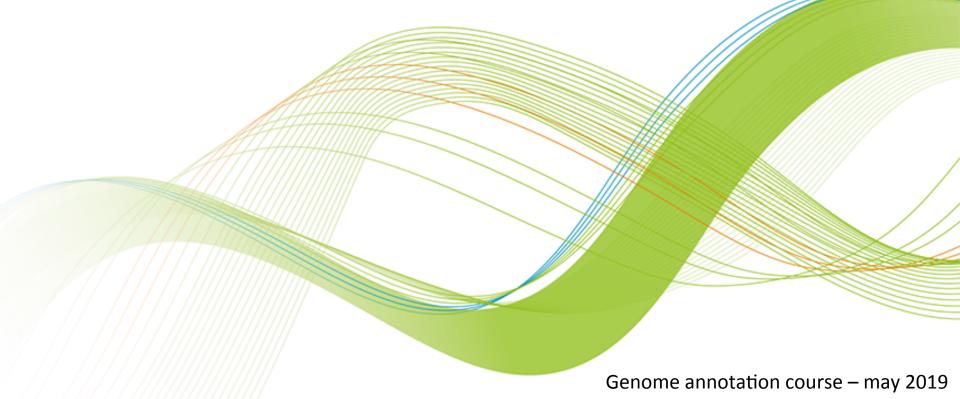




Bacterial Genome Annotation





Bacterial genome characteristics



- A bacterial genome is a single "circular" DNA molecule with several million base pairs in size
- Bacteria can contains plasmids (small and circular DNA molecules, that contain (usually) non-essential genes)
- Genomes contain a few thousand genes.
- "Gene density" is much higher than in humans, one million base pairs of bacterial DNA contains about 500 to 1000 genes.
 - bacterial genes have no introns,
 - the average number of codons in bacterial genes is less than in human genes,
 - neighboring genes are very close together throughout the genome



Bacterial feature types



- protein coding genes
 - promoter (-10, -35)
 - ribosome binding site (RBS)
 - coding sequence (CDS)
 - signal peptide, protein domains, structure
 - terminator
- non coding genes
 - transfer RNA (tRNA)
 - ribosomal RNA (rRNA)
 - non-coding RNA (ncRNA)
- other
 - o repeat patterns, operons, origin of replication, ...



Automatic annotation



Two strategies for identifying coding genes:

sequence alignment

- find known protein sequences in the contigs
 - transfer the annotation across
- will miss proteins not in your database
- may miss partial proteins

• ab initio gene finding

- find candidate open reading frames
 - build model of ribosome binding sites
 - predict coding regions
- may choose the incorrect start codon
- may miss atypical genes, overpredict small genes



Some good existing tools SciLifeLab



Software	ab initio	align- ment	Availability	Speed		
RAST	yes	yes	web only	12-24 hours		
xBASE	yes	no	web only	>4 hours		
BG7	no	yes	standalone	>10 hours		
PGAAP (NCBI)	yes	yes	email / we	>1 month		

Seemann T et al. Bacterial genome annotation, presentation 2016



Prokka



- Fast
 - exploits multi-core computers (aim < 15min)
- Convenient
 - Does structural and functional annotation in one go
 - Help submitting to NCBI and ENA
- Standards compliant
 - GFF3/GBK for viewing, TBL/FSA for Genbank.
- Provenance
 - Keep record of where/how/why it was annotated
- Also annotates archaea, mitochondria, and viruses



Prokka



- Complicated to install
 - many dependencies (available on conda and rackham)

Feature prediction tools used by Prokka:

Tool (reference)	Features predicted
Prodigal (Hyatt 2010) RNAmmer (Lagesen et al., 2007) Aragorn (Laslett and Canback, 2004) SignalP (Petersen et al., 2011) Infernal (Kolbe and Eddy, 2011)	Coding sequence (CDS) Ribosomal RNA genes (rRNA) Transfer RNA genes Signal leader peptides Non-coding RNA

Seemann T. *Prokka: rapid prokaryotic genome annotation.* **Bioinformatics**. 2014 Jul 15;30(14):2068-9. PMID:24642063



Prokka: method

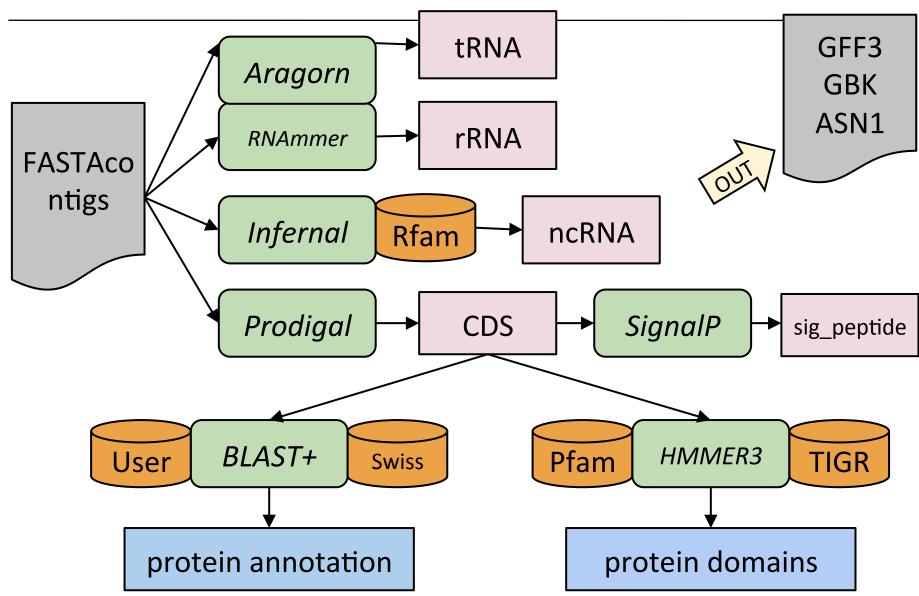


- Prodigal identifies the coordinates of candidates genes
- Compares with a database of known sequences
 - Small trustworthy database: the user provides a set of annotation proteins (optional)
 - Genus-specific proteome (optional)
 - Medium-size domain specific database: Uniprot-Swissprot
 - Curated model of protein families: all proteins from finished bacterial genomes in Refseq
 - HMMs profile: Pfam, TIGRFAMS (with HMMER)
 - If nothing is found, label as 'hypothetical protein'



Prokka pipeline (simplified)





Seemann T et al. Bacterial genome annotation, presentation 2016



Prokka options



- Only one parameter mandatory : Input fasta format
 - prokka [options] < contigs.fasta>
- More than 30 different options available
 - prokka --help



Command line options



```
General:
 --help
                    This help
 --version
                    Print version and exit
 --docs
                    Show full manual/documentation
 --citation
                    Print citation for referencing Prokka
 --quiet
                    No screen output (default OFF)
                    Debug mode: keep all temporary files (default OFF)
 --debug
Setup:
                    List all configured databases
 --listdb
                    Index all installed databases
 --setupdb
 --cleandb
                    Remove all database indices
 --depends
                    List all software dependencies
Outputs:
 --outdir [X]
                    Output folder [auto] (default '')
                    Force overwriting existing output folder (default OFF)
 --force
                    Filename output prefix [auto] (default '')
 --prefix [X]
 --addgenes
                    Add 'gene' features for each 'CDS' feature (default OFF)
 --locustag [X]
                    Locus tag prefix (default 'PROKKA')
 --increment [N]
                    Locus tag counter increment (default '1')
 --gffver [N]
                    GFF version (default '3')
 --compliant
                    Force Genbank/ENA/DDJB compliance: --genes --mincontiglen 200 --centre XXX (default OFF)
 --centre [X]
                    Sequencing centre ID. (default '')
Organism details:
                    Genus name (default 'Genus')
 --genus [X]
                    Species name (default 'species')
 --species [X]
 --strain [X]
                    Strain name (default 'strain')
 --plasmid [X]
                    Plasmid name or identifier (default '')
Annotations:
 --kingdom [X]
                    Annotation mode: Archaea|Bacteria|Mitochondria|Viruses (default 'Bacteria')
                    Genetic code / Translation table (set if --kingdom is set) (default '0')
 --gcode [N]
                    Gram: -/neg +/pos (default '')
 --gram [X]
                    Use genus-specific BLAST databases (needs --genus) (default OFF)
 --usegenus
 --proteins [X]
                    Fasta file of trusted proteins to first annotate from (default '')
 --hmms [X]
                    Trusted HMM to first annotate from (default '')
                    Improve gene predictions for highly fragmented genomes (default OFF)
 --metagenome
                    Do not clean up /product annotation (default OFF)
 --rawproduct
Computation:
 --fast
                    Fast mode - skip CDS /product searching (default OFF)
                    Number of CPUs to use [0=all] (default '8')
 --cpus [N]
 --mincontiglen [N] Minimum contig size [NCBI needs 200] (default '1')
 --evalue [n.n]
                   Similarity e-value cut-off (default '1e-06')
                    Enable searching for ncRNAs with Infernal+Rfam (SLOW!) (default '0')
 --rfam
 --norrna
                    Don't run rRNA search (default OFF)
                    Don't run tRNA search (default OFF)
  --notrna
                    Prefer RNAmmer over Barrnap for rRNA prediction (default OFF)
 --rnammer
```



Prokka output



Extension	Description
.gff	This is the master annotation in GFF3 format, containing both sequences and annotations. It can be viewed directly in Artemis or IGV.
.gbk	This is a standard Genbank file derived from the master .gff. If the input to prokka was a multi-FASTA, then this will be a multi-Genbank, with one record for each sequence.
.fna	Nucleotide FASTA file of the input contig sequences.
.faa	Protein FASTA file of the translated CDS sequences.
.ffn	Nucleotide FASTA file of all the prediction transcripts (CDS, rRNA, tRNA, tmRNA, misc_RNA)
.sqn	An ASN1 format "Sequin" file for submission to Genbank. It needs to be edited to set the correct taxonomy, authors, related publication etc.
.fsa	Nucleotide FASTA file of the input contig sequences, used by "tbl2asn" to create the .sqn file. It is mostly the same as the .fna file, but with extra Sequin tags in the sequence description lines.
.tbl	Feature Table file, used by "tbl2asn" to create the .sqn file.
.err	Unacceptable annotations - the NCBI discrepancy report.
.log	Contains all the output that Prokka produced during its run. This is a record of what settings you used, even if thequiet option was enabled.
.txt	Statistics relating to the annotated features found.
.tsv	Tab-separated file of all features: locus_tag,ftype,gene,EC_number,product



Prokka output



GFF format

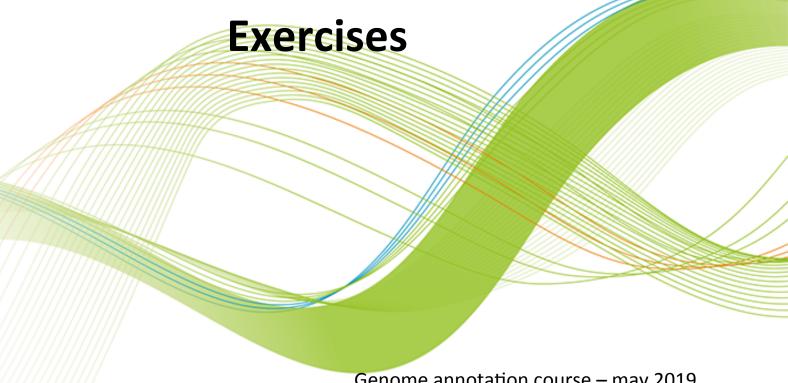
Sb_20131119_contig_1 Sb_20131119_contig_1 Sb_20131119_contig_1	Cufflinks Cufflinks Cufflinks	transc exon transc	1522	1522 2095 3626	2095 1000 4118	1000 1000	:	<pre>. gene_id "CUFF.1"; transcript_id "CUFF.1.1"; FPKM "2.6064385494"; frac "1.000000"; conf_lo "0.94 gene_id "CUFF.1"; transcript_id "CUFF.1.1"; exon_number "1"; FPKM "2.6064385494"; frac "1.000000"; conf . gene_id "CUFF.3"; transcript_id "CUFF.3.1"; FPKM "3.1548106029"; frac "1.000000"; conf_lo "0.82</pre>
Sb_20131119_contig_1	Cufflinks Cufflinks	exon		4118 4855	1000 5340	1000		gene_id "CUFF.3"; transcript_id "CUFF.3.1"; exon_number "1"; FPKM "3.1548106029"; frac "1.000000"; conf
Sb_20131119_contig_1 Sb_20131119_contig_1	Cufflinks	transc exon		5340	1000		:	gene_id "CUFF.5"; transcript_id "CUFF.5.1"; FPKM "7.0235237898"; frac "1.000000"; conf_lo "2.52 gene_id "CUFF.5"; transcript_id "CUFF.5.1"; exon_number "1"; FPKM "7.0235237898"; frac "1.000000"; conf
Sb_20131119_contig_1	Cufflinks	transc		5398	5975	1000		gene_id "CUFF.4"; transcript_id "CUFF.4.1"; FPKM "3.1706609980"; frac "1.000000"; conf_lo "1.17
Sb_20131119_contig_1 Sb_20131119_contig_10	Cufflinks Cufflinks	exon transc		5975 954	1000 2795	1000	:	<pre>gene_id "CUFF.4"; transcript_id "CUFF.4.1"; exon_number "1"; FPKM "3.1706609980"; frac "1.000000"; conf</pre>
Sb_20131119_contig_10	Cufflinks	exon		2795	1000	:		gene_id "CUFF.6"; transcript_id "CUFF.6.1"; exon_number "1"; FPKM "6.8195889357"; frac "1.000000"; conf
Sb_20131119_contig_1 Sb_20131119_contig_1	Cufflinks Cufflinks	transc exon		4502 4718	4718 1000	1000	:	gene_id "CUFF.2"; transcript_id "CUFF.2.1"; FPKM "37.5296486924"; frac "1.000000"; conf_lo "2.5 gene_id "CUFF.2"; transcript_id "CUFF.2.1"; exon_number "1"; FPKM "37.5296486924"; frac "1.000000"; con
Sb_20131119_contig_1	Cufflinks	transc		10522	13208	1000		gene_id "CUFF.23"; transcript_id "CUFF.23.1"; FPKM "55.6377793473"; frac "1.000000"; conf_lo "4
Sb_20131119_contig_1 Sb_20131119_contig_1	Cufflinks Cufflinks	exon transc		13208 13270	1000 14623	1000	:	<pre>gene_id "CUFF.23"; transcript_id "CUFF.23.1"; exon_number "1"; FPKM "55.6377793473"; frac "1.000000"; c</pre>
Sb_20131119_contig_1	Cufflinks	exon	13270	14623	1000			gene_id "CUFF.7"; transcript_id "CUFF.7.1"; exon_number "1"; FPKM "41.2374406123"; frac "1.000000"; con
Sb_20131119_contig_1000 Sb_20131119_contig_1000			transc exon	ript 3991	3991 4547	4547 1000	1000	. gene_id "CUFF.54"; transcript_id "CUFF.54.1"; FPKM "52.7578565123"; frac "1.000000"; co . gene_id "CUFF.54"; transcript_id "CUFF.54.1"; exon_number "1"; FPKM "52.7578565123"; frac "1.00
Sb 20131119_contig 100			transc		1097	2009	1000	. gene_id "CUFF.9"; transcript_id "CUFF.9.1"; FPKM "7.4263254644"; frac "1.000000"; conf

Seqid	source	type	start	end	score	strand	phase	attributes
Chr1	Prodig al	exon	234	1543	·	+	•	<pre>gene_id "gene1"; transcript_id "transcript1"; "prediction:, protein motif"</pre>
Chr1	Snap	CDS	577	1543		+	0	<pre>gene_id "gene1"; transcript_id "transcript1";</pre>





Bacterial Genome Annotation



Genome annotation course – may 2019