



Review

Short review on dopamine agonists: insight into clinical and research studies relevant to Parkinson's disease

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Abstract:

Parkinson's disease (PD) is a chronic and progressive neurological disorder characterized by selective degeneration of dopaminergic neurons (DAergic) in the substantia nigra pars compacta (SNpc) and subsequent decrease in dopamine (DA) levels in the striatum. Although levodopa replacement therapy is initially effective in symptomatic treatment of parkinsonian patients, its effectiveness often declines and various levodopa-related side effects appear after long-term treatment. The disabling side effects of levodopa therapy include motor fluctuations such as the wearing-off or on-off phenomena, dyskinesias and psychiatric symptoms. Nowadays, DA receptor agonists are often regarded as first choice in *de novo* and young parkinsonian patients to delay the onset of levodopa therapy. In advanced stages of the disease, they are also used as adjunct therapy together with levodopa to retard the development of motor complications. DA receptor agonists mimic the endogenous neurotransmitter, dopamine, and act by direct stimulation of presynaptic (autoreceptors) and postsynaptic DA receptors. Next to their clinical role in treating parkinsonian patients, laboratory studies reported antioxidative and neuron-rescuing effects of DA receptor agonists either *in vivo* or *in vitro*. This may involve reduced DA turnover following autoreceptor stimulation and direct free radical scavenging activity. In this review, we focus on and summarize the recently reported effects of the most commonly used DA agonists either in clinical or in research studies relevant to PD treatment.

Key words:

Parkinson's disease, levodopa, dopamine agonists, dopaminergic neurons

Abbreviations: AC – adenylyl cyclase, cAMP – cyclic adenosine monophosphate, DA – dopamine, DAergic – dopaminergic, DAT – dopamine transporter, GPe – globus pallidus pars externa, GPi – globus pallidus pars interna, GSH – glutathione, H₂O₂ – hydrogen peroxide, MAO-B – monoamine oxidase B, MPP⁺ – 1-methyl-4-phenylpyridinium, MPTP – 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine, NMDA – N-methyl-D-aspartate, 6-OHDA – 6-hydroxydopamine, PD – Parkinson's disease, SNpc – substantia nigra pars compacta, SNpr – substantia nigra pars reticulata, TH – tyrosine hydroxylase

Introduction

Parkinson's disease (PD) is a common progressive neurodegenerative disorder characterized by massive depletion of striatal dopamine (DA) as a result of degeneration of dopaminergic (DAergic) neurons in the substantia nigra. Clinically, the disease is manifested by bradykinesia, resting tremor, rigidity and distur-

bance of posture and gait [99]. To date, the etiopathogenesis of nigral DAergic neuron loss in PD is unclear. However, the presence of ongoing oxidative stress as the result of inefficient antioxidant defence mechanisms and generation of radical oxygen species in the substantia nigra pars compacta (SNpc) of the parkinsonian brain are important pathogenic mechanisms [2, 60, 142]. It should be noted that part of these free radicals are inevitably produced by dopamine metabolism in the brain either enzymatically through the action of monoamine oxidase-B (MAO-B) or by autooxidation [94]. Other sources of increased radical production may be endogenous neurotoxins occurring in the brain like tetrahydroisoquinolines or exogenously administered neurotoxins like the widely used herbicide paraquat which have similar neurochemical properties like the well-known neurotoxin, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) [7, 96]. Moreover, Antkiewicz-Michaluk [7] suggested that PD could be associated with excitotoxicity and apoptosis. Therefore, an effective anti-parkinsonian therapy should not only alleviate the disease-associated symptoms, but should also interfere with the progressive DAergic death in the substantia nigra.

The use of levodopa in PD

Treatment with levodopa

Since its introduction by Birkmayer and Hornykiewicz [15] levodopa remained the central pillar and the most effective drug for the symptomatic treatment of PD. Its prescription for parkinsonian patients is primarily based on its ability as a DA precursor to compensate for the decrease of DA in the brain. Although the initial use of levodopa replacement therapy is effective in symptomatic treatment of PD, the clinical efficacy often declines after long-term therapy and additionally disabling side-effects appear, most notably motor fluctuation such as the wearing-off or on-off phenomena and dyskinesia [23, 34]. These motor response complications appear in most patients with advanced PD treated with levodopa [23]. The precise mechanisms for the appearance of these treatment-related fluctuations are not clear. Nutt [91] reported that the long-duration response that characterizes the first few years of levodopa use in parkinsonian patients appeared to depend on the integrity of remaining DAergic nerve terminals in the striatum which retained the capacity to synthesize, release, reuptake

and store newly synthesized DA. After long-term use of levodopa and with progression of the disease, the short-duration response to levodopa and appearance of motor fluctuations are paralleled with more advanced DAergic denervation and loss of release and reuptake capacity [80]. Nonphysiological pulsatile stimulation of DA receptors seems to induce the development of motor fluctuations and dyskinesias whereas a more continuous stimulation might be associated with less fluctuation [136].

Effect of levodopa on dopaminergic cells

However DA replacement therapy with levodopa is successful to improve PD symptoms, it does not inhibit the progressive degeneration of DA neurons in SN [66]. Levodopa is not only ineffective against death of DAergic cells in PD patients, but there is also serious concern about possible toxic actions of levodopa on the remaining DAergic neurons. It has been reported that it was toxic to cultured DAergic neurons [4, 21, 46, 74, 90]. On the other hand, there is some evidence indicating that large doses of levodopa did not induce DA neuron degeneration in humans or normal mice and rats [101, 105]. In parkinsonian patients, it was speculated that the remaining DAergic neurons in the patient's brain could be particularly vulnerable to levodopa toxicity since they are hyperactive as a consequence of compensatory mechanisms [143]. In contrast, Dziewczapolski et al. [32] and Murer et al. [87] reported that treatment of rats with different degree of nigrostriatal damage for 6 months with oral levodopa was not toxic for the remaining DAergic neurons. Even when levodopa administration is started during an active degenerative process of DAergic neurons after intrastriatal 6-hydroxydopamine (6-OHDA) treatment, no aggravation of toxicity was found [40].

However, it is discussed that levodopa can enter other DAergic neurons, particularly of the mesolimbic pathway, which may result in psychotic symptoms and mood disturbance in some patients [141].

Mechanisms underlying levodopa toxicity

It was reported that increasing oxidative stress *via* autooxidation of levodopa plays an important role in levodopa toxicity. Spina and Cohen [120] and Fahn and Cohen [35] reported that the autooxidation and metabolism of levodopa can give rise to potentially

harmful free radical species, hydrogen peroxide (H_2O_2) and quinones. H_2O_2 played the most crucial role in the cascade of oxidative events induced by DA or levodopa towards SH-SY5Y cells [70]. Quinones were suggested to be responsible in part for the degeneration of non-DAergic neurons [97]. Pardo et al. [97] reported that the levels of quinones positively correlated with the severity of cell death in human neuroblastoma NB69 cells and at the same time the damage of DA neurons took place early before the rising of quinones. In addition to generation of H_2O_2 and quinone formation, levodopa-induced cell death may result from induction of apoptosis as evidenced by the increase in caspase-3 activity in Neuro-2A cells [100]. Taken together, levodopa-induced toxicity is related primarily to DA production. Excessive dopamine metabolism by high-dose levodopa therapy may promote oxidative stress and thereby accelerate the rate of neuronal degeneration either *in vivo* or *in vitro*. Interestingly, Muriel et al. [88, 89] observed that levodopa treatment of control and 6-OHDA-lesioned rats altered the localization of the D_1 DA receptor from the plasma membrane into the cytoplasm. They also reported that this effect was probably the result of D_1 DA receptor activation rather than that of DAergic denervation as such changes were observed on the lesioned and unlesioned side of the striatum of 6-OHDA-lesioned rats. The altered localization of D_1 receptors may participate in the occurrence of the side effects of levodopa therapy such as dyskinesia and fluctuations in motor performance.

Dopamine receptors and dopamine receptor agonists

Dopamine receptors

In brief, DA receptors belong to two classes of G protein-coupled receptors, the D_1 and D_2 classes. The classification of DA receptors was primarily based on their effects on adenylyl cyclase (AC) activity and cyclic adenosine monophosphate (cAMP) accumulation in the cells [63]. The D_1 receptor subtypes promote, whereas the D_2 subtypes inhibit AC activity and cAMP synthesis [84]. However, it has been reported that the D_2 receptors are mainly responsible for modulating the activity of voltage-sensitive Ca^{2+} and K^+ channels [119]. Moreover, molecular cloning confirmed that D_1 and D_2 receptors constituted two different classes of DA subtypes. In mammals, two re-

ceptor subtypes have been assigned to the D_1 receptor class (D_{1A}/D_1 and D_{1B}/D_5), but other types (D_{1C} and D_{1D}) exist in nonmammalian vertebrates [62] and similarly three types (D_2 , D_3 and D_4) of the D_2 class have been isolated [20]. DA receptor subtypes share similar sequences and structure [20]. Each type of DA receptors contains seven transmembrane domains with a unique binding site formed by the external loops of the protein [115].

The D_1 and D_2 receptors constitute the major DA receptor subtypes and they are differentially expressed in various regions of the human brain. The D_1 receptors are the most widespread receptors and are mainly expressed in the striatum, nucleus accumbens, olfactory tubercle, cerebral cortex and amygdala. It has also been detected in the island of Calleja and in the subthalamic nucleus [57, 58, 127]. The D_2 receptors are also mainly expressed in the striatum, nucleus accumbens and olfactory tubercle, but additionally in the SNpc and in the ventral tegmental area, where they presumably function as autoreceptors [58, 117, 127]. In the SNpr, binding of D_1 -specific ligands was demonstrated, but no mRNA was detected. Probably D_1 receptors are synthesized in the striatal neurons that project to the substantia nigra [58, 127]. Generally, D_1 receptors are mainly post-synaptic, while D_2 receptors are found post- and pre-synaptically (autoreceptor function) [57]. Functionally, there is some evidence that striatal D_1 receptors are expressed selectively by medium spiny neurons projecting to the globus pallidus pars interna (GPi) (direct pathway) [43, 53], while D_2 receptors are expressed by striatal medium spiny neurons projecting to the globus pallidus pars externa (GPe) (indirect pathway) [43, 53] as well as by both striatal cholinergic interneurons and dopaminergic nigrostriatal neurons (autoreceptors) [79, 85]. In accordance, occupation of D_1 receptors by DA or D_1 receptor agonists stimulates AC activity and stimulates the direct pathway. On the other hand, D_2 receptor occupation by DA or D_2 receptor agonists suppresses AC activity and inhibits neurons projecting from the striatum to GPe [3].

The D_5 receptor is localized in the SNpc, hypothalamus, striatum, cerebral cortex, nucleus accumbens and olfactory tubercle [64]. Also, it was reported that the D_5 receptor displayed higher affinity for DA than the D_1 subtype [49, 121]. The high affinity of the D_5 receptor for DA and its presence in the areas of DA pathways suggests that the D_5 receptor may participate in some activities of DAergic neurotransmission [67].

The D₃ receptor is localized in the forebrain limbic areas [119] and the largest receptor densities occur in granule cells of the islands of Calleja and in medium-sized spiny neurons of the rostral and ventromedial shell of the nucleus accumbens [29, 71]. D₃ receptor expression is low in the striatum. It is also expressed in the ventral pallidum, SNpc, the ventral tegmental area, mediodorsal thalamus, cerebral cortex, the cerebellum and amygdala in the rat brain [19, 29, 30]. There are some evident links between the alteration in dopamine D₃ receptor function and the etiology of a variety of CNS disorders, including schizophrenia and PD [75].

The D₄ receptor level is low in the basal ganglia and higher in the frontal cortex, medulla, amygdala, hypothalamus, hippocampus and mesencephalon [57].

In the rat central nervous system, the relative abundance of the DA receptors is D₁ > D₂ > D₃ > D₅ > D₄ [57].

Development of DA therapy with receptor agonists for treating PD

DA receptor agonists play an important role in anti-parkinsonian therapy and have become increasingly popular since the introduction of bromocriptine by Donald Calne and colleagues in 1974 [41]. Their development aimed at reducing the disabling motor complications produced by levodopa therapy [61]. More recently, DA receptor agonists are being used in the initial treatment of patients with *de novo* PD either as monotherapy or combined with low doses of levodopa [108]. Moreover, DA receptor agonists are advantageous in several aspects. They do not require carrier-mediated transport in the gut or brain. They act directly on the DA receptors without the need for metabolic modification, release or storage. They also have longer half-lives than levodopa and, therefore, they produce more persistent DA receptor stimulation than levodopa. Their metabolism does not generate free radicals which are considered one of the most important hazards in levodopa treatment particularly on DAergic neurons [46]. The most important DA receptor agonists which are currently approved and gained access into the clinical and research studies are ergoline derivatives such as bromocriptine, lisuride, pergolide, cabergoline and α -dihydroergocriptine as well as the non-ergoline derivatives like rotigotine, pramipexole, ropinirole and apomorphine. Ergolines, derivatives of ergot alkaloids, have a longer history in

anti-parkinsonian therapy and are as effective as non-ergolines, which were developed in the hope that they might provide the benefits of the ergoline agents without their side effects [17]. However, recently valvular heart disease was presented as a new complication of ergot derivative DA receptor agonists [55, 107, 118, 133] but non-ergot DA receptor agonists cannot be excluded at present due to lack of sufficient reliable pharmacoepidemiological data [24]. Partly, the individual DA receptor agonists show significant variation in their receptor affinity [44].

General effects of DA receptor agonists

DA receptor agonists are initially prescribed at the early stage of parkinsonism to postpone the onset of levodopa therapy. They are co-administered with low doses of levodopa to delay the development and to minimize the severity of levodopa-associated treatment complications [12, 110]. There is great evidence that stimulation of D₂ receptors at the post-synaptic level is primarily involved in alleviating levodopa-induced motor fluctuations. Additionally, direct receptor activation with an agonist might be expected to elicit more specific and controllable effects than those produced by the transmitter precursor levodopa, in patients with a damaged extrapyramidal system [41]. More recently, there is increasing evidence in the literature that DA agonists are not only beneficial to postpone levodopa therapy in early parkinsonism or to counteracting its complications after long-term use but they have also been suggested to be neuroprotective particularly in experimental models [72, 103]. The mechanisms and processes underlying the neuroprotective actions of DA receptor agonists appear to be interlaced. They spare levodopa, thereby reducing the formation of oxidative radicals from levodopa metabolism, act as radical scavengers, reduce DA synthesis, release and metabolism *via* activating pre-synaptic autoreceptors, ameliorate excitotoxicity by suppressing subthalamic nucleus overactivity and exert antiapoptotic effects [113].

Interestingly, DA receptor agonists have been reported to play an important neurogenic modulatory role in the central nervous system (CNS) development [129]. In line with this, Höglinger et al. [54] observed that experimental depletion of dopamine in rodents decreased precursor cell proliferation in both subependymal and subgranular zones in the adult. The highly proliferative precursors in subependymal zone

express DA receptors and receive DAergic afferents. More specifically, DA D₃ receptor mRNA expression in CNS occurs quite early in development and is predominantly found in the proliferative neuroepithelium [31]. In addition to *in vivo* reports, *in vitro* studies have shown that DA increased ³H thymidine incorporation (a marker of cell proliferation) in a neuroblastoma-glioma hybrid cell line transfected with the human D₃ receptor [102]. In contrast to the involvement of D₃ receptor early in neurogenesis, D₂ and D₁ receptor mRNAs occur later in the development and appear predominantly in migrating and differentiating neurons [31]. Moreover, in neurosphere cultures prepared from neural precursors in the adult subependymal zone, activation of dopamine D₂-like receptors directly increased the proliferation of these precursors [54].

Ergoline DA receptor agonists

Bromocriptine

Bromocriptine is the first DA receptor agonist that has been approved for anti-parkinsonian therapy since 1974 [41]. It was first used as adjunct therapy to levodopa in patients experiencing motor fluctuations and later was recommended as monotherapy in the early stage of the disease [134]. Bromocriptine has been shown to protect mice and DAergic cells against 6-OHDA and MPTP, and levodopa-induced cell loss, respectively. It also attenuated DA depletion in mouse striatum in response to methamphetamine [68, 92, 122]. The neuroprotective effect of bromocriptine is dependent on both, its action as a D₂ receptor agonist and its antioxidant capacity. In this context, it has been reported that bromocriptine is able to scavenge hydroxyl and superoxide radicals *in vitro* [92, 138] and to inhibit hydroxyl radical formation and lipid peroxidation *in vivo* [92].

Pergolide

Pergolide, a semi-synthetic ergoline derivative, is a potent DA receptor agonist used as adjunct to levodopa therapy to slow the clinical progression of PD or to prevent the levodopa-associated motor complications [47, 59]. However, its clinical use is already of minor importance now, since it was recently shown to be associated with the development of restrictive valvular heart disease [10, 55, 128]. Pergolide is ab-

sorbed rapidly following oral dosing reaching peak plasma concentration within 2–3 h, has a long half-life of about 21 h and is completely eliminated within 4–5 days [16]. Accordingly, it should produce a more physiological and continuous stimulation of DA receptors avoiding or delaying the induction of dyskinesia [16]. The neuroprotective effect of pergolide has been observed either *in vivo* or *in vitro*. For instance, chronic administration of pergolide preserved the integrity of nigrostriatal neurons in the aging rat's brain [38] and protected against reduction of striatal DA and its metabolites after 6-OHDA injection in mice [9]. Using cell culture models, pergolide has been shown to promote the survival of DAergic neurons, to exhibit partial protection against 1-methyl-4-phenylpyridinium iodide (MPP⁺) toxicity and to increase ³H-dopamine uptake by cultured cells after levodopa treatment [46]. Moreover, pergolide had protective effect against H₂O₂ in SH-SY5Y neuroblastoma cells [126]. The neuroprotective effect of pergolide has been shown to be mediated by free radical scavenging activity particularly hydroxyl radicals and nitric oxide and by decreasing phospholipid peroxidation [48, 95], suppressing apoptotic pathways through inhibiting of NF-κB nuclear translocation [125] and stabilizing the mitochondrial function [46]. In clinical trials pergolide was shown to reduce the long-term decline of striatal fluorodopa uptake compared to levodopa treatment, however, without reaching significance [116].

α-Dihydroergocriptine

α-Dihydroergocriptine is a synthetic hydrogenated ergot derivative with a strong dopamino-mimetic activity *in vivo* and *in vitro* [104]. It showed high efficacy and tolerance in treatment of PD [12, 13, 18, 77, 83] and revised PD-like symptoms in experimental models [27]. It was shown that α-dihydroergocriptine reduced abnormal motor behavior and neuronal degeneration induced by MPTP in monkeys [14], protected mice against convulsions induced by intracerebroventricular injection of glutamate and increased the survival time in mice submitted to total cerebral ischemia induced by MgCl₂ [27]. In cellular models, α-dihydroergocriptine protected cultured rat cerebral granule cells against age-dependent and glutamate-induced neuronal cell death [37] and increased the survival of primary cultured DAergic cells when co-administered with either levodopa or DA [45]. The neuroprotective

activity of α -dihydroergocriptine may be based on activation of dopamine D₂ receptors, interaction with excitatory amino acids, influencing the glutathione redox index, improving of cellular energy metabolism, decreasing of lipoperoxidative cellular degeneration, radical scavenging activity, and enhancement of antioxidant enzymes [27, 36, 45, 78]. Moreover, α -dihydroergocriptine increased the number of astrocytes in monkeys after MPTP treatment. Astrocyte activation is important for restoration of neuronal function by enhancing neuronal survival and axon growth in injured areas [86].

Cabergoline

Cabergoline, an ergoline DA receptor agonist, has a relatively long average elimination half-life of 65–110 h as estimated by data on urinary excretion rates in healthy volunteers and in parkinsonian patients, compared with other DA receptor agonists [137]. It was reported that cabergoline is well tolerated and effective in controlling parkinsonism particularly in the early stage of the disease. When combined with levodopa therapy, it was highly effective in improving motor disability without inducing hyperactivity or dyskinesia in cynomolgus monkeys [8]. Like other ergot derivative DA receptor agonists, cabergoline showed neuroprotective potential. It was reported that cabergoline protected nigrostriatal DAergic neurons against 6-OHDA in mice [139]. Activation of GSH, catalase, superoxide dismutase, direct free radical scavenging activity and stimulation of neurotrophic factors are important in mediating the neuroprotective action of cabergoline [93, 139].

Non-ergoline dopamine receptor agonists

Pramipexole

Pramipexole, a synthetic amino-benzothiazole derivative, has a potent agonist activity for D₂ and D₃ receptors [81, 82]. The rank order of receptor affinity of pramipexole within the D₂ subfamily is D₃ > D₂ according to saturation binding experiments [44]. On the other hand, pramipexole is lacking affinity for DA D₁ and D₅ receptors [67].

Clinically, it was reported that the safety profile of pramipexole is similar to that of the ergot-derived compounds and it was effective as monotherapy in early parkinsonism and as adjunct therapy with levo-

dopa in advanced stages of the disease [109]. Pramipexole has been found to slow the rate of loss of striatal dopamine transporter (DAT) density as measured in PD patients with the single photon emission computed tomography ligand ¹²³I β -carbomethoxy-3 β (4-iodophenyl)tropane [76, 98]. Furthermore, there are some clinical trials suggesting that pramipexole not only ameliorated motor symptoms, but also the depressive symptoms in PD [109].

Beside the effective role of pramipexole in treating parkinsonian symptoms, it showed a variety of neuroprotective effects in *in vivo* and *in vitro* experimental paradigms [6, 22, 39, 73, 132]. For example, pramipexole inhibited the depletion of striatal DA content in mice [106] and after MPTP treatment. In *in vitro* systems, it has been shown that pramipexole increased the survival of rat mesencephalic cells exposed to levodopa [21], protected SHSY-5Y neuroblastoma cells against MPP⁺ and rotenone [51] and rescued cerebellar granule cells from levodopa toxicity [131]. The neuroprotective effect of pramipexole seemed to be derived from different mechanisms. Several reports claimed that the neuroprotective effects of pramipexole are the result of the antioxidant properties [39, 73, 130, 144], inhibition of apoptotic pathways [1, 22, 65] and induction of neurotrophic factors [67]. Moreover, Carvey et al. [21] suggested that the neuroprotective effect of pramipexole may be mediated, in part, by interaction with the D₃ receptor. This hypothesis was further investigated *in vivo* by Ramirez et al. [106]. They reported that pramipexole was apparently less effective in inhibiting striatal DA depletion as a result of MPTP treatment in D₃ receptor knockout mice than in mice expressing the receptor. Moreover, the D₃ receptor antagonist, A-437203, was found to partially inhibit the protective effect of pramipexole against MPTP-induced striatal DA. Conclusively, D₃ receptor-dependent and -independent mechanisms seem to play a role against MPTP toxicity in this model.

Ropinirole

Ropinirole is a non-ergoline DA receptor agonist that exhibits a high affinity for the D₂ and D₃ receptors but little or no affinity for the D₁ receptor [25, 33]. Ropinirole is rapidly absorbed and well tolerated after oral administration [69]. Symptomatically, it was reported that ropinirole was as effective as bromocriptine in reducing motor complications and decreas-

ing levodopa dose without increasing adverse events including dyskinesia [26]. Also, ropinirole monotherapy was effective in treating resting tremor in early PD [114], in reducing periodic leg movements and in improving sleep efficiency in patients with restless legs syndrome [5, 25]. These positive effects of ropinirole in PD are believed to be due to stimulation of the post-synaptic dopamine D₂-type receptor [33]. In experimental models for PD, it has been found that ropinirole reversed the motor and behavioral deficits induced by MPTP in marmosets [33] and showed neuroprotective effect against 6-OHDA in mice [56]. Activation of GSH and GSH-regulating enzymes such as glutathione peroxidase, glutathione reductase and glutathione transferase as well as activation of catalase and superoxide dismutase were principal neuroprotective mechanisms mediated by ropinirole [123]. In clinical trials ropinirole reduced the long-term decline of striatal fluorodopa uptake compared to levodopa therapy indicating a preserving effect on terminal function of DAergic neurons [135].

Apomorphine

Apomorphine, a non-ergoline DA receptor agonist, is a short-acting and non-selective dopamine D₁/D₂ receptor agonist [28]. It was the first DA receptor agonist used for treating PD [28]. It has been recently reported that subcutaneous intermittent injections or continuous infusions of apomorphine are currently used for the management of sudden, unexpected and refractory levodopa-induced "off" states in fluctuating PD [28]. Like other DA receptor agonists, there is some evidence in the literature describing the neuroprotective potential of apomorphine in experimental models. It has been shown that apomorphine exhibited neuroprotection against DA depletion in 6-OHDA lesioned-rats [142] and MPTP-treated mice [50]. Furthermore, Battaglia et al. [11] and Yuan et al. [142] reported that continuous subcutaneous infusion of apomorphine rescued striatal DAergic terminals and increased the TH and DAT immunoreactivity against toxicity induced by MPTP in mice and enhanced the number of TH⁺ cells in the ventral tegmental area in partially 6-OHDA-lesioned rats, respectively. It has been observed that apomorphine increased the survival of cultured mesencephalic DAergic cells [52]. These findings provided an evidence for trophic effects of apomorphine either *in vivo* or *in vitro*. The neuroprotective effect of apomorphine could be

a consequence of antioxidant activity [42, 112], potent iron chelating action [124], inhibition of lipid peroxidation [140], induction of neurotrophic factors [111] and anti-inflammatory effects [50].

Concluding remarks

Since the introduction of bromocriptine in 1974, great attention has been paid to DA receptor agonists for treating PD. They are successfully used in the initial treatment of patients with *de novo* PD either as monotherapy or combined with low doses of levodopa. The beneficial effects of DA receptor agonists as anti-parkinsonian drugs is based on their ability to produce direct and continuous stimulation of both presynaptic (autoreceptors) and postsynaptic DA receptors. More recently, DA receptor agonists have been shown to produce neuroprotective activities in experimental models either *in vivo* or *in vitro*. However, there is shortage in the literature concerning the neuroprotective role of DA receptor agonists in parkinsonian patients as there is no objective method to check thoroughly for DAergic cell death. Taken together, we can say that there is no drug that has been unequivocally proven to be neuroprotective in the parkinsonian patient until now. On the other hand, DA receptor agonists are promising in this respect and ongoing research with more accurate neuroimaging devices should decrease the distance between the data obtained from experimental models and clinical studies.

References:

1. Abramova NA, Cassarino DS, Khan SM, Painter TW, Bennett JP Jr.: Inhibition by R(+) or S(-) pramipexole of caspase activation and cell death induced by methylpyridinium ion or beta amyloid peptide in SH-SY5Y neuroblastoma. *J Neurosci Res*, 2002, 67, 494–500.
2. Adams JD Jr., Odunze IN: Oxygen free radicals and Parkinson's disease. *Free Radic Biol Med*, 1991, 10, 161–169.
3. Albin RL, Young AB, Penney JB: The functional anatomy of basal ganglia disorders. *Trends Neurosci*, 1989, 12, 366–375.
4. Alexander T, Sortwell CE, Sladek CD, Roth RH, Steece-Collier K: Comparison of neurotoxicity following repeated administration of L-dopa, D-dopa and dopamine to embryonic mesencephalic dopamine neurons in

- cultures derived from Fisher 344 and Sprague-Dawley donors. *Cell Transplant*, 1997, 6, 309–315.
5. Allen R, Becker PM, Bogan R, Schmidt M, Kushida CA, Fry JM, Poceta JS, Winslow D: Ropinirole decreases periodic leg movements and improves sleep parameters in patients with restless legs syndrome. *Sleep*, 2004, 27, 907–914.
 6. Anderson DW, Neavin T, Smith JA, Schneider JS: Neuroprotective effects of pramipexole in young and aged MPTP-treated mice. *Brain Res*, 2001, 905, 44–53.
 7. Antkiewicz-Michaluk L: Endogenous risk factors in Parkinson's disease: dopamine and tetrahydroisoquinolines. *Pol J Pharmacol*, 2002, 54, 567–572.
 8. Arai N, Isaji M, Kojima M, Mizuta E, Kuno S: Combined effects of cabergoline and L-dopa on parkinsonism in MPTP-treated cynomolgus monkeys. *J Neural Transm*, 1996, 103, 1307–1316.
 9. Asanuma M, Ogawa N, Nishibayashi S, Kawai M, Kondo Y, Iwata E: Protective effects of pergolide on dopamine levels in the 6-hydroxydopamine-lesioned mouse brain. *Arch Int Pharmacodyn Ther*, 1995, 329, 221–230.
 10. Baseman DG, O'Suilleabhain PE, Reimold SC, Laskar SR, Baseman JG, Dewey RB Jr: Pergolide use in Parkinson's disease is associated with cardiac valve regurgitation. *Neurology*, 2004, 63, 301–304.
 11. Battaglia G, Busceti CL, Cuomo L, Giorgi FS, Orzi F, De Blasi A, Nicoletti F et al.: Continuous subcutaneous infusion of apomorphine rescues nigro-striatal dopaminergic terminals following MPTP injection in mice. *Neuropharmacology*, 2002, 42, 367–373.
 12. Battistin L, Bardin PG, Ferro-Milone F, Ravenna C, Toso V, Reboldi G: Alpha-dihydroergocryptine in Parkinson's disease: a multicentre randomized double blind parallel group study. *Acta Neurol Scand*, 1999, 99, 36–42.
 13. Bergamasco B, Fratola L, Muratorio A, Piccoli F, Mailand F, Parnetti L: Alpha-dihydroergocryptine in the treatment of *de novo* parkinsonian patients: results of a multicentre, randomized, double-blind, placebo-controlled study. *Acta Neurol Scand*, 2000, 101, 372–380.
 14. Bernocchi G, Gerzeli G, Scherini E, Vignola C: Neuroprotective effects of alpha-dihydroergocryptine against damages in the substantia nigra caused by severe treatment with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine. *Acta Neuropathol (Berl)*, 1993, 85, 404–413.
 15. Birkmayer W, Hornykiewicz O: The effect of 1-3,4-dihydroxyphenylalanine (= DOPA) on akinesia in parkinsonism. 1961. *Wien Klin Wochenschr*, 2001, 113, 851–854.
 16. Blin O: The pharmacokinetics of pergolide in Parkinson's disease. *Curr Opin Neurol*, 2003, 16, Suppl 1, S9–S12.
 17. Bonuccelli U: Comparing dopamine agonists in Parkinson's disease. *Curr Opin Neurol*, 2003, 16, Suppl 1, S13–S19.
 18. Bonuccelli U, D'Antonio P, D'Avino C, Piccini P, Muratorio A: Dihydroergocryptine in the treatment of Parkinson's disease. *J Neural Transm Suppl*, 1995, 45, 239–245.
 19. Bouthenet ML, Souil E, Martres MP, Sokoloff P, Giros B, Schwartz JC: Localization of dopamine D3 receptor mRNA in the rat brain using *in situ* hybridization histochemistry: comparison with dopamine D2 receptor mRNA. *Brain Res*, 1991, 564, 203–219.
 20. Callier S, Snappy M, Le Crom S, Prou D, Vincent JD, Vernier P: Evolution and cell biology of dopamine receptors in vertebrates. *Biol Cell*, 2003, 95, 489–502.
 21. Carvey PM, Pieri S, Ling ZD: Attenuation of levodopa-induced toxicity in mesencephalic cultures by pramipexole. *J Neural Transm*, 1997, 104, 209–228.
 22. Cassarino DS, Fall CP, Smith TS, Bennett JP Jr: Pramipexole reduces reactive oxygen species production *in vivo* and *in vitro* and inhibits the mitochondrial permeability transition produced by the parkinsonian neurotoxin methylpyridinium ion. *J Neurochem*, 1998, 71, 295–301.
 23. Chase TN, Mouradian MM, Engber TM: Motor response complications and the function of striatal efferent systems. *Neurology*, 1993, 43, S23–S27.
 24. Chaudhuri KR, Dhawan V, Basu S, Jackson G, Odin P: Valvular heart disease and fibrotic reactions may be related to ergot dopamine agonists, but non-ergot agonists may also not be spared. *Mov Disord*, 2004, 19, 1522–1523.
 25. Cheer SM, Bang LM, Keating GM: Ropinirole: for the treatment of restless legs syndrome. *CNS Drugs*, 2004, 18, 747–754.
 26. Clarke CE, Deane KH: Ropinirole versus bromocriptine for levodopa-induced complications in Parkinson's disease. *Cochrane Database Syst Rev*, 2001, CD001517.
 27. Coppi G: Neuroprotective activity of alpha-dihydroergocryptine in animal models. *J Neural Transm Suppl*, 1995, 45, 307–318.
 28. Deleu D, Hanssens Y, Northway MG: Subcutaneous apomorphine: an evidence-based review of its use in Parkinson's disease. *Drugs Aging*, 2004, 21, 687–709.
 29. Diaz J, Levesque D, Griffon N, Lammers CH, Martres MP, Sokoloff P, Schwartz JC: Opposing roles for dopamine D2 and D3 receptors on neurotensin mRNA expression in nucleus accumbens. *Eur J Neurosci*, 1994, 6, 1384–1387.
 30. Diaz J, Levesque D, Lammers CH, Griffon N, Martres MP, Schwartz JC, Sokoloff P: Phenotypical characterization of neurons expressing the dopamine D3 receptor in the rat brain. *Neuroscience*, 1995, 65, 731–745.
 31. Diaz J, Ridray S, Mignon V, Griffon N, Schwartz JC, Sokoloff P: Selective expression of dopamine D3 receptor mRNA in proliferative zones during embryonic development of the rat brain. *J Neurosci*, 1997, 17, 4282–4292.
 32. Dziewczapolski G, Murer G, Agid Y, Gershanik O, Raisman-Vozari R: Absence of neurotoxicity of chronic L-DOPA in 6-hydroxydopamine-lesioned rats. *Neuroreport*, 1997, 8, 975–979.
 33. Eden RJ, Costall B, Domeney AM, Gerrard PA, Harvey CA, Kelly ME, Naylor RJ, Owen DA, Wright A: Pre-clinical pharmacology of ropinirole (SK&F 101468-A) a novel dopamine D2 agonist. *Pharmacol Biochem Behav*, 1991, 38, 147–154.
 34. Fahn S: Controversies in the therapy of Parkinson's disease. *Adv Neurol*, 1996, 69, 477–486.
 35. Fahn S, Cohen G: The oxidant stress hypothesis in Parkinson's disease: evidence supporting it. *Ann Neurol*, 1992, 32, 804–812.
 36. Favit A, Sortino MA, Aleppo G, Scapagnini U, Canonico PL: Protection by dihydroergocryptine of

- glutamate-induced neurotoxicity. *Pharmacol Toxicol*, 1993, 73, 224–228.
37. Favit A, Sortino MA, Aleppo G, Scapagnini U, Canonico PL: The inhibition of peroxide formation as a possible substrate for the neuroprotective action of dihydroergocryptine. *J Neural Transm Suppl*, 1995, 45, 297–305.
 38. Felten DL, Felten SY, Fuller RW, Romano TD, Smalstig EB, Wong DT, Clemens JA: Chronic dietary pergolide preserves nigrostriatal neuronal integrity in aged-Fischer-344 rats. *Neurobiol Aging*, 1992, 13, 339–351.
 39. Ferger B, Teismann P, Mierau J: The dopamine agonist pramipexole scavenges hydroxyl free radicals induced by striatal application of 6-hydroxydopamine in rats: an *in vivo* microdialysis study. *Brain Res*, 2000, 883, 216–223.
 40. Ferrario JE, Delfino MA, Stefano AV, Zbarsky V, Douhou A, Murer MG, Raisman-Vozari R, Gershanik OS: Effects of orally administered levodopa on mesencephalic dopaminergic neurons undergoing a degenerative process. *Neurosci Res*, 2003, 47, 431–436.
 41. Foley P, Gerlach M, Double KL, Riederer P: Dopamine receptor agonists in the therapy of Parkinson's disease. *J Neural Transm*, 2004, 111, 1375–1446.
 42. Gassen M, Glinka Y, Pinchasi B, Youdim MB: Apomorphine is a highly potent free radical scavenger in rat brain mitochondrial fraction. *Eur J Pharmacol*, 1996, 308, 219–225.
 43. Gerfen CR, Engber TM, Mahan LC, Susel Z, Chase TN, Monsma FJ Jr, Sibley DR: D1 and D2 dopamine receptor-regulated gene expression of striatonigral and striatopallidal neurons. *Science*, 1990, 250, 1429–1432.
 44. Gerlach M, Double K, Arzberger T, Leblhuber F, Tatschner T, Riederer P: Dopamine receptor agonists in current clinical use: comparative dopamine receptor binding profiles defined in the human striatum. *J Neural Transm*, 2003, 110, 1119–1127.
 45. Gille G, Radad K, Reichmann H, Rausch WD: a-Dihydroergocryptine combined with L-DOPA or dopamine promotes the survival of dopaminergic neurons in culture. *J Neural Transm*, 2005, (in press).
 46. Gille G, Rausch WD, Hung ST, Moldzio R, Janetzky B, Hundemer HP, Kolter T, Reichmann H: Pergolide protects dopaminergic neurons in primary culture under stress conditions. *J Neural Transm*, 2002, 109, 633–643.
 47. Goetz CG, Diederich NJ: Dopaminergic agonists in the treatment of Parkinson's disease. *Neurol Clin*, 1992, 10, 527–540.
 48. Gomez-Vargas M, Nishibayashi-Asanuma S, Asanuma M, Kondo Y, Iwata E, Ogawa N: Pergolide scavenges both hydroxyl and nitric oxide free radicals *in vitro* and inhibits lipid peroxidation in different regions of the rat brain. *Brain Res*, 1998, 790, 202–208.
 49. Grandy DK, Zhang YA, Bouvier C, Zhou QY, Johnson RA, Allen L, Buck K et al.: Multiple human D5 dopamine receptor genes: a functional receptor and two pseudogenes. *Proc Natl Acad Sci USA*, 1991, 88, 9175–9179.
 50. Grunblatt E, Mandel S, Berkuzki T, Youdim MB: Apomorphine protects against MPTP-induced neurotoxicity in mice. *Mov Disord*, 1999, 14, 612–618.
 51. Gu M, Irvani M, Cooper JM, King D, Jenner P, Schapira AH: Pramipexole protects against apoptotic cell death by non-dopaminergic mechanisms. *J Neurochem*, 2004, 91, 1075–1081.
 52. Guo H, Tang Z, Yu Y, Xu L, Jin G, Zhou J: Apomorphine induces trophic factors that support fetal rat mesencephalic dopaminergic neurons in cultures. *Eur J Neurosci*, 2002, 16, 1861–1870.
 53. Harrison MB, Wiley RG, Wooten GF: Selective localization of striatal D1 receptors to striatonigral neurons. *Brain Res*, 1990, 528, 317–322.
 54. Hoglinger GU, Rizk P, Muriel MP, Duyckaerts C, Oertel WH, Caille I, Hirsch EC: Dopamine depletion impairs precursor cell proliferation in Parkinson disease. *Nat Neurosci*, 2004, 7, 726–735.
 55. Horvath J, Fross RD, Kleiner-Fisman G, Lerch R, Stalder H, Liaudat S, Raskoff WJ et al.: Severe multivalvular heart disease: a new complication of the ergot derivative dopamine agonists. *Mov Disord*, 2004, 19, 656–662.
 56. Iida M, Miyazaki I, Tanaka K, Kabuto H, Iwata-Ichikawa E, Ogawa N: Dopamine D2 receptor-mediated antioxidant and neuroprotective effects of ropinirole, a dopamine agonist. *Brain Res*, 1999, 838, 51–59.
 57. Jaber M, Robinson SW, Missale C, Caron MG: Dopamine receptors and brain function. *Neuropharmacology*, 1996, 35, 1503–1519.
 58. Jackson DM, Westlind-Danielsson A: Dopamine receptors: molecular biology, biochemistry and behavioral aspects. *Pharmacol Ther*, 1994, 64, 291–370.
 59. Jankovic J, Orman J: Parallel double-blind study of pergolide in Parkinson's disease. *Adv Neurol*, 1987, 45, 551–554.
 60. Jenner P: Dopamine agonists, receptor selectivity and dyskinesia induction in Parkinson's disease. *Curr Opin Neurol*, 2003, 16, S3–S7.
 61. Jenner P, Olanow CW: Oxidative stress and the pathogenesis of Parkinson's disease. *Neurology*, 1996, 47, S161–S170.
 62. Kapsimali M, Vidal B, Gonzalez A, Dufour S, Vernier P: Distribution of the mRNA encoding the four dopamine D(1) receptor subtypes in the brain of the European eel (*Anguilla anguilla*): comparative approach to the function of D(1) receptors in vertebrates. *J Comp Neurol*, 2000, 419, 320–343.
 63. Kebabian JW, Calne DB: Multiple receptors for dopamine. *Nature*, 1979, 277, 93–96.
 64. Khan ZU, Gutierrez A, Martin R, Penafiel A, Rivera A, de la CA: Dopamine D5 receptors of rat and human brain. *Neuroscience*, 2000, 100, 689–699.
 65. Kitamura Y, Kosaka T, Kakimura JI, Matsuoka Y, Kohno Y, Nomura Y, Taniguchi T: Protective effects of the antiparkinsonian drugs talipexole and pramipexole against 1-methyl-4-phenylpyridinium-induced apoptotic death in human neuroblastoma SH-SY5Y cells. *Mol Pharmacol*, 1998, 54, 1046–1054.
 66. Kitamura Y, Taniguchi T, Shimohama S, Akaike A, Nomura Y: Neuroprotective mechanisms of antiparkinsonian dopamine D2-receptor subfamily agonists. *Neurochem Res*, 2003, 28, 1035–1040.
 67. Kohno Y, Takeuchi S: Pharmacological profiles and clinical effects of antiparkinsonian agent, pramipexole (Japanese). *Nippon Yakurigaku Zasshi*, 2004, 123, 429–440.

68. Kondo T, Ito T, Sugita Y: Bromocriptine scavenges methamphetamine-induced hydroxyl radicals and attenuates dopamine depletion in mouse striatum. *Ann NY Acad Sci*, 1994, 738, 222–229.
69. Kuzel MD: Ropinirole: a dopamine agonist for the treatment of Parkinson's disease. *Am J Health Syst Pharm*, 1999, 56, 217–224.
70. Lai CT, Yu PH: Dopamine- and L-beta-3,4-dihydroxyphenylalanine hydrochloride (L-Dopa)-induced cytotoxicity towards catecholaminergic neuroblastoma SH-SY5Y cells. Effects of oxidative stress and antioxidative factors. *Biochem Pharmacol*, 1997, 53, 363–372.
71. Le Moine C, Bloch B: Expression of the D3 dopamine receptor in peptidergic neurons of the nucleus accumbens: comparison with the D1 and D2 dopamine receptors. *Neuroscience*, 1996, 73, 131–143.
72. Le WD, Jankovic J: Are dopamine receptor agonists neuroprotective in Parkinson's disease? *Drugs Aging*, 2001, 18, 389–396.
73. Le WD, Jankovic J, Xie W, Appel SH: Antioxidant property of pramipexole independent of dopamine receptor activation in neuroprotection. *J Neural Transm*, 2000, 107, 1165–1173.
74. Ling ZD, Pieri SC, Carvey PM: Comparison of the neurotoxicity of dihydroxyphenylalanine stereoisomers in cultured dopamine neurons. *Clin Neuropharmacol*, 1996, 19, 360–365.
75. Luedtke RR, Mach RH: Progress in developing D3 dopamine receptor ligands as potential therapeutic agents for neurological and neuropsychiatric disorders. *Curr Pharm Des*, 2003, 9, 643–671.
76. Marek K, Jennings D, Seibyl J: Single-photon emission tomography and dopamine transporter imaging in Parkinson's disease. *Adv Neurol*, 2003, 91, 183–191.
77. Martignoni E, Pacchetti C, Sibilla L, Bruggi P, Pedevilla M, Nappi G: Dihydroergocryptine in the treatment of Parkinson's disease: a six months' double-blind clinical trial. *Clin Neuropharmacol*, 1991, 14, 78–83.
78. Medico M, De Vivo S, Tomasello C, Grech M, Nicosia A, Castorina M, D'Agata MA et al.: Behavioral and neurochemical effects of dopaminergic drugs in models of brain injury. *Eur Neuropsychopharmacol*, 2002, 12, 187–194.
79. Mengod G, Martinez-Mir MI, Vilaro MT, Palacios JM: Localization of the mRNA for the dopamine D2 receptor in the rat brain by *in situ* hybridization histochemistry. *Proc Natl Acad Sci USA*, 1989, 86, 8560–8564.
80. Metman LV, Konitsiotis S, Chase TN: Pathophysiology of motor response complications in Parkinson's disease: hypotheses on the why, where, and what. *Mov Disord*, 2000, 15, 3–8.
81. Mierau J, Schingnitz G: Biochemical and pharmacological studies on pramipexole, a potent and selective dopamine D2 receptor agonist. *Eur J Pharmacol*, 1992, 215, 161–170.
82. Mierau J, Schneider FJ, Ensinger HA, Chio CL, Lajiness ME, Huff RM: Pramipexole binding and activation of cloned and expressed dopamine D2, D3 and D4 receptors. *Eur J Pharmacol*, 1995, 290, 29–36.
83. Minea D, Varga I, Falup-Pecurariu C, de Mey C, Retzow A, Althaus M: Influence of the dopamine agonist alpha-dihydroergocryptine on the pharmacokinetics of levodopa in patients with Parkinson's disease. *Clin Neuropharmacol*, 2001, 24, 235–238.
84. Missale C, Nash SR, Robinson SW, Jaber M, Caron MG: Dopamine receptors: from structure to function. *Physiol Rev*, 1998, 78, 189–225.
85. Morelli M, Mennini T, Di Chiara G: Nigral dopamine autoreceptors are exclusively of the D2 type: quantitative autoradiography of [125I]iodosulpride and [125I]SCH 23982 in adjacent brain sections. *Neuroscience*, 1988, 27, 865–870.
86. Muler HV, Matthiessen HP, Schmalenbach C, Schroeder WO: Glial support of CNS neuronal survival, neurite outgrowth and regeneration. *Restor Neurol Neurosci*, 1991, 2, 229–232.
87. Murer MG, Dziewczapolski G, Menalled LB, Garcia MC, Agid Y, Gershanik O, Raisman-Vozari R: Chronic levodopa is not toxic for remaining dopamine neurons, but instead promotes their recovery, in rats with moderate nigrostriatal lesions. *Ann Neurol*, 1998, 43, 561–575.
88. Muriel MP, Bernard V, Levey AI, Laribi O, Abrous DN, Agid Y, Bloch B, Hirsch EC: Levodopa induces a cytoplasmic localization of D1 dopamine receptors in striatal neurons in Parkinson's disease. *Ann Neurol*, 1999, 46, 103–111.
89. Muriel MP, Orieux G, Hirsch EC: Levodopa but not ropinirole induces an internalization of D1 dopamine receptors in parkinsonian rats. *Mov Disord*, 2002, 17, 1174–1179.
90. Mytilineou C, Han SK, Cohen G: Toxic and protective effects of L-dopa on mesencephalic cell cultures. *J Neurochem*, 1993, 61, 1470–1478.
91. Nutt JG: Clinical pharmacology of levodopa-induced dyskinesia. *Ann Neurol*, 2000, 47, S160–S164.
92. Ogawa N, Tanaka K, Asanuma M, Kawai M, Masumizu T, Kohno M, Mori A: Bromocriptine protects mice against 6-hydroxydopamine and scavenges hydroxyl free radicals *in vitro*. *Brain Res*, 1994, 657, 207–213.
93. Ohta K, Fujinami A, Kuno S, Sakakimoto A, Matsui H, Kawahara Y, Ohta M: Cabergoline stimulates synthesis and secretion of nerve growth factor, brain-derived neurotrophic factor and glial cell line-derived neurotrophic factor by mouse astrocytes in primary culture. *Pharmacology*, 2004, 71, 162–168.
94. Olanow CW: A radical hypothesis for neurodegeneration. *Trends Neurosci*, 1993, 16, 439–444.
95. Opacka-Juffry J, Wilson AW, Blunt SB: Effects of pergolide treatment on *in vivo* hydroxyl free radical formation during infusion of 6-hydroxydopamine in rat striatum. *Brain Res*, 1998, 810, 27–33.
96. Ossowska K, Wardas J, Kuter K, Nowak P, Dabrowska J, Bortel A, Labus L et al.: Influence of paraquat on dopaminergic transporter in the rat brain. *Pharmacol Rep*, 2005, 57, 330–335.
97. Pardo B, Mena MA, De Yebenes JG: L-dopa inhibits complex IV of the electron transport chain in catecholamine-rich human neuroblastoma NB69 cells. *J Neurochem*, 1995, 64, 576–582.
98. Parkinson Study Group: Dopamine transporter brain imaging to assess the effects of pramipexole vs. levodopa on Parkinson's disease progression. *JAMA*, 2002, 287, 1653–1661.

99. Paulus W, Jellinger K: The neuropathologic basis of different clinical subgroups of Parkinson's disease. *J Neuropathol Exp Neurol*, 1991, 50, 743–755.
100. Pedrosa R, Soares-da-Silva P: Oxidative and non-oxidative mechanisms of neuronal cell death and apoptosis by L-3,4-dihydroxyphenylalanine (L-DOPA) and dopamine. *Br J Pharmacol*, 2002, 137, 1305–1313.
101. Perry TL, Yong VW, Ito M, Foulks JG, Wall RA, Godin DV, Clavier RM: Nigrostriatal dopaminergic neurons remain undamaged in rats given high doses of L-DOPA and carbidopa chronically. *J Neurochem*, 1984, 43, 990–993.
102. Pilon C, Levesque D, Dimitriadou V, Griffon N, Martres MP, Schwartz JC, Sokoloff P: Functional coupling of the human dopamine D3 receptor in a transfected NG 108–15 neuroblastoma-glioma hybrid cell line. *Eur J Pharmacol*, 1994, 268, 129–139.
103. Pirtosek Z, Flisar D: Neuroprotection and dopamine agonists. *Adv Exp Med Biol*, 2004, 541, 55–74.
104. Popperl G, Tatsch K, Ruzicka E, Storch A, Gasser T, Schwarz J: Comparison of alpha-dihydroergocryptine and levodopa monotherapy in Parkinson's disease: assessment of changes in DAT binding with [(123)I]IPT SPECT. *J Neural Transm*, 2004, 111, 1041–1052.
105. Quinn N, Parkes D, Janota I, Marsden CD: Preservation of the substantia nigra and locus coeruleus in a patient receiving levodopa (2 kg) plus decarboxylase inhibitor over a four-year period. *Mov Disord*, 1986, 1, 65–68.
106. Ramirez AD, Wong SK, Menniti FS: Pramipexole inhibits MPTP toxicity in mice by dopamine D3 receptor dependent and independent mechanisms. *Eur J Pharmacol*, 2003, 475, 29–35.
107. Rascol O, Pathak A, Bagheri H, Montastruc JL: New concerns about old drugs: Valvular heart disease on ergot derivative dopamine agonists as an exemplary situation of pharmacovigilance. *Mov Disord*, 2004, 19, 611–613.
108. Reichmann H: Long-term treatment with dopamine agonists in idiopathic Parkinson's disease. *J Neurol*, 2000, 247, Suppl 4, IV 17–IV 19.
109. Reichmann H, Brecht MH, Koster J, Kraus PH, Lemke MR: Pramipexole in routine clinical practice: a prospective observational trial in Parkinson's disease. *CNS Drugs*, 2003, 17, 965–973.
110. Rinne UK, Bracco F, Chouza C, Dupont E, Gershanik O, Marti Masso JF, Montastruc JL, Marsden CD: Early treatment of Parkinson's disease with cabergoline delays the onset of motor complications. Results of a double-blind levodopa controlled trial. The PKDS009 Study Group. *Drugs*, 1998, 55, Suppl 1, 23–30.
111. Roceri M, Molteni R, Fumagalli F, Racagni G, Gennarelli M, Corsini G, Maggio R, Riva M: Stimulatory role of dopamine on fibroblast growth factor-2 expression in rat striatum. *J Neurochem*, 2001, 76, 990–997.
112. Sam EE, Verbeke N: Free radical scavenging properties of apomorphine enantiomers and dopamine: possible implication in their mechanism of action in parkinsonism. *J Neural Transm Park Dis Dement Sect*, 1995, 10, 115–127.
113. Schapira AH: Neuroprotection in PD – A role for dopamine agonists? *Neurology*, 2003, 61, S34–S42.
114. Schrag A, Keens J, Warner J: Ropinirole for the treatment of tremor in early Parkinson's disease. *Eur J Neurol*, 2002, 9, 253–257.
115. Schwartz JC, Giros B, Martres MP, Sokoloff P: The dopamine receptor family: molecular biology and pharmacology. *The Neurosciences*, 1992, 4, 99–108.
116. Schwarz J: Rationale for dopamine agonist use as monotherapy in Parkinson's disease. *Curr Opin Neurol*, 2003, 16, Suppl 1, S27–S33.
117. Sealfon SC, Olanow CW: Dopamine receptors: from structure to behavior. *Trends Neurosci*, 2000, 23, S34–S40.
118. Serratrice J, Disdier P, Habib G, Viallet F, Weiller PJ: Fibrotic valvular heart disease subsequent to bromocriptine treatment. *Cardiol Rev*, 2002, 10, 334–336.
119. Sokoloff P, Giros B, Martres MP, Bouthenet ML, Schwartz JC: Molecular cloning and characterization of a novel dopamine receptor (D3) as a target for neuroleptics. *Nature*, 1990, 347, 146–151.
120. Spina MB, Cohen G: Exposure of striatal [corrected] synaptosomes to L-dopa increases levels of oxidized glutathione. *J Pharmacol Exp Ther*, 1988, 247, 502–507.
121. Sunahara RK, Guan HC, O'Dowd BF, Seeman P, Laurier LG, Ng G, George SR et al.: Cloning of the gene for a human dopamine D5 receptor with higher affinity for dopamine than D1. *Nature*, 1991, 350, 614–619.
122. Takashima H, Tsujihata M, Kishikawa M, Freed WJ: Bromocriptine protects dopaminergic neurons from levodopa-induced toxicity by stimulating D(2)receptors. *Exp Neurol*, 1999, 159, 98–104.
123. Tanaka K, Miyazaki I, Fujita N, Haque ME, Asanuma M, Ogawa N: Molecular mechanism in activation of glutathione system by ropinirole, a selective dopamine D2 agonist. *Neurochem Res*, 2001, 26, 31–36.
124. Ubeda A, Montesinos C, Paya M, Alcaraz MJ: Iron-reducing and free-radical-scavenging properties of apomorphine and some related benzyloquinolines. *Free Radic Biol Med*, 1993, 15, 159–167.
125. Uberti D, Carsana T, Francisconi S, Toninelli GF, Canonico PL, Memo M: A novel mechanism for pergolide-induced neuroprotection: inhibition of NF-kappaB nuclear translocation. *Biochem Pharmacol*, 2004, 67, 1743–1750.
126. Uberti D, Piccioni L, Colzi A, Bravi D, Canonico PL, Memo M: Pergolide protects SH-SY5Y cells against neurodegeneration induced by H(2)O(2). *Eur J Pharmacol*, 2002, 434, 17–20.
127. Vallone D, Picetti R, Borrelli E: Structure and function of dopamine receptors. *Neurosci Biobehav Rev*, 2000, 24, 125–132.
128. Van Camp G, Flamez A, Cosyns B, Weytjens C, Muyldermans L, Van Zandijcke M, De Sutter J et al.: Treatment of Parkinson's disease with pergolide and relation to restrictive valvular heart disease. *Lancet*, 2004, 363, 1179–1183.
129. Van Kampen JM, Hagg T, Robertson HA: Induction of neurogenesis in the adult rat subventricular zone and neostriatum following dopamine D receptor stimulation. *Eur J Neurosci*, 2004, 19, 2377–2387.
130. Vincenzi FF, Hinds TR: Pramipexole has antioxidant properties and inhibits lipid peroxidation. *Proc West Pharmacol Soc*, 1998, 41, 43–46.

-
131. VonVoigtlander PF, Fici GJ, Althaus JS: Pharmacological approaches to counter the toxicity of Dopa. *Amino Acids*, 1998, 14, 189–196.
132. Vu TQ, Ling ZD, Ma SY, Robie HC, Tong CW, Chen EY, Lipton JW, Carvey PM: Pramipexole attenuates the dopaminergic cell loss induced by intraventricular 6-hydroxydopamine. *J Neural Transm*, 2000, 107, 159–176.
133. Waller EA, Kaplan J, Heckman MG: Valvular heart disease in patients taking pergolide. *Mayo Clin Proc*, 2005, 80, 1016–1020.
134. Watts RL: The role of dopamine agonists in early Parkinson's disease. *Neurology*, 1997, 49, S34–S48.
135. Whone AL, Watts RL, Stoessl AJ, Davis M, Reske S, Nahmias C, Lang AE et al.: Slower progression of Parkinson's disease with ropinirole versus levodopa: The REAL-PET study. *Ann Neurol*, 2003, 54, 93–101.
136. Widnell K: Pathophysiology of motor fluctuations in Parkinson's disease. *Mov Disord*, 2005, 20, Suppl 11, S17–S22.
137. Wiseman LR, Fitton A: Cabergoline. A review of its efficacy in the treatment of Parkinson's disease. *CNS Drugs*, 1999, 12, 485–497.
138. Yoshikawa T, Minamiyama Y, Naito Y, Kondo M: Antioxidant properties of bromocriptine, a dopamine agonist. *J Neurochem*, 1994, 62, 1034–1038.
139. Yoshioka M, Tanaka K, Miyazaki I, Fujita N, Higashi Y, Asanuma M, Ogawa N: The dopamine agonist cabergoline provides neuroprotection by activation of the glutathione system and scavenging free radicals. *Neurosci Res*, 2002, 43, 259–267.
140. Youdim MB, Grunblatt E, Mandel S: The pivotal role of iron in NF-kappa B activation and nigrostriatal dopaminergic neurodegeneration. Prospects for neuroprotection in Parkinson's disease with iron chelators. *Ann NY Acad Sci*, 1999, 890, 7–25.
141. Young BK, Camicioli R, Ganzini L: Neuropsychiatric adverse effects of antiparkinsonian drugs. Characteristics, evaluation and treatment. *Drugs Aging*, 1997, 10, 367–383.
142. Yuan H, Sarre S, Ebinger G, Michotte Y: Neuroprotective and neurotrophic effect of apomorphine in the striatal 6-OHDA-lesion rat model of Parkinson's disease. *Brain Res*, 2004, 1026, 95–107.
143. Zigmond MJ, Hastings TG, Perez RG: Increased dopamine turnover after partial loss of dopaminergic neurons: compensation or toxicity? *Parkinsonism Relat Disord*, 2002, 8, 389–393.
144. Zou L, Jankovic J, Rowe DB, Xie W, Appel SH, Le W: Neuroprotection by pramipexole against dopamine- and levodopa-induced cytotoxicity. *Life Sci*, 1999, 64, 1275–1285.

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