## SNV examination

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#### **Imports**

Import VCF files: gen\_vcf1 - VCF retrieved from genomic data, sample 1; gen\_vcf2 - VCF retrieved from genomic data, sample 2; gen\_merged\_vcf - genomic data, sample 1 and 2 merged on alignment step; nas\_vcf1 - nascent RNA data, sample 1; nas\_vcf2 - nascent RNA data, sample 2; nas\_vcf3 - nascent RNA data, sample 3; nas\_vcf\_merged - nascent RNA data, sample 1, 2 and 3 merged after mapping step.

```
fd <- '/home/nadzeya/praktika/'</pre>
gen_vcf1_path = file.path(fd, "gen1_snps_f.vcf.gz")
gen_vcf2_path = file.path(fd, "gen2_snps_f.vcf.gz")
gen_merged_vcf_path <- file.path(fd, "gen_merged_snps_f.vcf.gz")</pre>
nas_vcf1_path <- file.path(fd, "nas1_snps_f.vcf.gz")</pre>
nas_vcf2_path <- file.path(fd, "nas2_snps_f.vcf.gz")</pre>
nas_vcf3_path <- file.path(fd, "nas3_2_snps_f.vcf.gz")</pre>
nas_merged_vcf_path <- file.path(fd, "nas_merged_snps_f.vcf.gz")</pre>
filepaths <- c(gen_vcf1_path,
                gen_vcf2_path,
                gen_merged_vcf_path,
                nas vcf1 path,
                nas_vcf2_path,
                nas_vcf3_path,
                nas_merged_vcf_path)
labels <- c('Gen 1',
             'Gen 2',
             'Gen Merged',
             'Nas 1',
             'Nas 2',
             'Nas 3',
             'Nas Merged')
```

# Comparison of SNV quality metrics across files

Get the dataframe of QC metrics available and their descriptions.

```
gen1 <- readVcf(gen_vcf1_path, "hg38")

## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
## forced to unique rownames</pre>
```

```
## AC
                                                                                              Allele coun
## AF
                                                                                                        Al
## AN
                                                                                                         Z
## BaseQRankSum
## DP
## ExcessHet
## FS
                                                                                                      Phre
                                       Inbreeding coefficient as estimated from the genotype likelihoods
## InbreedingCoeff
## MLEAC
                      Maximum likelihood expectation (MLE) for the allele counts (not necessarily the s
                   Maximum likelihood expectation (MLE) for the allele frequency (not necessarily the s
## MLEAF
## MQ
## MQRankSum
                                                                                                Z-score F
```

Z-sco

Sym

Create information dataframes with QC metrics for all VCF files and plot these metrics densities.

## QD

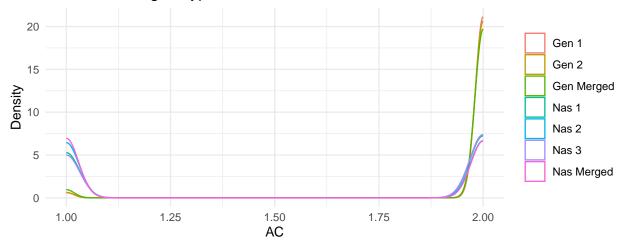
## SOR

## ReadPosRankSum

```
full_info_table <- create_full_info_table(filepaths)</pre>
```

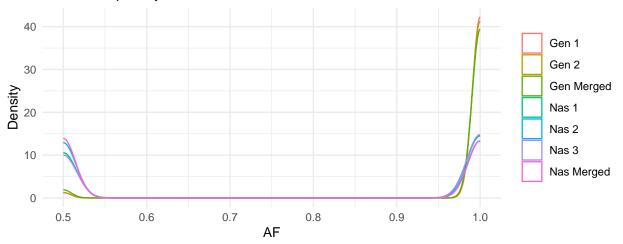
```
## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
## forced to unique rownames
## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
## forced to unique rownames
## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
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## forced to unique rownames
## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
## forced to unique rownames
## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
## forced to unique rownames
```

## Allele count in genotypes, for each ALT allele, in the same order as listed



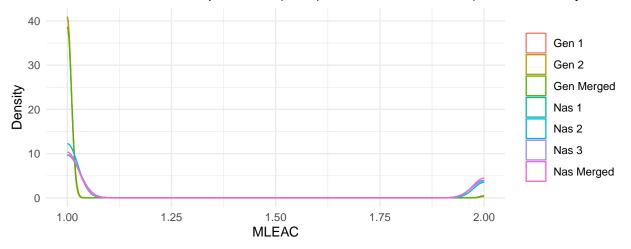
plot\_density(full\_info\_table, "AF", labels, descriptions)

# Allele Frequency, for each ALT allele, in the same order as listed



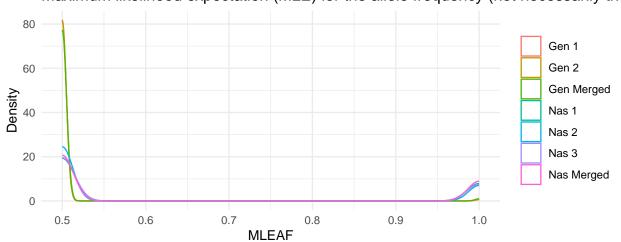
plot\_density(full\_info\_table, "MLEAC", labels, descriptions)

## Maximum likelihood expectation (MLE) for the allele counts (not necessarily the sa

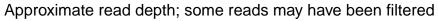


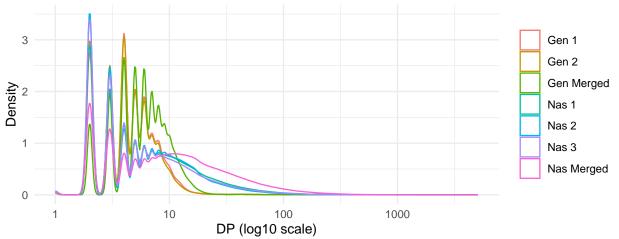
plot\_density(full\_info\_table, "MLEAF", labels, descriptions)

# Maximum likelihood expectation (MLE) for the allele frequency (not necessarily the

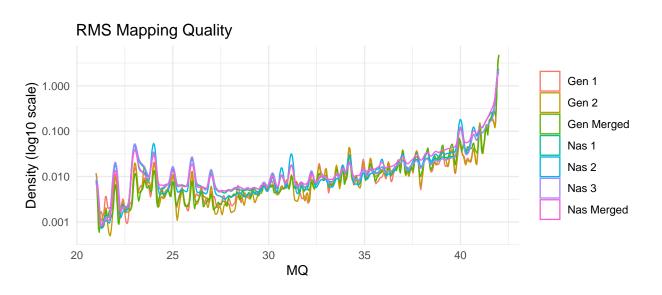


plot\_density(full\_info\_table, 'DP', labels, descriptions, x\_log10=TRUE)



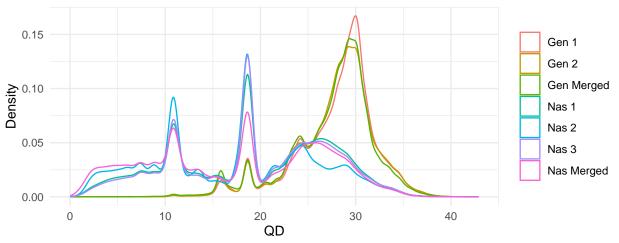


plot\_density(full\_info\_table, 'MQ', labels, descriptions, y\_log10=TRUE)



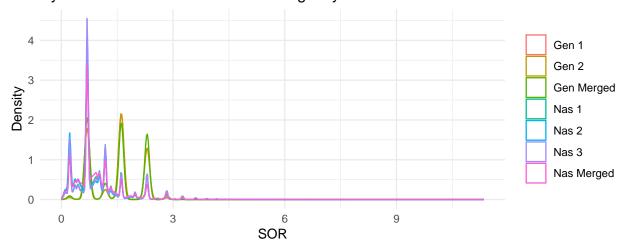
plot\_density(full\_info\_table, 'QD', labels, descriptions)

## Variant Confidence/Quality by Depth



plot\_density(full\_info\_table, 'SOR', labels, descriptions)

## Symmetric Odds Ratio of 2x2 contingency table to detect strand bias



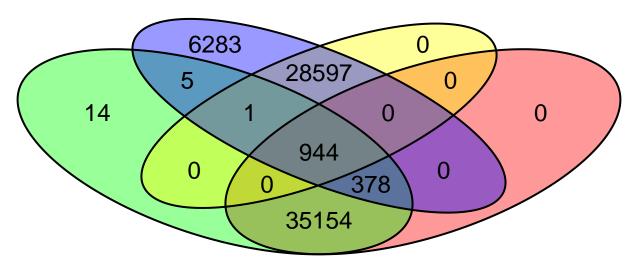
# Analysis of intersections

```
gen1_uf <- load_vcf(gen_vcf1_path, hardfilter=FALSE)
gen2_uf <- load_vcf(gen_vcf2_path, hardfilter=FALSE)
gen_merged_uf <- load_vcf(gen_merged_vcf_path, hardfilter=FALSE)
nas1_uf <- load_vcf(nas_vcf1_path, hardfilter=FALSE)
nas2_uf <- load_vcf(nas_vcf2_path, hardfilter=FALSE)
nas3_uf <- load_vcf(nas_vcf3_path, hardfilter=FALSE)
nas_merged_uf <- load_vcf(nas_merged_vcf_path, hardfilter=FALSE)

gen1 <- load_vcf(gen_vcf1_path)
gen2 <- load_vcf(gen_vcf2_path)
gen_merged <- load_vcf(gen_merged_vcf_path, stats=TRUE)</pre>
```

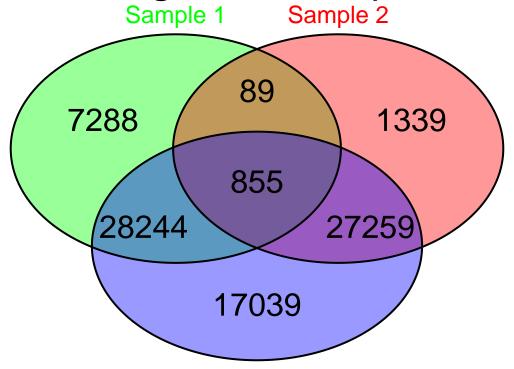
```
## [1] "Filtered by FS: 0"
## [1] "Filtered by SOR: 55"
## [1] "Filtered by QUAL: 0"
## [1] "Filtered by MQ: 16514"
## [1] "Filtered by ReadPosRankSum: 0"
## [1] "Filtered by MQRankSum: 0"
nas1 <- load_vcf(nas_vcf1_path, stats=TRUE)</pre>
## [1] "Filtered by FS: 0"
## [1] "Filtered by SOR: 3808"
## [1] "Filtered by QUAL: 0"
## [1] "Filtered by MQ: 114154"
## [1] "Filtered by ReadPosRankSum: 4"
## [1] "Filtered by MQRankSum: 0"
nas2 <- load_vcf(nas_vcf2_path, stats=TRUE)</pre>
## [1] "Filtered by FS: 0"
## [1] "Filtered by SOR: 3246"
## [1] "Filtered by QUAL: 0"
## [1] "Filtered by MQ: 163330"
## [1] "Filtered by ReadPosRankSum: 3"
## [1] "Filtered by MQRankSum: 2"
nas3 <- load_vcf(nas_vcf3_path, stats=TRUE)</pre>
## [1] "Filtered by FS: 0"
## [1] "Filtered by SOR: 3260"
## [1] "Filtered by QUAL: 0"
## [1] "Filtered by MQ: 105722"
## [1] "Filtered by ReadPosRankSum: 3"
## [1] "Filtered by MQRankSum: 0"
nas_merged <- load_vcf(nas_merged_vcf_path, stats=TRUE)</pre>
## [1] "Filtered by FS: 0"
## [1] "Filtered by SOR: 6291"
## [1] "Filtered by QUAL: 0"
## [1] "Filtered by MQ: 207597"
## [1] "Filtered by ReadPosRankSum: 34"
## [1] "Filtered by MQRankSum: 21"
SNVs from Genomic Data
```

# **SNV** from genomic data (filtered)



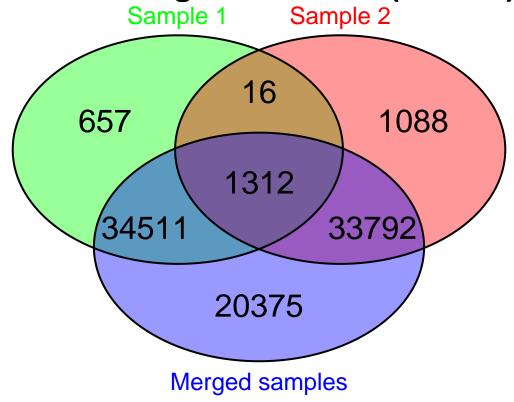
- Sample 1, unfiltered Sample 2, unfiltered
- Sample 1, filtered Sample 2, filtered

# SNVs from genomic data (unfiltered)



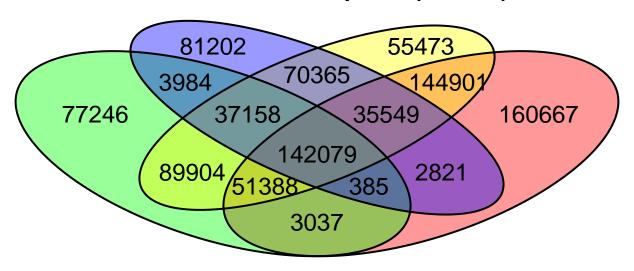
# Merged samples

# SNVs from genomic data (filtered)



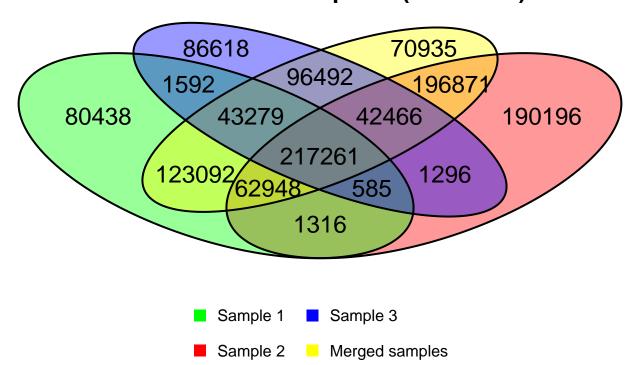
## SNVs from RNA-Seq Data

# **SNV from RNA-Seq data (filtered)**



- Sample 1 Sample 3

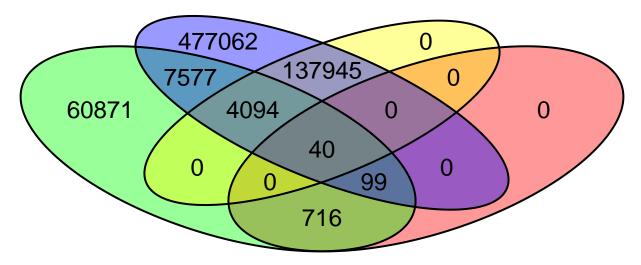
# SNV from RNA-Seq data (unfiltered)



#### SNVs from Genomic and RNA-Seq data

Out of 73 thousand SNVs detected in merged genomic data and 627 thousands of SNVs detected in merged nascent RNA data only 12 thousands SNVs are common:

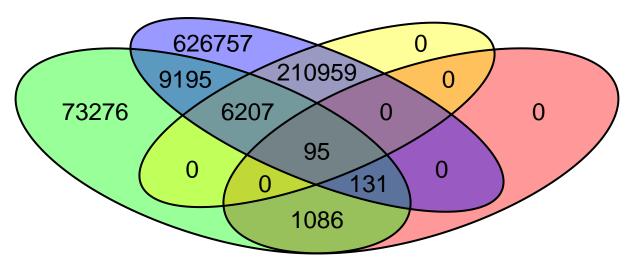
# SNV from genomic and RNA-Seq data



- Genomic (merged samples)RNA–Seq (merged samples)
- Genomic (common across samples) RNA–Seq (common across samples)

In case of unfiltered data, this intersection was only 95 SNVs:

# SNV from genomic and RNA-Seq data (unfiltered)



- Genomic (merged samples)
- RNA-Seq (merged samples)
- Genomic (common across samples)
- RNA-Seq (common across samples)

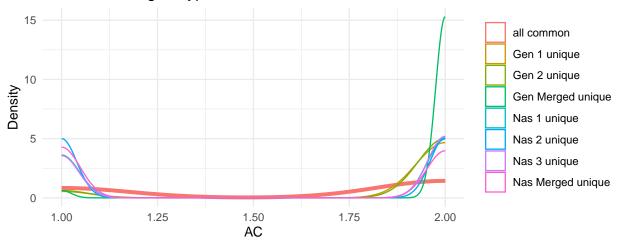
Therefore, SNVs which are common in all nascent RNA VCF files will be used further.

## Comparison of unique and intersecting SNVs

Define subsets of SNVs which are unique for all the files (e.g. SNVs which are present in gen\_vcf1, but absent in gen\_vcf2, gen\_merged, nas\_vcf1, nas\_vcf2, nas\_vcf3, nas\_merged) and visualise stats.

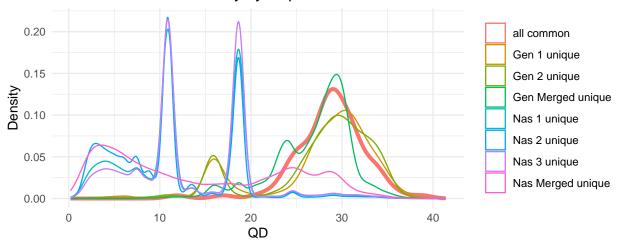
```
# Extract INFO tables and SNV positions (i.e. SNV identifiers)
info_tables <- lapply(filepaths, extract_info_and_positions)</pre>
## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
## forced to unique rownames
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## forced to unique rownames
## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
## forced to unique rownames
## Warning in .bcfHeaderAsSimpleList(header): duplicate keys in header will be
## forced to unique rownames
# Identify unique and common SNV sets based on positions
snv_set_positions <- identify_snv_sets_positions(lapply(info_tables, function(x) x$positions),</pre>
                                                  labels)
param_name <- "AC"</pre>
snv set with metric <- extract metric from info tables(info tables, param name, snv set positions)
plot_ac <- plot_snv_density(snv_set_with_metric, param_name, descriptions_df)</pre>
print(plot ac)
## Warning: Using 'size' aesthetic for lines was deprecated in ggplot2 3.4.0.
## i Please use 'linewidth' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.
```

#### Allele count in genotypes, for each ALT allele, in the same order as listed

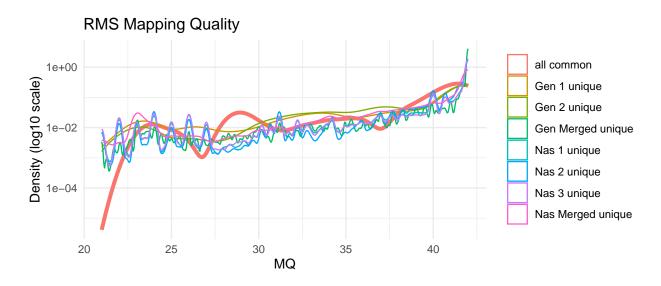


```
param_name <- "QD"
snv_set_with_metric <- extract_metric_from_info_tables(info_tables, param_name, snv_set_positions)
plot_qd <- plot_snv_density(snv_set_with_metric, param_name, descriptions_df)
print(plot_qd)</pre>
```

# Variant Confidence/Quality by Depth



param\_name <- "MQ"
snv\_set\_with\_metric <- extract\_metric\_from\_info\_tables(info\_tables, param\_name, snv\_set\_positions)
plot\_mq <- plot\_snv\_density(snv\_set\_with\_metric, param\_name, descriptions\_df, x\_log10 = FALSE, y\_log10 = print(plot\_mq)</pre>



```
param_name <- "SOR"
snv_set_with_metric <- extract_metric_from_info_tables(info_tables, param_name, snv_set_positions)
plot_sor <- plot_snv_density(snv_set_with_metric, param_name, descriptions_df, x_log10 = FALSE, y_log10
print(plot_sor)</pre>
```

