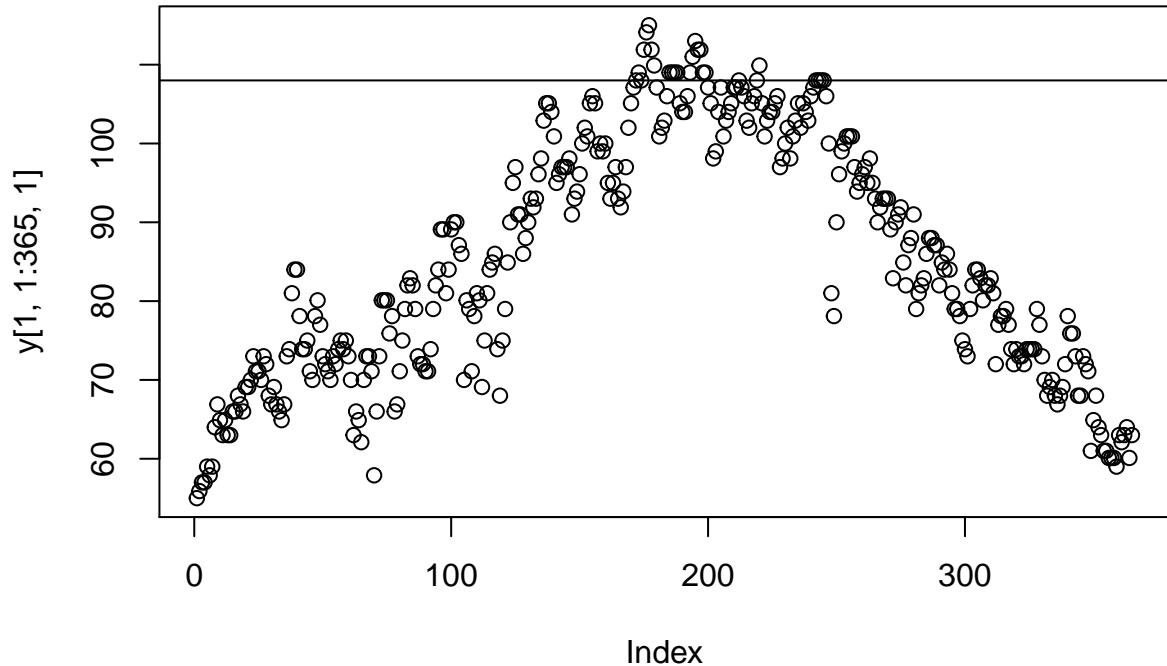


E3

HeatWave $W = 3$ consecutive days above 95th quantile for that year

```
rm(list = ls())
setwd("c:/e/brucebcampbell-git/bayesian-learning-with-R/E3")
load("heatwaves.RData")

plot(y[1,1:365,1])
abline(h = quantile(y[1,1:365,1], .95))
```



```
#####
# HW = 3 consecutive days above 95th quantile for that year
#####
X.num <- matrix(nrow = 41, ncol = 9 )
X.sev <- matrix(nrow = 41, ncol = 9 )

for(i in 1:41)
{
  for(k in 1:9)
  {
    data <- y[i,1:365,k]
```

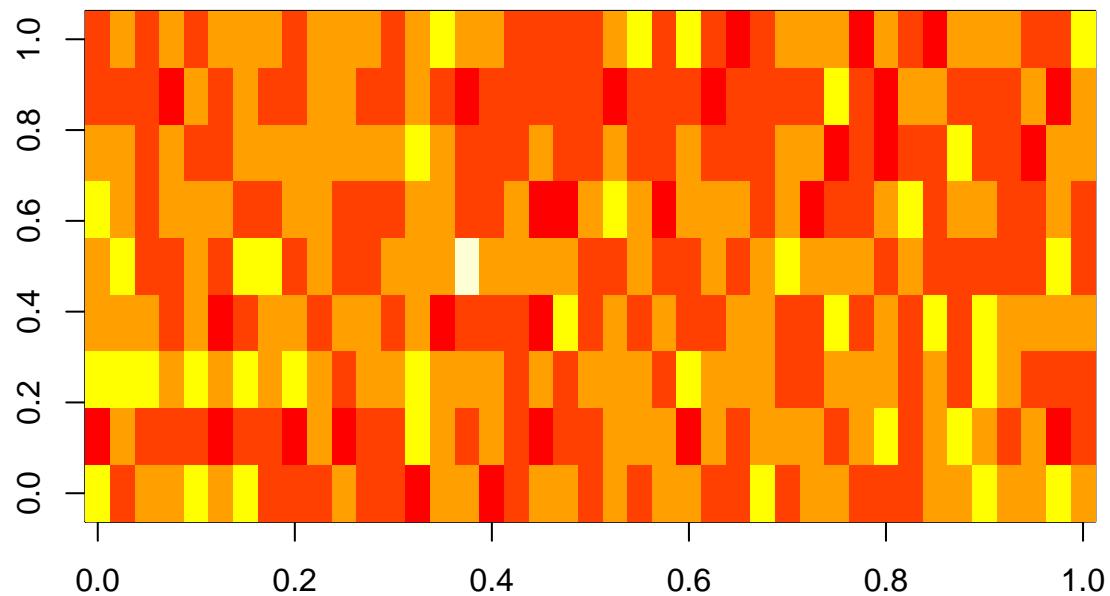
```

data <- na.exclude(data)
q_ik <- quantile(data,.95)
hot_ik <- data > q_ik
run_lengths <- rle(hot_ik)
df_rl <- data.frame(l = run_lengths$lengths,v = run_lengths$values)
hot_groups <- df_rl[df_rl$v==TRUE,]
heat_waves <- hot_groups[hot_groups$l>3,]
num_hw <- nrow(heat_waves)
sev_hw <- sum(heat_waves$l)

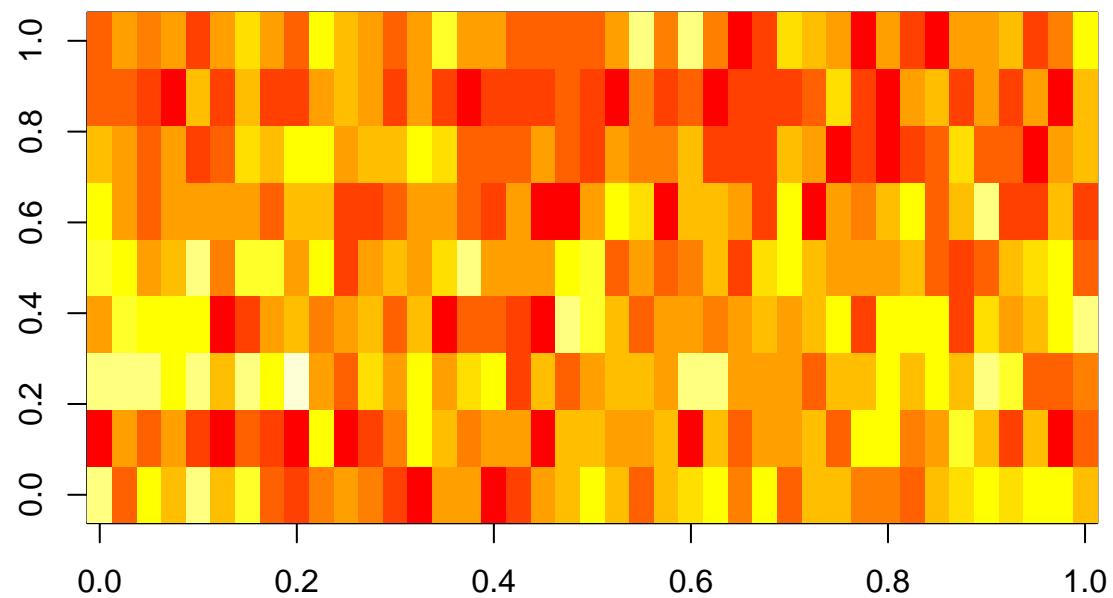
X.num[i,k] <- num_hw
X.sev[i,k] <- sev_hw
}

image(X.num)

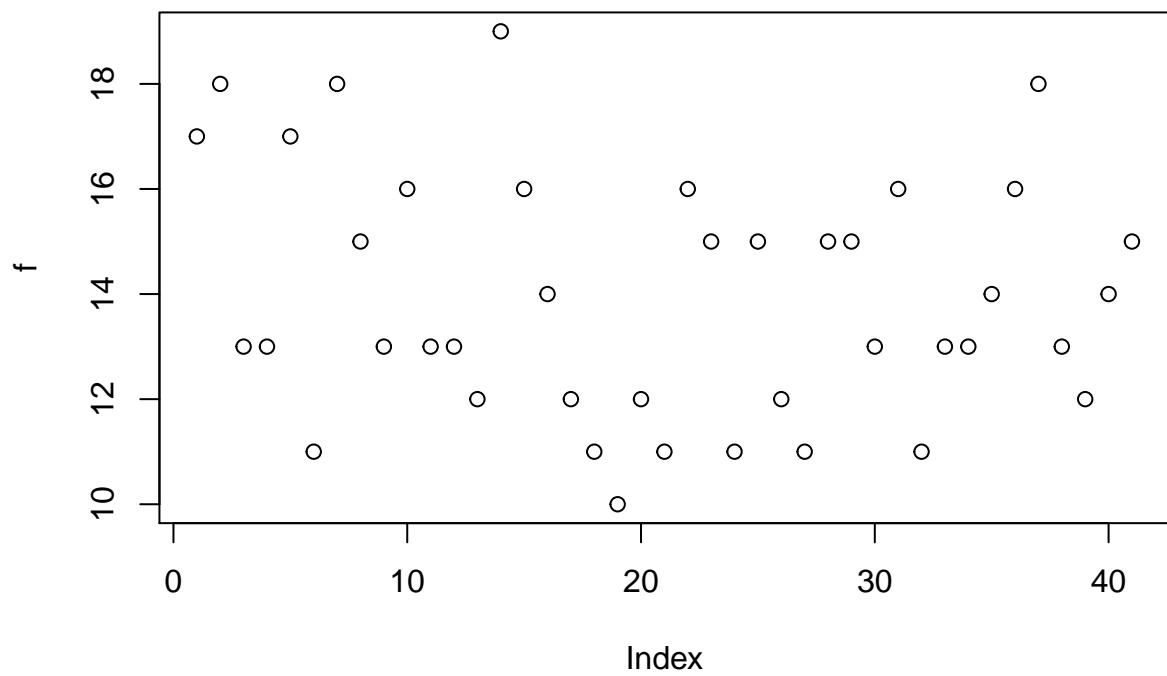
```



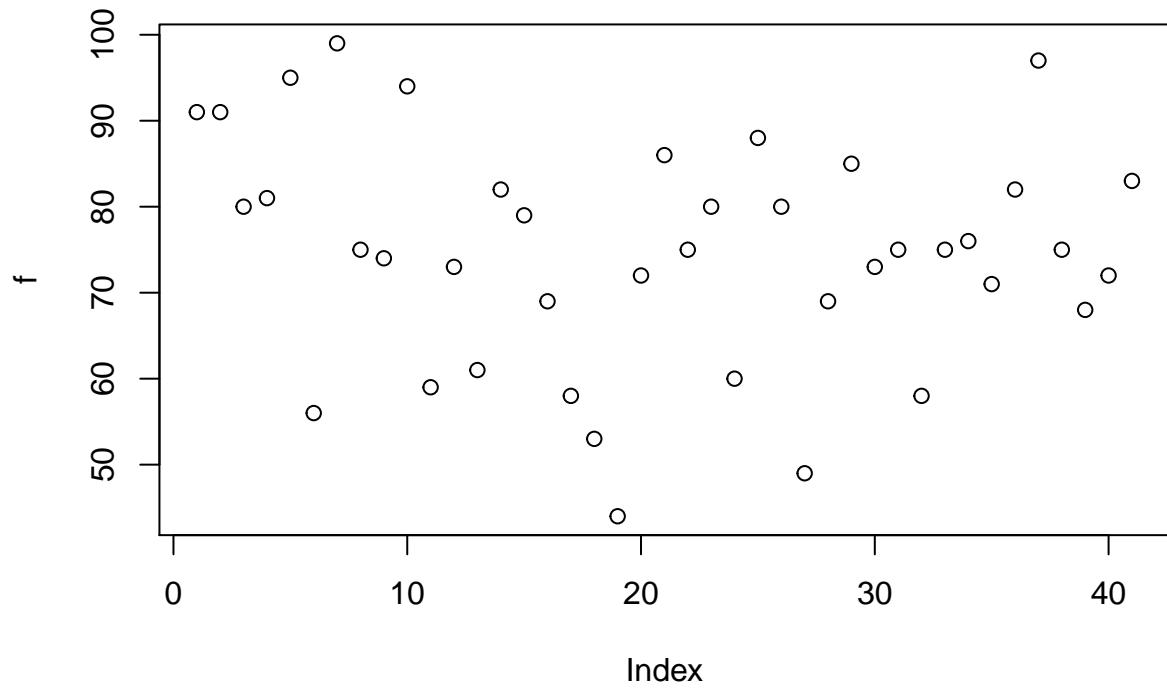
```
image(X.sev)
```



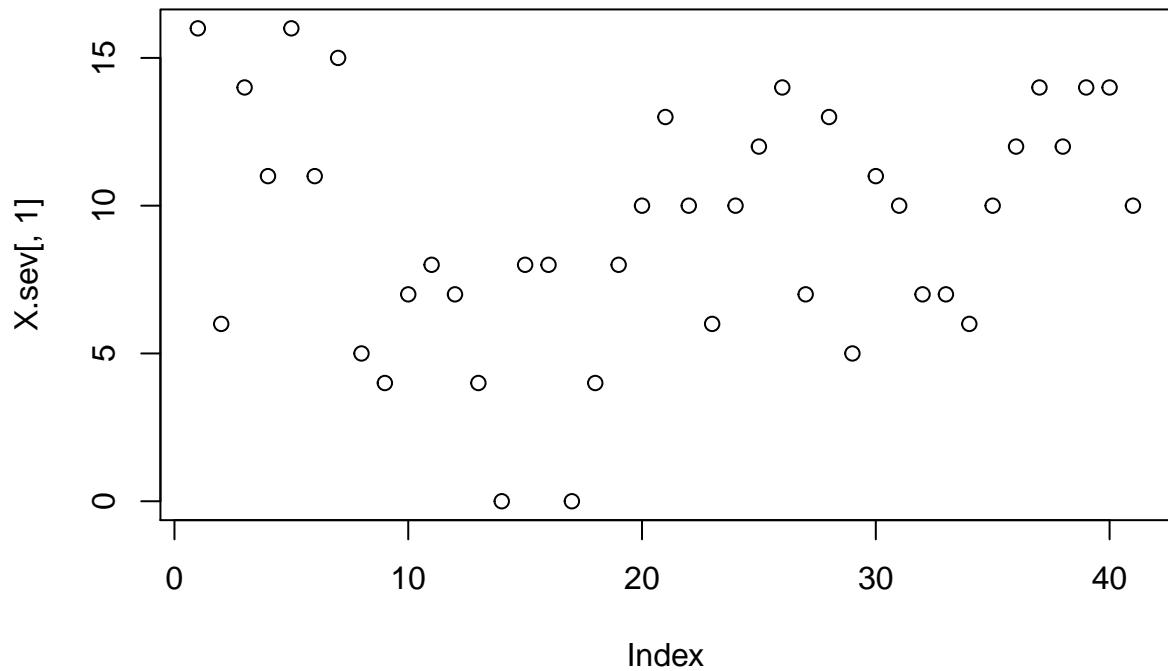
```
f<- rowSums(X.num)
plot(f)
```



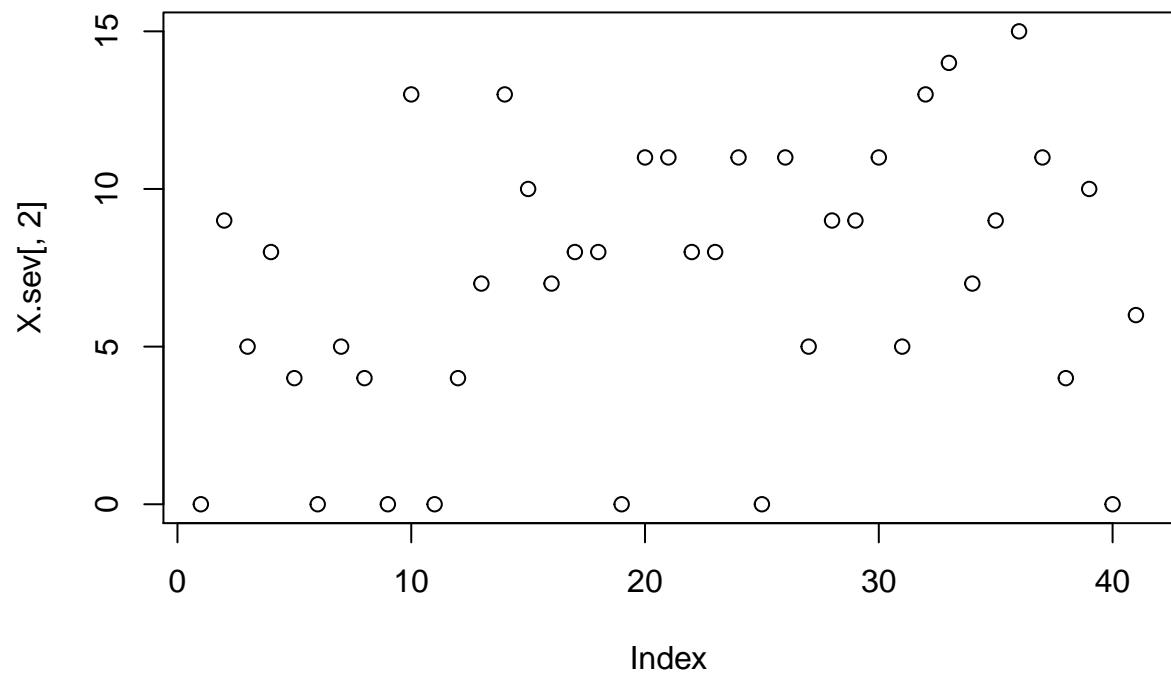
```
f<- rowSums(X.sev)
plot(f)
```



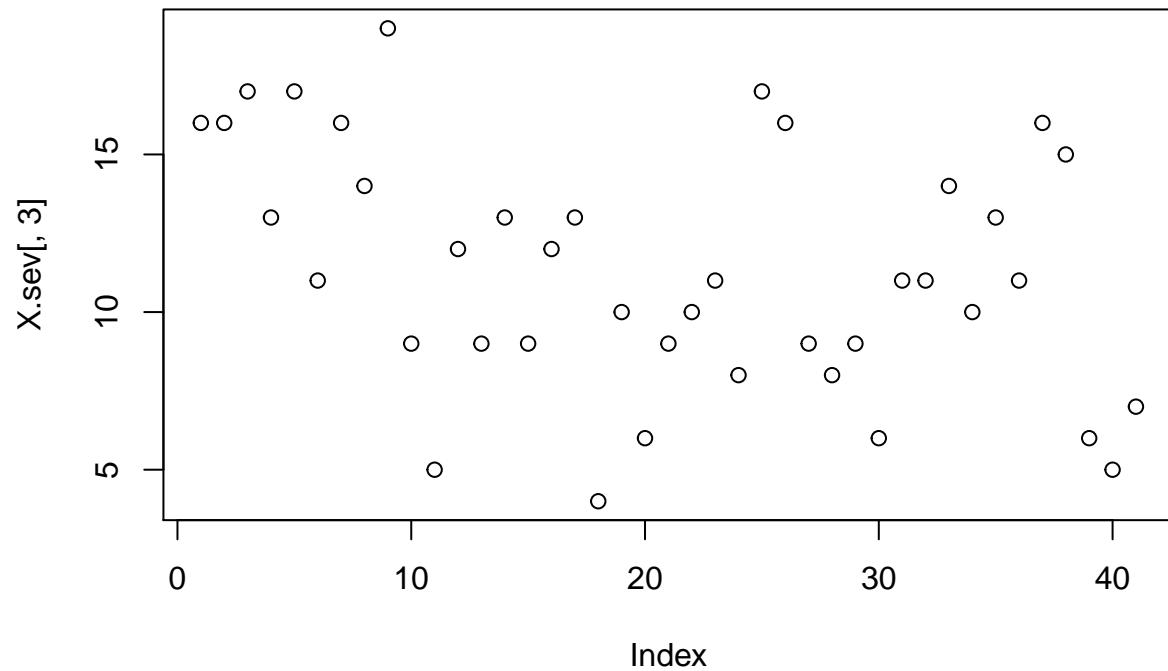
```
plot(X.sev[,1])
```



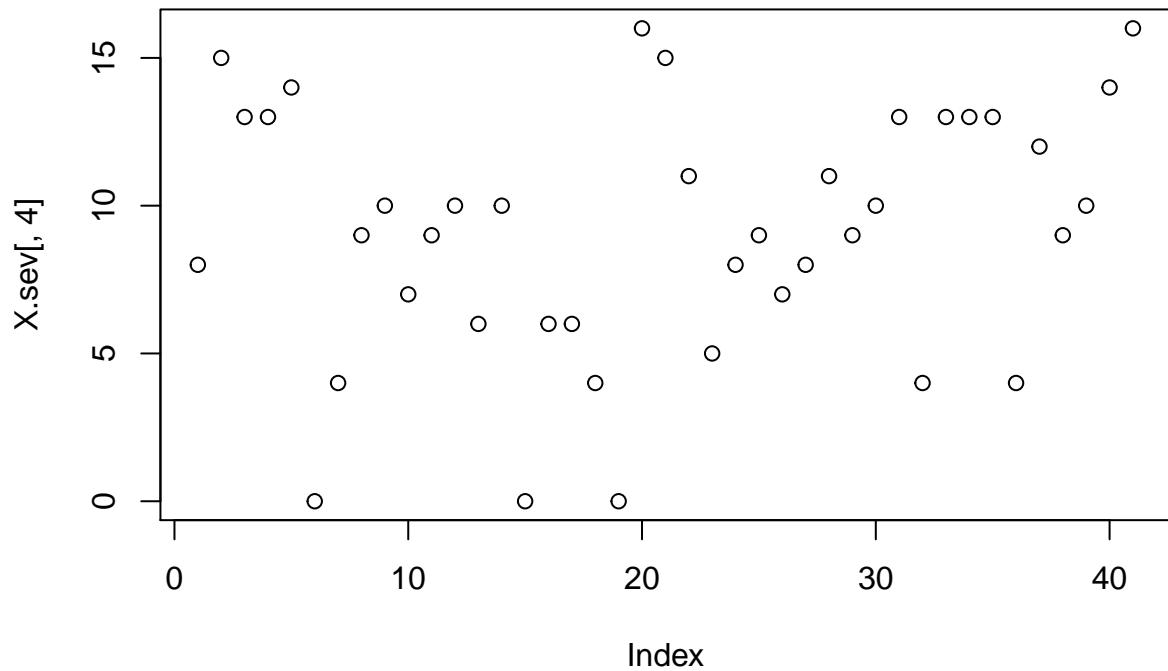
```
plot(X.sev[, 2])
```



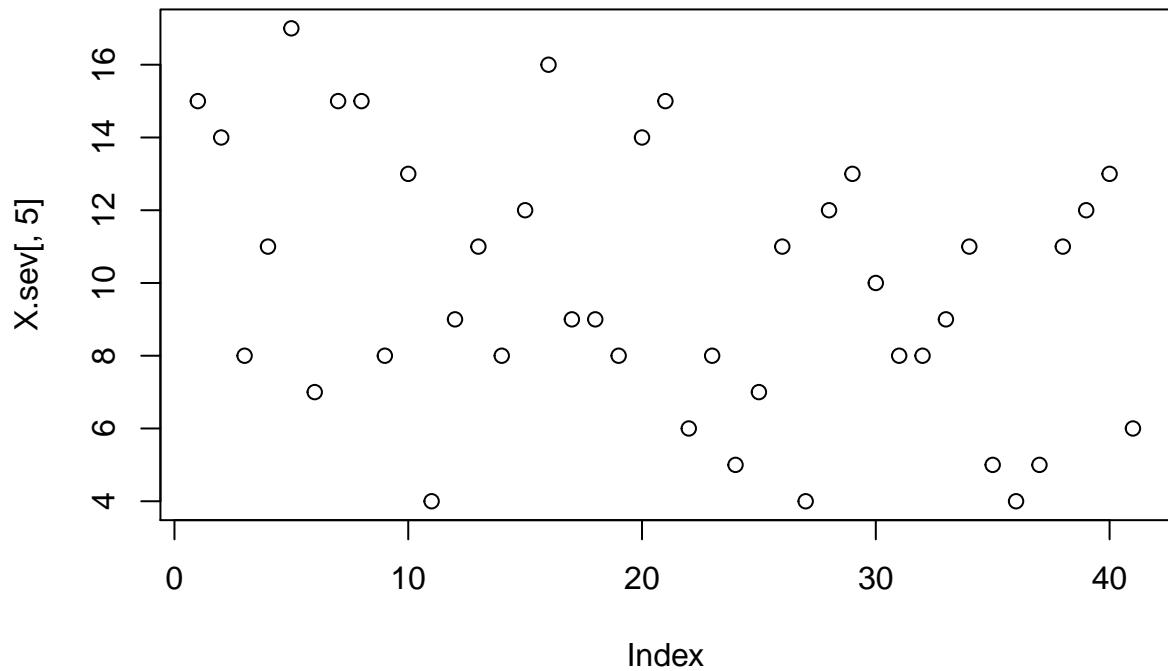
```
plot(X.sev[,3])
```



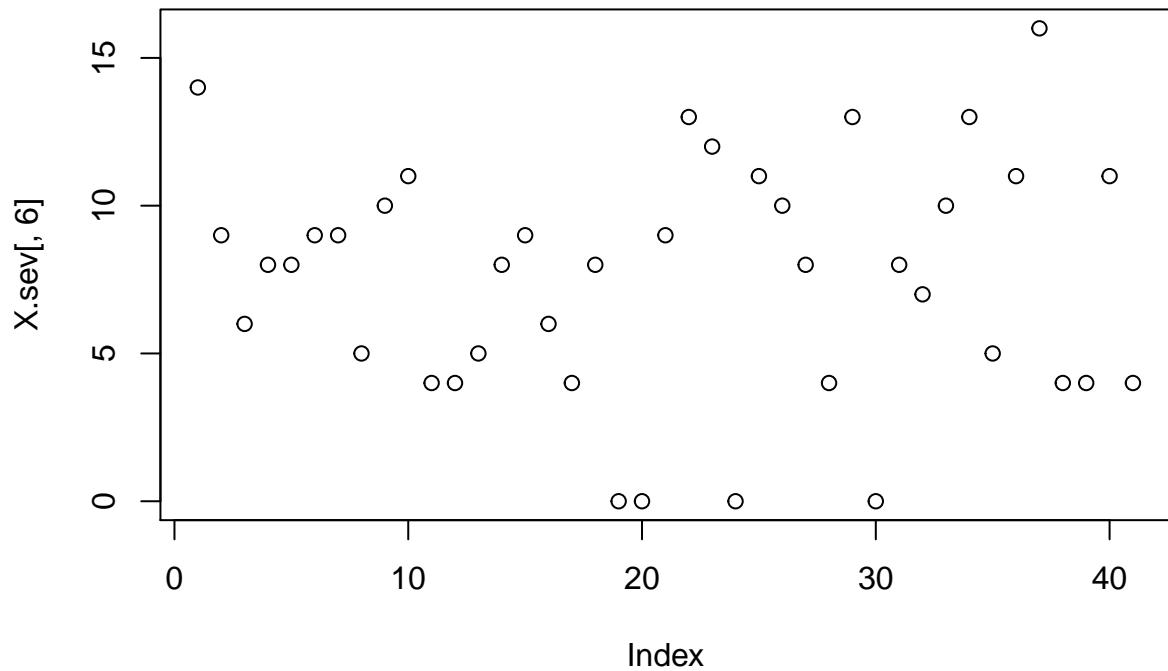
```
plot(X.sev[,4])
```



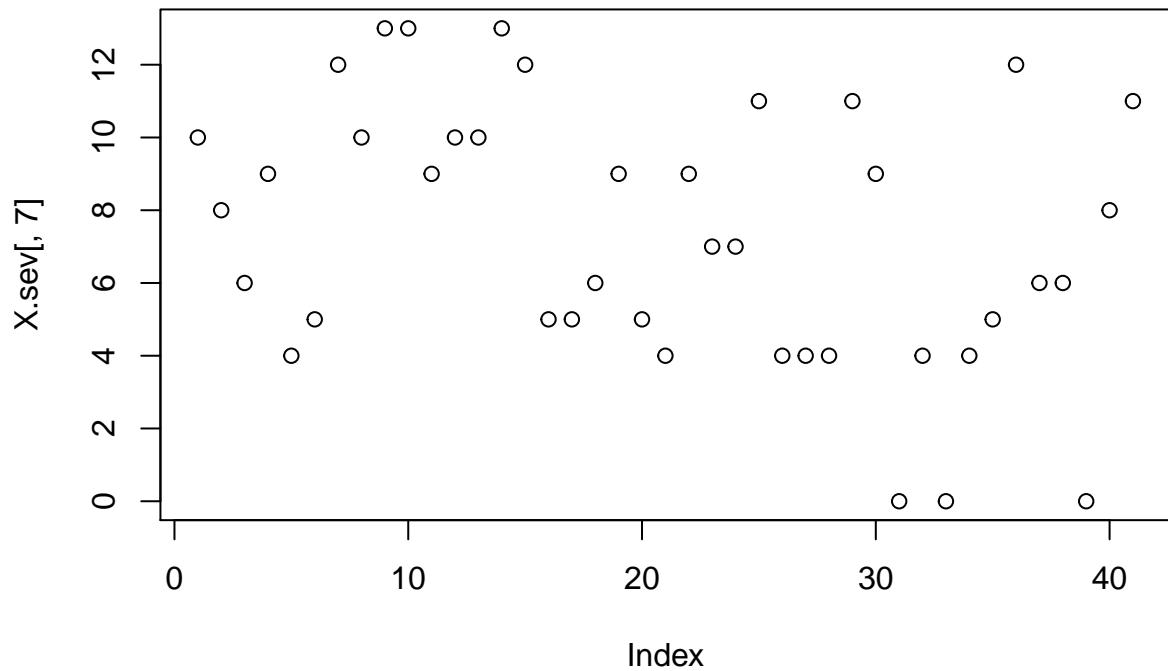
```
plot(X.sev[, 5])
```



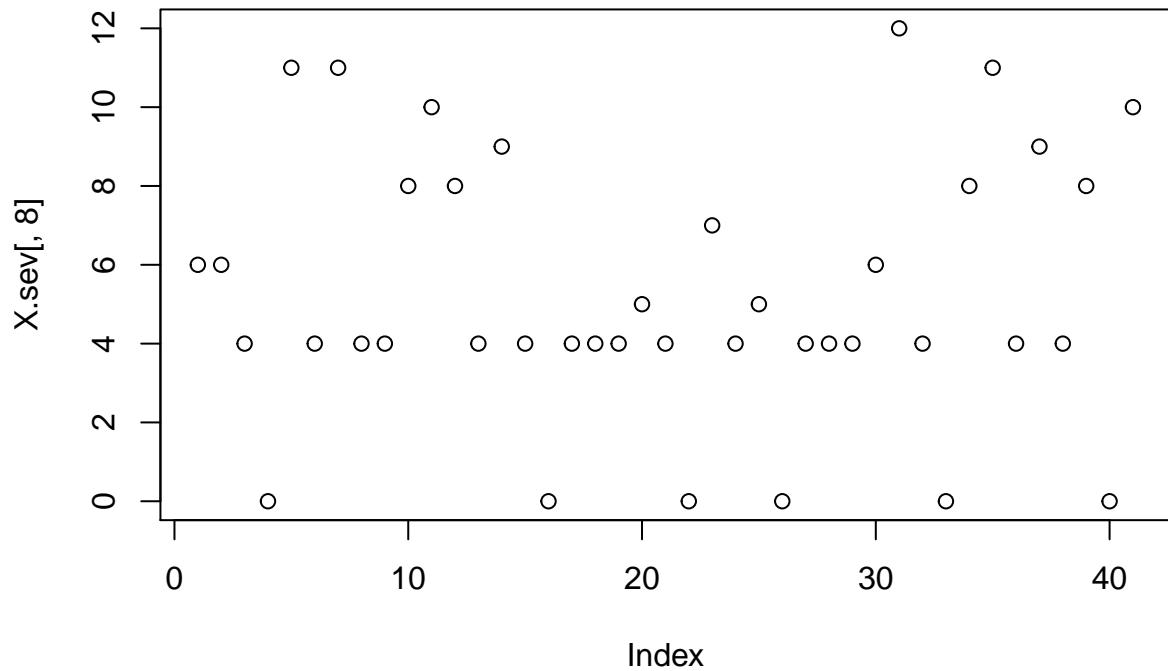
```
plot(X.sev[, 6])
```



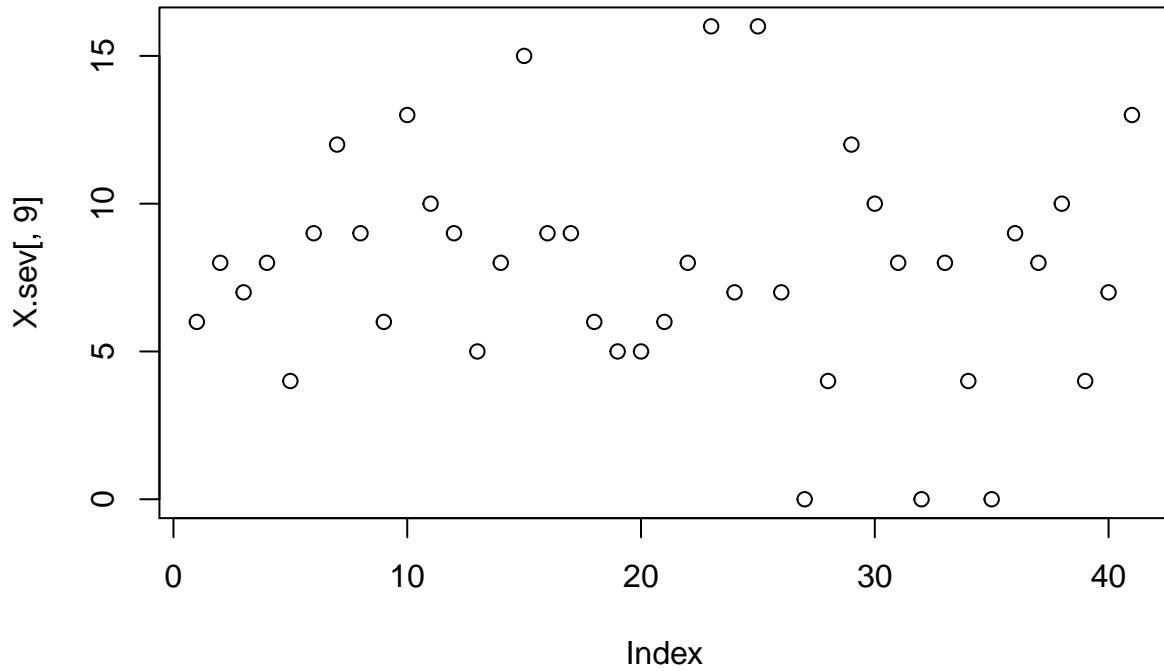
```
plot(X.sev[, 7])
```



```
plot(X.sev[,8])
```



```
plot(X.sev[, 9])
```

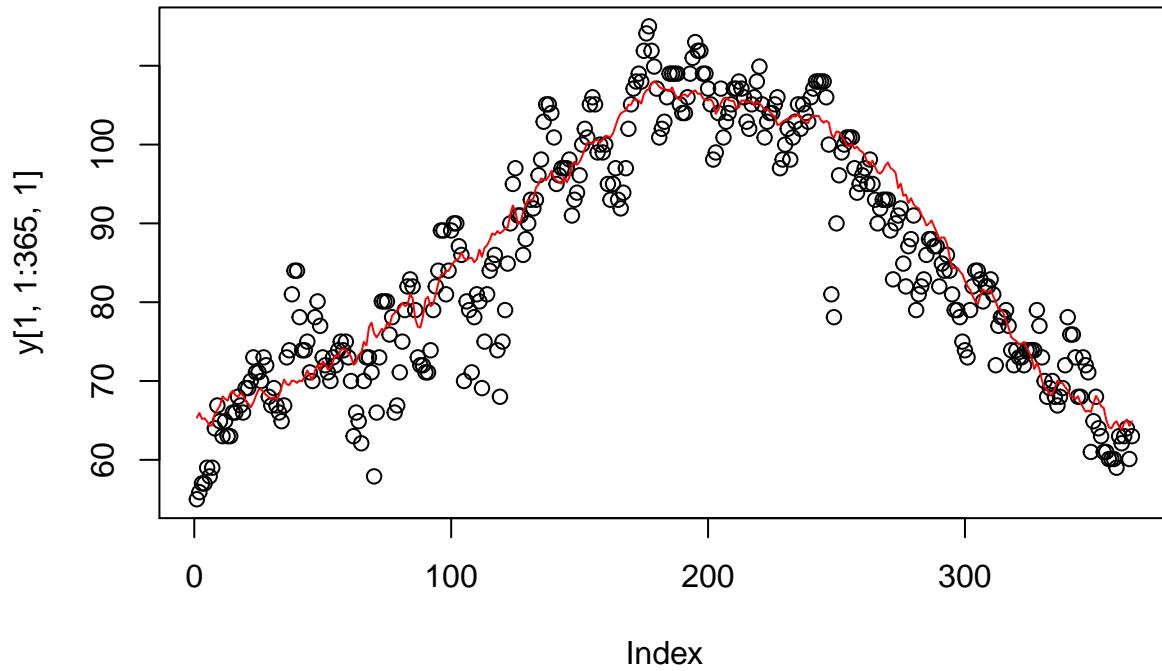


HeatWave = World Meteorological Organization definition

daily temperature for more than five consecutive days exceeds the average temperature by 9 degrees

```
#####
# HW = World Meteorological Organization definition
# daily temperature for more than five consecutive days exceeds the average temperature by 9 degrees
#####
X.num <- matrix(nrow = 41, ncol = 9 )
X.sev <- matrix(nrow = 41, ncol = 9 )

#Daily Averages
DA <- matrix(nrow = 365,ncol = 9)
for(j in 1:365)
{
  for(k in 1:9)
  {
    data <- y[1:41,j,k]
    data <- na.exclude(data)
    DA[j,k] <- mean(data)
  }
}
plot(y[1,1:365,1]); lines(DA[,1],col='red')
```



```

AboveDA.Idx <- y
for(i in 1:41)
{
  for(k in 1:9)
  {
    for(j in 1:365)
    {
      t <- y[i,j,k]
      A <- DA[j,k]
      isHot <- t >=A+9
      AboveDA.Idx[i,j,k] <- isHot
    }
  }
}

X.num <- matrix(nrow =41, ncol = 9 )
X.sev <- matrix(nrow =41, ncol = 9 )

for(i in 1:41)
{
  for(k in 1:9)
  {
    data <- AboveDA.Idx[i,1:365,k]
    data <- na.exclude(data)
    run_lengths <- rle(as.vector(data))
    df_rl <- data.frame(l = run_lengths$lengths, v = run_lengths$values)
  }
}

```

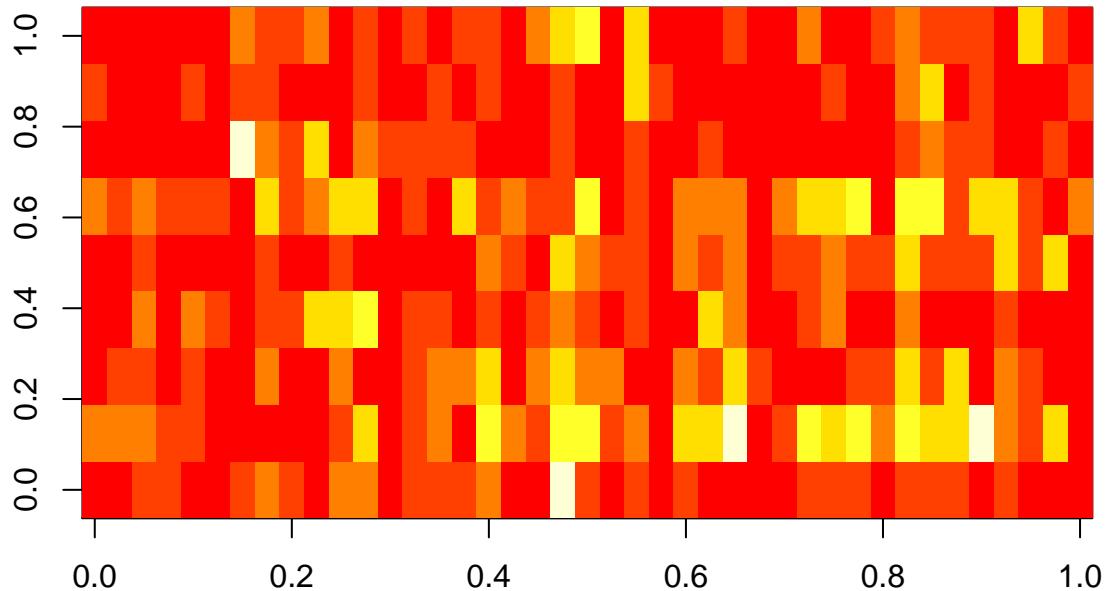
```

hot_groups <- df_rl[df_rl$v==TRUE,]
heat_waves <- hot_groups[hot_groups$l>5,]
num_hw <- nrow(heat_waves)
sev_hw <- sum(heat_waves$l)

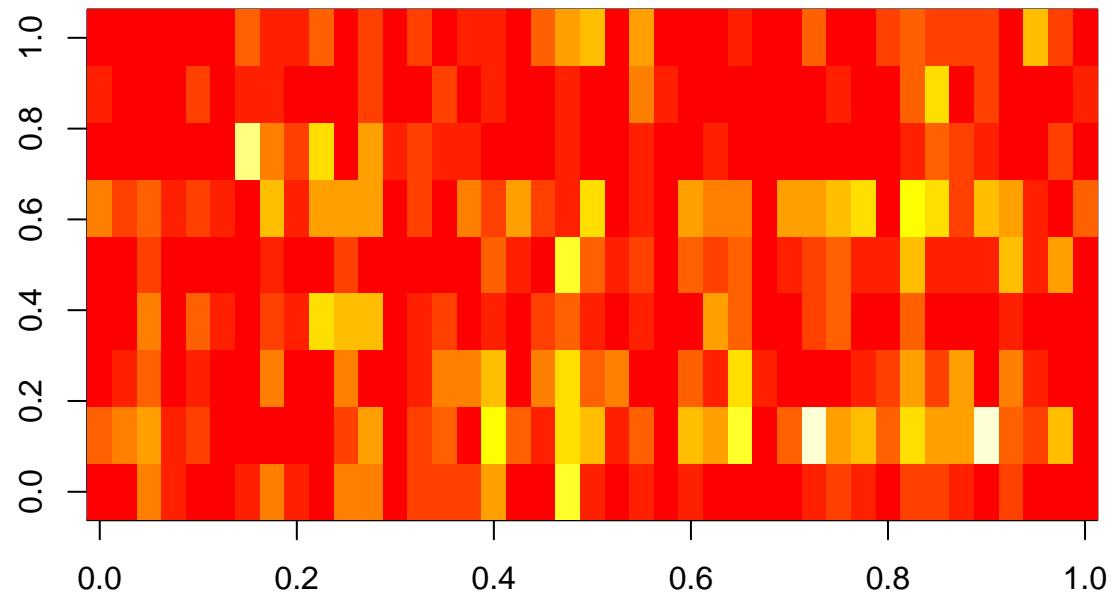
X.num[i,k] <- num_hw
X.sev[i,k] <- sev_hw
}

image(X.num)

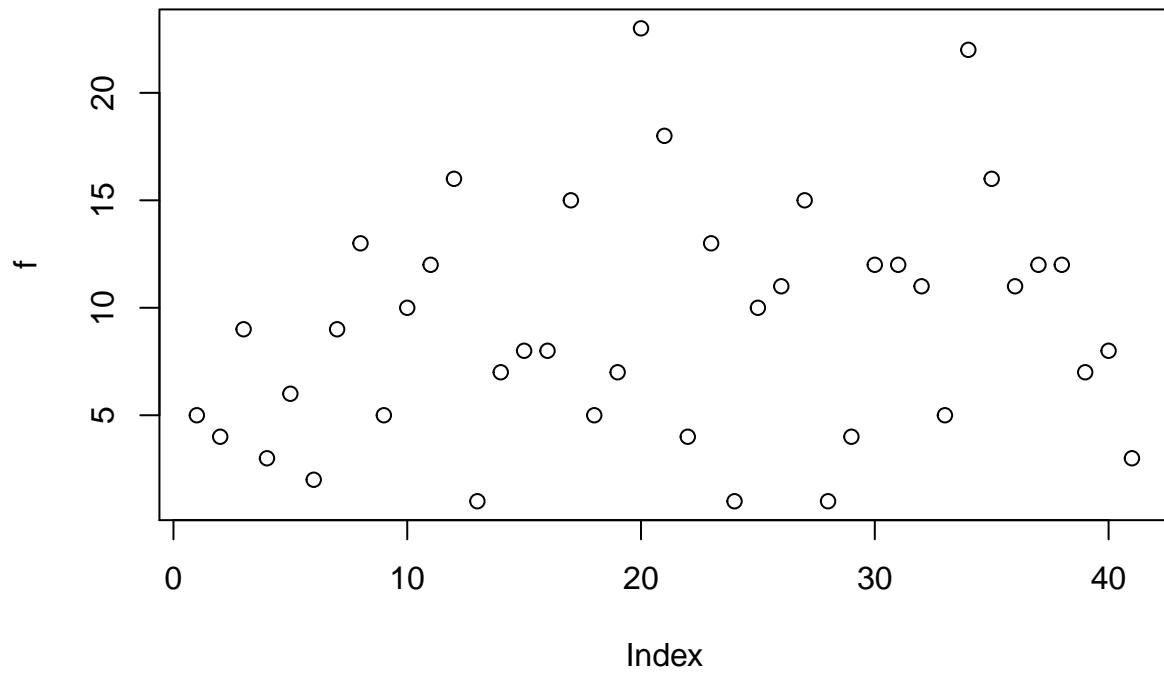
```



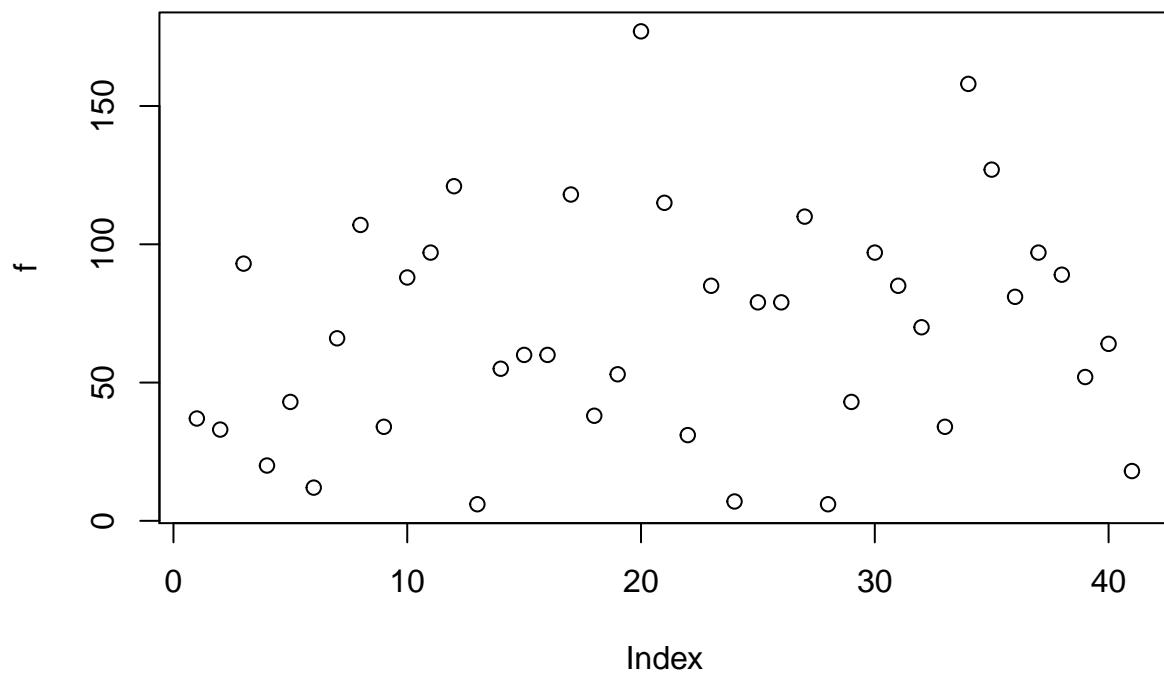
```
image(X.sev)
```



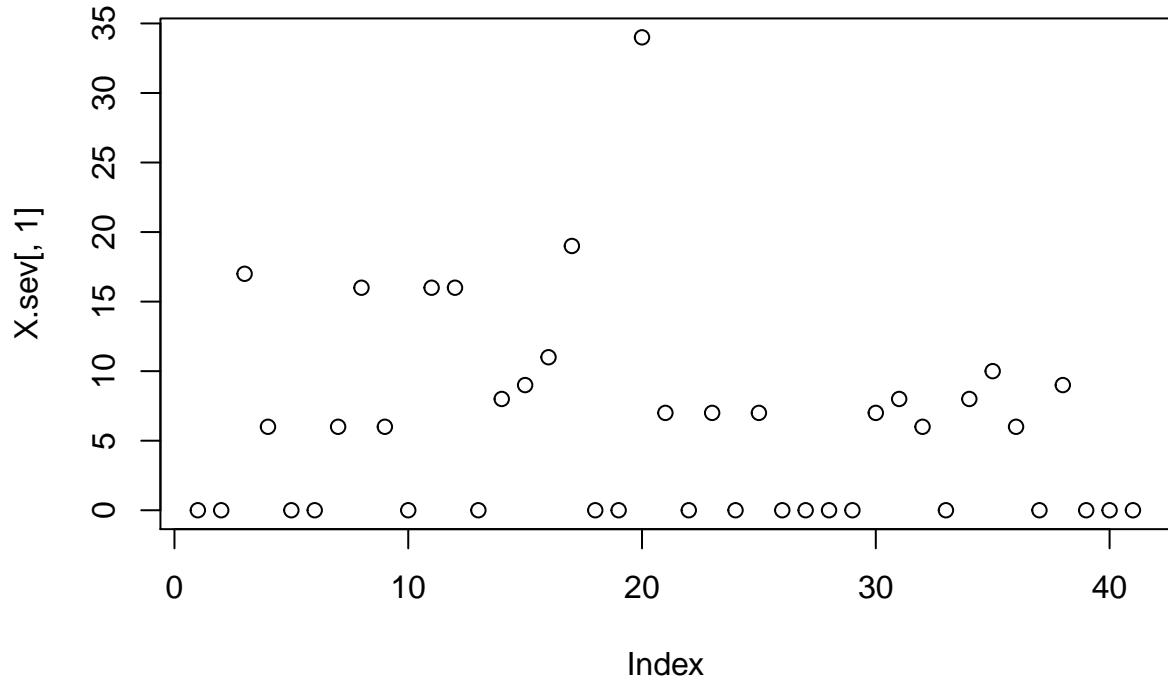
```
f<- rowSums(X.num)
plot(f)
```



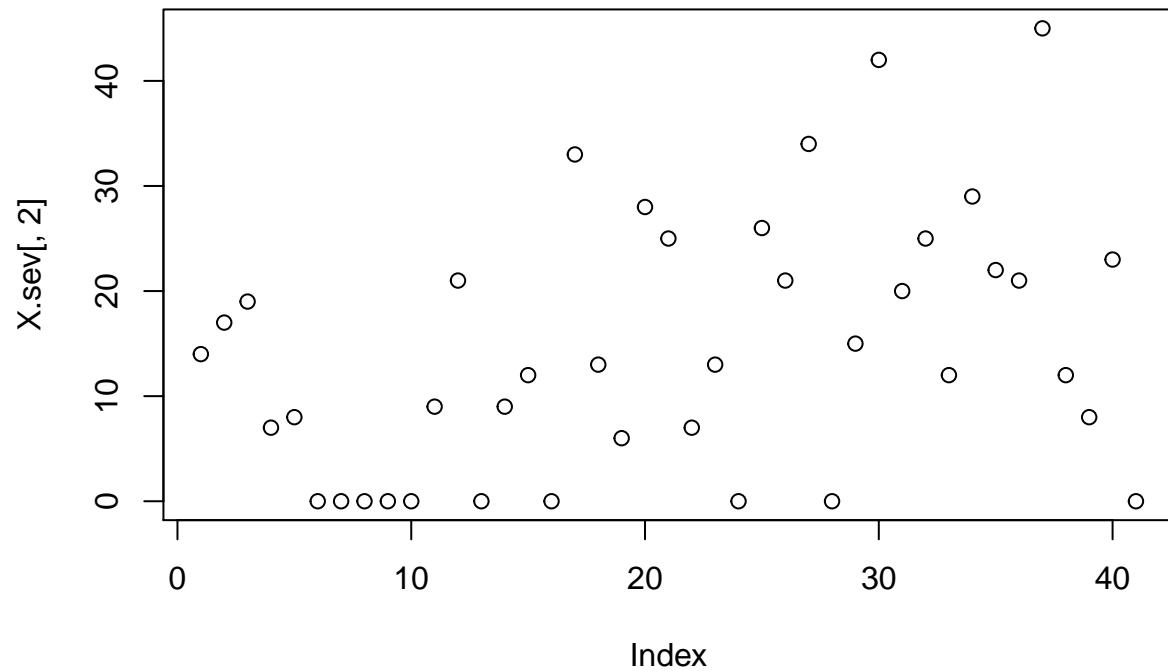
```
f<- rowSums(X.sev)
plot(f)
```



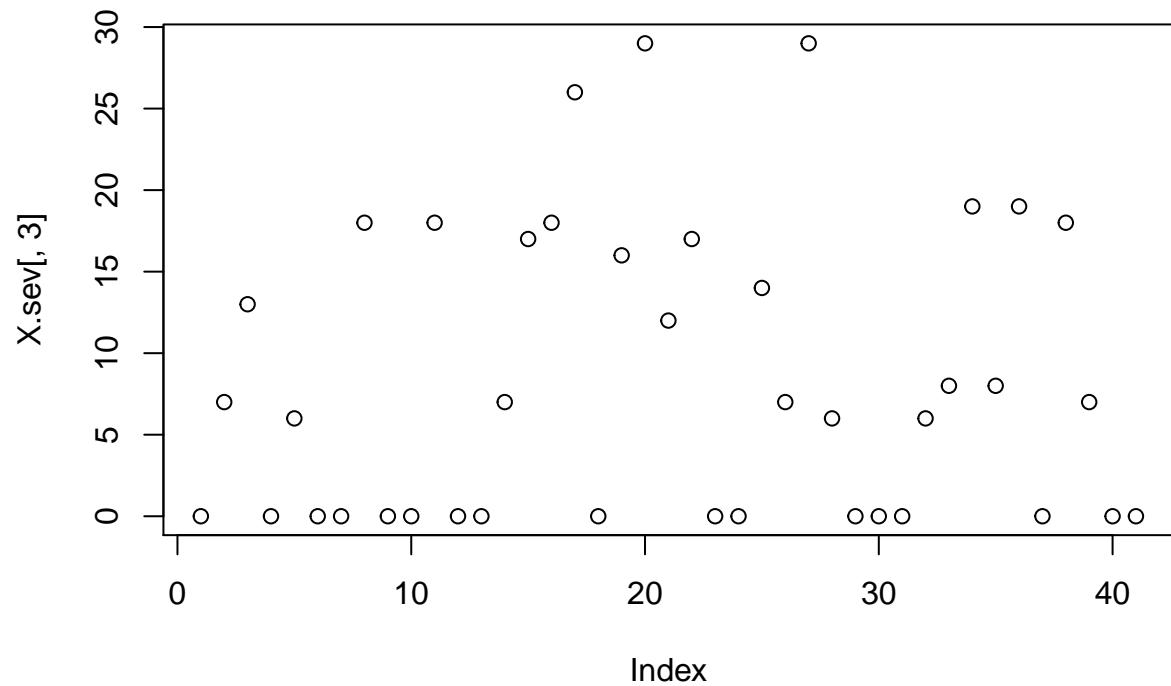
```
plot(X.sev[,1])
```



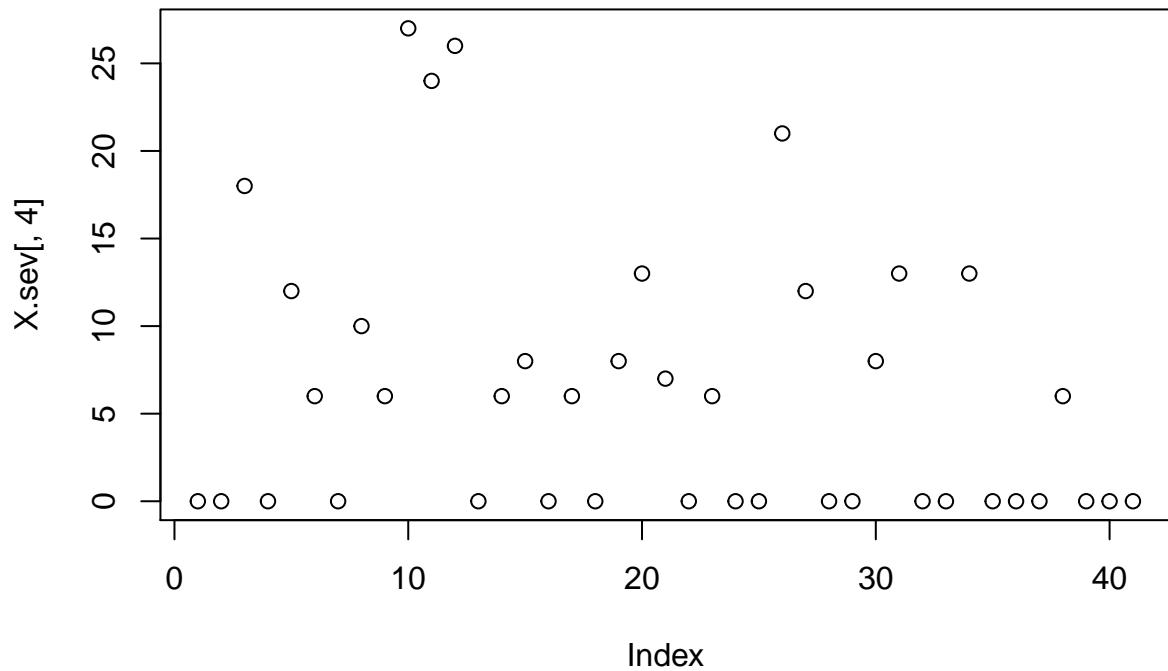
```
plot(X.sev[, 2])
```



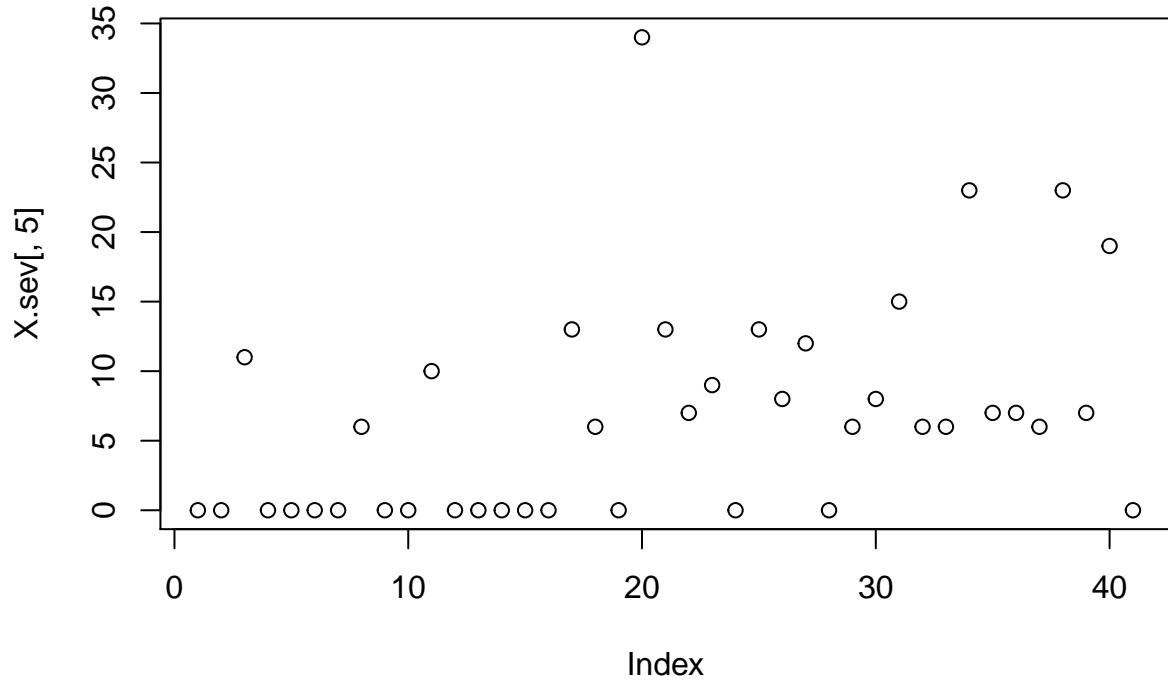
```
plot(X.sev[, 3])
```



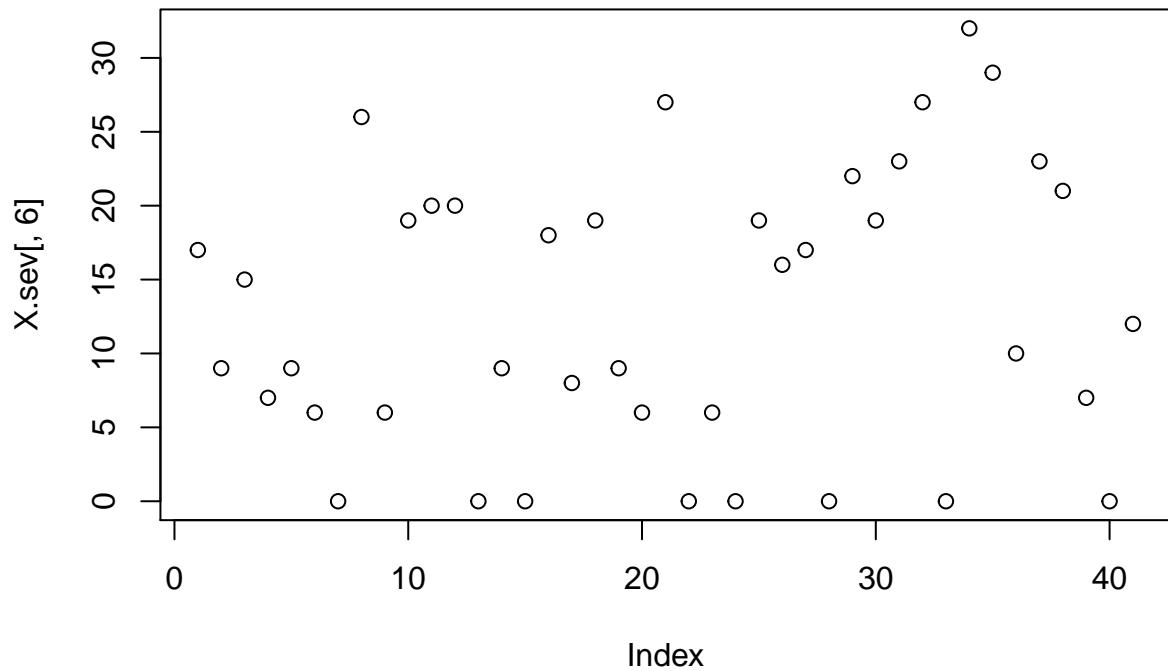
```
plot(X.sev[, 4])
```



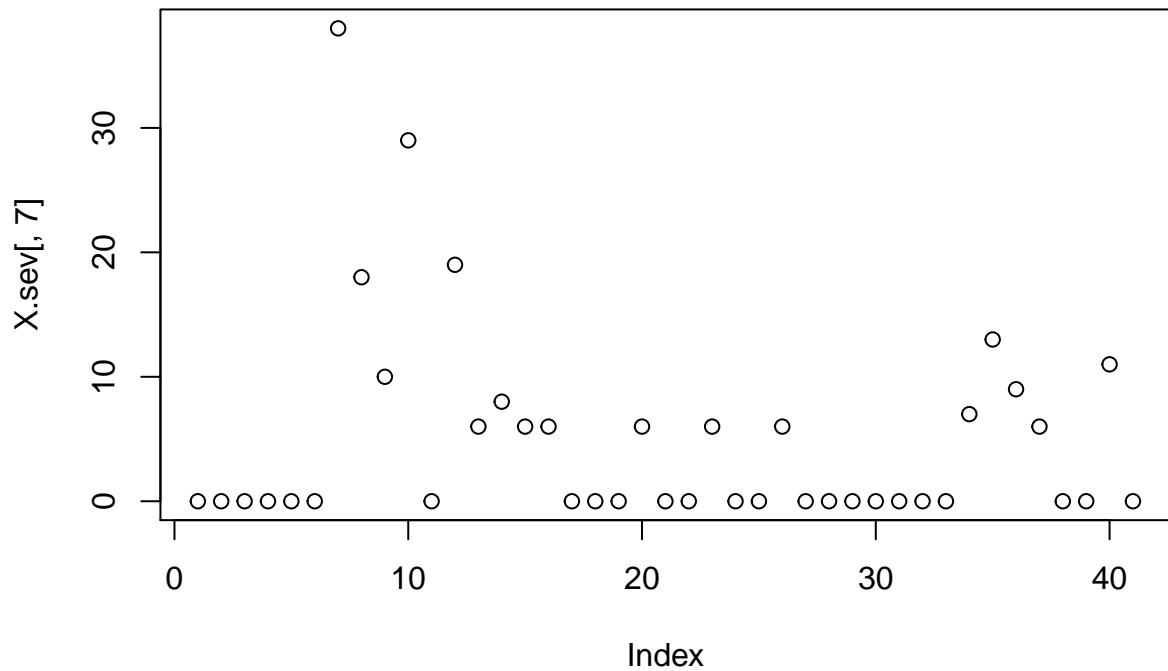
```
plot(X.sev[, 5])
```



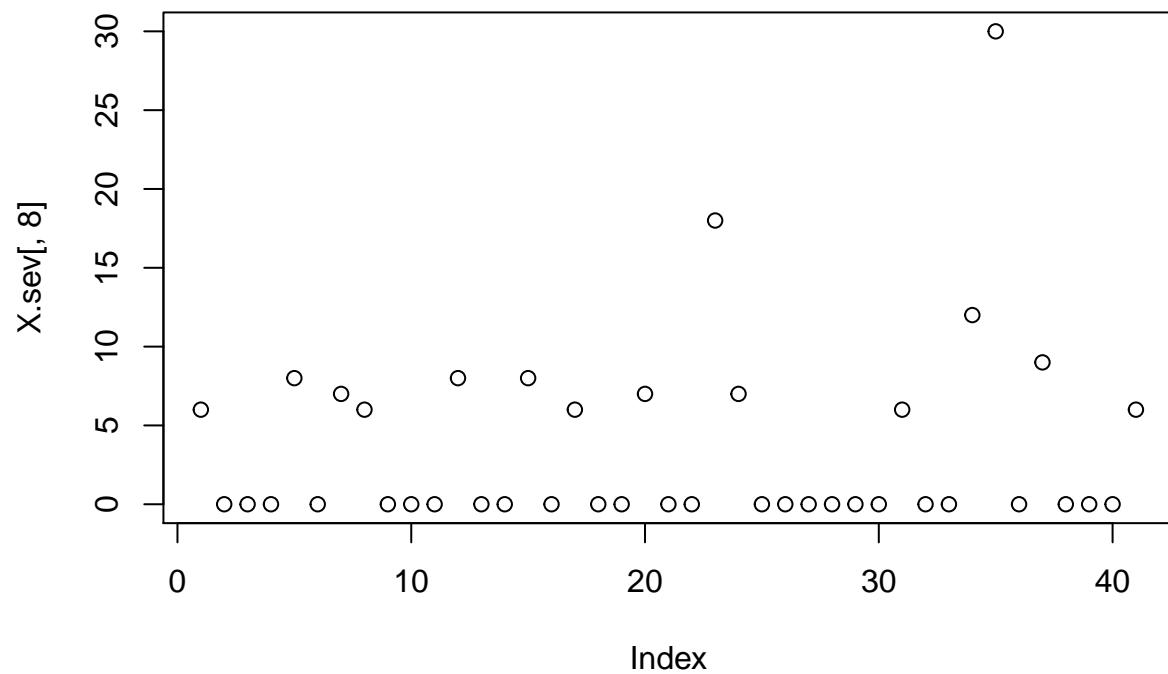
```
plot(X.sev[, 6])
```



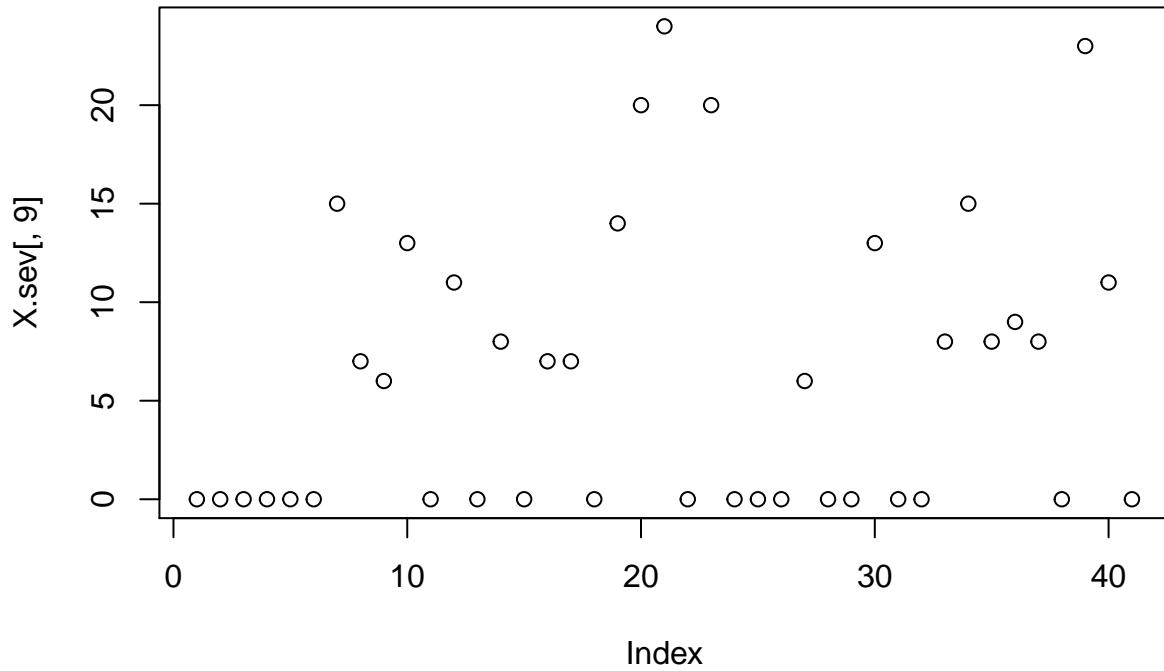
```
plot(X.sev[, 7])
```



```
plot(X.sev[,8])
```



```
plot(X.sev[, 9])
```

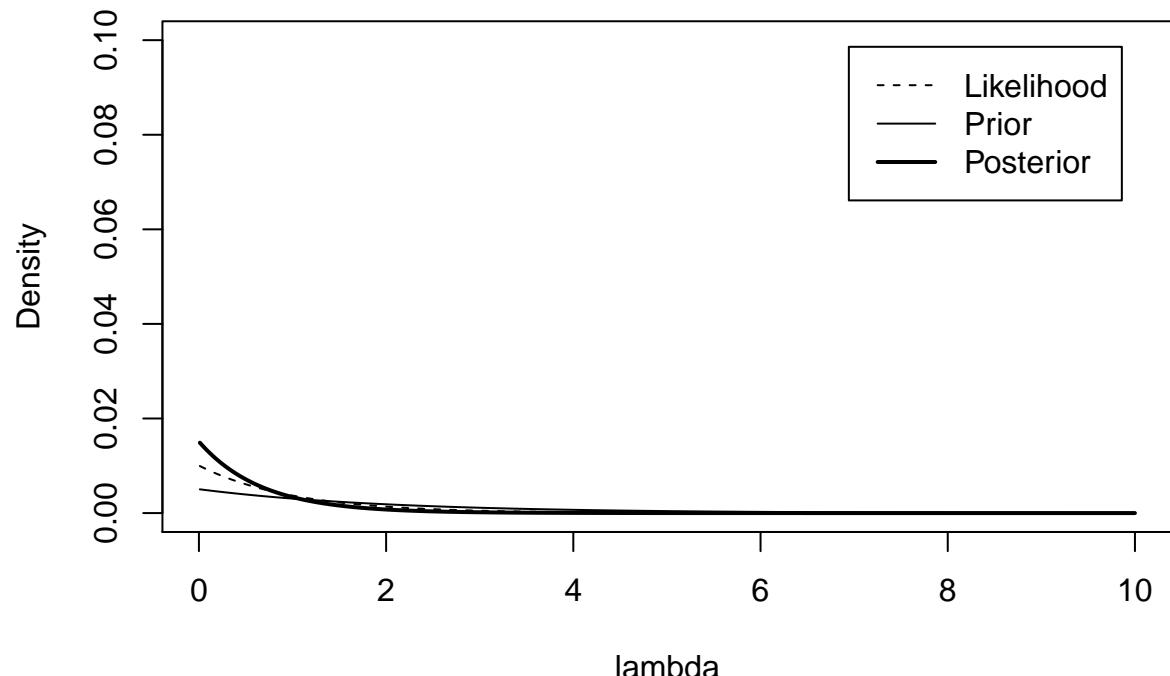


Plot the prior, likelihood, and posterior on a grid

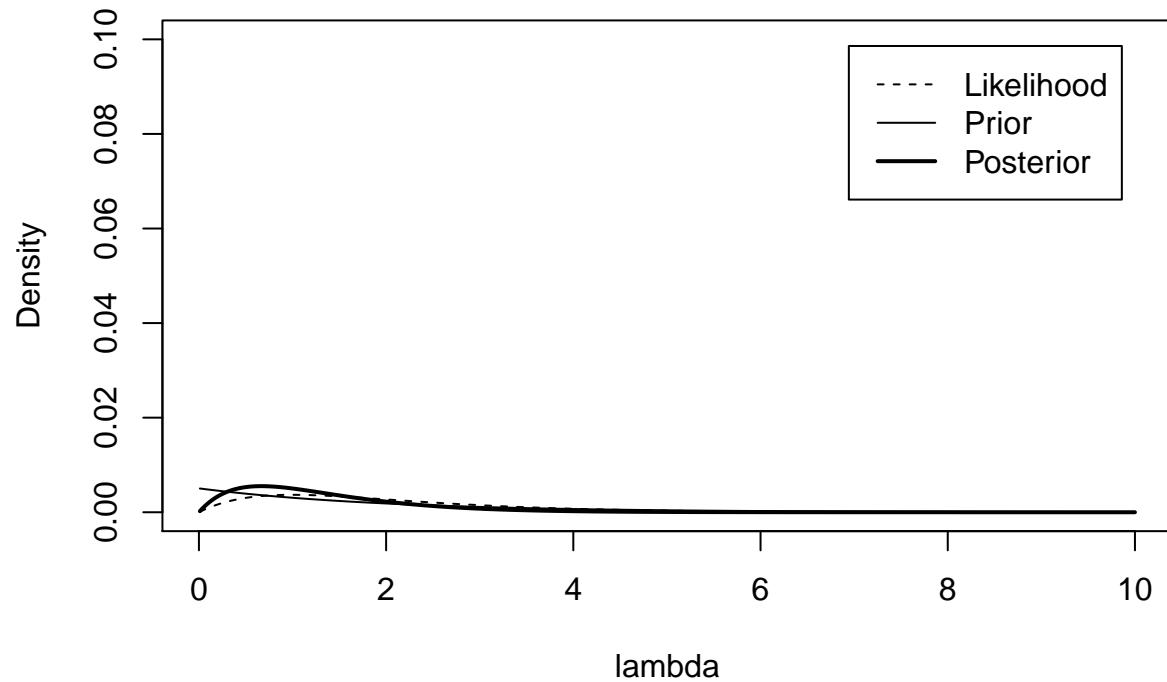
```
#####
#Plot the prior, likelihood, and posterior on a grid
k=1
for(i in 1:41)
{
  N      <- 1
  Y      <- X.num[i,k]
  a      <- 1
  b      <- 0.5
  grid   <- seq(0.01,10,.01)
  like   <- dpois(Y,N*grid)
  like   <- like/sum(like) #standardize
  prior  <- dgamma(grid,a,b)
  prior  <- prior/sum(prior) #standardize
  post   <- like*prior
  post   <- post/sum(post)
  ps.mean <-sum(post*grid)
  plot(grid,like,type="l",lty=2,
       xlab="lambda",ylab="Density",main=paste(Y," post mean = ",ps.mean,sep = ' '),
       ylim=c(0,.1))
  lines(grid,prior)
  lines(grid,post,lwd=2)
  legend("topright",c("Likelihood","Prior","Posterior"),lwd=c(1,1,2),lty=c(2,1,1),inset=0.05)
```

}

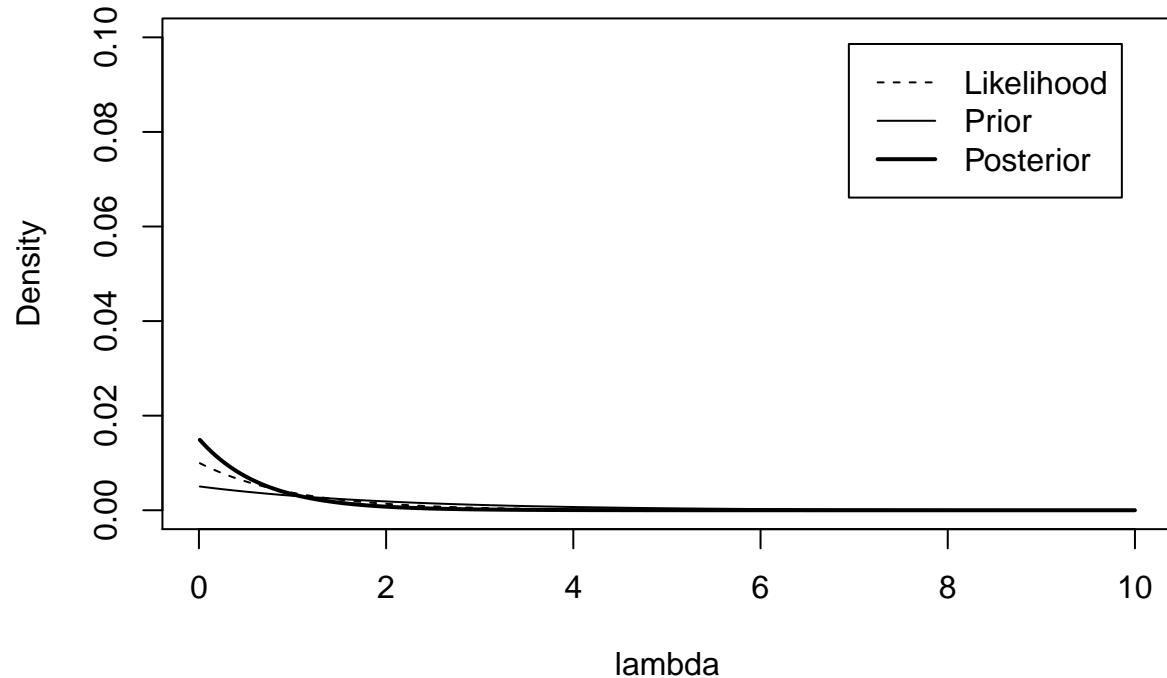
0 post mean = 0.671676107595651



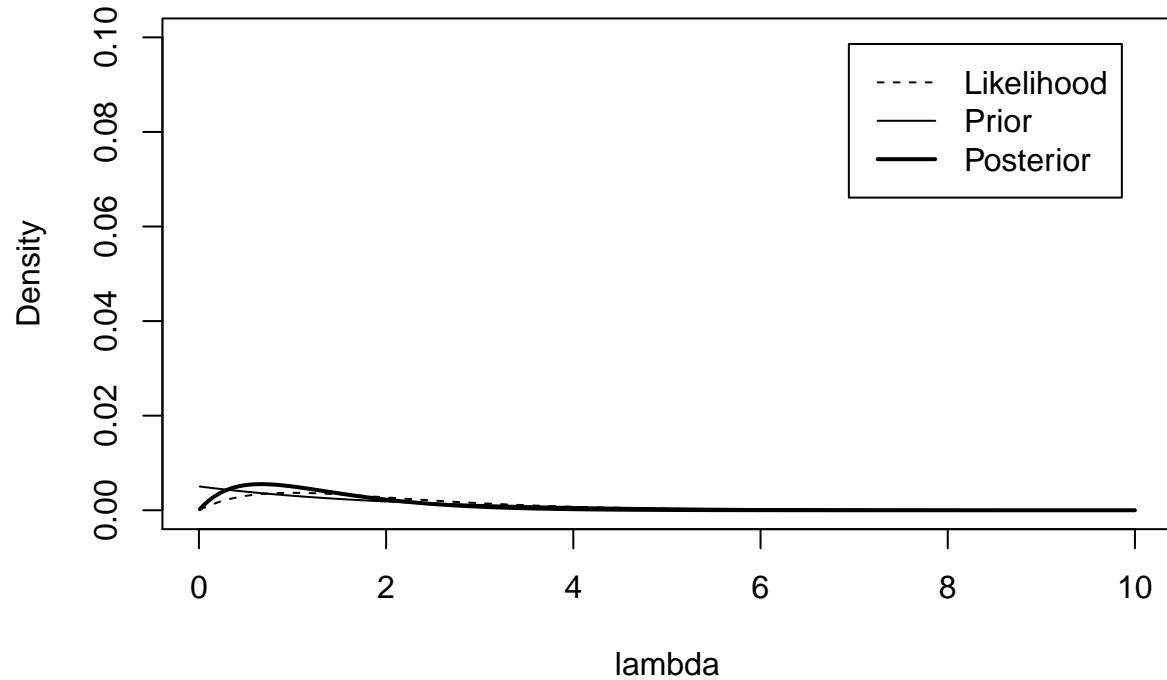
1 post mean = 1.33331274455286



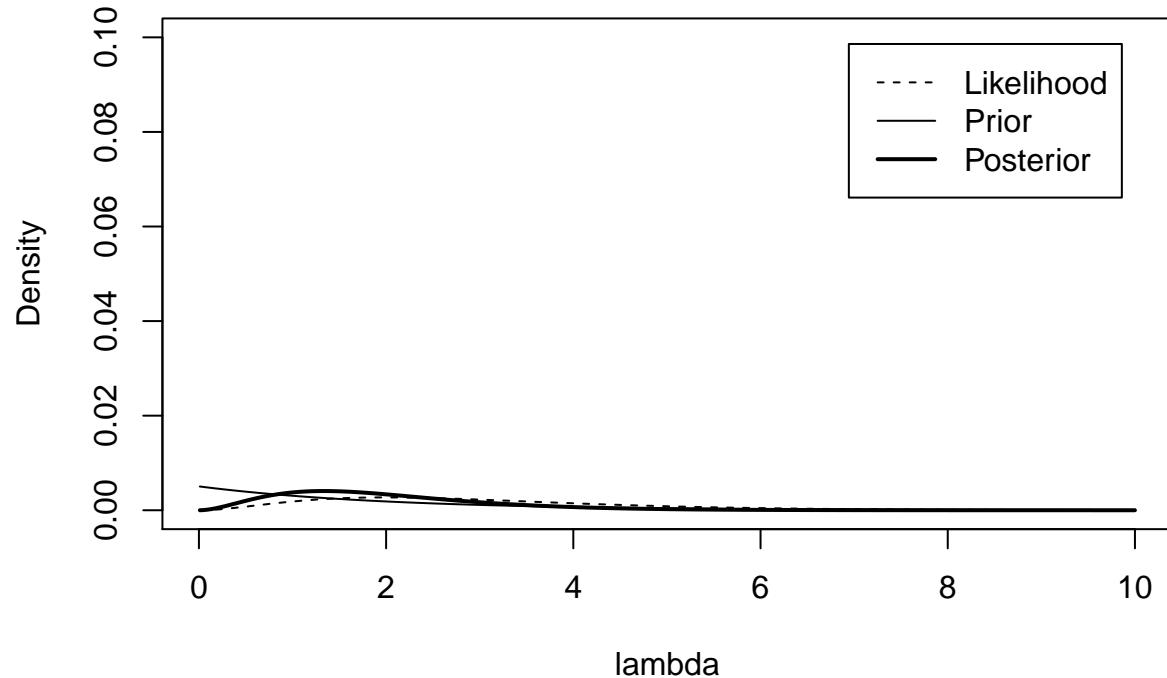
0 post mean = 0.671676107595651



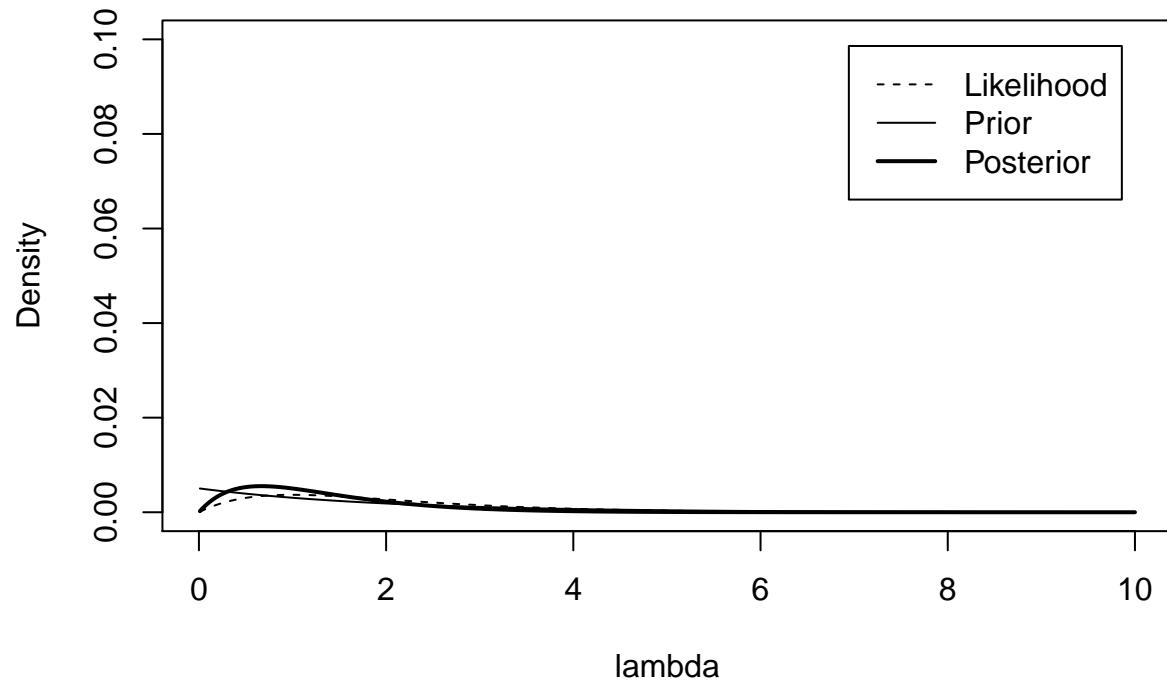
1 post mean = 1.33331274455286



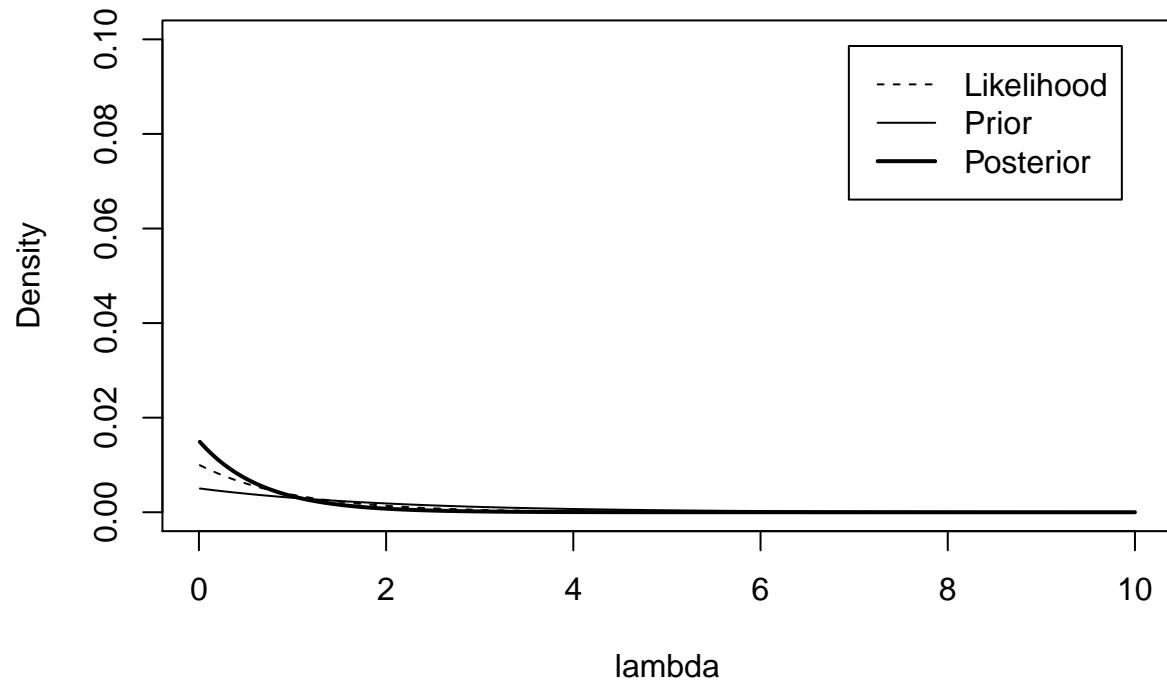
2 post mean = 1.99965790788987



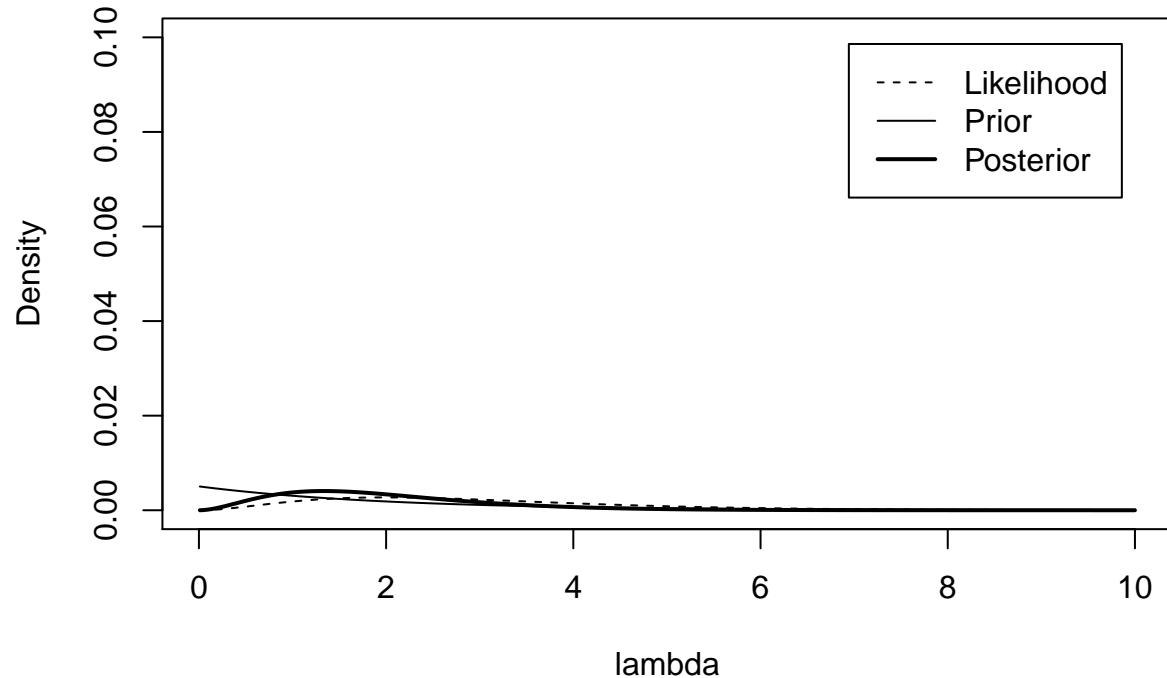
1 post mean = 1.33331274455286



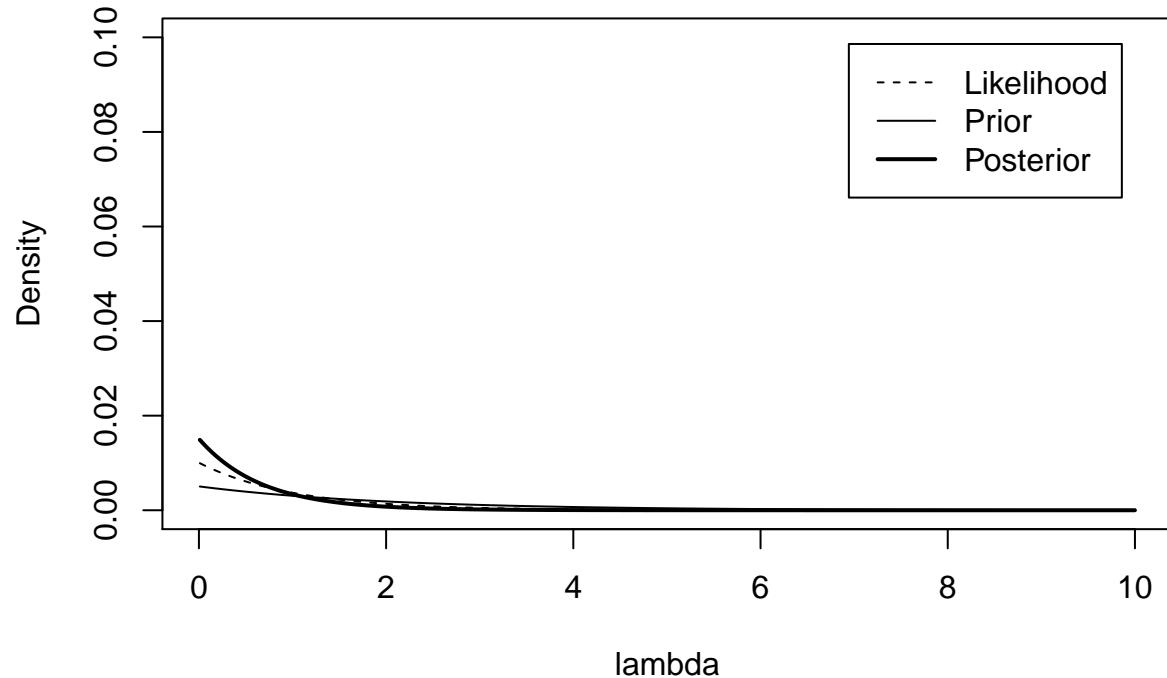
0 post mean = 0.671676107595651



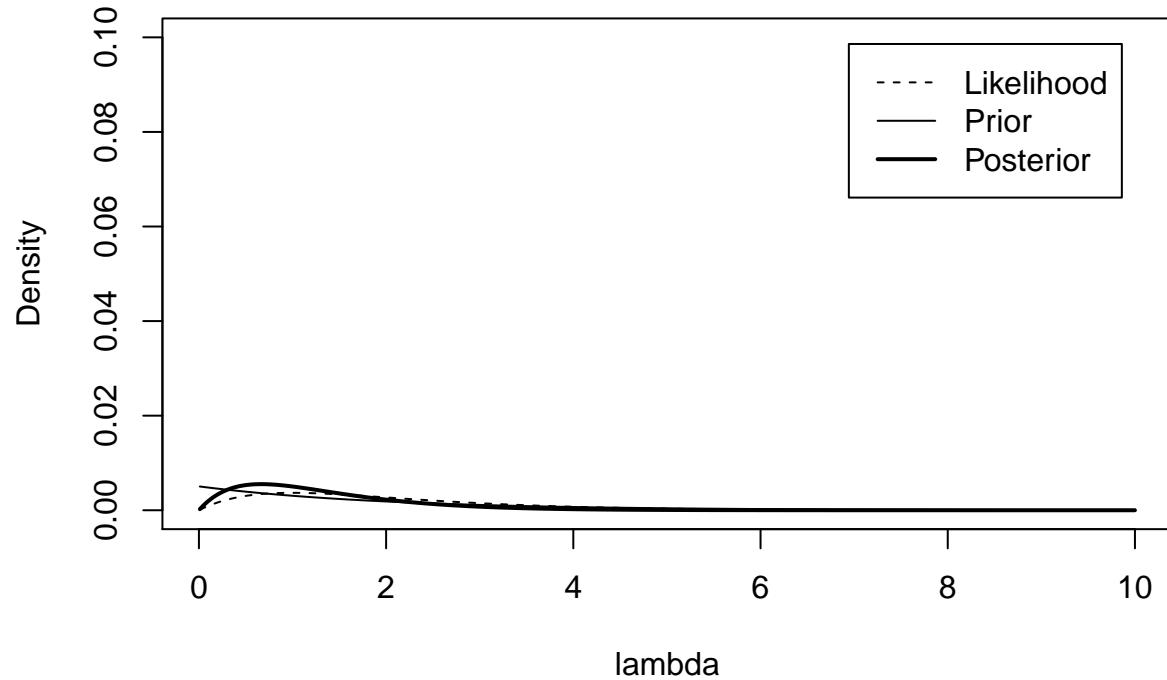
2 post mean = 1.99965790788987



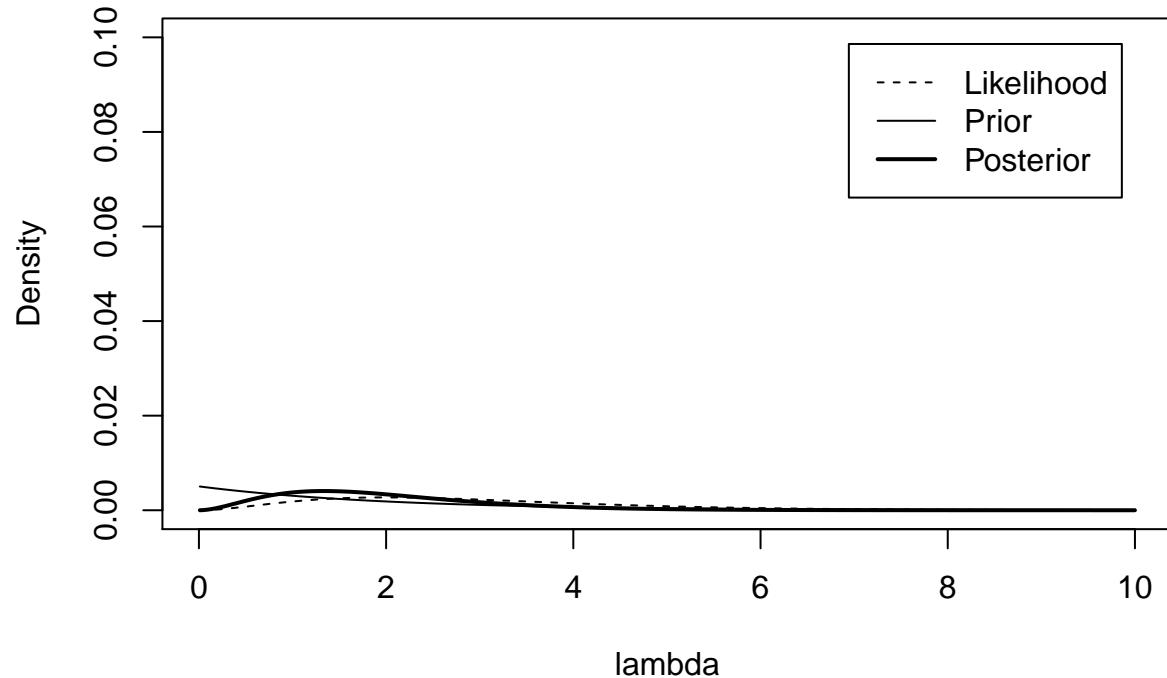
0 post mean = 0.671676107595651



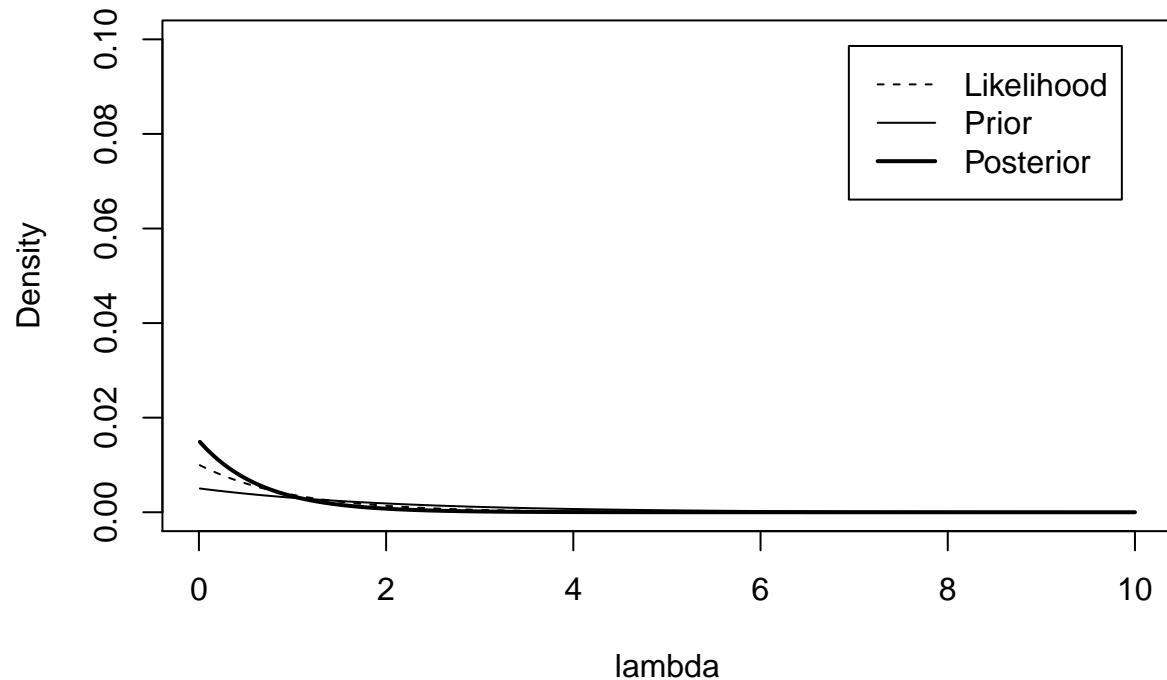
1 post mean = 1.33331274455286



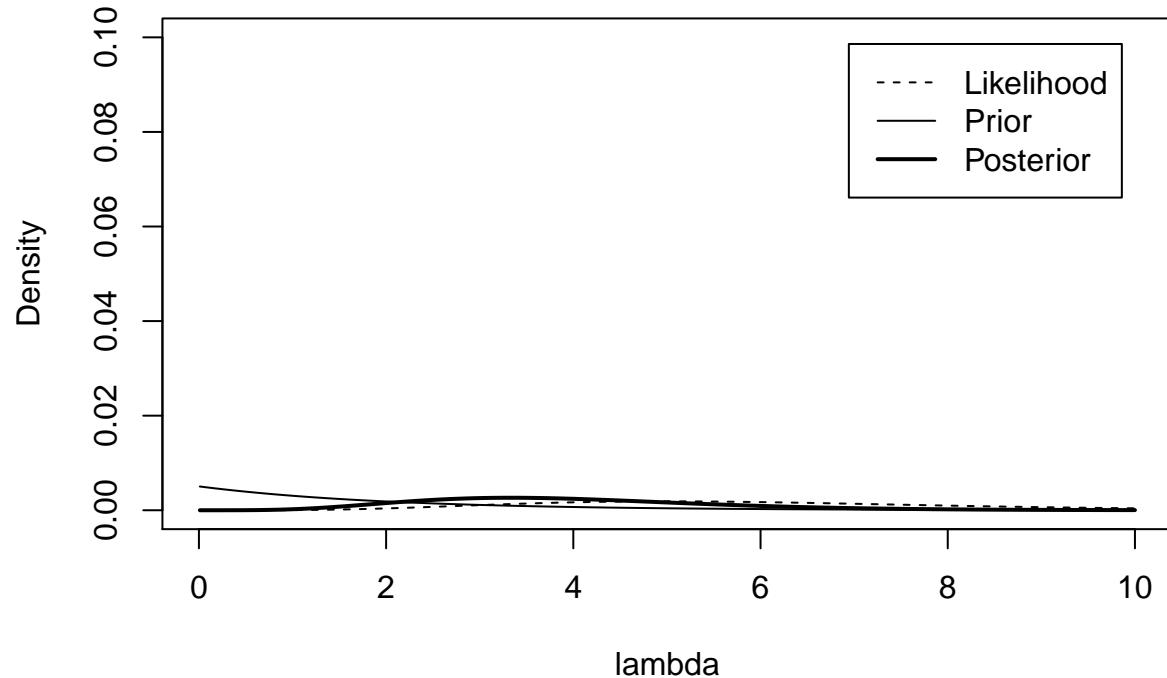
2 post mean = 1.99965790788987



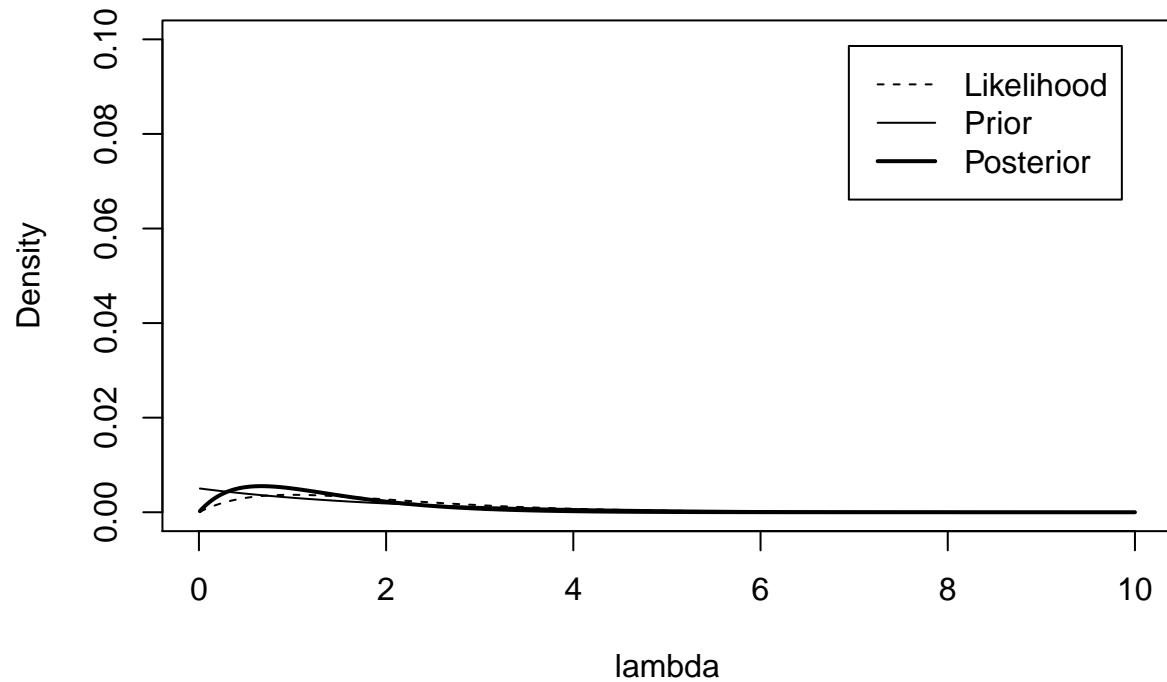
0 post mean = 0.671676107595651



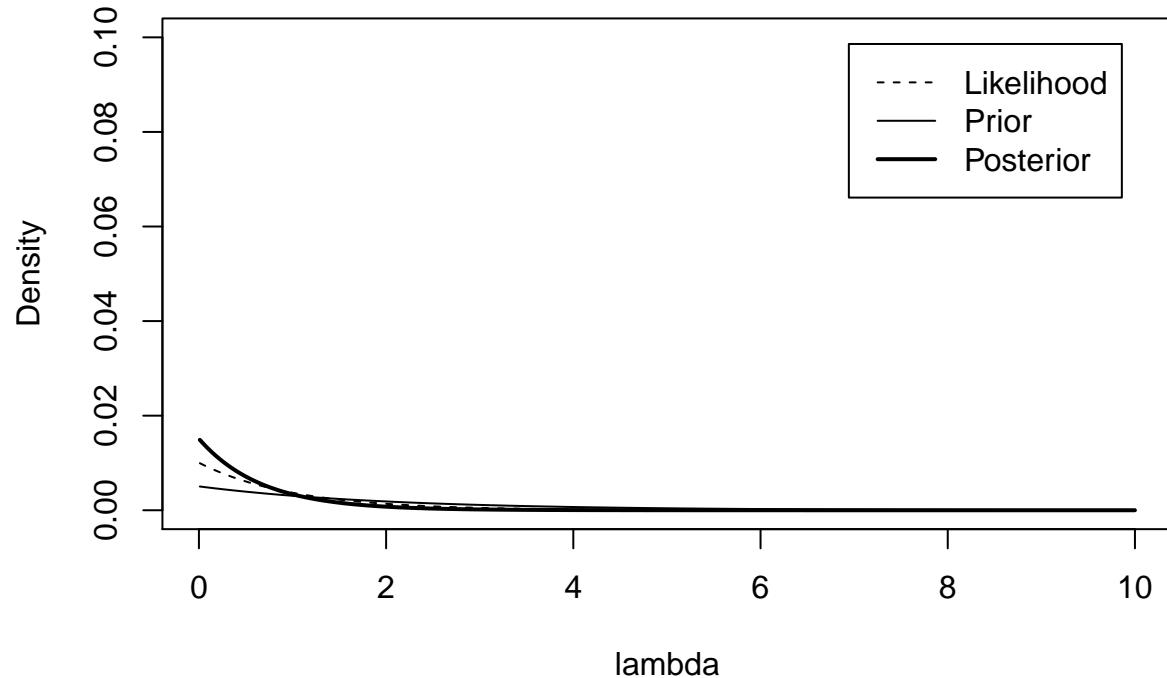
5 post mean = 3.98067542584636



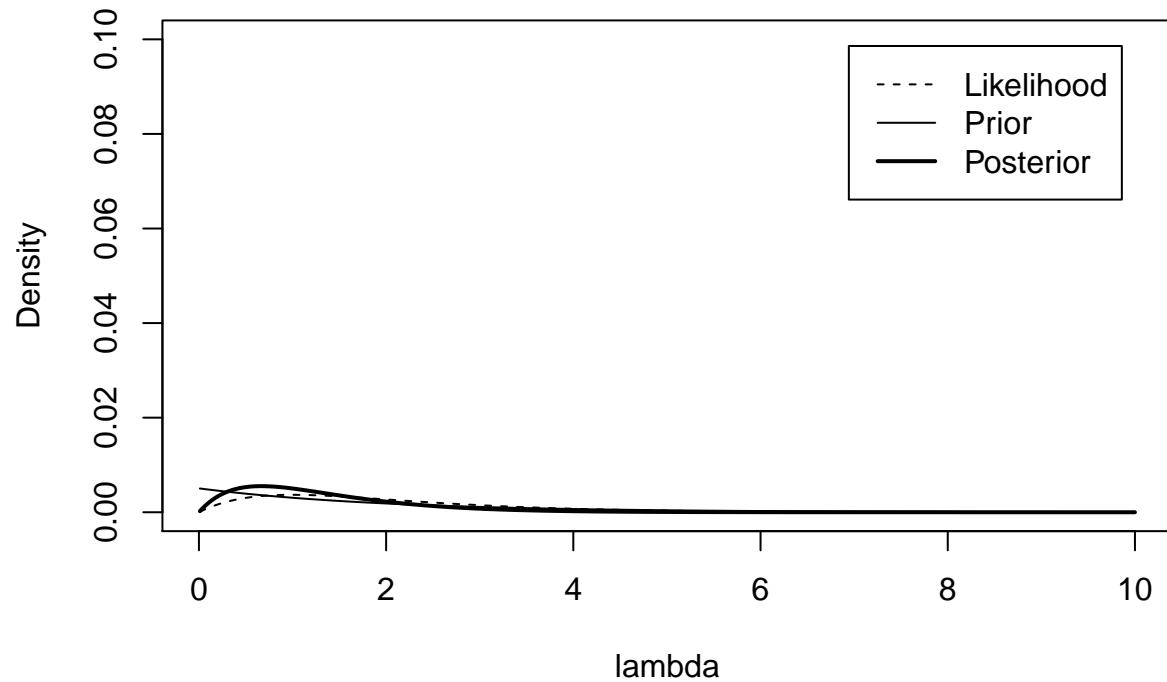
1 post mean = 1.33331274455286



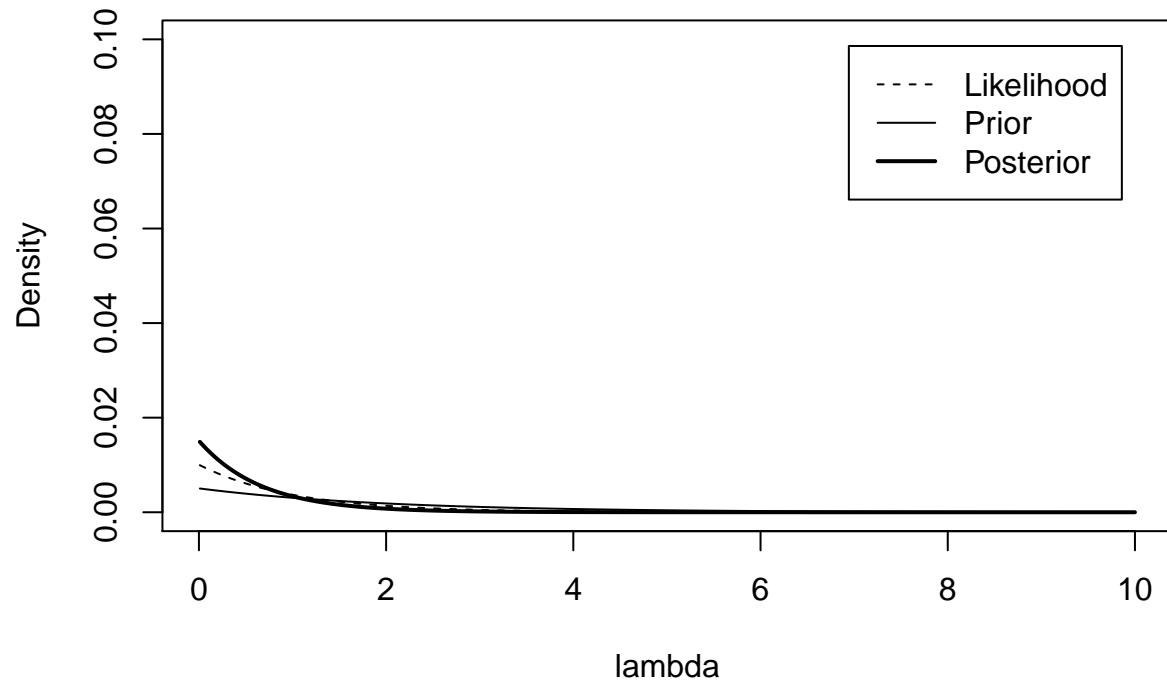
0 post mean = 0.671676107595651



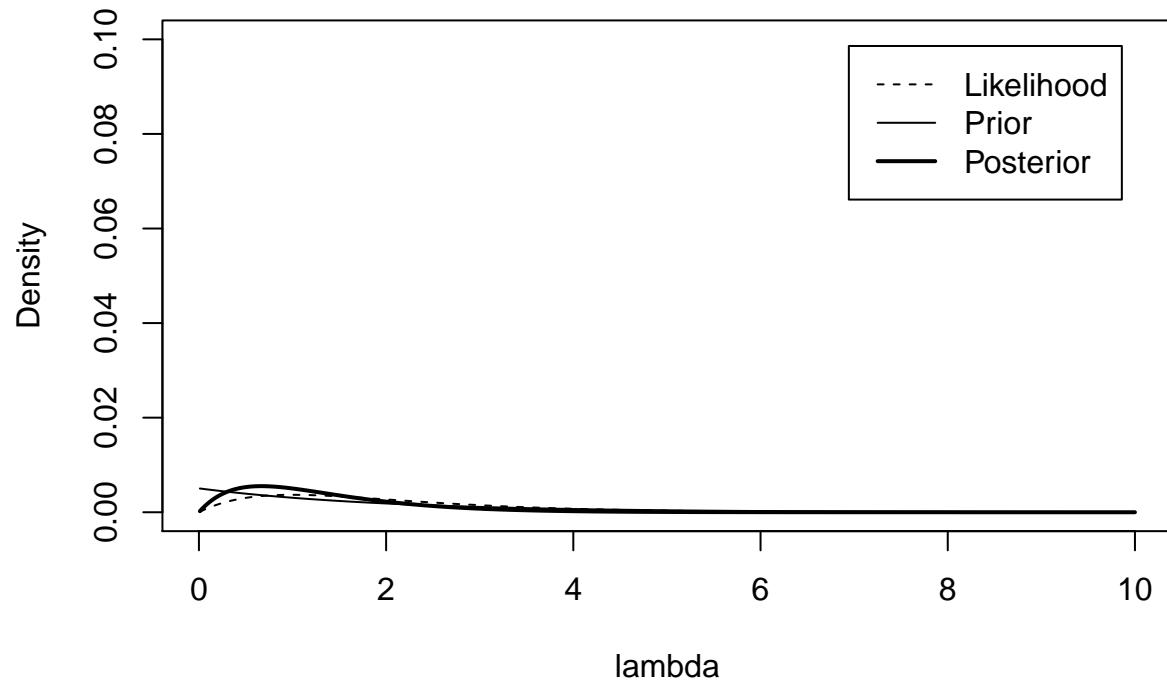
1 post mean = 1.33331274455286



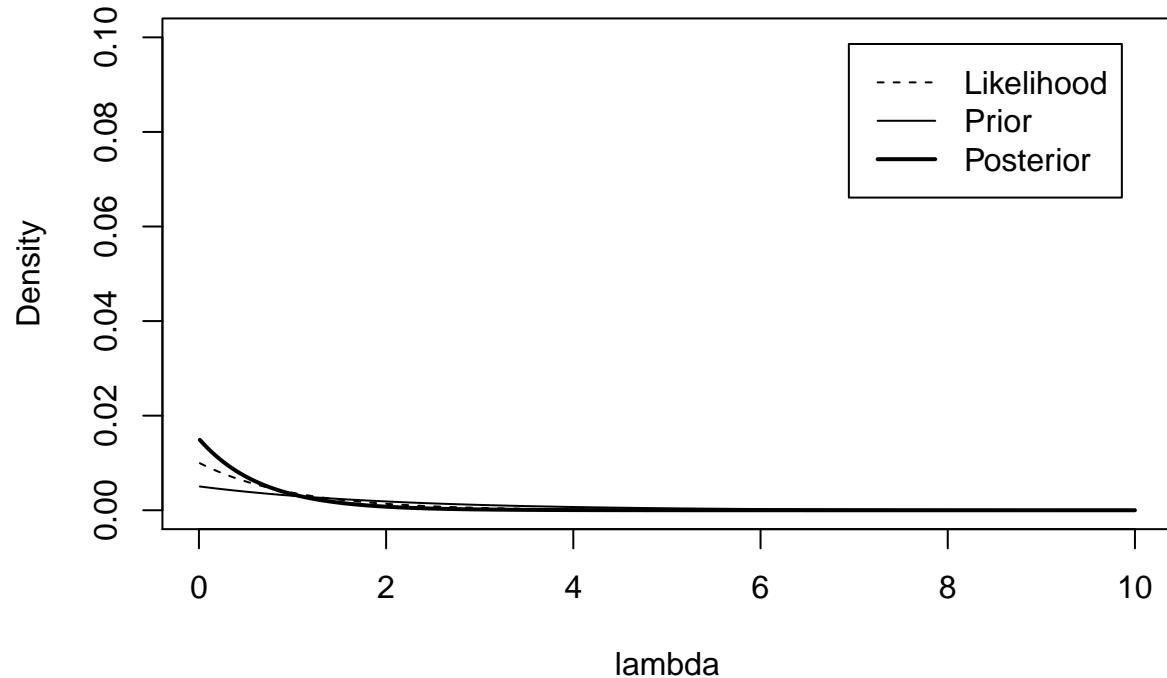
0 post mean = 0.671676107595651



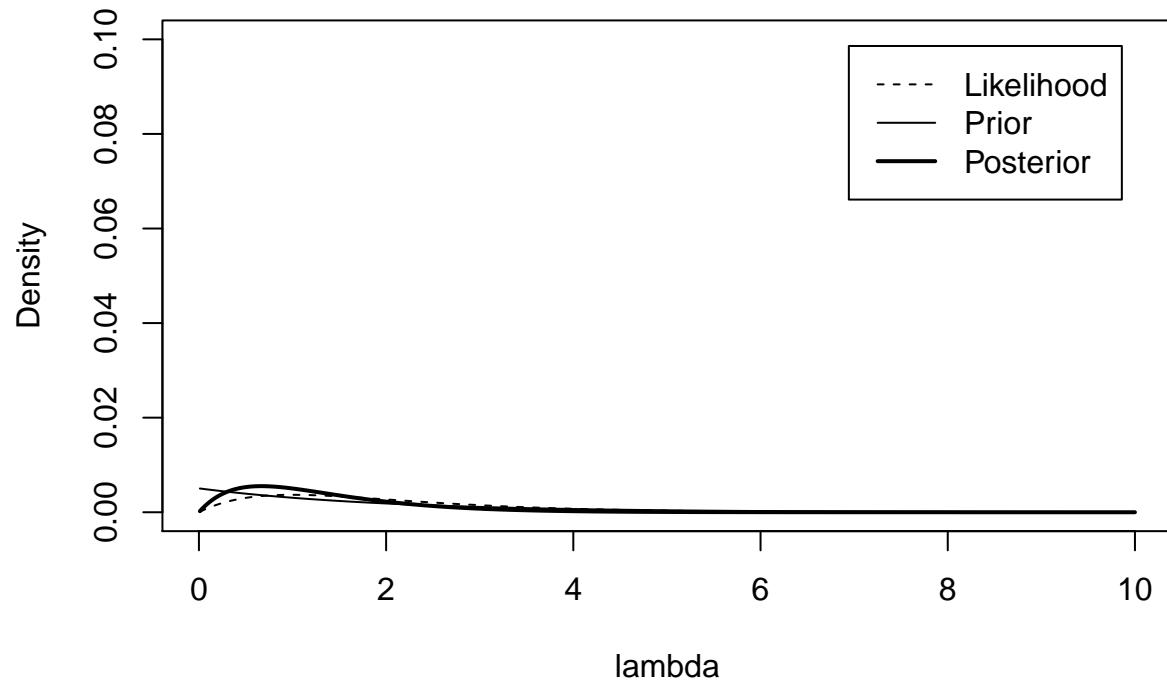
1 post mean = 1.33331274455286



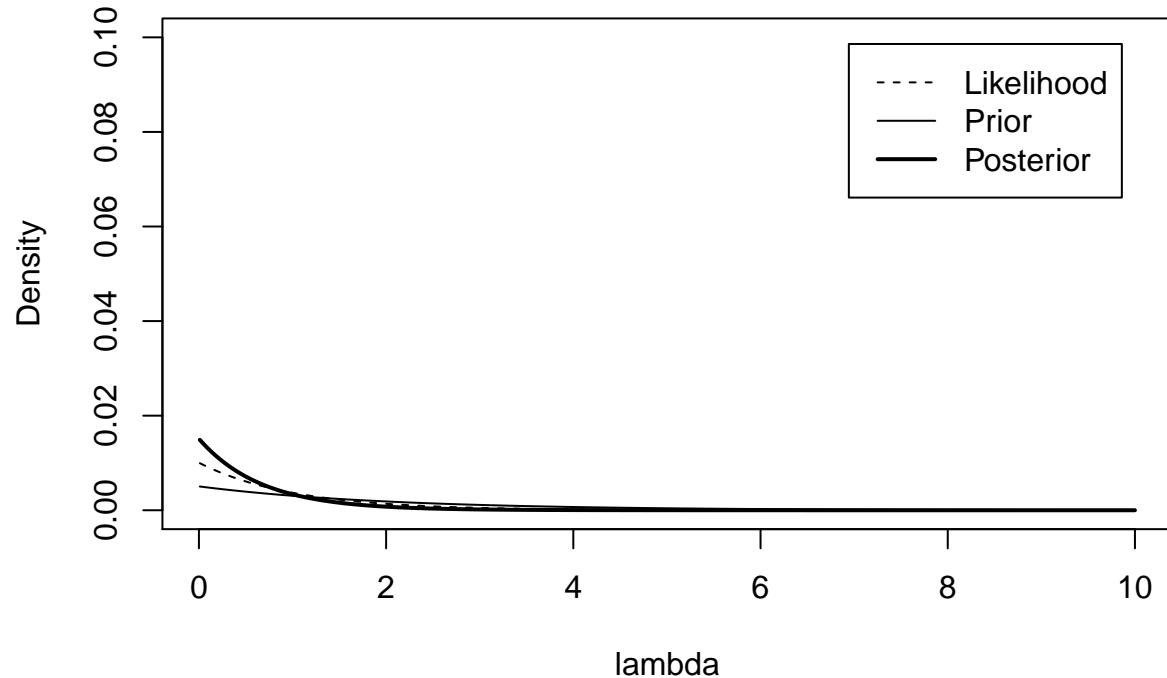
0 post mean = 0.671676107595651



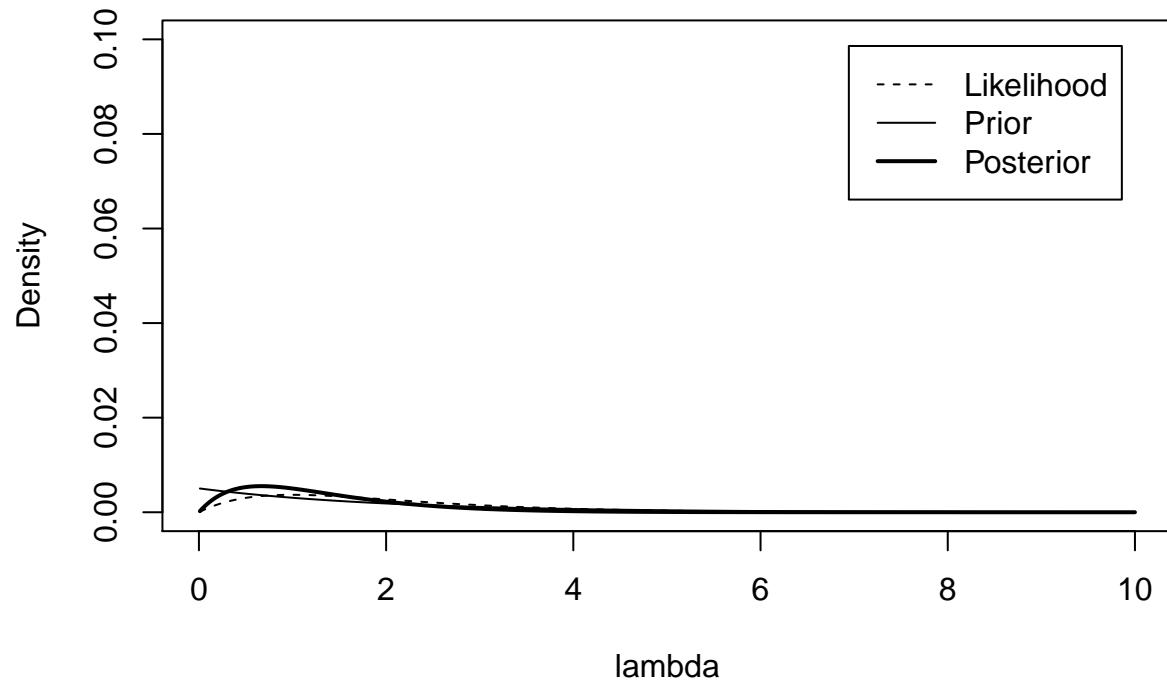
1 post mean = 1.33331274455286



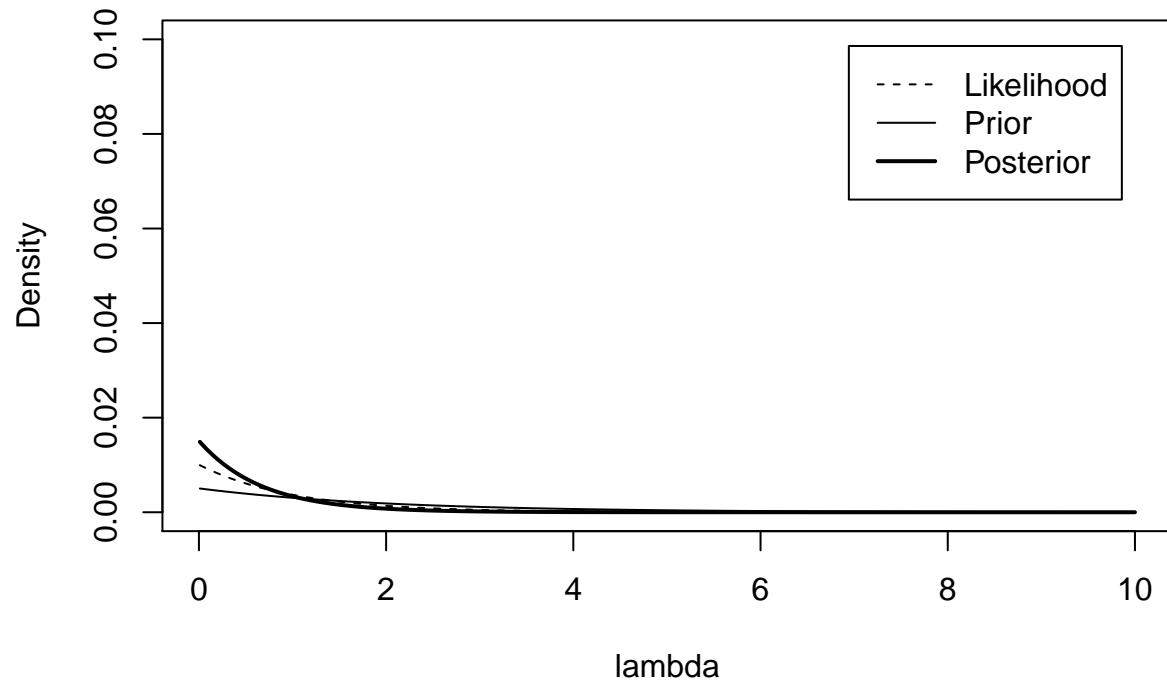
0 post mean = 0.671676107595651



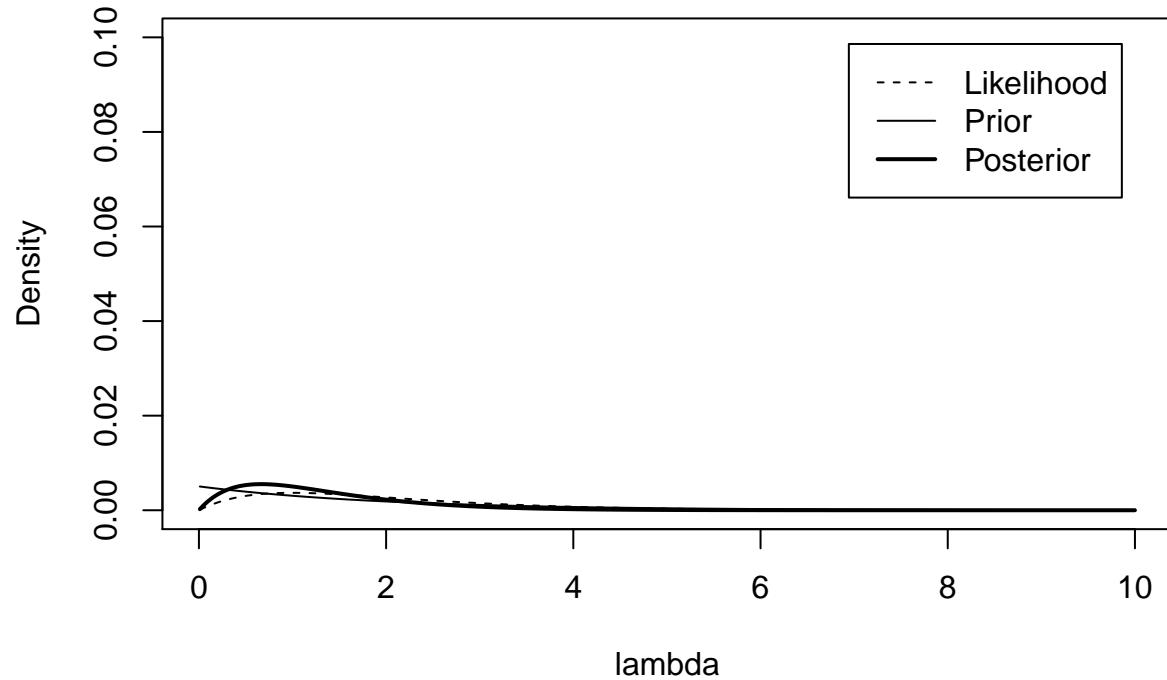
1 post mean = 1.33331274455286



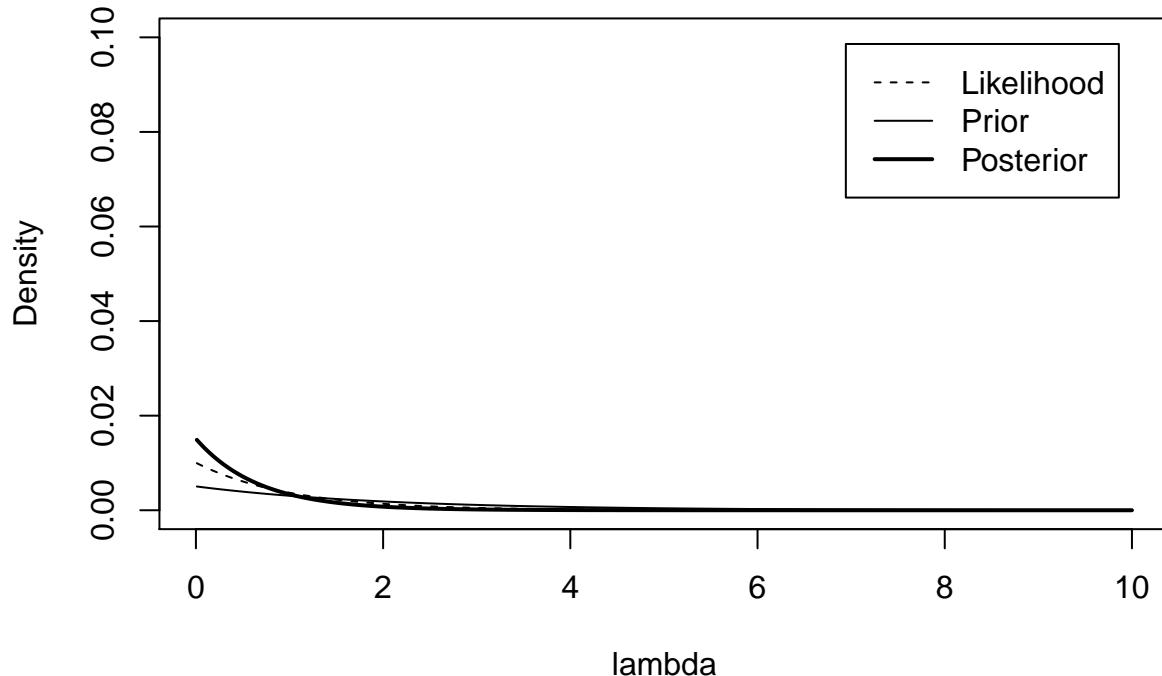
0 post mean = 0.671676107595651



1 post mean = 1.33331274455286



0 post mean = 0.671676107595651



Find MLE of fit to pois and negbinom, and fit glm's with time as predictor

```
#####
# Find MLE of fit to pois and negbinom, and fit glm's with time as predictor
#####
mle.nb.params <- matrix(nrow = 9, ncol = 2)
mle.pois.params <- matrix(nrow = 9, ncol = 1)

mle.nb.glm.params <- matrix(nrow = 9, ncol = 8)
mle.pois.glm.params <- matrix(nrow = 9, ncol = 8)
for(k in 1:9)
{
  hist(X.num[,k])
  library(fitdistrplus)
  library(gamlss)
  fit_nb <- fitdist(X.num[,k], 'nbinom', start = list(mu = 3, size = 0.1))
  mle.nb.params[k,1] <- fit_nb$estimate[1]
  mle.nb.params[k,2] <- fit_nb$estimate[2]
  plot(fit_nb)
  #gofstat(fit_nb)

  fit_pois <- fitdist(X.num[,k], 'pois', method = 'mle')
  mle.pois.params[k,1] <- fit_pois$estimate[1]
  plot(fit_pois)
```

```

#gofstat(fit_pois)

df <- data.frame(t=seq(1:41),Y=X.num[,k])
model.pois <- glm( Y~ t, family=poisson, df)
sp <- summary.glm(model.pois)

model.nb <- glm.nb(Y~t,data=df)
snb <- summary.glm(model.nb)

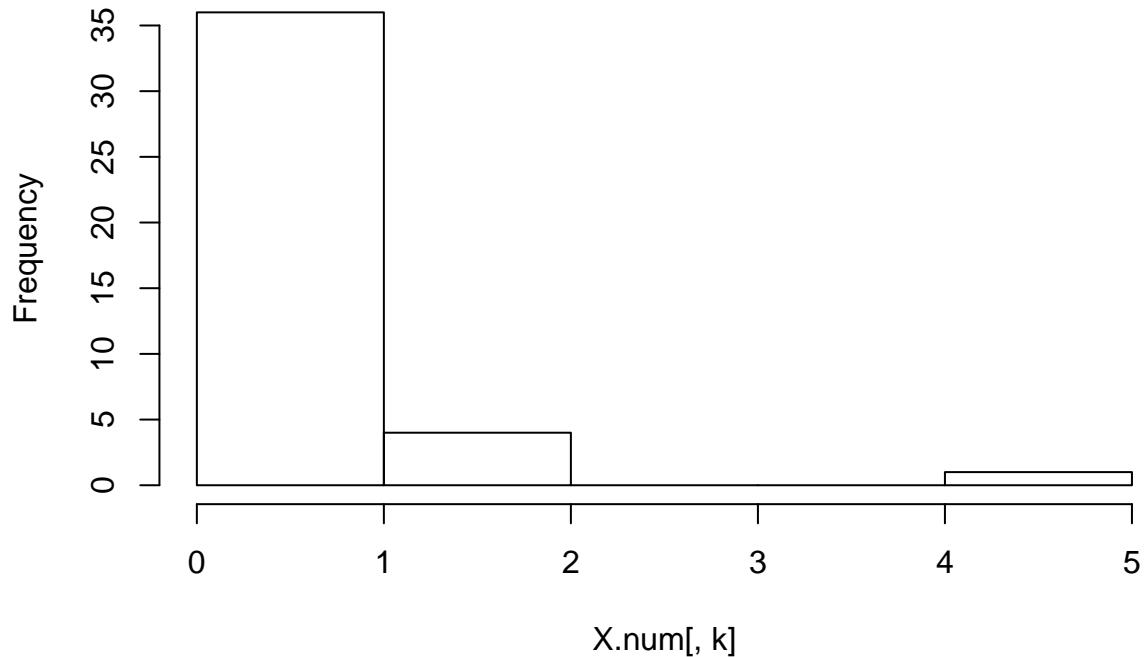
mle.nb.glm.params[k,1:4] <- snb$coefficients[1,]
mle.nb.glm.params[k,5:8] <- snb$coefficients[2,]

mle.pois.glm.params[k,1:4] <- sp$coefficients[1,]
mle.pois.glm.params[k,5:8] <- sp$coefficients[2,]
}

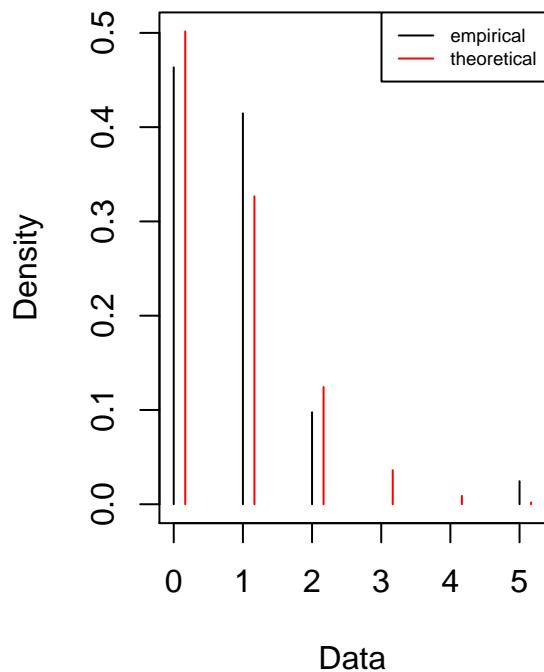
## Loading required package: MASS
## Loading required package: survival
## Loading required package: splines
## Loading required package: gamlss.data
## Loading required package: gamlss.dist
## Loading required package: nlme
## Loading required package: parallel
## **** GAMLSS Version 5.0-6 ****
## For more on GAMLSS look at http://www.gamlss.org/
## Type gamlssNews() to see new features/changes/bug fixes.

```

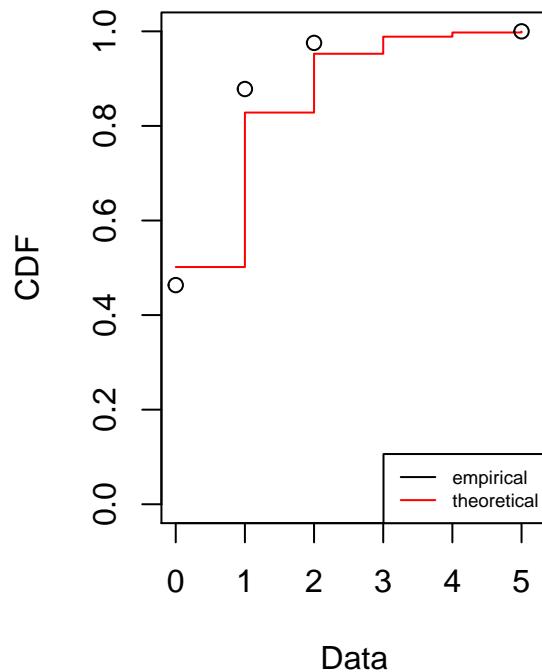
Histogram of X.num[, k]



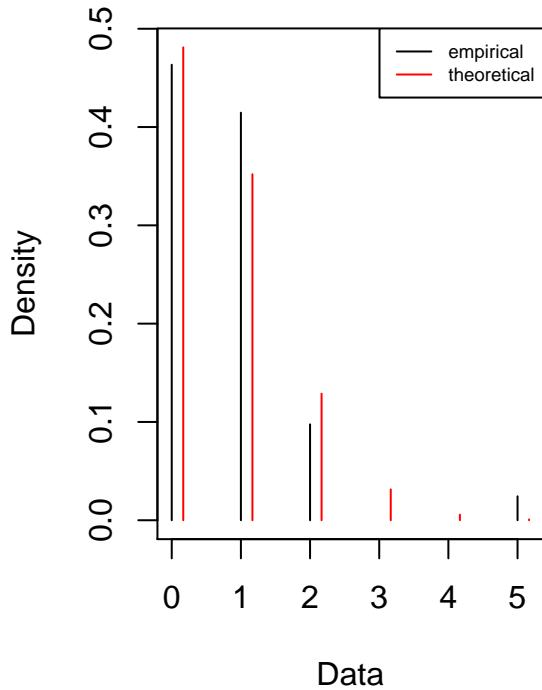
Emp. and theo. distr.



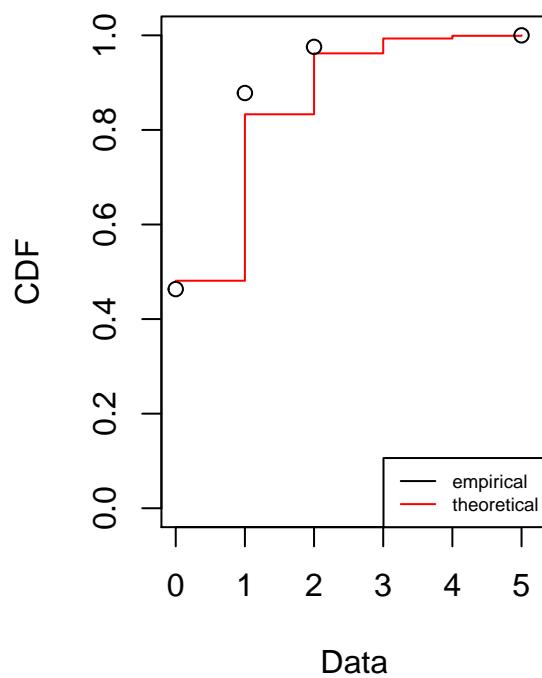
Emp. and theo. CDFs



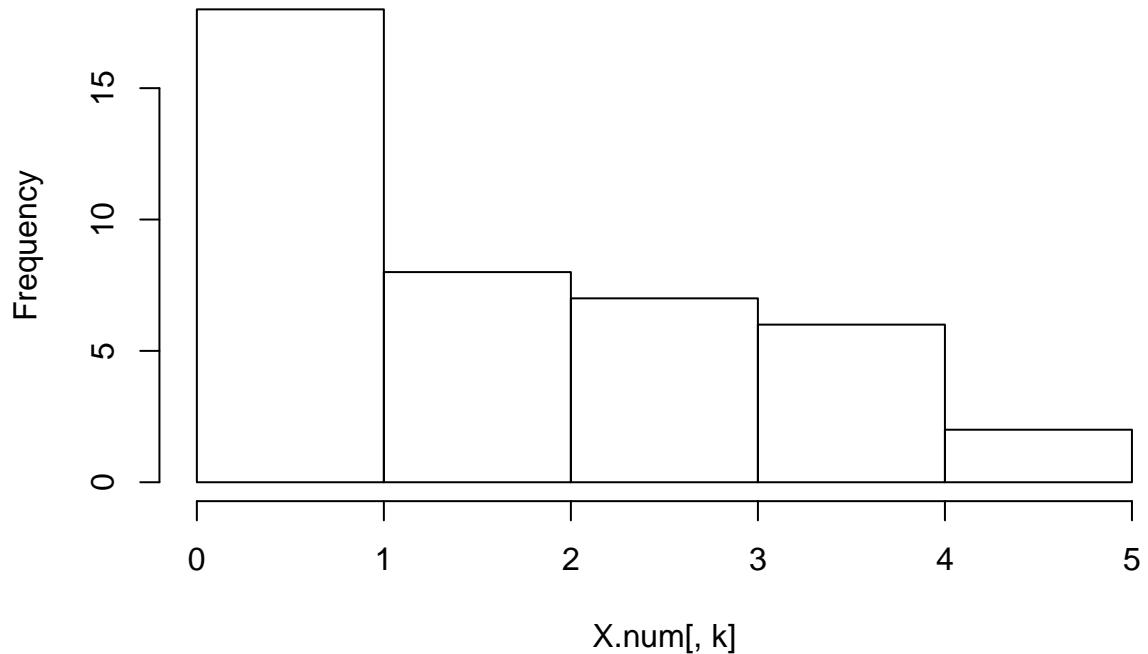
Emp. and theo. distr.



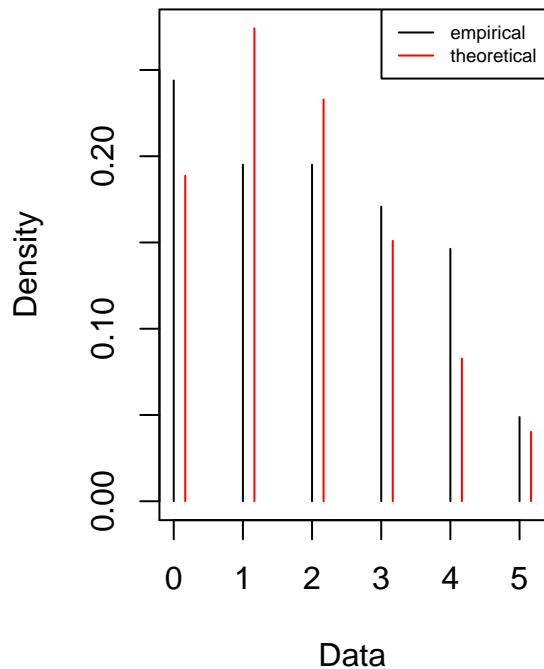
Emp. and theo. CDFs



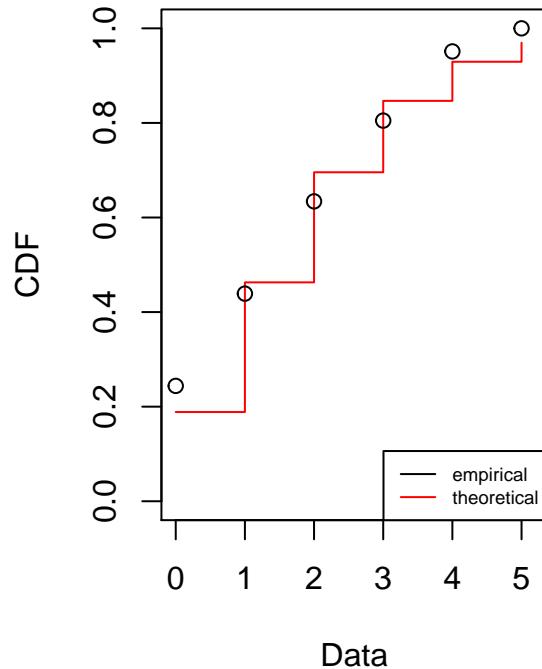
Histogram of X.num[, k]



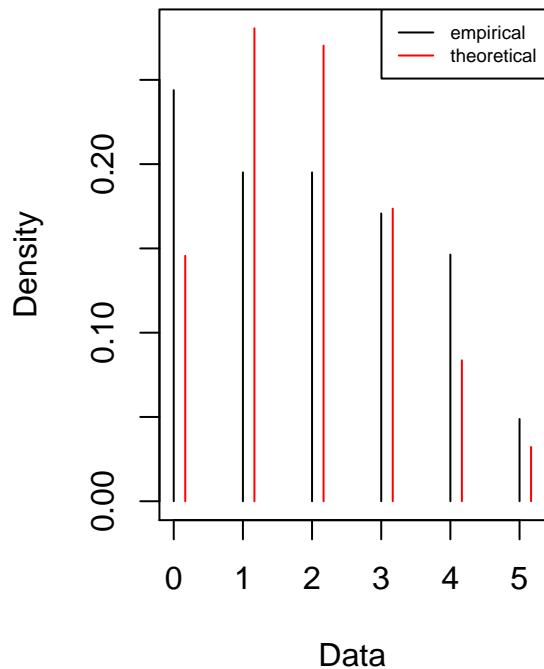
Emp. and theo. distr.



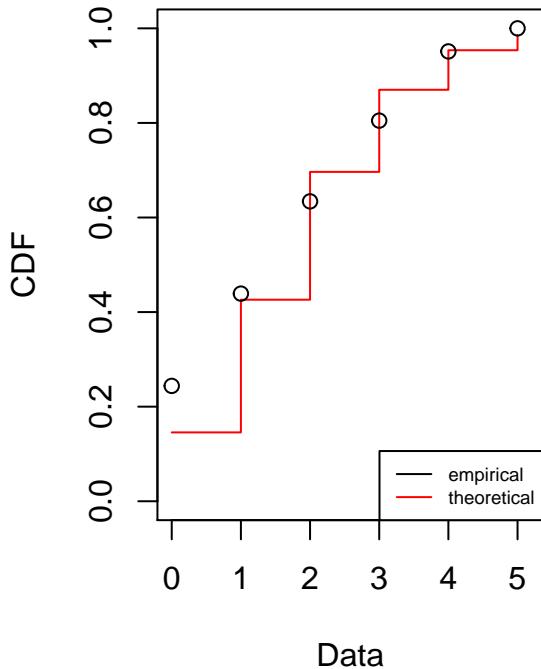
Emp. and theo. CDFs



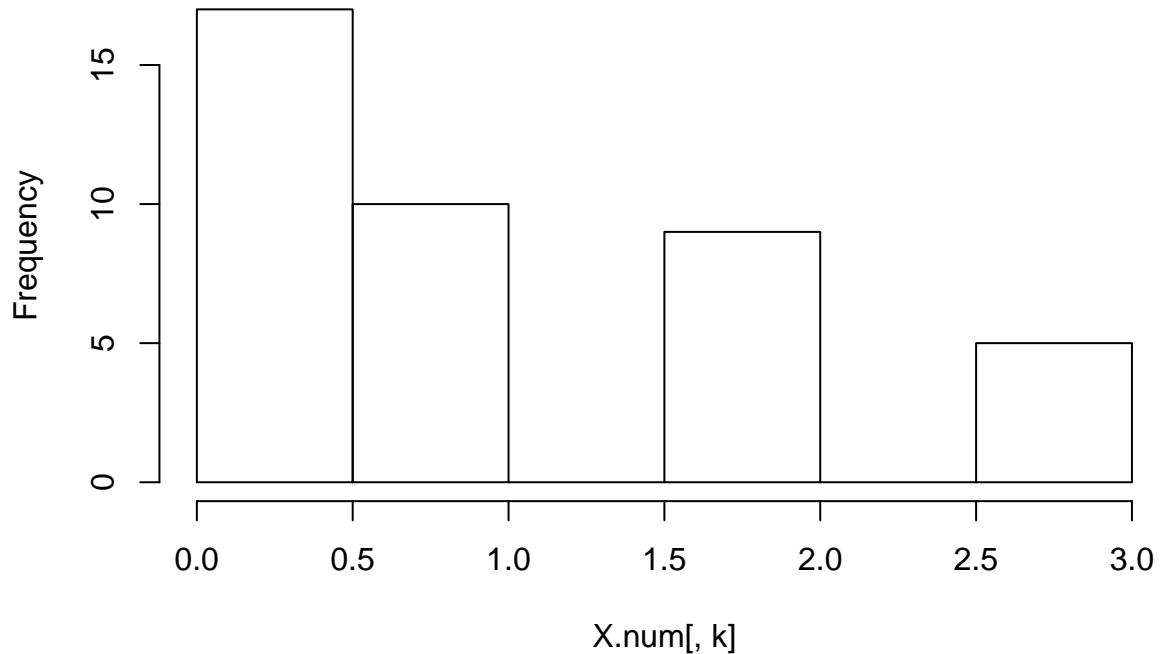
Emp. and theo. distr.



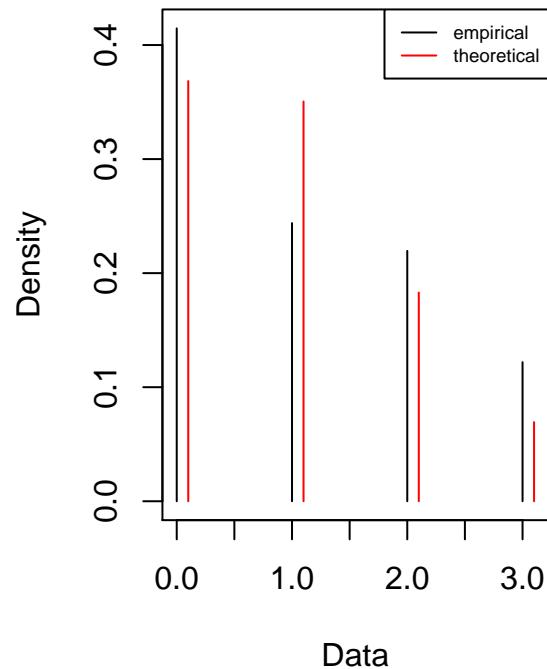
Emp. and theo. CDFs



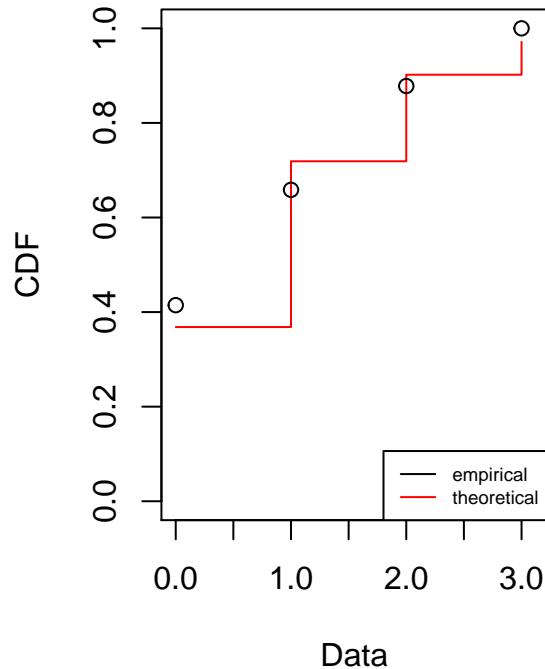
Histogram of X.num[, k]



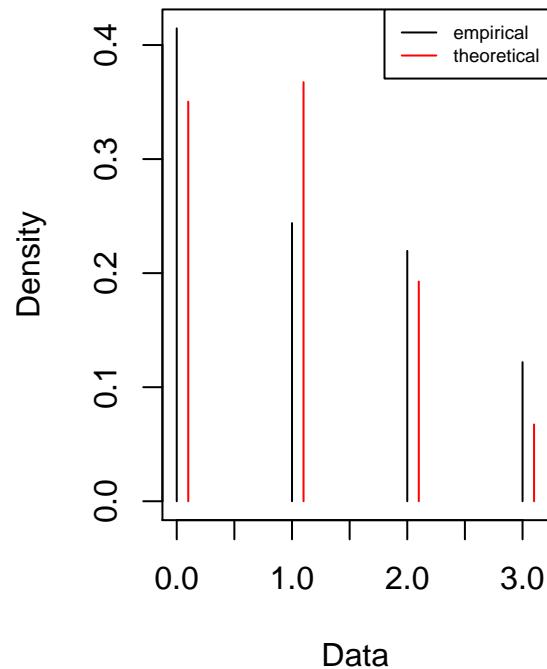
Emp. and theo. distr.



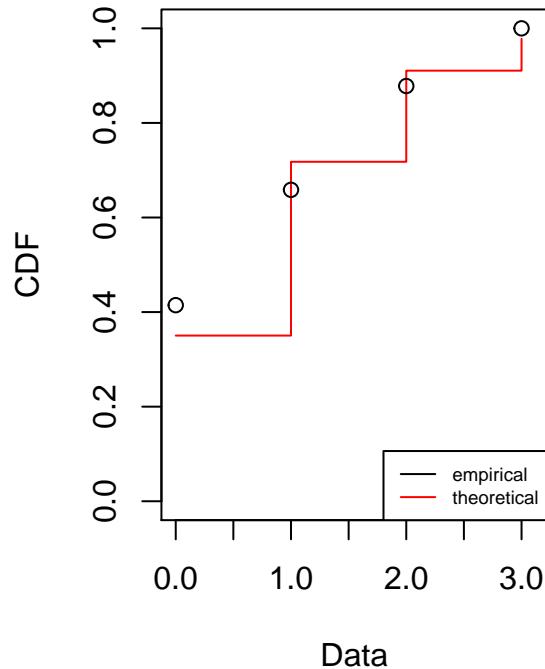
Emp. and theo. CDFs



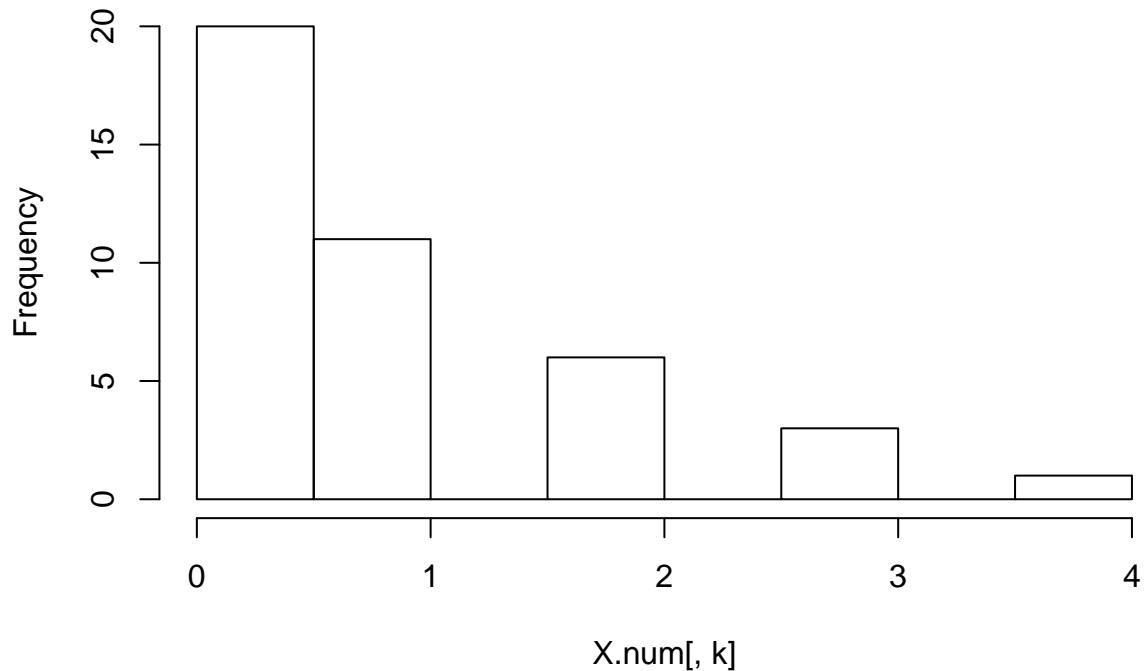
Emp. and theo. distr.



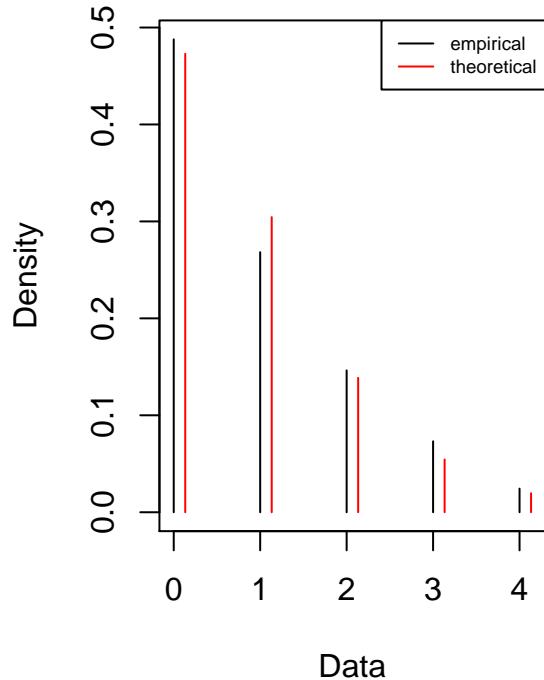
Emp. and theo. CDFs



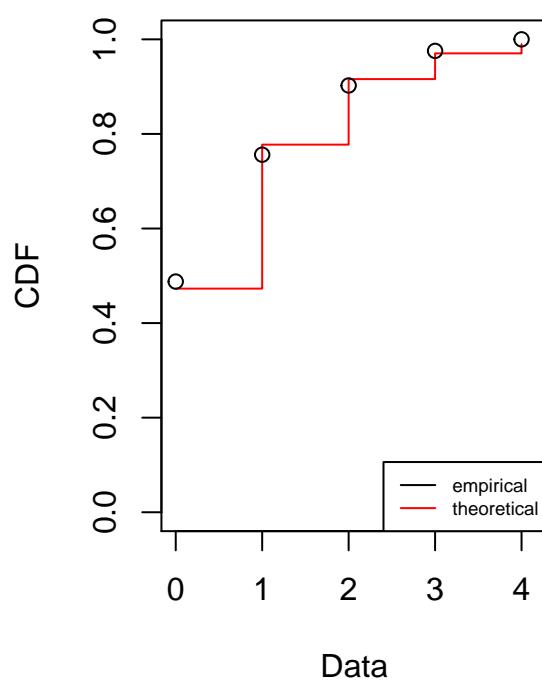
Histogram of X.num[, k]



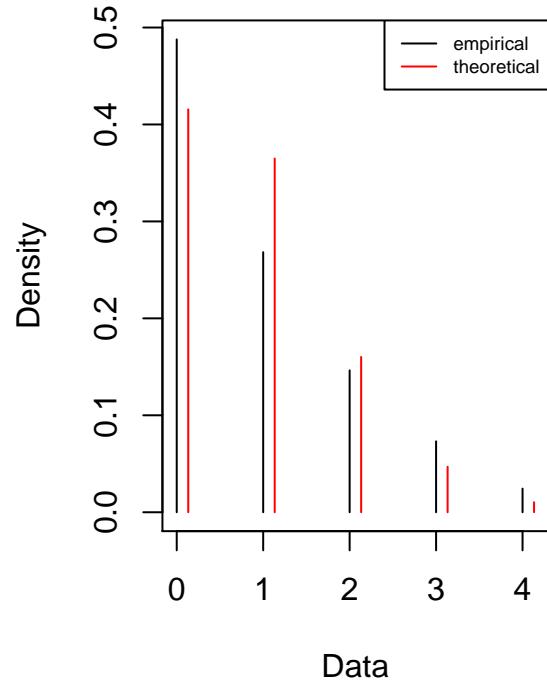
Emp. and theo. distr.



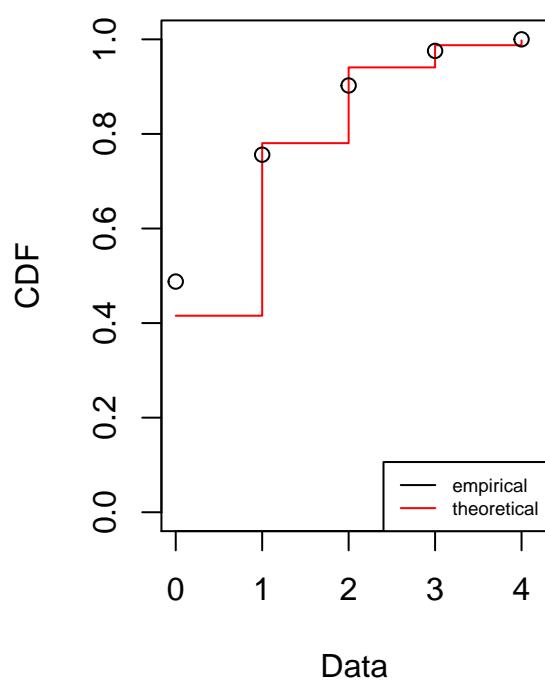
Emp. and theo. CDFs



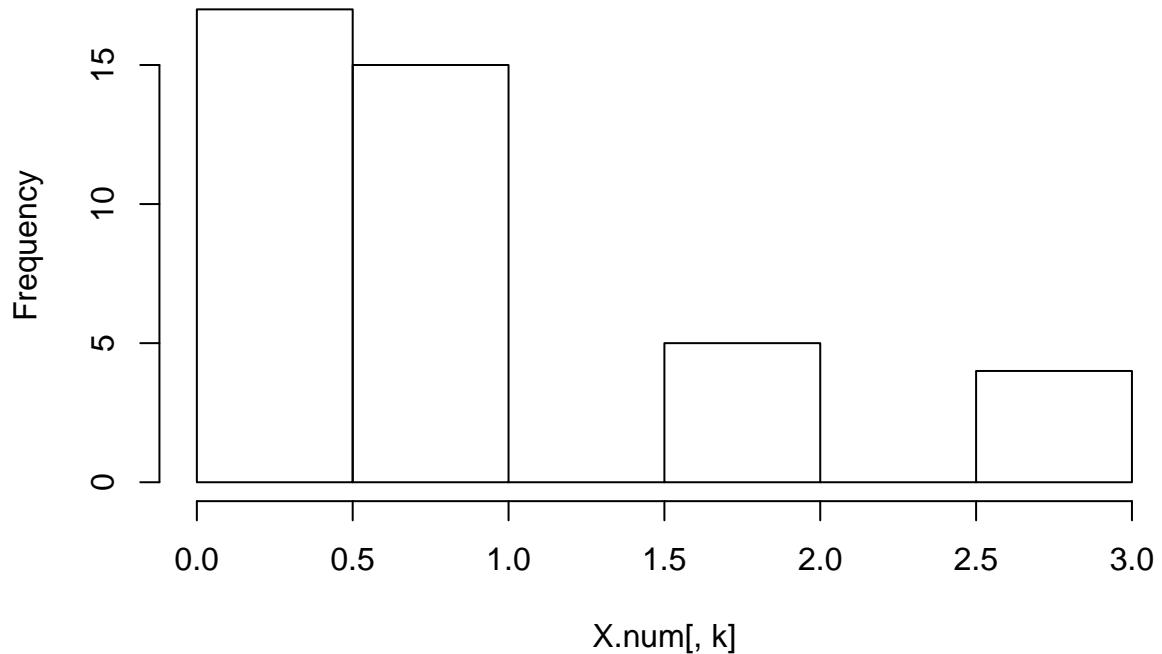
Emp. and theo. distr.



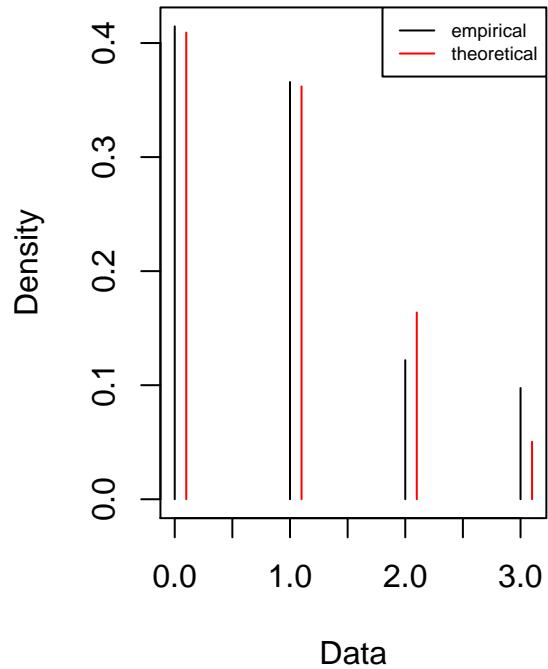
Emp. and theo. CDFs



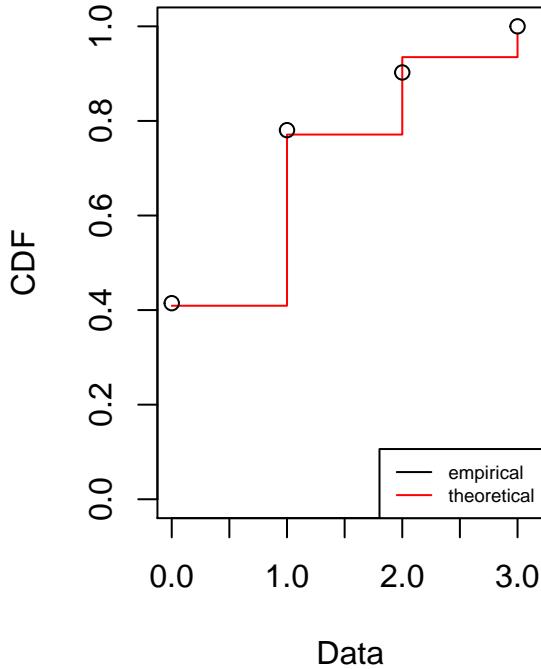
Histogram of X.num[, k]



Emp. and theo. distr.



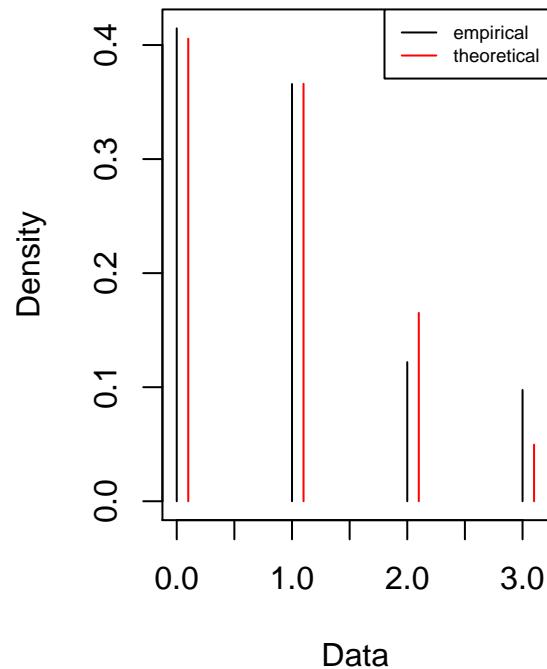
Emp. and theo. CDFs



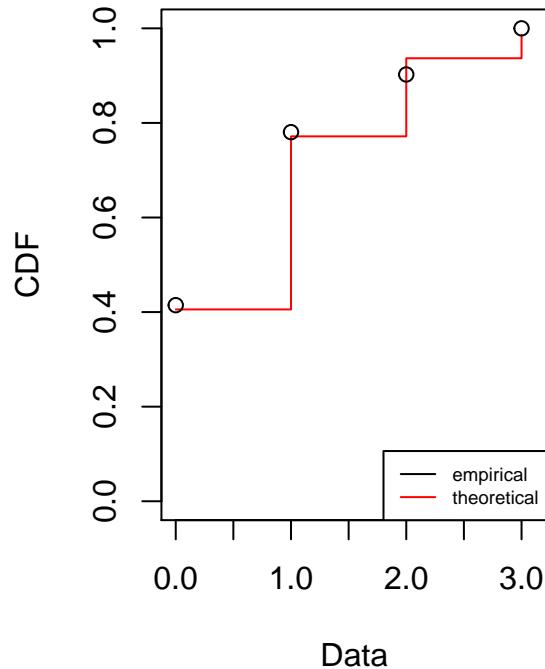
```
## Warning in theta.ml(Y, mu, sum(w), w, limit = control$maxit, trace =
## control$trace > : iteration limit reached

## Warning in theta.ml(Y, mu, sum(w), w, limit = control$maxit, trace =
## control$trace > : iteration limit reached
```

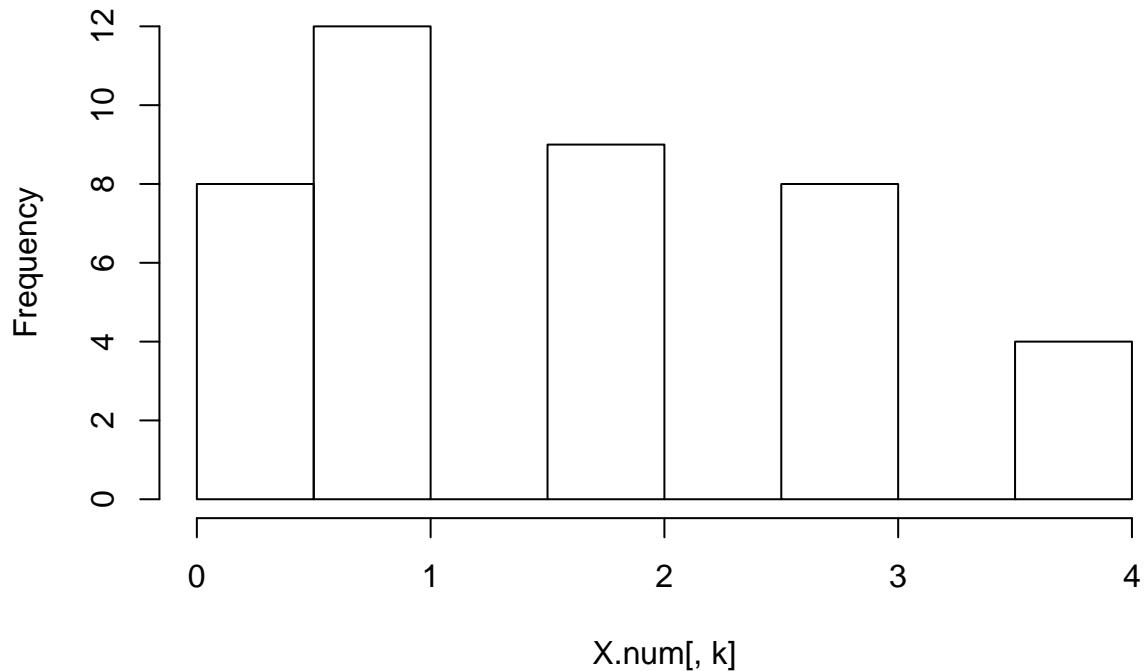
Emp. and theo. distr.



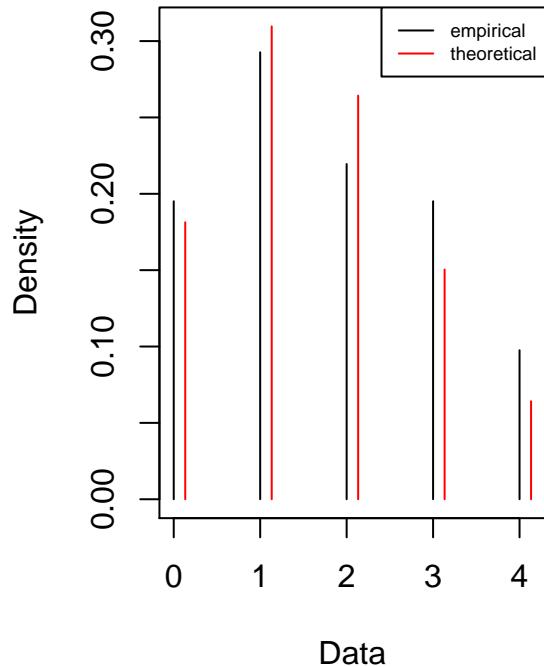
Emp. and theo. CDFs



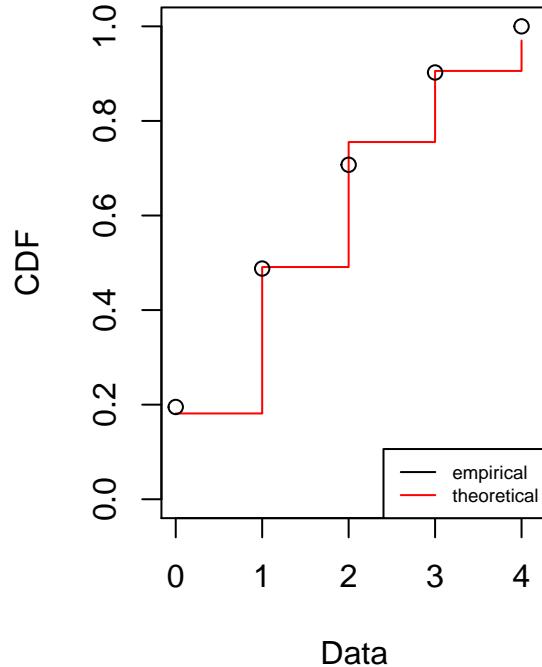
Histogram of X.num[, k]



Emp. and theo. distr.



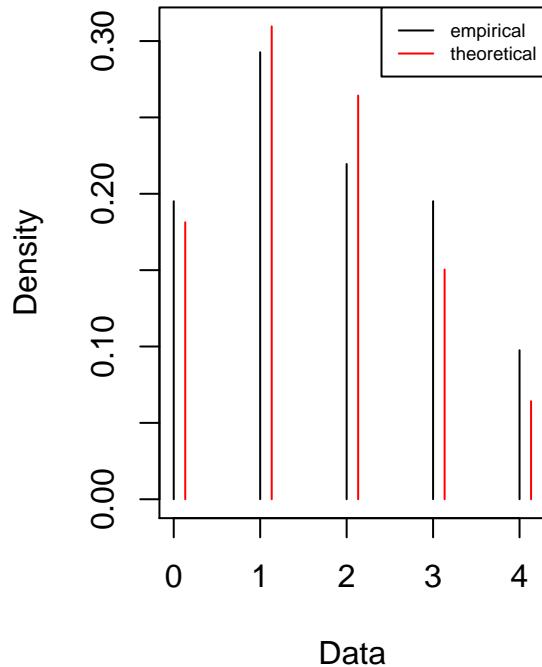
Emp. and theo. CDFs



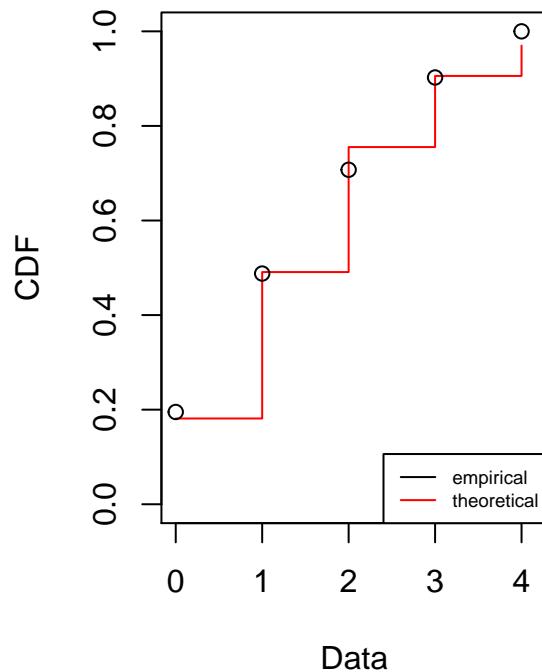
```
## Warning in theta.ml(Y, mu, sum(w), w, limit = control$maxit, trace =
## control$trace > : iteration limit reached

## Warning in theta.ml(Y, mu, sum(w), w, limit = control$maxit, trace =
## control$trace > : iteration limit reached
```

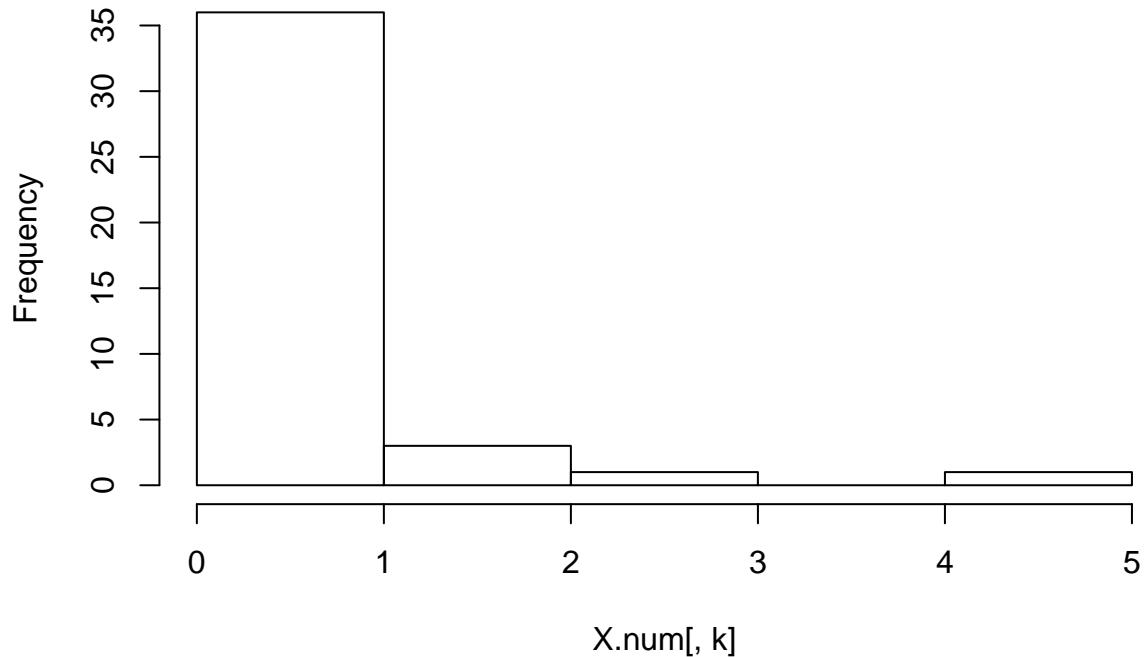
Emp. and theo. distr.



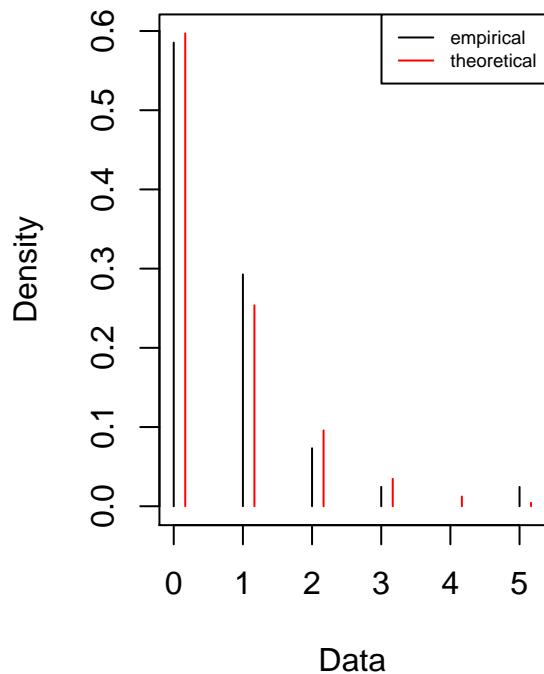
Emp. and theo. CDFs



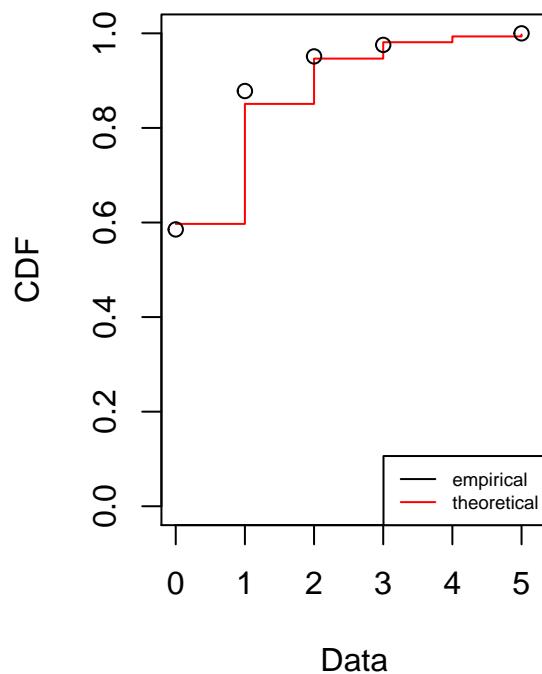
Histogram of X.num[, k]



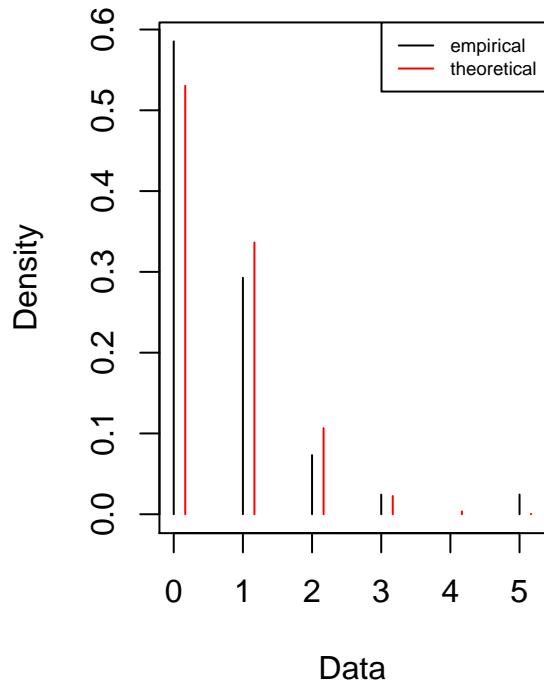
Emp. and theo. distr.



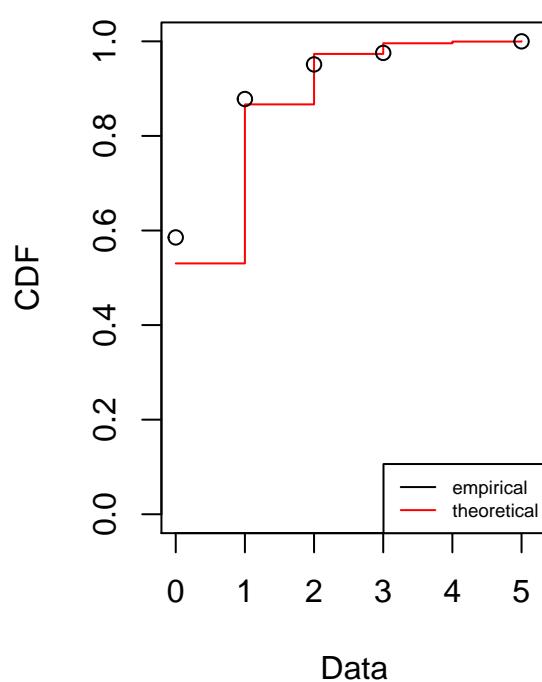
Emp. and theo. CDFs



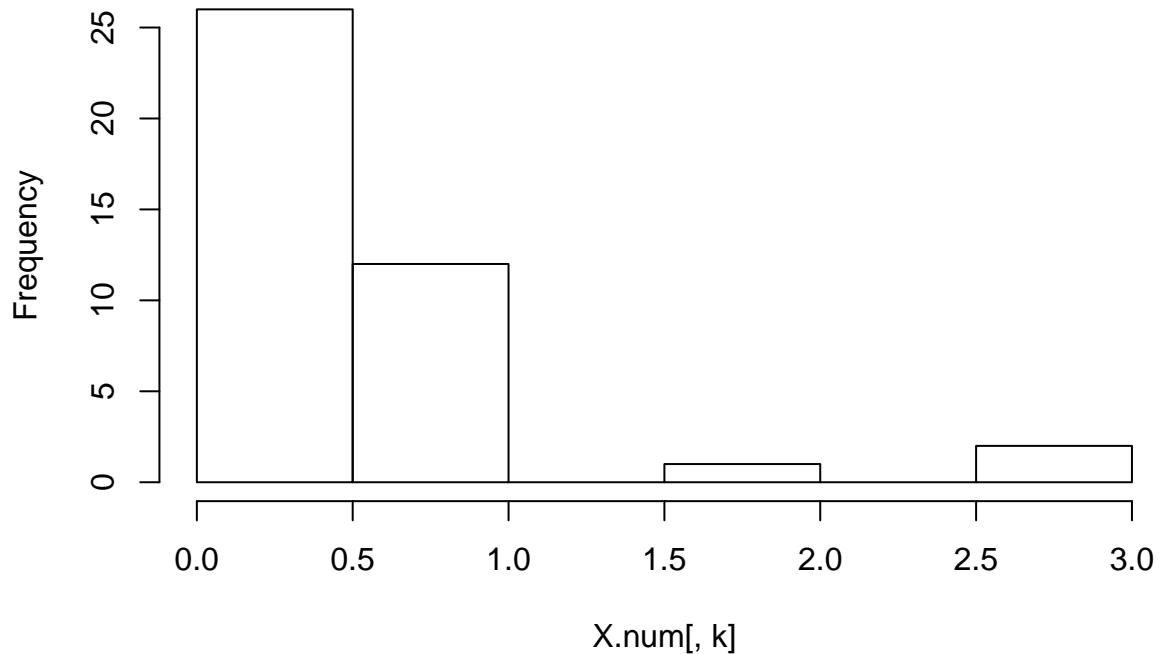
Emp. and theo. distr.



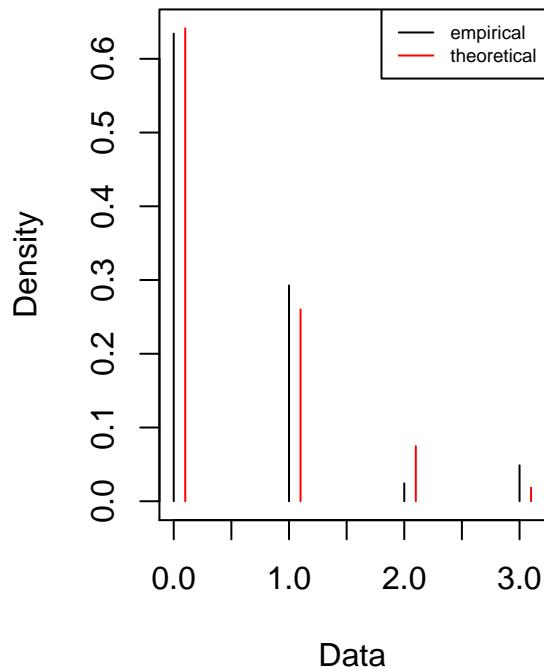
Emp. and theo. CDFs



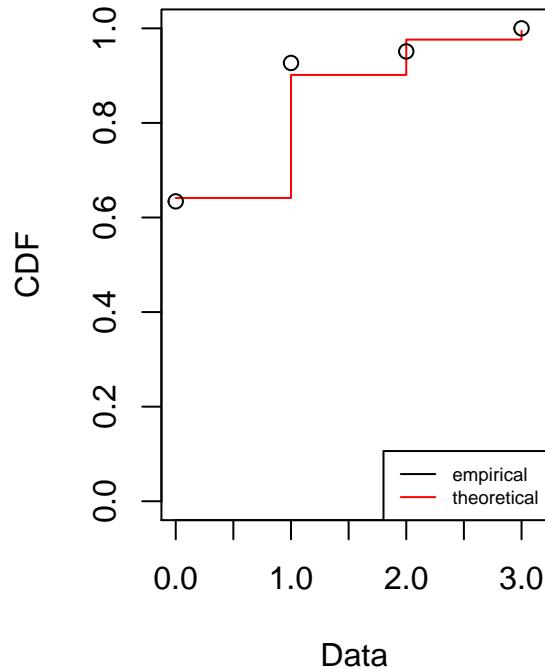
Histogram of X.num[, k]



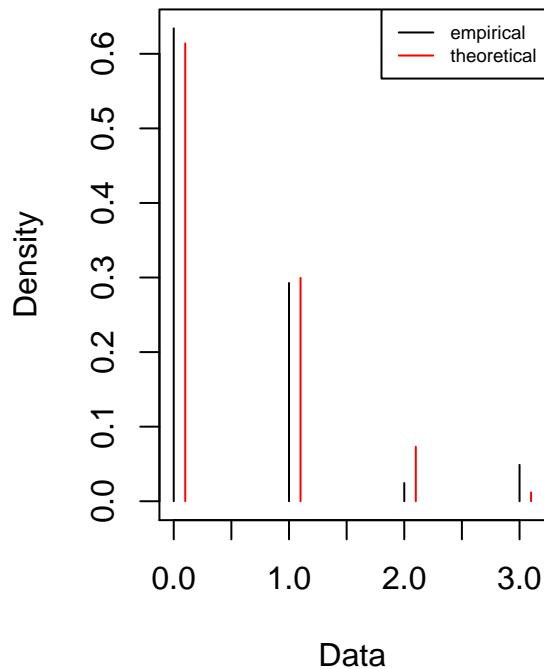
Emp. and theo. distr.



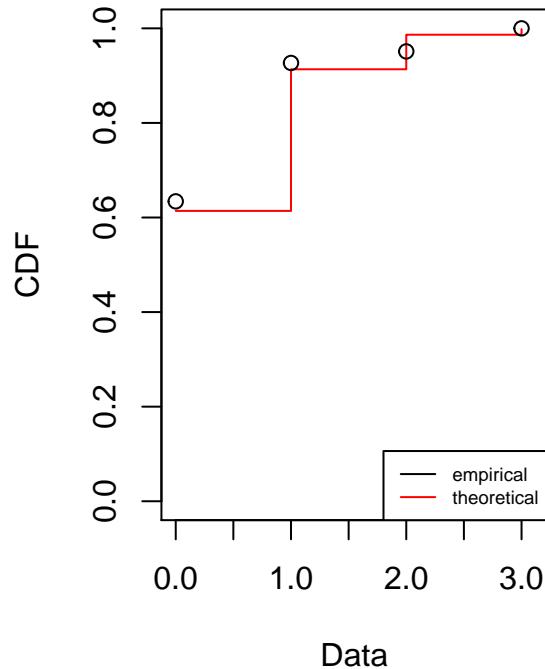
Emp. and theo. CDFs



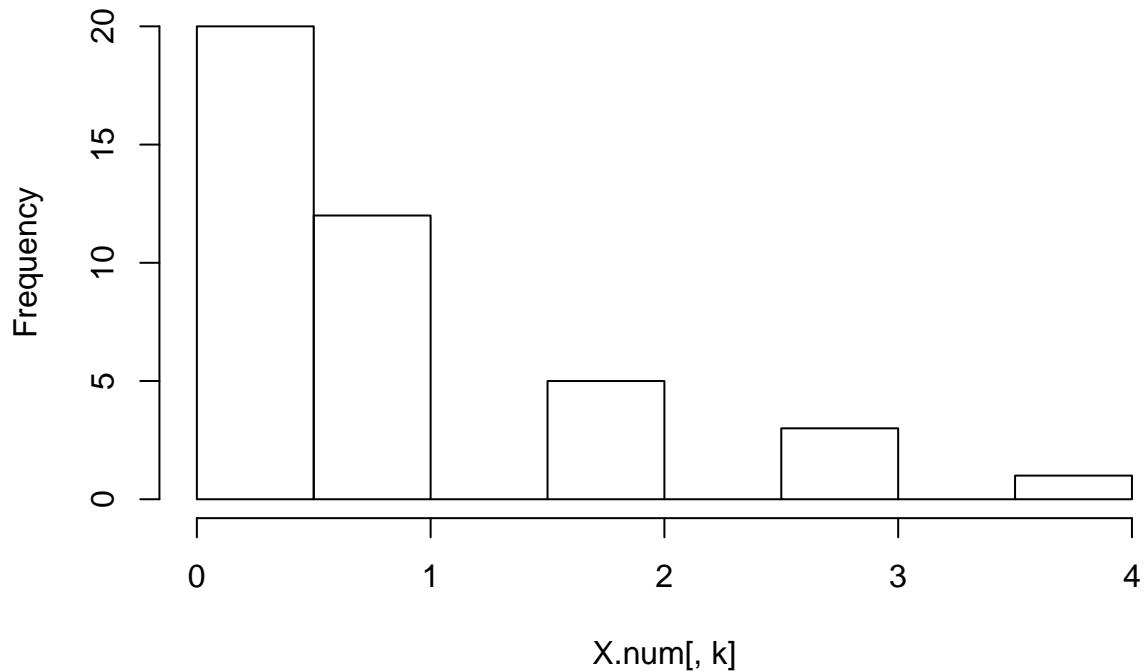
Emp. and theo. distr.



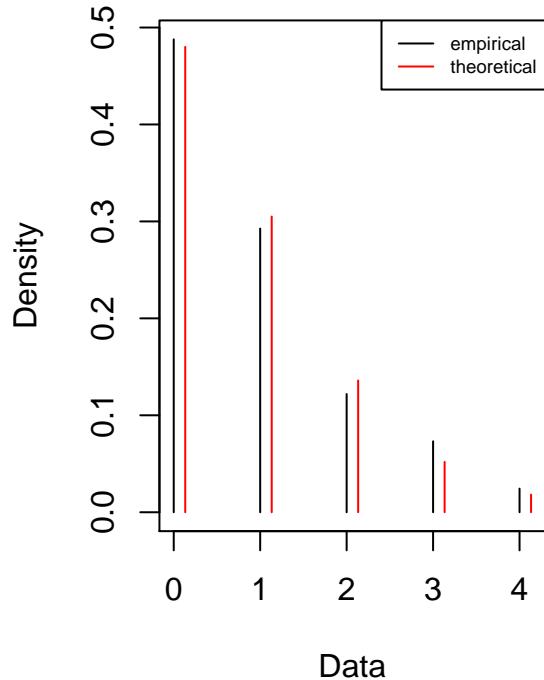
Emp. and theo. CDFs



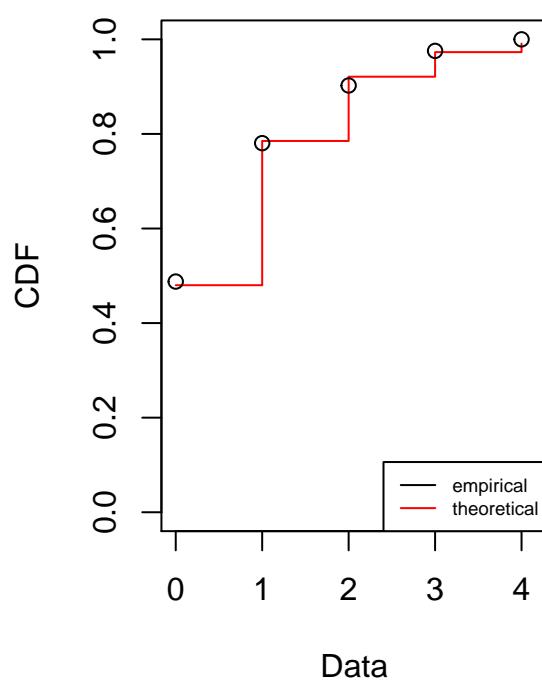
Histogram of X.num[, k]

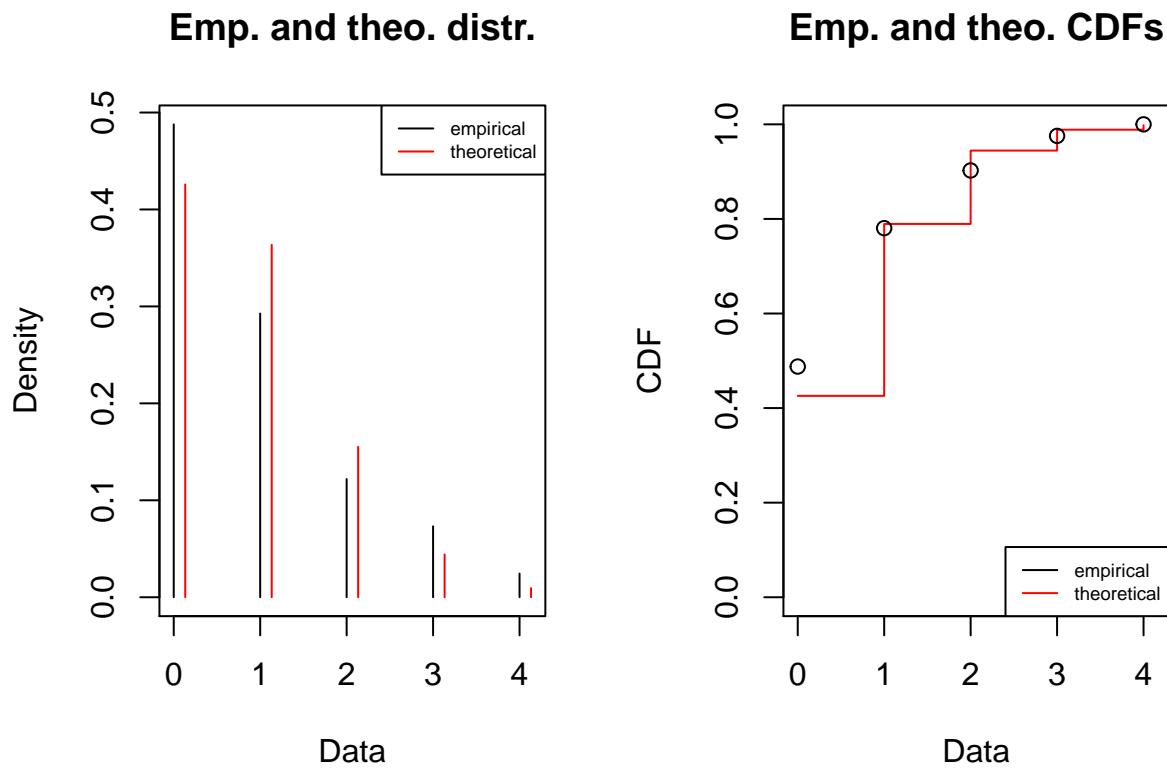


Emp. and theo. distr.



Emp. and theo. CDFs





```
colnames(mle.nb.glm.params) <- c(colnames(snb$coefficients), colnames(snb$coefficients))

colnames(mle.pois.glm.params) <- c(colnames(sp$coefficients), colnames(sp$coefficients))
library(pander)

pander(mle.nb.glm.params)
```

Table 1: Table continues below

Estimate	Std. Error	t value	Pr(> t)	Estimate	Std. Error	t value
-0.01112	0.3868	-0.02876	0.9772	-0.01508	0.01725	-0.8741
0.01262	0.2792	0.04522	0.9642	0.02806	0.01028	2.73
-0.1967	0.3437	-0.5724	0.5703	0.01123	0.01356	0.828
0.3951	0.3504	1.127	0.2665	-0.02732	0.01636	-1.67
-1.325	0.4159	-3.187	0.002834	0.05012	0.01406	3.564
0.2638	0.2475	1.066	0.2931	0.0124	0.009672	1.282
0.02877	0.436	0.06599	0.9477	-0.02511	0.01991	-1.261
-1.034	0.5317	-1.944	0.05909	0.01436	0.02081	0.69
-0.5377	0.429	-1.253	0.2176	0.01715	0.01672	1.026

Pr(> t)
0.3874
0.009462
0.4127

$\Pr(> t)$
0.1028
0.0009826
0.2074
0.2148
0.4943
0.3114

```
pander(mle.pois.glm.params)
```

Table 3: Table continues below

Estimate	Std. Error	z value	$\Pr(> z)$	Estimate	Std. Error	z value
-0.02045	0.3472	-0.05891	0.953	-0.01461	0.01557	-0.9385
0.01857	0.2683	0.0692	0.9448	0.0278	0.009817	2.832
-0.1918	0.3293	-0.5825	0.5602	0.011	0.01295	0.8492
0.3631	0.3029	1.199	0.2306	-0.02566	0.01447	-1.773
-1.325	0.4539	-2.919	0.003506	0.05012	0.01535	3.266
0.2638	0.2602	1.014	0.3106	0.0124	0.01017	1.22
0.01881	0.3579	0.05255	0.9581	-0.02458	0.017	-1.446
-1.034	0.4922	-2.101	0.03565	0.01437	0.01906	0.7538
-0.5177	0.376	-1.377	0.1686	0.01624	0.01444	1.124

$\Pr(> z)$
0.348
0.004629
0.3958
0.07627
0.001092
0.2226
0.1481
0.451
0.2608

Fit JAGS GLM Models for Poisson

```
library(rjags)

## Loading required package: coda
## Linked to JAGS 4.3.0
## Loaded modules: basemod,bugs

library(coda)
model_code = '
model
{
  ## Likelihood
```

```

for(i in 1:N){
  Y[i] ~ dpois(lambda[i])
  log(lambda[i]) <- mu[i]
  mu[i] <- intercept + beta*t[i]
}

## Priors
beta ~ dnorm(mu.beta,tau.beta)
intercept ~ dnorm(mu.intercept,tau.intercept)
}

for(k in 1:5)
{
  print(paste("JAGS GLM Models for Poisson ", k,sep = " "))
  # Set up the data
  model_data = list(N = 41, t=seq(1:41),Y=X.num[,k],mu.beta=0,tau.beta=.0001,mu.intercept=0,tau.intercept=0)
  # Choose the parameters to watch
  model_parameters = c("beta", "intercept")
  n.chains = 2;nSamples = 10000
  model <- jags.model(textConnection(model_code),data = model_data,n.chains = n.chains)#Compile Model G
  update(model, nSamples, progress.bar="none"); # Burnin
  out.coda <- coda.samples(model, variable.names=model_parameters,n.iter=2*nSamples)
  plot(out.coda)
  #assess the posteriors??? stationarity, by looking at the Heidelberg-Welch convergence diagnostic:
  heidel.diag(out.coda)
  # check that our chain???'s length is satisfactory.
  raftery.diag(out.coda)

  geweke.diag(out.coda)

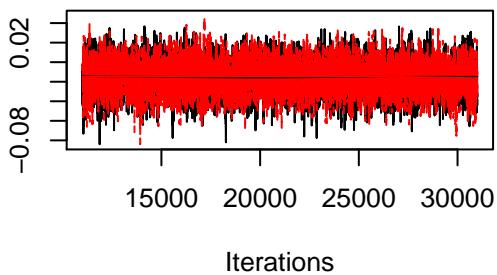
  if(n.chains > 1)
  {
    gelman.srf <-gelman.diag(out.coda)
    plot(gelman.srf$psrf,main = "Gelman Diagnostic")
  }

  chains.ess <- lapply(out.coda,effectiveSize)
  first.chain.ess <- chains.ess[1]
  plot(unlist(first.chain.ess), main="Effective Sample Size")
}

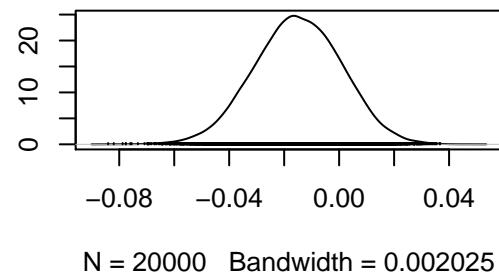
## [1] "JAGS GLM Models for Poisson 1"
## Compiling model graph
##   Resolving undeclared variables
##   Allocating nodes
## Graph information:
##   Observed stochastic nodes: 41
##   Unobserved stochastic nodes: 2
##   Total graph size: 212
##
## Initializing model

```

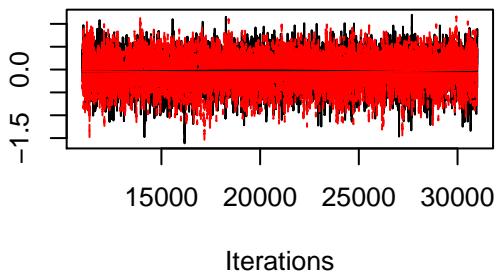
Trace of beta



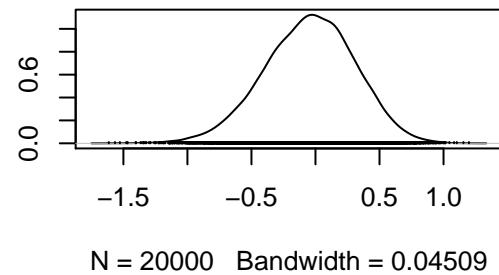
Density of beta



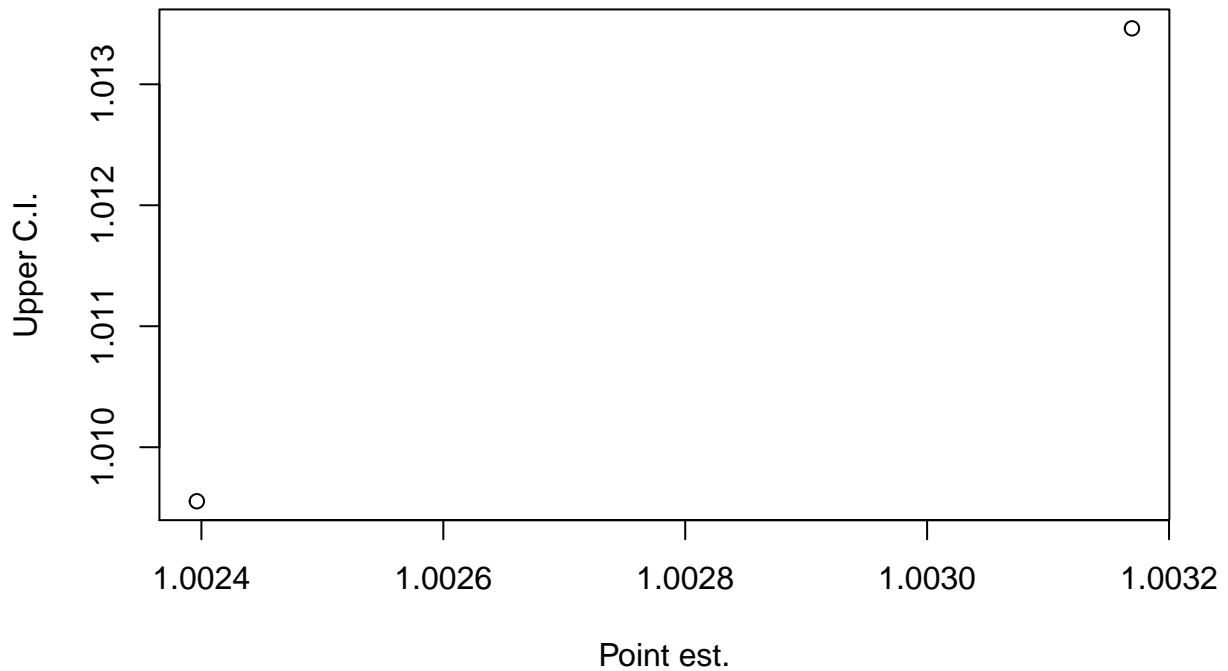
Trace of intercept



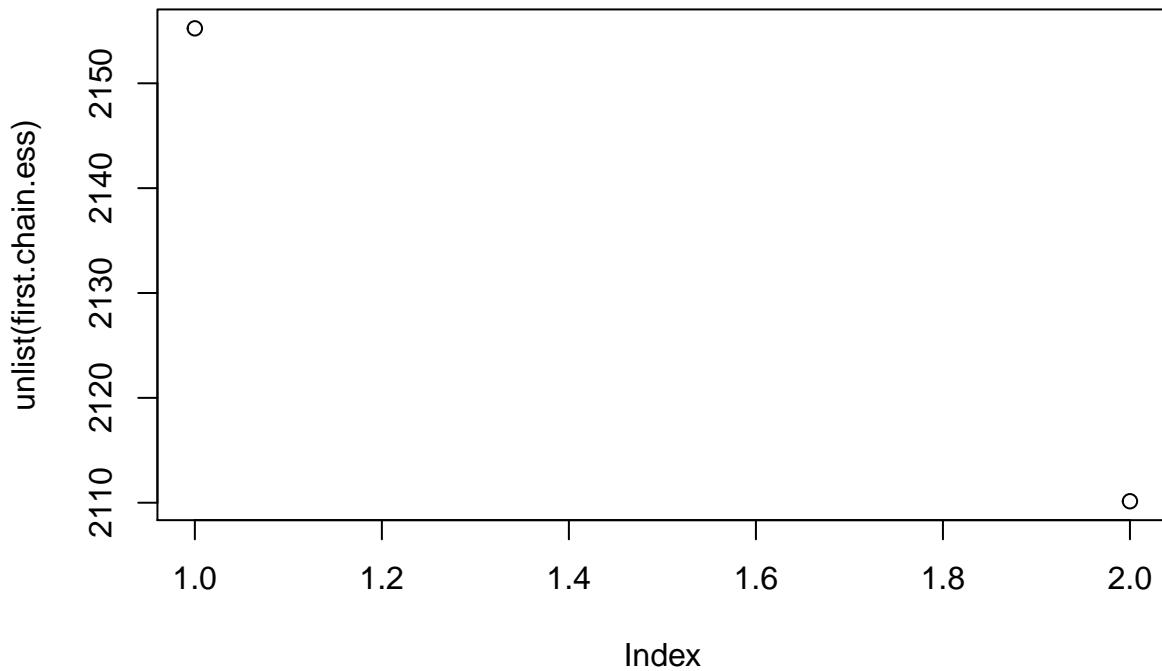
Density of intercept



Gelman Diagnostic

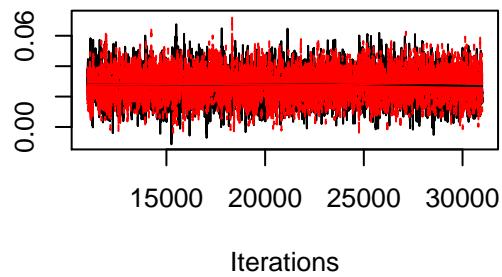


Effective Sample Size

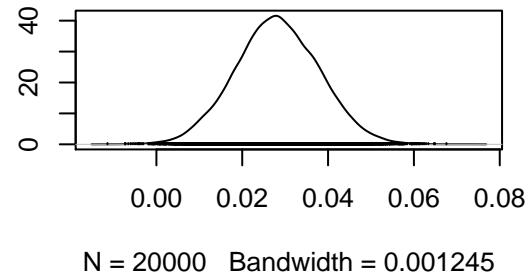


```
## [1] "JAGS GLM Models for Poisson 2"
## Compiling model graph
##    Resolving undeclared variables
##    Allocating nodes
## Graph information:
##    Observed stochastic nodes: 41
##    Unobserved stochastic nodes: 2
##    Total graph size: 212
##
## Initializing model
```

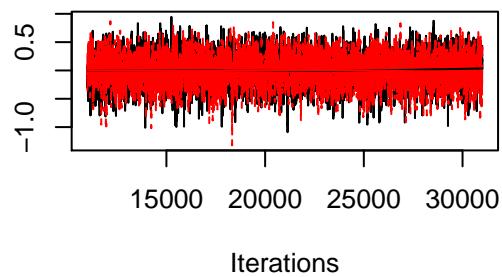
Trace of beta



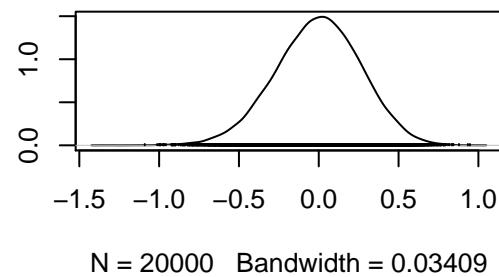
Density of beta



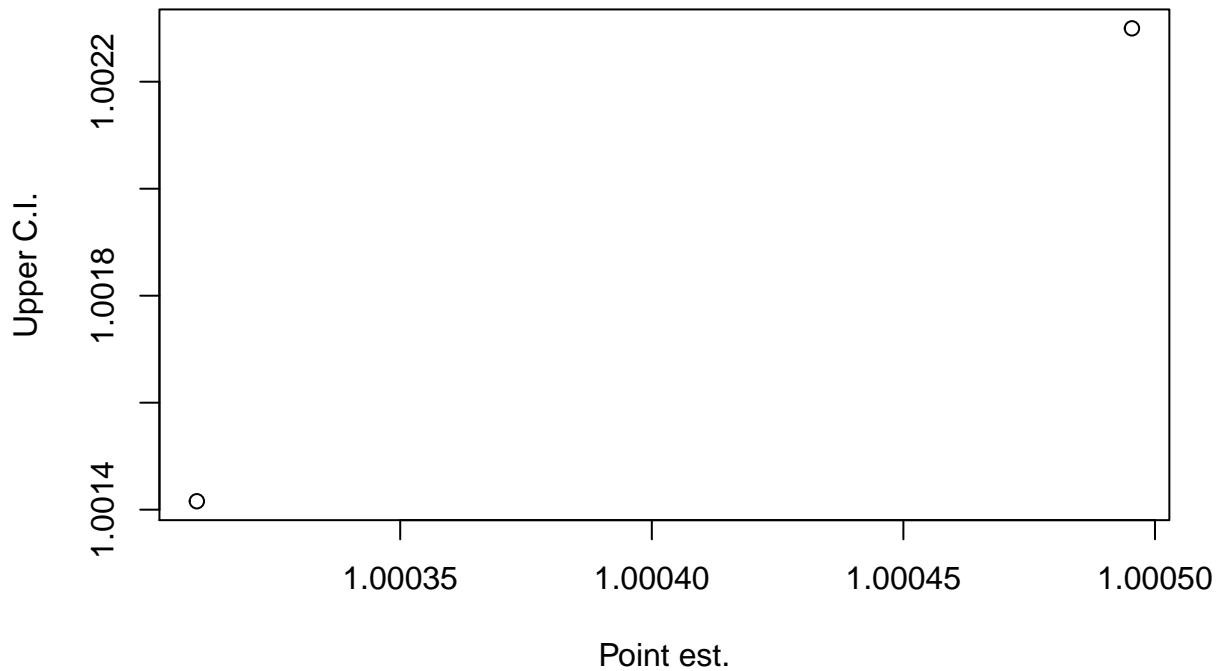
Trace of intercept



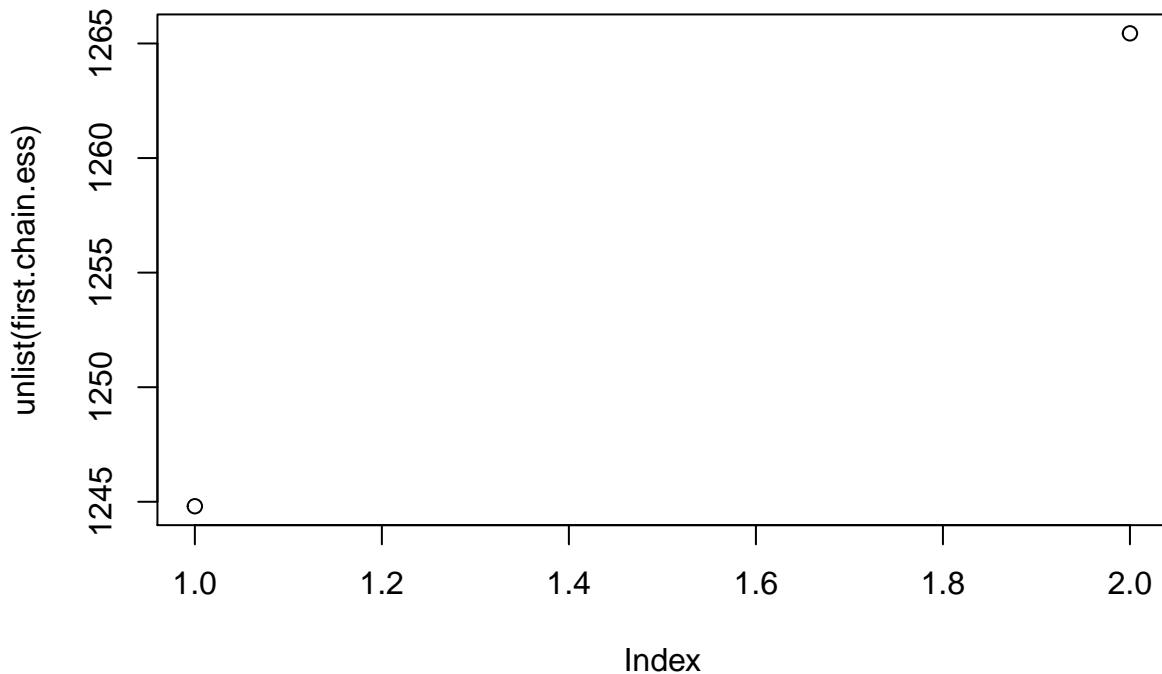
Density of intercept



Gelman Diagnostic

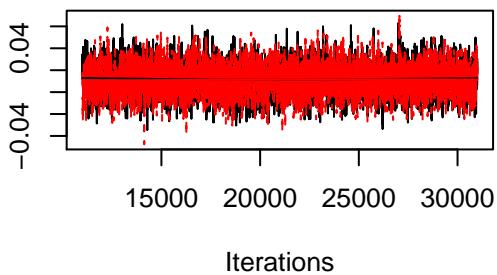


Effective Sample Size

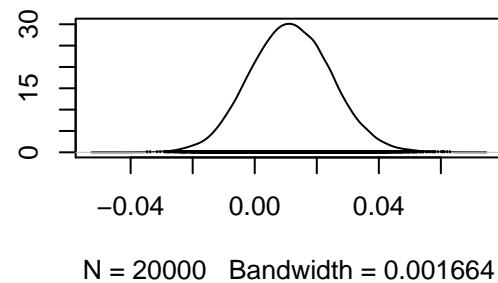


```
## [1] "JAGS GLM Models for Poisson 3"
## Compiling model graph
##    Resolving undeclared variables
##    Allocating nodes
## Graph information:
##    Observed stochastic nodes: 41
##    Unobserved stochastic nodes: 2
##    Total graph size: 212
##
## Initializing model
```

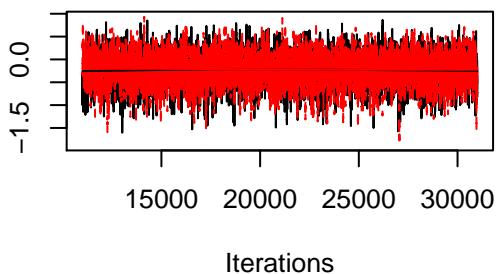
Trace of beta



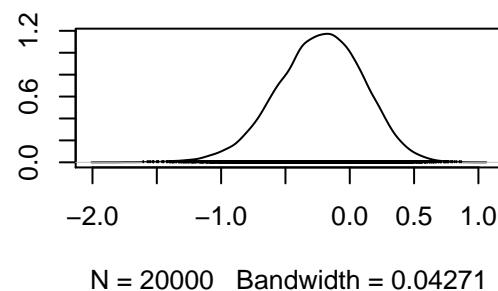
Density of beta



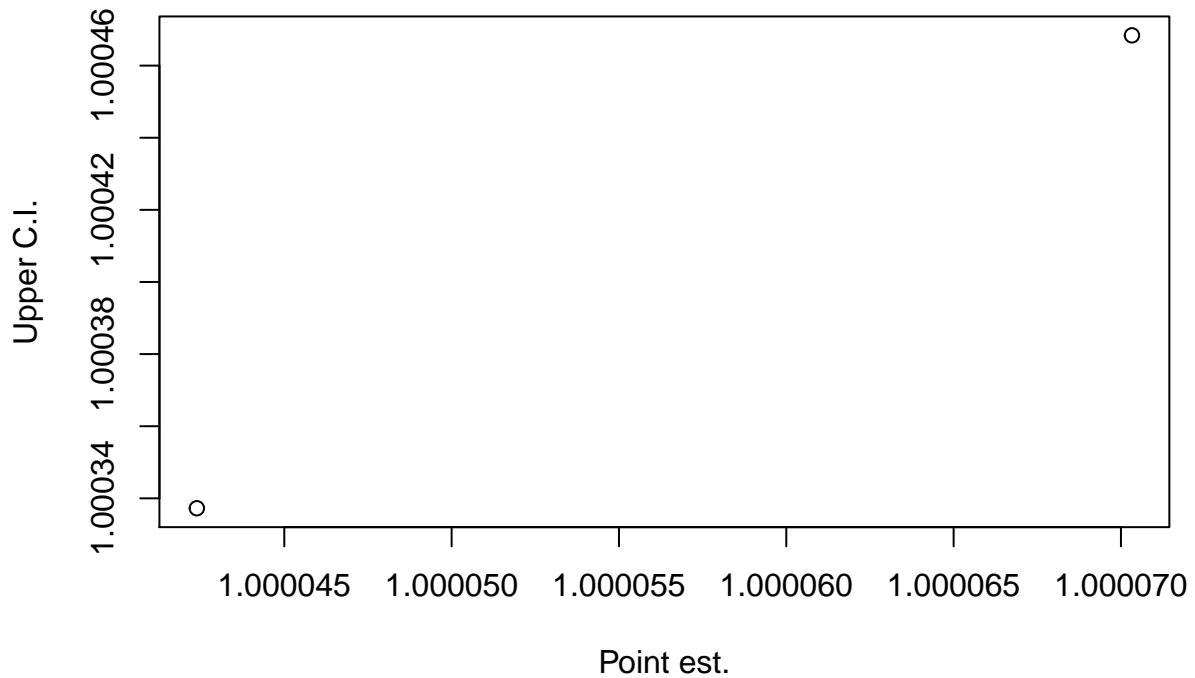
Trace of intercept



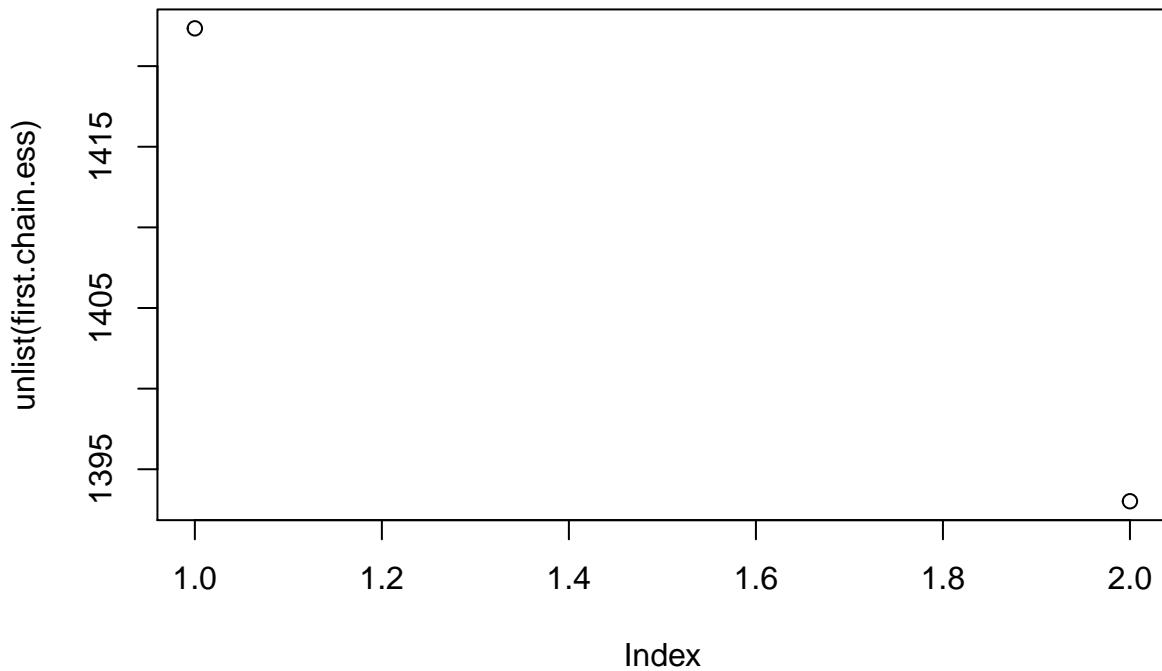
Density of intercept



Gelman Diagnostic

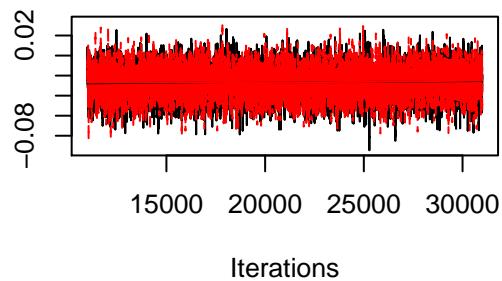


Effective Sample Size

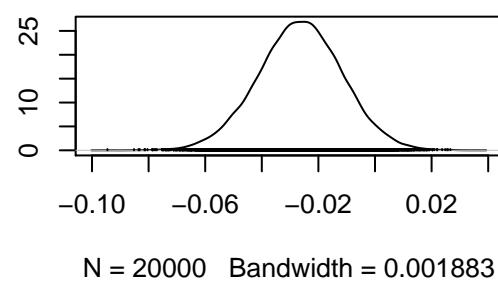


```
## [1] "JAGS GLM Models for Poisson 4"
## Compiling model graph
##    Resolving undeclared variables
##    Allocating nodes
## Graph information:
##    Observed stochastic nodes: 41
##    Unobserved stochastic nodes: 2
##    Total graph size: 212
##
## Initializing model
```

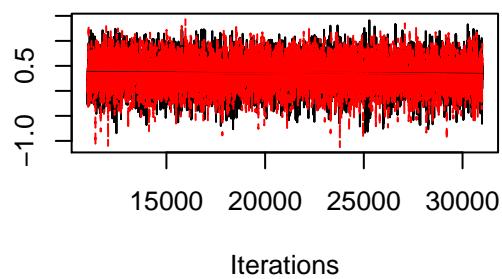
Trace of beta



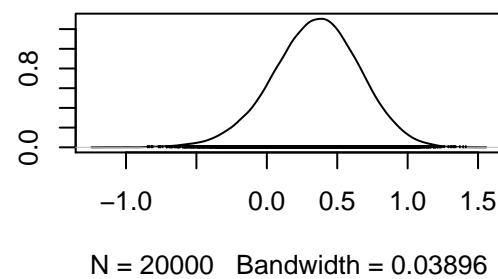
Density of beta



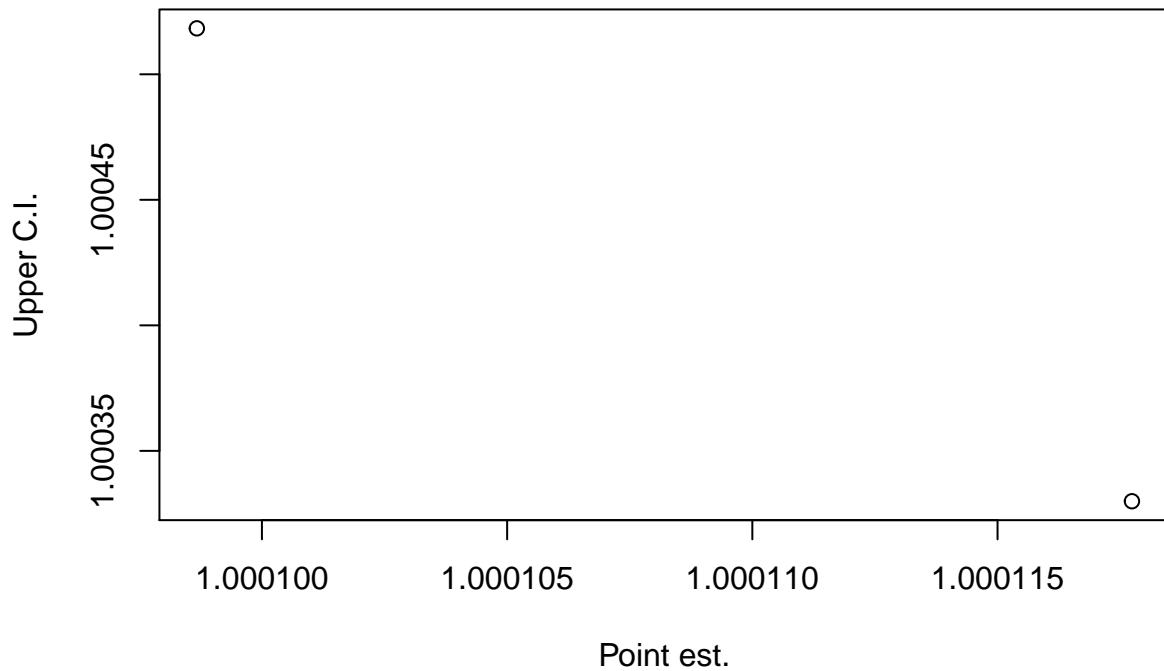
Trace of intercept



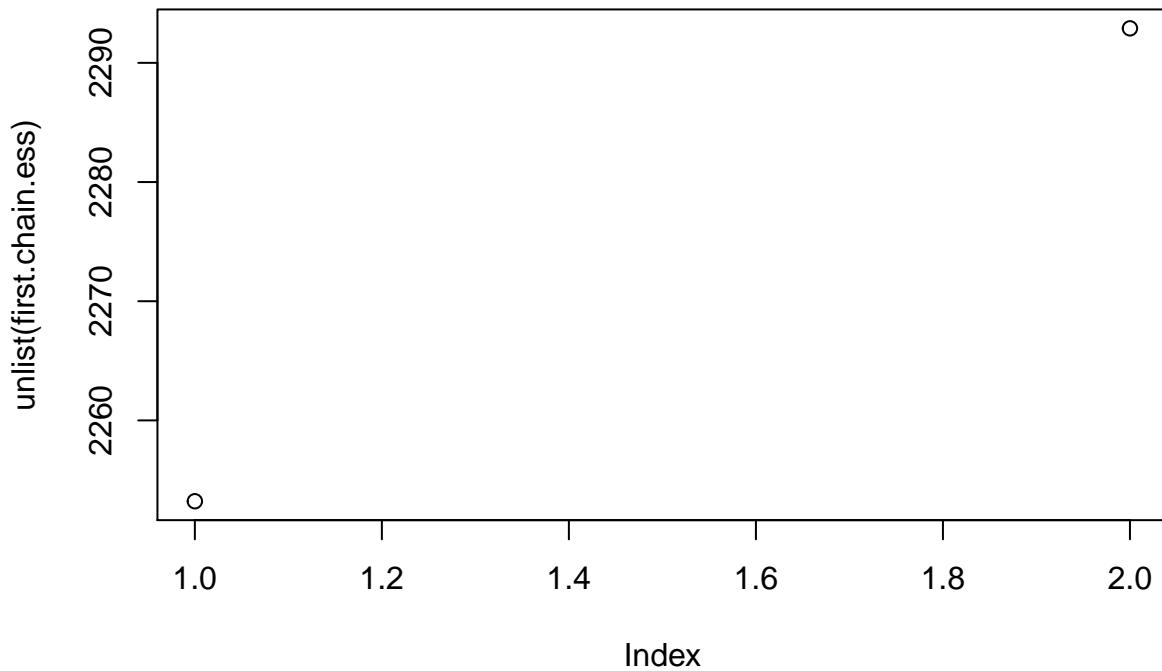
Density of intercept



Gelman Diagnostic

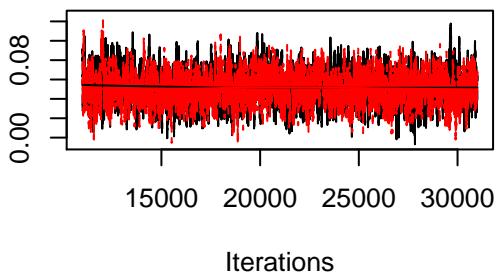


Effective Sample Size

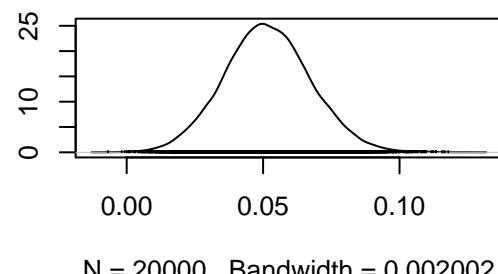


```
## [1] "JAGS GLM Models for Poisson 5"
## Compiling model graph
##    Resolving undeclared variables
##    Allocating nodes
## Graph information:
##    Observed stochastic nodes: 41
##    Unobserved stochastic nodes: 2
##    Total graph size: 212
##
## Initializing model
```

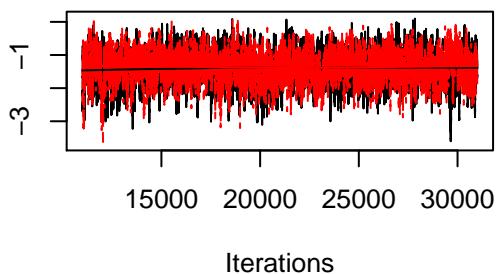
Trace of beta



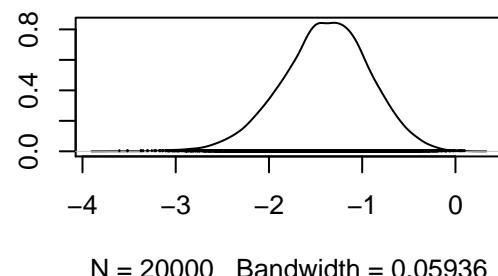
Density of beta



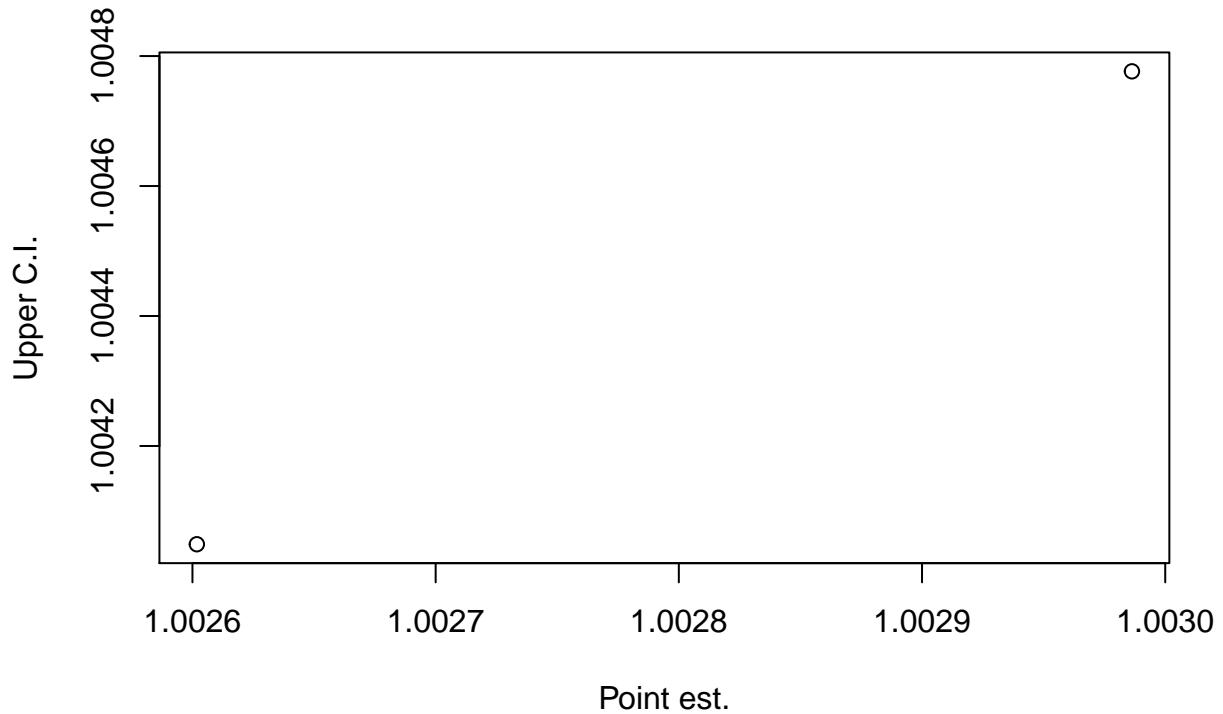
Trace of intercept



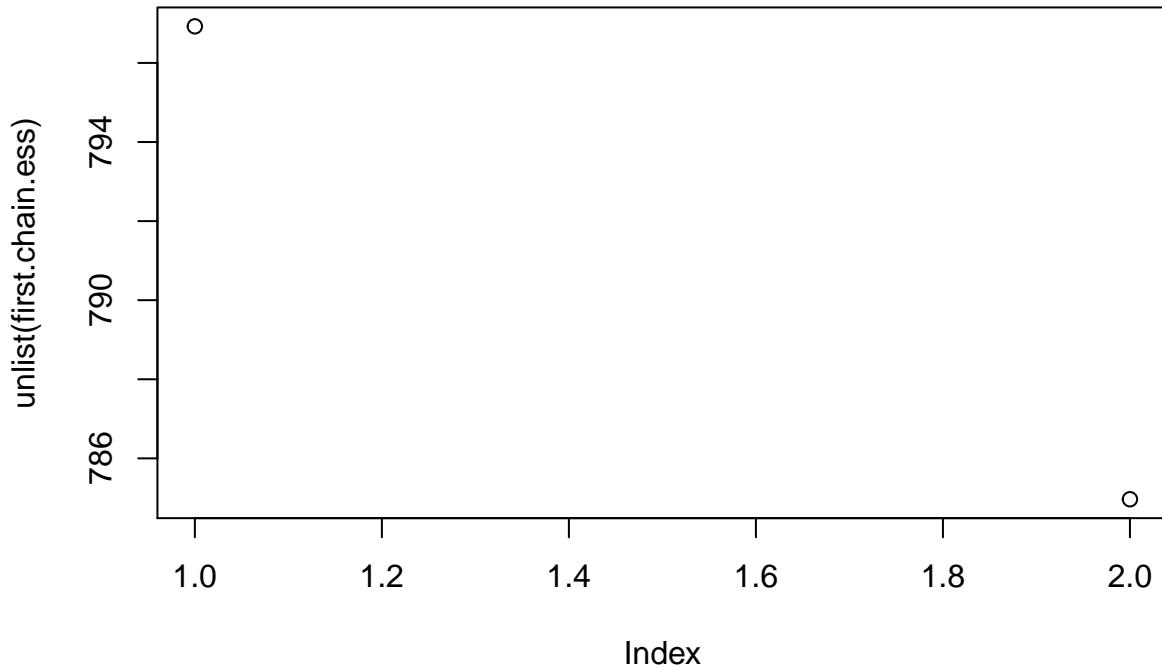
Density of intercept



Gelman Diagnostic



Effective Sample Size



Fit JAGS GLM Models for Negative Binomial

```
#####
##### Fit JAGS GLM Models for Negative Binomial

#
# Direct estimation of the p and r parameters in a negative binomial distribution could involve bad auto
#
# The model specifications:
#
# Parameterized by p and r:
# model {
#   for( i in 1 : N ) {
#     y[i] ~ dnegbin( p , r )
#   }
#   p ~ dbeta(1.001,1.001)
#   r ~ dgamma(0.01,0.01)
#   m <- r*(1-p)/p
#   v <- r*(1-p)/(p*p)
# }
#
# Parameterized by m and r:
# model {
#   for( i in 1 : N ) {
#     y[i] ~ dnegbin( p , r )
#   }
# }
```

```

#      }
#      p <- r/(r+m)
#      )
#      m ~ dgamma(0.01,0.01)
#      v <- r*(1-p)/(p*p)
# }

library(rjags)
library(coda)
model_code = '
model
{
  ## Likelihood
  for(i in 1:N){
    Y[i] ~ dnegbin(p[i],r)

    p[i] <- r/(r+lambda[i])
    log(lambda[i]) <- mu[i]
    mu[i] <- intercept + beta*t[i]
  }

  ## Priors
  beta ~ dnorm(mu.beta,tau.beta)
  intercept ~ dnorm(mu.intercept,tau.intercept)
  r ~ dunif(0,20)
  #r ~ dgamma(0.01,0.01)
}

'
for(k in 1:5)
{
  print(paste("JAGS GLM Models for Negative Binomial ", k,sep = " "))

  # Set up the data
  model_data = list(N = 41, t=seq(1:41),Y=X.num[,k],mu.beta=0,tau.beta=.0001,mu.intercept=0,tau.intercept=0)
  # Choose the parameters to watch
  model_parameters = c("r","beta", "intercept")
  n.chains =2;nSamples = 10000
  model <- jags.model(textConnection(model_code),data = model_data,n.chains = n.chains) #Compile Model G
  update(model, nSamples, progress.bar="none"); # Burnin
  out.coda <- coda.samples(model, variable.names=model_parameters,n.iter=2*nSamples)
  plot(out.coda)

  #assess the posteriors??? stationarity, by looking at the Heidelberg-Welch convergence diagnostic:
  heidel.diag(out.coda)
  # check that our chain???s length is satisfactory.
  raftery.diag(out.coda)

  geweke.diag(out.coda)

  if(n.chains > 1)
  {
    gelman.srf <-gelman.diag(out.coda)
    plot(gelman.srf$psrf,main = "Gelman Diagnostic")
  }

  chains.ess <- lapply(out.coda,effectiveSize)
}

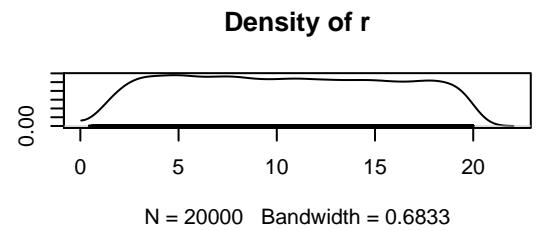
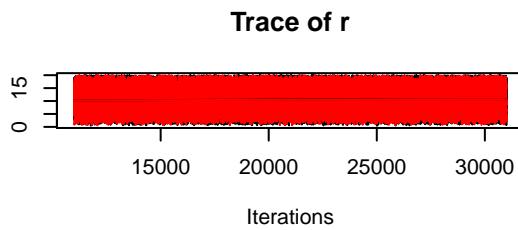
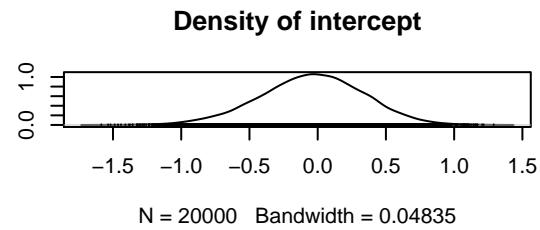
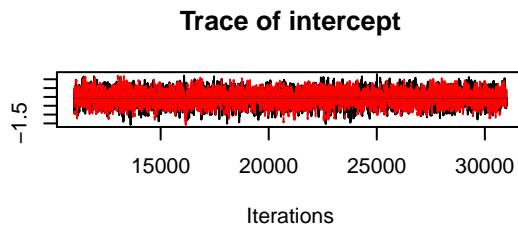
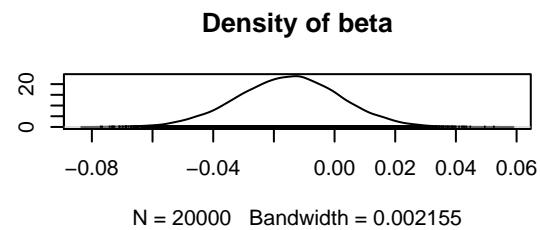
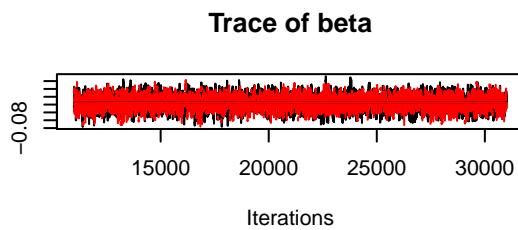
```

```

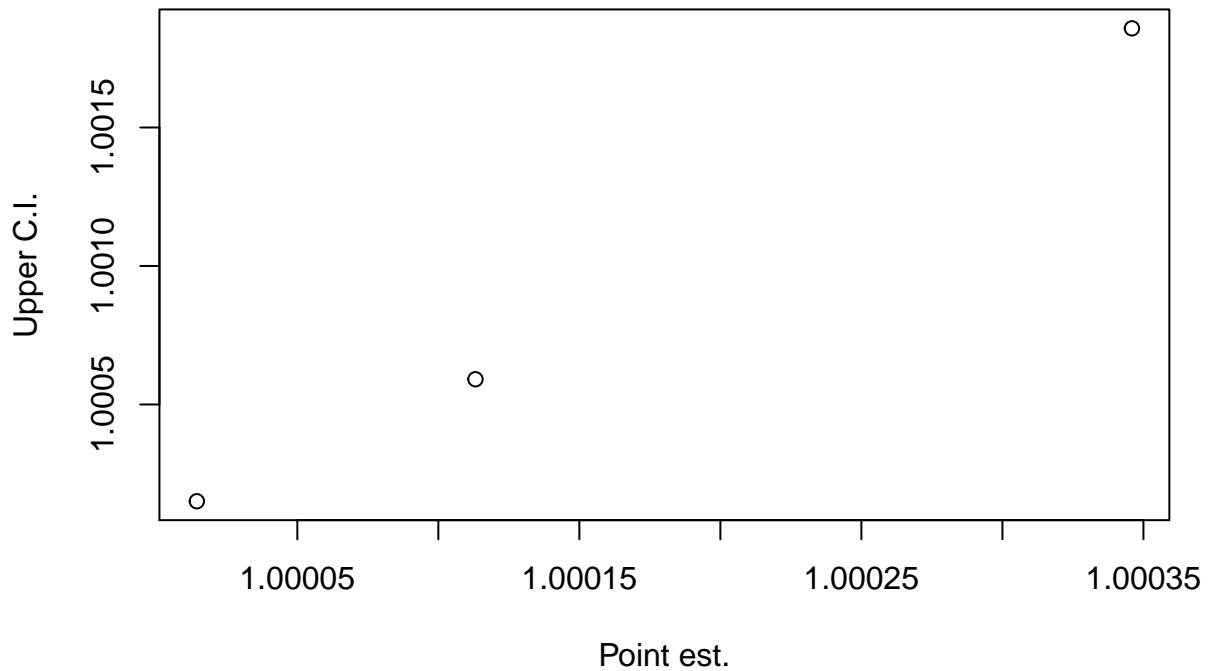
first.chain.ess <- chains.ess[1]
plot(unlist(first.chain.ess), main="Effective Sample Size")
}

## [1] "JAGS GLM Models for Negative Binomial 1"
## Compiling model graph
## Resolving undeclared variables
## Allocating nodes
## Graph information:
##   Observed stochastic nodes: 41
##   Unobserved stochastic nodes: 3
##   Total graph size: 297
##
## Initializing model

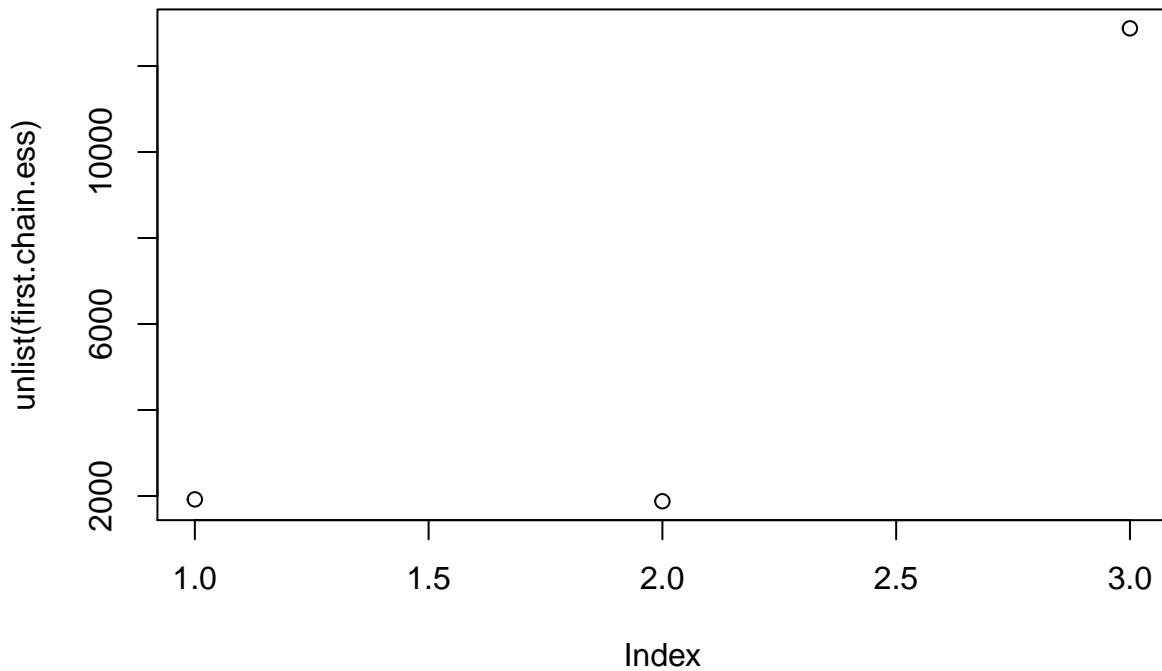
```



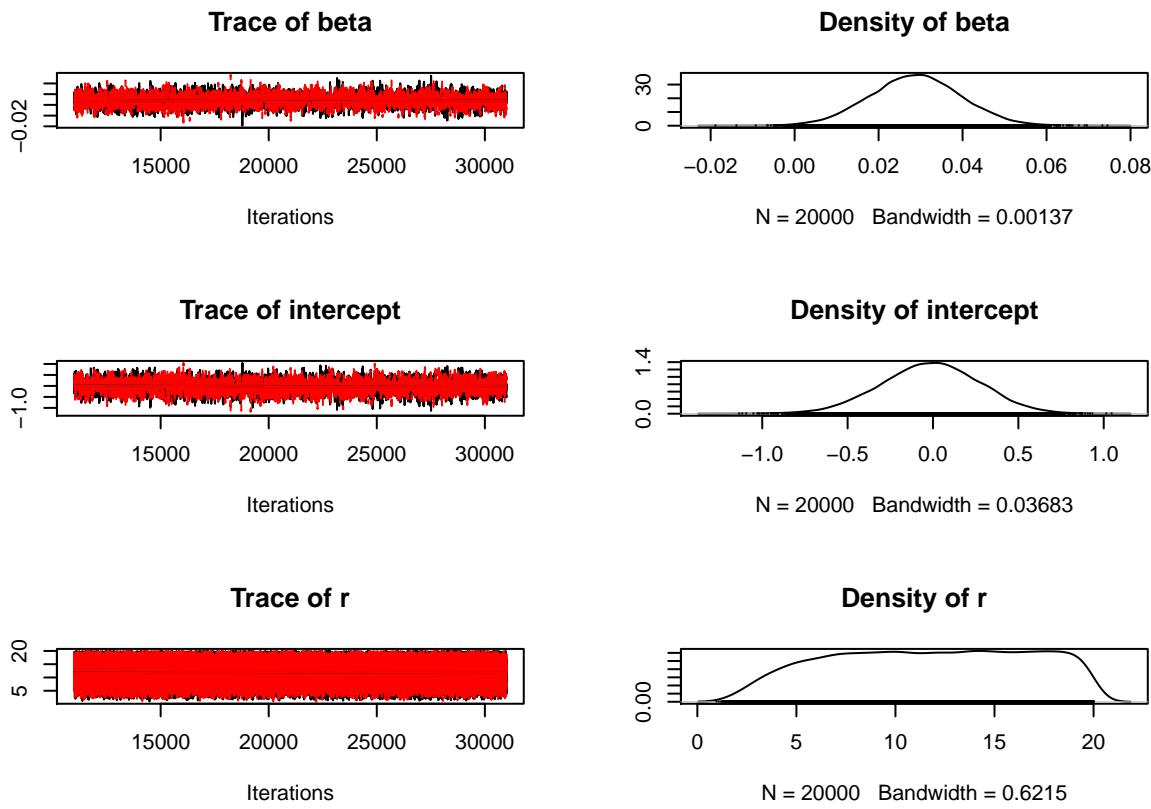
Gelman Diagnostic



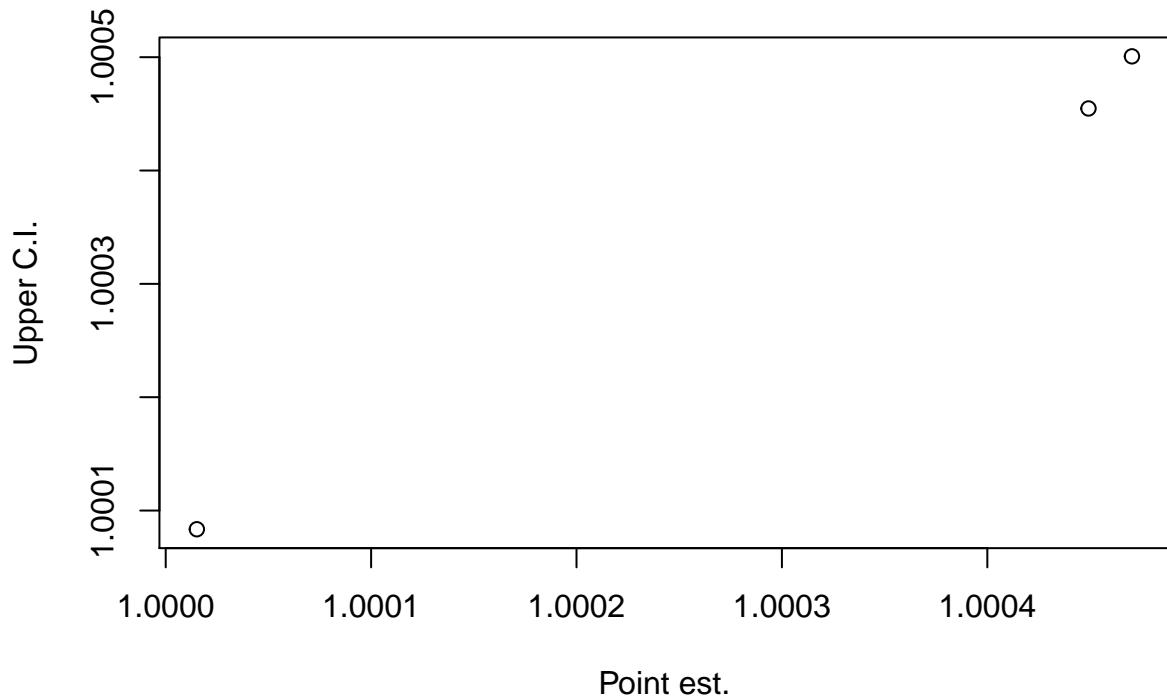
Effective Sample Size



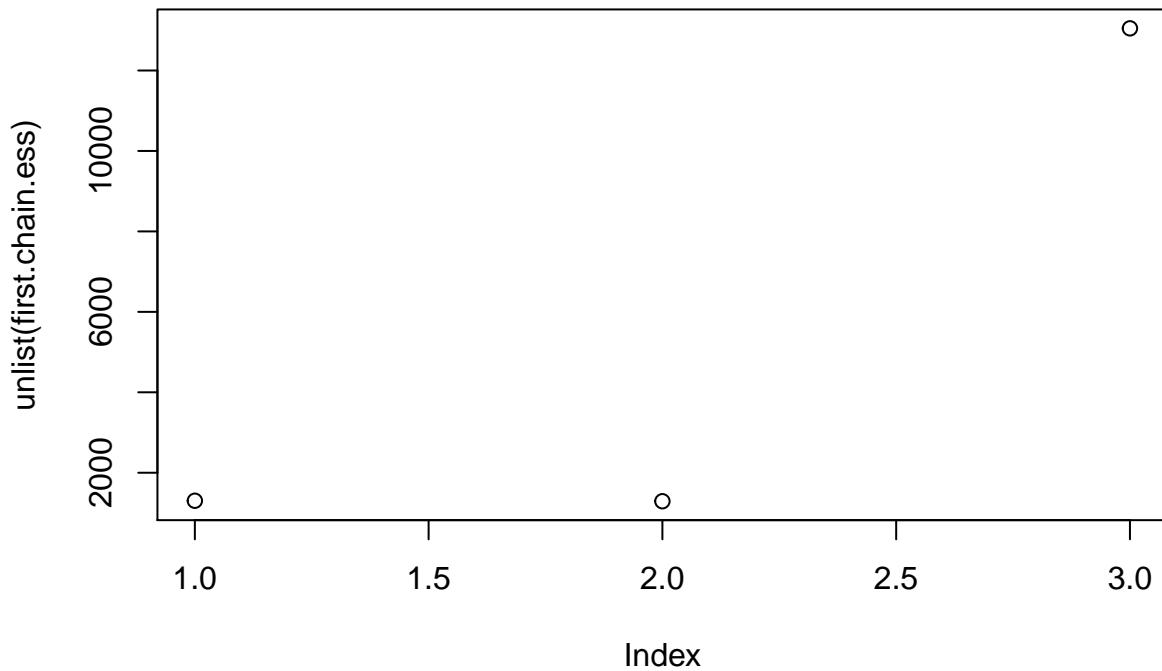
```
## [1] "JAGS GLM Models for Negative Binomial 2"
## Compiling model graph
##    Resolving undeclared variables
##    Allocating nodes
## Graph information:
##    Observed stochastic nodes: 41
##    Unobserved stochastic nodes: 3
##    Total graph size: 297
##
## Initializing model
```



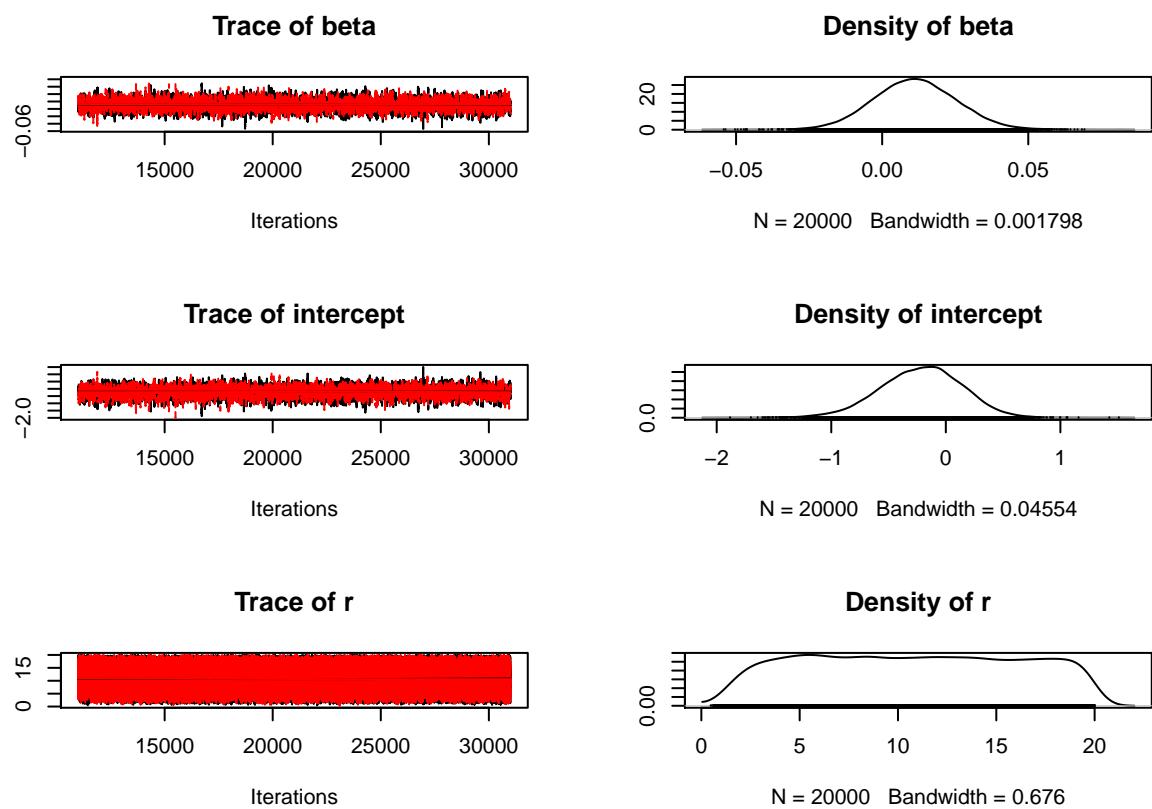
Gelman Diagnostic



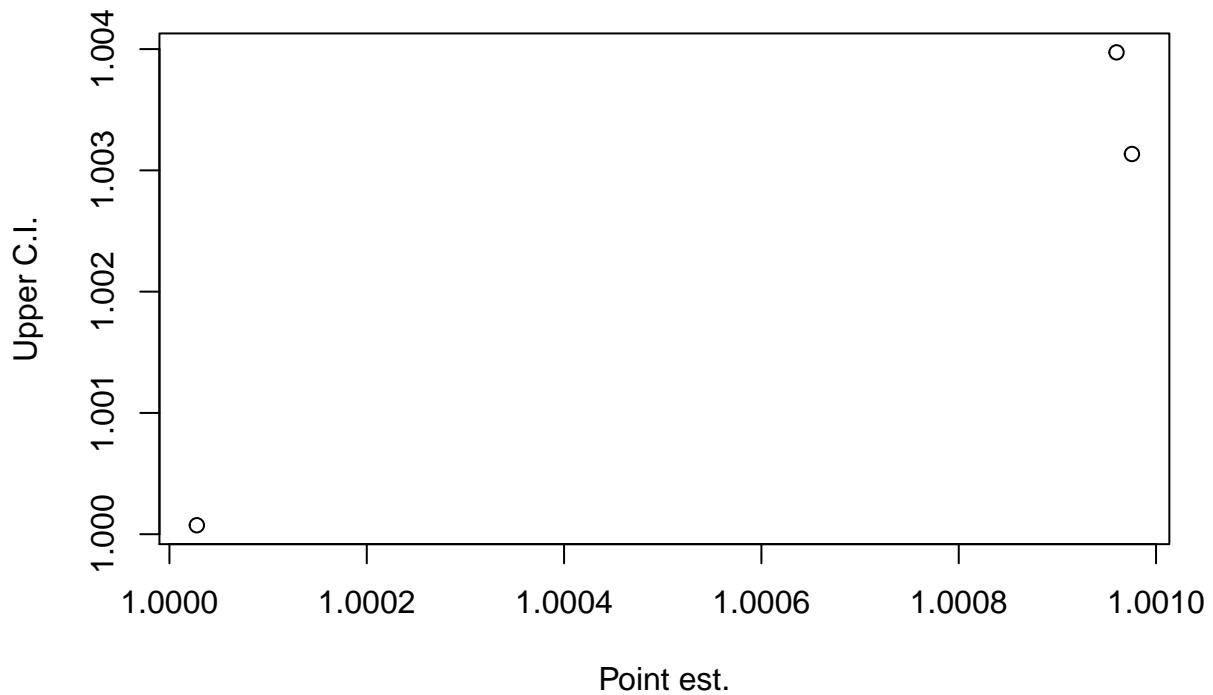
Effective Sample Size



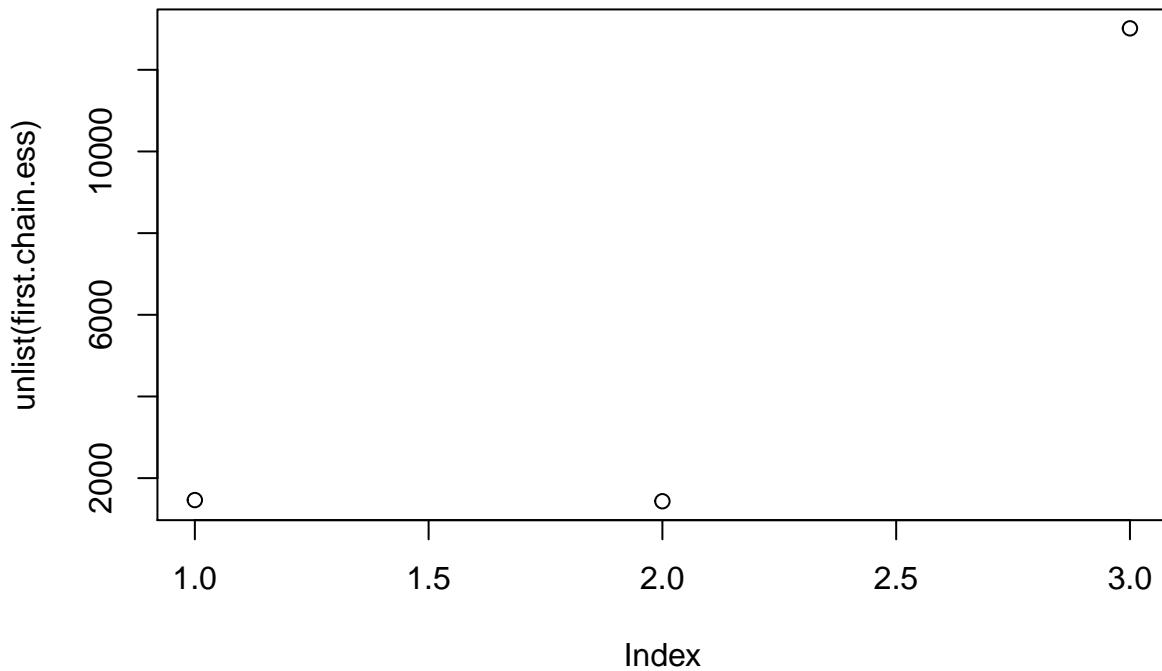
```
## [1] "JAGS GLM Models for Negative Binomial 3"
## Compiling model graph
##    Resolving undeclared variables
##    Allocating nodes
## Graph information:
##    Observed stochastic nodes: 41
##    Unobserved stochastic nodes: 3
##    Total graph size: 297
##
## Initializing model
```



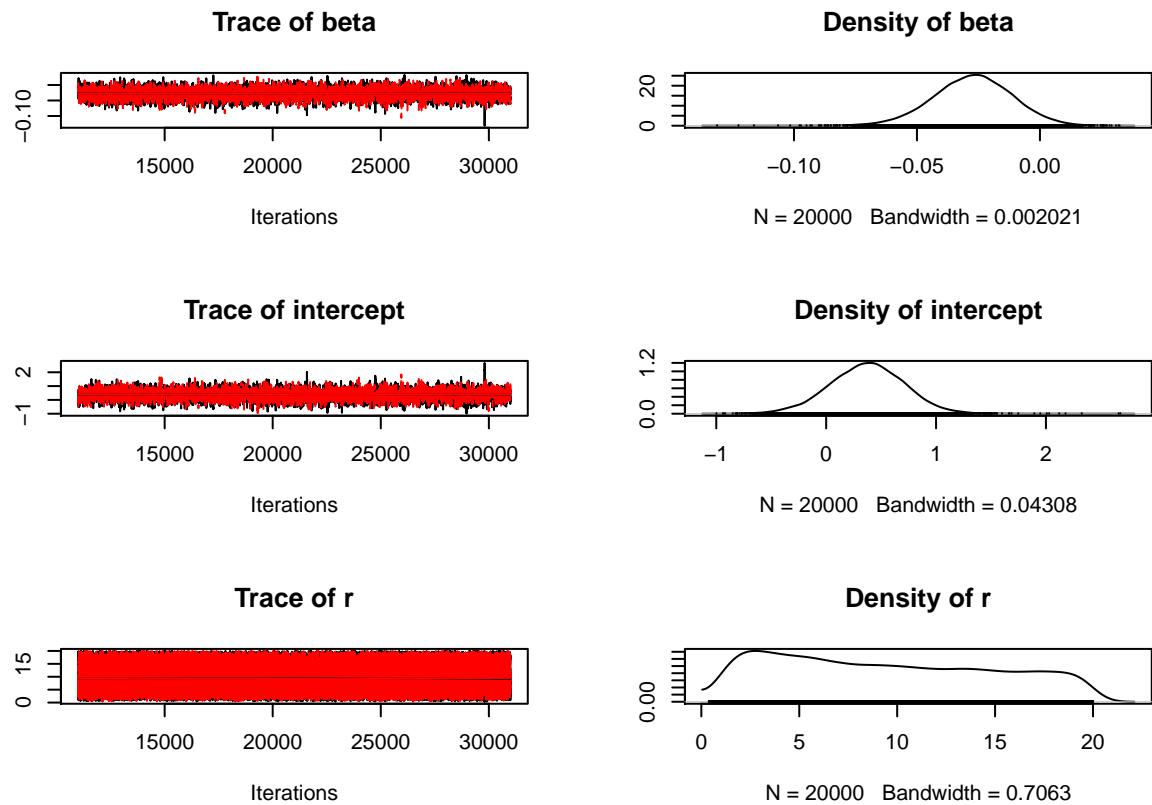
Gelman Diagnostic



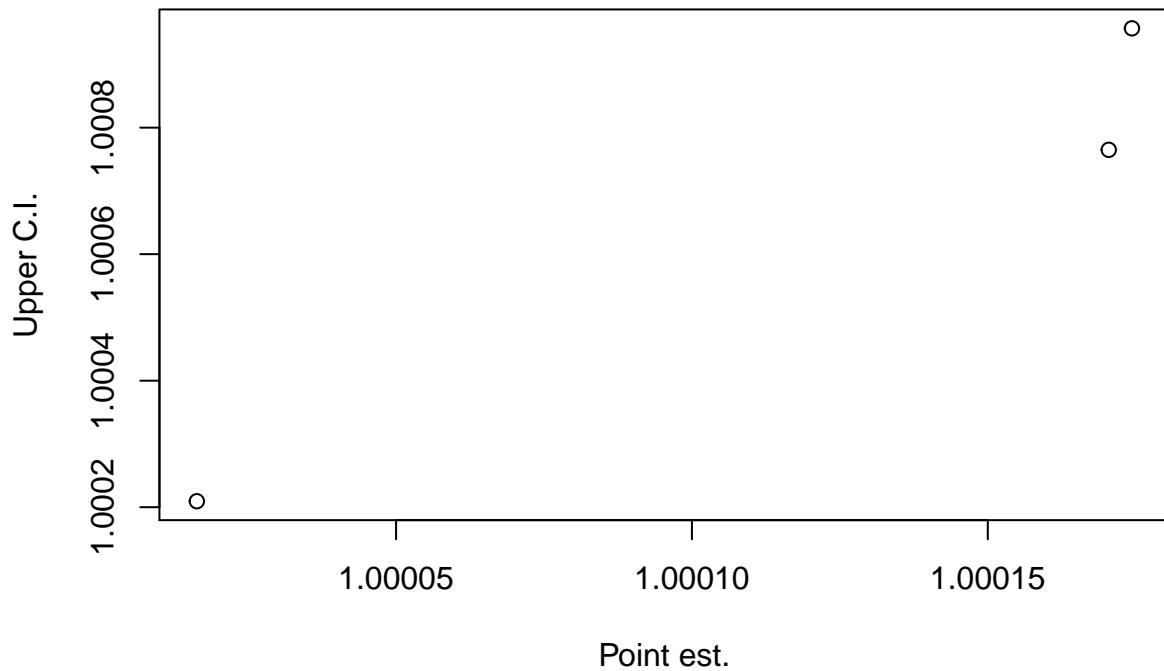
Effective Sample Size



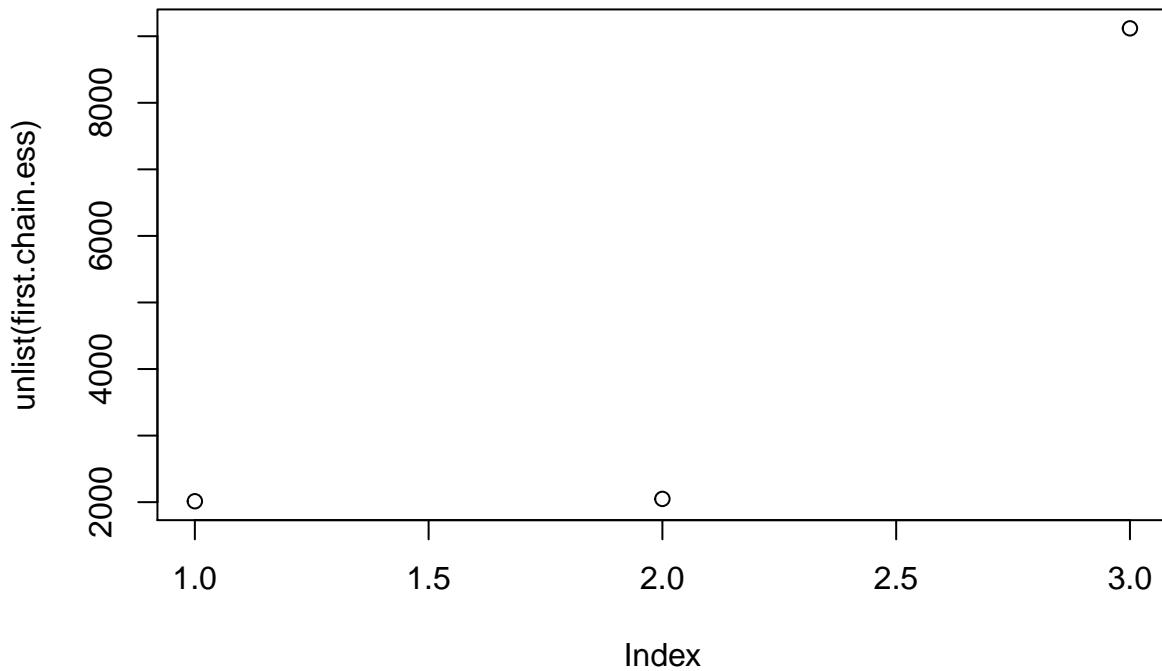
```
## [1] "JAGS GLM Models for Negative Binomial 4"
## Compiling model graph
##    Resolving undeclared variables
##    Allocating nodes
## Graph information:
##    Observed stochastic nodes: 41
##    Unobserved stochastic nodes: 3
##    Total graph size: 297
##
## Initializing model
```



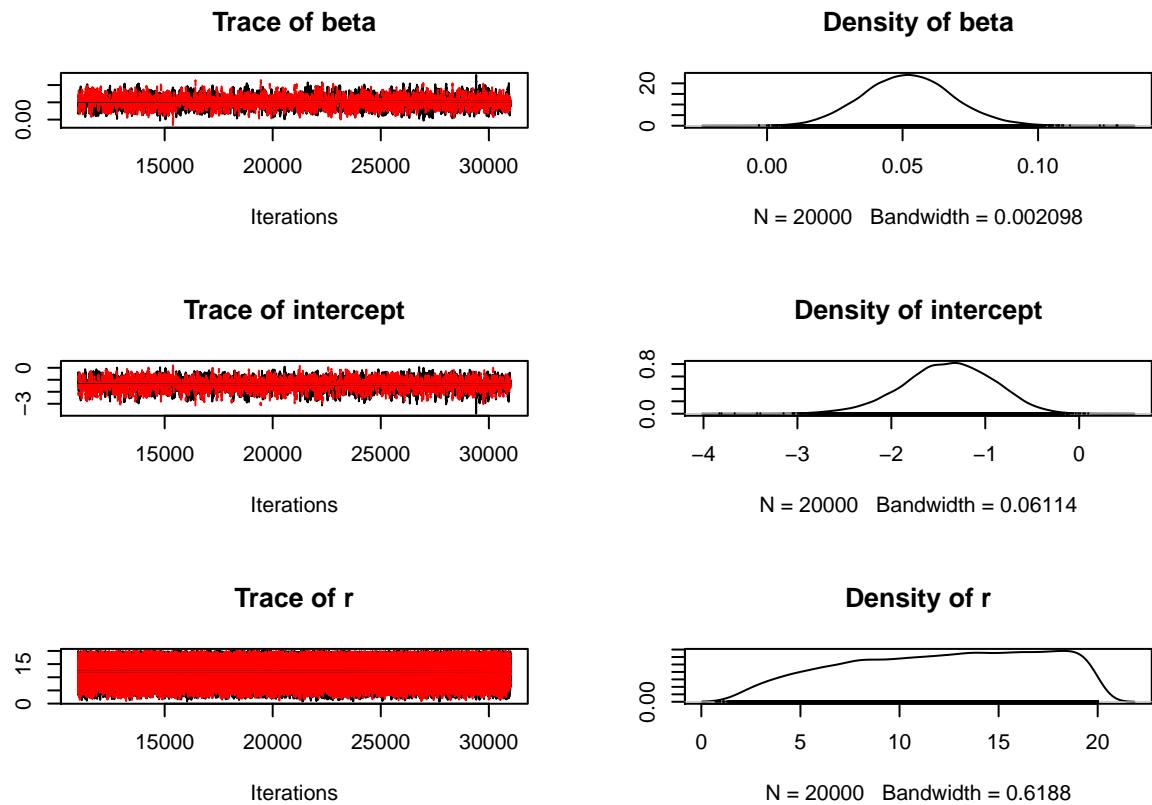
Gelman Diagnostic



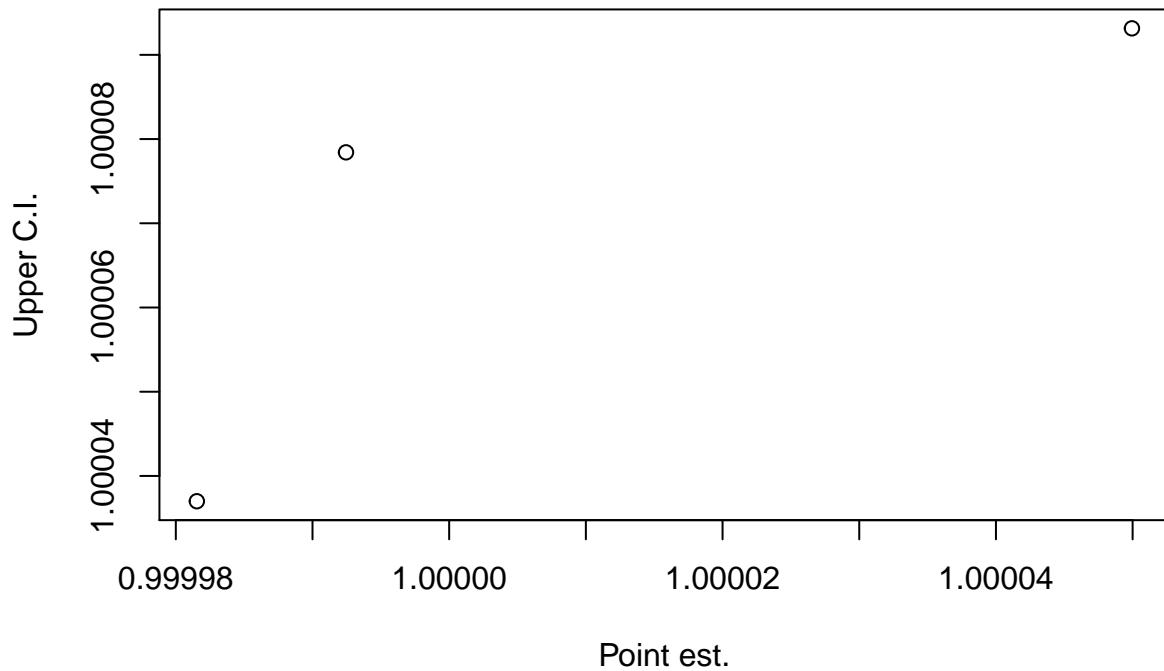
Effective Sample Size



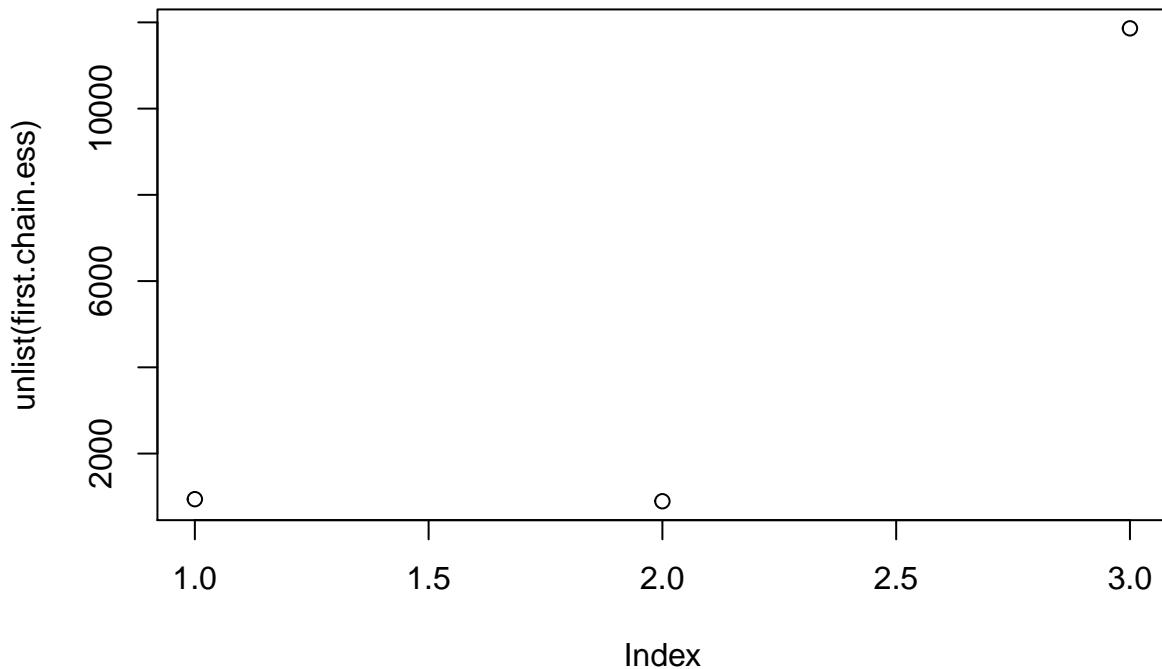
```
## [1] "JAGS GLM Models for Negative Binomial 5"
## Compiling model graph
##    Resolving undeclared variables
##    Allocating nodes
## Graph information:
##    Observed stochastic nodes: 41
##    Unobserved stochastic nodes: 3
##    Total graph size: 297
##
## Initializing model
```



Gelman Diagnostic



Effective Sample Size



Fit JAGS Poisson Random Effects

```
#####
##### Fit JAGS Poisson Random Effects
library(rjags)
library(coda)
model_code =
model
{
  ## Likelihood
  for(i in 1:N){
    for(j in 1:9){
      Y[i,j] ~ dpois(lambda[i,j])
      log(lambda[i,j]) <- mu[i,j]
      mu[i,j] <- intercept + beta*t[i] + alpha[j]
    }
  }

  ## Priors
  # Random effects
  for(i in 1:9){
    alpha[i] ~ dnorm(0,taus)
  }
  taus ~ dgamma(0.1,0.1)
  beta ~ dnorm(mu.beta,tau.beta)
```

```

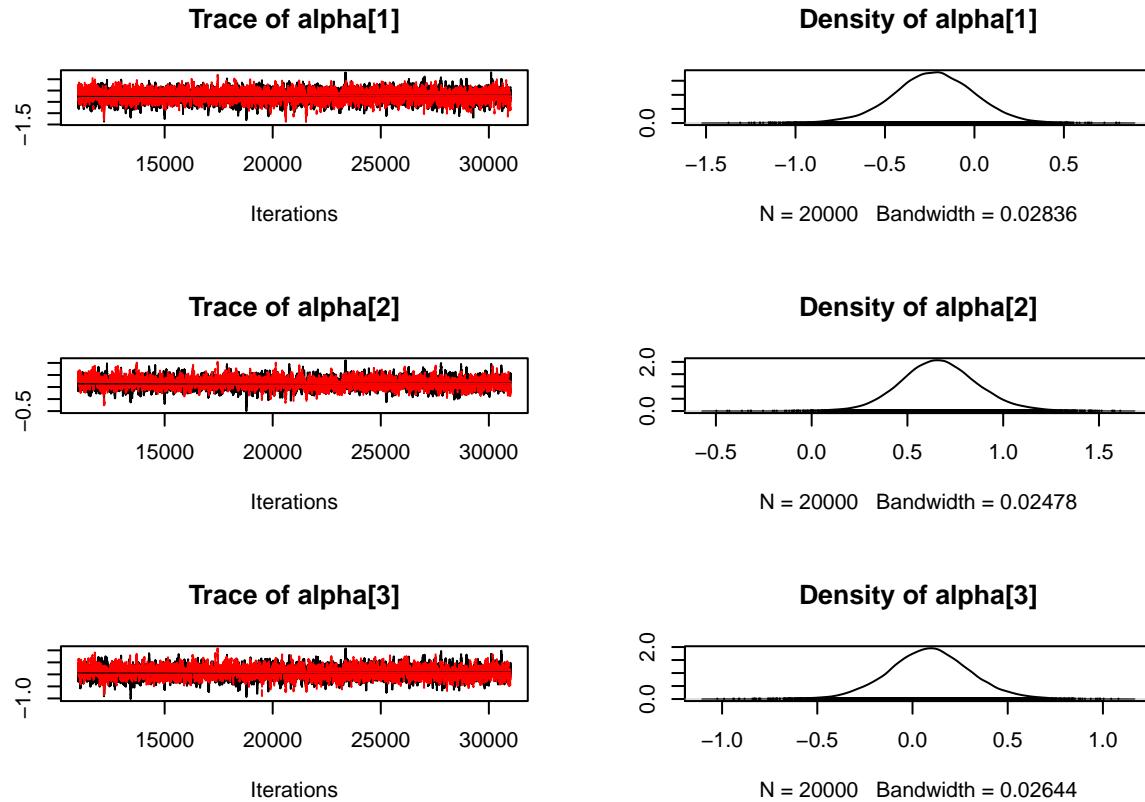
    intercept ~ dnorm(mu.intercept,tau.intercept)
}

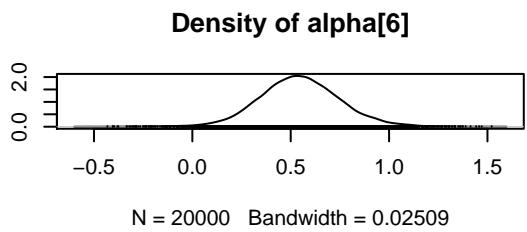
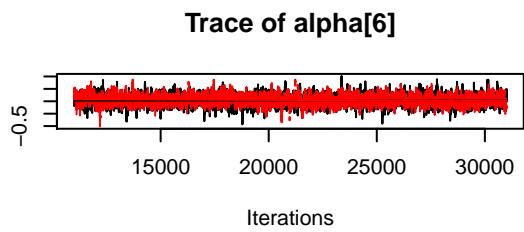
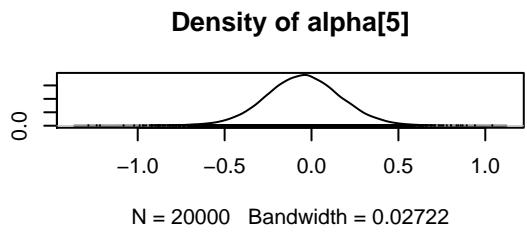
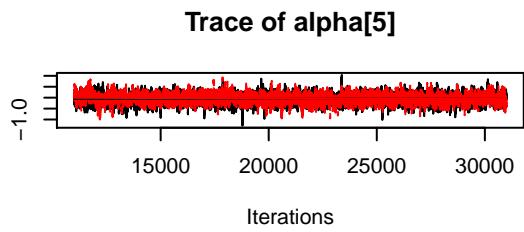
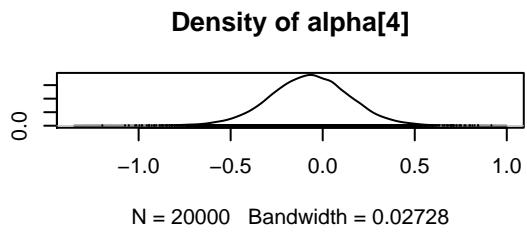
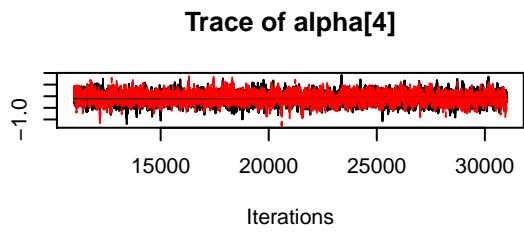
#
# Set up the data
model_data = list(N = 41, t=seq(1:41),Y=X.num,mu.beta=0,tau.beta=.0001,mu.intercept=0,tau.intercept=.0001)
# Choose the parameters to watch
model_parameters = c("r","beta", "intercept", "alpha")
n.chains =2;nSamples = 10000
model <- jags.model(textConnection(model_code),data = model_data,n.chains = n.chains)#Compile Model G

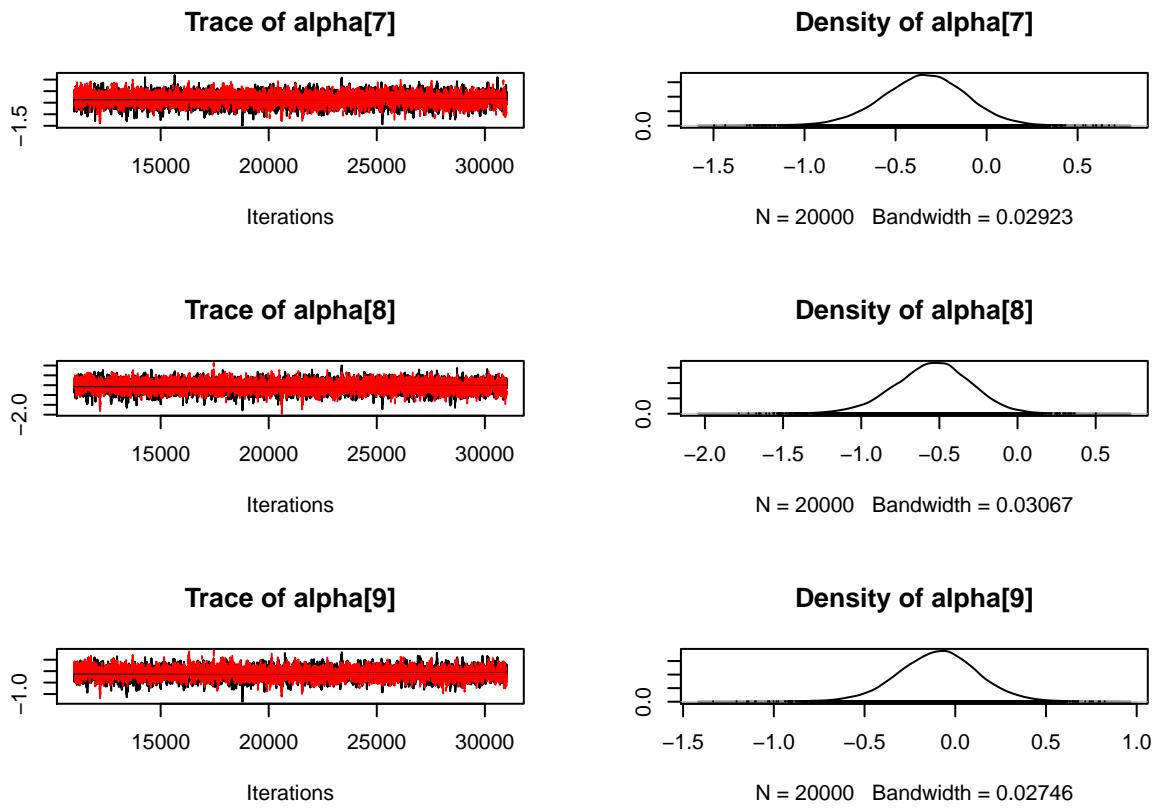
## Compiling model graph
## Resolving undeclared variables
## Allocating nodes
## Graph information:
##   Observed stochastic nodes: 369
##   Unobserved stochastic nodes: 12
##   Total graph size: 1208
##
## Initializing model
update(model, nSamples, progress.bar="none"); # Burnin
out.coda <- coda.samples(model, variable.names=model_parameters,n.iter=2*nSamples)

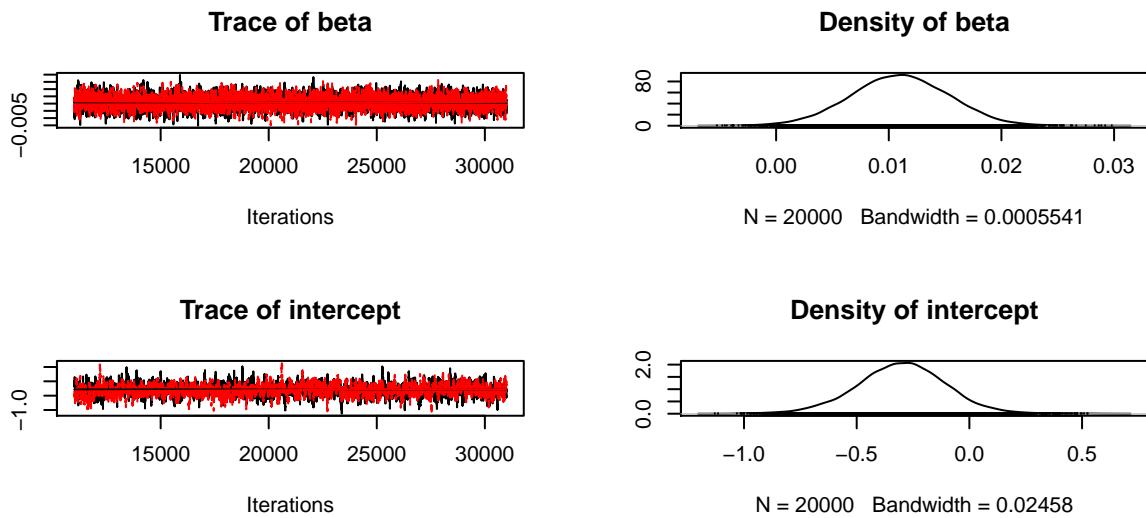
## Warning in jags.samples(model, variable.names, n.iter, thin, type = "trace", : Failed to set trace m
## Variable r not found
plot(out.coda)

```









```
#assess the posteriors??? stationarity, by looking at the Heidelberg-Welch convergence diagnostic:
heidel.diag(out.coda)
```

```
## [[1]]
##
##           Stationarity start      p-value
##           test      iteration
## alpha[1]  passed       1       0.105
## alpha[2]  passed       1       0.236
## alpha[3]  passed       1       0.282
## alpha[4]  passed       1       0.274
## alpha[5]  passed       1       0.593
## alpha[6]  passed       1       0.329
## alpha[7]  passed       1       0.431
## alpha[8]  passed       1       0.179
## alpha[9]  passed       1       0.209
## beta     passed       1       0.861
## intercept passed       1       0.476
##
##           Halfwidth Mean      Halfwidth
##           test
## alpha[1]  passed    -0.2376 0.014224
## alpha[2]  passed     0.6583 0.014674
## alpha[3]  failed     0.0798 0.014422
## alpha[4]  failed    -0.0792 0.013646
## alpha[5]  failed    -0.0559 0.014255
```

```

## alpha[6] passed 0.5376 0.014355
## alpha[7] passed -0.3520 0.013533
## alpha[8] passed -0.5563 0.013940
## alpha[9] failed -0.1020 0.013191
## beta passed 0.0109 0.000205
## intercept passed -0.2911 0.017849
##
## [[2]]
##
##           Stationarity start      p-value
##           test          iteration
## alpha[1] passed       6001      0.0530
## alpha[2] passed       6001      0.0649
## alpha[3] passed       6001      0.0673
## alpha[4] passed       4001      0.0871
## alpha[5] passed       6001      0.1634
## alpha[6] passed       6001      0.0510
## alpha[7] passed       8001      0.5321
## alpha[8] passed       6001      0.1846
## alpha[9] passed       6001      0.0973
## beta passed          1        0.1801
## intercept passed     2001      0.0529
##
##           Halfwidth Mean      Halfwidth
##           test
## alpha[1] passed    -0.2288 0.016439
## alpha[2] passed     0.6651 0.017896
## alpha[3] failed     0.0874 0.017894
## alpha[4] failed    -0.0700 0.015631
## alpha[5] failed    -0.0512 0.017550
## alpha[6] passed     0.5460 0.016334
## alpha[7] passed    -0.3566 0.017698
## alpha[8] passed    -0.5477 0.016362
## alpha[9] failed    -0.0972 0.017726
## beta passed        0.0110 0.000209
## intercept passed   -0.3055 0.018346

# check that our chain???'s length is satisfactory.
raftery.diag(out.coda)

```

```

## [[1]]
##
## Quantile (q) = 0.025
## Accuracy (r) = +/- 0.005
## Probability (s) = 0.95
##
##           Burn-in Total Lower bound Dependence
##           (M)    (N)  (Nmin) factor (I)
## alpha[1] 30    33978 3746      9.07
## alpha[2] 30    34140 3746      9.11
## alpha[3] 28    29992 3746      8.01
## alpha[4] 25    31705 3746      8.46
## alpha[5] 25    28360 3746      7.57
## alpha[6] 49    50379 3746     13.40
## alpha[7] 30    39582 3746     10.60

```

```

##   alpha[8] 25      27400 3746      7.31
##   alpha[9] 20      26525 3746      7.08
##   beta     18      19641 3746      5.24
##   intercept 54      54870 3746     14.60
##
##
## [[2]]
##
## Quantile (q) = 0.025
## Accuracy (r) = +/- 0.005
## Probability (s) = 0.95
##
##          Burn-in  Total Lower bound Dependence
##          (M)      (N)   (Nmin) factor (I)
##   alpha[1] 28      34818 3746      9.29
##   alpha[2] 48      52452 3746     14.00
##   alpha[3] 42      46536 3746     12.40
##   alpha[4] 30      32850 3746      8.77
##   alpha[5] 28      35917 3746      9.59
##   alpha[6] 35      40572 3746     10.80
##   alpha[7] 32      41912 3746     11.20
##   alpha[8] 40      55904 3746     14.90
##   alpha[9] 42      50463 3746     13.50
##   beta     18      18009 3746      4.81
##   intercept 50      51735 3746     13.80

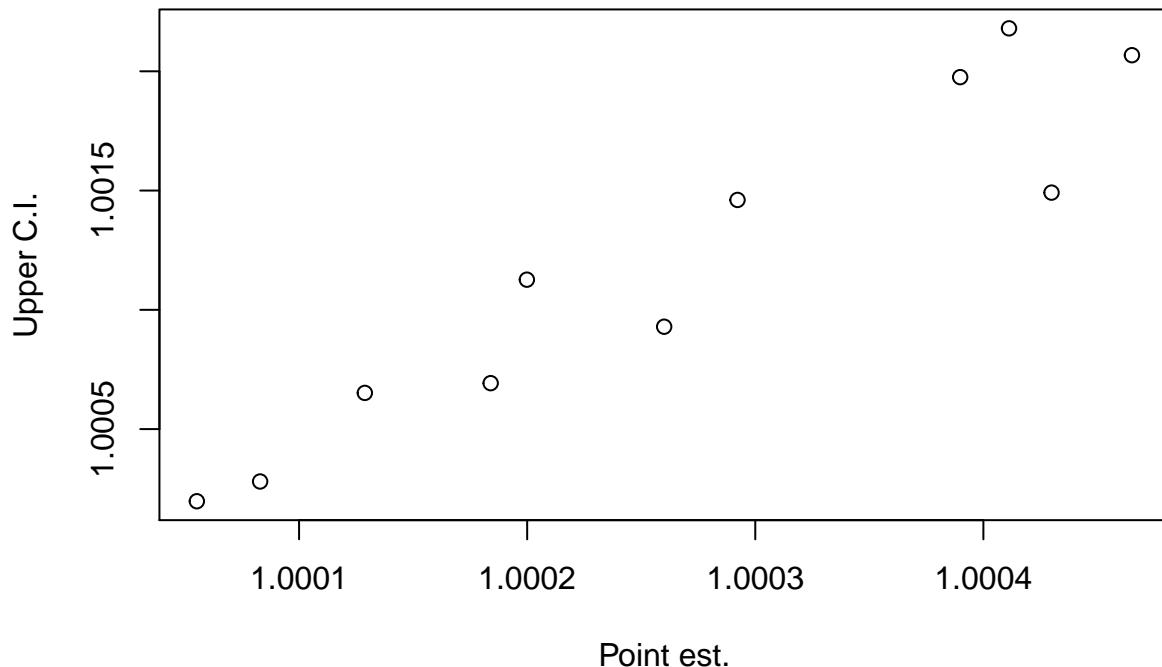
geweke.diag(out.coda)

## [[1]]
##
## Fraction in 1st window = 0.1
## Fraction in 2nd window = 0.5
##
##   alpha[1]  alpha[2]  alpha[3]  alpha[4]  alpha[5]  alpha[6]  alpha[7]
##   -0.18363 -0.32794  0.06228  0.01257  0.29577  0.05159  0.30095
##   alpha[8]  alpha[9]      beta intercept
##   0.18407 -0.22364  0.96630 -0.25604
##
##
## [[2]]
##
## Fraction in 1st window = 0.1
## Fraction in 2nd window = 0.5
##
##   alpha[1]  alpha[2]  alpha[3]  alpha[4]  alpha[5]  alpha[6]  alpha[7]
##   1.973    2.086    1.916    1.668    2.071    1.705    2.030
##   alpha[8]  alpha[9]      beta intercept
##   1.730    1.619    1.588   -1.899

if(n.chains > 1)
{
  gelman.srf <- gelman.diag(out.coda)
  plot(gelman.srf$psrf, main = "Gelman Diagnostic")
}

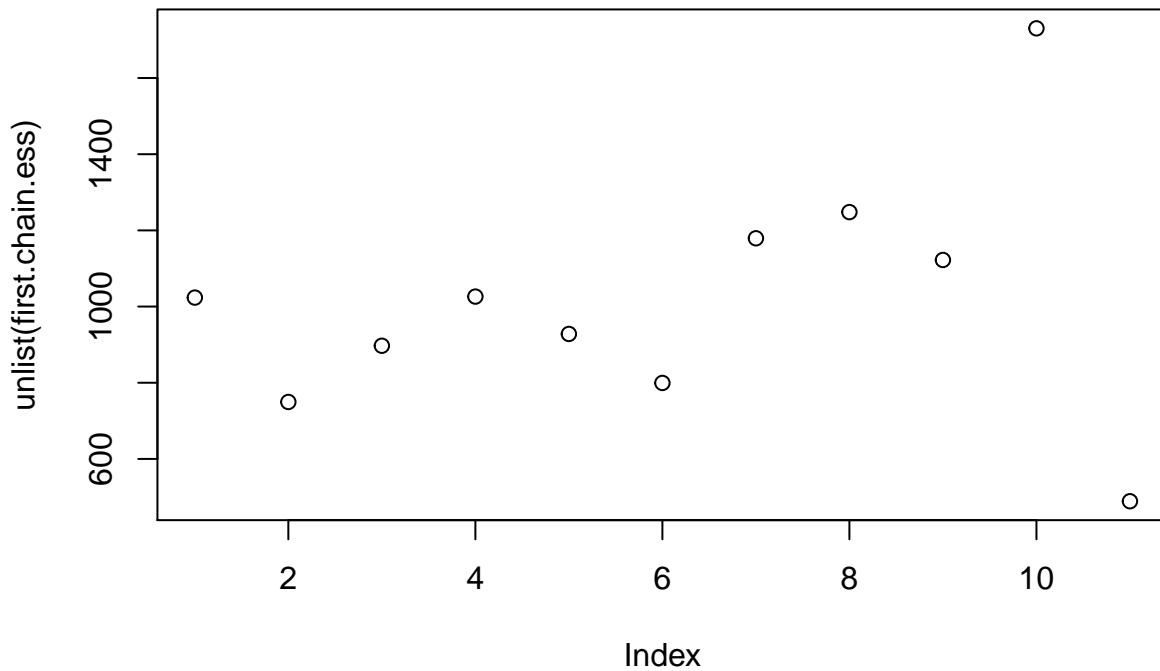
```

Gelman Diagnostic



```
chains.ess <- lapply(out.coda, effectiveSize)
first.chain.ess <- chains.ess[1]
plot(unlist(first.chain.ess), main="Effective Sample Size")
```

Effective Sample Size



Fit JAGS Negative Binomial Random Effects

```
#####
# Fit JAGS Negative Binomial Random Effects
library(rjags)
library(coda)
model_code = '
model
{
  ## Likelihood
  for(i in 1:N){
    for(j in 1:9){
      Y[i,j] ~ dnegbin(p[i,j],r)
      p[i,j] <- r/(r+lambda[i,j])
      log(lambda[i,j]) <- mu[i,j]
      mu[i,j] <- intercept + beta*t[i] + alpha[j]
    }
  }

  ## Priors
  # Random effects
  for(i in 1:9){
    alpha[i] ~ dnorm(0,taus)
  }
  taus ~ dgamma(0.1,0.1)
```

```

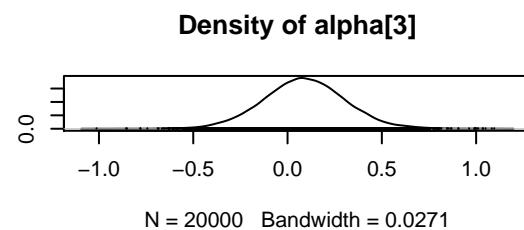
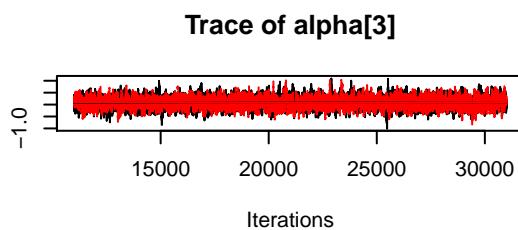
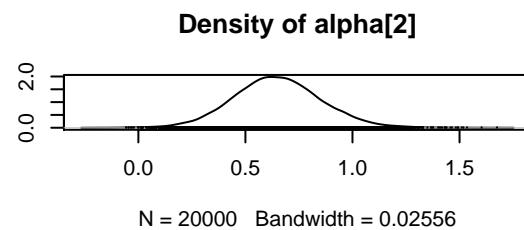
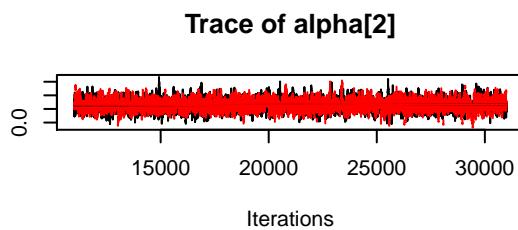
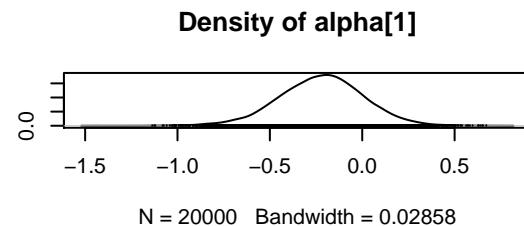
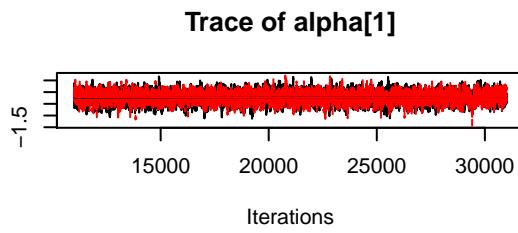
r ~ dunif(0,20)
beta ~ dnorm(mu.beta,tau.beta)
intercept ~ dnorm(mu.intercept,tau.intercept)
}

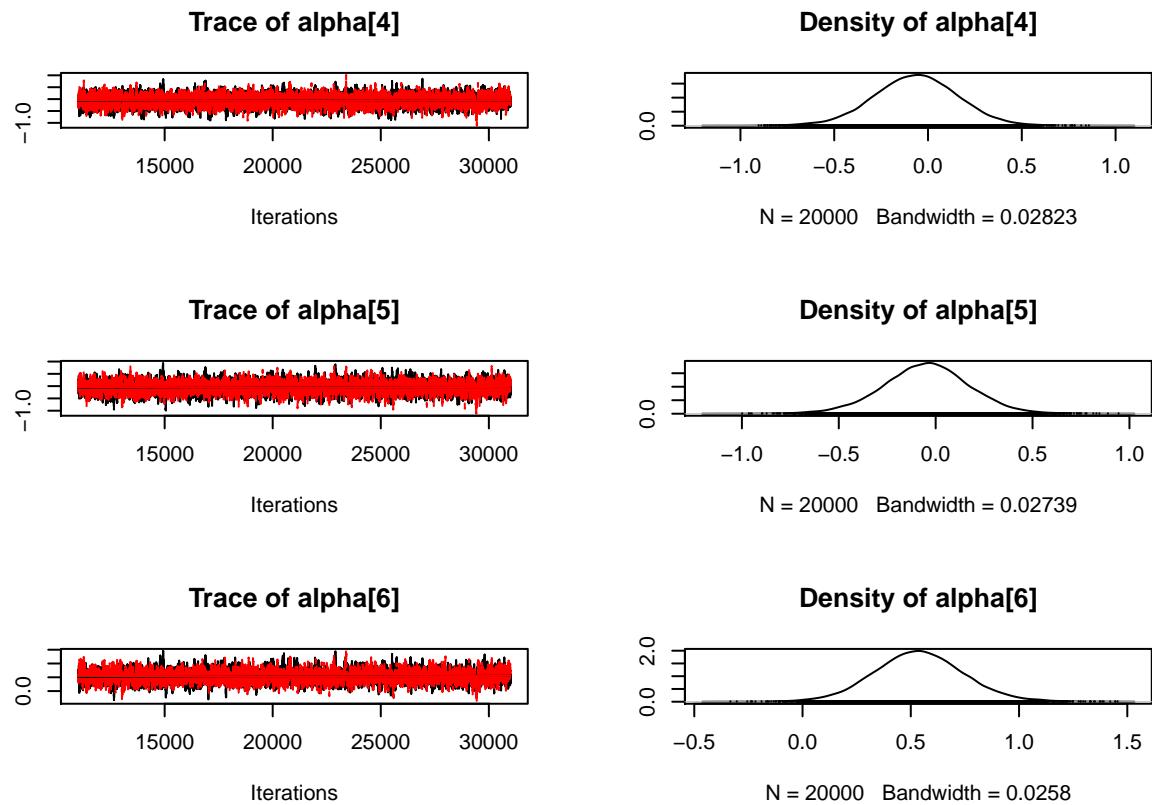
# Set up the data
model_data = list(N = 41, t=seq(1:41),Y=X.num, mu.beta=0, tau.beta=.0001, mu.intercept=0, tau.intercept=.0001)

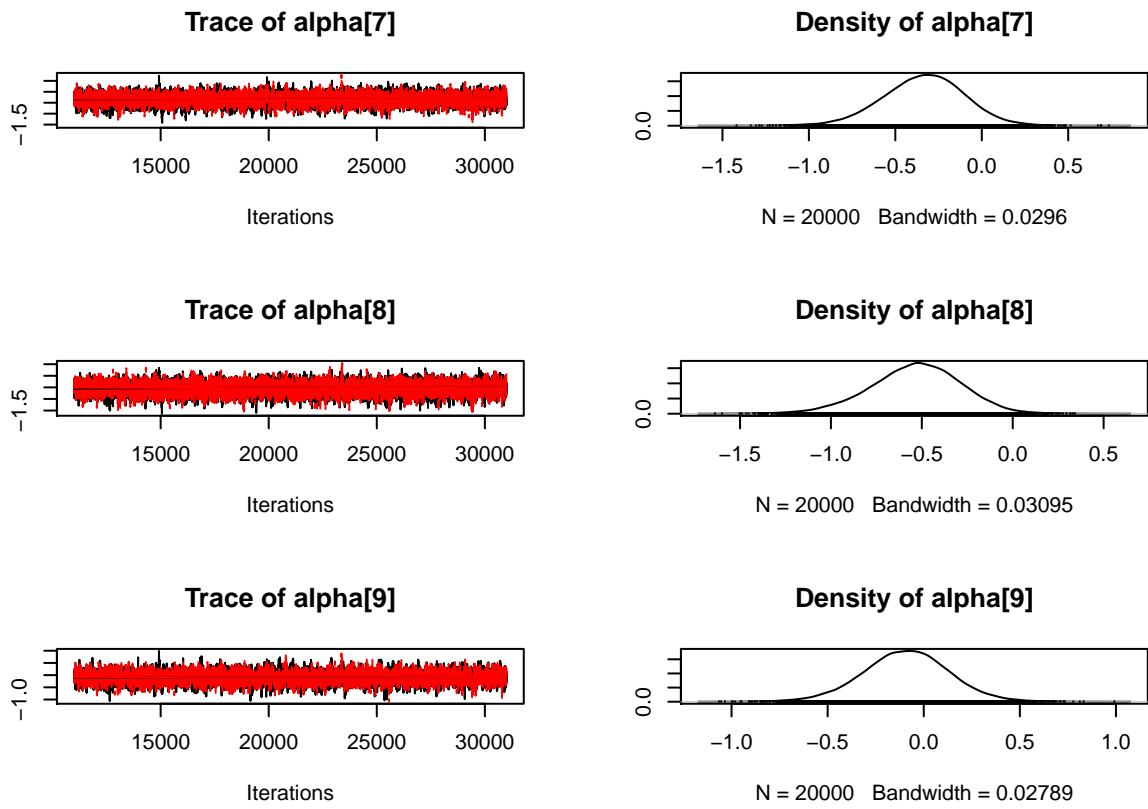
# Choose the parameters to watch
model_parameters = c("r", "beta", "intercept", "alpha")
n.chains = 2; nSamples = 10000
model <- jags.model(textConnection(model_code), data = model_data, n.chains = n.chains) #Compile Model G

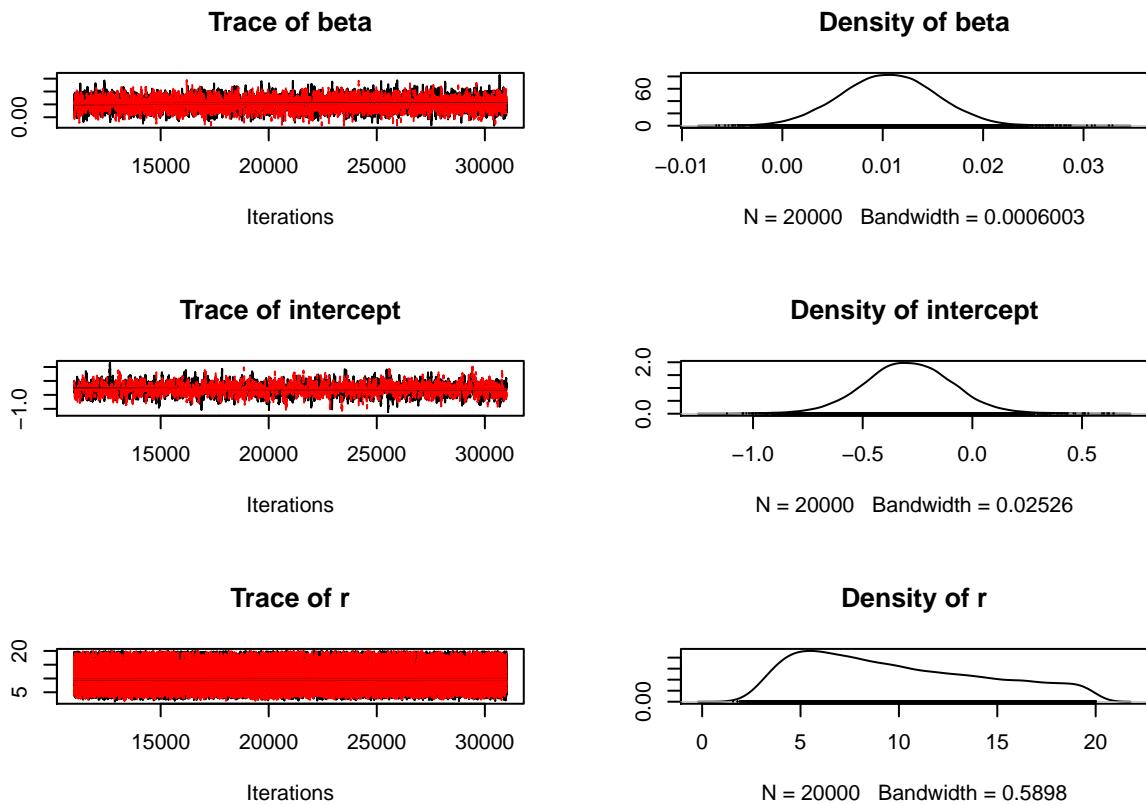
## Compiling model graph
## Resolving undeclared variables
## Allocating nodes
## Graph information:
##   Observed stochastic nodes: 369
##   Unobserved stochastic nodes: 13
##   Total graph size: 1948
##
## Initializing model
update(model, nSamples, progress.bar="none"); # Burnin
out.coda <- coda.samples(model, variable.names=model_parameters, n.iter=2*nSamples)
plot(out.coda)

```









```
#assess the posteriors??? stationarity, by looking at the Heidelberg-Welch convergence diagnostic:
heidel.diag(out.coda)
```

```
## [1]
##
##          Stationarity start      p-value
##          test      iteration
## alpha[1] passed      1      0.1324
## alpha[2] passed      1      0.4053
## alpha[3] passed      1      0.2897
## alpha[4] passed      1      0.4878
## alpha[5] passed      1      0.3330
## alpha[6] passed      1      0.1299
## alpha[7] passed      1      0.2904
## alpha[8] passed      1      0.1111
## alpha[9] passed      1      0.3589
## beta     passed    2001      0.0651
## intercept passed      1      0.1132
## r        passed      1      0.6124
##
##          Halfwidth Mean      Halfwidth
##          test
## alpha[1] passed   -0.2220 0.011530
## alpha[2] passed    0.6500 0.012891
## alpha[3] failed   0.0901 0.012442
## alpha[4] failed  -0.0637 0.012137
```

```

## alpha[5] failed -0.0485 0.012543
## alpha[6] passed 0.5383 0.012539
## alpha[7] passed -0.3355 0.011454
## alpha[8] passed -0.5335 0.011363
## alpha[9] failed -0.0918 0.012007
## beta passed 0.0107 0.000228
## intercept passed -0.2947 0.016526
## r passed 9.8348 0.092130
##
## [[2]]
##
##           Stationarity start      p-value
##           test          iteration
## alpha[1] passed        1        0.983
## alpha[2] passed        1        0.999
## alpha[3] passed        1        1.000
## alpha[4] passed        1        0.950
## alpha[5] passed        1        0.996
## alpha[6] passed        1        0.998
## alpha[7] passed        1        0.835
## alpha[8] passed        1        0.825
## alpha[9] passed        1        0.988
## beta passed          1        0.977
## intercept passed      1        0.996
## r passed            1        0.647
##
##           Halfwidth Mean      Halfwidth
##           test
## alpha[1] passed    -0.2210 0.013116
## alpha[2] passed    0.6488 0.013165
## alpha[3] failed    0.0881 0.013383
## alpha[4] failed   -0.0648 0.013001
## alpha[5] failed   -0.0499 0.013399
## alpha[6] passed    0.5389 0.013779
## alpha[7] passed   -0.3381 0.012567
## alpha[8] passed   -0.5368 0.012682
## alpha[9] failed   -0.0932 0.012821
## beta passed       0.0105 0.000227
## intercept passed -0.2881 0.017536
## r passed          9.8491 0.098147
# check that our chain???'s length is satisfactory.
raftery.diag(out.coda)

```

```

## [[1]]
##
## Quantile (q) = 0.025
## Accuracy (r) = +/- 0.005
## Probability (s) = 0.95
##
##           Burn-in Total Lower bound Dependence
##           (M)     (N)  (Nmin)    factor (I)
## alpha[1] 21    27286 3746      7.28
## alpha[2] 30    32030 3746      8.55
## alpha[3] 24    26166 3746      6.99

```

```

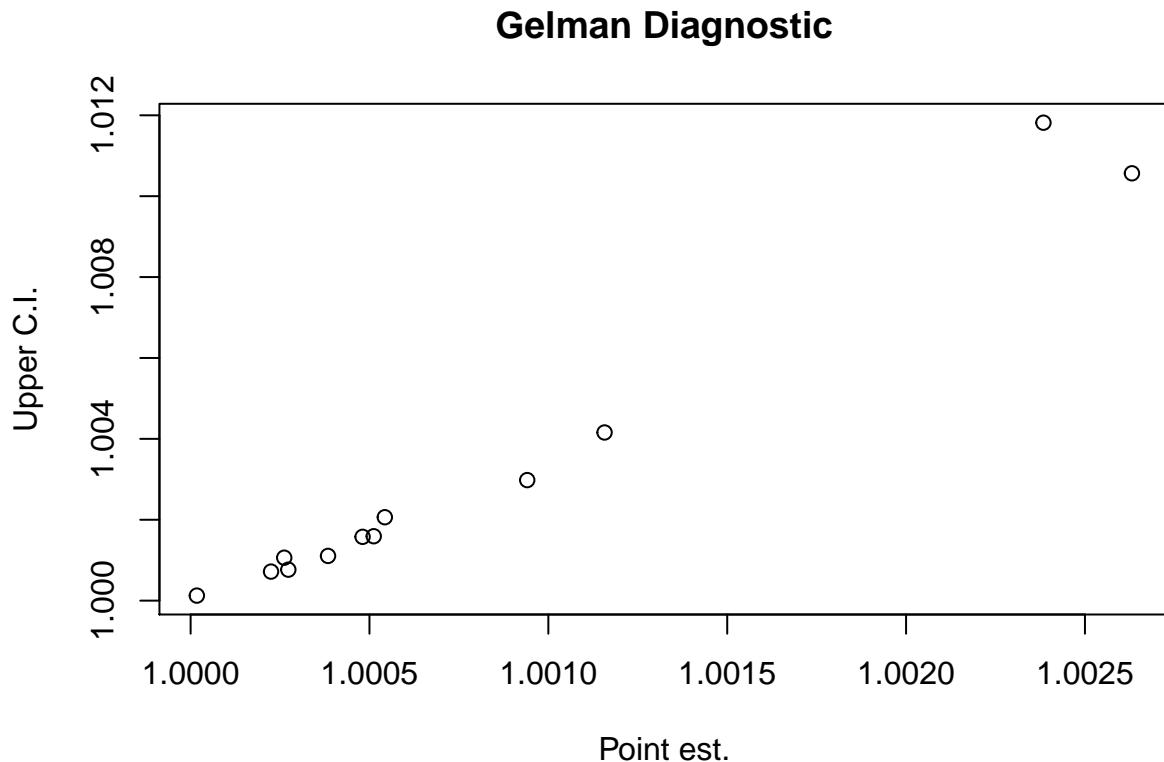
## alpha[4] 24      28494 3746      7.61
## alpha[5] 16      17588 3746      4.70
## alpha[6] 24      29322 3746      7.83
## alpha[7] 12      15006 3746      4.01
## alpha[8] 16      18496 3746      4.94
## alpha[9] 16      18172 3746      4.85
## beta     15      18408 3746      4.91
## intercept 42    40614 3746      10.80
## r        3       4428  3746      1.18
##
##
## [[2]]
##
## Quantile (q) = 0.025
## Accuracy (r) = +/- 0.005
## Probability (s) = 0.95
##
##          Burn-in Total Lower bound Dependence
##          (M)   (N)   (Nmin) factor (I)
## alpha[1] 20      23880 3746      6.37
## alpha[2] 35      40103 3746     10.70
## alpha[3] 30      29900 3746      7.98
## alpha[4] 25      25560 3746      6.82
## alpha[5] 25      33760 3746      9.01
## alpha[6] 24      26332 3746      7.03
## alpha[7] 20      24335 3746      6.50
## alpha[8] 20      23965 3746      6.40
## alpha[9] 30      40242 3746     10.70
## beta     15      18486 3746      4.93
## intercept 42    43972 3746     11.70
## r        3       4373  3746      1.17

geweke.diag(out.coda)

## [[1]]
##
## Fraction in 1st window = 0.1
## Fraction in 2nd window = 0.5
##
## alpha[1]  alpha[2]  alpha[3]  alpha[4]  alpha[5]  alpha[6]  alpha[7]
## -1.4251  -0.3992  -1.2804  -1.0409  -0.9972  -1.1202  -1.1090
## alpha[8]  alpha[9]      beta intercept      r
## -1.9664  -0.4029  -1.4711   1.1859   0.6916
##
##
## [[2]]
##
## Fraction in 1st window = 0.1
## Fraction in 2nd window = 0.5
##
## alpha[1]  alpha[2]  alpha[3]  alpha[4]  alpha[5]  alpha[6]  alpha[7]
## 1.6304   1.0406   1.0067   1.2836   1.4355   1.2204   1.8260
## alpha[8]  alpha[9]      beta intercept      r
## 2.0767   1.1904   0.4412  -1.2131  -0.5063

```

```
if(n.chains > 1)
{
  gelman.srf <- gelman.diag(out.coda)
  plot(gelman.srf$psrf, main = "Gelman Diagnostic")
}
```



```
chains.ess <- lapply(out.coda, effectiveSize)
first.chain.ess <- chains.ess[1]
plot(unlist(first.chain.ess), main="Effective Sample Size")
```

Effective Sample Size

