

s[1] # Subset by position

Combine two scores' objects:



CC BY SA Ramiro Magno • Learn more at <https://maialab.org/quincunx> • quincunx version 0.0.1 • Updated: 2021-05-05

# randomizr:: CHEAT SHEET

## Two Arm Trials

**Simple** random assignment is like flipping coins for each unit separately.

```
simple_ra(N = 100, prob = 0.5)
```

**Complete** random assignment allocates a fixed number of units to each condition.

```
complete_ra(N = 100, m = 50)
complete_ra(N = 100, prob = 0.5)
```

**Block** random assignment conducts complete random assignment separately for groups of units.

```
blocks <- rep(c("A", "B", "C"),
              c(50, 100, 200))

# defaults to half of each block
block_ra(blocks = blocks)

# can change with block_m
block_ra(blocks = blocks,
         block_m = c(20, 30, 40))
```

**Cluster** random assignment allocates whole groups of units to conditions together.

```
clusters <- rep(letters, times = 1:26)
cluster_ra(clusters = clusters)
```

**Block and cluster** random assignment conducts cluster random assignment separately for groups of clusters.

```
clusters <- rep(letters, times = 1:26)
blocks <- rep(paste0("block_", 1:5),
              c(15, 40, 65, 90, 141))
block_and_cluster_ra(blocks = blocks,
                     clusters = clusters)
```

randomizr is part of the DeclareDesign suite of packages for designing, implementing, and analyzing social science research designs.

## Multi Arm Trials

Set the number of arms with `num_arms` or with `conditions`.

```
complete_ra(N = 100, num_arms = 3)
complete_ra(N = 100, conditions = c("control",
                                    "placebo", "treatment"))
```

The `*_each` arguments in randomizr functions specify design parameters for each arm separately.

```
complete_ra(N = 100, m_each = c(20, 30, 50))
complete_ra(N = 100,
           prob_each = c(0.2, 0.3, 0.5))
```

If the design is the **same** for all blocks, use `prob_each`:

```
blocks <- rep(c("A", "B", "C"),
              c(50, 100, 200))
block_ra(blocks = blocks,
         prob_each = c(.1, .1, .8))
```

If the design is **different** in different blocks, use `block_m_each` or `block_prob_each`:

```
block_m_each <- rbind(c(10, 20, 20),
                      c(30, 50, 20),
                      c(50, 75, 75))
block_ra(blocks = blocks,
         block_m_each = block_m_each)

block_prob_each <- rbind(c(.1, .1, .8),
                         c(.2, .2, .6),
                         c(.3, .3, .4))
block_ra(blocks = blocks,
         block_prob_each = block_prob_each)
```

If `conditions` is numeric, the output will be **numeric**.

If `conditions` is not numeric, the output will be a **factor** with levels in the order provided to `conditions`.

```
complete_ra(N = 100, conditions = -2:2)
complete_ra(N = 100, conditions = c("A", "B"))
```

## Declaration

Learn about assignment procedures by “declaring” them with `declare_ra()`

```
declaration <-
  declare_ra(N = 100, m_each = c(30, 30, 40))
```

```
declaration # print design information
```

Conduct a random assignment:

```
conduct_ra(declaration)
```

Obtain observed condition probabilities (useful for inverse probability weighting if probabilities of assignment are not constant)

```
Z <- conduct_ra(declaration)
obtain_condition_probabilities(declaration, Z)
```

## Sampling

All assignment functions have sampling analogues: Sampling is identical to a two arm trial where the treatment group is sampled.

### Assignment

```
simple_ra()
```

```
complete_ra()
```

```
block_ra()
```

```
cluster_ra()
```

```
block_and_cluster_ra()
```

```
declare_ra()
```

```
conduct_ra()
```

### Sampling

```
simple_rs()
```

```
complete_rs()
```

```
strata_rs()
```

```
cluster_rs()
```

```
strata_and_cluster_rs()
```

```
declare_rs()
```

```
draw_rs()
```

## Stata

A Stata version of randomizr is available, with the same arguments but different syntax:

```
ssc install randomizr
set obs 100
complete_ra, m(50)
```

# Best Practice for R :: CHEAT SHEET

## Software

- Write code in the **RStudio** IDE
- Use **quarto** for literate programming
- Use **git** to version-control your code and analysis
- Use **GitHub** to collaborate with other people

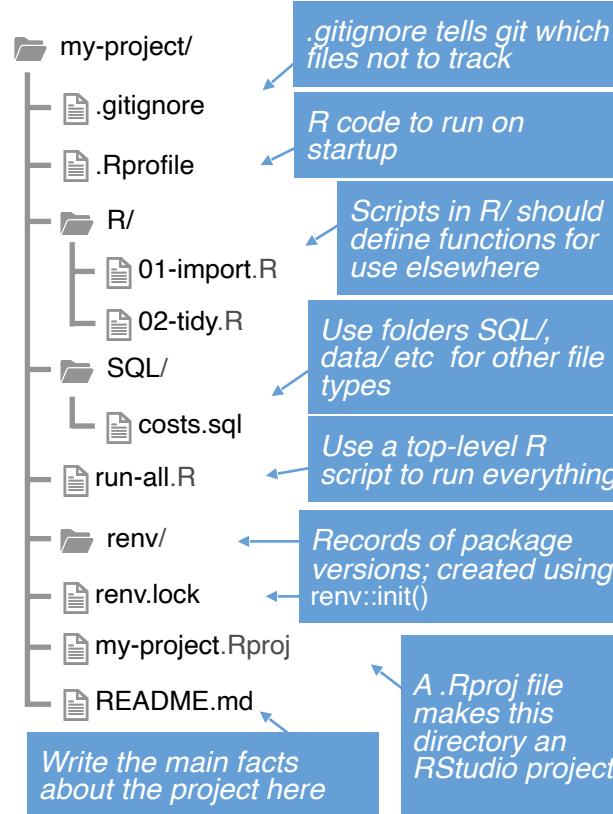
## Projects

### PROJECT CREATION

- Create** a new project in RStudio using File > New Project > New Directory
- Do** put projects in a single, local folder like C:\Users\your-name\Documents
- Don't** put projects in locations controlled by OneDrive / iCloud (these don't play well with Git)

### PROJECT STRUCTURE

Most projects should be structured like this:



NB, usethis::use\_description() + usethis::use\_namespace() will turn this structure into a package!



## Packages

Packages should be loaded in one place with successive calls to library()



- Use the **tidyverse** for normal wrangling, plotting etc
- Use **tidymodels** for modelling and machine learning
- Use **{shiny}**, **{bslib}** and **{bs4Dash}** for app development
- Use **r-lib** packages like **{rlang}**, **{cli}** & **{glue}** for low-level programming
- Use **{renv}** in long-term projects to track dependency packages

GitHub stars are a good proxy for a package's quality. Not sure whether to use a package? If it has >200 stars on GitHub it's probably good!

## Getting Help



### CREATE A REPREX

- A **minimal, reproducible example** should demonstrate the issue as simply as possible
- Copy your example code and run **reprex::reprex()** to embed errors/messages/outputs as comments
- Use your reprex in a question on Teams or Stackoverflow

```
print("Hello " + "world!")
#> Error in "Hello " + "world!": non-numeric argument to
#> binary operator
```

This reprex minimally demonstrates an error when attempting to use + for Python-style string concatenation

### ETIQUETTE WHEN ASKING QUESTIONS

#### Don't

Post screenshots of your code

#### Do

Use **reprex::reprex()** and paste your code as text

Include big files

Use **dput()** or **tibble::tribble()** to include a data sample

Ignore messages or warnings

**Ensure your code only fails where you're expecting it to**

## Databases

- Use **{DBI}** and **{odbc}** to connect to SQL
  - Use **helper functions** to create connections
- ```
connect_to_db <- function(db) {
  DBI::dbConnect(
    odbc::odbc(), Database = db,
    # Hard-code common options here
  )
  # Connect using the helper
  con <- connect_to_db("DWH")
```

## Learning More

- For common data science tasks, see [R for Data Science \(2e\)](#)
- For package development, see [R Packages \(2e\)](#)
- For advanced programming, see [Advanced R \(2e\)](#)
- For app development, see [Mastering Shiny](#)



## WRITING FUNCTIONS: WORKFLOW

a <-	complex operation on a
b <-	complex operation on b
c <-	complex operation on c
d <-	complex operation on d

1. Repetitive, complex code; purpose clarified by comments

```
operate_on  on(x) {
  complex operation on x
}
```

2. Complex logic abstracted into functions

a <-	operate_on
b <-	operate_on
c <-	operate_on
d <-	operate_on

3. Repetition reduced; clearer code; less need for comments

For other styling guidance, refer to the [Tidyverse style guide](#)

## NAMING THINGS

- Use **lower\_snake\_case** for most objects (functions, variables etc)
- Title\_Snake\_Case** may be used for *column names*
- Use only **syntactic** names where possible (include only *numbers, letters, underscores* and *periods*, and don't start with a number)

## WHITESPACE

- Add spaces** after commas and around operators like **>**, **%>%**, **+**, **-**, **\***, **/**, **=** and **<**
- Indentation increases** should always be by exactly 2 spaces
- Add linebreaks** when lines get longer than **80** characters.
- When there are many arguments in a call, **give each argument its own line** (including the first one!)

```
# Good (lower_snake_case everywhere):
add1      <- function(x) x + 1
first_letters <- letters[1:3]
iris_sample <- slice_sample(iris, n = 5)
```

```
# Bad (non-syntactic, not lower_snake_case):
`add 1`    <- function(x) x + 1
FirstLetters <- letters[1:3]
iris.sample <- slice_sample(iris, n = 5)
```

```
# Good (lots of spaces, indents always by +2):
df <- iris |>
  mutate(
    Sepal.Area = Sepal.Width * Sepal.Length,
    Petal.Area = Petal.Width * Petal.Length
  )
```

```
# Bad (inconsistent spacing and indentation):
df<-iris |>
  mutate(Sepal.Area=Sepal.Width*Sepal.Length,
    Petal.Area=Petal.Width*Petal.Length)
```

# Basic Regular Expressions in R

## Cheat Sheet

### Character Classes

<code>[:digit:]</code> or <code>\d</code>	Digits; [0-9]
<code>\D</code>	Non-digits; [^0-9]
<code>[:lower:]</code>	Lower-case letters; [a-z]
<code>[:upper:]</code>	Upper-case letters; [A-Z]
<code>[:alpha:]</code>	Alphabetic characters; [A-z]
<code>[:alnum:]</code>	Alphanumeric characters [A-z0-9]
<code>\w</code>	Word characters; [A-z0-9_]
<code>\W</code>	Non-word characters
<code>[:xdigit:]</code> or <code>\x</code>	Hexadec. digits; [0-9A-Fa-f]
<code>[:blank:]</code>	Space and tab
<code>[:space:]</code> or <code>\s</code>	Space, tab, vertical tab, newline, form feed, carriage return
<code>\S</code>	Not space; [^[:space:]]
<code>[:punct:]</code>	Punctuation characters; !#\$%&'()*+, -./; <=>?@[]^_`{ }~
<code>[:graph:]</code>	Graphical characters; [[:alnum:][:punct:]]
<code>[:print:]</code>	Printable characters; [[:alnum:][:punct:]\s]
<code>[:cntrl:]</code> or <code>\c</code>	Control characters; \n, \r etc.

### Special Metacharacters

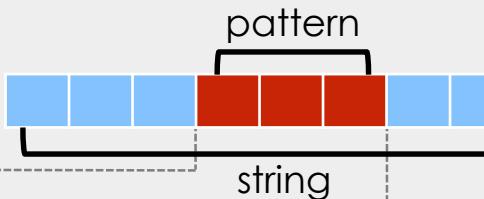
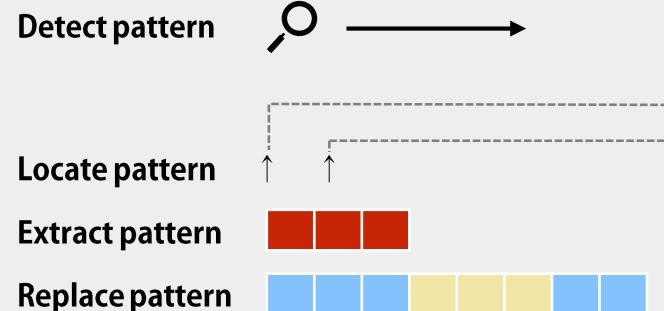
<code>\n</code>	New line
<code>\r</code>	Carriage return
<code>\t</code>	Tab
<code>\v</code>	Vertical tab
<code>\f</code>	Form feed

### Lookarounds and Conditionals\*

<code>(?=)</code>	Lookahead (requires PERL = TRUE), e.g. (?=yx): position followed by 'xy'
<code>(?!)</code>	Negative lookahead (PERL = TRUE); position NOT followed by pattern
<code>(?&lt;=)</code>	Lookbehind (PERL = TRUE), e.g. (?<=yx): position following 'xy'
<code>(?&lt;!)</code>	Negative lookbehind (PERL = TRUE); position NOT following pattern
<code>?(if)then</code>	If-then-condition (PERL = TRUE); use lookaheads, optional char. etc in if-clause
<code>?(if)then else</code>	If-then-else-condition (PERL = TRUE)

\*see, e.g. <http://www.regular-expressions.info/lookaround.html>  
<http://www.regular-expressions.info/conditional.html>

## Functions for Pattern Matching



```
> string <- c("Hipopopotamus", "Rhymenoceros", "time for bottomless lyrics")
> pattern <- "t.m"
```

### Detect Patterns

```
grep(pattern, string)
[1] 1 3
grep(pattern, string, value = TRUE)
[1] "Hipopopotamus"
[2] "time for bottomless lyrics"
grepl(pattern, string)
[1] TRUE FALSE TRUE
stringr::str_detect(string, pattern)
[1] TRUE FALSE TRUE
```

### Split a String using a Pattern

```
strsplit(string, pattern) or stringr::str_split(string, pattern)
```

### Locate Patterns

```
regexpr(pattern, string)
find starting position and length of first match
gregexpr(pattern, string)
find starting position and length of all matches
stringr::str_locate(string, pattern)
find starting and end position of first match
stringr::str_locate_all(string, pattern)
find starting and end position of all matches
```

### Extract Patterns

```
regmatches(string, regexpr(pattern, string))
extract first match [1] "tam" "tim"
regmatches(string, gregexpr(pattern, string))
extract all matches, outputs a list [[1]] "tam" [[2]] character(0) [[3]] "tim" "tom"
stringr::str_extract(string, pattern)
extract first match [1] "tam" NA "tim"
stringr::str_extract_all(string, pattern)
extract all matches, outputs a list
stringr::str_extract_all(string, pattern, simplify = TRUE)
extract all matches, outputs a matrix
stringr::str_match(string, pattern)
extract first match + individual character groups
stringr::str_match_all(string, pattern)
extract all matches + individual character groups
```

### Replace Patterns

```
sub(pattern, replacement, string)
replace first match
gsub(pattern, replacement, string)
replace all matches
stringr::str_replace(string, pattern, replacement)
replace first match
stringr::str_replace_all(string, pattern, replacement)
replace all matches
```

### Character Classes and Groups

- . Any character except \n
- | Or, e.g. (a|b)
- [...] List permitted characters, e.g. [abc]
- [a-z] Specify character ranges
- [^...] List excluded characters
- (...) Grouping, enables back referencing using \\N where N is an integer

### Anchors

- ^ Start of the string
- \$ End of the string
- \b Empty string at either edge of a word
- \B NOT the edge of a word
- \B Beginning of a word
- \B End of a word

### Quantifiers

- \* Matches at least 0 times
- + Matches at least 1 time
- ? Matches at most 1 time; optional string
- {n} Matches exactly n times
- {n,} Matches at least n times
- {n,m} Matches between n and m times

### General Modes

By default R uses *extended regular expressions*. You can switch to *PCRE regular expressions* using `PERL = TRUE` for base or by wrapping patterns with `perl()` for stringr.

All functions can be used with literal searches using `fixed = TRUE` for base or by wrapping patterns with `fixed()` for stringr.

All base functions can be made case insensitive by specifying `ignore.case = TRUE`.

### Escaping Characters

Metacharacters (. \* + etc.) can be used as literal characters by escaping them. Characters can be escaped using `\\\` or by enclosing them in `\Q...\E`.

### Case Conversions

Regular expressions can be made case insensitive using `(?i)`. In backreferences, the strings can be converted to lower or upper case using `\L` or `\U` (e.g. `\L\1`). This requires `PERL = TRUE`.

### Greedy Matching

By default the asterisk \* is greedy, i.e. it always matches the longest possible string. It can be used in lazy mode by adding ?, i.e. \*?.

Greedy mode can be turned off using `(?U)`. This switches the syntax, so that `(?U)a*` is lazy and `(?U)a*?` is greedy.

### Note

Regular expressions can conveniently be created using e.g. the packages `rex` or `rebus`.

# Use Python with R with reticulate :: CHEATSHEET



The **reticulate** package lets you use Python and R together seamlessly in R code, in R Markdown documents, and in the RStudio IDE.

## Python in R Markdown

(Optional) Build Python env to use.

knitr versions >= 1.18 will automatically use the reticulate engine for Python chunks. See `?reticulate::eng_python` for a listing of supported knitr chunk options.

Suggest the Python environment to use, in your setup chunk.

Begin Python chunks with ````{python}`. Chunk options like `echo`, `include`, etc. all work as expected.

Use the `py` object to access objects created in Python chunks from R chunks.

Python chunks all execute within a **single** Python session so you have access to all objects created in previous chunks.

Use the `r` object to access objects created in R chunks from Python chunks.

Output displays below chunk, including matplotlib plots.

```
python.Rmd x
1 ````{r setup, include = FALSE}
2 library(reticulate)
3 virtualenv_create("fmri-proj")
4 py_install("seaborn", envname = "fmri-proj")
5 use_virtualenv("fmri-proj")
6
7 ````{python, echo = FALSE}
8 import seaborn as sns
9 fmri = sns.load_dataset("fmri")
10
11 ````{r}
12 f1 <- subset(py$fmri, region == "parietal")
13
14 ````{python}
15 import matplotlib as mpl
16 sns.lmplot("timepoint", "signal", data=r.f1)
17 mpl.pyplot.show()
18
19 ````{r}
20
21
```

```
python.R x
1 library(reticulate)
2 py_install("seaborn")
3 use_virtualenv("r-reticulate")
4
5 sns <- import("seaborn")
6
7 fmri <- sns$load_dataset("fmri")
8 dim(fmri)
9
10 # creates tips
11 source_python("python.py")
12 dim(tips)
13
14 # creates tips in main
15 py_run_file("python.py")
16 dim(py$tips)
17
18 py_run_string("print(tips.shape)")
19
```

## Object Conversion

**Tip:** To index Python objects begin at 0, use integers, e.g. `0L`

Reticulate provides **automatic** built-in conversion between Python and R for many Python types.

R	↔	Python
Single-element vector		Scalar
Multi-element vector		List
List of multiple types		Tuple
Named list		Dict
Matrix/Array		NumPy ndarray
Data Frame		Pandas DataFrame
Function		Python function
NULL, TRUE, FALSE		None, True, False

Or, if you like, you can convert manually with

`py_to_r(x)` Convert a Python object to an R object. Also `r_to_py()`.

`tuple(..., convert = FALSE)` Create a Python tuple. `tuple("a", "b", "c")`

`dict(..., convert = FALSE)` Create a Python dictionary object. Also `py_dict()` to make a dictionary that uses Python objects as keys. `dict(foo = "bar", index = 42L)`

`np_array(data, dtype = NULL, order = "C")` Create NumPy arrays. `np_array(c(1:8), dtype = "float16")`

`array_reshape(x, dim, order = c("C", "F"))` Reshape a Python array. `x <- 1:4; array_reshape(x, c(2, 2))`

`py_func(f)` Wrap an R function in a Python function with the same signature. `py_func(xor)`

`py_main_thread_func(f)` Create a function that will always be called on the main thread.

`iterate(it, f = base::identity, simplify = TRUE)` Apply an R function to each value of a Python iterator or return the values as an R vector, draining the iterator as you go. Also `iter_next()` and `as_iterator()`.

`py_iterator(fn, completed = NULL)` Create a Python iterator from an R function. `seq_gen <- function(x){n <- x; function() {n <- n + 1; n}}; py_iterator(seq_gen(9))`

## Helpers

`py_capture_output(expr, type = c("stdout", "stderr"))` Capture and return Python output. Also `py_suppress_warnings()`.

`py_get_attr(x, name, silent = FALSE)` Get an attribute of a Python object. Also `py_set_attr()`, `py_has_attr()`, and `py_list_attributes()`.

`py_help(object)` Open the documentation page for a Python object. `py_help(sns)`

`py_last_error()` Get the last Python error encountered. Also `py_clear_last_error()` to clear the last error. `py_last_error()`

`py_save_object(object, filename, pickle = "pickle", ...)` Save and load Python objects with pickle. Also `py_load_object()`. `py_save_object(x, "x.pickle")`

`with(data, expr, as = NULL, ...)` Evaluate an expression within a Python context manager.  
`py <- import_builtins(); with(py$open("output.txt", "w") %as% file, {file$write("Hello, there!")})`

## Python in R

Call Python from R code in three ways:

### IMPORT PYTHON MODULES

Use `import()` to import any Python module. Access the attributes of a module with `$`.

- `import(module, as = NULL, convert = TRUE, delay_load = FALSE)` Import a Python module. If `convert = TRUE`, Python objects are converted to their equivalent R types. Also `import_from_path()`. `import("pandas")`
- `import_main(convert = TRUE)` Import the main module, where Python executes code by default. `import_main()`
- `import_builtins(convert = TRUE)` Import Python's built-in functions. `import_builtins()`

### SOURCE PYTHON FILES

Use `source_python()` to source a Python script and make the Python functions and objects it creates available in the calling R environment.

- `source_python(file, envir = parent.frame(), convert = TRUE)` Run a Python script, assigning objects to a specified R environment. `source_python("file.py")`

### RUN PYTHON CODE

Execute Python code into the **main** Python module with `py_run_file()` or `py_run_string()`.

- `py_run_string(code, local = FALSE, convert = TRUE)` Run Python code (passed as a string) in the main module. `py_run_string("x = 10"); py$x`
- `py_run_file(file, local = FALSE, convert = TRUE)` Run Python file in the main module. `py_run_file("script.py")`
- `py_eval(code, convert = TRUE)` Run a Python expression, return the result. `py_eval("1 + 1")`

Access the results, and anything else in Python's **main** module, with `py`.

- `py` An R object that contains the Python main module and the results stored there. `py$x`



# Python in the IDE

Syntax highlighting for Python scripts and chunks.

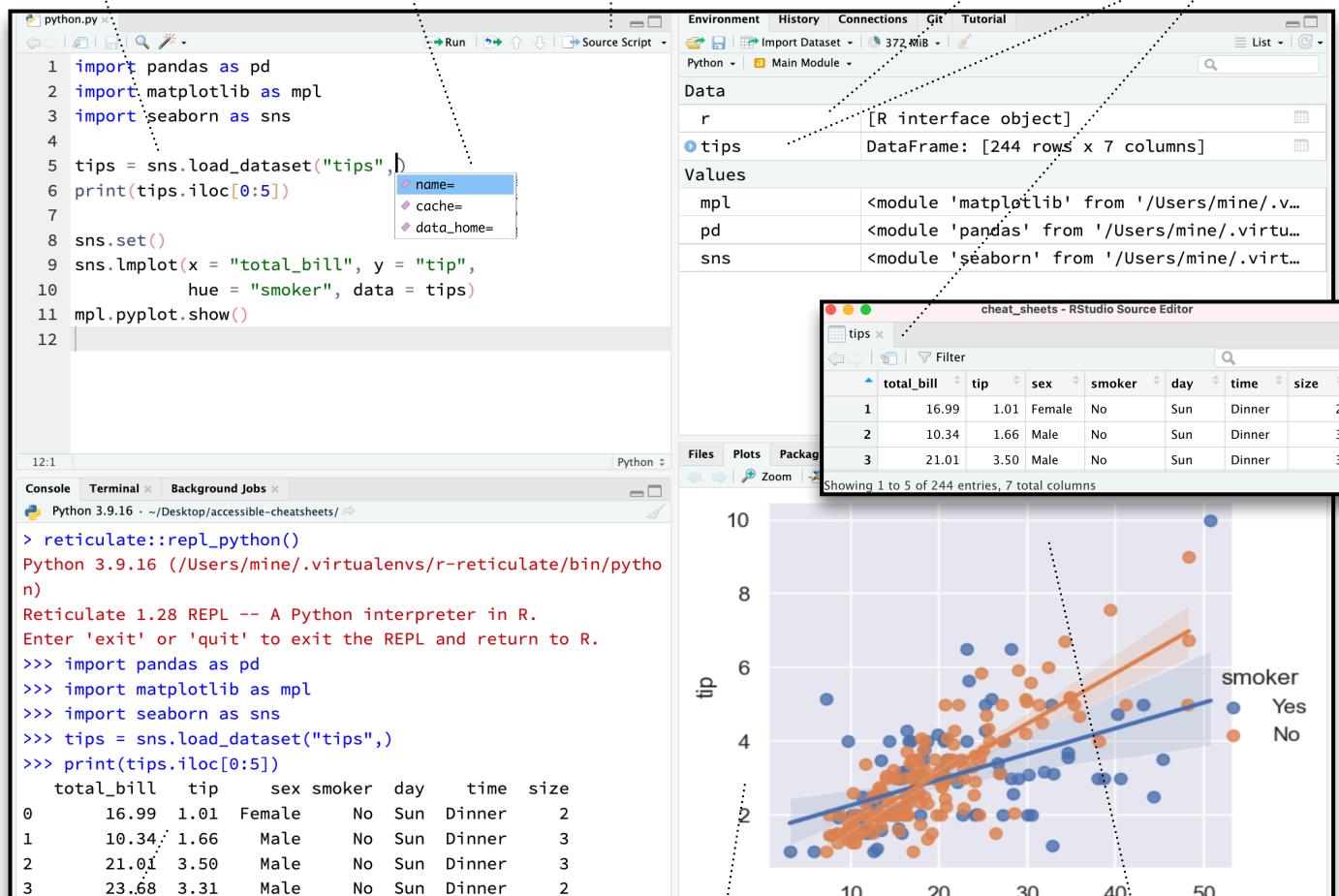
Tab completion for Python functions and objects (and Python modules imported in R scripts).

Source Python scripts.

Execute Python code line by line with **Cmd + Enter** (**Ctrl + Enter**).

View Python objects in the Environment Pane.

View Python objects in the Data Viewer.



A Python REPL opens in the console when you run Python code with a keyboard shortcut. Type **exit** to close.

## Python REPL

A REPL (Read, Eval, Print Loop) is a command line where you can run Python code and view the results.

1. Open in the console with **repl\_python()**, or by running code in a Python script with **Cmd + Enter** (**Ctrl + Enter**).
2. Type commands at **>>>** prompt.
3. Press **Enter** to run code.
4. Type **exit** to close and return to R console.

```
Console Terminal x Background Jobs x
> reticulate::repl_python()
Python 3.9.16 (/Users/mine/.virtualenvs/r-reticulate/bin/python)
Reticulate 1.28 REPL -- A Python interpreter in R.
Enter 'exit' or 'quit' to exit the REPL and return to R.

>>> import seaborn as sns
>>> tips = sns.load_dataset("tips")
>>> tips.shape
(244, 7)
>>> exit
> |
```

# Configure Python

Reticulate binds to a local instance of Python when you first call **import()** directly or implicitly from an R session. To control the process, find or build your desired Python instance. Then suggest your instance to reticulate. **Restart R to unbind**.

## Find Python

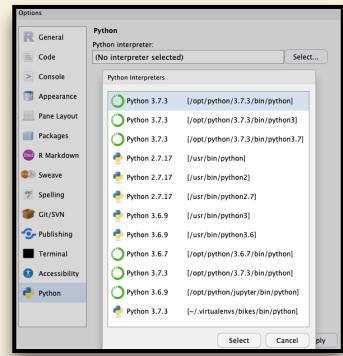
- **install\_python(version, list = FALSE, force = FALSE)** Download and install Python. `install_python("3.9.16")`
- **py\_available(initialize = FALSE)** Check if Python is available on your system. Also **py\_module\_available()** and **py\_numpy\_module()**. `py_available()`
- **py\_discover\_config()** Return the detected installation of Python. Use **py\_config()** to check which version has been loaded. `py_config()`
- **virtualenv\_list()** List all available virtual environments. Also **virtualenv\_root()**. `virtualenv_list()`
- **conda\_list(conda = "auto")** List all available conda environments. Also **conda\_binary()** and **conda\_version()**. `conda_list()`

## Suggest an env to use

Set a default Python interpreter in the RStudio IDE Global or Project Options.

Go to **Tools > Global Options... > Python** for Global Options.

Within a project, go to **Tools > Project Options... > Python**.



Otherwise, reticulate scans the instances on your computer in the following order:

1. The instance referenced by the environment variable **RETICULATE\_PYTHON** (if specified). **Tip: set in .Renviron file.**
  - **Sys.setenv(RETICULATE\_PYTHON = PATH)** Set default Python binary. Persists across sessions! Undo with **Sys.unsetenv()**. `Sys.setenv(RETICULATE_PYTHON = "/usr/local/bin/python")`
2. The instances referenced by **use\_** functions if called before **import()**.
  - **use\_python(python)** Path to a Python binary. `use_python("/usr/local/bin/python")`
  - **use\_virtualenv(virtualenv)** Path to or name of a Python virtualenv. `use_virtualenv("~/myenv")` `use_virtualenv("r-keras")`
3. A virtual env found in the current working directory: `"/.venv"`
4. Environments that are named after the imported module. e.g. `~/.virtualenvs/r-scipy/` for `import("scipy")`
5. The package default virtualenv, "r-reticulate".
6. At the location of the Python binary discovered on the system PATH (i.e. `Sys.which("python")`)

## Create a Python env

- **virtualenv\_create(envname = NULL, ...)** Create a new virtual environment. `virtualenv_create("r-pandas")`
- **conda\_create(envname = NULL, ...)** Create a new conda environment. `conda_create("r-pandas", packages = "pandas")`

## Install Packages

Install Python packages with R (below) or the shell:  
**pip install SciPy**  
**conda install SciPy**

- **py\_install(packages, envname, ...)** Installs Python packages into a Python env. `py_install("pandas")`
- **virtualenv\_install(envname, packages, ...)** Install a package within a virtualenv. Also **virtualenv\_remove()**. `virtualenv_install("r-pandas", packages = "pandas")`
- **conda\_install(envname, packages, ...)** Install a package within a conda env. Also **conda\_remove()**. `conda_install("r-pandas", packages = "plotly")`

# Google Earth Engine with rgee :: CHEAT SHEET



## Mission

The goal of rgee is to offer a user-friendly interface for analyzing spatial data on the Google Earth Engine (GEE) platform using the R programming language.

## Installation

	■ It is necessary to have <code>Rtools</code> installed. ▶ <code>install.package("rgee")</code>
	■ In terminal execute, as follow:  user:~\$ sudo apt install libjq-dev user:~\$ sudo apt install libprotobuf-dev user:~\$ sudo apt install protobuf-compiler ▶ <code>install.package("rgee")</code>
	■ In terminal execute, as follow:  user:~\$ docker run -d -p 8787:8787 -e USER=rgee -e PASSWORD=rgee --name rgee-dev csaybar/rgee

For **Python requirements installation**, use `ee_install`:

```
> rgee::ee_install()
```

only run once  
rgee is installed

See the **Python** section in [rgeebok](#) for more details.

## Hello world Earth Engine

```
> library("rgee")
> ee_Initialize(user, drive, gcs)

GEE username (Optional)
Connect GEE with GD.
Connect GEE with GCS.

# Earth Engine API style (chaining methods)
> ee$string("Hello World from Earth Engine!")$getInfo()

Fetch and return information. From GEE server to local.

> [1] "Hello World from Earth Engine!"
```

## Pipe integration %>%

Pipe operator has been included into rgee to provide functional programming style.

```
# Earth Engine API with pipes style
> ee$string("Hello World from Earth Engine!")%>%
  ee$string$getInfo()

> [1] "Hello World from Earth Engine!"
```

## Basic classes

Basic data structures available in GEE..

Type	Class	Example
Number	ee\$Number	> ee\$Number(2021)
String	ee\$String	> ee\$String("Hello")
List	ee\$List	> ee\$List(c("Hi", "amy"))
Dictionary	ee\$Dictionary	> ee\$Dictionary(list(year = 2021))
Array	ee\$Array	> ee\$Array(26, 9, 2021)
Date	ee\$Date	> ee\$Date("1990-01-01")

## ee\$Geometry

A collection of geometric forms that describe an object spatially.

Type	Geom	Function
Point	•	<b>ee\$Geometry\$Point</b> <code>sf::st_point</code>
LineString	—	<b>ee\$Geometry\$LineString</b> <code>sf::st_linestring</code>
LineRing	—	<b>ee\$Geometry\$LineRing</b> <code>sf::st_linestring</code>
Polygon	—	<b>ee\$Geometry\$Polygon</b> <code>sf::st_polygon</code>
Multipoint	••	<b>ee\$Geometry\$Multipoint</b> <code>sf::st_multipoint</code>
MultiLineString	—	<b>ee\$Geometry\$MultiLineString</b> <code>sf::st_multilinestring</code>
MultiGeometry	—	<b>ee\$Geometry\$MultiGeometry</b> <code>sf::st_geometrycollection</code>

## Geometric operations

Type	Function
Buffer	<code>*\$buffer</code>
Intersection	<code>*\$intersection</code>
Union	<code>*\$union</code>
Difference	<code>*\$difference</code>
Symmetric difference	<code>*\$symmetricdifference</code>

(\* : The symbol mean is a type of GEE geometry, for example : a ee\$Geometry\$Polygon)

## Data catalog

The Earth Engine catalogue can be accessed interactively from R with rgee.

Function	Example
<code>ee_utils_dataset_display</code>	> ee_utils_dataset_display("Landsat")

## Visualization

rgee supports the visualization of spatial Earth Engine objects such as Image, ImageCollection, Feature, FeatureCollection, and allows users to customize the legend using the `Map$addLegend` method.

Object	Geom	Method	Arguments
Image		<b>Map\$addLayer</b>	<ul style="list-style-type: none"> <li>■ <code>eeObject*</code></li> <li>■ <code>VisParams</code></li> <li>■ <code>name</code></li> <li>■ <code>show</code></li> <li>■ <code>opacity</code></li> </ul>
Feature			
FeatureCollection			
ImageCollection			
		<b>Map\$addLayers</b>	■ <code>nmax</code>

\* `eeObject` can also be a Cloud Optimized GeoTIFF (COG) file.

`Map$Legend` needs that users pass the same `visParams` used in `Map$addLayer`.

Data	Function	Type
Categorical	<code>Map\$addLegend(...)</code>	color_mapping = "categorical"
Continue		color_mapping = "continue"
Discrete		color_mapping = "discrete"
Customize		color_mapping = "character"

## Example

```
> image <- ee$Image$Dataset$CGIAR_SRTM90_V4
> visparams <- list(min = 0, max = 3000)
> m1 <- Map$addLayer(image, visparams, "DEM")
> m1 + Map$addLegend(visparams, "DEM", "bottomright", 8)

rgee also supports the metadata display of GEE spatial objects (ee_print).

> ee$Image("CGIAR/SRTM90_V4") %>% ee_print(srtm)
```

# Google Earth Engine with rgee :: CHEAT SHEET

## Considerations

Some issues can occurs when reticulate translate the R code into Python. We detected four cases:

1. map method in ee\$List objects.

**Solution:** Use `ee_utils_pyfunc`.

2. Strict integer number data type.

**Solution:** Add "L" at the end. For instance: `> ee$Number(20L)`

3. Be careful with ee\$Date objects.

**Solution:** Use `eedate_to_rdate` and `rdate_to_eedate`.

4. Reserved words.

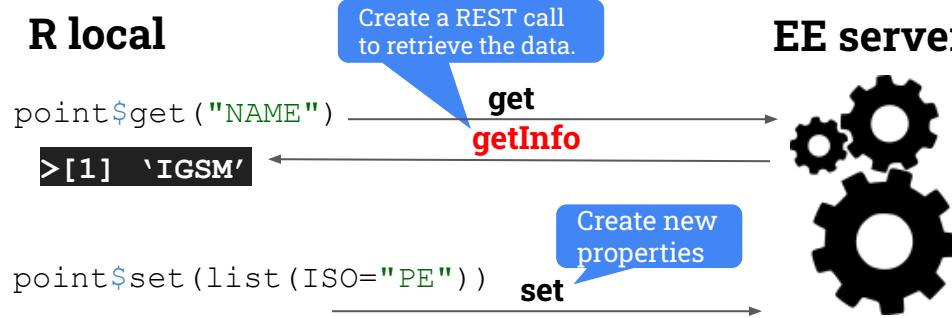
**Solution:** Use quotation marks. For instance: `> x$'repeat'(20, 2)`

## ee\$Feature

It is an GEE geometry + properties.

```
> xy <- c(-77.08643, -12.05536)           Longitude and
> geom <- ee$Geometry$Point(xy)             latitude in a vector
> props <- list(ID = 1, NAME = "IGSM")
> point <- ee$Feature(geom, props)
```

### R local



## ee\$FeatureCollection

It is an set of GEE features + properties.

```
> minmax <- c(-77.08, -12.05, -77.08, -12.05)
> box <- ee$Geometry$Rectangle(minmax)
> lf1 <- ee$Feature(box, list(ISO="PE"))
> lf2 <- ee$Feature(box, list(ISO="RU"))
> prps <- list(ID=1, NAME="polygons")
> fc <- ee$FeatureCollection(c(lf1, lf2), prps)
> print(fc)
```

```
> EarthEngine Object: FeatureCollection
```

## ee\$image

It is an set of bands. An band is array of values + properties.

```
> image1 <- ee$image(1)           Create a constant image
> image1
> EarthEngine Object: Image
> image2 <- ee$image(2)
> list_img <- list(image1, image2)
> image3 <- ee$image(list_img)
> image3
```

**EarthEngine Object: Image**

Create a constant image

Concatenate two single-band images into one multi-band image

## Image I/O

Functions	FROM	TO	RETURN
ee_as_raster	EE server	Local	R object
ee_image_to_asset	EE server	EE asset	Unstarted task
ee_image_to_gcs	EE server	GCS	Unstarted task
ee_image_to_drive	EE server	GD	Unstarted task
ee_as_stars	EE server	Local	R object
raster_as_ee	Local	EE server	GEE object
stars_as_ee	Local	EE server	GEE object

## ee\$imageCollection

It is an set of GEE images + properties.

```
> ic <- ee$imageCollection(list_img)
> ic
> EarthEngine Object: ImageCollection
```

## ImageCollection I/O

Functions	FROM	TO	RETURN
ee_get_date_ic	EE server	Local	R data.frame
ee_imagecollection_to_local	EE server	Local	R object

## FeatureCollection Export (Table)

Set of functions to fetch and return GEE FeatureCollections.

Functions	FROM	TO	RETURN
gcs_to_ee_table	GCS	EE server	Unstarted task
ee_as_sf	EE server	Local	R object
ee_table_to_drive	EE server	GD	Unstarted task
ee_table_to_gcs	EE server	GCS	Unstarted task
ee_table_to_asset	EE server	EE asset	Unstarted task
sf_as_ee	Local	EE server	GEE object

## GEE Asset Manager

Set of functions to interact with the GEE asset manager.

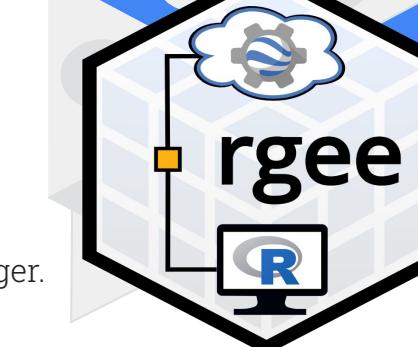
Batch operations are supported.

### FUNCTIONS

```
ee_manage_create
ee_manage_delete
ee_manage_assetlist
ee_manage_quota
ee_manage_copy
ee_manage_move
ee_manage_set_properties
ee_manage_delete_properties
ee_manage_asset_access
ee_manage_task
ee_manage_cancel_all_running_task
```

### DESCRIPTION

Create an empty folder or ic.  
Delete an GEE asset.  
List files in a folder or ic.  
Show user GEE quota.  
Copy a paste GEE asset.  
Cut and paste a GEE asset.  
Set GEE asset properties.  
Delete GEE asset properties.  
Change IAM policy.  
Show the task's user history.  
Cancel all the running task.



## Custom Animations

Auxiliary functions to create GIF files with Earth Engine. They depend of the magick package. `rgeeExtra` now include these functions.

### FUNCTIONS

```
ee_utils_gif_annotation
ee_utils_gif_creator
ee_utils_gif_save
```

### DESCRIPTION

Add text to a GIF.  
From ee\$imageCollection to GIF.  
Write a magick object as a GIF file.

## Miscellaneous

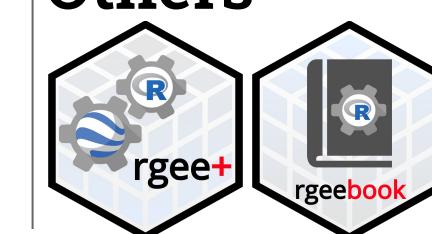
### FUNCTIONS

```
ee_utils_create_json
ee_utils_create_manifest_image
ee_utils_create_manifest_table
ee_utils_dataset_display
ee_utils_future_value
ee_utils_get_crs
ee_utils_py_to_r
ee_utils_pyfunc
ee_utils_shp_to_zip
ee_utils_cog_metadata
```

### DESCRIPTION

Convert a R list into a JSON.  
GEE Image manifest creator.  
GEE Table manifest creator.  
Search into the GEE Data Catalog.  
Return the future values object.  
Convert SR-ORG into a OGC WKT.  
Translate Python objects to R.  
Wrap a R function in Python.  
Create a zip from an sf object.  
Metadata of a COG tile server.

## Others



This cheatsheet was created using the `rgee` reference manual and the `rgee vignettes`. Visit the `rgeebook` for additional information about this package.

# rmarkdown :: CHEATSHEET

## What is rmarkdown?



**.Rmd files** • Develop your code and ideas side-by-side in a single document. Run code as individual chunks or as an entire document.

**Dynamic Documents** • Knit together plots, tables, and results with narrative text. Render to a variety of formats like HTML, PDF, MS Word, or MS Powerpoint.

**Reproducible Research** • Upload, link to, or attach your report to share. Anyone can read or run your code to reproduce your work.

## Workflow

- 1 Open a **new .Rmd file** in the RStudio IDE by going to *File > New File > R Markdown*.
- 2 **Embed code** in chunks. Run code by line, by chunk, or all at once.
- 3 **Write text** and add tables, figures, images, and citations. Format with Markdown syntax or the RStudio Visual Markdown Editor.
- 4 **Set output format(s) and options** in the YAML header. Customize themes or add parameters to execute or add interactivity with Shiny.
- 5 **Save and render** the whole document. Knit periodically to preview your work as you write.
- 6 **Share your work!**

## Embed Code with knitr

### CODE CHUNKS

Surround code chunks with `{{r}}` and `{{` or use the Insert Code Chunk button. Add a chunk label and/or chunk options inside the curly braces after {{r}}.

```
```{r chunk-label, include=FALSE}
summary(mtcars)
```
```

### SET GLOBAL OPTIONS

Set options for the entire document in the first chunk.

```
```{r include=FALSE}
knitr::opts_chunk$message = FALSE
```
```

### INLINE CODE

Insert `{{r <code>}}` into text sections. Code is evaluated at render and results appear as text.

"Built with `r getRversion()`" --> "Built with 4.1.0"



The screenshot shows the RStudio IDE with the Source Editor open. A numbered callout path highlights the workflow: 1. New File (File menu), 2. Embed Code (Knit on Save button), 3. Write Text (Visual tab), 4. Set Output Format(s) and Options (YAML header), 5. Save and Render (Knit button), and 6. Share (Publish dropdown).

The screenshot shows the RStudio IDE with the Visual Editor tab selected. A numbered callout path highlights the workflow: 1. New File (File menu), 2. Embed Code (Knit on Save button), 3. Write Text (Visual tab), 4. Set Output Format(s) and Options (YAML header), 5. Save and Render (Knit button), and 6. Share (Publish dropdown).

The screenshot shows the RStudio IDE with the Render tab selected. A numbered callout path highlights the workflow: 1. New File (File menu), 2. Embed Code (Knit on Save button), 3. Write Text (Visual tab), 4. Set Output Format(s) and Options (YAML header), 5. Save and Render (Knit button), and 6. Share (Publish dropdown).

## Insert Citations

Create citations from a bibliography file, a Zotero library, or from DOI references.

### BUILD YOUR BIBLIOGRAPHY

- Add BibTeX or CSL bibliographies to the YAML header.
 

```
---
title: "My Document"
bibliography: references.bib
link-citations: TRUE
---
```
- If Zotero is installed locally, your main library will automatically be available.
- Add citations by DOI by searching "from DOI" in the **Insert Citation** dialog.

### INSERT CITATIONS

- Access the **Insert Citations** dialog in the Visual Editor by clicking the @ symbol in the toolbar or by clicking **Insert > Citation**.
- Add citations with markdown syntax by typing **[@cite]** or **@cite**.

## Insert Tables

Output data frames as tables using **kable**(data, caption).

```
```{r}
data <- faithful[1:4, ]
knitr::kable(data,
             caption = "Table with kable")
```
```

Other table packages include **flextable**, **gt**, and **kableExtra**.



## Write with Markdown

The syntax on the left renders as the output on the right.

Plain text.

Plain text.

End a line with two spaces to start a new paragraph.

End a line with two spaces to start a new paragraph.

Also end with a backslash\ to make a new line.

Also end with a backslash\ to make a new line.

**italics\*** and **\*\*bold\*\***

**italics** and **bold**

**superscript**<sup>2</sup>/**subscript**<sub>2</sub>

**superscript**<sup>2</sup>/**subscript**<sub>2</sub>

**~~strikethrough~~**

**strikethrough**

**escaped:** `* \_ \_`

**escaped:** `* \_ \_`

**endash:** `--`, **emdash:** `---`

**endash:** `--`, **emdash:** `---`

## Header 1 Header 2

...

**Header 6**

unordered list

unordered list

- item 2

- item 2

- item 2a (indent 1 tab)

- item 2b

1. ordered list

1. ordered list

2. item 2

2. item 2

- item 2a (indent 1 tab)

- item 2b

<link url>

<http://www.posit.co/>

[This is a link.](link url)

This is a link.

[This is another link][id].

This is another link.

At the end of the document:

[id]: link url

![Caption](image.png)

or !![Caption][id2]

At the end of the document:

[id2]: image.png

`verbatim code`

```

multiple lines of verbatim code

```

> block quotes

block quotes

equation:  $e^{i\pi} + 1 = 0$

equation block:

$$E = mc^2$$

horizontal rule:

---

| Right | Left | Default | Center |

|-----:|-----:|-----:|-----:|

| 12 | 12 | 12 | 12 |

| 123 | 123 | 123 | 123 |

| 1 | 1 | 1 | 1 |

| 1 | 1 | 1 | 1 |

| Right | Left | Default | Center |
|-------|------|---------|--------|
| 12    | 12   | 12      | 12     |
| 123   | 123  | 123     | 123    |
| 1     | 1    | 1       | 1      |

### HTML Tabsets

## Results {tabset}

## Plots

text

## Tables

more text

### Results

Plots

Tables

text



# Set Output Formats and their Options in YAML

Use the document's YAML header to set an **output format** and customize it with **output options**.

```
---
```

```
title: "My Document"
author: "Author Name"
output:
  html_document:
    toc: TRUE
```

**Indent format 2 characters,  
indent options 4 characters**

| OUTPUT FORMAT           | CREATES                      |
|-------------------------|------------------------------|
| html_document           | .html                        |
| pdf_document*           | .pdf                         |
| word_document           | Microsoft Word (.docx)       |
| powerpoint_presentation | Microsoft Powerpoint (.pptx) |
| odt_document            | OpenDocument Text            |
| rtf_document            | Rich Text Format             |
| md_document             | Markdown                     |
| github_document         | Markdown for Github          |
| ioslides_presentation   | ioslides HTML slides         |
| slidy_presentation      | Slidy HTML slides            |
| beamer_presentation*    | Beamer slides                |

\* Requires LaTeX, use `tinytex::install_tinytex()`  
Also see `flexdashboard`, `bookdown`, `distill`, and `blogdown`.

| IMPORTANT OPTIONS   | DESCRIPTION                                                                            | HTML    | PDF | MS Word | MS PPT |
|---------------------|----------------------------------------------------------------------------------------|---------|-----|---------|--------|
| anchor_sections     | Show section anchors on mouse hover (TRUE or FALSE)                                    | X       |     |         |        |
| citation_package    | The LaTeX package to process citations ("default", "natbib", "biblatex")               | X       |     |         |        |
| code_download       | Give readers an option to download the .Rmd source code (TRUE or FALSE)                | X       |     |         |        |
| code_folding        | Let readers to toggle the display of R code ("none", "hide", or "show")                | X       |     |         |        |
| css                 | CSS or SCSS file to use to style document (e.g. "style.css")                           | X       |     |         |        |
| dev                 | Graphics device to use for figure output (e.g. "png", "pdf")                           | X X     |     |         |        |
| df_print            | Method for printing data frames ("default", "kable", "tibble", "paged")                | X X X X |     |         |        |
| fig_caption         | Should figures be rendered with captions (TRUE or FALSE)                               | X X X X |     |         |        |
| highlight           | Syntax highlighting ("tango", "pygments", "kate", "zenburn", "textmate")               | X X X   |     |         |        |
| includes            | File of content to place in doc ("in_header", "before_body", "after_body")             | X X     |     |         |        |
| keep_md             | Keep the Markdown .md file generated by knitting (TRUE or FALSE)                       | X X X X |     |         |        |
| keep_tex            | Keep the intermediate TEX file used to convert to PDF (TRUE or FALSE)                  | X       |     |         |        |
| latex_engine        | LaTeX engine for producing PDF output ("pdflatex", "xelatex", or "lualatex")           | X       |     |         |        |
| reference_docx/_doc | docx/pptx file containing styles to copy in the output (e.g. "file.docx", "file.pptx") | X X     |     |         |        |
| theme               | Theme options (see Bootswatch and Custom Themes below)                                 | X       |     |         |        |
| toc                 | Add a table of contents at start of document (TRUE or FALSE)                           | X X X X |     |         |        |
| toc_depth           | The lowest level of headings to add to table of contents (e.g. 2, 3)                   | X X X X |     |         |        |
| toc_float           | Float the table of contents to the left of the main document content (TRUE or FALSE)   | X       |     |         |        |

Use `?<output format>` to see all of a format's options, e.g. `?html_document`

## More Header Options

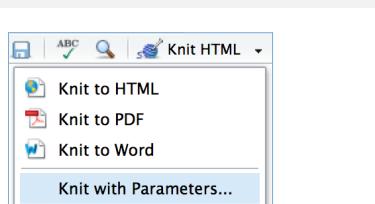
### PARAMETERS

Parameterize your documents to reuse with new inputs (e.g., data, values, etc.).

1. **Add parameters** in the header as sub-values of `params`.
2. **Call parameters** in code using `params$<name>`.
3. **Set parameters** with Knit with Parameters or the `params` argument of `render()`.

### REUSABLE TEMPLATES

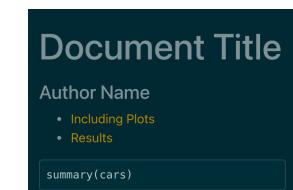
1. **Create a new package** with a `inst/rmarkdown/templates` directory.
2. **Add a folder** containing `template.yaml` (below) and `skeleton.Rmd` (template contents).
3. **Install** the package to access template by going to **File > New R Markdown > From Template**.



### BOOTSWATCH THEMES

Customize HTML documents with Bootswatch themes from the `bslib` package using the theme output option.

Use `bslib::bootswatch_themes()` to list available themes.



```
---
```

```
title: "Document Title"
author: "Author Name"
output:
  html_document:
    theme:
      bootswatch: solar
```

### CUSTOM THEMES

Customize individual HTML elements using `bslib` variables. Use `?bs_theme` to see more variables.

```
---
```

```
output:
  html_document:
    theme:
      bg: "#121212"
      fg: "#E4E4E4"
      base_font:
        google: "Prompt"
```

More on `bslib` at [pkgs.rstudio.com/bslib/](https://pkgs.rstudio.com/bslib/).

### STYLING WITH CSS AND SCSS

Add CSS and SCSS to your document by adding a path to a file with the `css` option in the YAML header.

```
---
```

```
title: "My Document"
author: "Author Name"
output:
  html_document:
    css: "style.css"
```

Apply CSS styling by writing HTML tags directly or:

- Use markdown to apply style attributes inline.

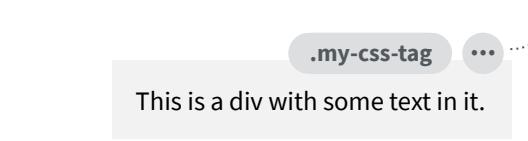
Bracketed Span  
A [green]{.my-color} word.

A green word.

Fenced Div  
:::{.my-color}  
All of these words  
are green.  
:::

All of these words  
are green.

- Use the Visual Editor. Go to **Format > Div/Span** and add CSS styling directly with Edit Attributes.



This is a div with some text in it.

## Render

When you render a document, rmarkdown:

1. Runs the code and embeds results and text into an .md file with knitr.
2. Converts the .md file into the output format with Pandoc.



**Save**, then **Knit** to preview the document output. The resulting HTML/PDF/MS Word/etc. document will be created and saved in the same directory as the .Rmd file.

Use `rmarkdown::render()` to render/knit in the R console. See `?render` for available options.

## Share

### Publish on Posit Connect

to share R Markdown documents securely, schedule automatic updates, and interact with parameters in real-time.

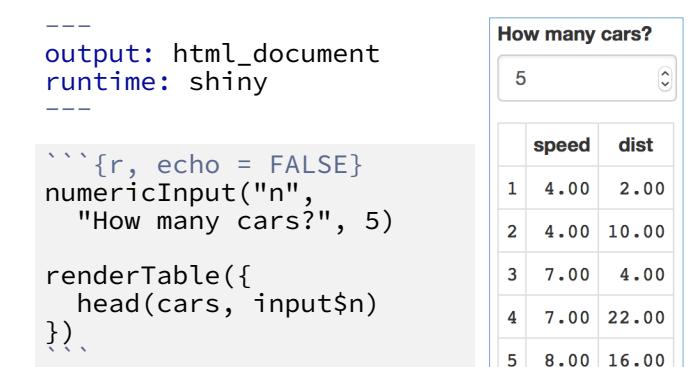
[posit.co/products/enterprise/connect](https://posit.co/products/enterprise/connect).



### INTERACTIVITY

Turn your report into an interactive Shiny document in 4 steps:

1. Add `runtime: shiny` to the YAML header.
2. Call Shiny input functions to embed input objects.
3. Call Shiny render functions to embed reactive output.
4. Render with `rmarkdown::run()` or click **Run Document** in RStudio IDE.



Also see Shiny Prerendered for better performance. [rmarkdown.rstudio.com/authoring\\_shiny\\_prerendered](https://rmarkdown.rstudio.com/authoring_shiny_prerendered).

Embed a complete app into your document with `shiny::shinyAppDir()`. More at [bookdown.org/yihui/rmarkdown/shiny-embedded.html](https://bookdown.org/yihui/rmarkdown/shiny-embedded.html).

# rmarkdown :: CHEAT SHEET

## What is rmarkdown?



**.Rmd files** • Develop your code and ideas side-by-side in a single document. Run code as individual chunks or as an entire document.

**Dynamic Documents** • Knit together plots, tables, and results with narrative text. Render to a variety of formats like HTML, PDF, MS Word, or MS Powerpoint.

**Reproducible Research** • Upload, link to, or attach your report to share. Anyone can read or run your code to reproduce your work.

## Workflow

- 1 Open a **new .Rmd file** in the RStudio IDE by going to *File > New File > R Markdown*.
- 2 **Embed code** in chunks. Run code by line, by chunk, or all at once.
- 3 **Write text** and add tables, figures, images, and citations. Format with Markdown syntax or the RStudio Visual Markdown Editor.
- 4 **Set output format(s) and options** in the YAML header. Customize themes or add parameters to execute or add interactivity with Shiny.
- 5 **Save and render** the whole document. Knit periodically to preview your work as you write.
- 6 **Share your work!**

## Embed Code with knitr

### CODE CHUNKS

Surround code chunks with `{{r}}` and `{{` or use the Insert Code Chunk button. Add a chunk label and/or chunk options inside the curly braces after **r**.

```
```{r chunk-label, include=FALSE}
summary(mtcars)
```
```

### SET GLOBAL OPTIONS

Set options for the entire document in the first chunk.

```
```{r include=FALSE}
knitr::opts_chunk$message = FALSE
```
```

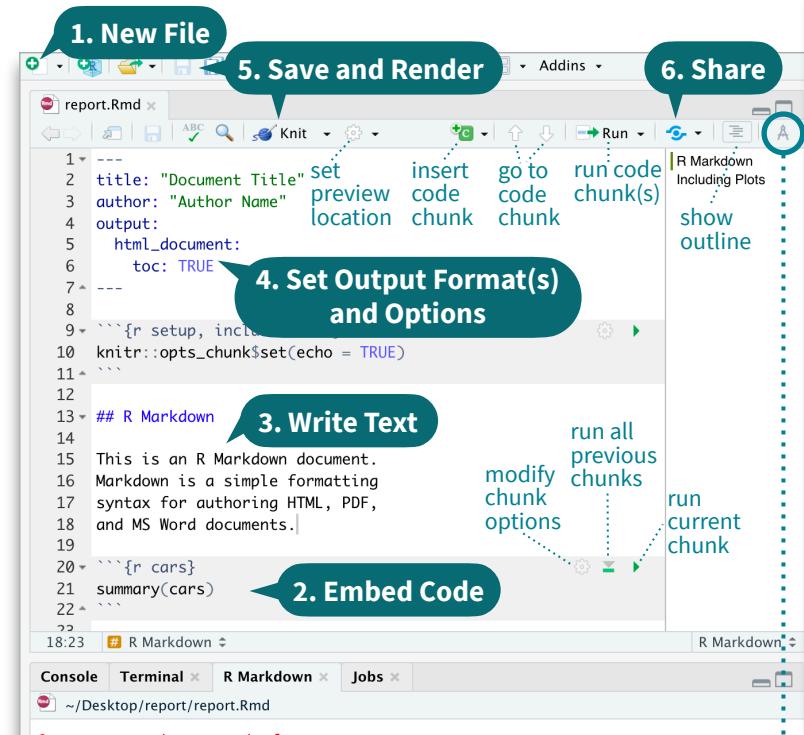
### INLINE CODE

Insert `r <code>` into text sections. Code is evaluated at render and results appear as text.

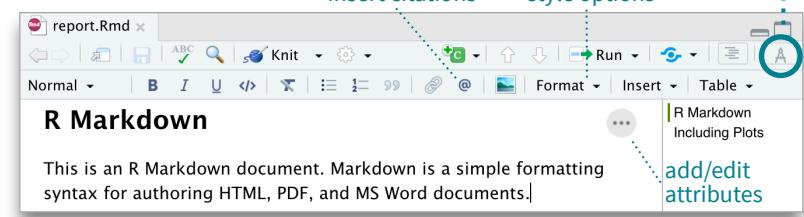
"Built with `r getRversion()`" --> "Built with 4.1.0"



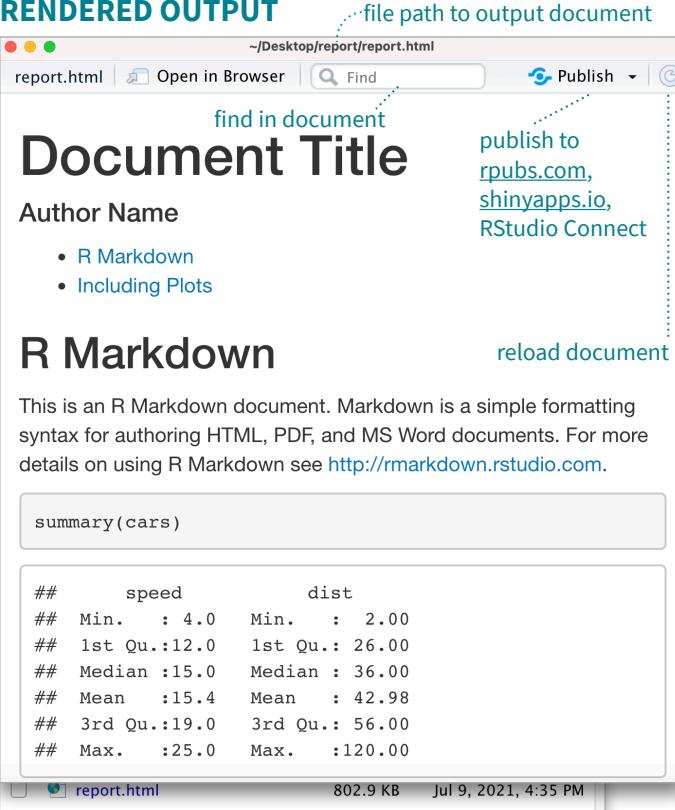
### SOURCE EDITOR



### VISUAL EDITOR



### RENDERED OUTPUT



## Write with Markdown

The syntax on the left renders as the output on the right.

Plain text.

Plain text.

End a line with two spaces to start a new paragraph.

End a line with two spaces to start a new paragraph.

Also end with a backslash\ to make a new line.

Also end with a backslash\ to make a new line.

**italics\*** and **\*\*bold\*\***

**italics** and **bold**

superscript<sup>2</sup>/subscript<sub>2</sub>

superscript<sup>2</sup>/subscript<sub>2</sub>

~~strikethrough~~

strikethrough

escaped: `\*`\\`\_`

escaped: \* \\\_

endash: --, emdash: ---

endash: -, emdash: -

### Header 1 Header 2

...

Header 6

- unordered list

• item 2

- item 2a (indent 1 tab)

• item 2b

1. ordered list

2. item 2

- item 2a (indent 1 tab)

• item 2b

<link url>

[This is a link.](link url)

[This is another link][id].

This is another link.

<http://www.rstudio.com/>

This is a link.

This is another link.



Caption.

verbatim code

multiple lines of verbatim code

> block quotes

block quotes

equation:  $e^{i\pi} + 1 = 0$

equation block:

$$E = mc^2$$

horizontal rule:

| Right | Left | Default | Center |
|-------|------|---------|--------|
| 12    | 12   | 12      | 12     |
| 123   | 123  | 123     | 123    |
| 1     | 1    | 1       | 1      |

### HTML Tabsets

```
# Results {.tabset}
## Plots text
text
```

## Tables
more text

### Results

|       |        |
|-------|--------|
| Plots | Tables |
|-------|--------|

text





# Set Output Formats and their Options in YAML

Use the document's YAML header to set an **output format** and customize it with **output options**.

```
---
```

```
title: "My Document"
author: "Author Name"
output:
  html_document:
    toc: TRUE
---
```

**Indent format 2 characters,  
indent options 4 characters**

| OUTPUT FORMAT           | CREATES                      |
|-------------------------|------------------------------|
| html_document           | .html                        |
| pdf_document*           | .pdf                         |
| word_document           | Microsoft Word (.docx)       |
| powerpoint_presentation | Microsoft Powerpoint (.pptx) |
| odt_document            | OpenDocument Text            |
| rtf_document            | Rich Text Format             |
| md_document             | Markdown                     |
| github_document         | Markdown for Github          |
| ioslides_presentation   | ioslides HTML slides         |
| slidy_presentation      | Slidy HTML slides            |
| beamer_presentation*    | Beamer slides                |

\* Requires LaTeX, use `tinytex::install_tinytex()`  
Also see `flexdashboard`, `bookdown`, `distill`, and `blogdown`.

| IMPORTANT OPTIONS   | DESCRIPTION                                                                            | HTML    | PDF | MS Word | MS PPT |
|---------------------|----------------------------------------------------------------------------------------|---------|-----|---------|--------|
| anchor_sections     | Show section anchors on mouse hover (TRUE or FALSE)                                    | X       |     |         |        |
| citation_package    | The LaTeX package to process citations ("default", "natbib", "biblatex")               | X       |     |         |        |
| code_download       | Give readers an option to download the .Rmd source code (TRUE or FALSE)                | X       |     |         |        |
| code_folding        | Let readers to toggle the display of R code ("none", "hide", or "show")                | X       |     |         |        |
| css                 | CSS or SCSS file to use to style document (e.g. "style.css")                           | X       |     |         |        |
| dev                 | Graphics device to use for figure output (e.g. "png", "pdf")                           | X X     |     |         |        |
| df_print            | Method for printing data frames ("default", "kable", "tibble", "paged")                | X X X X |     |         |        |
| fig_caption         | Should figures be rendered with captions (TRUE or FALSE)                               | X X X X |     |         |        |
| highlight           | Syntax highlighting ("tango", "pygments", "kate", "zenburn", "textmate")               | X X X   |     |         |        |
| includes            | File of content to place in doc ("in_header", "before_body", "after_body")             | X X     |     |         |        |
| keep_md             | Keep the Markdown .md file generated by knitting (TRUE or FALSE)                       | X X X X |     |         |        |
| keep_tex            | Keep the intermediate TEX file used to convert to PDF (TRUE or FALSE)                  | X       |     |         |        |
| latex_engine        | LaTeX engine for producing PDF output ("pdflatex", "xelatex", or "lualatex")           | X       |     |         |        |
| reference_docx/_doc | docx/pptx file containing styles to copy in the output (e.g. "file.docx", "file.pptx") | X X     |     |         |        |
| theme               | Theme options (see Bootswatch and Custom Themes below)                                 | X       |     |         |        |
| toc                 | Add a table of contents at start of document (TRUE or FALSE)                           | X X X X |     |         |        |
| toc_depth           | The lowest level of headings to add to table of contents (e.g. 2, 3)                   | X X X X |     |         |        |
| toc_float           | Float the table of contents to the left of the main document content (TRUE or FALSE)   | X       |     |         |        |

Use `?<output format>` to see all of a format's options, e.g. `?html_document`

## More Header Options

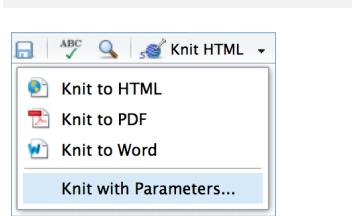
### PARAMETERS

Parameterize your documents to reuse with new inputs (e.g., data, values, etc.).

1. **Add parameters** in the header as sub-values of `params`.
2. **Call parameters** in code using `params$<name>`.
3. **Set parameters** with Knit with Parameters or the `params` argument of `render()`.

### REUSABLE TEMPLATES

1. **Create a new package** with a `inst/rmarkdown/templates` directory.
2. **Add a folder** containing `template.yaml` (below) and `skeleton.Rmd` (template contents).
3. **Install** the package to access template by going to **File > New R Markdown > From Template**.



### BOOTSWATCH THEMES

Customize HTML documents with Bootswatch themes from the `bslib` package using the theme output option.

Use `bslib::bootswatch_themes()` to list available themes.



```
---
```

```
title: "Document Title"
author: "Author Name"
output:
  html_document:
    theme:
      bootswatch: solar
---
```

### CUSTOM THEMES

Customize individual HTML elements using `bslib` variables. Use `?bs_theme` to see more variables.

```
---
```

```
output:
  html_document:
    theme:
      bg: "#121212"
      fg: "#E4E4E4"
      base_font:
        google: "Prompt"
---
```

More on `bslib` at [pkgs.rstudio.com/bslib/](http://pkgs.rstudio.com/bslib/).

### STYLING WITH CSS AND SCSS

Add CSS and SCSS to your document by adding a path to a file with the `css` option in the YAML header.

```
---
```

```
title: "My Document"
author: "Author Name"
output:
  html_document:
    css: "style.css"
---
```

Apply CSS styling by writing HTML tags directly or:

- Use markdown to apply style attributes inline.

Bracketed Span  
A [green]{.my-color} word.

A green word.

Fenced Div  
:::{.my-color}  
All of these words  
are green.  
:::

All of these words  
are green.

- Use the Visual Editor. Go to **Format > Div/Span** and add CSS styling directly with Edit Attributes.

.my-css-tag ...  
This is a div with some text in it.

## Render

When you render a document, rmarkdown:

1. Runs the code and embeds results and text into an .md file with knitr.
2. Converts the .md file into the output format with Pandoc.



**Save**, then **Knit** to preview the document output. The resulting HTML/PDF/MS Word/etc. document will be created and saved in the same directory as the .Rmd file.

Use `rmarkdown::render()` to render/knit in the R console. See `?render` for available options.

## Share

### Publish on RStudio Connect

to share R Markdown documents securely, schedule automatic updates, and interact with parameters in real time.

[rstudio.com/products/connect/](https://rstudio.com/products/connect/)



### INTERACTIVITY

Turn your report into an interactive Shiny document in 4 steps:

1. Add `runtime: shiny` to the YAML header.
2. Call Shiny input functions to embed input objects.
3. Call Shiny render functions to embed reactive output.
4. Render with `rmarkdown::run()` or click **Run Document** in RStudio IDE.

```
---
```

```
output: html_document
runtime: shiny
---
```

```
```{r, echo = FALSE}
numericInput("n",
  "How many cars?", 5)
renderTable({
  head(cars, input$n)
})
```

	speed	dist
1	4.00	2.00
2	4.00	10.00
3	7.00	4.00
4	7.00	22.00
5	8.00	16.00

Also see Shiny Prerendered for better performance.

[rmarkdown.rstudio.com/authoring\\_shiny\\_prerendered](https://rmarkdown.rstudio.com/authoring_shiny_prerendered)

Embed a complete app into your document with `shiny::shinyAppDir()`. More at [bookdown.org/yihui/rmarkdown/shiny-embedded.html](https://bookdown.org/yihui/rmarkdown/shiny-embedded.html).

# Add silhouettes with rphylopic :: CHEAT SHEET

## Install rphylopic

**rphylopic** allows you to add species' silhouettes from phylopic to ggplot2 or base plots:

CRAN version  
install.packages("rphylopic")

Development version  
install.packages("remotes")  
remotes::install\_github("sckott/rphylopic")  
library('rphylopic')

## uuid

Universally unique identifier (uuid) is a 128-bit number. It has 32 alphanumeric characters in the form of 8-4-4-4-12. Every silhouette has a uuid to uniquely identify it.

## Find silhouettes

### 1. Work with names.

- **name\_search**(text, options)[[1]]

Searches the uuid code based on common name or taxonomy of a species. The options can be namebankID, type, names, root, uri.

- **name\_get**(uuid, options)

Get information on a name using the uuid code. The options can be citationStart, html, namebankID, root.

- **name\_images**(uuid, options = 'credit')

Searches for different images for a taxonomic name.

- **name\_taxonomy**(uuid, options, as)

Returns taxonomic name based on uuid code. Options can be string, and as can be list, table, json.

- **name\_taxonomy\_many**(uuid, options, as)

Returns taxonomic names for two or more concatenated (c()) uuid codes.

- **name\_taxonomy\_sources**(uuid)

Gives information on the sources for a name's taxonomy given a uuid.

### 2. Work with uBio data

- **ubio\_get**(namebankID)

Retrieve the uuid code based on the namebankID number.

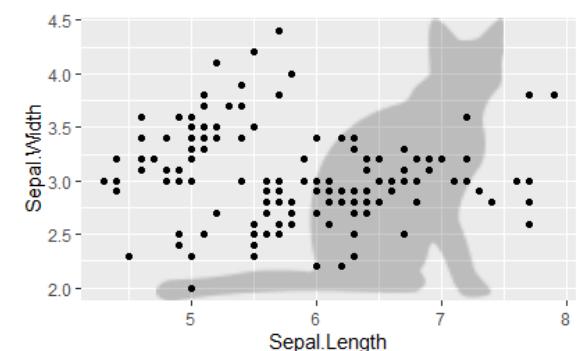
## Plot silhouettes

### 1. Plot a silhouette behind a plot

- **ggplot**

```
library(ggplot2)  
cat <- image_data("23cd6aa4-9587-4a2e-  
8e26-de42885004c9", size = 128)[[1]]
```

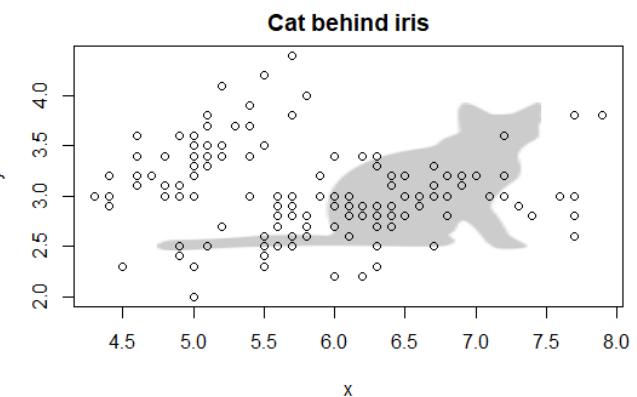
```
ggplot(data = iris,  
       aes(x = Sepal.Length,  
            y = Sepal.Width))+  
  geom_point() +  
  add_phylopic(cat, alpha = 0.2)
```



- **Base plot**

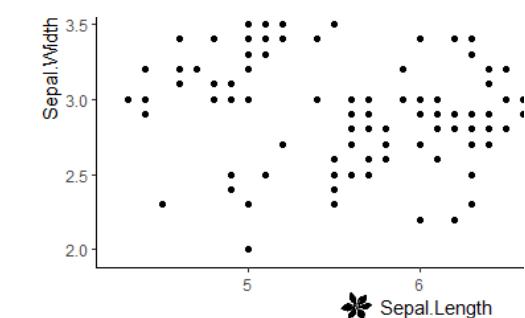
```
cat <- image_data("23cd6aa4-9587-4a2e-  
8e26-de42885004c9", size = 128)[[1]]
```

```
plot(1, 1,  
     type = 'n',  
     main = "Cat behind iris")  
add_phylopic_base(cat,  
                  x = 0.5,  
                  y = 0.5,  
                  ysize = 0.8,  
                  alpha = 0.2)
```



### 2. Plot a silhouette anywhere in a plot

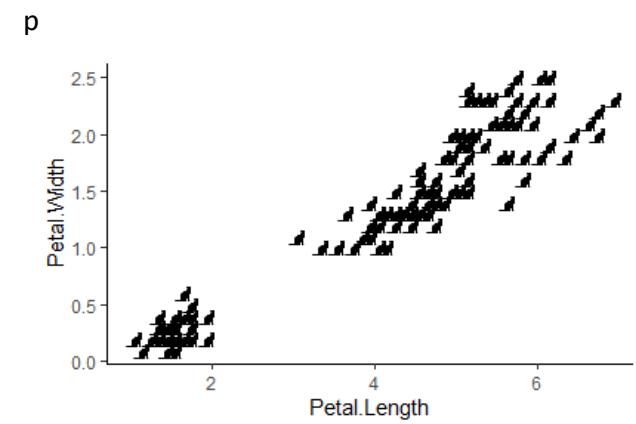
```
ggpubr::ggarrange(plot) +  
  add_phylopic(irisimg,  
              alpha = 1,  
              x = 0.43,  
              y = 0.05,  
              ysize = 0.06)
```



### 3. Plot silhouettes as points in a plot

- **ggplot2**

```
p <- ggplot(iris,  
            aes(Petal.Length,  
                 Petal.Width)) +  
  geom_blank() +  
  theme_classic()  
  
for (i in 1:nrow(iris)) {  
  p <- p +  
    add_phylopic(cat,  
                 alpha = 1,  
                 iris$Petal.Length[i],  
                 iris$Petal.Width[i],  
                 ysize = 0.2)  
}
```

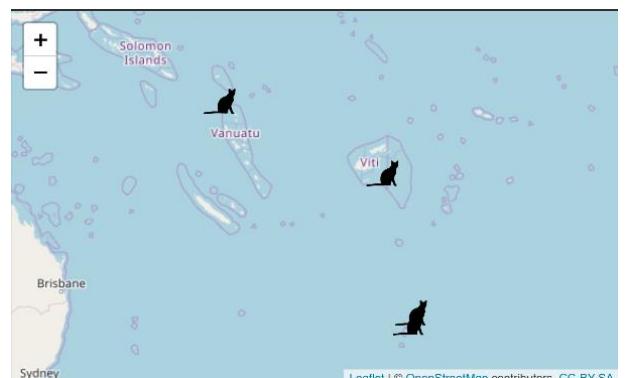


### 4. Save PNG file to disk

- Download a silhouette from <http://phylopic.org/> and save it in your working directory.  
img <- png::readPNG("img.png")

### 5. Use silhouettes as icons in leaflet plots

```
library(leaflet)  
data(quakes) ## this is a table  
# get an image  
cat <- image_data("23cd6aa4-9587-4a2e-  
8e26-de42885004c9", size = 128)[[1]]  
# save to disk  
catimg <- save_png(cat)  
# make an icon. See ?makeIcon for more  
# iconwidth is in pixels  
cat_icon <- makeIcon(iconurl = catimg,  
                      iconwidth = 30)  
# make the plot, just 7:10 rows  
leaflet(data = quakes[7:10,]) %>%  
  addTiles() %>%  
  addMarkers(~long, ~lat,  
            icon = cat_icon)
```



#### Citation

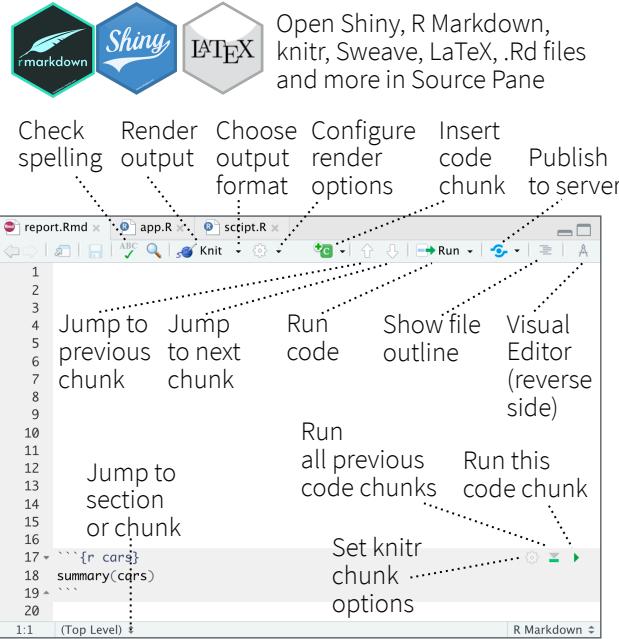
Don't forget to cite rphylopic. See how here:

```
citation("rphylopic")
```

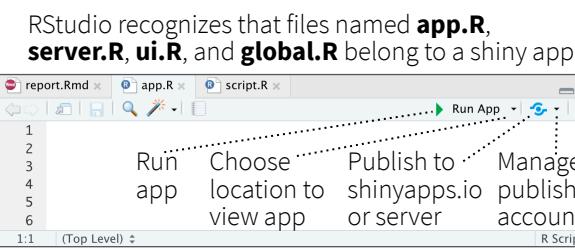
# RStudio IDE :: CHEATSHEET



## Documents and Apps



Access markdown guide at **Help > Markdown Quick Reference**  
See reverse side for more on **Visual Editor**



## Package Development

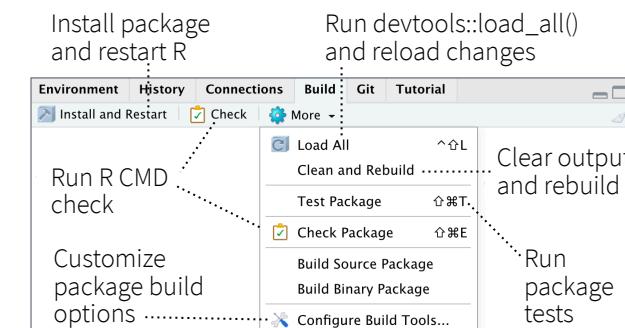


Create a new package with **File > New Project > New Directory > R Package**

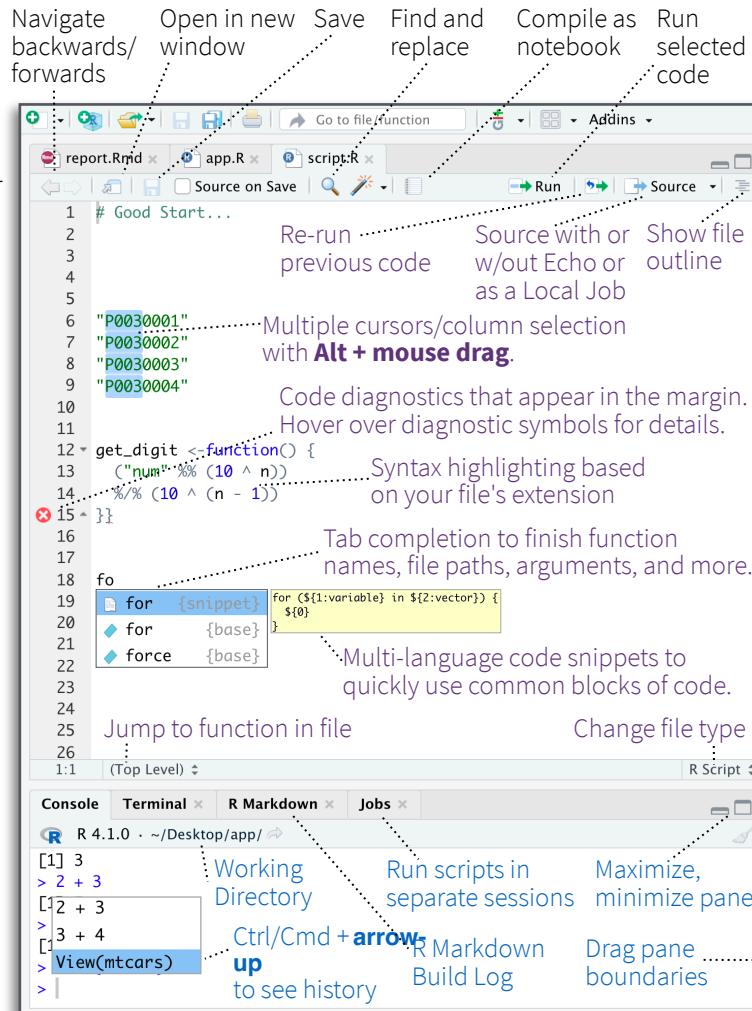
Enable roxygen documentation with **Tools > Project Options > Build Tools**

Roxygen guide at **Help > Roxygen Quick Reference**

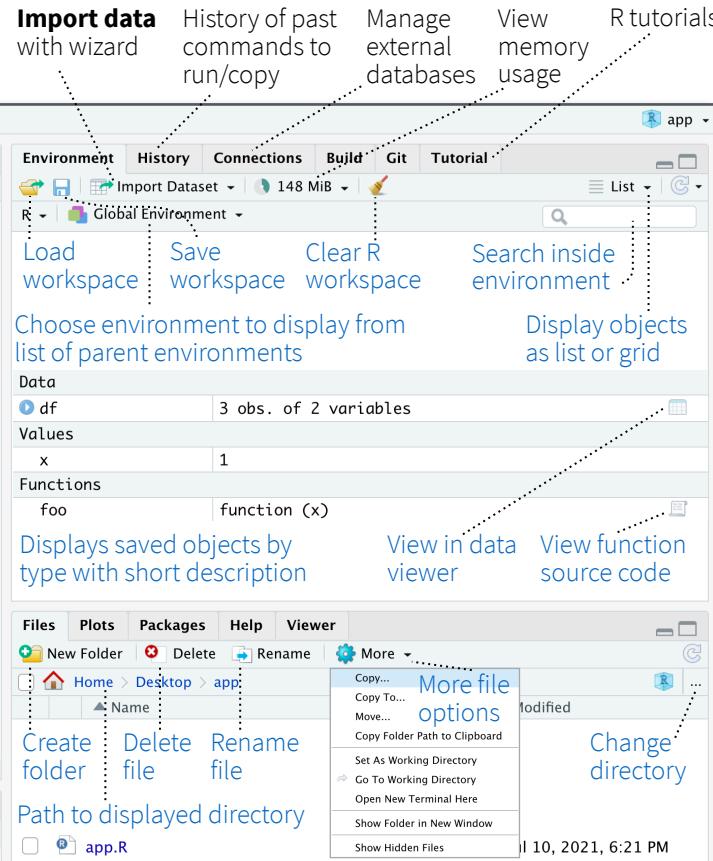
See package information in the **Build Tab**



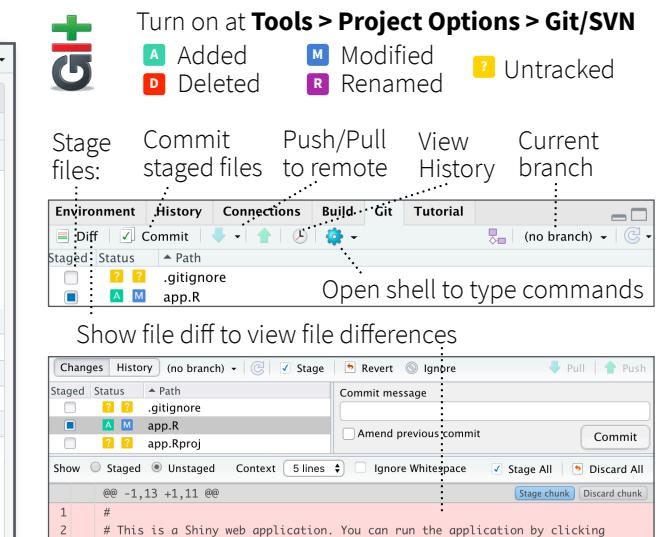
## Source Editor



## Tab Panes

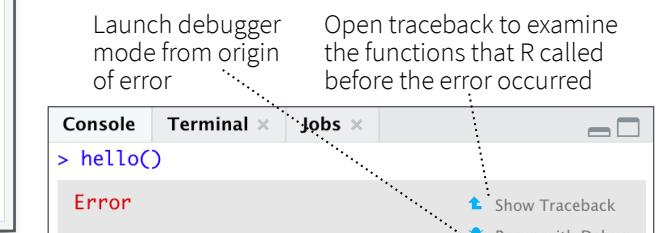


## Version Control



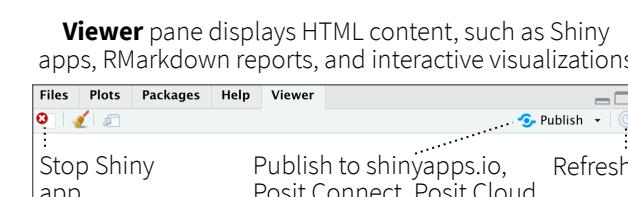
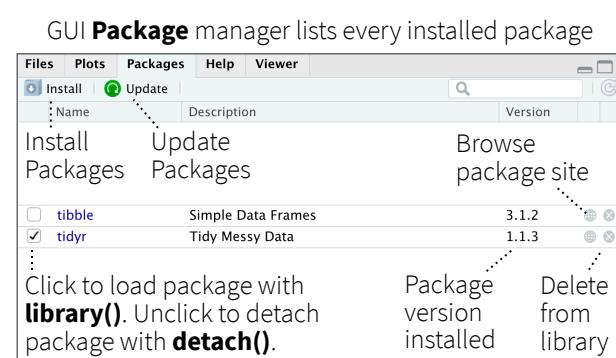
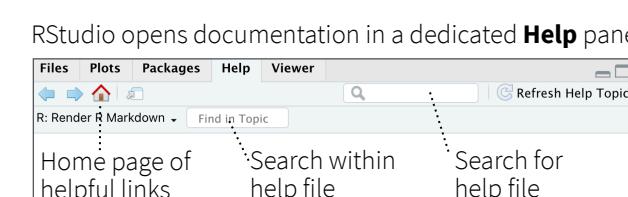
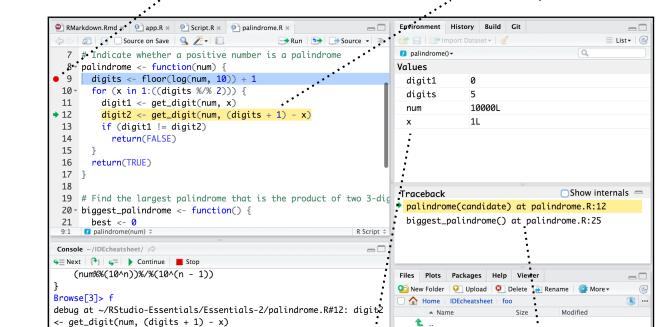
## Debug Mode

Use **debug()**, **browser()**, or a breakpoint and execute your code to open the debugger mode.



Click next to line number to add/remove a breakpoint.

Highlighted line shows where execution has paused





# Keyboard Shortcuts

## RUN CODE

Search command history  
Interrupt current command  
Clear console

	Windows/Linux	Mac
Ctrl+arrow-up	Ctrl+arrow-up	Cmd+arrow-up
Esc	Esc	Esc
Ctrl+L	Ctrl+L	Ctrl+L

## NAVIGATE CODE

Go to File/Function

Ctrl+.	Ctrl+.
--------	--------

## WRITE CODE

Attempt completion

Insert <- (assignment operator)  
Insert |> or %>% (pipe operator)  
(Un)Comment selection

	Windows/Linux	Mac
Alt+-	Option+-	Cmd+Shift+M
Ctrl+Shift+M	Ctrl+Shift+M	Cmd+Shift+C
Ctrl+Shift+C	Ctrl+Shift+C	Cmd+Shift+C

## MAKE PACKAGES

Load All (devtools)  
Test Package (Desktop)  
Document Package

	Windows/Linux	Mac
Ctrl+Shift+L	Ctrl+Shift+L	Cmd+Shift+L
Ctrl+Shift+T	Ctrl+Shift+T	Cmd+Shift+T
Ctrl+Shift+D	Ctrl+Shift+D	Cmd+Shift+D

## DOCUMENTS AND APPS

Knit/Render Document (knitr)  
Insert chunk (Sweave & Knitr)  
Run from start to current line

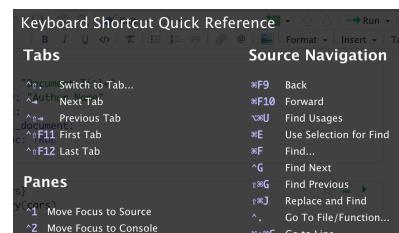
Windows/Linux	Mac
Ctrl+Shift+K	Cmd+Shift+K
Ctrl+Alt+I	Cmd+Option+I
Ctrl+Alt+B	Cmd+Option+B

## MORE KEYBOARD SHORTCUTS

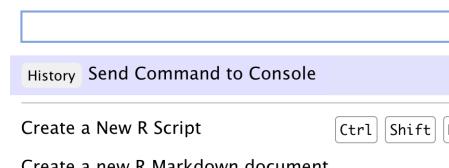
Keyboard Shortcuts Help  
Show Command Palette

Alt+Shift+K	Option+Shift+K
Ctrl+Shift+P	Cmd+Shift+P

View the Keyboard Shortcut Quick Reference with **Tools > Keyboard Shortcuts** or **Alt/Option + Shift + K**



Search for keyboard shortcuts with **Tools > Show Command Palette** or **Ctrl/Cmd + Shift + P**.



# Visual Editor

## R Markdown

This is an R Markdown document. Markdown is a simple formatting syntax for authoring HTML, PDF, and MS Word documents.

```
{r cars}
summary(cars)
```

Jump to chunk or header

# Posit Workbench

## WHY POSIT WORKBENCH?

Extend the open source server with a commercial license, support, and more:

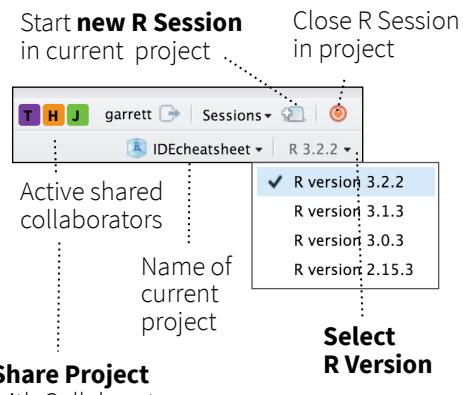
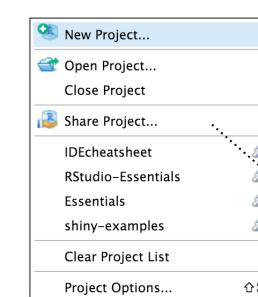
- open and run multiple R sessions at once
- tune your resources to improve performance
- administrative tools for managing user sessions
- collaborate real-time with others in shared projects
- switch easily from one version of R to a different version
- integrate with your authentication, authorization, and audit practices
- work in the RStudio IDE, JupyterLab, Jupyter Notebooks, or VS Code

Download a free 45 day evaluation at [posit.co/products/enterprise/workbench/](https://posit.co/products/enterprise/workbench/)

# Share Projects

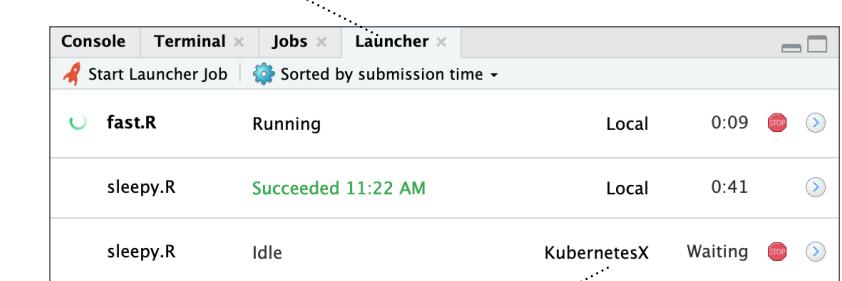
## File > New Project

RStudio saves the call history, workspace, and working directory associated with a project. It reloads each when you re-open a project.



# Run Remote Jobs

Run R on remote clusters (Kubernetes/Slurm) via the Job Launcher



Run launcher jobs remotely

# SamplingStrata: : CHEAT SHEET

## Optimal stratification

Given a sampling frame, SamplingStrata allows to optimize its stratification when designing a sampling survey, given precision constraints on target estimates.

### Three different methods

The optimization can be run by indicating three different methods, on the basis of the following:

- A. if stratification variables are categorical (or reduced to) then the method is the "**atomic**";
- B. if stratification variables are continuous, then the method is the "**continuous**";
- C. if stratification variables are continuous, and there is spatial correlation among units in the sampling frame, then the required method is the "**spatial**".

### A. Method "atomic"

Different steps:

1. define the sampling frame;
2. set precision constraints;
3. build atomic strata;
4. run optimization;
5. perform evaluation;
6. select the sample.

#### Sampling frame

```
library(SamplingStrata)
data("swissmunicipalities")
swissmunicipalities$id <- c(1:nrow(swissmunicipalities))
frame <- buildFrameDF(
  df = swissmunicipalities,
  id = "id",
  domainvalue = "REG",
  X = c("POPTOT", "HApoly"),
  Y = c("Surfacesbois", "Airind"))
```

Data on 2896 Swiss municipalities

Stratification variables

Target variables

#### Precision constraints

```
ndom <- length(unique(frame$domainvalue))
cv <- as.data.frame(list(
  DOM = rep("DOM1", ndom),
  CV1 = rep(0.10, ndom),
  CV2 = rep(0.10, ndom),
  domainvalue = c(1:ndom)))
```

10% of maximum expected CV

#### Atomic strata

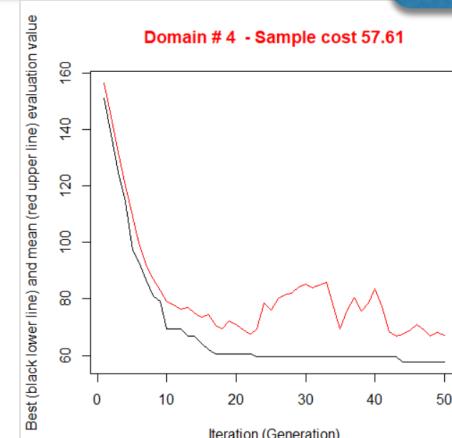
```
strata <- buildStrataDF(frame)
```

#### Optimization

```
solution <-
optimStrata(method="atomic",
            framesamp = frame,
            errors = cv,
            iter = 50,
            pops = 10)
```

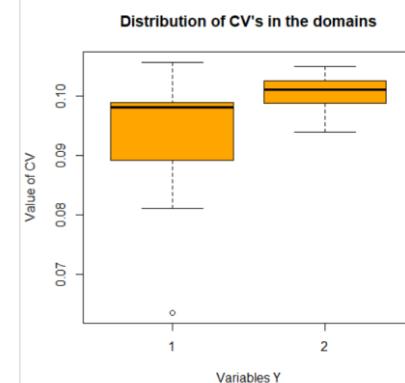
Number of iterations

Number of solutions per iteration



#### Evaluation

```
outstrata <- solution$aggr_strata
framenew <- solution$framenew
eval <- evalSolution(framenew, outstrata)
eval$coeff_var
```



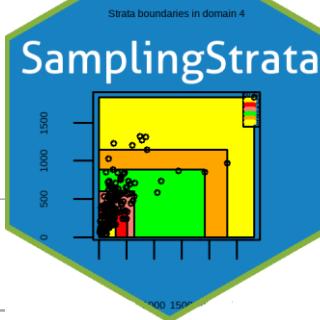
#### Sample selection

```
s <- selectSample(framenew, outstrata)
head(s)
```

DOMAINVALUE	STRATO	ID	X1	X2	Y1	Y2	LABEL	WEIGHTS
1	1	2398	241	294	101	0	1	21.38462
2	1	2331	267	449	215	1	1	21.38462
3	1	2410	237	935	471	0	1	21.38462
4	1	2112	370	330	98	0	1	21.38462
5	1	2563	173	178	16	0	1	21.38462
6	1	2091	382	594	338	0	1	21.38462

To install last available release:

```
library(devtools)
install_github("barcaroli/SamplingStrata")
```



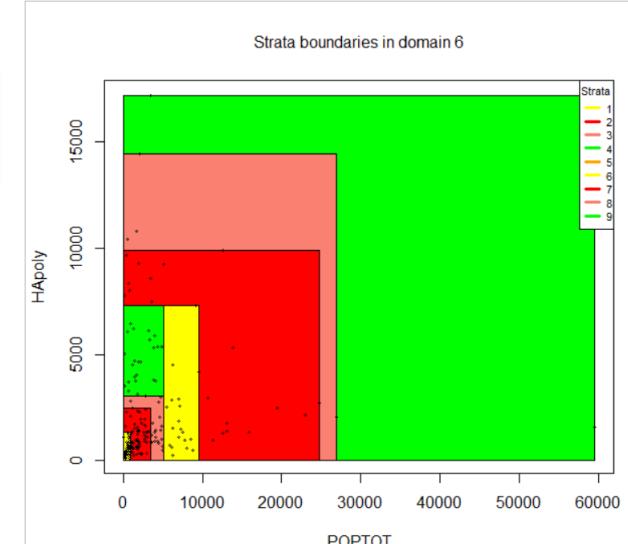
## Evaluation

```
framenew <- solution$framenew
outstrata <- solution$aggr_strata
ss <- summaryStrata(framenew, outstrata)
head(ss)
```

Domain	Stratum	Population	Allocation	SamplingRate
1	1	278	13	0.047370
2	1	49	6	0.113769
3	1	70	5	0.070377
4	1	69	10	0.145075
5	1	33	6	0.173466
6	1	22	9	0.424133

Lower_X1	Upper_X1	Lower_X2	Upper_X2	
1	27	780	32	986
2	65	1422	48	1018
3	95	1562	198	2288
4	78	1963	159	11907
5	1992	2711	107	8925
6	2759	3567	185	11378

```
plotStrata2d(framenew,
              outstrata,
              domain = 6,
              vars = c("X1", "X2"),
              labels = c("POPTOT", "HApoly"))
```



```
eval <-
evalSolution(framenew, outstrata)
eval$coeff_var
```

## Sample selection

```
s <- selectSample(framenew, outstrata)
head(s)
```

DOMAINVALUE	STRATO	ID	X1	X2	Y1	Y2	LABEL	WEIGHTS
1	1	2398	241	294	101	0	1	21.38462
2	1	2331	267	449	215	1	1	21.38462
3	1	2410	237	935	471	0	1	21.38462
4	1	2112	370	330	98	0	1	21.38462
5	1	2563	173	178	16	0	1	21.38462
6	1	2091	382	594	338	0	1	21.38462

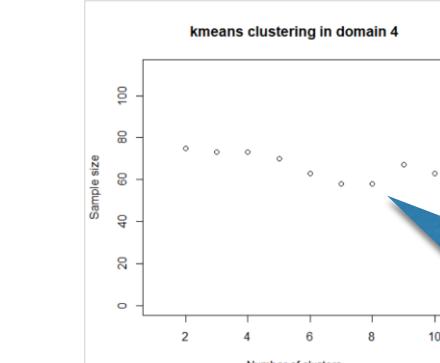
## B. Method "continuous"

Same steps with the exception of strata building, not necessary.

Frame definition and precision constraints settings are done in the same way than in method "atomic". One more step is in determination of the most promising number of strata with kmeans clustering.

#### Kmeans clustering

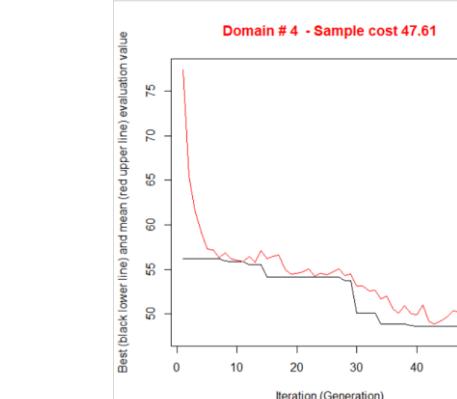
```
kmean <- KmeansSolution2(frame=frame,
                           errors=cv,
                           maxclusters = 10)
nstrat <- tapply(kmean$suggestions,
                  kmean$domainvalue,
                  FUN=function(x)
                    length(unique(x)))
sugg <- prepareSuggestion(
  kmean = kmean,
  frame = frame,
  nstrat = nstrat)
```



Visualization of strata by couples of X's  
Suggested number of strata (8) for domain 4

#### Optimization

```
solution <- optimStrata (
  method = "continuous",
  framesamp = frame,
  errors = cv,
  nStrata = nstrat,
  iter = 50,
  pops = 10,
  suggestions = sugg)
```



## C. Method "spatial"

In cases where units in the sampling frame are geo-referenced and there is spatial correlation among them, it is possible to apply the "spatial" method in the optimization of the frame stratification.

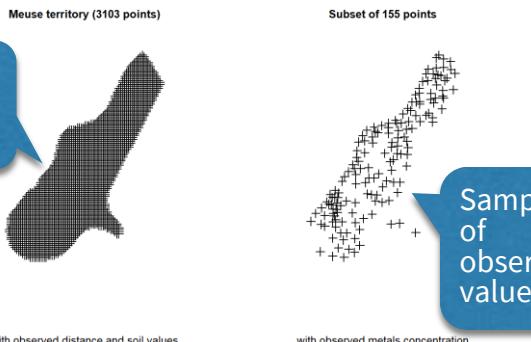
Different steps:

1. perform a preliminary spatial analysis and fit spatial models on target variables
2. define the sampling frame and add predicted values, prediction errors and coordinates;
3. set precision constraints;
4. run optimization;
5. select the sample.

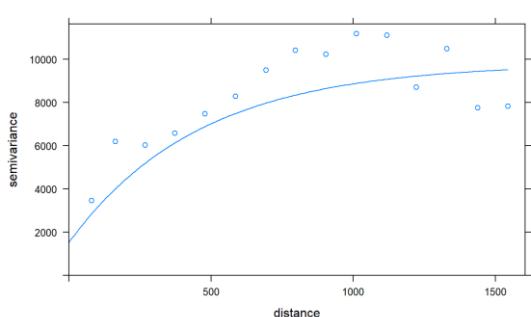
### Spatial analysis

We make use of the «Meuse river» datasets, reporting measures of 4 metals concentration.

```
library(sp)
# locations (155 observed points)
data("meuse")
# grid of points (3103)
data("meuse.grid")
meuse.grid$id <- c(1:nrow(meuse.grid))
coordinates(meuse)<-c('x','y')
coordinates(meuse.grid)<-c('x','y')
```



```
library(gstat)
library(automap)
v <- variogram(lead~dist+soil,data=meuse)
fit.vgm.lead <- autofitVariogram(
  lead ~dist+soil,meuse,model="Exp")
plot(v, fit.vgm.lead$var_model)
```



```
prediction
lead.kr <- krige(lead~dist+soil,
  meuse, meuse.grid,
  model=fit.vgm.lead$var_model)
lead.pred <- ifelse(lead.kr[1]$var1.pred<0,
  0,lead.kr[1]$var1.pred)
lead.var <- ifelse(lead.kr[2]$var1.var < 0,
  0,lead.kr[2]$var1.var)
```

### Sampling frame

```
df <- as.data.frame(list(
  dom=rep(1,nrow(meuse.grid)),
  lead.pred=lead.pred,
  lead.var=lead.var,
  lon=meuse.grid$x,
  lat=meuse.grid$y,
  id=c(1:nrow(meuse.grid))))
frame <- buildFrameSpatial(df=df,
  id="id",
  X=c("lead.pred"),
  Y=c("lead.pred"),
  variance=c ("lead.var"),
  lon="lon",
  lat="lat",
  domainvalue = "dom")
```

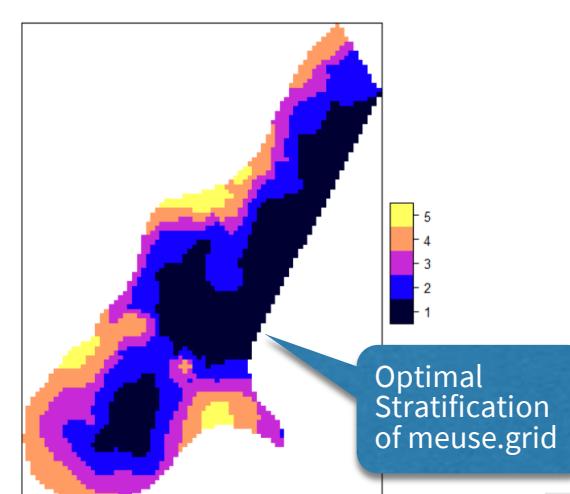
### Precision constraints

```
cv2 <- as.data.frame(list(
  DOM=rep("DOM1",1),
  CV1=rep(0.05,1),
  domainvalue=c(1:1) ))
```

### Optimization

```
solution <- optimStrata(method="spatial",
  errors=cv2, framesamp=frame, iter=25,
  nStrata=5, fitting=1, kappa=1,
  range=fit.vgm.lead$var_model$range[2])
```

```
framenew <- solution$framenew
outstrata <- solution$aggr_strata
frameres <- SpatialPixelsDataFrame(
  points=framenew[c("LON","LAT")],
  data=framenew)
frameres$LABEL <-
  as.factor(frameres$LABEL)
spplot(frameres,c("LABEL"),
  col.regions=bpy.colors(5))
```



## Use of models

Usually, values of target variables are not available in sampling frames, but only of co-variates. In order to calculate correctly the variance of target variables in strata, we can make use of models. When applying methods 'atomic' and 'continuous', it is possible to declare linear or log-linear models linking each target variable to one co-variate available in the sampling frame.

Consider the case with 'swissmunicipalities' dataset. Suppose that for all units we only have values for POPTOT and HApolY, while only on a subset (500) of it the values for Surfacesbois and Airbat are also available.

We fit the following models:

```
k <- sample(c(1:2896),500)
s <- swissmunicipalities[k,]
Airind_POPTOT <-
  lm(Airind~POPTOT, data=s)
Bois_HApoly <-
  lm(Surfacesbois~HApoly,data=s)
```

For both models we calculate heteroscedasticity indexes and variance:

```
airind <-
  computeGamma(Airind_POPTOT$residuals,
  s$POPTOT,nbins = 14)
airind
# gamma sigma r.square
# 0.59235109 0.06794055 0.87070106
bois <-
  computeGamma(Bois_HApoly$residuals,
  s$HApoly,nbins = 14)
bois
# gamma sigma r.square
# 0.8547931 0.4483606 0.9732122 )
```

We can now instantiate the values in the 'model' dataframe:

```
model <- NULL
model$beta[1] <-
  Airind_POPTOT$coefficients[2]
model$sig2[1] <- airind[2]^2
model$type[1] <- "linear"
model$gamma[1] <- airind[1]
model$beta[2] <-
  Bois_HApoly$coefficients[2]
model$sig2[2] <- bois[2]^2
model$type[2] <- "linear"
model$gamma[2] <- bois[1]
model <- as.data.frame(model)
model
# beta sig2 type gamma
# 0.01109583 0.1708807 linear 0.4703953
# 0.26068155 0.2010272 linear 0.8547931
```

## Sampling frame

```
frame <- buildFrameDF(
  df=swissmunicipalities,
  id="id",
  X=c("POPTOT", "HApoly"),
  Y=c("POPTOT", "HApoly"),
  domainvalue = "REG")
```

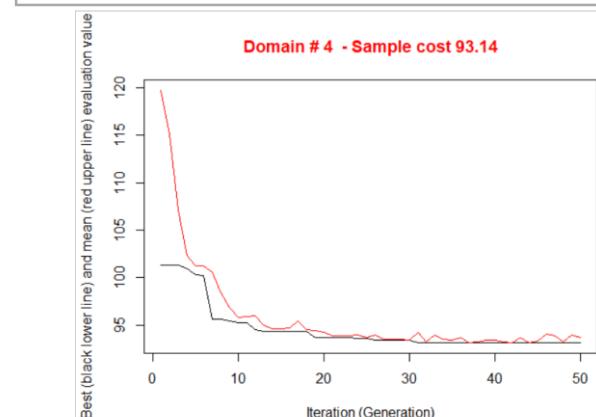
```
frame$airind <-
  swissmunicipalities$Airind
frame$surfacesbois <-
  swissmunicipalities$Surfacesbois
```

### Optimization

With the same precision constraints of 10% for both target variables we run the optimization step:

```
solution <-
  optimStrata(
    method = "continuous",
    errors = cv,
    framesamp = frame,
    model = model,
    nStrata = rep(5,7),
    iter = 50,
    pops = 10)
```

'model' dataframe previously defined



### Evaluation

```
framenew <- solution$framenew
outstrata <- solution$aggr_strata
framenew$Y3 <- framenew$AIRIND
framenew$Y4 <- framenew$SURFACESBOIS
val <- evalSolution(framenew,outstrata)
val$coeff_var
# CV1 CV2 CV3 CV4 dom
# 0.0107 0.0706 0.0316 0.0603 DOM1
# 0.0073 0.0364 0.0220 0.0426 DOM2
# 0.0062 0.0252 0.0253 0.0332 DOM3
# 0.0071 0.0328 0.0303 0.0572 DOM4
# 0.0055 0.0646 0.0171 0.0541 DOM5
# 0.0037 0.0745 0.0173 0.0606 DOM6
# 0.0036 0.0753 0.0145 0.0541 DOM7
```

Notice that both the CV's of the co-variates (CV1 and CV2) and the CV's of the real target variables (CV3 and CV4) are compliant to the 10% precision constraints.

# SAS <-> R :: CHEAT SHEET

## Introduction

This guide aims to familiarise SAS users with R.  
R examples make use of tidyverse collection of packages.

Install tidyverse: `install.packages("tidyverse")`  
Attach tidyverse packages for use: `library(tidyverse)`

R data here in 'data frames', and occasionally vectors (via `c()`)  
Other R structures (lists, matrices...) are not explored here.

Keyboard shortcuts: `<-` `Alt + -` `%>%` `Ctrl + Shift + m`

## Datasets; drop, keep & rename variables

```
data new_data;
set old_data;
run;
```

```
new_data <- old_data
```

```
data new_data (keep=id);
set old_data (drop=job_title);
run;
```

```
new_data <- old_data %>%
select(-job_title) %>%
select(id)
```

```
data new_data (drop= temp: );
set old_data;
run;
```

```
new_data <- old_data %>%
select( -starts_with("temp"))
C.f. contains(), ends_with()
```

```
data new_data;
set old_data;
rename old_name = new_name;
run;
```

```
new_data <- old_data %>%
rename(new_name = old_name)
```

Note order differs

## Conditional filtering

```
data new_data;
set old_data;
if Sex = "M";
run;
```

```
new_data <- old_data %>%
filter(Sex == "M")
```

```
data new_data;
set old_data;
if year in (2010,2011,2012);
run;
```

```
new_data <- old_data %>%
filter(year %in% c(2010,2011,2012))
```

```
data new_data;
set old_data;
by id;
if first.id;
run;
```

```
new_data <- old_data %>%
group_by( id ) %>%
slice(1)
```

Could use slice(n()) for last

```
data new_data;
set old_data;
if dob > "25APR1990'd";
run;
```

```
new_data <- old_data %>%
filter(dob > as.Date("1990-04-25"))
```

## New variables, conditional editing

```
data new_data;
set old_data;
total_income = wages + benefits ;
run;
```

```
new_data <- old_data %>%
mutate(total_income = wages + benefits)
```

```
data new_data;
set old_data;
if hours > 30 then full_time = "Y";
else full_time = "N";
run;
```

```
new_data <- old_data %>%
mutate(full_time = if_else(hours > 30 , "Y" , "N"))
```

```
data new_data;
set old_data;
if temp > 20 then weather = "Warm";
else if temp > 10 then weather = "Mild";
else weather = "Cold";
run;
```

```
new_data <- old_data %>%
mutate(weather = case_when(
temp > 20 ~ "Warm",
temp > 10 ~ "Mild",
TRUE ~ "Cold" ))
```

## Counting and Summarising

```
proc freq data = old_data ;
table job_type ;
run;
```

```
old_data %>%
count(job_type)
```

For percent, add:  
%>% mutate(percent = n\*100/sum(n))

```
proc freq data = old_data ;
table job_type*region ;
run;
```

```
old_data %>%
count(job_type , region )
```

```
proc summary data = old_data nway ;
class job_type region ;
output out = new_data ;
run;
```

```
new_data <- old_data %>%
group_by( job_type , region ) %>%
summarise( Count = n() )
```

Equivalent without nway not trivially produced

```
proc summary data = old_data nway ;
class job_type region ;
var salary ;
output out = new_data
sum( salary ) = total_salaries ;
run;
```

```
new_data <- old_data %>%
group_by( job_type , region ) %>%
summarise( total_salaries = sum( salary ) ,
Count = n() )
```

Lots of summary functions in both languages

Swap summarise() for mutate() to add summary data to original data

## Combining datasets

```
data new_data ;
set data_1 data_2 ;
run;
```

```
new_data <- bind_rows( data_1 , data_2 )
```

C.f. rbind() which produces error if columns are not identical

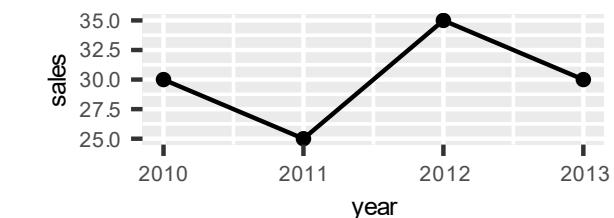
```
data new_data ;
merge data_1 (in= in_1) data_2 ;
by id ;
if in_1 ;
run;
```

```
new_data <- left_join( data_1 , data_2 , by = "id")
```

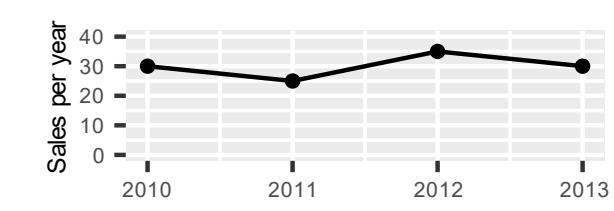
C.f. full\_join(), right\_join(), inner\_join()

## Some plotting in R

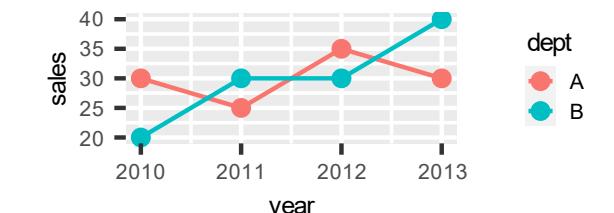
```
ggplot( my_data , aes( year , sales ) ) +
geom_point() + geom_line()
```



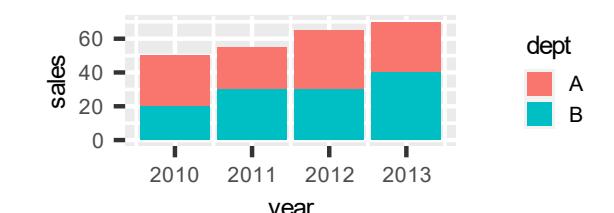
```
ggplot( my_data , aes( year , sales ) ) +
geom_point() + geom_line() + ylim(0, 40) +
labs(x = "" , y = "Sales per year")
```



```
ggplot(my_data, aes( year, sales, colour = dept ) ) +
geom_point() + geom_line()
```

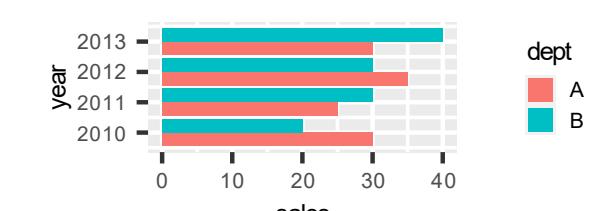


```
ggplot( my_data , aes( year, sales, fill = dept ) ) +
geom_col()
```



Note 'colour' for lines & points, 'fill' for shapes

```
ggplot( my_data , aes( year, sales, fill = dept ) ) +
geom_col( position = "dodge" ) + coord_flip()
```



C.f. position = "fill" for 100% stacked bars/cols

## Sorting and Row-Wise Operations

```

proc sort data=old_data out=new_data;
  by id descending income ;
run;

proc sort data=old_data nodup;
  by id job_type;
run;

Note nodup relies on adjacency of duplicate rows, distinct( ) does not

proc sort data=old_data nodupkey;
  by id ;
run;

data new_data;
  set old_data;
  by id descending income ;
  if first.id ;
run;

data new_data;
  set old_data;
  prev_id= lag( id );
run;

C.f. lead( ) for subsequent rows

data new_data;
  set old_data;
  by id;
  counter +1;
  if first.id then counter = 1;
run;

```

## Converting and Rounding

```

data new_data;
  set old_data ;
  num_var = input("5" , 8. );
  text_var = put( 5 , 8. );
run;

data new_data ;
  set old_data;
  nearest_5 = round( x , 5 )
  two_decimals = round( x , 0.01)
run;

```

## Creating functions to modify datasets

```

%macro add_variable(dataset_name);
data &dataset_name;
  set &dataset_name;
  new_variable = 1;
run;
%mend;
%add_variable( my_data );

add_variable <- function( dataset_name ){
  dataset_name <- dataset_name %>%
    mutate(new_variable = 1)
  return( dataset_name )
}

my_data <- add_variable( my_data )

Note SAS can modify within the macro,
whereas R creates a copy within the function

```

## String Manipulation

```

data new_data;
  set old_data;
  if find(job_title , "Health" );
run;

data new_data;
  set old_data;
  substring = substr( big_string , 3 , 4 );
run;

data new_data;
  set old_data;
  address = tranwrd( address , "Street" , "St" );
run;

data new_data;
  set old_data;
  house_number = compress( address , , "dk" );
run;

```

*Returns characters 3 to 6. Note SAS uses <start>, <length>, R uses <start>, <end>*

*C.f. str\_replace( ) for first instance of pattern only*

*Wide range of regexps in both languages, this example extracts digits only*

## Transpose/Pivot

```

proc transpose data=long_data out=wide_data;
  by student <-
  id subject ;
  var grade ;
run;

Add NOTSORTED if long_data
is not sorted by student

proc transpose data=wide_data
  out=long_data(rename=(col1=grade)) name=subject;
  by student ;
  var English Irish Maths;
run;

```

*wide\_data <- long\_data %>%  
pivot\_wider(names\_from = subject , values\_from = grade)*

*long\_data <- wide\_data %>%  
pivot\_longer(c(English, Irish, Maths) ,  
names\_to = "subject", values\_to = "grade")*

## File operations

Operate in 'Work' library.  
Use libname to define file locations

Operate in a particular 'working directory' (identify using getwd( ))  
Move to other locations using setwd()

```

libname library_name "file_location";
data library_name.saved_data;
  set data_in_use;
run;

libname library_name "file_location";
data data_in_use ;
  set library_name.saved_data ;
run;

proc export data = my_data
  outfile = "my_file.csv" dbms = csv replace;
run;

proc import datafile = "my_file.csv"
  out = my_data dbms = csv;
run;

```

*saveRDS(data\_in\_use , file= "file\_location/saved\_data.rds")  
or  
setwd("file\_location")  
saveRDS( data\_in\_use , file = "saved\_data.rds")*

*data\_in\_use <- readRDS("file\_location/saved\_data.rds" )  
or  
setwd("file\_location")  
data\_in\_use <- readRDS("saved\_data.rds")*

*write\_csv(my\_data , "my\_file.csv")*

*my\_data <- read\_csv("my\_file.csv")*

*Both examples assume column headers in csv file*

# SAS Vs R in Pharma: : CHEAT

## Introduction

This cheat sheet mainly focus on data manipulation techniques frequently used in pharmaceutical industry. Run the below codes while starting R.

```
install.packages("tidyverse", "lubridate", "flextable","officer")
library(tidyverse,lubridate,flextable,officer)
```

## Data Inputs

```
data DM;
infile datalines delimiter=';';
input subjid $ strata sex $ armcd $;
age height weight ;
datalines;
101, 1 , M, B, 43, 150, 75
102, 2 , F, A, 53, 178, 65
103, 2 , F, B, 67, 157, 64
104, 1 , M, A, 34, 168, 72
105, 2 , M, B, 76, 145, 61
;
run;

data VS;
infile datalines delimiter=';';
input subjid $ strata armcd $ visit $;
visitnum paramcd $ aval ;
datalines;
101, 1, B, visit 1, 100, SYSBP, 120
101, 1, B, visit 19, 1900, SYSBP, 128
101, 1, B, visit 1, 100, DIABP, 65
101, 1, B, visit 19, 1900, DIABP, 78
102, 2, A, visit 1, 100, SYSBP, 156
102, 2, A, visit 19, 1900, SYSBP, 127
102, 2, A, visit 1, 100, DIABP, 74
102, 2, A, visit 19, 1900, DIABP, 72
105, 2, B, visit 1, 100, SYSBP, 136
105, 2, B, visit 19, 1900, SYSBP, 125
105, 2, B, visit 1, 100, DIABP, 59
105, 2, B, visit 19, 1900, DIABP, 64
;
run;

data EX;
infile datalines delimiter=';';
input subjid $ visitnum visit $ exstdtc $23-49 ;
datalines;
101, 100, visit 1, 2021-12-22T08:25
101, 1900, visit 19, 2021-12-29T08:55
104, 100, visit 1, 2021-12-16T11:02
104, 1900, visit 19, 2022-01-06T13:45
;
run;
```

```
subjid <- c('101','102','103', '104','105')
strata <- c(1,2,2,1,2)
sex <- c('M','F','F','M','M')
armcd <- c('B','A','B','A','B')
age <- c(43,53,67,34,76)
height <- c(150,178,157,168,145)
weight <- c(75,65,64,72,61)
DM <- data.frame(subjid,strata,sex,
                  armcd,age,height, weight)
View(DM)
```

```
subjid <- c('101','101','101','101',
           '102','102','102','102',
           '105','105','105','105')
strata <- c(1,1,1,2,2,2,2,2,2,2)
armcd <- c('B','B','B','B','A','A',
           'A','A','B','B','B','B')
visit <- c('Visit 1','Visit 19','Visit 1', 'Visit 19',
           'Visit 1','Visit 19','Visit 1', 'Visit 19',
           'Visit 1','Visit 19','Visit 1', 'Visit 19')
visitnum <- c(100,1900,100,1900,100,1900,
              100,1900,100,1900,100,1900)
paramcd <- c('SYSBP','SYSBP','DIABP','DIABP',
             'SYSBP','SYSBP','DIABP','DIABP',
             'SYSBP','SYSBP','DIABP','DIABP')
aval <- c(120,128,65,78,156,127,
          74,72,136,125,59,64)
VS <- data.frame(subjid,strata,armcd,visit,
                  visitnum,paramcd,aval)
View(VS)
```

```
subjid <- c('101','101','104','104')
Visitnum <- c(100,1900, 100,1900)
visit <- c('Visit 1','Visit 19', 'Visit 1', 'Visit 19')
exstdtc <- c("2021-12-22T08:25",
            "2021-12-29T08:55",
            "2021-12-16T11:02",
            "2022-01-06T13:45")
EX <- data.frame(subjid,visitnum,visit,exstdtc)
View(EX)
```

## Variable operation

### Variable Sorting

```
proc sort data=VS
  out=ADVS_SRT1;
  by subjid descending paramcd
    visitnum;
run;
```

```
ADVS_SRT1 <- VS %>%
  arrange(subjid,desc(paramcd),
         visitnum)
View(ADVS_SRT1)
```

### Data Filtering

```
data ADSL_FL1;
  set DM;
  if strata = 2;
run;
```

```
ADSL_FL <- DM %>%
  filter(strata==2 )
View(ADSL_FL)
```

```
data ADSL_FL2;
  set DM;
  if strata = 2 & armcd = 'A';
run;
```

```
ADSL_FL2 <- DM %>%
  filter(strata==2 & armcd=='A')
View(ADSL_FL2)
```

```
data ADSL_FL3;
  set DM;
  if subjid in ('101','102');
run;
```

```
ADSL_FL3 <- ADSL %>%
  filter(subjid %in% c('101', '102'))
View(ADSL_FL3)
```

### Data operations (keep, drop, and rename)

```
data ADSL_DO;
  set DM;
  keep subjid armcd ;
  drop age;
  rename subjid=usubjid;
run;
```

```
ADSL_DO <- DM %>%
  select(subjid,age,armcd)%>%
  select(-age) %>%
  rename(usubjid=subjid)
View(ADSL_DO)
```

### Variable creation

```
data ADSL_MT1;
  set DM;
  height_m= height/100;
  BMI=weight/(height_m**2);
run;
```

```
ADSL_MT1 <- DM %>%
  mutate(height_m=height/100) %>%
  mutate(BMI=weight/(height_m^2))
View(ADSL_MT1)
```

### Remove duplicate records

```
proc sort data=VS
  out=ADVS_SRT1 nodupkey;
  by subjid paramcd;
run;
```

```
ADVS_SRT1 <- VS %>%
  arrange( subjid , paramcd )%>%
  group_by ( subjid, paramcd ) %>%
  slice( 1 )
view(ADVS_SRT1)
```

## Data Transformation

### Data transpose (long to wide)

```
proc transpose data=VS
  out=ADVS_TR;
  by subjid strata armcd visit;
  id paramcd;
  var aval;
run;
```

\*Sort the dataset before transpose  
(wide to long)

```
proc transpose data=ADVS_TR
  out=ADVS_TR2;
  by subjid strata armcd visit;
  var SYSBP DIABP;
run;
```

\*Sort the dataset before transpose

### Data Appending

```
data ADVS_APD;
  set VS EX;
run;
```

```
ADVS_TR <-VS %>%
  pivot_wider(
    names_from=paramcd,
    values_from=aval)
view(ADVS_TR)
```

```
ADVS_TR2 <- ADVS_TR %>%
  pivot_longer(
    cols=SYSBP:DIABP,
    names_to='paramcd',
    values_to='aval')
view(ADVS_TR2)
```

### Data merging single-dataset

```
proc sql;
  create table ADVS_IJ as
  select distinct a.* , b.exstdtc
  from VS as a
  inner join EX as b
  on a.subjid = b.subjid and
  a.visitnum= b.visitnum;
quit;
```

```
ADVS_IJ <- VS %>%
  inner_join(EX,
             by = c("subjid","visitnum"))
view(ADVS_IJ)
```

### Multiple-dataset

```
proc sql;
  create table ADSL_FJ as
  select distinct a.* , b.visitnum,
  b.paramcd,b.aval,c.exstdtc
  from DM as a
  full join VS as b
  on a.subjid = b.subjid
  full join EX as c
  on a.subjid = c.subjid;
quit;
```

```
ADVS_FJ <- VS %>%
  full_join(EX,
            by = c("subjid","visitnum")) %>%
  full_join(DM, by = "subjid")
view(ADVS_FJ)
```

# SAS Vs R in Pharma: : CHEAT

## Character operation

### Variable conversion:

#### Numeric to character:

```
data ADVS_CHAR;
  set VS;
  avalc=put(aval,8.);
  view(ADVS_CHAR)
run;
*SAS has formats to handle digits
```

#### Character to numeric

```
data ADVS_NUM;
  set ADVS_CHAR;
  aval_num=input(avalc, 8.);
  View(ADVS_NUM)
run;
*SAS has various informats
```

#### String operations:

```
data ADSL_STR1;
  set VS;
  substring=substr(visit,7,2);
  scanstring=scan(visit,2);
run;
View(ADSL_STR1)
```

#### If and else if command

```
data ADSL_IF;
  set DM;
  length age_r $12;
  if age < 18
    then age_r= "<18";
  else if 18 <= age <=64
    then age_r = "18-64";
  else if age > 64
    then age_r = ">65";
run;
View(ADSL_IF)
```

#### Remove leading/trailing spaces and Concatenation

```
data ADVS_RB;
  set VS;
  group_t=strip(subjid)||"/"||strip(armcd)||"/"||strip(strata);
  view(ADVS_RB)
*See Date/time section for handling in-between spaces
```

## Plotting

```
proc sgplot data=DM;
  scatter x=height y=weight;
  xaxis values=
    (140 to 180 by 10);
  yaxis values=
    (50 to 80 by 10);
run;
```

```
proc sgplot data=ADSL_IF;
  vbar age_r;
run;
```

```
proc sgpanel data=VS;
  panelby paramcd subjid;
  scatter x=visit
    y=aval/group=armcd;
  series x=visit
    y=aval/group=armcd;
run;
```

```
ggplot(data=DM,
  aes(x=height, y=weight)) +
  geom_point()+
  lims(x=c(140,180),y=c(50,80)) +
  ggtitle("Height Vs. weight") +
  theme_classic()
```

```
ggplot(data=ADSL_IF,
  aes(x=age_r)) +
  geom_bar()+
  xlab("Age category")
theme_classic()
```

```
ggplot(data=VS,
  aes(x=visitnum,
    y=aval,colour=armcd))+geom_point()+
  geom_line()+
  facet_wrap(~ paramcd + subjid)+scale_x_continuous(
    labels=c("Visit1","Visit19"),
    breaks=c(100, 1900))
```

## Data Summary

### Summary

```
proc summary data=
  ADVS_SRT;
  by paramcd visitnum visit;
  var aval;
  output out=summary;
run;
*Sort the data with "by" variables before summarise
```

### frequency

```
proc freq data=DM;
  table armcd*strata
  / out=ADSL_FREQ;
run;
```

```
ADVS_SM <-VS %>%
group_by(paramcd,armcd,visit)%>%
summarise(mean=mean(aval),
sd= sd(aval),
min=min(aval),
max=max(aval),
n=length(aval))
View(ADVS_SM)
```

```
ADSL_FQ <- DM %>%
count(armcd,strata)
view(ADSL_FQ)
ADSL_FREQ <- ADSL_FQ %>%
mutate(percent=n/(sum(n)))
View(ADSL_FREQ)
```

## Date/time operations

```
data ADEX_DTM;
  set EX;
  format ADTM datetime18. ADT date9.
  ATM Time5.;
  ADTM = input(exstdtc, e8601DT.);
  ADT = datepart(ADTM);
  ATM = timepart(ADTM);
  visit=compress(visit);
run;
```

```
proc transpose data=ADEX_DTM
  out=ADEX_DTM1;
  by subjid;
  id visit;
  var ADT;
run;
```

```
data ADEX_DUR;
  set ADEX_DTM1;
  ADUR=visit19 - visit1;
run;
```

```
ADEX_DTM <- EX %>%
  mutate(ADTM=ymd_hm(exstdtc)) %>%
  mutate(ADT=date(ADTM)) %>%
  mutate(hours=hour(ADTM)) %>%
  mutate(mins=minute(ADTM)) %>%
  mutate(ATM=paste(hours,":",mins))
View(ADEX_DTM)
```

```
ADEX_DTM1 <- ADEX_DTM %>%
  mutate(visit_=str_replace_all(
    visit," ",""))
select(subjid,visit_,ADT) %>%
  spread(visit_,ADT)
```

```
ADEX_DTM2 <- ADEX_DTM1 %>%
  mutate(diff=difftime(
    Visit19,Visit1,unit='days'))%>%
  mutate(adur=as.numeric(word(diff,1))) %>%
  select(subjid,Visit1,Visit19,adur)
```

## Reporting

```
proc report data=ADVS_RB headline
split='#' spacing=0;
  columns (group_t paramcd visit aval)%>%
  define group_t/ 'subjid/Armcd/Strata'=group_t,
  "Parameter"=paramcd,
  "Visit"=visit,
  "Value"=aval)%>%
  regularable()%>%
  autofit();
run;
```

```
report<- ADVS_RB %>%
  select(group_t,paramcd,visit,aval)%>%
  rename("Usubjid/Armcd/Strata"=group_t,
  "Parameter"=paramcd,
  "Visit"=visit,
  "Value"=aval)%>%
  regularable()%>%
  autofit()
report <- merge_v(report)
report <- valign(report, valign="top")
```

## Data import and export

```
proc import datafile ="ADSL.csv"
  out = ADSL dbms= csv;
run;
```

```
read.csv(ADSL , "ADSL.csv")
```

```
proc export data = ADSL
  outfile = "ADSL.csv" dbms = csv replace;
run;
```

```
write.csv(ADSL , "ADSL.csv")
```

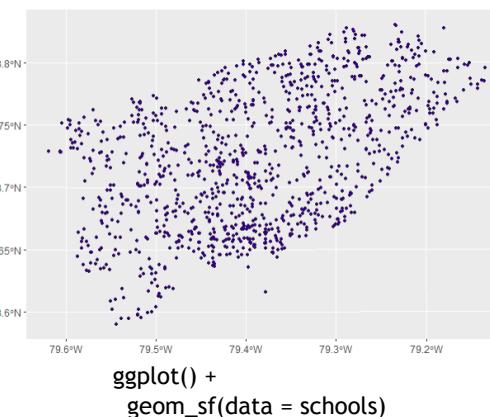
# Spatial manipulation with sf: : CHEAT SHEET



The sf package provides a set of tools for working with geospatial vectors, i.e. points, lines, polygons, etc.

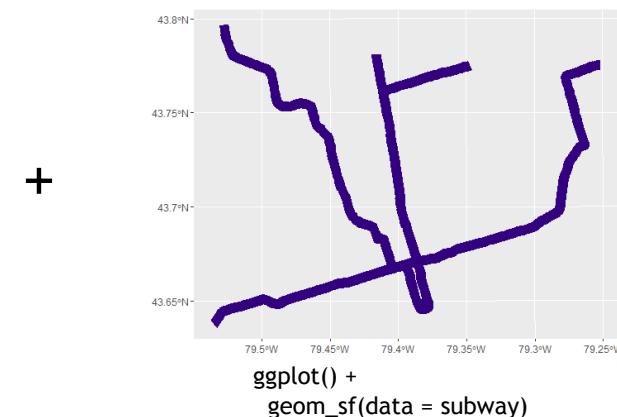
## Geometric confirmation

- st\_contains(x, y, ...) Identifies if y is within x (i.e. point within polygon)
- st\_covered\_by(x, y, ...) Identifies if x is completely within y (i.e. polygon completely within polygon)
- st\_covers(x, y, ...) Identifies if any point from x is outside of y (i.e. polygon outside polygon)
- st\_crosses(x, y, ...) Identifies if any geometry of x have commonalities with y
- st\_disjoint(x, y, ...) Identifies when geometries from x do not share space with y
- st\_equals(x, y, ...) Identifies if x and y share the same geometry
- st\_intersects(x, y, ...) Identifies if x and y geometry share any space
- st\_overlaps(x, y, ...) Identifies if geometries of x and y share space, are of the same dimension, but are not completely contained by each other
- st\_touches(x, y, ...) Identifies if geometries of x and y share a common point but their interiors do not intersect
- st\_within(x, y, ...) Identifies if x is in a specified distance to y



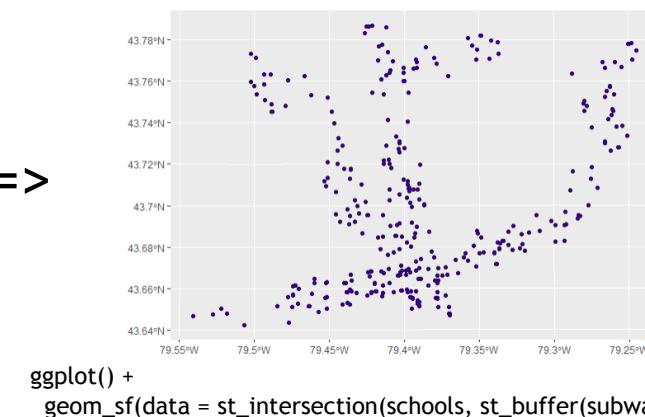
## Geometric operations

- st\_boundary(x) Creates a polygon that encompasses the full extent of the geometry
- st\_buffer(x, dist, nQuadSegs) Creates a polygon covering all points of the geometry within a given distance
- st\_centroid(x, ..., of\_largest\_polygon) Creates a point at the geometric centre of the geometry
- st\_convex\_hull(x) Creates geometry that represents the minimum convex geometry of x
- st\_line\_merge(x) Creates linestring geometry from sewing multi linestring geometry together
- st\_node(x) Creates nodes on overlapping geometry where nodes do not exist
- st\_point\_on\_surface(x) Creates a point that is guaranteed to fall on the surface of the geometry
- st\_polygonize(x) Creates polygon geometry from linestring geometry
- st\_segmentize(x, dfMaxLength, ...) Creates linestring geometry from x based on a specified length
- st\_simplify(x, preserveTopology, dTolerance) Creates a simplified version of the geometry based on a specified tolerance



## Geometry creation

- st\_triangulate(x, dTolerance, bOnlyEdges) Creates polygon geometry as triangles from point geometry
- st\_voronoi(x, envelope, dTolerance, bOnlyEdges) Creates polygon geometry covering the envelope of x, with x at the centre of the geometry
- st\_point(x, c(numeric vector), dim = "XYZ") Creating point geometry from numeric values
- st\_multipoint(x = matrix(numeric values in rows), dim = "XYZ") Creating multi point geometry from numeric values
- st\_linestring(x = matrix(numeric values in rows), dim = "XYZ") Creating linestring geometry from numeric values
- st\_multilinestring(x = list(numeric matrices in rows), dim = "XYZ") Creating multi linestring geometry from numeric values
- st\_polygon(x = list(numeric matrices in rows), dim = "XYZ") Creating polygon geometry from numeric values
- st\_multipolygon(x = list(numeric matrices in rows), dim = "XYZ") Creating multi polygon geometry from numeric values



# Spatial manipulation with sf: : CHEAT SHEET



The sf package provides a set of tools for working with geospatial vectors, i.e. points, lines, polygons, etc.

## Geometry operations

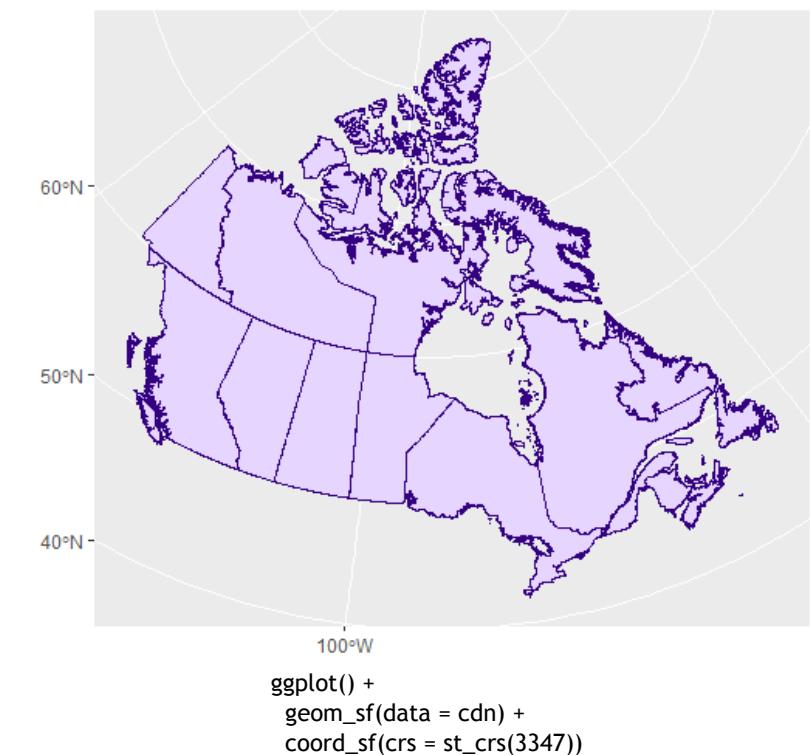
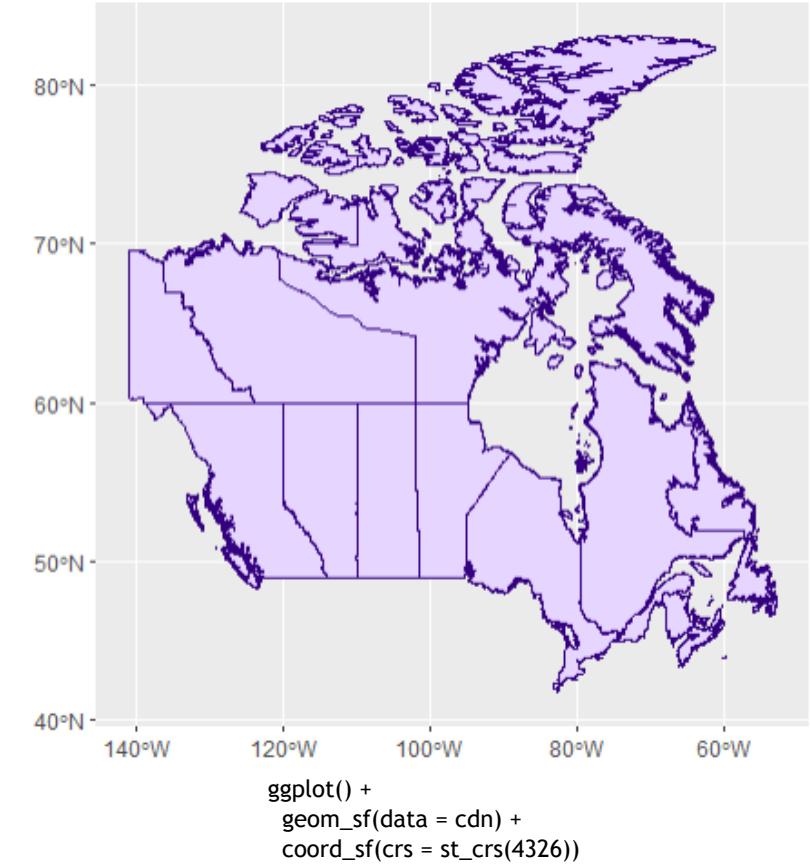
- ↳ `st_contains(x, y, ...)` Identifies if y is within x (i.e. point within polygon)
- ↳ `st_crop(x, y, ..., xmin, ymin, xmax, ymax)` Creates geometry of x that intersects a specified rectangle
- ↳ `st_difference(x, y)` Creates geometry from x that does not intersect with y
- ↳ `st_intersection(x, y)` Creates geometry of the shared portion of x and y
- ↳ `st_sym_difference(x, y)` Creates geometry representing portions of x and y that do not intersect
- ↳ `st_snap(x, y, tolerance)` Snap nodes from geometry x to geometry y
- ↳ `st_union(x, y, ..., by_feature)` Creates multiple geometries into a single geometry, consisting of all geometry elements

## Geometric measurement

- `st_area(x)` Calculate the surface area of a polygon geometry based on the current coordinate reference system
- `st_distance(x, y, ..., dist_fun, by_element, which)` Calculates the 2D distance between x and y based on the current coordinate system
- `st_length(x)` Calculates the 2D length of a geometry based on the current coordinate system

## Misc operations

- ↳ `st_as_sf(x, ...)` Create a sf object from a non-geospatial tabular data frame
- ↳ `st_cast(x, to, ...)` Change x geometry to a different geometry type
- ↳ `st_coordinates(x, ...)` Creates a matrix of coordinate values from x
- ↳ `st_crs(x, ...)` Identifies the coordinate reference system of x
- ↳ `st_join(x, y, join, FUN, suffix, ...)` Performs a spatial left or inner join between x and y
- ↳ `st_make_grid(x, cellsize, offset, n, crs, what)` Creates rectangular grid geometry over the bounding box of x
- ↳ `st_nearest_feature(x, y)` Creates an index of the closest feature between x and y
- ↳ `st_nearest_points(x, y, ...)` Returns the closest point between x and y
- ↳ `st_read(dsn, layer, ...)` Read file or database vector dataset as a sf object
- ↳ `st_transform(x, crs, ...)` Convert coordinates of x to a different coordinate reference system



# Shiny for R :: CHEATSHEET



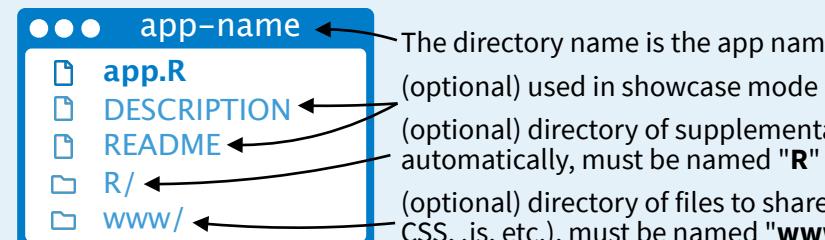
## Building an App

A **Shiny** app is a web page (**ui**) connected to a computer running a live R session (**server**).



Users can manipulate the UI, which will cause the server to update the UI's displays (by running R code).

Save your template as **app.R**. Keep your app in a directory along with optional extra files.



Launch apps stored in a directory with **runApp(<path to directory>)**.

## Share

Share your app in three ways:

1. **Host it on shinyapps.io**, a cloud based service from Posit. To deploy Shiny apps:

Create a free or professional account at [shinyapps.io](#)

Click the Publish icon in RStudio IDE, or run: `rsconnect::deployApp("<path to directory>")`

2. **Purchase Posit Connect**, a publishing platform for R and Python. [posit.co/products/enterprise/connect/](#)

3. **Build your own Shiny Server** [posit.co/products/open-source/shinyserver/](#)

To generate the template, type **shinyapp** and press **Tab** in the RStudio IDE or go to **File > New Project > New Directory > Shiny Application**

```
# app.R
library(shiny)

ui <- fluidPage(
  numericInput(inputId = "n",
    "Sample size", value = 25),
  plotOutput(outputId = "hist")
)

server <- function(input, output, session) {
  output$hist <- renderPlot({
    hist(rnorm(input$n))
  })
}

shinyApp(ui = ui, server = server)
```

**In ui** nest R functions to build an HTML interface

**Customize the UI with Layout Functions**

**Add Inputs with \*Input() functions**

**Add Outputs with \*Output() functions**

**Tell the server how to render outputs and respond to inputs with R**

**Wrap code in render\*() functions before saving to output**

**Refer to UI inputs with input\$<id> and outputs with output\$<id>**

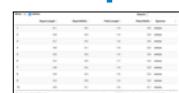
**Call shinyApp() to combine ui and server into an interactive app!**

See annotated examples of Shiny apps by running **runExample(<example name>)**. Run **runExample()** with no arguments for a list of example names.

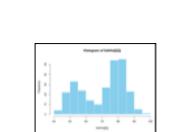
Sample size: 1000  
Histogram of rnorm(input\$n)  
Frequency vs. mnorm(input\$n)

## Outputs

**render\*()** and **\*Output()** functions work together to add R output to the UI.



**DT::renderDataTable(expr, options, searchDelay, callback, escape, env, quoted, outputArgs)**



**renderImage(expr, env, quoted, deleteFile, outputArgs)**

**renderPrint(expr, env, quoted, width, outputArgs)**

**renderTable(expr, striped, hover, bordered, spacing, width, align, rownames, colnames, digits, na, ..., env, quoted, outputArgs)**

**renderText(expr, env, quoted, outputArgs, sep)**

**renderUI(expr, env, quoted, outputArgs)**

**dataTableOutput(outputId)**

**imageOutput(outputId, width, height, click, dblclick, hover, brush, inline)**

**plotOutput(outputId, width, height, click, dblclick, hover, brush, inline)**

**verbatimTextOutput(outputId, placeholder)**

**tableOutput(outputId)**

**textOutput(outputId, container, inline)**

**uiOutput(outputId, inline, container, ...)**

**htmlOutput(outputId, inline, container, ...)**

## Inputs

Collect values from the user.

Access the current value of an input object with **input\$<inputId>**. Input values are **reactive**.

Action

**actionButton(inputId, label, icon, width, ...)**

Link

**actionLink(inputId, label, icon, ...)**

checkbox

**checkboxGroupInput(inputId, label, choices, selected, inline, width, choiceNames, choiceValues)**

checkbox

**checkboxInput(inputId, label, value, width)**

date

**dateInput(inputId, label, value, min, max, format, startview, weekstart, language, width, autoclose, datesdisabled, daysofweekdisabled)**

dateRange

**dateRangeInput(inputId, label, start, end, min, max, format, startview, weekstart, language, separator, width, autoclose)**

file

**fileInput(inputId, label, multiple, accept, width, buttonLabel, placeholder)**

number

**numericInput(inputId, label, value, min, max, step, width)**

password

**passwordInput(inputId, label, value, width, placeholder)**

radio

**radioButtons(inputId, label, choices, selected, inline, width, choiceNames, choiceValues)**

select

**selectInput(inputId, label, choices, selected, multiple, selectize, width, size)**  
Also **selectizeInput()**

slider

**sliderInput(inputId, label, min, max, value, step, round, format, locale, ticks, animate, width, sep, pre, post, timeFormat, timezone, dragRange)**

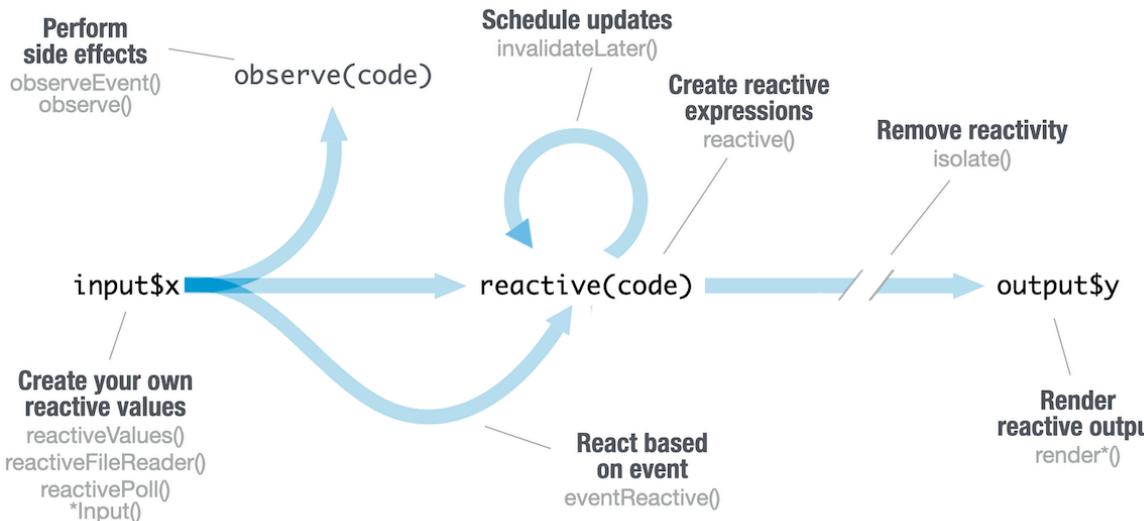
text

**textInput(inputId, label, value, width, placeholder)** Also **textAreaInput()**

These are the core output types. See [htmlwidgets.org](#) for many more options.

# Reactivity

Reactive values work together with reactive functions. Call a reactive value from within the arguments of one of these functions to avoid the error **Operation not allowed without an active reactive context**.



## CREATE YOUR OWN REACTIVE VALUES

```
# *Input() example
ui <- fluidPage(
  textInput("a","","A")
)
```

```
#reactiveVal example
server <- function(input,output){
  rv <- reactiveVal()
  rv$number <- 5
}
```

**\*Input()** functions  
Each input function creates a reactive value stored as `input$<inputId>`.

**reactiveVal(...)**  
Creates a single reactive values object.  
**reactiveValues(...)**  
Creates a list of names reactive values.

## CREATE REACTIVE EXPRESSIONS

```
ui <- fluidPage(
  textInput("a","","A"),
  textInput("z","","Z"),
  textOutput("b"))

server <- function(input,output){
  re <- reactive({
    paste(input$a,input$z)
  })
  output$b <- renderText({
    re()
  })
}
shinyApp(ui, server)
```

**reactive(x, env, quoted, label, domain)**  
**Reactive expressions:**

- cache their value to reduce computation
- can be called elsewhere
- notify dependencies when invalidated

Call the expression with function syntax, e.g. `re()`.

## REACT BASED ON EVENT

```
ui <- fluidPage(
  textInput("a","","A"),
  actionButton("go","Go"),
  textOutput("b"))

server <- function(input,output){
  re <- eventReactive(
    input$go, input$a)
  output$b <- renderText({
    re()
  })
}
shinyApp(ui, server)
```

**eventReactive(eventExpr, valueExpr, event.env, event.quoted, value.env, value.quoted, ..., label, domain, ignoreNULL, ignoreInit)**  
Creates reactive expression with code in 2nd argument that only invalidates when reactive values in 1st argument change.

## RENDERS REACTIVE OUTPUT

```
ui <- fluidPage(
  textInput("a","","A"),
  textOutput("b"))

server <- function(input,output){
  output$b <- renderText({
    input$a
  })
}
shinyApp(ui, server)
```

**render\*() functions**  
Builds an object to display. Will rerun code in body to rebuild the object whenever a reactive value in the code changes.  
Save the results to `output$<outputId>`.

## PERFORM SIDE EFFECTS

```
ui <- fluidPage(
  textInput("a","","A"),
  actionButton("go","Go"))

server <- function(input,output){
  observeEvent(
    input$go,
    print(input$a)
  )
}
shinyApp(ui, server)
```

**observe(x, env)**  
Creates an observer from the given expression.  
**observeEvent(eventExpr, handlerExpr, event.env, event.quoted, handler.env, handler.quoted, ..., label, suspended, priority, domain, autoDestroy, ignoreNULL, ignoreInit, once)**  
Runs code in 2nd argument when reactive values in 1st argument change.

## REMOVE REACTIVITY

```
ui <- fluidPage(
  textInput("a","","A"),
  textOutput("b"))

server <- function(input,output){
  output$b <- renderText({
    isolate({input$a})
  })
}
shinyApp(ui, server)
```

**isolate(expr)**  
Runs a code block. Returns a non-reactive copy of the results.

# UI

 - An app's UI is an HTML document.

Use Shiny's functions to assemble this HTML with R.

```
fluidPage(
  textInput("a","",)
)
## <div class="container-fluid">
## <div class="form-group shiny-input-container">
##   <label for="a"></label>
##   <input id="a" type="text"
##   class="form-control" value="">
## </div>
## </div>
```

Returns HTML

**HTML** Add static HTML elements with `tags`, a list of functions that parallel common HTML tags, e.g. `tags$a()`. Unnamed arguments will be passed into the tag; named arguments will become tag attributes.

Run `names(tags)` for a complete list.  
`tags$h1("Header")` → `<h1>Header</h1>`

The most common tags have wrapper functions. You do not need to prefix their names with `tags$`

```
ui <- fluidPage(
  h1("Header 1"),
  hr(),
  br(),
  p(strong("bold")),
  p(em("italic")),
  p(code("code")),
  a(href="", "link"),
  HTML("<p>Raw html</p>")
)
```

**Header 1**

bold  
italic  
code  
link  
Raw html

**CSS** To include a CSS file, use `includeCSS()`, or  
1. Place the file in the `www` subdirectory  
2. Link to it with:

`tags$head(tags$link(rel = "stylesheet", type = "text/css", href = "<file name>"))`

**JS** To include JavaScript, use `includeScript()` or  
1. Place the file in the `www` subdirectory  
2. Link to it with:

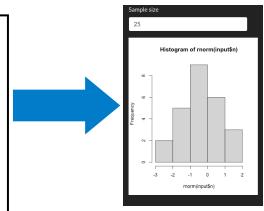
`tags$head(tags$script(src = "<file name>"))`

**IMAGES** To include an image:  
1. Place the file in the `www` subdirectory  
2. Link to it with `img(src = "<file name>")`

# Themes

Use the `bslib` package to add existing themes to your Shiny app ui, or make your own.

```
library(bslib)
ui <- fluidPage(
  theme = bs_theme(
    bootswatch = "darkly",
    ...
  )
)
```



**bootswatch\_themes()** Get a list of themes.

# Layouts



Use the `bslib` package to lay out your app and its components.

## PAGE LAYOUTS

### Dashboard layouts

`page_sidebar()` A sidebar page

`page_navbar()` Multi-page app with a top navigation bar

`page_fillable()` A screen-filling page layout

### Basic layouts

`page()`   `page_fluid()`   `page_fixed()`

## USER INTERFACE LAYOUTS

### Multiple columns

`layout_columns()`

Organize UI elements into Bootstrap's 12-column CSS grid

`layout_column_wrap()`

Organize elements into a grid of equal-width columns

### Multiple panels

`navset_tab()`

One Two Three

First tab content.

`navset_pill()`

One Two Three

First tab content.

`navset_underline()`

One Two Three

First tab content.

`nav_panel()` Content to display when given item is selected

`nav_menu()` Create a menu of nav items

`nav_item()` Place arbitrary content in the nav panel

`nav_spacer()` Add spacing between nav items

Also dynamically update nav containers with `nav_select()`,

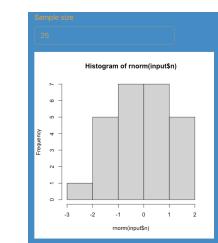
`nav_insert()`, `nav_remove()`, `nav_show()`, `nav_hide()`.

### Sidebar layout

`sidebar()`   `layout_sidebar()`   `toggle_sidebar()`

Build your own theme by customizing individual arguments.

**bs\_theme(bg = "#558AC5", fg = "#F9B02D", ...)**



**?bs\_theme** for a full list of arguments.

**bs\_themer()** Place within the server function to use the interactive theming widget.

# Shiny for Python :: CHEAT SHEET



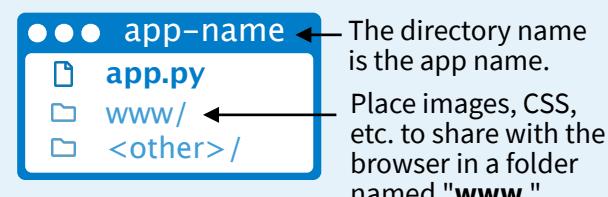
## Build an App

A **Shiny** app is an interactive web page (**ui**) powered by a live Python session run by a **server** (or by a browser with Shinylive).



Users can manipulate the UI, which will cause the server to update the UI's displays (by running Python code).

Save your app as **app.py** in a directory with the files it uses.



## Share

Share your app in three ways:

1. **Host it on shinyapps.io**, a cloud based service from Posit. To deploy Shiny apps:

Create a free or professional account at [shinyapps.io](https://shinyapps.io)

Use the reconnect-python package to publish with **rsconnect deploy shiny <path to directory>**

2. **Purchase Posit Connect**, a publishing platform for R and Python.

[posit.co/connect](https://posit.co/connect)

3. **Use open source deployment options**

[shiny.posit.co/py/docs/deploy.html](https://shiny.posit.co/py/docs/deploy.html)

Nest Python functions to build an HTML interface

Add Inputs with `ui.input_*` functions

Add Outputs with `ui.output_*` functions

For each output, define a function that generates the output

Call the values of UI inputs with `input.<id>()`

Run `shiny create .` in the terminal to generate a template `app.py` file

```
from shiny import App, render, ui
import matplotlib.pyplot as plt
import numpy as np

app_ui = ui.page_fluid(
    ui.input_slider(
        "n", "Sample Size", 0, 1000, 20
    ),
    ui.output_plot("dist")
)

def server(input, output, session):
    @render.plot
    def dist():
        x = np.random.normal(0, 1, input.n)
        plt.hist(x, range=[-3, 3])

app = App(app_ui, server)
```

Layout the UI with Layout Functions

Specify the type of output with a `@render` decorator

Call App() to combine `app_ui` and `server` into an interactive app

Launch apps with `shiny run app.py --reload`

## Shinylive

Shinylive apps use WebAssembly to run entirely in a browser—no need for a special server to run Python.



- Edit and/or host Shinylive apps at [shinylive.io](https://shinylive.io)
- Create a Shinylive version of an app to deploy with `shinylive export myapp site`. Then deploy to a hosting site like Github or Netlify
- Embed Shinylive apps in Quarto sites, blogs, etc.

```
---  
filters:  
- shinylive  
---  
An embedded Shinylive app:  
```{shinylive-python}  
# standalone: true  
# [App.py code here...]  
```
```

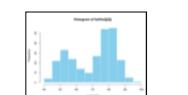
To embed a Shinylive app in a Quarto doc, include the bold syntax.

## Outputs

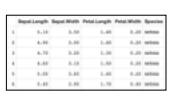
Match `ui.output_*` functions to `@render.*` decorators to link Python output to the UI.



`ui.output_data_frame(id)`  
`@render.data_frame`



`ui.output_image(id, width, height, click, dblclick, hover, brush, inline)`  
`@render.image`



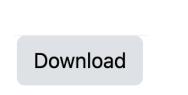
`ui.output_plot(id, width, height, click, dblclick, hover, brush, inline)`  
`@render.plot`



`ui.output_table(id)`  
`@render.table`



`ui.output_text_verbatim(id, ...)`  
`ui.output_text(id, container, inline)`  
`@render.text`



`ui.output_ui(id, inline, container, ...)`  
`ui.output_html(id, inline, container, ...)`  
`@render.ui`

`ui.download_button(id, label, icon, ...)`  
`@session.download`

## Inputs

Use a `ui.` function to make an input widget that saves a value as `<id>`. Input values are *reactive* and need to be called as `<id>()`.

Action

`ui.input_action_button(id, label, icon, width, ...)`

Link

`ui.input_action_link(id, label, icon, ...)`

Check me

`ui.input_checkbox(id, label, value, width)`

Choice 1

`ui.input_checkbox_group(id, label, choices, selected, inline, width)`

Choice 2

`ui.input_date(id, label, value, min, max, format, startview, weekstart, language, width, autoclose, datesdisabled, daysofweekdisabled)`

Choice 3

`ui.input_date_range(id, label, start, end, min, max, format, startview, weekstart, language, separator, width, autoclose)`

Choose File

`ui.input_file(id, label, multiple, accept, width, buttonLabel, placeholder, capture)`

1

`ui.input_numeric(id, label, value, min, max, step, width)`

.....

`ui.input_password(id, label, value, width, placeholder)`

Choice A

`ui.input_radio_buttons(id, label, choices, selected, inline, width)`

Choice B

`ui.input_select(id, label, choices, selected, multiple, selectize, width, size)`

Choice C

`ui.input_selectize(id, label, choices, selected, multiple, selectize, width, size)`

Choice 1

`ui.input_slider(id, label, min, max, value, step, ticks, animate, width, sep, pre, post, dateFormat, timezone, dragRange)`

Enter text

`ui.input_switch(id, label, value, width)`

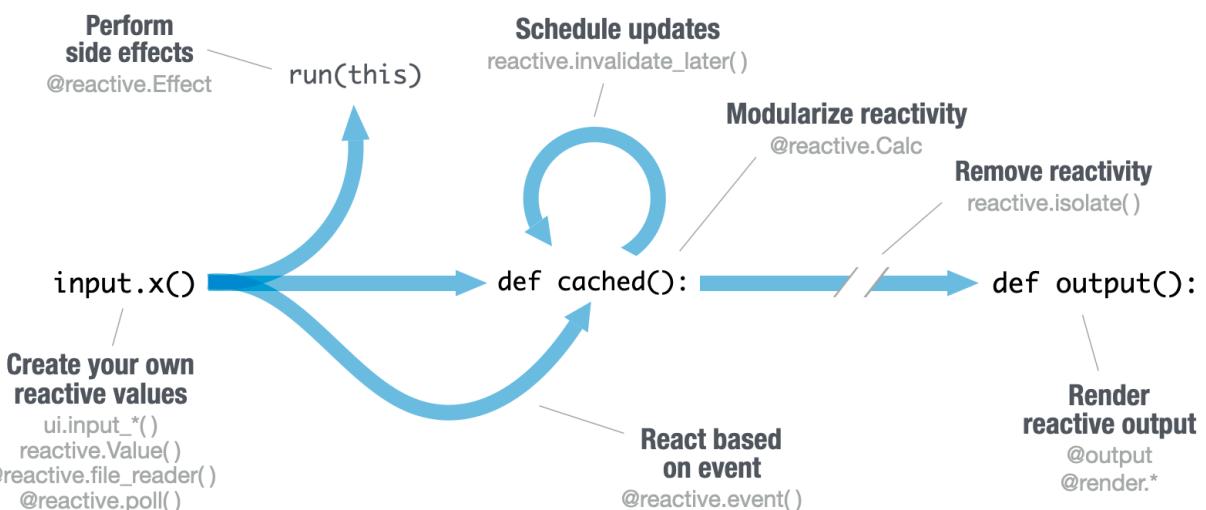
Enter text

`ui.input_text(id, label, value, width, placeholder, autocomplete, spellcheck)`

`ui.input_text_area(id, label, value, width, placeholder, rows, cols, spellcheck)`

# Reactivity

Reactive values work together with reactive functions. Call a reactive value from within the arguments of one of these functions to avoid the error **No current reactive context**.



## CREATE YOUR OWN REACTIVE VALUES

```
# ...
app_ui = ui.page_fluid(
  ui.input_text("a", "A")
)

def server(
  input, output, session
):
  rv = reactive.Value()
  rv.set(5)
# ...
```

**ui.input\_\***() makes an input widget that saves a reactive value as **input.<id>()**.

**reactive.value()** Creates an object whose value you can set.

## DISPLAY REACTIVE OUTPUT

```
app_ui = ui.page_fluid(
  ui.input_text("a", "A"),
  ui.output_text("b"),
)
def server(
  input, output, session
):
  @render.text
  def b():
    return input.a()
```

**ui.output\_\***() adds an output element to the UI.

**@render.\*** Decorator to identify and render outputs

**def <id>()**: Code to generate the output

## CREATE REACTIVE EXPRESSIONS

```
# ...
def server(
  input, output, session
):
  @reactive.calc
  def re():
    return input.a() + input.b()
# ...
```

**@reactive.calc** Makes a function a reactive expression. Shiny notifies functions that use the expression when it becomes invalidated, triggering recomputation. Shiny caches the value of the expression while it is valid to avoid unnecessary computation.

## PERFORM SIDE EFFECTS

```
# ...
def server(
  input, output, session
):
  @reactive.effect
  @reactive.event(input.a)
  def print():
    print("Hi!")
# ...
```

**@reactive.effect** Reactively trigger a function with a side effect. Call a reactive value or use **@reactive.event** to specify when the function will rerun.

## REACT BASED ON EVENT

```
# ...
def server(
  input, output, session
):
  @reactive.Calc
  @reactive.event(input.a)
  def re():
    return input.b()
# ...
```

**@reactive.event()** Makes a function react *only when* a specified value is invalidated, here **input.a**.

## REMOVE REACTIVITY

```
# ...def server(
  input, output, session
):
  @render.text
  def a():
    with reactive.isolate():
      return input.a()
# ...
```

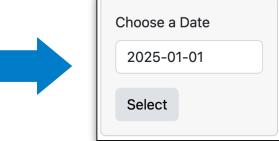
**reactive.isolate()** Create non-reactive context within a reactive function. Calling a reactive value within this context will *not* cause the calling function to re-execute should the value become invalid.

# Layouts

Combine multiple elements into a "single element" that has its own properties with a panel function:

|                        |                        |
|------------------------|------------------------|
| ui.panel_absolute()    | ui.panel_sidebar()     |
| ui.panel_conditional() | ui.panel_title()       |
| ui.panel_fixed()       | ui.panel_well()        |
| ui.panel_main()        | ui.row() / ui.column() |

ui.panel\_well(  
ui.input\_date(...),  
ui.input\_action\_button(...)  
)



Layout panels with a layout function. Add elements as arguments of the layout functions.

## ui.layout\_sidebar()

title panel  
side panel main panel

## ui.row()

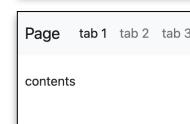
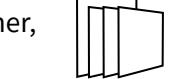
column col  
column

Layer **ui.nav()**s on top of each other, and navigate between them, with:

ui.page\_fluid(ui.navset\_tab(  
 ui.nav("tab 1", "contents"),  
 ui.nav("tab 2", "contents"),  
 ui.nav("tab 3", "contents")))

ui.page\_fluid(ui.navset\_pill\_list(  
 ui.nav("tab 1", "contents"),  
 ui.nav("tab 2", "contents"),  
 ui.nav("tab 3", "contents")))

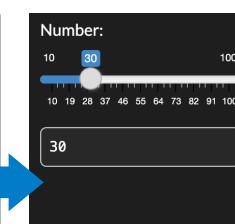
ui.page\_navbar(  
 ui.nav("tab 1", "contents"),  
 ui.nav("tab 2", "contents"),  
 ui.nav("tab 3", "contents"),  
 title = "Page")



# Themes

Use the **shinyswatch** package to add existing bootstrap themes to your Shiny app ui.

```
import shinyswatch
app_ui = ui.page_fluid(
  shinyswatch.theme.darkly(),
...
)
```



# Shiny for R Comparison



Shiny for Python is quite similar to Shiny for R with a few important differences:

1. Call inputs as **input.<id>()**

|          |           |
|----------|-----------|
| input\$x | input.x() |
|----------|-----------|

2. Define outputs as decorated functions **def <id>()**:

|                              |                                       |
|------------------------------|---------------------------------------|
| output\$y <- renderText(z()) | @renderText<br>def y():<br>return z() |
|------------------------------|---------------------------------------|

3. To create a reactive expression, use **@reactive.calc**

|                                       |  |
|---------------------------------------|--|
| z <- reactive({<br>input\$x + 1<br>}) | @reactive.calc<br>def z():<br>return input.x()+1 |
|---------------------------------------|--|

4. To create an observer, use **@reactive.effect**

|   |  |
|---|--|
| a <- observe({<br>print(input\$x)<br>}) | @reactive.effect<br>def a():<br>print(input.x()) |
|---|--|

5. Combine these with **@reactive.event**

|   |   |
|---|---|
| b <- eventReactive(<br>input\$goCue,<br>{input.go_cue<br>}) | @reactive.event<br>def b():<br>return input.x()+1 |
|---|---|

6. Use **reactive.value()** instead of **reactiveVal()**

|                |                   |
|----------------|-------------------|
| reactiveVal(1) | reactive.value(1) |
|----------------|-------------------|

7. Use **nav\_\***() instead of **\*Tab()**

|                                    |                                      |
|------------------------------------|--------------------------------------|
| insertTab()<br>appendTab()<br>etc. | nav_insert()<br>nav_append()<br>etc. |
|------------------------------------|--------------------------------------|

8. Functions are intuitively organized into submodules

|                                    |  |
|------------------------------------|--|
| dateInput()<br>textInput()<br>etc. | ui.input_date()<br>ui.input_text()<br>etc. |
|------------------------------------|--|

# Data & Variable Transformation with sjmisc Cheat Sheet



sjmisc complements dplyr, and helps with data transformation tasks and recoding *variables*.

sjmisc works together seamlessly with dplyr and pipes. All functions are designed to support labelled data.



## Design Philosophy

The design of sjmisc functions follows the tidyverse-approach: first argument is always the data (either a *data frame* or *vector*), followed by variable names to be processed by the functions.

The returned object for each function *equals the type of the data-argument*.

### Vector input

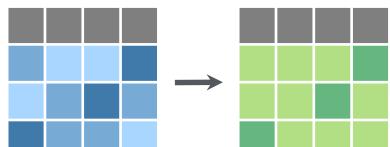
- If the data-argument is a *vector*, functions return a *vector*.



```
rec(mtcars$carb, rec = "1,2=1; 3,4=2; else=3")
```

### Data frame input

- If the data-argument is a *data frame*, functions return a *data frame*.



```
rec(mtcars, carb, rec = "1,2=1; 3,4=2; else=3")
```

## The ...-ellipses Argument

Apply functions to a single variable, selected variables or to a complete data frame.

Variable selection is powered by dplyr's `select()`: Separate variables with comma, or use dplyr's select-helpers to select variables, e.g. `?rec`:

```
rec(mtcars, one_of(c("gear", "carb")),  
    rec = "min:3=1; 4:max=2")
```

```
rec(mtcars, gear, carb, rec = "min:3=1; 4:max=2")
```

## Descriptives and Summaries

Most of the sjmisc functions (including recode-functions) also work on grouped data frames:

```
library(dplyr)  
efc %>%  
  group_by(e16sex, c172code) %>%  
  frq(e42dep)
```

### Frequency Tables

```
frq(x, ..., sort.frq = c("none", "asc", "desc"),  
     weight.by = NULL, auto.grp = NULL, ...)
```

Print frequency tables of (labelled) vectors. Uses variable labels as table header.

```
data(efc); frq(efc, e42dep, c161sex)
```

Use this data set in examples!

```
flat_table(data, ..., margin = c("counts",  
  "cell", "row", "col"), digits = 2,  
  show.values = FALSE)
```

Print contingency tables of (labelled) vectors. Uses value labels.

```
flat_table(efc, e42dep, c172code, e16sex)
```

```
count_na(x, ...)
```

Print frequency table of tagged NA values.

```
library(haven); x <- labelled(c(1:3,  
  tagged_na("a", "a", "z")), labels =  
  c("Refused" = tagged_na("a"), "N/A" =  
  tagged_na("z")))  
count_na(x)
```

### Descriptive Summary

```
descr(x, ..., max.length = NULL)
```

Descriptive summary of data frames, including variable labels in output.

```
descr(efc, contains("cop"), max.length = 20)
```

## Finding Variables in a Data Frame

Use `find_var()` to search for variables by names, value or variable labels. Returns vector/data frame.

```
# variables with "cop" in names and variable labels  
find_var(efc, pattern = "cop", out = "df")
```

```
# variables with "level" in names and value labels  
find_var(efc, "level", search = "name_value")
```

## Recode and Transform Variables

Recode functions add a *suffix* to new variables, so original variables are preserved.

By default, original input data frame and new created variables are returned. Use `append = FALSE` to return the recoded variables only.

```
rec(x, ..., rec, as.num = TRUE, var.label =  
  NULL, val.labels = NULL, append = TRUE,  
  suffix = "_r")
```

Recode values, return result as numeric, character or categorical (factor).

```
rec(mtcars, carb, rec = "1,2=1; 3,4=2; else=3")
```

```
dicho(x, ..., dich.by = "median", as.num =  
  FALSE, var.label = NULL, val.labels = NULL,  
  append = TRUE, suffix = "_d")
```

Dichotomise variable by median, mean or specific value.

```
dicho(mtcars, disp)
```

```
split_var(x, ..., n, as.num = FALSE,  
  val.labels = NULL, var.label = NULL,  
  inclusive = FALSE, append = TRUE,  
  suffix = "_g")
```

Split variable into equal sized groups. Unlike `dplyr::ntile()`, does not split original categories into different values (see examples in `?split_var`).

```
split_var(mtcars, mpg, disp, n = 3)
```

```
group_var(x, ..., size = 5, as.num = TRUE,  
  right.interval = FALSE, n = 30, append =  
  TRUE, suffix = "_gr")
```

Split variable into groups with equal value range, or into a max. # of groups (value range per group is adjusted to match # of groups).

```
group_var(mtcars, mpg, disp, size = 5)
```

```
group_var(mtcars, mpg, size = "auto", n = 4)
```

```
std(x, ..., robust = "sd", include.fac = FALSE,  
  append = TRUE, suffix = "_z")
```

Z-standardise variables. Also `center()`.

```
std(efc, e17age, c160age)
```

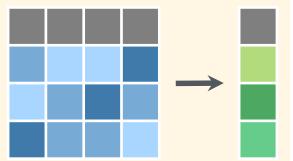
```
recode_to(x, ..., lowest = 0, highest = -1,  
  append = TRUE, suffix = "_r0")
```

Shift ("renumber") categories or values.

```
recode_to(mtcars$gear)
```

## Summarise Variables and Cases

The summary functions mostly mimic base R equivalents, but are designed to work together with pipes and dplyr.



```
row_sums(x, ..., na.rm = TRUE, var =  
  "rowsums", append = FALSE)
```

Row sums of data frames.

```
row_sums(efc, c82cop1:c90cop9)
```

```
row_means(x, ..., n, var = "rowmeans",  
  append = FALSE)
```

Row means, for at least `n` valid (non-NA) values.

```
row_means(efc, c82cop1:c90cop9, n = 7)
```

```
row_count(x, ..., count, var = "rowcount",  
  append = FALSE)
```

Row-wise count # of values in data frames.

```
row_count(efc, c82cop1:c90cop9, count = 2)
```

## Other Useful Functions

- `add_columns()` and `replace_columns()` to combine data frames, but either replace or preserve existing columns.

- `set_na()` and `replace_na()` to convert regular into missing values, or vice versa. `replace_na()` also replaces specific `tagged NA` values only.

- `remove_var()` and `var_rename()` to remove variables from data frames, or rename variables.

- `group_str()` to group similar string values. Useful for variables with similar, but not identically spelled string values that should be "merged".

- `merge_df()` to full join data frames and preserve value and variable labels.

- `to_long()` to gather multiple columns in data frames from wide into long format.

## Use with %>% and dplyr

```
# use sjmisc-functions in pipes  
mtcars %>% select(gear, carb) %>%  
  rec(rec = "min:3=1; 4:max=2")
```

```
# use sjmisc-function inside mutate  
mtcars %>% select(gear, carb) %>% mutate(  
  carb2 = rec(carb, rec = "1,2=0;3:8=1"),  
  gear2 = rec(gear, rec = "3=1;4:max=2"))
```



# slackr :: CHEAT SHEET

Send files, messages, R objects, and images to Slack directly from R

## Installation

### CRAN version

```
install.packages("slackr")
```

### Development version

```
devtools::install_github("mrkaye97/slackr")
```

## Setup

- Go to <https://api.slack.com/apps>

- Click “Create New App” and then follow the setup instructions

### Multi-channel bot

- Click “OAuth & Permissions” under “Features”
- Enable the following scopes in order to get all of the functionality:
  - channels:read
  - chat:write
  - users:read
  - chat:write.customize
  - files:read
  - chat:write.public
  - groups:read
  - im:write
  - groups:write
  - incoming-webhook
  - channels:history
- Click “Install to Workspace”
- Select a channel (for webhook messages)
- Copy the Bot User OAuth Access Token
- Click “Incoming Webhooks” under “Features”
- Copy the Webhook URL

### Single-channel bot

- Click “Incoming “Webhooks” under “Features”
- Turn the “Activate Incoming Webhooks” switch on
- Click “Add New Webhook to Workspace”
- Select the channel you’d like the bot to post to
- Copy the Webhook URL

### Setup environment variables for Slack API access

```
slackr_setup(
  channel = "#general",
  username = "slackr",
  icon_emoji = "",
  incoming_webhook_url = "",
  bot_user_oauth_token = "",
  config_file = "~/.slackr",
  echo = FALSE,
  cacheChannels = TRUE,
  cache_dir = "")
```

*Generate Config Text (.txt) File (optional)*  
 bot\_user\_oauth\_token: xoxb-[...]  
 channel: #yourchannel  
 username: yourusername  
 incoming\_webhook\_url: https://hooks.slack.com/services/XXXXXX/XXXXX/XXXXXX

## Common Functions

### Post a ggplot to a Slack channel

```
ggslackr(
  plot = last_plot(),
  channels = Sys.getenv("SLACK_CHANNEL"),
  scale = 1,
  width = par("din")[1],
  height = par("din")[2],
  units = c("in", "cm", "mm"),
  dpi = 300,
  limitsize = TRUE,
  bot_user_oauth_token =
    Sys.getenv("SLACK_BOT_USER_OAUTH_TOKEN"),
  file = "ggplot",
  ...)
```

### Save R objects to an RData file on Slack

```
save_slackr(
  ...,
  channels = Sys.getenv("SLACK_CHANNEL"),
  file = "slackr",
  bot_user_oauth_token =
    Sys.getenv("SLACK_BOT_USER_OAUTH_TOKEN"),
  plot_text = ""
)
```

### Send result of R expressions to a Slack channel via webhook API

```
slackr_bot(
  ...,
  channel = "",
  username = "",
  icon_emoji = "",
  incoming_webhook_url =
    Sys.getenv("SLACK_INCOMING_URL_PREFIX"))
```

### Delete the specified number of messages from the channel

```
slackr_delete(
  count,
  channel = Sys.getenv("SLACK_CHANNEL"),
  bot_user_oauth_token =
    Sys.getenv("SLACK_BOT_USER_OAUTH_TOKEN"))
```

### Send a message to a Slack channel

```
slackr_msg(
  txt = "",
  channel = Sys.getenv("SLACK_CHANNEL"),
  username = Sys.getenv("SLACK_USERNAME"),
  icon_emoji = Sys.getenv("SLACK_ICON_EMOJI"),
  bot_user_oauth_token =
    Sys.getenv("SLACK_BOT_USER_OAUTH_TOKEN"),
  ...)
```

### Send a file to Slack

```
slackr_upload(
  filename,
  title = basename(filename),
  initial_comment = basename(filename),
  channels = Sys.getenv("SLACK_CHANNEL"),
  bot_user_oauth_token =
    Sys.getenv("SLACK_BOT_USER_OAUTH_TOKEN"))
```

## Other Usages

|                              |  |
|------------------------------|--|
| auth_test()                  | Checks authentication & identity against the Slack API                                   |
| call_slack_api()             | A wrapper function to call the Slack API with authentication and pagination              |
| convert_response_to_tibble() | Convert Slack API json response to tibble  |
| register_onexit()            | Append text_slackr as on.exit to functions   |
| slackr_census_fun()          | Create a cache of the users and channels in the workspace in order to limit API requests |
| slackr_channels()            | Get a data frame of Slack channels   |
| slackr_chtrans()             | Translate vector of channel names to channel IDs for API                                 |
| slackr_dev()                 | Send the graphics contents of the current device to a Slack channel                      |
| slackr_history()             | Reads history of a channel   |
| slackr_ims()                 | Get a data frame of Slack IM IDs   |
| slackr_users()               | Get a data frame of Slack users  |
| text_slackr()                | Sends basic text to a slack channel  |
| tex_slackr()                 | Post a tex output to a Slack channel   |
| with_pagination()            | Calls the slack API with pagination using cursors  |

## Vignettes

|  |  |
|--|--|
| <i>Creating a single-channel bot</i>                 | vignette('webhook-setup', package = 'slackr')    |
| <i>Creating a fully-functional multi-channel bot</i> | vignette('scoped-bot-setup', package = 'slackr') |
| <i>Usage</i>   | vignette('using-slackr', package = 'slackr')     |

# Data Science in Spark with sparklyr :: CHEAT SHEET



## Connect

### DATABRICKS CONNECT (v2)

1. Open your .Renviron file: `usethis::edit_r_environ()`
2. In the .Renviron file add your Databricks Host Url and Token (PAT):
  - `DATABRICKS_HOST = [Your Host URL]`
  - `DATABRICKS_TOKEN = [Your PAT]`
3. Install extension: `install.packages("pysparklyr")`
4. Open connection:
 

```
sc <- spark_connect(
  cluster_id = "[Your cluster's ID]",
  method = "databricks_connect"
)
```

= Supported in Databricks Connect v2

### STANDALONE CLUSTER

1. Install RStudio Server on one of the existing nodes or a server in the same LAN
2. Open a connection
 

```
spark_connect(master="spark://host:port",
  version = "3.2",
  spark_home = [path to Spark])
```

### YARN CLIENT

1. Install RStudio Server on an edge node
2. Locate path to the cluster's Spark Home Directory, it normally is `"/usr/lib/spark"`
3. Basic configuration example
 

```
conf <- spark_config()
conf$spark.executor.memory <- "300M"
conf$spark.executor.cores <- 2
conf$spark.executor.instances <- 3
conf$spark.dynamicAllocation.enabled<-"false"
```
4. Open a connection
 

```
sc <- spark_connect(master = "yarn",
  spark_home = "/usr/lib/spark/",
  version = "2.1.0", config = conf)
```

### YARN CLUSTER

1. Make sure to have copies of the `yarn-site.xml` and `hive-site.xml` files in the RStudio Server
2. Point environment variables to the correct paths
 

```
Sys.setenv(JAVA_HOME="[Path]")
Sys.setenv(SPARK_HOME ="[Path]")
Sys.setenv(YARN_CONF_DIR ="[Path]")
```
3. Open a connection
 

```
sc <- spark_connect(master = "yarn-cluster")
```

## KUBERNETES

1. Use the following to obtain the Host and Port
 

```
system2("kubectl", "cluster-info")
```
2. Open a connection
 

```
sc <- spark_connect(config =
  spark_config_kubernetes(
    "k8s://https://[HOST]:[PORT]",
    account = "default",
    image = "docker.io/owner/repo:version"
))
```

## LOCAL MODE

No cluster required. [Use for learning purposes only](#)

1. Install a local version of Spark: `spark_install()`
2. Open a connection

```
sc <- spark_connect(master="local")
```

## CLOUD

Azure - `spark_connect(method = "synapse")`  
Qubole- `spark_connect(method = "qubole")`

## Import



### READ A FILE INTO SPARK

Arguments that apply to all functions:  
`sc, name, path, options=list(), repartition=0, memory=TRUE, overwrite=TRUE`

#### CSV

```
spark_read_csv(header = TRUE,
  columns=NULL, infer_schema=TRUE,
  delimiter = ";", quote = "\"", escape = "\\",
  charset = "UTF-8", null_value = NULL)
```

#### JSON

```
spark_read_json()
```

#### PARQUET

```
spark_read_parquet()
```

#### TEXT

```
spark_read_text()
```

#### DELTA

```
spark_read_delta()
```

## FROM A TABLE

`dplyr::tbl(scr, ...)` - Creates a reference to the table without loading its data into memory

`dbplyr::in_catalog()` - Enables a three part table address

```
x <- tbl(sc,in_catalog("catalog", "schema", "table"))
```

## Import

- From R (`copy_to()`)
- Read a file (`spark_read_`)
- Read Hive table (`tbl()`)

## Wrangle

- `dplyr` verb
- `tidy` commands
- Feature transformer (`ft_`)
- Direct Spark SQL (`DBI`)

[R for Data Science](#), [Wickham](#), [Çetinkaya-Rundel](#), [Grolmusz](#)

## R DATA FRAME INTO SPARK

`dplyr::copy_to(dest, df, name)`

Apache Arrow accelerates data transfer between R and Spark. To use, simply load the library

library(sparklyr)
library(arrow)

## Wrangle

### DPLYR VERBS

Translates into Spark SQL statements

```
copy_to(sc, mtcars) |>
  mutate(trm = ifelse(am == 0,
    "auto", "man")) |>
  group_by(trm) |>
  summarise_all(mean)
```

### TIDYR

`pivot_longer()` - Collapse several columns into two.

`pivot_wider()` - Expand two columns into several.

`nest()` / `unnest()` - Convert groups of cells into list-columns, and vice versa.

`unite()` / `separate()` - Split a single column into several columns, and vice versa.

`fill()` - Fill NA with the previous value

## FEATURE TRANSFORMERS

`ft_binarizer()` - Assigned values based on threshold

`ft_bucketizer()` - Numeric column to discretized column

`ft_count_vectorizer()` - Extracts a vocabulary from document

`ft_discrete_cosine_transform()` - 1D discrete cosine transform of a real vector

`ft_elementwise_product()` - Element-wise product between 2 cols

`ft_hashing_tf()` - Maps a sequence of terms to their term frequencies using the hashing trick.

`ft_idf()` - Compute the Inverse Document Frequency (IDF) given a collection of documents.

`ft_imputer()` - Imputation estimator for completing missing values, uses the mean or the median of the columns.

`ft_index_to_string()` - Index labels back to label as strings

`ft_interaction()` - Takes in Double and Vector columns and outputs a flattened vector of their feature interactions.

`ft_max_abs_scaler()` - Rescale each feature individually to range [-1, 1]

`ft_min_max_scaler()` - Rescale each feature to a common range [min, max] linearly

`ft_ngram()` - Converts the input array of strings into an array of n-grams

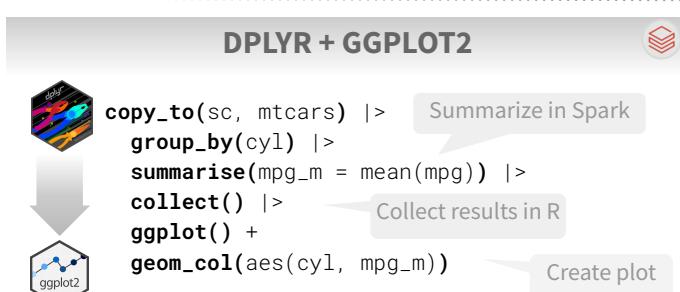
`ft_bucketed_random_projection_lsh()`  
`ft_minhash_lsh()` - Locality Sensitive Hashing functions for Euclidean distance and Jaccard distance (MinHash)

# Data Science in Spark with sparklyr :: CHEAT SHEET



|  |  |
|--|--|
|  | <code>ft_norm()</code> - Normalize a vector to have unit norm using the given p-norm   |
|  | <code>ft_one_hot_encoder()</code> - Continuous to binary vectors   |
|  | <code>ft_pca()</code> - Project vectors to a lower dimensional space of top k principal components.                                |
|  | <code>ft_quantile_discretizer()</code> - Continuous to binned categorical values.  |
|  | <code>ft_regex_tokenizer()</code> - Extracts tokens either by using the provided regex pattern to split the text.                  |
|  | <code>ft_robust_scaler()</code> - Removes the median and scales according to standard scale.                                       |
|  | <code>ft_standard_scaler()</code> - Removes the mean and scaling to unit variance using column summary statistics                  |
|  | <code>ft_stop_words_remover()</code> - Filters out stop words from input   |
|  | <code>ft_string_indexer()</code> - Column of labels into a column of label indices.  |
|  | <code>ft_tokenizer()</code> - Converts to lowercase and then splits it by white spaces   |
|  | <code>ft_vectorAssembler()</code> - Combine vectors into single row-vector   |
|  | <code>ft_vector_indexer()</code> - Indexing categorical feature columns in a dataset of Vector                                     |
|  | <code>ft_vector_slicer()</code> - Takes a feature vector and outputs a new feature vector with a subarray of the original features |
|  | <code>ft_word2vec()</code> - Word2Vec transforms a word into a code  |

## Visualize



## Modeling

### REGRESSION

`ml_linear_regression()` - Linear regression.  
`ml_aft_survival_regression()` - Parametric survival regression model named accelerated failure time (AFT) model  
`ml_generalized_linear_regression()` - GLM  
`ml_isotonic_regression()` - Uses parallelized pool adjacent violators algorithm.  
`ml_random_forest_regressor()` - Regression using random forests.

### CLASSIFICATION

`ml_linear_svc()` - Classification using linear support vector machines  
`ml_logistic_regression()` - Logistic regression  
`ml_multilayer_perceptron_classifier()` - Based on the Multilayer Perceptron.  
`ml_naive_bayes()` - It supports Multinomial NB which can handle finitely supported discrete data  
`ml_one_vs_rest()` - Reduction of Multiclass, performs reduction using one against all strategy.

### TREE

`ml_decision_tree_classifier()` | `ml_decision_tree()` | `ml_decision_tree_regressor()` - Classification and regression using decision trees  
`ml_gbt_classifier()` | `ml_gradient_boosted_trees()` | `ml_gbt_regressor()` - Binary classification and regression using gradient boosted trees  
`ml_random_forest_classifier()` - Classification and regression using random forests.  
`ml_feature_importances()` | `ml_tree_feature_importance()` - Feature Importance for Tree Models

### CLUSTERING

`ml_bisecting_kmeans()` - A bisecting k-means algorithm based on the paper  
`ml_lda()` | `ml_describe_topics()` | `ml_log_likelihood()` | `ml_log_perplexity()` | `ml_topics_matrix()` - LDA topic model designed for text documents.  
`ml_gaussian_mixture()` - Expectation maximization for multivariate Gaussian Mixture Models (GMMs)

`ml_kmeans()` | `ml_compute_cost()`  
`| ml_compute_silhouette_measure()` - Clustering with support for k-means

`ml_power_iteration()` - For clustering vertices of a graph given pairwise similarities as edge properties.

### RECOMMENDATION

`ml_als()` | `ml_recommend()` - Recommendation using Alternating Least Squares matrix factorization

### EVALUATION

`ml_clustering_evaluator()` - Evaluator for clustering  
`ml_evaluate()` - Compute performance metrics  
`ml_binary_classification_evaluator()` | `ml_binary_classification_eval()` | `ml_classification_eval()` - A set of functions to calculate performance metrics for prediction models.

### FREQUENT PATTERN

`ml_fpgrowth()` | `ml_association_rules()` | `ml_freq_itemsets()` - A parallel FP-growth algorithm to mine frequent itemsets.

`ml_freq_seq_patterns()` | `ml_prefixspan()` - PrefixSpan algorithm for mining frequent itemsets.

### STATS

`ml_summary()` - Extracts a metric from the summary object of a Spark ML model

`ml_corr()` - Compute correlation matrix

### FEATURE

`ml_chisquare_test(x,features,label)` - Pearson's independence test for every feature against the label  
`ml_default_stop_words()` - Loads the default stop words for the given language

### UTILITIES

`ml_call_constructor()` - Identifies the associated sparklyr ML constructor for the JVM  
`ml_model_data()` - Extracts data associated with a Spark ML model  
`ml_standardize_formula()` - Generates a formula string from user inputs  
`ml_uid()` - Extracts the UID of an ML object.

## ML Pipelines

*Easily create a formal Spark Pipeline models using R. Save the Pipeline in native Scala. It will have no dependencies on R.*

### INITIALIZE AND TRAIN

`ml_pipeline()` - Initializes a new Spark Pipeline

`ml_fit()` - Trains the model, outputs a Spark Pipeline Model.

### SAVE AND RETRIEVE

`ml_save()` - Saves into a format that can be read by Scala and PySpark .

`ml_read()` - Reads Spark object into sparklyr.

```

ft_dplyr_transformer() ml_linear_regression()
ml_pipeline() > ft_bucketizer() ml_fit() ml_save()
  
```

[spark.posit.co/guides/pipelines](http://spark.posit.co/guides/pipelines)

## Distributed R

Run arbitrary R code at scale inside your cluster with `spark_apply()`. Useful when there you need functionality only available in R, and to solve 'embarrassingly parallel problems'

`spark_apply(x, f, columns = NULL, memory = TRUE, group_by = NULL, name = NULL, barrier = NULL, fetch_result_as_sdf = TRUE)`

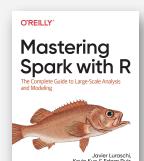
```

copy_to(sc, mtcars) |>
spark_apply(
  nrow, # R only function
  group_by = "am",
  columns = "am double, x long"
)
  
```

## More Info



[spark.posit.co](http://spark.posit.co)



[therinspark.com](http://therinspark.com)

# SqueakR :: CHEAT SHEET



## Adding new data

### 1. Load an Excel file using :

```
my_file <-  
add_timepoint_data(data_path =  
"path_to_data", t1 = 2, t2 = 12)
```

### 2. Score the loaded dataset using:

```
my_file <-  
score_timepoint_data(data_subset =  
my_file, group = "Drug",  
experimenter = "My Name")
```

### 3. Add the processed dataset to your experiment object:

```
experiment <-  
add_to_experiment(my_file)
```

Save the experiment to a specified directory:

```
experiment <-  
save_experiment(experiment =  
experiment, save_path = "save-dir")
```

## Experiment.RData

Your experiment object will be saved to a specified directory using the `save_experiment( )` function.

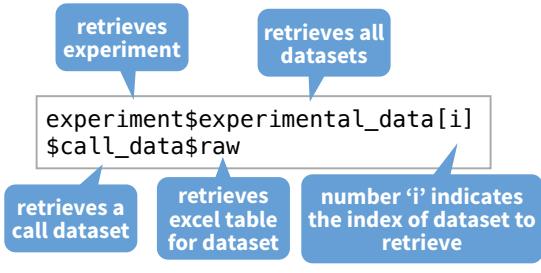
The file will be saved as an .RData file, and can be loaded into R at any point to continue analysis, or add or remove more data.

Experiment object files can also be loaded into the **SqueakR Dashboard** for visualization.

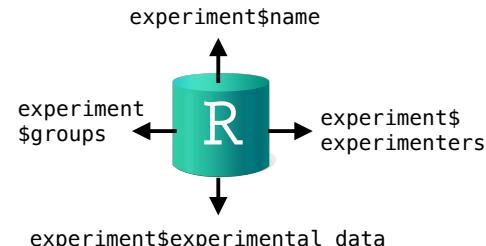
## Managing Experiment

### CODE

How to navigate the experiment object:



### EXPERIMENT OBJECT



### EXPERIMENT CONTENTS

| item              | description  |
|-------------------|--|
| name              | the name of the experiment                         |
| last_saved        | the date the object was last saved                 |
| groups            | the experimental groups                            |
| experimenters     | the experimenters who collected data               |
| experimental_data | the full dataset of scored data for the experiment |

## SqueakR Pipeline(s)

```
experiment <- semisqueakRpipeline()
```

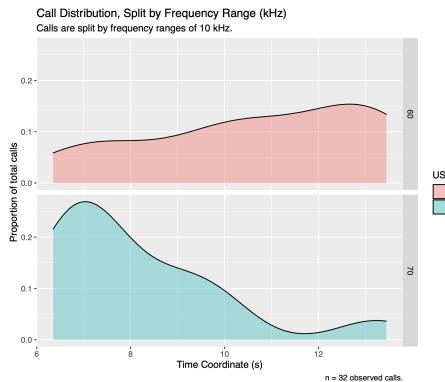
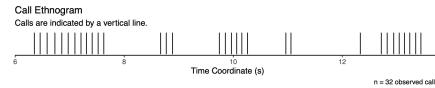
creates an experiment by looping through a folder of DeepSqueak data, prompting the user for each file's metadata

```
experiment <- autosqueakRpipeline()
```

creates an experiment by looping through a folder of DeepSqueak data, pulling metadata from a Google Sheet

## Visualizations

Graphs which can be generated using SqueakR:



## Learning SqueakR

### DOWNLOAD AND INSTALLATION

To install the package, run the following in RStudio:

```
install.packages("SqueakR")  
library(SqueakR)
```

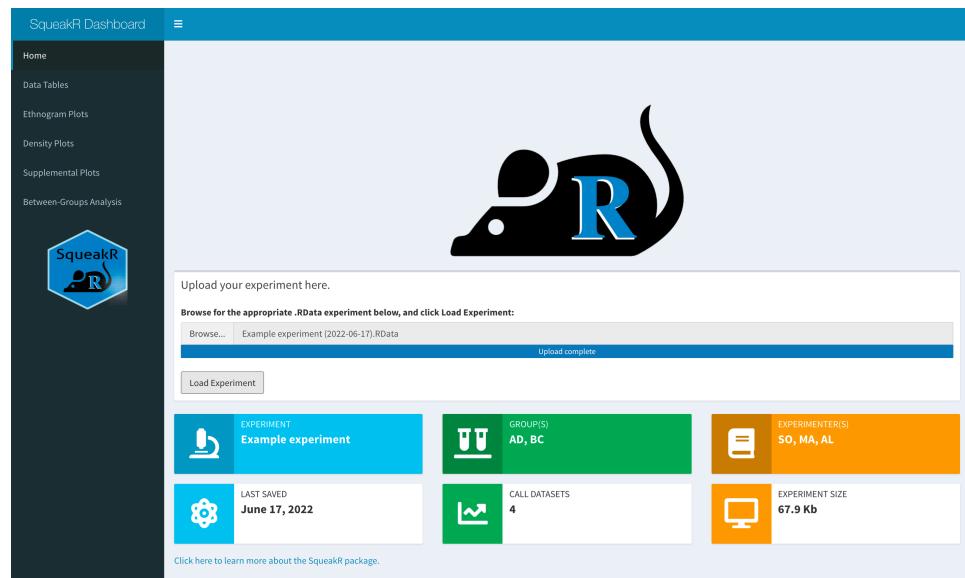
### SQUEAKR ON SWIRL

SqueakR can be learned on Swirl! Learn more about using Swirl to interactively learn about SqueakR directly from the RStudio console on the [SqueakR on Swirl](#) repository.

Alternatively, you can check the [SqueakR website](#) for extended documentation.

## SqueakR Dashboard

A Shiny dashboard for visualizing and conducting data analysis without any code!



# SqueakR :: CHEAT SHEET



## SqueakR :: Workflow

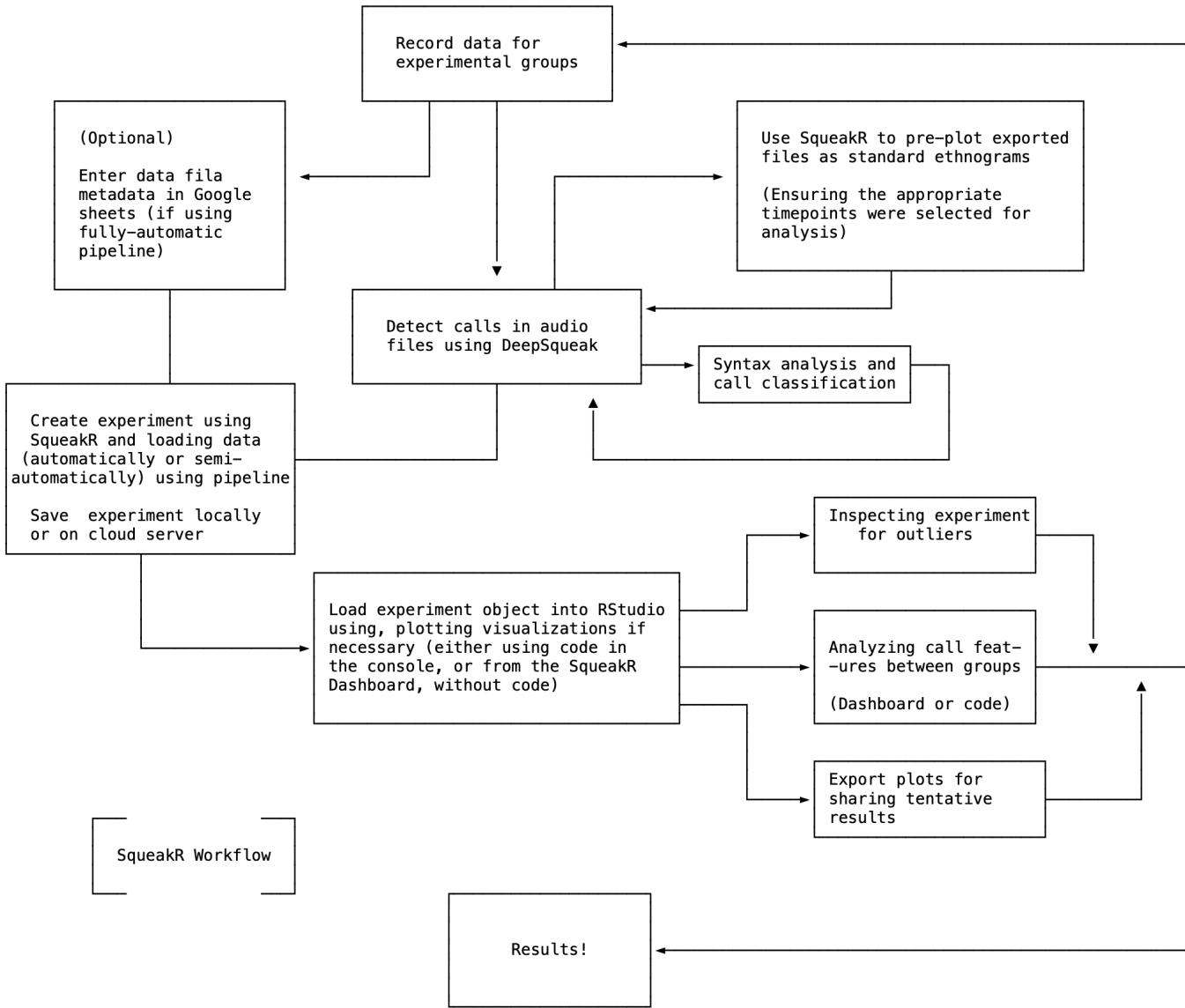
How to use the SqueakR Dashboard for your experiment:

1. Collect data using a recording software capable of making **.WAV**, **.FLAC**, or **.UVF** audio files.
2. **Detect calls** in the audio files using DeepSqueak (the wiki on how to use the software can be found [here](#)). Analysis can be based on either automated or manual review.
3. **Export the detected call files** as Excel documents to a folder containing all of your experimental data.
4. **Run a SqueakR pipeline** to semi-automatically or fully-automatically generate your experiment .RData file.
5. **Load** your experiment into the **SqueakR Dashboard** to visualize and analyze your experiment in a Shiny interface, without any coding required. Data can also be plotted directly into RStudio without the interface, using SqueakR functions.

| Name              | Type                         | Value                |
|-------------------|------------------------------|----------------------|
| experiment        | list [5]                     | List of length 5     |
| name              | character [1]                | 'Example experiment' |
| last_saved        | double (S3: POSIXct, POSIXt) | 2022-06-17 10:45:14  |
| groups            | character [2]                | 'AD' 'BC'            |
| experimenters     | character [3]                | 'SO' 'MA' 'AL'       |
| experimental_data | list [4]                     | List of length 4     |
| call_data         | list [14]                    | List of length 14    |
| call_data         | list [14]                    | List of length 14    |
| call_data         | list [14]                    | List of length 14    |
| call_data         | list [14]                    | List of length 14    |

```
> describe_experiment(experiment)
Experimental name: Example experiment
Last saved: 2022-06-17 10:45:14
Experimenters: SO, MA, AL
Experimental groups: AD, BC
Total call datapoints: 4
Data for AD: 2
Data for BC: 2
>
```

## Workflow Diagram



## SqueakR Website

<https://osimon81.github.io/SqueakR>

# SqueakR :: CHEAT SHEET



## Adding new data

1. Load an Excel file using:

```
my_file <-  
  add_timepoint_data(data_path =  
    "path_to_data", t1 = 2, t2 = 12)
```

2. Score the loaded dataset using:

```
my_file <-  
  score_timepoint_data(data_subset =  
    my_file, group = "Drug",  
    experimenter = "My Name")
```

3. Add the processed dataset to your experiment object:

```
experiment <-  
  add_to_experiment(my_file)
```

Save the experiment to a specified directory:

```
experiment <-  
  save_experiment(experiment =  
    experiment, save_path = "save-dir")
```

## Experiment.RData

Your experiment object will be saved to a specified directory using the `save_experiment()` function.

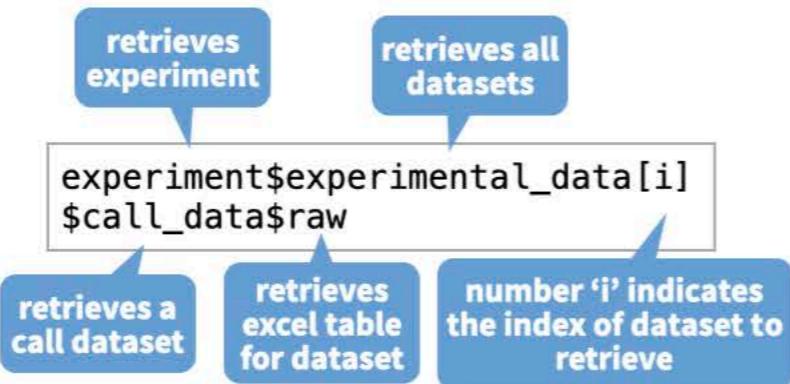
The file will be saved as an .RData file, and can be loaded into R at any point to continue analysis, or add or remove more data.

Experiment object files can also be loaded into the **SqueakR Dashboard** for visualization.

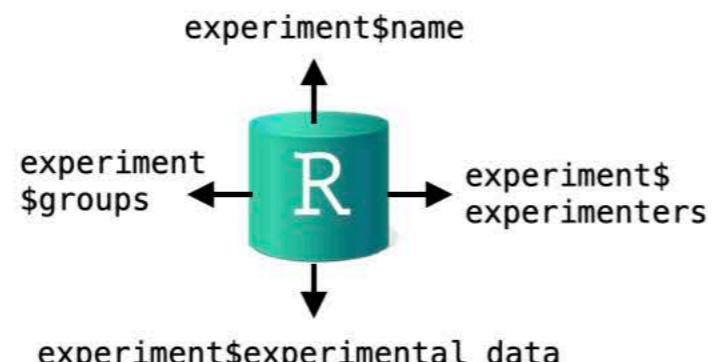
## Managing Experiment

### CODE

How to navigate the experiment object:



### EXPERIMENT OBJECT



### EXPERIMENT CONTENTS

| item                           | description  |
|--------------------------------|--|
| <code>name</code>              | the name of the experiment                         |
| <code>last_saved</code>        | the date the object was last saved                 |
| <code>groups</code>            | the experimental groups                            |
| <code>experimenters</code>     | the experimenters who collected data               |
| <code>experimental_data</code> | the full dataset of scored data for the experiment |

## SqueakR Pipeline(s)

```
experiment <- semisqueakRpipeline()
```

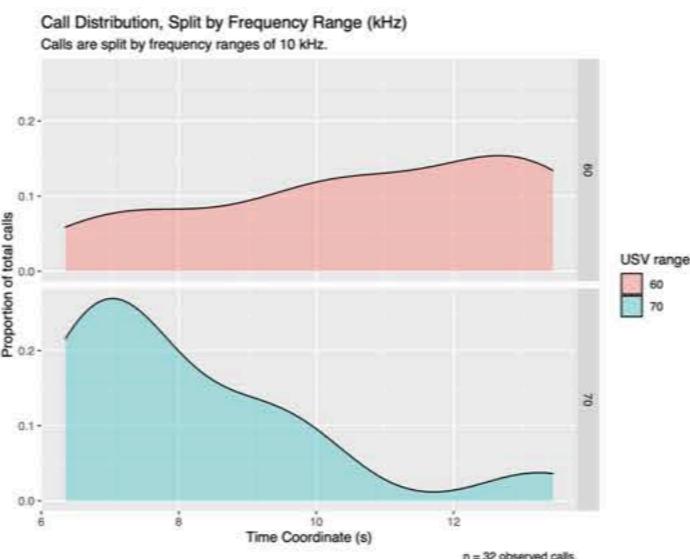
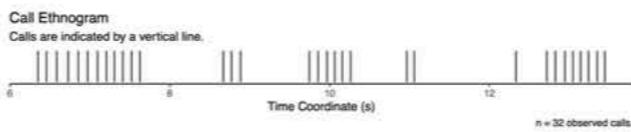
**creates an experiment by looping through a folder of DeepSqueak data, prompting the user for each file's metadata**

```
experiment <- autosqueakRpipeline()
```

**creates an experiment by looping through a folder of DeepSqueak data, pulling metadata from a Google Sheet**

## Visualizations

Graphs which can be generated using SqueakR:



## Learning SqueakR

### DOWNLOAD AND INSTALLATION

To install the package, run the following in RStudio:

```
install.packages("SqueakR")
library(SqueakR)
```

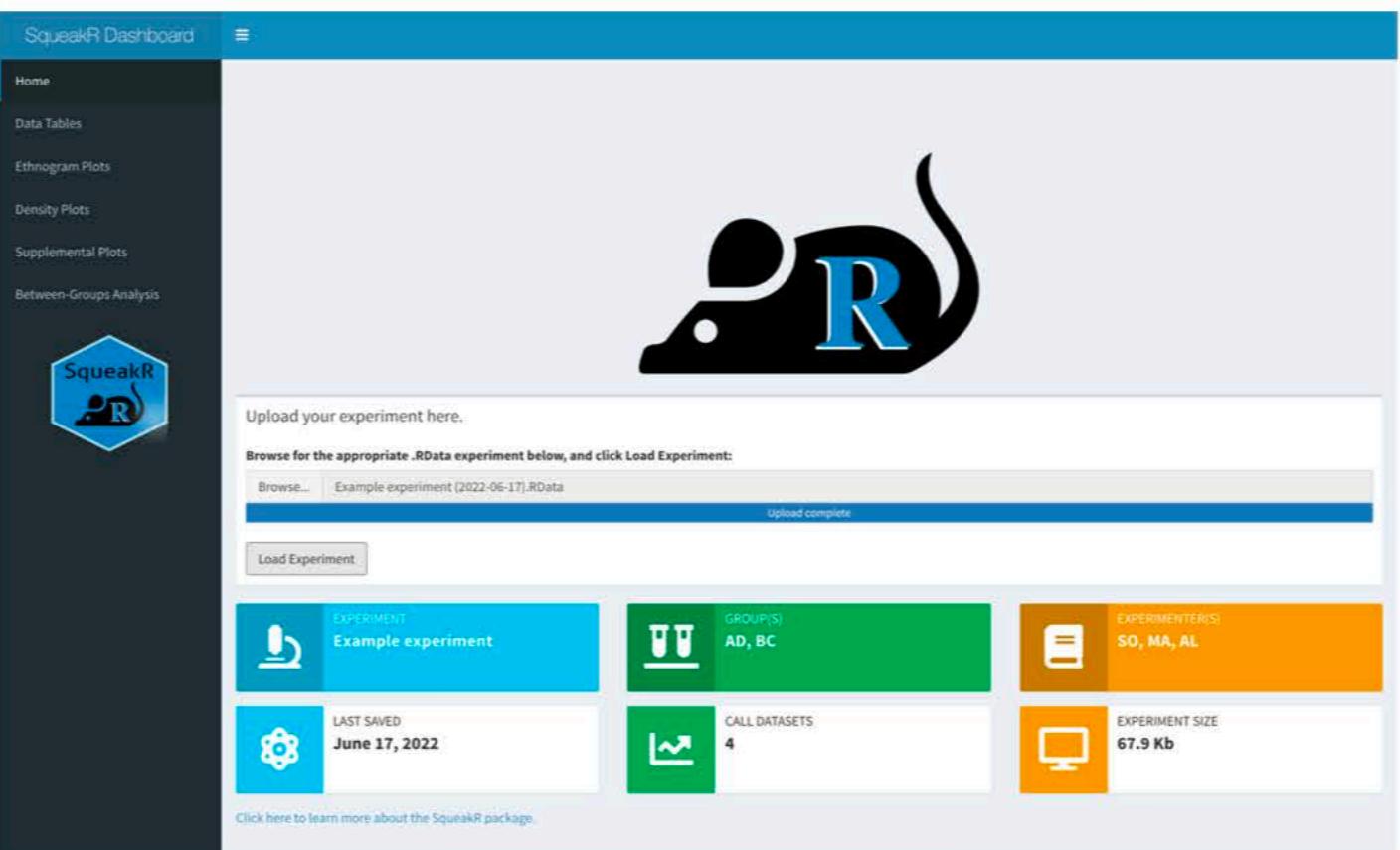
### SQUEAKR ON SWIRL

SqueakR can be learned on Swirl! Learn more about using Swirl to interactively learn about SqueakR directly from the RStudio console on the [SqueakR on Swirl](#) repository.

Alternatively, you can check the [SqueakR website](#) for extended documentation.

## SqueakR Dashboard

A Shiny dashboard for visualizing and conducting data analysis without any code!



# Stata to R :: CHEAT SHEET



## Introduction

This cheat sheet summarizes common Stata commands for econometric analysis and provides their equivalent expression in R.

References for importing/cleaning data, manipulating variables, and other basic commands include Hanck et al. (2019), [Econometrics with R](#), and Wickham and Grolemund (2017), [R for Data Science](#).

Example data comes from Wooldridge *Introductory Econometrics: A Modern Approach*. Download Stata data sets [here](#). R data sets can be accessed by installing the [`wooldridge` package](#) from CRAN.

All R commands written in base R, unless otherwise noted.

## Setup

Note: While it is common to create a `log` file in Stata to store the commands and output of Stata sessions, the equivalent does not exist in R. A more savvy version in R is to create a [R-markdown](#) file to capture code and output.

```
ssc install outreg2 // install `outreg2` package. Note: unlike R packages, Stata packages do not have to be loaded each time once installed.
```

```
install.packages("wooldridge") # install `wooldridge` package  
  
data(package = "wooldridge") # list datasets in `wooldridge` package  
  
load(wage1) # load `wage1` dataset into session  
  
?wage1 # consult documentation on `wage1` dataset
```

## Basic plots

example data: `wage1`

```
hist(wage) // histogram of `wage`  
hist(wage), by(nonwhite) //  
scatter(wage educ) // scatter plot of `wage` by `educ`  
twoway (scatter wage educ) (lfit wage educ) // scatter plot with fitted line  
graph box wage, by(nonwhite) // boxplot of wage by `nonwhite`
```

## Summarize Data

example data: `wage1`

Where Stata only allows one to work with one data set at a time, multiple data sets can be loaded into the R environment simultaneously, and hence must be specified with each function call. Note: R does not have an equivalent to Stata's `codebook` command.

```
browse // open browser for loaded data  
  
describe // describe structure of loaded data  
summarize // display summary statistics for all variables in dataset  
list in 1/6 // display first 6 rows  
  
tabulate educ // tabulate `educ` variable frequencies  
tabulate educ female // cross-tabulate `educ` and `female` frequencies
```

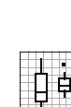


```
view(wage1) # open browser for loaded `wage1` data  
  
str(wage1) # describe structure of `wage1` data  
summary(wage1) # display summary statistics for `wage1` variables  
head(wage1) # display first 6 (default) rows data  
tail(wage1) # display last 6 rows  
  
table(wage1$educ) #tabulate `educ` frequencies  
table("yrs_edu" = wage1$educ, "female" = wage1$female) # tabulate `educ` frequencies name table columns
```

**Tip:** The {AER} package will automatically load other useful dependent packages, including: {car}, {lmtest}, {sandwich} which are used for many of the commands listed in this cheat sheet.



```
hist(wage1$wage) # histogram of `wage`  
  
{  
  plot(y = wage1$wage, x = wage1$educ) # scatter plot  
  abline(lm(wage1$wage~wage1$educ), col="red") # add fitted line to scatterplot  
  
  boxplot(wage1$wage~wage1$nonwhite) # boxplot of `wage` by `nonwhite`
```



## Estimate Models, 1/2

### OLS

example data: `wage1`

```
reg wage educ // simple regression of `wage` by `educ` (Results printed automatically).
```

```
reg wage educ if nonwhite==1 // add condition with if statement
```

```
reg wage educ exper, robust // multiple regression using HC1 robust standard errors
```

```
reg wage educ exper, cluster(numdep) // use clustered standard errors
```

**Tip:** An alternate way to compute robust standard errors in R for any models not covered by {estimatr} package is load the {AER} package and run:

```
coeftest(mod1, vcov. = vcovHC, type = "HC1")
```

### MLE (Logit/Probit/Tobit)

example data: `mroz`

```
logit inlf nwifeinc educ // estimate logistic regression
```

```
probit inlf nwifeinc educ // estimate logistic regression
```

```
tobit hours nwifeinc educ, ll(0) // estimate tobit regression, lower-limit of y censored at zero
```



```
mod1 <- lm(wage ~ educ, data = wage1) # simple regression of `wage` by `educ`, store results in mod1  
summary(mod1) # print summary of `mod1` results
```

```
mod2 <- lm(wage ~ educ, data = wage1[wage1$nonwhite==1, ]) # add condition with if statement
```

```
mod3 <- estimatr::lm_robust(wage ~ educ + exper, data = wage1, se_type = "stata") # multiple regression with HC1 (Stata default) robust standard errors, use {estimatr} package
```

```
mod4 <- estimatr::lm_robust(wage ~ educ + exper, data = wage1, clusters = numdep) # use clustered standard errors.
```



```
mod_log <- glm(inlf~nwifeinc + educ + family=binomial(link="logit"), data=mroz) # estimate logistic regression
```

```
mod_pro <- glm(inlf~nwifeinc + educ + family=binomial(link="probit"), data=mroz) # estimate logistic regression
```

```
mod_tob <- AER::tobit(hours ~ nwifeinc + educ, left = 0, data = mroz) # estimate tobit regression, lower-limit of y censored at zero, use {AER} package
```

## Postestimation, 1/2

example data: `wage1`

Note: Postestimation commands in Stata apply to the most recently run estimation commands.

```
reg wage educ // estimation used for the following post-estimation commands
```

```
predict yhat // get predicted values from last estimation, store as `yhat`
```

```
predict e, res // get residuals from last estimation, store as `e`
```



```
mod1 <- lm(wage ~ educ, data = wage1) # estimation used for the following post-estimation commands  
yhat <- predict(mod1) # get predicted values
```

```
e <- residuals(mod1) # get residual values
```



## Create/Edit Variables

*example data: `wage1`*

Note: where Stata only allows one to work with one data set at a time, multiple data sets can be loaded into the R environment simultaneously, hence the data set must be specified for each command.

```
gen exper2 = exper^2 // create
`exper` squared variable
egen wage_avg = mean(wage) // create
average wage variable
```

```
drop tenursq // drop `tenursq`
variable
```

```
keep wage educ exper nonwhite // keep
selected variables
```

```
tab numdep, gen(numdep) // create
dummy variables for `numdep`
```

```
recode exper (1/20 = 1 "1 to 20
years") (21/40 = 2 "21 to 40 years")
(41/max = 3 "41+ years"),
gen(experlvl) // recode `exper` and
gen new variable
```

```
wage1$exper2 <- wage1$exper^2 # 
create `exper` squared variable
wage1$wage_avg <- mean(wage1$wage) #
create average wage variable
```

```
wage1$tenursq <- NULL #drop `tenursq`
```

```
wage1 <- wage1[, c("wage", "educ",
"exper", "nonwhite")] # keep selected
variables
```

```
wage1 <-
fastDummies::dummy_cols(wage1,
select_columns = "numdep") # create
dummy variables for `numdep`, use
{fastDummies} package
```

```
{ wage1$experlvl <- 3 # recode `exper`#
wage1$experlvl[wage1$exper < 41] <- 2
wage1$experlvl[wage1$exper < 21] <- 1
```

## Statistical tests / diagnostics

```
reg lwage educ exper // estimation
used for examples below
estat hettest // Breusch-Pagan /
Cook-Weisberg test for
heteroskedasticity
estat ovtest // Ramsey RESET test
for omitted variables
ttest wage, by(nonwhite) //
independent group t-test, compare
means of same variable between
groups
```

```
example data: `wage1`#
mod <- lm(lwage ~ educ exper, data =
wage1) # estimate used for examples
below
lmtest::bptest(mod) # Breusch-Pagan
/ Cook-Weisberg test for hetero-
skedasticity using the {lmtest}
package
lmtest::resettest(mod) # Ramsey
RESET test
t.test(wage ~ nonwhite, data =
wage1) # independent group t-test
```

## Interactions, categorical/continuous variables

In Stata, it is common to use special operators to specify the treatment of variables as continuous (`c.`) or categorical(`i.`). Similarly, the `#` operator denotes different ways to return the interaction of those variables. Here we show some common uses of these operators as well as their R equivalents.

```
reg lwage i.numdep // treat
`numdep` as a factor variable
reg lwage c.educ#c.exper // return
interaction term only
reg lwage c.educ##c.exper // return
full factorial specification
reg lwage c.exper##i.numdep // 
return full, interact continuous
and categorical
```

```
lm(lwage ~ as.factor(numdep), data
= wage1) # treat `numdep` as factor
lm(lwage ~ educ:exper, data =
wage1) # return interaction term
only
lm(lwage ~ educ*exper, data =
wage1) # return full factorial
specification
lm(wage ~ exper*as.factor(numdep),
data = wage1) # return full,
interact continuous and categorical
```

## Estimate Models, 2/2

### Panel/Longitudinal

*example data: `murder`*

```
xtset id year // set `id` as
entities (panel) and `year` as
time variable
xtdescribe // describe pattern of
xt data
xtsum // summarize xt data
xtreg mrd rte unem, fe // fixed
effects regression
```



```
plm::is.pbalanced(murder$id,
murder$year) # check panel balance
with {plm} package
modfe <- plm::plm(mrd rte ~ unem,
index = c("id", "year"), model =
"within", data = murder) # estimate
fixed effects ("within") model
summary(modfe) # display results
```

### Instrumental Variables (2SLS)

*example data: `mroz`*

```
ivreg lwage (educ = fatheduc),
first // show results of first
stage regression
est first // test IV and
endogenous variable
ivreg lwage(educ = fatheduc) //
show results of 2SLS directly
```



```
modiv <- AER::ivreg(lwage ~ educ |
fatheduc, data = mroz) # estimate
2SLS with {AER} package
summary(modiv, diagnostics = TRUE)
# get diagnostic tests of IV and
endogenous variable
```

## Post-estimation, 2/2

*example data: `wage1`*

Note: Postestimation commands in Stata apply to the most recently run estimation commands.

```
reg lwage educ exper##exper // 
estimation used for following post-
estimation commands
estimates store mod1 // stores in
memory the last estimation results
to `mod1`
```

```
margins // get average predictive
margins
margins, dydx(*) // get average
marginal effects for all variables
marginsplot // plot marginal
effects
```

```
margins, dydx(exper) // average
marginal effects of experience
margins, at(exper=(1(10)51)) //
average predictive margins over
`exper` range at 10-year increments
```

```
estimates use mod1 // loads `mod1` 
back into working memory
estimates table mod1 mod2 // 
display table with stored
estimation results
```



```
margins::prediction(mod1) # get
average predictive margins with
{margins} package
m1 <- margins::margins(mod1) # get
average marginal effects for all
variables
plot(m) # plot marginal effects
```

```
summary(m) # get detailed summary of
marginal effects
margins::prediction(mod1, at =
list(exper = seq(1,51,10))) #
predictive margins over `exper` range
at 10-year increments
```

```
stargazer::stargazer(mod1, mod2, type
= "text") # use {stargazer} package,
with `type=text` to display results
within R. Note: `type` also can be
changed for LaTeX and HTML output.
```



# String manipulation with stringr :: CHEATSHEET



The **stringr** package provides a set of internally consistent tools for working with character strings, i.e. sequences of characters surrounded by quotation marks.

## Detect Matches

|  |  |
|--|--|
|  | <b>str_detect(string, pattern, negate = FALSE)</b><br>Detect the presence of a pattern match in a string. Also <b>str_like()</b> . str_detect(fruit, "a")                  |
|  | <b>str_starts(string, pattern, negate = FALSE)</b><br>Detect the presence of a pattern match at the beginning of a string. Also <b>str_ends()</b> . str_starts(fruit, "a") |
|  | <b>str_which(string, pattern, negate = FALSE)</b><br>Find the indexes of strings that contain a pattern match. str_which(fruit, "a")                                       |
|  | <b>str_locate(string, pattern)</b> Locate the positions of pattern matches in a string. Also <b>str_locate_all()</b> . str_locate(fruit, "a")                              |
|  | <b>str_count(string, pattern)</b> Count the number of matches in a string. str_count(fruit, "a")   |

## Subset Strings

|  |  |
|--|--|
|  | <b>str_sub(string, start = 1L, end = -1L)</b> Extract substrings from a character vector. str_sub(fruit, 1, 3); str_sub(fruit, -2)   |
|  | <b>str_subset(string, pattern, negate = FALSE)</b> Return only the strings that contain a pattern match. str_subset(fruit, "p")  |
|  | <b>str_extract(string, pattern)</b> Return the first pattern match found in each string, as a vector. Also <b>str_extract_all()</b> to return every pattern match. str_extract(fruit, "[aeiou]")                     |
|  | <b>str_match(string, pattern)</b> Return the first pattern match found in each string, as a matrix with a column for each () group in pattern. Also <b>str_match_all()</b> . str_match(sentences, "(a the) ([^ +])") |

## Manage Lengths

|  |  |
|--|--|
|  | <b>str_length(string)</b> The width of strings (i.e. number of code points, which generally equals the number of characters). str_length(fruit)                                |
|  | <b>str_pad(string, width, side = c("left", "right", "both"), pad = " ")</b> Pad strings to constant width. str_pad(fruit, 17)  |
|  | <b>str_trunc(string, width, side = c("right", "left", "center"), ellipsis = "...")</b> Truncate the width of strings, replacing content with ellipsis. str_trunc(sentences, 6) |
|  | <b>str_trim(string, side = c("both", "left", "right"))</b> Trim whitespace from the start and/or end of a string. str_trim(str_pad(fruit, 17))                                 |
|  | <b>str_squish(string)</b> Trim whitespace from each end and collapse multiple spaces into single spaces. str_squish(str_pad(fruit, 17, "both"))                                |

## Mutate Strings

|  |   |
|--|---|
|  | <b>str_sub()</b> <- value. Replace substrings by identifying the substrings with str_sub() and assigning into the results. str_sub(fruit, 1, 3) <- "str"          |
|  | <b>str_replace(string, pattern, replacement)</b> Replace the first matched pattern in each string. Also <b>str_remove()</b> . str_replace(fruit, "p", "-")        |
|  | <b>str_replace_all(string, pattern, replacement)</b> Replace all matched patterns in each string. Also <b>str_remove_all()</b> . str_replace_all(fruit, "p", "-") |
|  | <b>str_to_lower(string, locale = "en")<sup>1</sup></b> Convert strings to lower case. str_to_lower(sentences)   |
|  | <b>str_to_upper(string, locale = "en")<sup>1</sup></b> Convert strings to upper case. str_to_upper(sentences)   |
|  | <b>str_to_title(string, locale = "en")<sup>1</sup></b> Convert strings to title case. Also <b>str_to_sentence()</b> . str_to_title(sentences)                     |

## Join and Split

|  |   |
|--|---|
|  | <b>str_c(..., sep = "", collapse = NULL)</b> Join multiple strings into a single string. str_c(letters, LETTERS)  |
|  | <b>str_flatten(string, collapse = "")</b> Combines into a single string, separated by collapse. str_flatten(fruit, ",")   |
|  | <b>str_dup(string, times)</b> Repeat strings times times. Also <b>str_unique()</b> to remove duplicates. str_dup(fruit, times = 2)  |
|  | <b>str_split_fixed(string, pattern, n)</b> Split a vector of strings into a matrix of substrings (splitting at occurrences of a pattern match). Also <b>str_split()</b> to return a list of substrings and <b>str_split_i()</b> to return the ith substring. str_split_fixed(sentences, " ", n=3) |
|  | <b>str_glue(..., .sep = "", .envir = parent.frame())</b> Create a string from strings and {expressions} to evaluate. str_glue("Pi is {pi}")   |
|  | <b>str_glue_data(.x, ..., .sep = "", .envir = parent.frame(), .na = "NA")</b> Use a data frame, list, or environment to create a string from strings and {expressions} to evaluate. str_glue_data(mtcars, "[rownames(mtcars)] has {hp} hp")   |

## Order Strings

|  |  |
|--|--|
|  | <b>str_order(x, decreasing = FALSE, na_last = TRUE, locale = "en", numeric = FALSE, ...)<sup>1</sup></b> Return the vector of indexes that sorts a character vector. fruit[str_order(fruit)] |
|  | <b>str_sort(x, decreasing = FALSE, na_last = TRUE, locale = "en", numeric = FALSE, ...)<sup>1</sup></b> Sort a character vector. str_sort(fruit)   |

## Helpers

|  |  |
|--|--|
|  | <b>str_conv(string, encoding)</b> Override the encoding of a string. str_conv(fruit, "ISO-8859-1")   |
|  | <b>str_view(string, pattern, match = NA)</b> View HTML rendering of all regex matches. str_view(sentences, "[aeiou])")                                   |
|  | <b>str_equal(x, y, locale = "en", ignore_case = FALSE, ...)<sup>1</sup></b> Determine if two strings are equivalent. str_equal(c("a", "b"), c("a", "c")) |
|  | <b>str_wrap(string, width = 80, indent = 0, exdent = 0)</b> Wrap strings into nicely formatted paragraphs. str_wrap(sentences, 20)                       |

<sup>1</sup> See [bit.ly/ISO639-1](https://bit.ly/ISO639-1) for a complete list of locales.

# Need to Know

Pattern arguments in string are interpreted as regular expressions *after any special characters have been parsed*.

In R, you write regular expressions as *strings*, sequences of characters surrounded by quotes ("") or single quotes('').

Some characters cannot be represented directly in an R string. These must be represented as **special characters**, sequences of characters that have a specific meaning., e.g.

| Special Character | Represents |
|-------------------|------------|
| \\\               | \          |
| '"                | "          |
| \n                | new line   |

Run `?"""` to see a complete list

Because of this, whenever a \ appears in a regular expression, you must write it as \\ in the string that represents the regular expression.

Use `writeLines()` to see how R views your string after all special characters have been parsed.

```
writeLines("|\.")  
# \.
```

```
writeLines("\\| is a backslash")  
# \| is a backslash
```

## INTERPRETATION

Patterns in stringr are interpreted as regexs. To change this default, wrap the pattern in one of:

`regex(pattern, ignore_case = FALSE, multiline = FALSE, comments = FALSE, dotall = FALSE, ...)`  
Modifies a regex to ignore cases, match end of lines as well of end of strings, allow R comments within regex's , and/or to have . match everything including \n.  
`str_detect("i", regex("i", TRUE))`

`fixed()` Matches raw bytes but will miss some characters that can be represented in multiple ways (fast). `str_detect("\u0130", fixed("i"))`

`coll()` Matches raw bytes and will use locale specific collation rules to recognize characters that can be represented in multiple ways (slow). `str_detect("\u0130", coll("i", TRUE, locale = "tr"))`

`boundary()` Matches boundaries between characters, line\_breaks, sentences, or words. `str_split(sentences, boundary("word"))`

# Regular Expressions -

Regular expressions, or *regexp*s, are a concise language for describing patterns in strings.

## MATCH CHARACTERS

| string<br>(type this)  | regexp<br>(to mean this) | matches<br>(which matches this)            | example           |
|------------------------|--------------------------|--|-------------------|
| a (etc.)               | a (etc.)                 | a (etc.)                                   | see("a")          |
| \.                     | \.                       | .  | see("\.")         |
| \!                     | \!                       | !  | see("\!")         |
| \?                     | \?                       | ?  | see("\?")         |
| \\\                    | \\\                      | \  | see("\\\\")       |
| \(                     | \(                       | (  | see("\()")        |
| \)                     | \)                       | )  | see("\)")         |
| \{                     | \{                       | {  | see("\{")         |
| \}                     | \}                       | }  | see("\}")         |
| \n                     | \n                       | new line (return)                          | see("\n")         |
| \t                     | \t                       | tab  | see("\t")         |
| \s                     | \s                       | any whitespace (\\$ for non-whitespaces)   | see("\s")         |
| \d                     | \d                       | any digit (\D for non-digits)              | see("\d")         |
| \w                     | \w                       | any word character (\W for non-word chars) | see("\w")         |
| \b                     | \b                       | word boundaries                            | see("\b")         |
| [:digit:] <sup>1</sup> | [:digit:] <sup>1</sup>   | digits                                     | see("[ :digit:]") |
| [:alpha:] <sup>1</sup> | [:alpha:] <sup>1</sup>   | letters                                    | see("[ :alpha:]") |
| [:lower:] <sup>1</sup> | [:lower:] <sup>1</sup>   | lowercase letters                          | see("[ :lower:]") |
| [:upper:] <sup>1</sup> | [:upper:] <sup>1</sup>   | uppercase letters                          | see("[ :upper:]") |
| [:alnum:] <sup>1</sup> | [:alnum:] <sup>1</sup>   | letters and numbers                        | see("[ :alnum:]") |
| [:punct:] <sup>1</sup> | [:punct:] <sup>1</sup>   | punctuation                                | see("[ :punct:]") |
| [:graph:] <sup>1</sup> | [:graph:] <sup>1</sup>   | letters, numbers, and punctuation          | see("[ :graph:]") |
| [:space:] <sup>1</sup> | [:space:] <sup>1</sup>   | space characters (i.e. \s)                 | see("[ :space:]") |
| [:blank:] <sup>1</sup> | [:blank:] <sup>1</sup>   | space and tab (but not new line)           | see("[ :blank:]") |
| .                      | .                        | every character except a new line          | see(".")          |

<sup>1</sup> Many base R functions require classes to be wrapped in a second set of [ ], e.g. [[:digit:]]

## ALTERNATES

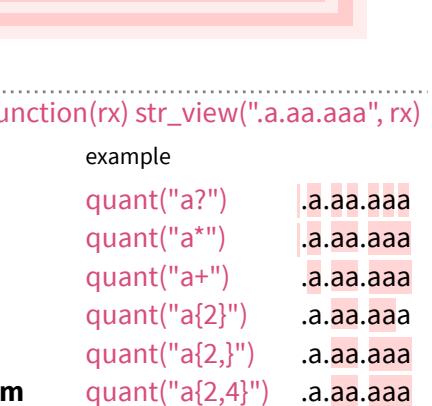
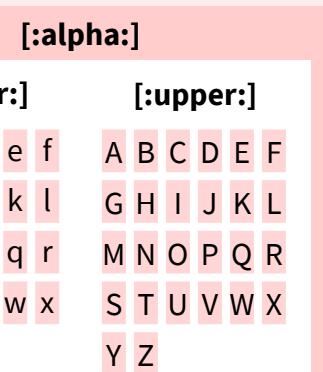
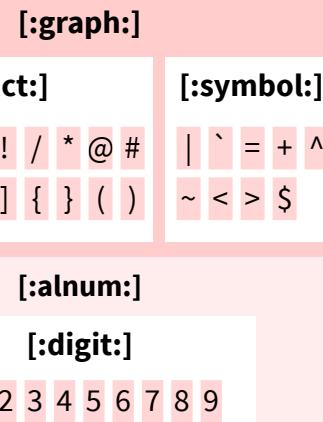
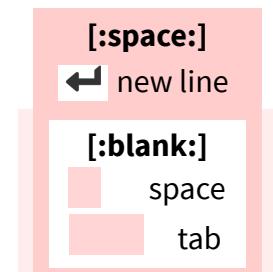
| regexp | matches      | example       |
|--------|--------------|---------------|
| ab d   | or           | alt("ab d")   |
| [abe]  | one of       | alt("[abe]")  |
| [^abe] | anything but | alt("[^abe]") |
| [a-c]  | range        | alt("[a-c]")  |

## ANCHORS

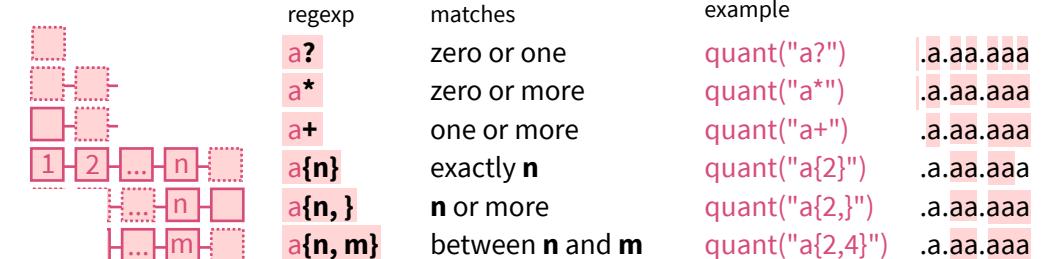
| regexp | matches         | example       |
|--------|-----------------|---------------|
| ^a     | start of string | anchor("^a")  |
| a\$    | end of string   | anchor("a\$") |

## LOOK AROUNDS

| regexp  | matches         | example         |
|---------|-----------------|-----------------|
| a(?=c)  | followed by     | look("a(?=c)")  |
| a(?!c)  | not followed by | look("a(?!c)")  |
| (?<=b)a | preceded by     | look("(?<=b)a") |
| (?<!b)a | not preceded by | look("(?<!b)a") |



## QUANTIFIERS



## GROUPS

Use parentheses to set precedent (order of evaluation) and create groups

| regexp  | matches         | example        |
|---------|-----------------|----------------|
| (ab d)e | sets precedence | alt("(ab d)e") |

Use an escaped number to refer to and duplicate parentheses groups that occur earlier in a pattern. Refer to each group by its order of appearance

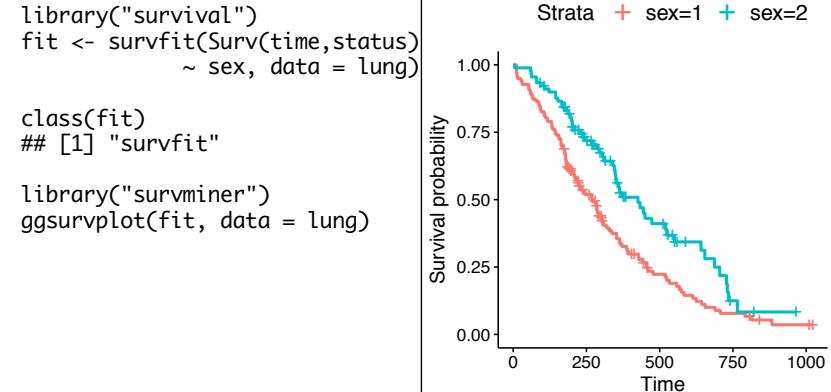
| string<br>(type this) | regexp<br>(to mean this) | matches              | example             |
|-----------------------|--------------------------|----------------------|---------------------|
| \1                    | \1 (etc.)                | first () group, etc. | ref("(a)(b)\1\2\1") |

# Creating Survival Plots

## Informative and Elegant with survminer

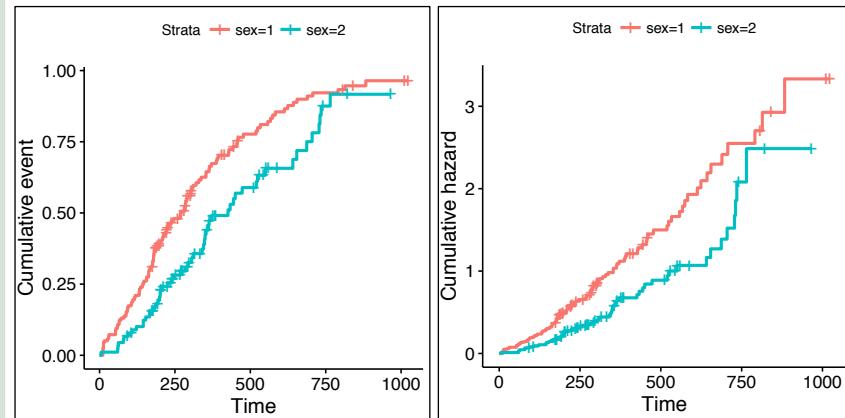
### Survival Curves

The **ggsurvplot()** function creates **ggplot2** plots from **survfit** objects.



Use the **fun** argument to set the transformation of the survival curve. E.g. "**event**" for cumulative events, "**cumhaz**" for the cumulative hazard function or "**pct**" for survival probability in percentage.

```
ggsurvplot(fit, data = lung, fun = "event")
ggsurvplot(fit, data = lung, fun = "cumhaz")
```



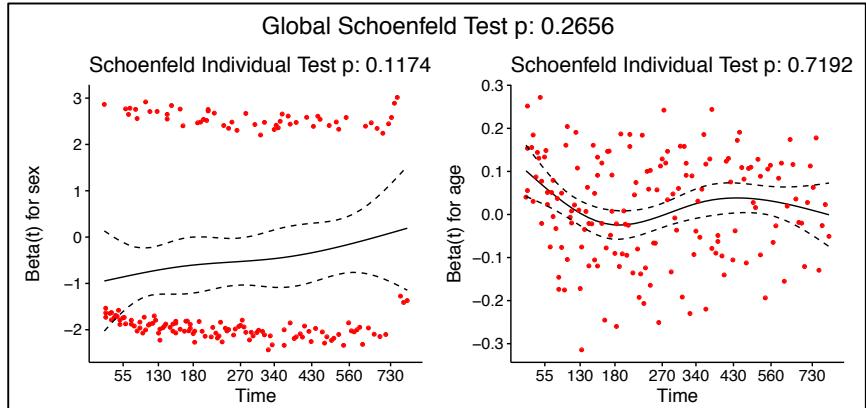
With lots of graphical parameters you have full control over look and feel of the survival plots; position and content of the legend; additional annotations like p-value, title, subtitle.

```
ggsurvplot(fit, data = lung,
conf.int = TRUE,
pval = TRUE,
fun = "pct",
risk.table = TRUE,
size = 1,
linetype = "strata",
palette = c("#E7B800",
"#2E9DFD"),
legend = "bottom",
legend.title = "Sex",
legend.labs = c("Male",
"Female"))
```

### Diagnostics of Cox Model

The function **cox.zph()** from **survival** package may be used to test the proportional hazards assumption for a Cox regression model fit. The graphical verification of this assumption may be performed with the function **ggcooxzph()** from the **survminer** package. For each covariate it produces plots with scaled Schoenfeld residuals against the time.

```
library("survival")
fit <- coxph(Surv(time, status) ~ sex + age, data = lung)
ftest <- cox.zph(fit)
ftest
##          rho chisq      p
## sex     0.1236 2.452 0.117
## age    -0.0275 0.129 0.719
## GLOBAL   NA 2.651 0.266
library("survminer")
ggcooxzph(ftest)
```



The function **ggcoxdiagnostics()** plots different types of residuals as a function of time, linear predictor or observation id. The type of residual is selected with **type** argument. Possible values are "martingale", "deviance", "score", "schoenfeld", "dfbeta", "dfbetas", and "scaledsch".

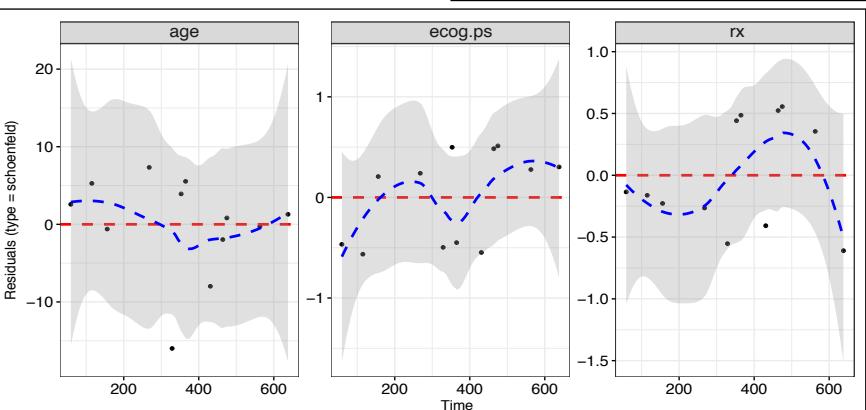
The **ox.scale** argument defines what shall be plotted on the OX axis. Possible values are "linear.predictions", "observation.id", "time".

Logical arguments **hline** and **sline** may be used to add horizontal line or smooth line to the plot.

```
library("survival")
library("survminer")
fit <- coxph(Surv(time, status) ~ sex + age, data = lung)
```

```
ggcoxdiagnostics(fit,
type = "deviance",
ox.scale = "linear.predictions")
```

```
ggcoxdiagnostics(fit,
type = "schoenfeld",
ox.scale = "time")
```



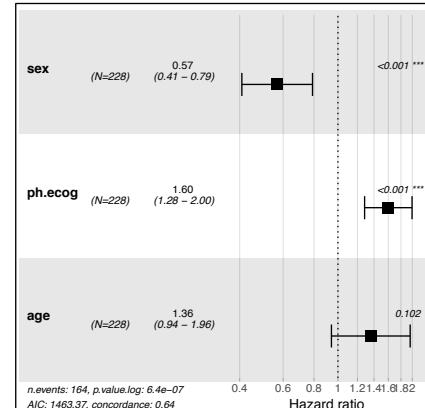
### Summary of Cox Model

The function **ggforest()** from the **survminer** package creates a forest plot for a Cox regression model fit. Hazard ratio estimates along with confidence intervals and p-values are plotted for each variable.

```
library("survival")
library("survminer")
lung$age <- ifelse(lung$age > 70, ">70", "<= 70")
fit <- coxph( Surv(time, status) ~ sex + ph.ecog + age, data = lung)
fit
```

```
## Call:
## coxph(formula = Surv(time, status) ~ sex+ph.ecog+age, data=lung)
##
##            coef exp(coef) se(coef)      z      p
## sex     -0.567    0.567    0.168 -3.37 0.00075
## ph.ecog  0.470    1.600    0.113  4.16 3.1e-05
## age>70   0.307    1.359    0.187  1.64 0.10175
##
## Likelihood ratio test=31.6 on
## n= 227, number of events= 164
```

```
ggforest(fit)
```



The function **ggadjustedcurves()** from the **survminer** package plots Adjusted Survival Curves for Cox Proportional Hazards Model. Adjusted Survival Curves show how a selected factor influences survival estimated from a Cox model.

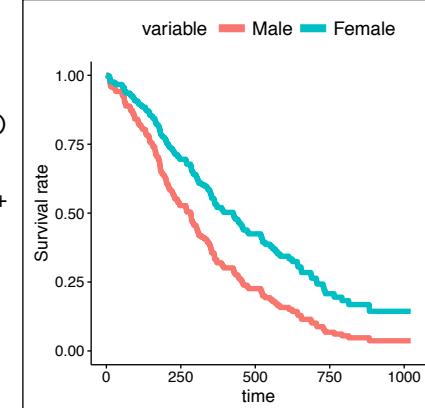
Note that these curves differ from Kaplan Meier estimates since they present expected survival based on given Cox model.

```
library("survival")
library("survminer")
```

```
lung$sex <- ifelse(lung$sex == 1,
"Male", "Female")
```

```
fit <- coxph(Surv(time, status) ~
sex + ph.ecog + age +
strata(sex),
data = lung)
```

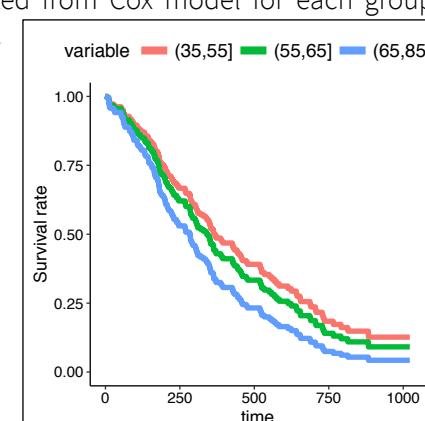
```
ggadjustedcurves(fit, data=lung)
```



Note that it is not necessary to include the grouping factor in the Cox model. Survival curves are estimated from Cox model for each group defined by the factor independently.

```
lung$age3 <- cut(lung$age,
c(35, 55, 65, 85))
```

```
ggadjustedcurves(fit, data=lung,
variable="age3")
```



# R Syntax Comparison :: CHEAT SHEET

## Dollar sign syntax

```
goal(data$x, data$y)
```

### SUMMARY STATISTICS:

one continuous variable:

```
mean(mtcars$mpg)
```

one categorical variable:

```
table(mtcars$cyl)
```

two categorical variables:

```
table(mtcars$cyl, mtcars$am)
```

one continuous, one categorical:

```
mean(mtcars$mpg [mtcars$cyl==4])
```

```
mean(mtcars$mpg [mtcars$cyl==6])
```

```
mean(mtcars$mpg [mtcars$cyl==8])
```

### PLOTTING:

one continuous variable:

```
hist(mtcars$disp)
```

```
boxplot(mtcars$disp)
```

one categorical variable:

```
barplot(table(mtcars$cyl))
```

two continuous variables:

```
plot(mtcars$disp, mtcars$mpg)
```

two categorical variables:

```
mosaicplot(table(mtcars$am, mtcars$cyl))
```

one continuous, one categorical:

```
histogram(mtcars$disp[mtcars$cyl==4])
```

```
histogram(mtcars$disp[mtcars$cyl==6])
```

```
histogram(mtcars$disp[mtcars$cyl==8])
```

```
boxplot(mtcars$disp[mtcars$cyl==4])
boxplot(mtcars$disp[mtcars$cyl==6])
boxplot(mtcars$disp[mtcars$cyl==8])
```

### WRANGLING:

subsetting:

```
mtcars[mtcars$mpg>30, ]
```

making a new variable:

```
mtcars$efficient[mtcars$mpg>30] <- TRUE
```

```
mtcars$efficient[mtcars$mpg<30] <- FALSE
```

## Formula syntax

```
goal(y~x|z, data=data, group=w)
```

### SUMMARY STATISTICS:

one continuous variable:

```
mosaic::mean(~mpg, data=mtcars)
```

one categorical variable:

```
mosaic::tally(~cyl, data=mtcars)
```

two categorical variables:

```
mosaic::tally(cyl~am, data=mtcars)
```

one continuous, one categorical:

```
mosaic::mean(mpg~cyl, data=mtcars)
```

tilde

### PLOTTING:

one continuous variable:

```
lattice::histogram(~disp, data=mtcars)
```

```
lattice::bwplot(~disp, data=mtcars)
```

one categorical variable:

```
mosaic::bargraph(~cyl, data=mtcars)
```

two continuous variables:

```
lattice::xyplot(mpg~disp, data=mtcars)
```

two categorical variables:

```
mosaic::bargraph(~am, data=mtcars, group=cyl)
```

one continuous, one categorical:

```
lattice::histogram(~disp|cyl, data=mtcars)
```

```
lattice::bwplot(cyl~disp, data=mtcars)
```

The variety of R syntaxes give you many ways to “say” the same thing

read across the cheatsheet to see how different syntaxes approach the same problem

## Tidyverse syntax

```
data %>% goal(x)
```

### SUMMARY STATISTICS:

one continuous variable:

```
mtcars %>% dplyr::summarize(mean(mpg))
```

one categorical variable:

```
mtcars %>% dplyr::group_by(cyl) %>% dplyr::summarize(n())
```

the pipe

two categorical variables:

```
mtcars %>% dplyr::group_by(cyl, am) %>% dplyr::summarize(n())
```

one continuous, one categorical:

```
mtcars %>% dplyr::group_by(cyl) %>% dplyr::summarize(mean(mpg))
```

### PLOTTING:

one continuous variable:

```
ggplot2::qplot(x=mpg, data=mtcars, geom = "histogram")
```

```
ggplot2::qplot(y=disp, x=1, data=mtcars, geom="boxplot")
```

one categorical variable:

```
ggplot2::qplot(x=cyl, data=mtcars, geom="bar")
```

two continuous variables:

```
ggplot2::qplot(x=disp, y=mpg, data=mtcars, geom="point")
```

two categorical variables:

```
ggplot2::qplot(x=factor(cyl), data=mtcars, geom="bar") + facet_grid(.~am)
```

one continuous, one categorical:

```
ggplot2::qplot(x=disp, data=mtcars, geom = "histogram") + facet_grid(.~cyl)
```

```
ggplot2::qplot(y=disp, x=factor(cyl), data=mtcars, geom="boxplot")
```

### WRANGLING:

subsetting:

```
mtcars %>% dplyr::filter(mpg>30)
```

making a new variable:

```
mtcars <- mtcars %>% dplyr::mutate(efficient = if_else(mpg>30, TRUE, FALSE))
```

# R Syntax Comparison :: CHEAT SHEET

**Syntax** is the set of rules that govern what code works and doesn't work in a programming language. Most programming languages offer one standardized syntax, but R allows package developers to specify their own syntax. As a result, there is a large variety of (equally valid) R syntaxes.

The three most prevalent R syntaxes are:

1. The **dollar sign syntax**, sometimes called **base R syntax**, expected by most base R functions. It is characterized by the use of `dataset$variablename`, and is also associated with square bracket subsetting, as in `dataset[1, 2]`. Almost all R functions will accept things passed to them in dollar sign syntax.
2. The **formula syntax**, used by modeling functions like `lm()`, lattice graphics, and `mosaic` summary statistics. It uses the tilde (~) to connect a response variable and one (or many) predictors. Many base R functions will accept formula syntax.
3. The **tidyverse syntax** used by `dplyr`, `tidyverse`, and more. These functions expect data to be the first argument, which allows them to work with the "pipe" (%>%) from the `magrittr` package. Typically, `ggplot2` is thought of as part of the tidyverse, although it has its own flavor of the syntax using plus signs (+) to string pieces together. `ggplot2` author Hadley Wickham has said the package would have had different syntax if he had written it after learning about the pipe.

**Educators often try to teach within one unified syntax, but most R programmers use some combination of all the syntaxes.**

## Internet research tip:

If you are searching on google, StackOverflow, or another favorite online source and see code in a syntax you don't recognize:

- Check to see if the code is using one of the three common syntaxes listed on this cheatsheet
- Try your search again, using a keyword from the syntax name ("tidyverse") or a relevant package ("mosaic")



Sometimes particular syntaxes work, but are considered dangerous to use, because they are so easy to get wrong. For example, passing variable names without assigning them to a named argument.

## Even more ways to say the same thing

Even within one syntax, there are often variations that are equally valid. As a case study, let's look at the `ggplot2` syntax. `ggplot2` is the plotting package that lives within the tidyverse. If you read `down` this column, all the code here produces the same graphic.

### quickplot

`qplot()` stands for quickplot, and allows you to make quick plots. It doesn't have the full power of `ggplot2`, and it uses a slightly different syntax than the rest of the package.

```
ggplot2::qplot(x=disp, y=mpg, data=mtcars, geom="point")
```

```
ggplot2::qplot(x=disp, y=mpg, data=mtcars) ?
```

```
ggplot2::qplot(disp, mpg, data=mtcars) ?? ??
```

read down this column for many pieces of code in one syntax that look different but produce the same graphic

### ggplot

To unlock the power of `ggplot2`, you need to use the `ggplot()` function (which sets up a plotting region) and add `geoms` to the plot.

```
ggplot2::ggplot(mtcars) +  
  geom_point(aes(x=disp, y=mpg))
```

```
ggplot2::ggplot(data=mtcars) +  
  geom_point(mapping=aes(x=disp, y=mpg))
```

plus adds layers

```
ggplot2::ggplot(mtcars, aes(x=disp, y=mpg)) +  
  geom_point()
```

```
ggplot2::ggplot(mtcars, aes(x=disp)) +  
  geom_point(aes(y=mpg))
```

### ggformula

The "third and a half way" to use the formula syntax, but get `ggplot2`-style graphics

```
ggformula::gf_point(mpg~disp, data= mtcars)
```

### formulas in base plots

Base R plots will also take the formula syntax, although it's not as commonly used

```
plot(mpg~disp, data=mtcars)
```

# The teachR's :: CHEAT SHEET

Getting ready to teach some R? Use our cheat sheet to **prepare, teach and debrief**



## Before the course (design)

Use these to prepare your lecture/course:

**Who are your learners? (Persona Analysis)**  
(change according to requirements...[1])

The R novice

**Background:** some statistics, some programming

**Prior knowledge:** basic R course, base R syntax

**Goals:** understand tidy concepts,  
expose to tidyverse practices

**Special needs:** First successes, mitigate fears, encourage learning

The R “false expert”

**Background:** working with R for some time, but doesn't keep-up

**Prior knowledge:** been using base R syntax, loops, and functions

**Goals:** strengthen tidyverse familiarity, apply dplyr workflow

**Special needs:** switch from obsolete methods to state-of-the-art R

**Define goals using Bloom's Taxonomy [2]**

Design your classes to move your learners “up the pyramid”

Keep “realistic goals” for each persona

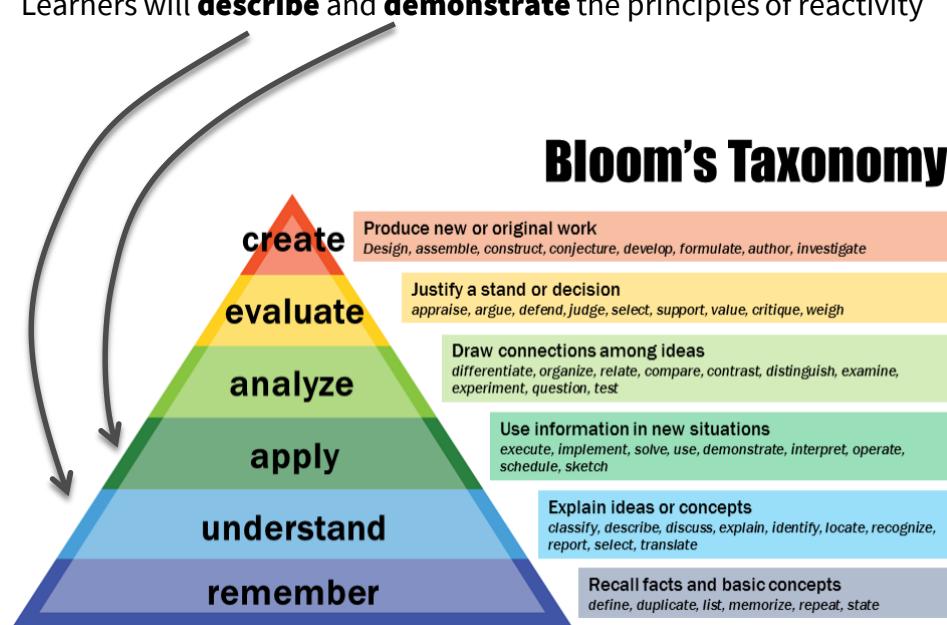


**For example (R shiny - novice):**

Learners will **describe** and **demonstrate** the principles of reactivity

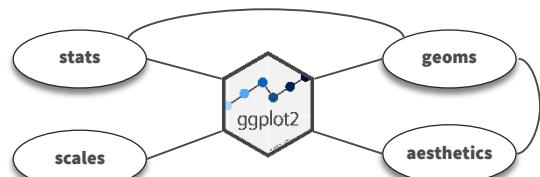


### Bloom's Taxonomy



## Design your lecture using Conceptual maps

Keep the number of elements small (up to ~7 items), e.g.:



## Write the “final exam”

How are you going to test knowledge after the lecture?

What should learners be able to answer?

## Turn the concepts into slides



## Add faded examples (exercises) and check-in slides

**Check-ins**, e.g.: multiple choice quick questions”

**Faded examples** = fill in the blanks, e.g.:

ggplot(data = \_\_\_, mapping = aes(x = \_\_\_, y = \_\_\_)) +  
geom\_ \*() +

## After the course (learn/improve)

Make sure you make the most to improve your next lecture



Use feedback to understand what went well, and what you need to improve.



Measure the time each lecture takes you (or where did you get to), so that next time your time estimates will be better

## Useful tips and tricks

### Useful tips for preparations



Use github to upload course materials

RMarkdown for exercises

Recommended reading materials/references for R courses:

**R for Data Science** / Garrett Grolemund and Hadley Wickham

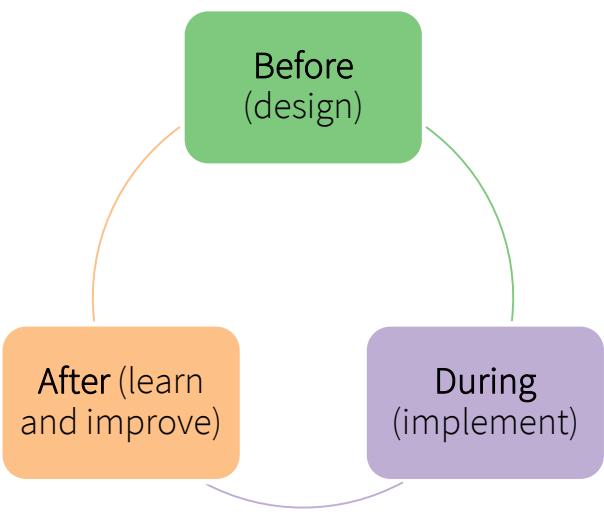
([r4ds.had.co.nz](http://r4ds.had.co.nz))

**Advanced R** / Hadley Wickham ([adv-r.had.co.nz](http://adv-r.had.co.nz))

RStudio Cheat sheets:

<https://www.rstudio.com/resources/cheatsheets/>

## Iterative work flow



## Additional sources

[1] Dreyfus, Stuart E., and Hubert L. Dreyfus. *A five stage model of the mental activities involved in directed skill acquisition*. No. ORC-80-2. California Univ Berkeley Operations Research Center, 1980.

[2] Content downloaded from <https://cft.vanderbilt.edu/guides-sub-pages/blooms-taxonomy/>  
(CC-BY-SA Vanderbilt University Center for Teaching)

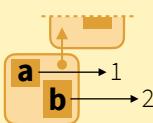
# Tidy evaluation with rlang :: CHEAT SHEET



## Vocabulary

**Tidy Evaluation (Tidy Eval)** is not a package, but a framework for doing non-standard evaluation (i.e. delayed evaluation) that makes it easier to program with tidyverse functions.

**pi**



**Symbol** - a name that represents a value or object stored in R. `is_symbol(expr(pi))`

**Environment** - a list-like object that binds symbols (names) to objects stored in memory. Each env contains a link to a second, **parent** env, which creates a chain, or search path, of environments. `is_environment(current_env())`

`rlang::caller_env(n = 1)` Returns calling env of the function it is in.

`rlang::child_env(.parent, ...)` Creates new env as child of .parent. Also **env**.

`rlang::current_env()` Returns execution env of the function it is in.

**1**

**abs (1)**

**pi** — code  
3.14 — result

**Constant** - a bare value (i.e. an atomic vector of length 1). `is_bare_atomic(1)`

**Call object** - a vector of symbols/constants/calls that begins with a function name, possibly followed by arguments. `is_call(expr(abs(1)))`

**Code** - a sequence of symbols/constants/calls that will return a result if evaluated. Code can be:

1. Evaluated immediately (**Standard Eval**)
2. Quoted to use later (**Non-Standard Eval**)

`is_expression(expr(pi))`

**Expression** - an object that stores quoted code without evaluating it. `is_expression(expr(a + b))`

**Quosure**- an object that stores both quoted code (without evaluating it) and the code's environment. `is_quosure(quo(a + b))`

`a` **rlang::quo\_get\_env(quo)** Return the environment of a quosure.

`a` **rlang::quo\_set\_env(quo, expr)** Set the environment of a quosure.

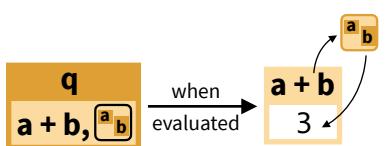
`a + b` **rlang::quo\_get\_expr(quo)** Return the expression of a quosure.

**Expression Vector** - a list of pieces of quoted code created by base R's `expression` and `parse` functions. Not to be confused with **expression**.

## Quoting Code

Quote code in one of two ways (if in doubt use a quosure):

### QUOSURES



**Quosure**- An expression that has been saved *with an environment* (aka a closure).

A quosure can be evaluated later in the stored environment to return a predictable result.

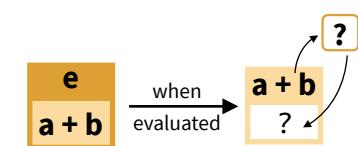
`rlang::quo(expr)` Quote contents as a quosure. Also **quos** to quote multiple expressions. `a <- 1; b <- 2; q <- quo(a + b); qs <- quos(a, b)`

`rlang::enquo(arg)` Call from within a function to quote what the user passed to an argument as a quosure. Also **enquos** for multiple args.  
`quote_this <- function(x) enquo(x)`  
`quote_these <- function(...) enquos(...)`

`rlang::new_quosure(expr, env = caller_env())` Build a quosure from a quoted expression and an environment.  
`new_quosure(expr(a + b), current_env())`



### EXPRESSION



**Quoted Expression** - An expression that has been saved by itself.

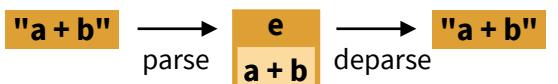
A quoted expression can be evaluated later to return a result that will depend on the environment it is evaluated in

`rlang::expr(expr)` Quote contents. Also **exprs** to quote multiple expressions. `a <- 1; b <- 2; e <- expr(a + b); es <- exprs(a, b, a + b)`

`rlang::enexpr(arg)` Call from within a function to quote what the user passed to an argument. Also **enexprs** to quote multiple arguments.  
`quote_that <- function(x) enexpr(x)`  
`quote_those <- function(...) enexprs(...)`

`rlang::ensym(x)` Call from within a function to quote what the user passed to an argument as a symbol, accepts strings. Also **ensyms**.  
`quote_name <- function(name) ensym(name)`  
`quote_names <- function(...) ensyms(...)`

## Parsing and Deparsing



**Parse** - Convert a string to a saved expression.

• • •

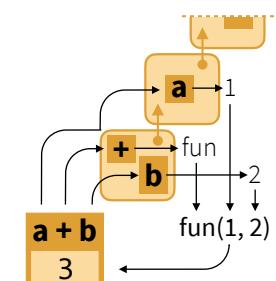
`rlang::parse_expr(x)` Convert a string to an expression. Also **parse\_exprs**, **sym**, **parse\_quo**, **parse\_quos**. `e <- parse_expr("a+b")`

**Deparse** - Convert a saved expression to a string.

• • •

`rlang::expr_text(expr, width = 60L, nlines = Inf)` Convert expr to a string. Also **quo\_name**. `expr_text(e)`

## Evaluation



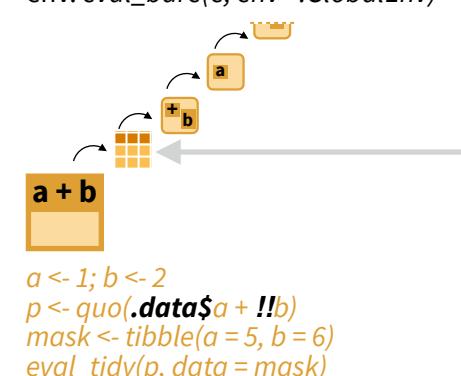
To evaluate an expression, R :

1. Looks up the symbols in the expression in the active environment (or a supplied one), followed by the environment's parents
2. Executes the calls in the expression

**The result of an expression depends on which environment it is evaluated in.**

### QUOTED EXPRESSION

`rlang::eval_bare(expr, env = parent.frame())` Evaluate expr in env. `eval_bare(e, env = GlobalEnv)`



### QUOSURES (and quoted exprs)

`rlang::eval_tidy(expr, data = NULL, env = caller_env())` Evaluate expr in env, using data as a **data mask**. Will evaluate quosures in their stored environment. `eval_tidy(q)`

**Data Mask** - If data is non-NULL, `eval_tidy` inserts data into the search path before env, matching symbols to names in data.

Use the pronoun **.data\$** to force a symbol to be matched in data, and **!!** (see back) to force a symbol to be matched in the environments.

## Building Calls

`rlang::call2(fn, ..., .ns = NULL)` Create a call from a function and a list of args. Use **exec** to create and then evaluate the call. (See back page for !!!)  
`args <- list(x = 4, base = 2)`

`log (x = 4, base = 2)`

**2**

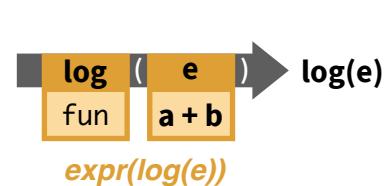
```
call2("log", x = 4, base = 2)
call2("log", !!!args)
exec("log", x = 4, base = 2)
exec("log", !!!args)
```



# Quasiquotation (!!, !!!, :=)

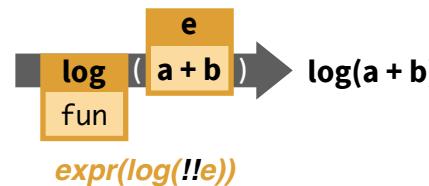
## QUOTATION

Storing an expression without evaluating it.  
`e <- expr(a + b)`



## QUASIUOTATION

Quoting some parts of an expression while evaluating and then inserting the results of others (**unquoting** others).  
`e <- expr(a + b)`

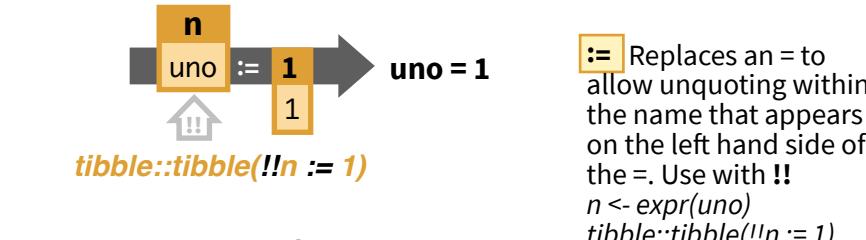
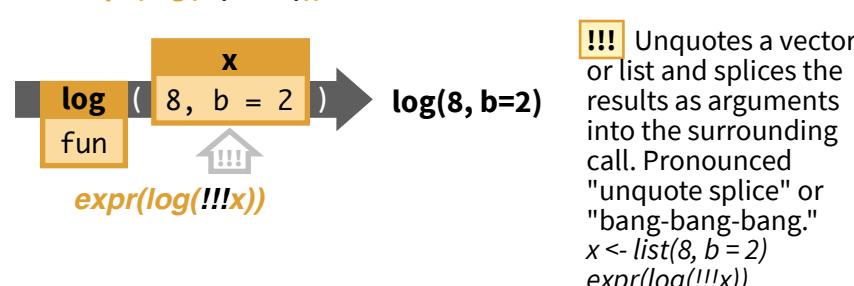
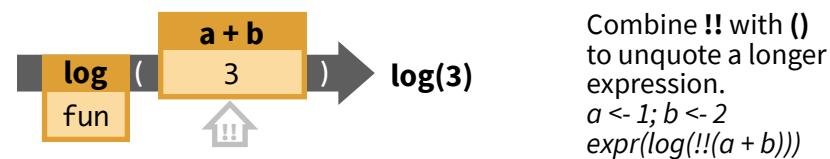
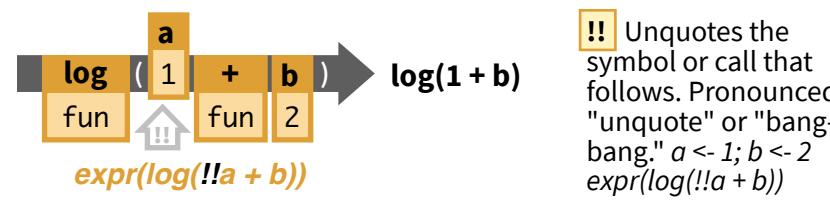


`rlang` provides **!!**, **!!!**, and **:=** for doing quasiquotation.

**!!**, **!!!**, and **:=** are not functions but syntax (symbols recognized by the functions they are passed to). Compare this to how

- . is used by `magrittr::%>%()`
- . is used by `stats::lm()`
- .x is used by `purrr::map()`, and so on.

**!!**, **!!!**, and **:=** are only recognized by some `rlang` functions and functions that use those functions (such as tidyverse functions).



# Programming Recipes

**Quoting function**- A function that quotes any of its arguments internally for delayed evaluation in a chosen environment. You must take **special steps to program safely** with a quoting function.

**How to spot a quoting function?**  
A function quotes an argument if the argument returns an error when run on its own.

Many tidyverse functions are quoting functions: e.g. `filter`, `select`, `mutate`, `summarise`, etc.

```
dplyr::filter(cars, speed == 25)
      speed dist
      1     25    85
```

|                    |
|--------------------|
| <b>speed == 25</b> |
| Error!             |

## PROGRAM WITH A QUOTING FUNCTION

```
data_mean <- function(data, var) {
  require(dplyr)
  var <- rlang::enquo(var) 1
  data %>%
    summarise(mean = mean (!!var)) 2
}
```

1. Capture user argument that will be quoted with `rlang::enquo`.
2. Unquote the user argument into the quoting function with **!!**.

## PASS MULTIPLE ARGUMENTS TO A QUOTING FUNCTION

```
group_mean <- function(data, var, ...) {
  require(dplyr)
  var <- rlang::enquo(var)
  group_vars <- rlang::enquos(...) 1
  data %>%
    group_by (!!group_vars) %>%
    summarise(mean = mean (!!var)) 2
}
```

1. Capture user arguments that will be quoted with `rlang::enquo`.
2. Unquote splice the user arguments into the quoting function with **!!!**.

## MODIFY USER ARGUMENTS

```
my_do <- function(f, v, df) {
  f <- rlang::enquo(f)
  v <- rlang::enquo(v)
  todo <- rlang::quo (!!f)( !!v) 2
  rlang::eval_tidy(todo, df) 3
}
```

1. Capture user arguments with `rlang::enquo`.
2. **Unquote** user arguments into a new expression or quoture to use
3. **Evaluate** the new expression/ quoture instead of the original argument

## APPLY AN ARGUMENT TO A DATA FRAME

```
subset2 <- function(df, rows) {
  rows <- rlang::enquo(rows) 1
  vals <- rlang::eval_tidy(rows, data = df)
  df[vals, , drop = FALSE] 2
}
```

1. Capture user argument with `rlang::enquo`.
2. Evaluate the argument with `rlang::eval_tidy`. Pass the data frame to `data` to use as a data mask.
3. **Suggest** in your documentation that your users use the `.data` and `.env` pronouns.

## WRITE A FUNCTION THAT RECOGNIZES QUASIUOTATION (!!, !!!, :=)

1. Capture the quasiquotation-aware argument with `rlang::enquo`.
2. Evaluate the arg with `rlang::eval_tidy`.

```
add1 <- function(x) {
  q <- rlang::enquo(x)
  rlang::eval_tidy(q) + 1
}
```

1  
2

## PASS TO ARGUMENT NAMES OF A QUOTING FUNCTION

```
named_m <- function(data, var, name) {
  require(dplyr)
  var <- rlang::enquo(var)
  name <- rlang::ensym(name) 1
  data %>%
    summarise (!!name := mean (!!var)) 2
}
```

1. Capture user argument that will be quoted with `rlang::ensym`.
2. Unquote the name into the quoting function with **!!** and **:=**.

## PASS CRAN CHECK

```
#' @importFrom rlang .data
  mutate_y <- function(df) {
    dplyr::mutate(df, y = .data$a + 1)
  }
```

1  
2

Quoted arguments in tidyverse functions can trigger an **R CMD check** NOTE about undefined global variables. To avoid this:

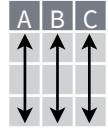
1. Import `rlang::data` to your package, perhaps with the roxygen2 tag `@importFrom rlang .data`
2. Use the `.data$` pronoun in front of variable names in tidyverse functions

# Data tidying with `tidyr` :: CHEATSHEET



**Tidy data** is a way to organize tabular data in a consistent data structure across packages.

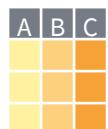
A table is tidy if:



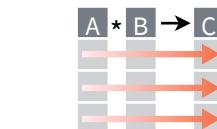
Each **variable** is in its own **column**



Each **observation**, or **case**, is in its own row



Access **variables** as **vectors**



Preserve **cases** in vectorized operations

## Tibbles

### AN ENHANCED DATA FRAME

Tibbles are a table format provided by the **tibble** package. They inherit the data frame class, but have improved behaviors:

- **Subset** a new tibble with `[`, a vector with `[[` and `$`.
- **No partial matching** when subsetting columns.
- **Display** concise views of the data on one screen.

`options(tibble.print_max = n, tibble.print_min = m, tibble.width = Inf)` Control default display settings.

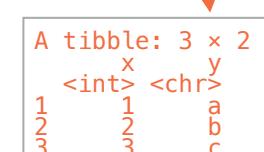
`View()` or `glimpse()` View the entire data set.

### CONSTRUCT A TIBBLE

**tibble(...)** Construct by columns.

`tibble(x = 1:3, y = c("a", "b", "c"))`

Both make this tibble



**tibble(...)** Construct by rows.

`tibble(~x, ~y, 1, "a", 2, "b", 3, "c")`

**as\_tibble(x, ...)** Convert a data frame to a tibble.

**enframe(x, name = "name", value = "value")**

Convert a named vector to a tibble. Also **deframe()**.

**is\_tibble(x)** Test whether x is a tibble.



## Reshape Data

- Pivot data to reorganize values into a new layout.

table4a

| country | 1999 | 2000 |
|---------|------|------|
| A       | 0.7K | 2K   |
| B       | 37K  | 80K  |
| C       | 212K | 213K |



| country | year | cases |
|---------|------|-------|
| A       | 1999 | 0.7K  |
| B       | 1999 | 37K   |
| C       | 1999 | 212K  |
| A       | 2000 | 2K    |
| B       | 2000 | 80K   |
| C       | 2000 | 213K  |

table2

| country | year | type  | count |
|---------|------|-------|-------|
| A       | 1999 | cases | 0.7K  |
| A       | 1999 | pop   | 19M   |
| A       | 2000 | cases | 2K    |
| A       | 2000 | pop   | 20M   |
| B       | 1999 | cases | 37K   |
| B       | 1999 | pop   | 172M  |
| B       | 2000 | cases | 80K   |
| B       | 2000 | pop   | 174M  |
| C       | 1999 | cases | 212K  |
| C       | 1999 | pop   | 1T    |
| C       | 2000 | cases | 213K  |
| C       | 2000 | pop   | 1T    |

| country | year | cases | pop  |
|---------|------|-------|------|
| A       | 1999 | 0.7K  | 19M  |
| A       | 2000 | 2K    | 20M  |
| B       | 1999 | 37K   | 172M |
| B       | 2000 | 80K   | 174M |
| C       | 1999 | 212K  | 1T   |
| C       | 2000 | 213K  | 1T   |

## Split Cells

- Use these functions to split or combine cells into individual, isolated values.

table5

| country | century | year |
|---------|---------|------|
| A       | 19      | 99   |
| A       | 20      | 00   |
| B       | 19      | 99   |
| B       | 20      | 00   |

| country | year |
|---------|------|
| A       | 1999 |
| A       | 2000 |
| B       | 1999 |
| B       | 2000 |

table3

| country | year | rate     |
|---------|------|----------|
| A       | 1999 | 0.7K/19M |
| A       | 2000 | 2K/20M   |
| B       | 1999 | 37K/172M |
| B       | 2000 | 80K/174M |

| country | year | rate |
|---------|------|------|
| A       | 1999 | 0.7K |
| A       | 1999 | 19M  |
| A       | 2000 | 2K   |
| A       | 2000 | 20M  |
| B       | 1999 | 37K  |
| B       | 1999 | 172M |
| B       | 2000 | 80K  |
| B       | 2000 | 174M |

**pivot\_longer**(data, cols, names\_to = "name", values\_to = "value", values\_drop\_na = FALSE)

"Lengthen" data by collapsing several columns into two. Column names move to a new names\_to column and values to a new values\_to column.

```
pivot_longer(table4a, cols = 2:3, names_to = "year", values_to = "cases")
```

**pivot\_wider**(data, names\_from = "name", values\_from = "value")

The inverse of pivot\_longer(). "Widen" data by expanding two columns into several. One column provides the new column names, the other the values.

```
pivot_wider(table2, names_from = type, values_from = count)
```

## Expand Tables

Create new combinations of variables or identify implicit missing values (combinations of variables not present in the data).

| x | x1 | x2 | x3 |
|---|----|----|----|
| A | 1  | 3  |    |
| B | 1  | 4  |    |
| B | 2  | 3  |    |

| x | x1 | x2 | x3 |
|---|----|----|----|
| A | 1  | 3  |    |
| B | 1  | 4  |    |
| B | 2  | 3  |    |
| B | 2  | 3  | NA |

**expand**(data, ...) Create a new tibble with all possible combinations of the values listed in ... Drop other variables.

```
expand(mtcars, cyl, gear, carb)
```

**complete**(data, ..., fill = list()) Add missing possible combinations of values of variables listed in ... Fill remaining variables with NA.

```
complete(mtcars, cyl, gear, carb)
```

| x | x1 | x2 |
|---|----|----|
| A | 1  | 3  |
| B | NA |    |
| C | NA |    |
| D | 3  |    |
| E | NA |    |

**drop\_na**(data, ...) Drop rows containing NA's in ... columns.

```
drop_na(x, x2)
```

| x | x1 | x2 |
|---|----|----|
| A | 1  |    |
| B | NA |    |
| C | NA |    |
| D | 3  |    |
| E | NA |    |

**fill**(data, ..., .direction = "down") Fill in NA's in ... columns using the next or previous value.

```
fill(x, x2)
```

| x | x1 |
| --- | --- |



# Nested Data

A **nested data frame** stores individual tables as a list-column of data frames within a larger organizing data frame. List-columns can also be lists of vectors or lists of varying data types.

Use a nested data frame to:

- Preserve relationships between observations and subsets of data. Preserve the type of the variables being nested (factors and datetimes aren't coerced to character).
- Manipulate many sub-tables at once with **purrr** functions like `map()`, `map2()`, or `pmap()` or with **dplyr** `rowwise()` grouping.

## CREATE NESTED DATA

**nest(data, ...)** Moves groups of cells into a list-column of a data frame. Use alone or with `dplyr::group_by()`:

1. Group the data frame with `group_by()` and use `nest()` to move the groups into a list-column.

```
n_storms <- storms |>
  group_by(name) |>
  nest()
```

2. Use `nest(new_col = c(x, y))` to specify the columns to group using `dplyr::select()` syntax.

```
n_storms <- storms |>
  nest(data = c(year:long))
```

| name | yr   | lat  | long  |
|------|------|------|-------|
| Amy  | 1975 | 27.5 | -79.0 |
| Amy  | 1975 | 28.5 | -79.0 |
| Amy  | 1975 | 29.5 | -79.0 |
| Bob  | 1979 | 22.0 | -96.0 |
| Bob  | 1979 | 22.5 | -95.3 |
| Bob  | 1979 | 23.0 | -94.6 |
| Zeta | 2005 | 23.9 | -35.6 |
| Zeta | 2005 | 24.2 | -36.1 |
| Zeta | 2005 | 24.7 | -36.6 |

| name | yr   | lat  | long  |
|------|------|------|-------|
| Amy  | 1975 | 27.5 | -79.0 |
| Amy  | 1975 | 28.5 | -79.0 |
| Amy  | 1975 | 29.5 | -79.0 |
| Bob  | 1979 | 22.0 | -96.0 |
| Bob  | 1979 | 22.5 | -95.3 |
| Bob  | 1979 | 23.0 | -94.6 |
| Zeta | 2005 | 23.9 | -35.6 |
| Zeta | 2005 | 24.2 | -36.1 |
| Zeta | 2005 | 24.7 | -36.6 |

| name  | data            |
|-------|-----------------|
| Luke  | <tibble [50x3]> |
| C-3PO | <tibble [50x3]> |
| R2-D2 | <tibble [50x3]> |

| name  | films     |
|-------|-----------|
| Luke  | <chr [5]> |
| C-3PO | <chr [6]> |
| R2-D2 | <chr[7]>  |

Index list-columns with `[[[]]]`. `n_storms$data[[1]]`

## CREATE TIBBLES WITH LIST-COLUMNS

**tibble::tribble(...)** Makes list-columns when needed.

```
tribble(~max, ~seq,
       3, 1:3,
       4, 1:4,
       5, 1:5)
```

| max | seq       |
|-----|-----------|
| 3   | <int [3]> |
| 4   | <int [4]> |
| 5   | <int [5]> |

**tibble::tibble(...)** Saves list input as list-columns.

```
tibble(max = c(3, 4, 5), seq = list(1:3, 1:4, 1:5))
```

**tibble::enframe(x, name="name", value="value")**

Converts multi-level list to a tibble with list-cols.  
`enframe(list('3'=1:3, '4'=1:4, '5'=1:5), 'max', 'seq')`

## OUTPUT LIST-COLUMNS FROM OTHER FUNCTIONS

**dplyr::mutate(), transmute(), and summarise()** will output list-columns if they return a list.

```
mtcars |>
  group_by(cyl) |>
  summarise(q = list(quantile(mpg)))
```

## RESHAPE NESTED DATA

**unnest(data, cols, ..., keep\_empty = FALSE)** Flatten nested columns back to regular columns. The inverse of `nest()`.  
`n_storms |> unnest(data)`

**unnest\_longer(data, col, values\_to = NULL, indices\_to = NULL)**  
Turn each element of a list-column into a row.

```
starwars |>
  select(name, films) |>
  unnest_longer(films)
```

| name | yr   | lat  | long  |
|------|------|------|-------|
| Amy  | 1975 | 27.5 | -79.0 |
| Amy  | 1975 | 28.5 | -79.0 |
| Amy  | 1975 | 29.5 | -79.0 |
| Bob  | 1979 | 22.0 | -96.0 |
| Bob  | 1979 | 22.5 | -95.3 |
| Bob  | 1979 | 23.0 | -94.6 |
| Zeta | 2005 | 23.9 | -35.6 |
| Zeta | 2005 | 24.2 | -36.1 |
| Zeta | 2005 | 24.7 | -36.6 |

| name  | films     |
|-------|-----------|
| Luke  | <chr [5]> |
| C-3PO | <chr [6]> |
| R2-D2 | <chr[7]>  |

| name  | films                |
|-------|----------------------|
| Luke  | The Empire Strik...  |
| Luke  | Revenge of the S...  |
| Luke  | Return of the Jed... |
| C-3PO | The Empire Strik...  |
| C-3PO | Attack of the Cl...  |
| C-3PO | The Phantom M...     |
| R2-D2 | The Empire Strik...  |
| R2-D2 | Attack of the Cl...  |
| R2-D2 | The Phantom M...     |

**unnest\_wider(data, col)** Turn each element of a list-column into a regular column.

```
starwars |>
  select(name, films) |>
  unnest_wider(films, names_sep = "_")
```

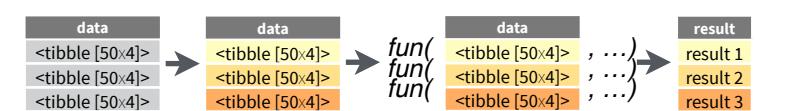
| name  | films     |
|-------|-----------|
| Luke  | <chr [5]> |
| C-3PO | <chr [6]> |
| R2-D2 | <chr[7]>  |

| name  | films_1       | films_2       | films_3        |
|-------|---------------|---------------|----------------|
| Luke  | The Empire... | Revenge of... | Return of...   |
| C-3PO | The Empire... | Attack of...  | The Phantom... |
| R2-D2 | The Empire... | Attack of...  | The Phantom... |

## TRANSFORM NESTED DATA

A vectorized function takes a vector, transforms each element in parallel, and returns a vector of the same length. By themselves vectorized functions cannot work with lists, such as list-columns.

**dplyr::rowwise(.data, ...)** Group data so that each row is one group, and within the groups, elements of list-columns appear directly (accessed with `[]`, not as lists of length one. **When you use `rowwise()`, dplyr functions will seem to apply functions to list-columns in a vectorized fashion.**



Apply a function to a list-column and **create a new list-column**.

```
n_storms |>
  rowwise() |>
  mutate(n = list(dim(data)))
```

**dim()** returns two values per row  
wrap with `list` to tell `mutate` to create a list-column

Apply a function to a list-column and **create a regular column**.

```
n_storms |>
  rowwise() |>
  mutate(n = nrow(data))
```

**nrow()** returns one integer per row

Collapse **multiple list-columns** into a single list-column.

```
starwars |>
  rowwise() |>
  mutate(transport = list(append(vehicles, starships)))
```

**append()** returns a list for each row, so col type must be list

Apply a function to **multiple list-columns**.

```
starwars |>
  rowwise() |>
  mutate(n_transports = length(c(vehicles, starships)))
```

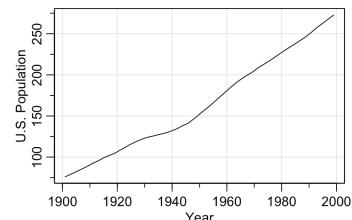
**length()** returns one integer per row

See **purrr** package for more list functions.

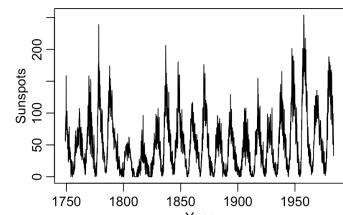
# Time Series Cheat Sheet

## Plot Time Series

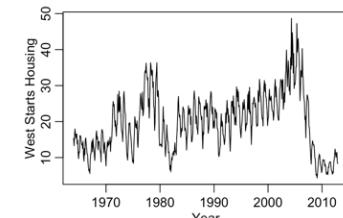
1. `tsplot(x=time, y=data)`



2. `plot(ts(data, start=start_time, frequency=gap))`



3. `ts.plot(ts(data, start=start_time, frequency=gap))`



## Simulation

### Autoregression of Order p

$$X_t = \phi_1 X_{t-1} + \phi_2 X_{t-2} + \dots + \phi_p X_{t-p} + W_t$$

### Moving Average of Order q

$$X_t = Z_t + \theta_1 Z_{t-1} + \theta_2 Z_{t-2} + \dots + \theta_q Z_{t-q}$$

### ARMA (p, q)

$$X_t = \phi_1 X_{t-1} + \phi_2 X_{t-2} + \dots + \phi_p X_{t-p} + Z_t + \theta_1 Z_{t-1} + \theta_2 Z_{t-2} + \dots + \theta_q Z_{t-q}$$

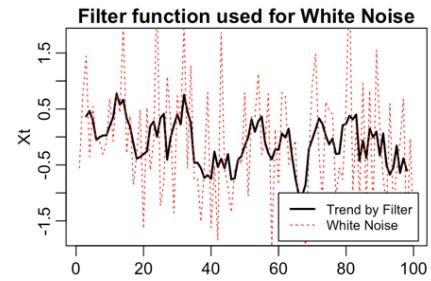
### Simulation of ARMA (p, q)

`arima.sim(model=list(ar=c(phi1, ..., phip), ma=c(theta1, ..., thetaq)), n=n)`

## Filters

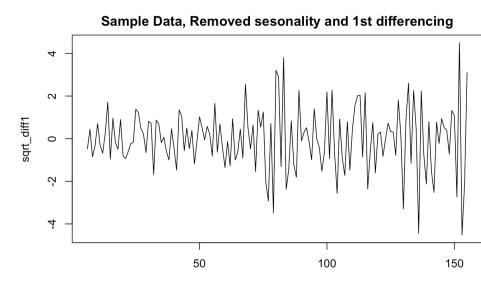
### Linear Filter: `filter()`

`filter(data, filter=filter_coefficients, sides=2, method="convolution", circular=F)`



### Differencing Filter: `diff()`

`diff(data, lag=4, differences=1)`

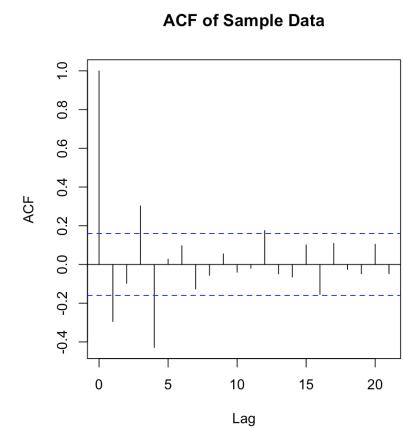


## Auto-correlation

### Use ACF and PACF to detect model

### (Complete) Auto-correlation function: `acf()`

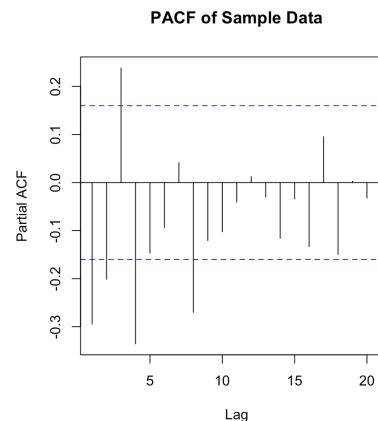
`acf(data, type='correlation', na.action=na.pass)`



### Partial Auto-correlation function: `pacf()`

`pacf(data, na.action=na.pass)`

**OR:** `acf(data, type='partial', na.action=na.pass)`



## Forecasting

### Forecasting future observations given a fitted ARMA model

**predict():** Predict future observations given a fitted ARMA model

`predict(arima_model, number_to_predict)`

### Plot Predicted values and Confidence Interval:

`fit<-predict(arima_model, number_to_predict)`

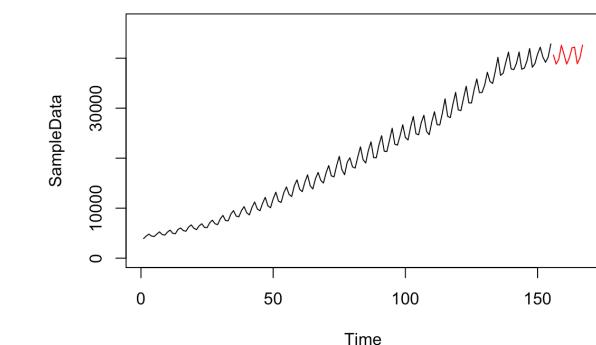
`ts.plot(data,`

`xlim=c(1, length(data)+number_to_predict),`

`ylim=c(0, max(fit$pred+1.96*fit$se)))`

`lines(length(data)+1:length(data)+`

`number_to_predict, fit$pred)`



### arima(): To estimate parameters of an AR model

`ar(x=data, aic=T, order.max = NULL, c("yule-walker", "burg", "ols", "mle", "yw"))`

```
Call:
ar(x = sqrt1, aic = TRUE, order.max = NULL, method = c("yule-walker", "burg", "ols", "mle", "yw"))

Coefficients:
1 -0.3066 -0.1903 0.0793 -0.5065 -0.1873 -0.1149 -0.0580 -0.3031 -0.1207
2 -0.1903 0.0793 -0.5065 -0.1873 -0.1149 -0.0580 -0.3031 -0.1207
3 0.0793 -0.5065 -0.1873 -0.1149 -0.0580 -0.3031 -0.1207
4 -0.5065 -0.1873 -0.1149 -0.0580 -0.3031 -0.1207
5 -0.1873 -0.1149 -0.0580 -0.3031 -0.1207
6 -0.1149 -0.0580 -0.3031 -0.1207
7 -0.0580 -0.3031 -0.1207
8 -0.3031 -0.1207
9 -0.1207

Order selected 9 sigma^2 estimated as 1.52
```

**arima():** To estimate parameters of an AM or ARMA model, and build model

`arima(data, order=c(p, o, q), method=c('ML'))`

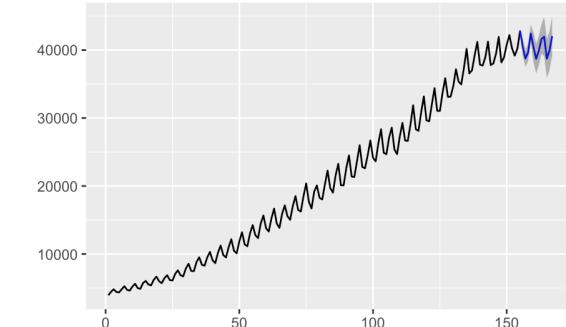
```
Call:
arima(x = sqrt1, order = c(2, 0, 6), method = c("ML"))

Coefficients:
ar1 ar2 ma1 ma2 ma3 ma4 ma5 ma6 intercept
-0.1658 -0.7951 -0.1536 0.7524 -0.2101 -0.7680 0.0605 -0.6812 -0.0048
s.e. 0.0918 0.0754 0.0916 0.1047 0.0794 0.0743 0.0812 0.0895 0.0062
sigma^2 estimated as 1.193: log likelihood = -230.78, aic = 481.55
```

### AICc(): Compare models using AICC

`AICc(fittedModel)`

### Predicted value and Conf Interval of ARIMA



# Deep Learning with {torch} CHEAT SHEET



## Intro

{torch} is based on Pytorch, a framework popular among deep learning researchers.

{torch}'s GPU acceleration allows to implement fast machine learning algorithms using its convenient interface, as well as a vast range of use cases, not only for deep learning, according to its flexibility and its low level API.

It is part of an ecosystem of packages to interface with specific dataset like {torchaudio} for timeseries-like, {torchvision} for image-like, and {tabnet} for tabular data. It is complemented by {luz} for a higher-level programming interface

## Working with torch models

### DEFINE A NN MODULE

```
dense <- nn_module(
  "no_bias_dense_layer",
  initialize = function(in_f, out_f) {
    self$w <- nn_parameter(torch_randn(in_f, out_f))
  },
  forward = function(x) {
    torch_mm(x, self$w)
  }
)
Create a nn module names no_bias_dense_layer
```

### ASSEMBLE MODULES INTO NETWORK

```
model <- dense(4, 3)
Instantiate a network from a single module

model <- nn_sequential(
  dense(4,3), nn_relu(), nn_dropout(0.4),
  dense(3,1), nn_sigmoid())
Instantiate a sequential network with multiple layers
```

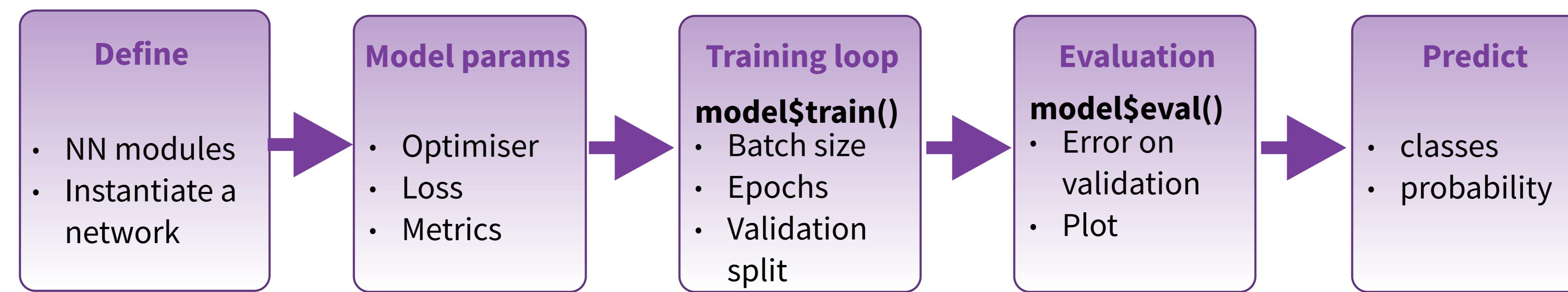
### MODEL FIT

```
model$train()
Turns on gradient update

with_enable_grad({
  y_pred <- model(trainset)
  loss <- (y_pred - y)$pow(2)$mean()
  loss$backward()
})
Detailed training loop step (alternative)
```

### EVALUATE A MODEL

```
model$eval()
or
with_no_grad({
  model(validationset)
})
Perform forward operation with no gradient update
```



<https://torch.mlverse.org/>

<https://mlverse.shinyapps.io/torch-tour/>

## INSTALLATION

The torch R package uses the C++ libtorch library. You can install the prerequisites directly from R.

<https://torch.mlverse.org/docs/articles/installation.html>

```
install.packages("torch")
library(torch)
install_torch()
```

See `?install_torch` for GPU instructions

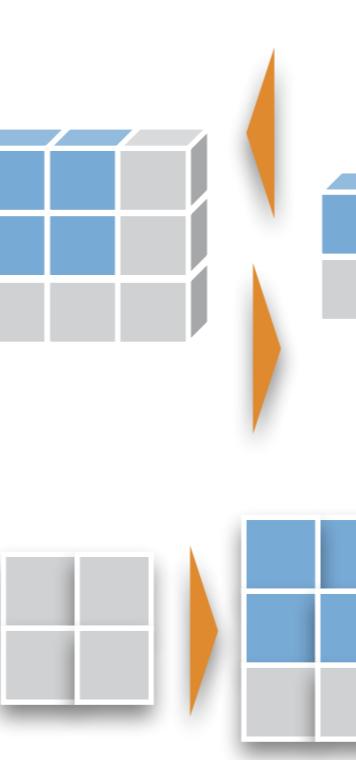
## Neural-network layers

### CORE LAYERS

|  |  |
|--|--|
|  | <b>nn_linear()</b><br>Add a linear transformation NN layer to an input   |
|  | <b>nn_bilinear()</b> to two inputs   |
|  | <b>nn_sigmoid()</b> , <b>nn_relu()</b><br>Apply an activation function to an output  |
|  | <b>nn_dropout()</b><br><b>nn_dropout2d()</b><br><b>nn_dropout3d()</b><br>Applies Dropout to the input                          |
|  | <b>nn_batch_norm1d()</b><br><b>nn_batch_norm2d()</b><br><b>nn_batch_norm3d()</b><br>Applies batch normalisation to the weights |

### CONVOLUTIONAL LAYERS

|  |   |
|--|---|
|  | <b>nn_conv1d()</b> 1D, e.g. temporal convolution  |
|  | <b>nn_conv_transpose2d()</b><br>Transposed 2D (deconvolution)   |
|  | <b>nn_conv2d()</b> 2D, e.g. spatial convolution over images   |
|  | <b>nn_conv_transpose3d()</b><br>Transposed 3D (deconvolution)<br><b>nn_conv3d()</b> 3D, e.g. spatial convolution over volumes |



**nnf\_pad()**  
Zero-padding layer

### ACTIVATION LAYERS

|  |   |
|--|---|
|  | <b>nn_leaky_relu()</b><br>Leaky version of a rectified linear unit                    |
|  | <b>nn_relu6()</b><br>rectified linear unit clamped by 6                               |
|  | <b>nn_rrelu()</b><br>Randomized leaky rectified linear unit                           |
|  | <b>nn_elu()</b> , <b>nn_selu()</b><br>Exponential linear unit, Scaled Exp lineal unit |

### POOLING LAYERS

|  |  |
|--|--|
|  | <b>nn_max_pool1d()</b>                                 |
|  | <b>nn_max_pool2d()</b>                                 |
|  | <b>nn_max_pool3d()</b><br>Maximum pooling for 1D to 3D |
|  | <b>nn_avg_pool1d()</b>                                 |
|  | <b>nn_avg_pool2d()</b>                                 |
|  | <b>nn_avg_pool3d()</b><br>Average pooling for 1D to 3D |

|  |   |
|--|---|
|  | <b>nn_adaptive_max_pool1d()</b>                             |
|  | <b>nn_adaptive_max_pool2d()</b>                             |
|  | <b>nn_adaptive_max_pool3d()</b><br>Adaptive maximum pooling |

|  |   |
|--|---|
|  | <b>nn_adaptive_avg_pool1d()</b>                             |
|  | <b>nn_adaptive_avg_pool2d()</b>                             |
|  | <b>nn_adaptive_avg_pool3d()</b><br>Adaptive average pooling |

### RECURRENT LAYERS

|  |  |
|--|--|
|  | <b>nn_rnn()</b><br>Fully-connected RNN where the output is to be fed back to input |
|  | <b>nn_gru()</b><br>Gated recurrent unit - Cho et al                                |
|  | <b>nn_lstm()</b><br>Long-Short Term Memory unit - Hochreiter 1997                  |





# Class Agnostic Time Series with tsbox :: CHEAT SHEET

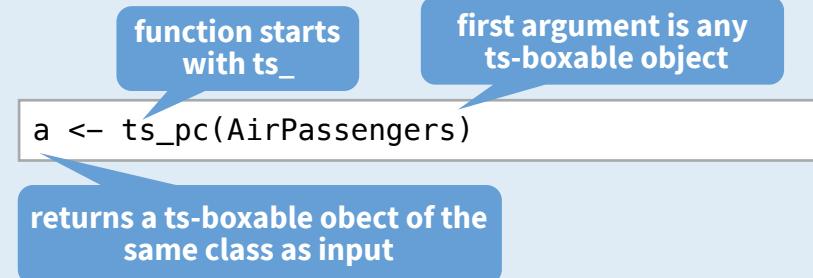
## Basics

### IDEA

tsbox provides a time series toolkit which:

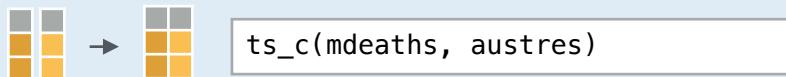
1. works identically with most time series **classes**
2. handles regular and irregular **frequencies**
3. **converts** between classes and frequencies

Most functions in tsbox have the same structure:

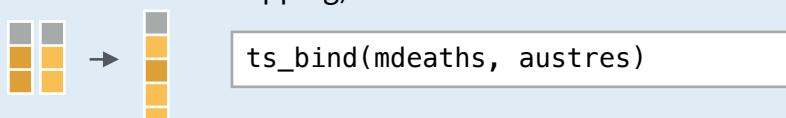


### COMBINE TIME SERIES

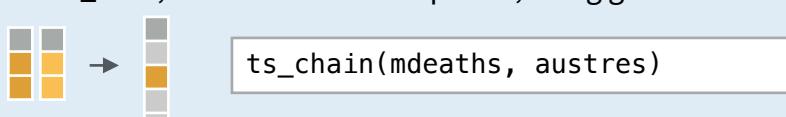
collect time series of **all classes** and **frequencies** as multiple time series



combine time series to a new, single time series (first series wins if overlapping)

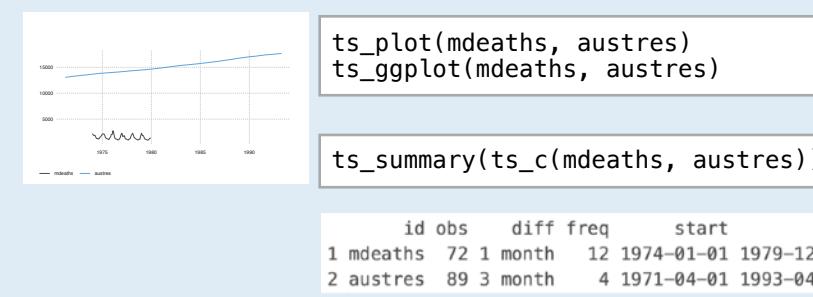


like ts\_bind, but extra- and retropolate, using growth rates



### PLOT AND SUMMARIZE

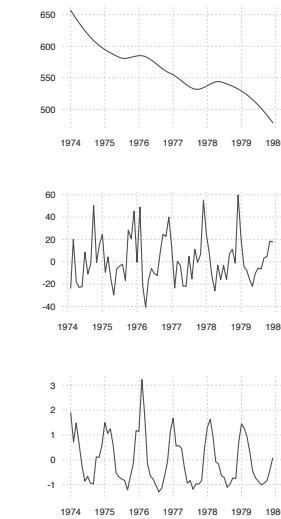
Plot time series of **all classes** and **frequencies**



## Helper Functions

Transform time series of **all classes** and **frequencies**

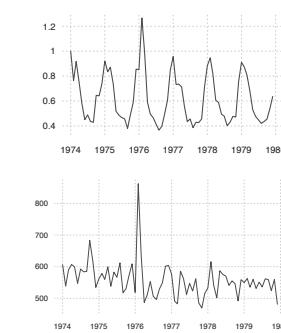
### TRANSFORM



**ts\_trend()**: Trend estimation based on loess

**ts\_pc()**, **ts\_pcy()**, **ts\_pca()**, **ts\_diff()**, **ts\_difffy()**: (annualized) Percentage change rates or differences to previous period, year

**ts\_scale()**: normalize mean and variance



**ts\_index()**: Index, based on levels

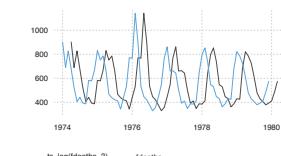
**ts\_compound()**: Index, based on growth rates

**ts\_index(fdeaths, base = 1976)**

**ts\_seas()**: seasonal adjustment using X-13

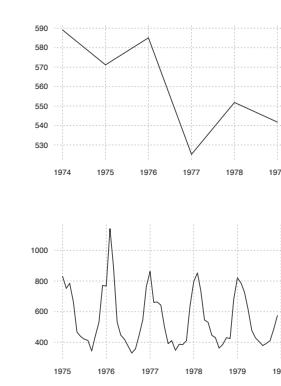
**ts\_seas(fdeaths)**

### SPAN AND FREQUENCY



**ts\_lag()**: Lag or lead of time series

**ts\_lag(fdeaths, 4)**



**ts\_frequency()**: convert to frequency

**ts\_frequency(fdeaths, "year")**

**ts\_span()**: filter time series for a time span.

**ts\_span(fdeaths, "1976-01-01")**

**ts\_span(fdeaths, "-5 year")**

## Class Conversion

tsbox is built around a set of converters, which convert time series of the following **supported classes** to each other:

| converter function                      | ts-boxable class       |
|---|------------------------|
| <b>ts_ts()</b>                          | ts, mts                |
| <b>ts_data.frame()</b> , <b>ts_df()</b> | data.frame             |
| <b>ts_data.table()</b> , <b>ts_dt()</b> | data.table             |
| <b>ts_tbl()</b>                         | df_tbl, "tibble"       |
| <b>ts_xts()</b>                         | xts                    |
| <b>ts_zoo()</b>                         | zoo                    |
| <b>ts_tibbletime()</b>                  | tibbletime             |
| <b>ts_timeSeries()</b>                  | timeSeries             |
| <b>ts_tsibble()</b>                     | tsibble                |
| <b>ts_tslist()</b>                      | a list with ts objects |

## Time Series in data frames

### LONG STRUCTURE

Default structure to store multiple time series in long data frames (or data tables, or tibbles)

**ts\_df(ts\_c(fdeaths, mdeaths))**

| id      | time       | value |
|---------|------------|-------|
| fdeaths | 1974-01-01 | 901   |
| fdeaths | 1974-02-01 | 689   |
| fdeaths | 1974-03-01 | 827   |
| ...     | ...        | ...   |

### AUTO-DETECT COLUMN NAMES

tsbox auto-detects a **value**-, a **time**- and zero, one or several **id**-columns. Alternatively, the **time**- and the **value**-column can be explicitly named **time** and **value**.

**ts\_default()**: standardize column names in data frames

### RESHAPE

**ts\_wide()**: convert default long structure to wide

**ts\_long()**: convert wide structure to default long

### USE WITH PIPE

tsbox plays well with tibbles and with **%>%**, so it can be easily integrated into a dplyr/pipe workflow

```
library(dplyr)
ts_c(fdeaths, mdeaths) %>%
  ts_tbl() %>%
  ts_trend() %>%
  ts_pc()
```

pass return value as first argument to the next function

# Cheat Sheet :: VEGAN



## What is VEGAN?

The **vegan** package provides tools for descriptive community ecology. It has basic functions of **community ordination**, **diversity analysis** and **dissimilarity analysis**. Most of its multivariate tools can be used for other data types as well.

Examples using : **data(dune)**

### Unconstrained Ordination

**metaMDS(data, ...)** Nonmetric Multidimensional Scaling

All ordination results can be displayed with

**plot(data, type = "")**

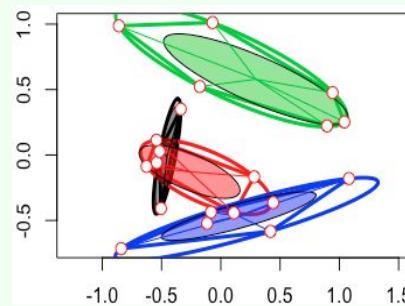
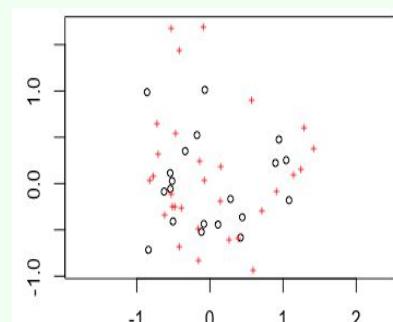
**type = "p"** results with points of black circles to indicate sites and red pluses to show species

**type = "t"** results with text

**ordihull()** adds convex hulls

**ordielipse()** adds ellipses of standard deviation, standard error or confidence areas

**ordispider()** draws items to their center



### Constrained Ordination

**cca(formula, data, ...)** Constrained Correspondence Analysis

Displays only the variation that can be explained by used constraints

**rda(formula, data, scale=FALSE, ...)** Redundancy Analysis

**capscale(formula, data, distance = "", ...)** Distance based Redundancy Analysis

**formula()** Model formula must be either community data matrix or dissimilarity matrix

OR

**distance = "name of dissimilarity index"** if formula is not specified

### Analysis of constraints

**anova.cca(object, permutations = "", ...)** Permutation Test for CCA & RDA to assess the significance of constraints

**object** specifies one or several result objects from cca, rda, or capscale

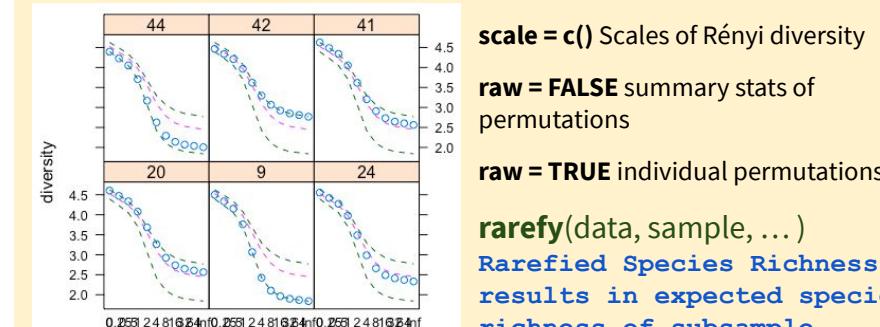
**permutations** = control values, or permutation index

### Diversity Analysis of Eco Communities

**diversity(data, index = "", MARGIN = 1, base = exp(1), ...)**

Shannon, Simpson, and Fisher diversity indices and species richness.

**renyi(data, scale = c(), raw = FALSE, ...)** Rényi Diversity index



**scale = c()** Scales of Rényi diversity

**raw = FALSE** summary stats of permutations

**raw = TRUE** individual permutations

**rarefy(data, sample, ...)**

Rarefied Species Richness results in expected species richness of subsample

### Taxonomic Diversity

**taxondive(data, distance, match.force = FALSE)** Taxonomic diversity indices

**taxa2dist(data, varstep = FALSE, check = TRUE, ...)** Converts class tables to taxonomic distances

### Ranked Abundance Distribution

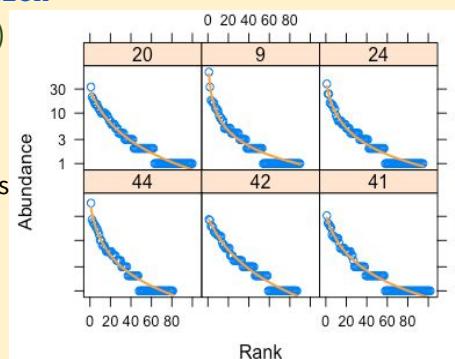
**radfit(data, ...)** Fits the most popular model to data using maximum likelihood estimation

**rad.null(data, family = poisson)**

Fits broken stick model to expected abundance of species

**type = "b"** Plots both observed points and fitted lines

**family =** Error distribution; poisson default is used for counts, gaussian may be appropriate for abundance



### Beta Diversity

**betadiver(data, method = NA, ...)** Estimates beta diversity

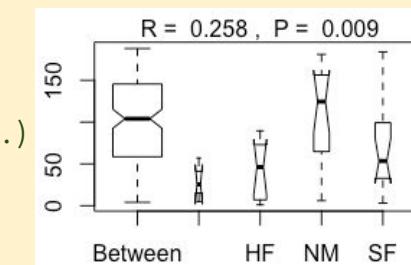
**method = ""** can specify which beta index to use (24 options)

**betadiver(help=TRUE)** list all 24 indices available

### Analysis of Diversity in Groups

**anosim(data, grouping, permutations = "", distance = "", ...)**

Analysis of similarities between two or more groups



### Dissimilarity Analysis

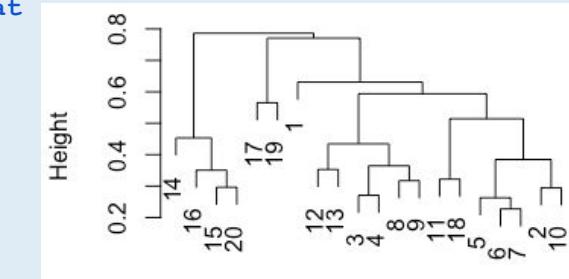
**vegdist(data, method = "", na.rm = FALSE, ...)** Dissimilarity indices

**method = "dissimilarity index"**

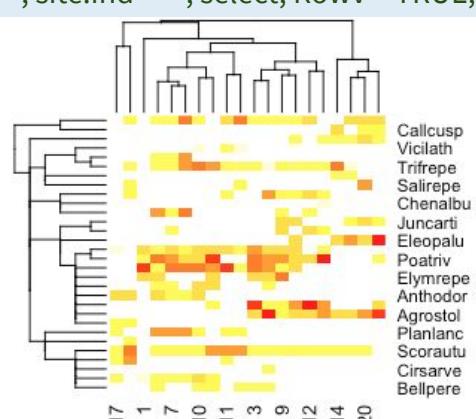
- "manhattan", "euclidean", "canberra", "clark", "bray", "kulczynski", "jaccard", "gower", "altGower", "morisita", "horn", "mountford", "raup", "binomial", "chao", "cao" or "mahalanobis".

### Other Fun Features

**vegemitte(data, use, scale, sp.ind = "", site.ind = "", select, ...)**  
Creates a compact ordered community tree in text format



**tabasco(data, use, sp.ind = "", site.ind = "", select, Rowv = TRUE, Colv = TRUE, scale, col = heat.colors(12), ...)**  
Creates a community table using heat map, abundances are coded by color



**use** is either a vector or object

**sp.ind / site.ind** species and site indices

**select** a subset of plots

**Rowv / Colv** = reorder rows and columns, if TRUE it is ordered by correspondence analysis

**beals(data, species = NA, reference = data, include = TRUE)**

Beals Smoothing and Degree of Absence Analysis determines probability of a species occurring in a site based on joint occurrences with other species

**species = NA** will compute for all species, or can specify single

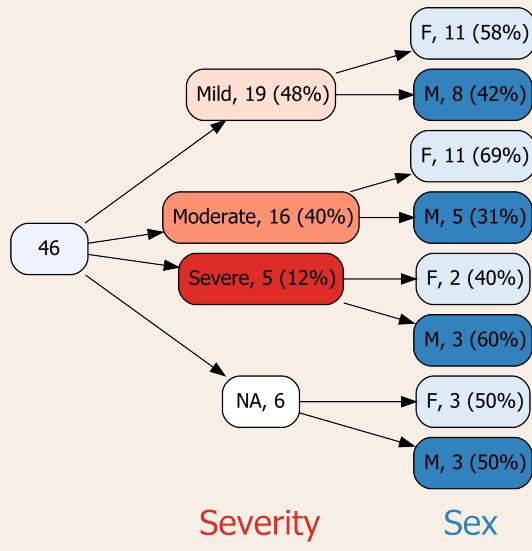
**reference =** data to be used to compare for joint analysis

**include = TRUE** to include target species in computations

\*\*VEGAN uses quantitative data but setting **binary = TRUE** will make data presence/absence\*\*

# Examining nested subsets with vtree: cheat sheet

`vtree(FakeData, "Severity_Sex", sameline=T)`



Severity

Sex

Pruning

| Parameter               | Effect  |
|-------------------------|---|
| <code>prune</code>      | Remove identified nodes and their descendants.      |
| <code>keep</code>       | Only retain identified nodes and their descendants. |
| <code>prunebelow</code> | Remove descendants of identified nodes.             |
| <code>follow</code>     | Only retain descendants of identified nodes.        |

Example: `prune=list(Severity=c("Moderate","Severe"))`

Labels

| Parameter setting   |   |
|---|---|
| <code>labelvar=c(variable="Label")</code>                         | Assign <i>Label</i> to <i>variable</i>                  |
| <code>labelnode=list(variable=c(New="Old"))</code>                | In <i>variable</i> , replace <i>Old</i> with <i>New</i> |
| <code>tlabelnode=list(c(Group="A", Sex="F", label="girl"))</code> | Change the label of a specific node                     |
| <code>varnamepointsiz=30</code>                                   | Set font size for variable names                        |
| <code>shownodelabels=FALSE</code>                                 | Do not show node labels                                 |
| <code>showvarnames=FALSE</code>                                   | Do not show variable names                              |
| <code>showlegend=TRUE</code>                                      | Show a legend   |
| <code>title="All businesses"</code>                               | Show a title for the root node                          |

## Summaries

| Type                   | Parameter setting                      |
|------------------------|--|
| Simple                 | <code>summary="variable"</code>        |
| Custom                 | <code>summary="variable format"</code> |
| Code                   | Produces                               |
| <code>%mean%</code>    | mean                                   |
| <code>%SD%</code>      | standard deviation                     |
| <code>%sum%</code>     | sum                                    |
| <code>%range%</code>   | range                                  |
| <code>%median%</code>  | median                                 |
| <code>%IQR%</code>     | inter-quartile range                   |
| <code>%freqpct%</code> | frequency and %                        |
| <code>%freq%</code>    | just frequency                         |
| <code>%npct%</code>    | frequency and %                        |
| <code>%pct%</code>     | %                                      |
| <code>%list%</code>    | list values                            |
| <code>%trunc=n%</code> | truncation at <i>n</i> characters      |

## Control code summary restricted to:

|                         |                            |
|-------------------------|----------------------------|
| <code>%noroot%</code>   | all nodes except the root  |
| <code>%leafonly%</code> | leaf nodes                 |
| <code>%var=v%</code>    | nodes of variable <i>v</i> |
| <code>%node=n%</code>   | nodes named <i>n</i>       |

## Image settings

| Parameter setting              | Effect          |
|--------------------------------|-----------------|
| <code>imagedwidth="3in"</code> | 3 inches wide   |
| <code>imageheight="4in"</code> | 4 inches tall   |
| <code>pxwidth=800</code>       | 800 pixels wide |
| <code>pxheight=2000</code>     | 200 pixels high |

## Frequencies and percentages

| Parameter setting            | Effect             |
|------------------------------|--------------------|
| <code>vp=FALSE</code>        | Full denominator   |
| <code>showpct=FALSE</code>   | Do not show %      |
| <code>showcount=FALSE</code> | Do not show counts |

## Variable specification

| Suffix | Effect                                  |
|--------|---|
| #      | Variable names ending in numeric digits |
| *      | Variable names ending in any character  |
| @      | REDCap checklist variable names         |

| Prefix              | Effect                                       |
|---------------------|--|
| <code>is.na:</code> | Missing value?                               |
| <code>r:</code>     | REDCap checklist variable                    |
| <code>i:</code>     | Intersection of group of variables           |
| <code>any:</code>   | Are any of a group of variables affirmative? |
| <code>all:</code>   | Are all of a group of variables affirmative? |

## Text

| Parameter setting  |                                 |
|--|---------------------------------|
| <code>text=list(Category=c(triple="*"))</code>                     | Add * to all nodes of this type |
| <code>ttext=list(c(Group="A", Category="triple", text="*"))</code> | Add * to a specific node        |

## Formatting

`\n` line break    `*italics*`    `**bold**`    `%red ...%`

## Pattern trees and tables

| <code>vtree(FakeData, "Severity_Sex", pattern=T, varnamebold=T)</code> | <code>vtree(FakeData, "Severity_Sex", ptable=T)</code>  |          |     |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
|--|---|----------|-----|----------|-----|---|---|--------|---|---|---|------|---|---|---|------|---|---|---|--------|---|---|----|----------|---|---|----|------|---|----|----|------|---|----|----|----------|---|
|  | <table border="1"> <thead> <tr> <th>n</th> <th>pct</th> <th>Severity</th> <th>Sex</th> </tr> </thead> <tbody> <tr> <td>2</td> <td>4</td> <td>Severe</td> <td>F</td> </tr> <tr> <td>3</td> <td>7</td> <td>&lt;NA&gt;</td> <td>F</td> </tr> <tr> <td>3</td> <td>7</td> <td>&lt;NA&gt;</td> <td>M</td> </tr> <tr> <td>3</td> <td>7</td> <td>Severe</td> <td>M</td> </tr> <tr> <td>5</td> <td>11</td> <td>Moderate</td> <td>M</td> </tr> <tr> <td>8</td> <td>17</td> <td>Mild</td> <td>M</td> </tr> <tr> <td>11</td> <td>24</td> <td>Mild</td> <td>F</td> </tr> <tr> <td>11</td> <td>24</td> <td>Moderate</td> <td>F</td> </tr> </tbody> </table> | n        | pct | Severity | Sex | 2 | 4 | Severe | F | 3 | 7 | <NA> | F | 3 | 7 | <NA> | M | 3 | 7 | Severe | M | 5 | 11 | Moderate | M | 8 | 17 | Mild | M | 11 | 24 | Mild | F | 11 | 24 | Moderate | F |
| n  | pct   | Severity | Sex |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| 2  | 4   | Severe   | F   |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| 3  | 7   | <NA>     | F   |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| 3  | 7   | <NA>     | M   |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| 3  | 7   | Severe   | M   |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| 5  | 11  | Moderate | M   |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| 8  | 17  | Mild     | M   |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| 11   | 24  | Mild     | F   |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| 11   | 24  | Moderate | F   |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| <code>pattern</code>   | <code>Severity</code>   |          |     |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |
| <code>Severity</code>  | <code>Sex</code>  |          |     |          |     |   |   |        |   |   |   |      |   |   |   |      |   |   |   |        |   |   |    |          |   |   |    |      |   |    |    |      |   |    |    |          |   |

## Splitting text across lines

| Parameter setting          | Effect  |
|----------------------------|---|
| <code>splitwidth=50</code> | Split text in nodes after 50 characters         |
| <code>vsplitwidth=5</code> | Split text in variable names after 5 characters |

# xplain Cheat Sheet

## Important Links

- xplain package on CRAN <https://cran.r-project.org/web/packages/xplain/index.html>
- xplain web tutorial <http://www.zuckarelli.de/xplain/index.html>
- xplain cheat sheet [http://www.zuckarelli.de/xplain/xplain\\_cheatsheet.pdf](http://www.zuckarelli.de/xplain/xplain_cheatsheet.pdf)
- xplain on GitHub <https://www.github.com/jsugarelli/xplain>

## Purpose & Application

- xplain allows to **write interpretation/explanation texts** for statistical functions in the form of XML files.
- The user of the functions can read these explanations **while working on his/her specific problems**.
- xplain explanations **can react to the user's results** and provide meaningful insights related to the user's problem.
- For this, the xplain **XML files can contain R code** and can **work with the return object** of the user's function call.

> `xplain("lm(education ~ young + income + urban)")`  
 > Your R<sup>2</sup> is 0.11 which is quite low. There is a serious risk your model is misspecified. You should reconsider the selection of variables included in your model.

### xplain XML files

**1** Any valid xplain XML must be enclosed in an `<xplain>` block. Multiple `<xplain>` blocks per XML file are possible.

`<package>`

**2** A `<package>` block combines all functions from the same package.

`<function>`

**3** Within a `<function>` block, explanations/interpretations for the function as such or for specific elements of the return object can be provided.

`<result>`

**4** Packages explanations/ interpretations related to one element of the function's return object.

```

<xplain>
  1 <xplain>
    2 <package name = "stats">
      3 <function name = "lm">
        4 <title>This is about lm</title>
        5 <text>...</text>
        6 <result name = "coefficients">
          4 <title>...<title>
          5 <text>...</text>
        </result>
      </function>
    </package>
  </xplain>
</xml>
  
```

Not case-sensitive

**5** Structures explanations with headers.

`<text>`

**6** The actual explanations/interpretations. Can include R code with references to the function's return object.

### Main attributes: Overview

|              |  |
|--------------|--|
| <b>name</b>  | Name of the element (package, function, result).   |
| <b>lang</b>  | Language (ISO code) of the explanation (e.g. "EN").  |
| <b>level</b> | Complexity level; integer number; cumulative, i.e. Level=1 explanations will also be presented when Level=2 or Level=3 are called. |

### Attributes: Inheritance and necessity

- Elements **inherit attributes from higher-level** elements; e.g., if only one language, definition on `<xplain>` level suffices. Lower-level attributes overrule higher-level.
- name** attribute required for `<package>`, `<function>` and `<result>` elements.
- All levels shown, if no **level** is given to `xplain()`.

### Including R code

R code can be easily integrated into `<text></text>` elements:

```

<text> !%< R code %! </text>
  ↑   ↑
  R code delimiter tags
  
```

Access the explained function's (`<function name="...">`) return object:

- Access the full return object with `@`. Example: `summary(@)`.
- Access the current `<result name="...">` item of the return object with `##`. Example: `mean(##)`.

### Using placeholders

```

<define name= "placeholder" > !%< R code %! </define>
  ↓
</text> Text... !** "placeholder" **! Text... </text>
  ↑   ↑
  Placeholder name delimiter tags
  
```

Example: `<define name="s">!%< summary(@) %!</define>`  
`<text>And here is the summary !**! for your model</text>`

### Iterating through (items of) the return object

- To apply a `<text>` element to a whole matrix, data frame, vector or list, use the `foreach` attribute.
- Value of `foreach` defines what is iterated over and (for 2D structures) in which sequence; `items` is for lists.
- \$ is a placeholder for the index of the current element.
- Example** (shows all 1<sup>st</sup> column elements of the coefficient matrix):  
`<text foreach="rows">!%< $coefficients[,1] %!</text>`

`foreach =`  
 "rows"  
 "columns"  
 "rows, columns"  
 "columns, rows"  
 "items"  
 "items"

### Calling xplain()

|          |                    |  |
|----------|--------------------|--|
| <b>1</b> | <code>call</code>  | Call of the explained function as string                               |
|          | <code>xml</code>   | Path of the XML file providing the explanations                        |
|          | <code>lang</code>  | Language of the explanations to be shown (default means English)       |
|          | <code>level</code> | Complexity level of the explanations (cumulative! Default means "all") |

**2**  
 Wrapper function with  
`xplain.getcall()`

`Example: lm`  
`lm.xplain <- function(formula, data, subset, weights, na.action, method = "qr", model = TRUE, x = FALSE, y = FALSE, qr = TRUE, singular.ok = TRUE, contrasts = NULL, offset, ...) {`  
 `call <- xplain.getcall("lm")`  
 `xplain(call, xml = "http://www.zuckarelli.de/example_lm.xml")`