

ClearanceRate_F1Scallops

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Merge to master file 'ClearRate_Master'

Date from: - 20210914 - 20210930

```
# 20210914 DATA ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
# formatting for merge
# call the 914 data and remove blanks
clear.rate_914_Scallops <- merge( # note - merging with th elengths removed the blanks from the clearan
  (clear.rate_914 %>% dplyr::filter(!Chamber_tank %in% 'Blank')),
  (length.resp.clear %>%
    dplyr::filter(Date %in% 20210914) %>%
    dplyr::mutate(Chamber_tank = sub("_", "", Chamber_tank))) ) %>%
  dplyr::select(!c('Food'))

# 20210930 DATA ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
# a bit more work with the 930 data to merge with the 914
clear.rate_930_Scallops <- merge( (data.frame(melt(clear.rate_930, id.vars = c('Date', 'Chamber_tank', '
  dplyr::filter(!Fed_Unfed %in% 'blank') %>%
  dplyr::mutate(Time._min = (gsub(".*_", "", variable))) %>%
  dplyr::rename(Count = value) %>%
  dplyr::arrange(Run, Plate, pH, Replicate, Number, Fed_Unfed, Time._min) ), #merge with...
  (length.resp.clear %>%
    dplyr::filter(Date %in% 20210930) %>%
    dplyr::mutate(Chamber_tank = sub("_", "", Chamber_tank)) %>%
    dplyr::mutate(Fed_Unfed = ifelse(Food == 1, 'fed', 'unfed')))
  ) %>%
  dplyr::select(!c('variable', 'Food', 'Sample.ID'))

ClearRate_Master <- rbind(clear.rate_914_Scallops, clear.rate_930_Scallops)

# calculate the clearance rate normalized for shell length
ClearRate_Master$Cells_ml <- (ClearRate_Master$Count)*(1000/33)

print(head(ClearRate_Master))
```

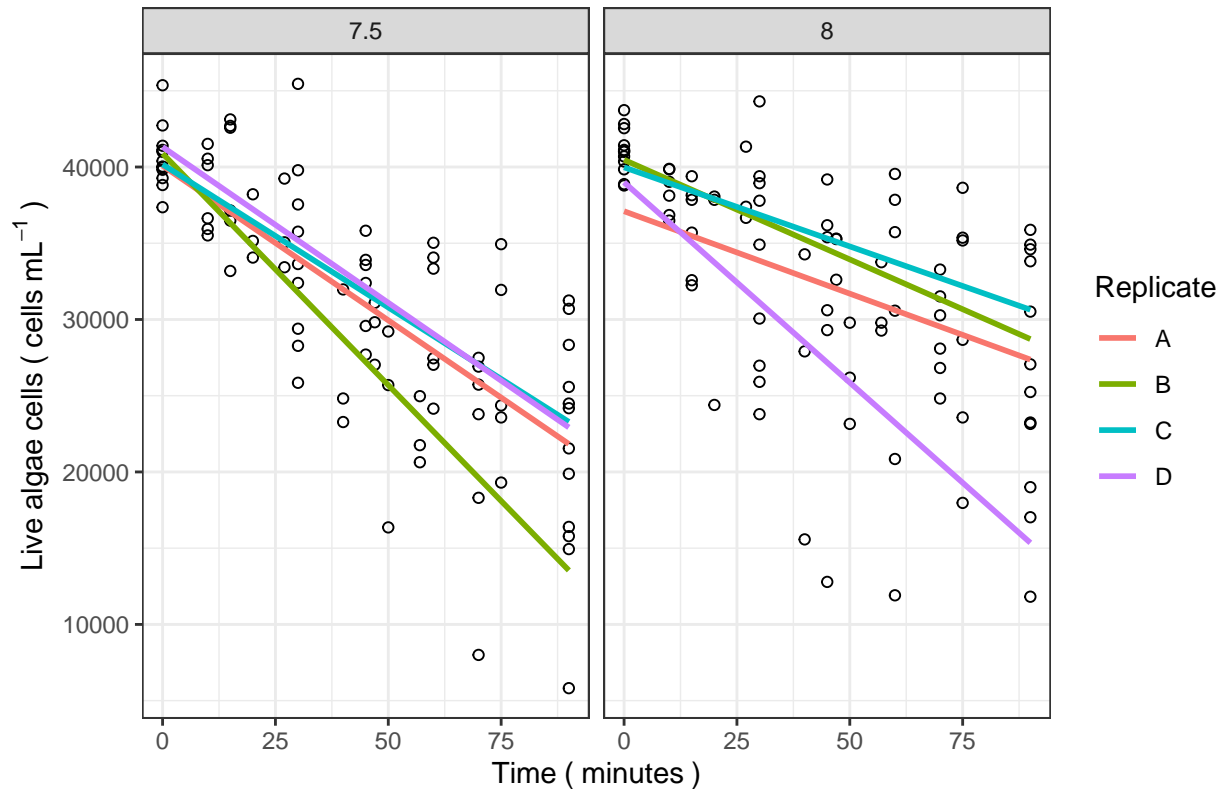
##	Date	Run	pH	Chamber_tank	Fed_Unfed	Time._min	Count	Plate	Replicate
## 1	20210914	1	7.5	7.5A	<NA>	0	1314	NA	A
## 2	20210914	1	7.5	7.5A	<NA>	10	1186	NA	A
## 3	20210914	1	7.5	7.5A	<NA>	27	1295	NA	A

Visualize raw data & simple slopes/algae loss time-1

- note: some data points were omitted in the worksheet, review the 'raw' vs 'worksheet_R' data files

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RAW DATA: F1 Scallops, Clearance Rate 20210914

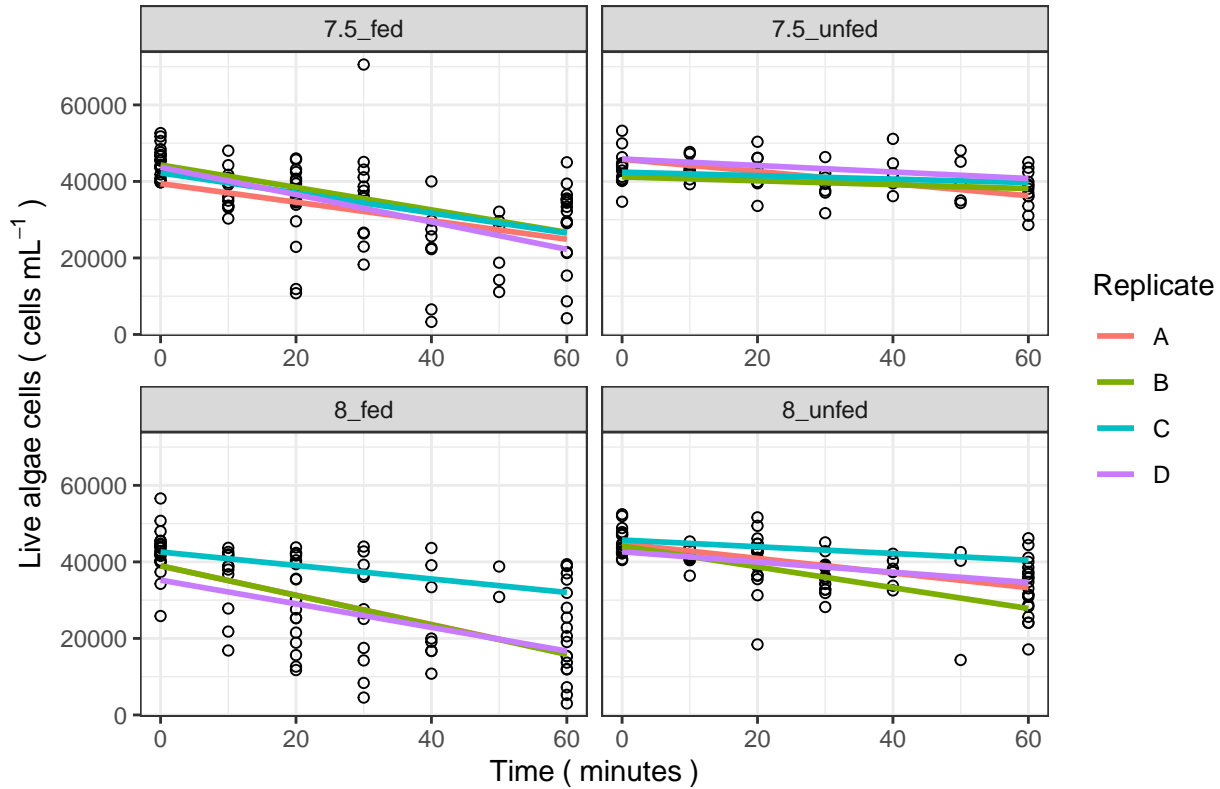


```
# Clearance rate analysis for 9/30 data
ClearRate_Master.914 <- ClearRate_Master %>%
  dplyr::filter(Date %in% 20210914) %>%
  dplyr::mutate(unique_Identifier = paste(pH, "Run", Run, "Rep", Replicate, "Num", Number, sep='_'))

loop_914 <- as.data.frame(unique(ClearRate_Master.914$unique_Identifier)) %>% dplyr::rename(ID = "unique_Identifier")

SlopeTable_914 <- data.frame() # run this before the loop
for(i in 1:nrow(loop_914)){
  dat <- ClearRate_Master.914 %>% filter(unique_Identifier %in% loop_914[i,])
  slope <- summary(lm((dat$Cells_ml) ~ as.numeric(dat$Time_min)))$coef[2, "Estimate"]
  SLOPE <- summary(lm((dat$Cells_ml) ~ as.numeric(dat$Time_min)))$r.squared
  pval <- summary(lm((dat$Cells_ml) ~ as.numeric(dat$Time_min)))$coef[2, "Pr(>|t|)"]
  mod <- lm(as.numeric(dat$Time_min) ~ dat$Cells_ml)
  norm_assum <- shapiro.test(resid(mod))
  shapiro_pval <- norm_assum$p.value
  # assign the data table
  SLOPE.loop <- data.frame(matrix(nrow = 1, ncol = 5)) # create a new data table
  colnames(SLOPE.loop) <- c('pH', 'Replicate', 'slope', 'SLOPE', 'pval') # assign headers
  SLOPE.loop$pH <- gsub("_.*", "\\1", loop_914[i,])
  SLOPE.loop$Replicate <- gsub("^(?:[^\_+]{4}([^\_+]).*)", "\\1", loop_914[i,])
  SLOPE.loop$slope <- slope * 60 # cells per mL per hour
  SLOPE.loop$Number <- gsub("^(?:[^\_+]{6}([^\_+]).*)", "\\1", loop_914[i,])
  SLOPE.loop$SLOPE <- SLOPE
  SLOPE.loop$pval <- pval
  SLOPE.loop$shapiro_pval <- shapiro_pval
}
```

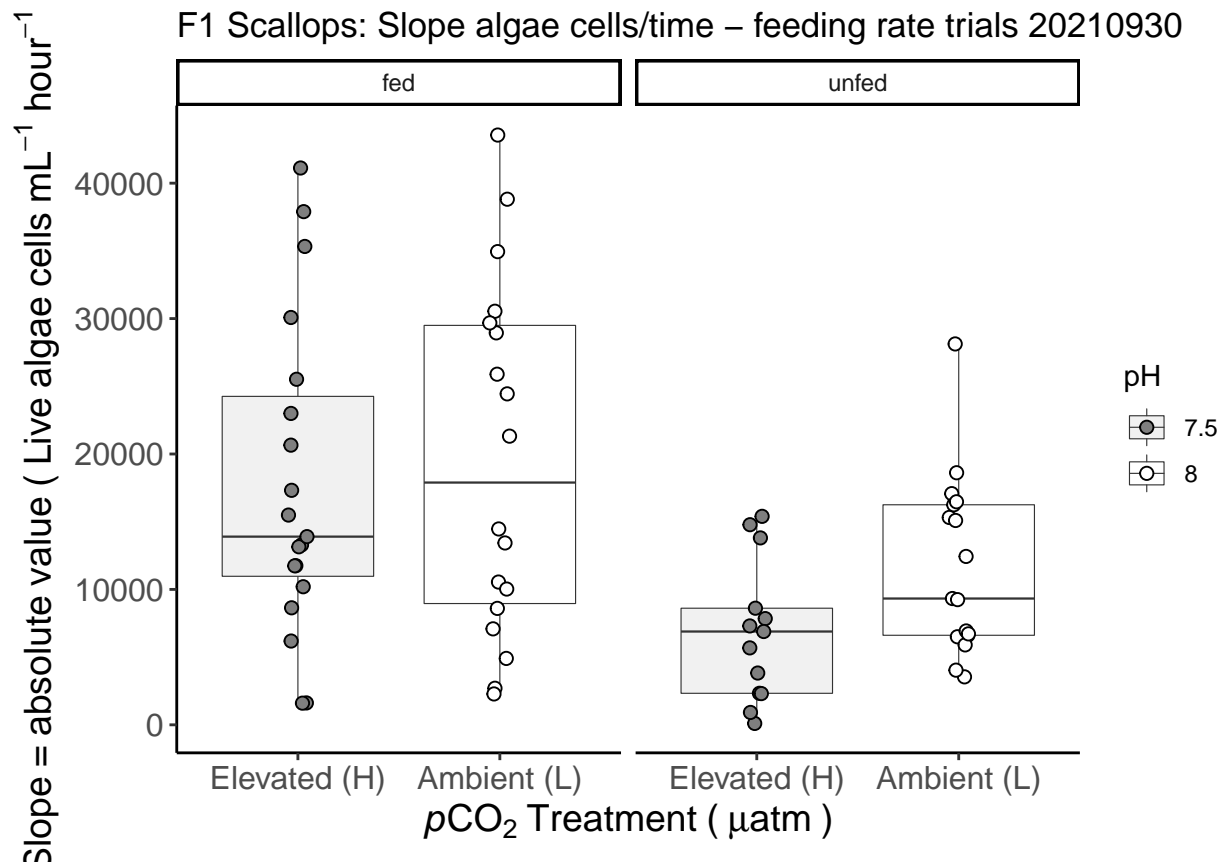

RAW DATA: F1 Scallops, Clearance Rate 20210930



```
# Clearance rate analysis for 9/30 data
ClearRate_Master.930 <- ClearRate_Master %>%
  dplyr::filter(Date %in% 20210930) %>%
  dplyr::mutate(unique_Identifier = paste(pH, Fed_Unfed, "Run", Run, "Rep", Replicate, "Num", Number, sep=

loop_930 <- as.data.frame(unique(ClearRate_Master.930$unique_Identifier)) %>% dplyr::rename(ID = "unique(

SlopeTable_930 <- data.frame() # run this before the loop
for(i in 1:nrow(loop_930)){
  dat <- ClearRate_Master.930 %>% filter(unique_Identifier %in% loop_930[i,])
  slope<- summary(lm((dat$Cells_ml) ~ as.numeric(dat$Time._min)))$coef[2,"Estimate"]
  SLOPE <- summary(lm((dat$Cells_ml) ~ as.numeric(dat$Time._min)))$r.squared
  pval <- summary(lm((dat$Cells_ml) ~ as.numeric(dat$Time._min)))$coef[2,"Pr(>|t|)"]
  mod <- lm(as.numeric(dat$Time._min) ~ dat$Cells_ml)
  norm_assum <- shapiro.test(resid(mod))
  shapiro_pval <- norm_assum$p.value
  # assign the data table
  SLOPE.loop <- data.frame(matrix(nrow = 1, ncol = 6)) # create a new data table
  colnames(SLOPE.loop) <- c('pH', 'Replicate', 'Fed_Unfed','slope', 'SLOPE', 'pval') # assign headers
  SLOPE.loop$pH <- gsub("_.*", "\\1", loop_930[i,])
  SLOPE.loop$Replicate <- gsub("^(:[^_+]{5}([^_+]).*", "\\1", loop_930[i,])
  SLOPE.loop$Fed_Unfed <- gsub("^(:[^_+]{1}([^_+]).*", "\\1", loop_930[i,])
  SLOPE.loop$slope <- slope * 60 # cells per mL per hour
  SLOPE.loop$Number <- gsub("^(:[^_+]{7}([^_+]).*", "\\1", loop_930[i,])
  SLOPE.loop$SLOPE <- SLOPE
  SLOPE.loop$pval <- pval
```

```
# Save plots ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
```

```
pdf(paste0("C:/Users/samjg/Documents/Github_repositories/Airradians_OA/RAnalysis/Output/FeedingRates/FL",
ggarrange(SlopePlots_914, SlopeFig914_geombox,
          labels = c("A", "B"),
          ncol = 1, nrow = 2)
```

```
## 'geom_smooth()' using formula 'y ~ x'
```

```
dev.off()
```

```
## pdf
```

```
## 2
```

```
pdf(paste0("C:/Users/samjg/Documents/Github_repositories/Airradians_OA/RAnalysis/Output/FeedingRates/FL",
ggarrange(SlopePlots_930, SlopeFig930_geombox,
          labels = c("A", "B"),
          ncol = 1, nrow = 2)
```

```
## 'geom_smooth()' using formula 'y ~ x'
```

```
dev.off()
```

```
## pdf
```

```
## 2
```


Calculate Clearance Rate

Note: the following clusters calculate CR for the start/end Algae counts (i.e. 20210914 time 0 and time 90 minutes; 20210930 time 0 and time 60 minutes)

CLEARANCE RATE EQUATION

$$FR = (V/t * (\ln(C_0/C_t) - A)) / L$$

- **V** == the volume of the vessel
- **t** == time of the trial interval (i.e. 60 minutes or 1 hour)
- **ln(C₀/C_t)** == ratio of the live algae concentration (cells ml⁻¹) at time 0 (C₀) and at the elapsed time interval(s) (C_t) - take the natural log of this number
- **A** == ln(C₀/C_t) for the 'blank' values in each treatment, accounts for the sink or stuck algae cells
- **L** == normalization factor between individuals - here we will use the shell length in mm

'A' Calculate the blank values for each trial

'summarise()' has grouped output by 'pH'. You can override using the '.groups' argument.

```
##      pH Time._min meanBlank_ln_AlgaeLoss sdBlank_ln_AlgaeLoss
## 1  7.5      10      0.005925943      NA
## 2  7.5      15      0.008379498      0.001565165
## 3  7.5      20      0.083124754      NA
## 4  7.5      30      0.066622217      0.034126783
## 5  7.5      40      0.065622318      NA
## 6  7.5      50      0.094424309      NA
## 7  7.5      57      0.059777204      NA
## 8  7.5      60      0.098939948      NA
## 9  7.5      70      0.101756318      NA
## 10 7.5      75      0.075223421      NA
## 11 7.5      90      0.089593318      0.053841127
## 12 8.0      10      0.015870864      0.001415206
## 13 8.0      15      0.081890972      NA
## 14 8.0      30      0.112614144      NA
## 15 8.0      40      0.067635835      NA
## 16 8.0      45      0.076359279      0.067200682
## 17 8.0      47      0.046063258      NA
## 18 8.0      50      0.040346822      NA
## 19 8.0      57      0.044518856      NA
## 20 8.0      60      0.158305235      NA
## 21 8.0      70      0.024924890      0.015757351
## 22 8.0      75      0.112731849      NA
## 23 8.0      90      0.068248862      0.056330183
##      seBlank_ln_AlgaeLoss n
## 1              NA 1
## 2      0.001106739 2
## 3              NA 1
## 4      0.019703108 3
## 5              NA 1
## 6              NA 1
## 7              NA 1
```

```
## 8          NA 1
## 9          NA 1
## 10         NA 1
## 11      0.031085189 3
## 12      0.001000702 2
## 13         NA 1
## 14         NA 1
## 15         NA 1
## 16      0.047518058 2
## 17         NA 1
## 18         NA 1
## 19         NA 1
## 20         NA 1
## 21      0.011142130 2
## 22         NA 1
## 23      0.028165092 4
```

'summarise()' has grouped output by 'pH'. You can override using the '.groups' argument.

```
##    pH Time._min meanBlank_ln_AlgaeLoss sdBlank_ln_AlgaeLoss
## 1 7.5      20      0.07899175      0.09373166
## 2 7.5      30      0.01338708      NA
## 3 7.5      40      0.14527004      NA
## 4 7.5      60      0.06080047      0.07900592
## 5 8.0      20      0.06528042      0.03669390
## 6 8.0      30      0.04766821      0.00226676
## 7 8.0      40      0.12783337      NA
## 8 8.0      60      0.06835669      0.03782168
##    seBlank_ln_AlgaeLoss n
## 1      0.066278294 2
## 2          NA 1
## 3          NA 1
## 4      0.045614090 3
## 5      0.021185232 3
## 6      0.001602841 2
## 7          NA 1
## 8      0.021836358 3
```

‘A’ Call the blank values for each trial - continued...

- Call the blank value **specifically for the final timepoint** of the clearance rate trial(s)!
- this value will be called in the for loop when calculating CR in following cluster(s)

```
# 20210914 DATA ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
Blank_914_pH7.5 <- (A_914_BlankMeans %>% dplyr::filter(pH == 7.5) %>% dplyr::arrange(desc(Time._min)))$
Blank_914_pH8.0 <- (A_914_BlankMeans %>% dplyr::filter(pH == 8.0) %>% dplyr::arrange(desc(Time._min)))$

# 20210930 DATA ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
Blank_930_pH7.5 <- (A_930_BlankMeans %>% dplyr::filter(pH == 7.5) %>% dplyr::arrange(desc(Time._min)))$
Blank_930_pH8.0 <- (A_930_BlankMeans %>% dplyr::filter(pH == 8.0) %>% dplyr::arrange(desc(Time._min)))$
```

20210914 Clearance Rate data

```

df_total.914 <- data.frame() # start dataframe
for (i in 1:nrow(loop_914)) {
  dat <- ClearRate_Master.914 %>%
    dplyr::filter(unique_Identifier == loop_914[i,]) %>%
    dplyr::arrange(Time._min)
  C0 <- (dat %>% dplyr::filter(Time._min == 0))$Cells_ml[1]
  dat2 <- dat %>%
    #dplyr::mutate(diff = as.numeric(Time._min) - lag(as.numeric(Time._min), default = first(as.numeric
    dplyr::filter(Time._min %in% 90) %>%
    dplyr::mutate(Blank = if(pH == 7.5) Blank_914_pH7.5 else Blank_914_pH8.0 ) %>%
    dplyr::mutate(AlgaeLossRatio = C0 / as.numeric(Cells_ml) ) %>%
    dplyr::filter(!AlgaeLossRatio < 1) %>%
    dplyr::mutate(ln_AlgaeLossRatio = ln(AlgaeLossRatio)) %>%
    # dplyr::mutate(ClearanceRate = ( (25/1000) / (diff/60) * # V / t == Volume of the vessel (in Lite
    # ( ln_AlgaeLossRatio ) / Length.um. ) ) %>%
    dplyr::mutate(ClearanceRate_L_hour_mm = ( (25/1000) * # V / t == Volume of the vessel (in Liters as
    (( ln_AlgaeLossRatio / (as.numeric(Time._min)/60) - Blank))) /
    dplyr::mutate(Time_period = paste((as.numeric(substr(Time._min,1,1)) -1), "0-", Time._min, sep = ''))
    dat2OM <- dat2 %>% dplyr::filter(!Time._min == 0)

  if (nrow(dat2OM) > 0) {
    ClearRate.table <- data.frame(matrix(nrow = nrow(dat2OM), ncol = 9)) # create dataframe t
    colnames(ClearRate.table) <- c('Date', 'ID', 'pH', 'Replicate', 'Num', 'Run', 'Time_period', 'AlgaeLo

    ClearRate.table$Date <- dat2OM$Date
    ClearRate.table$ID <- loop_914[i,]
    ClearRate.table$pH <- gsub("_.*", "\\1", ClearRate.table$ID)
    ClearRate.table$Replicate <- gsub("^(?:[^\_]+\_){4}([^\_]+).*", "\\1", ClearRate.table
    ClearRate.table$Num <- gsub("^(?:[^\_]+\_){6}([^\_]+).*", "\\1", ClearRate.table
    ClearRate.table$Run <- gsub("^(?:[^\_]+\_){2}([^\_]+).*", "\\1", ClearRate.table
    ClearRate.table$Time_period <- paste((as.numeric(substr(dat2OM$Time._min,1,1)) -1),
    ClearRate.table$AlgaeLossRatio <- dat2OM$AlgaeLossRatio
    ClearRate.table$ClearanceRate_L_hour_mm <- dat2OM$ClearanceRate_L_hour_mm

    df <- data.frame(ClearRate.table) # name dataframe for this single row
    df_total.914 <- rbind(df_total.914,df) #bind to a cumulative list dataframe
    #print(df_total.914) # print to monitor progress
  }
  else {}
}

ClearRates_914_Means <- df_total.914 %>%
  #dplyr::filter(!ClearanceRate_L_hour_meter %in% '-Inf') %>%
  dplyr::group_by(pH) %>%
  dplyr::summarise(
    meanCR = mean(ClearanceRate_L_hour_mm),
    sdCR = sd(ClearanceRate_L_hour_mm),
    seCR = sd(ClearanceRate_L_hour_mm) / sqrt(length(ClearanceRate_L_hour_mm)),
    n = n()) %>%
  na.omit()

```

```
print(ClearRates_914_Means)
```

```
## # A tibble: 2 x 5
##   pH      meanCR      sdCR      seCR      n
##   <chr>    <dbl>    <dbl>    <dbl> <int>
## 1 7.5     0.00378 0.00288 0.000831    12
## 2 8       0.00236 0.00170 0.000490    12
```

```
# summary(lmer(meanCR~pH+ (1|Time_period), data=ClearRates_914_Means))
# summary(aov(lm(meanCR~pH*Time_period, data=ClearRates_914_Means)))
```

```
mod914CR <- aov(lm(ClearanceRate_L_hour_mm~ pH , data = (df_total.914 %>% dplyr::filter(!ClearanceRate.
pander(summary(mod914CR), style='markdown')
```

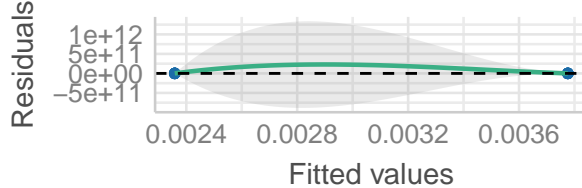
Table 1: Analysis of Variance Model

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
pH	1	1.206e-05	1.206e-05	2.158	0.156
Residuals	22	0.0001229	5.588e-06	NA	NA

```
check_model(mod914CR) # observe the diagnostics of the model
```

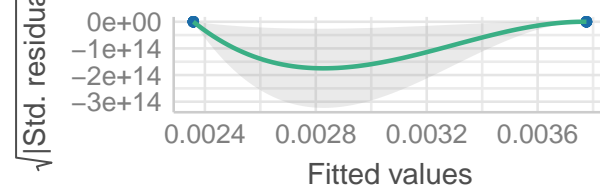
Linearity

Reference line should be flat and horizontal



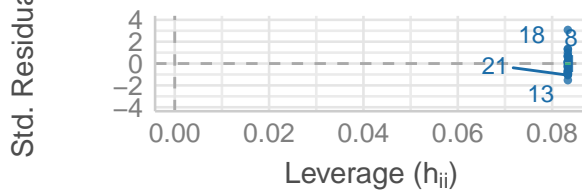
Homogeneity of Variance

Reference line should be flat and horizontal



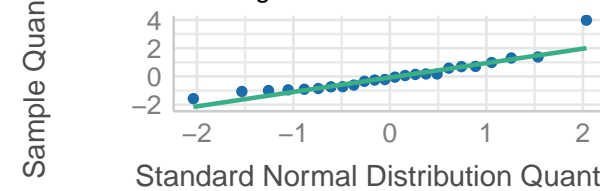
Influential Observations

Points should be inside the contour lines



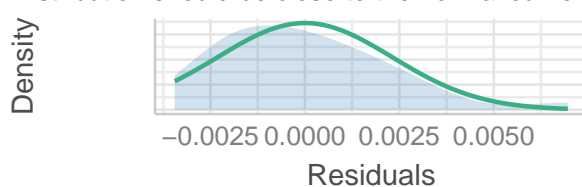
Normality of Residuals

Dots should fall along the line



Normality of Residuals

Distribution should be close to the normal curve



```
shapiro.test(residuals(mod914CR)) # non normal
```

```
##  
## Shapiro-Wilk normality test  
##  
## data: residuals(mod914CR)  
## W = 0.92006, p-value = 0.05859
```

```
leveneTest(mod914CR) # good
```

```
## Warning in leveneTest.default(y = y, group = group, ...): group coerced to  
## factor.
```

```
## Levene's Test for Homogeneity of Variance (center = median)  
##      Df F value Pr(>F)  
## group 1  1.2472 0.2761  
##      22
```

```
#summary(lmer(ClearanceRate_L_hour_mm~ pH + (1|Replicate), data = (df_total.914 %>% dplyr::filter(!Cle
```

```
CR_914_barplot <- ClearRates_914_Means %>%  
  #dplyr::filter(!Time_period %in% c('40-50', '50-60')) %>%  
  ggplot(aes(x=pH , y=meanCR, fill = pH)) +  
  geom_bar(position=position_dodge(), aes(y=meanCR), stat="identity", alpha=0.5) +  
  scale_fill_manual("Treatment", values = c("8" = "grey50", "7.5" = "black")) +  
  geom_errorbar(position=position_dodge(width=0.9), aes(ymin=meanCR+seCR, ymax=meanCR+seCR), width=0.2,  
  geom_linerange(aes(ymin = meanCR, ymax = meanCR+seCR)) +  
  geom_point(position=position_dodge(width=0.9), aes(y=meanCR)) +  
  theme_classic() +  
  ggtitle("Clearance Rate, F1 Scallops 20210914 (0-90 minutes)") # +  
  # facet_wrap(~Time_period)  
CR_914_barplot
```

A bar chart comparing the meanCR (y-axis) across two pH levels (7.5 and 8, x-axis) for two treatments. The y-axis ranges from 0.000 to 0.004. The x-axis is labeled 'pH' and has two categories: 7.5 and 8. The legend, titled 'Treatment', shows that the dark gray bar represents Treatment 7.5 and the light gray bar represents Treatment 8. Error bars are present on top of each bar, indicating variability or confidence intervals.

pH	Treatment	meanCR (approx.)
7.5	7.5 (Dark Gray)	0.0037
8	8 (Light Gray)	0.0023

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A box plot comparing the ClearanceRate_L_hour_mm for two pH levels: 7.5 and 8. The y-axis represents the ClearanceRate_L_hour_mm, ranging from 0.000 to 0.009. The x-axis is labeled 'pH'. The legend indicates that pH 7.5 is represented by black dots and pH 8 is represented by grey dots. The box plot for pH 7.5 shows a median around 0.003, with a range from approximately 0.0015 to 0.0055. The box plot for pH 8 shows a median around 0.002, with a range from approximately 0.0005 to 0.003. Individual data points are overlaid on the box plots.

pH	ClearanceRate_L_hour_mm
7.5	0.0003
7.5	0.0012
7.5	0.0015
7.5	0.0018
7.5	0.0020
7.5	0.0022
7.5	0.0030
7.5	0.0031
7.5	0.0035
7.5	0.0055
7.5	0.0068
7.5	0.0105
8	0.0005
8	0.0006
8	0.0008
8	0.0010
8	0.0012
8	0.0018
8	0.0022
8	0.0023
8	0.0024
8	0.0028
8	0.0055

```
## pdf
## 2
```

```
df_total.930 <- data.frame() # start dataframe
for (i in 1:nrow(loop_930)) {
  dat <- ClearRate_Master.930 %>%
    dplyr::filter(uniq_Identifier == loop_930[i,]) %>%
    dplyr::arrange(Time._min)
  C0 <- (dat %>% dplyr::filter(Time._min == 0))$Cells_ml[1]
  dat2 <- dat %>%
    #dplyr::mutate(diff = as.numeric(Time._min) - lag(as.numeric(Time._min), default = first(as.numeric
    dplyr::filter(Time._min %in% 60) %>%
    dplyr::mutate(Blank = if(pH == 7.5) Blank_930_pH7.5 else Blank_930_pH8.0 ) %>%
```

```

dplyr::mutate(AlgaeLossRatio = C0 / as.numeric(Cells_ml) ) %>%
dplyr::filter(!AlgaeLossRatio < 1) %>%
dplyr::mutate(ln_AlgaeLossRatio = ln(AlgaeLossRatio)) %>%
# dplyr::mutate(ClearanceRate = ( (25/1000) / (diff/60) ) * # V / t == Volume of the vessel (in Lit
# ( ln_AlgaeLossRatio ) / Length.um. ) ) %>%
dplyr::mutate(ClearanceRate_L_hour_mm = ( (25/1000) * # V / t == Volume of the vessel (in Liters as
(( ln_AlgaeLossRatio / (as.numeric(Time._min)/60) - Bl
dplyr::mutate(Time_period = paste((as.numeric(substr(Time._min,1,1)) -1), "0-", Time._min, sep = ''))
# dat20M <- dat2 %>% dplyr::filter(!Time._min == 0)

if (nrow(dat2) > 0) {
ClearRate.table <- data.frame(matrix(nrow = nrow(dat2), ncol = 10)) # create dataframe to
colnames(ClearRate.table) <- c('Date', 'ID', 'pH', 'Fed_Unfed', 'Replicate', 'Num', 'Run', 'Time_peri

ClearRate.table$Date <- dat2$Date
ClearRate.table$ID <- loop_930[i,]
ClearRate.table$pH <- gsub("_.*", "\\1", ClearRate.table$ID)
ClearRate.table$Fed_Unfed <- gsub("(?:[^\_]+\_){1}([^\_]+).*", "\\1", ClearRate.table
ClearRate.table$Replicate <- gsub("(?:[^\_]+\_){5}([^\_]+).*", "\\1", ClearRate.table
ClearRate.table$Num <- gsub("(?:[^\_]+\_){7}([^\_]+).*", "\\1", ClearRate.table
ClearRate.table$Run <- gsub("(?:[^\_]+\_){3}([^\_]+).*", "\\1", ClearRate.table
ClearRate.table$Time_period <- paste((as.numeric(substr(dat2$Time._min,1,1)) -1), "0
ClearRate.table$AlgaeLossRatio <- dat2$AlgaeLossRatio
ClearRate.table$ClearanceRate_L_hour_mm <- dat2$ClearanceRate_L_hour_mm

df <- data.frame(ClearRate.table) # name dataframe for this single row
df_total.930 <- rbind(df_total.930,df) #bind to a cumulative list dataframe
#print(df_total.930) # print to monitor progress
}
else {}
}

```

```

## Warning in if (pH == 7.5) Blank_930_pH7.5 else Blank_930_pH8.0: the condition
## has length > 1 and only the first element will be used

```

```

## Warning in if (pH == 7.5) Blank_930_pH7.5 else Blank_930_pH8.0: the condition
## has length > 1 and only the first element will be used

```

```

ClearRates_930_Means <- df_total.930 %>%
dplyr::filter(!ClearanceRate_L_hour_mm < 0) %>%
dplyr::group_by(pH, Fed_Unfed, Time_period) %>%
dplyr::summarise(
  meanCR = mean(ClearanceRate_L_hour_mm),
  sdCR = sd(ClearanceRate_L_hour_mm),
  seCR = sd(ClearanceRate_L_hour_mm) / sqrt(length(ClearanceRate_L_hour_mm)),
  n = n()) %>%
na.omit()

```

```

## 'summarise()' has grouped output by 'pH', 'Fed_Unfed'. You can override using the '.groups' argument

```

```

ClearRates_930_Means

```



```
## # A tibble: 4 x 7
## # Groups:   pH, Fed_Unfed [4]
##   pH    Fed_Unfed Time_period meanCR    sdCR    seCR    n
##   <chr> <chr>      <chr>      <dbl>    <dbl>    <dbl> <int>
## 1 7.5    fed        50-60      0.00364 0.00339 0.000846 16
## 2 7.5    unfed      50-60      0.00142 0.000604 0.000201 9
## 3 8      fed        50-60      0.00480 0.00383 0.000903 18
## 4 8      unfed      50-60      0.00204 0.00146 0.000334 19

# summary(lmer(meanCR~pH*Fed_Unfed + (1/Time_period), data=ClearRates_930_Means))
# summary(aov(lm(meanCR~pH*Fed_Unfed, data=ClearRates_930_Means)))

mod930CR <- aov(lm(ClearanceRate_L_hour_mm~ pH * Fed_Unfed, data = (df_total.930 %>% dplyr::filter(!CL
pander(summary(mod930CR), style='markdown')
```

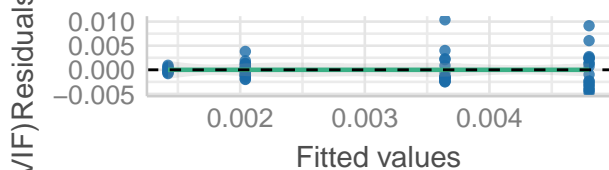
Table 2: Analysis of Variance Model

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
pH	1	4.353e-06	4.353e-06	0.5458	0.463
Fed_Unfed	1	9.774e-05	9.774e-05	12.26	0.0008984
pH:Fed_Unfed	1	1.018e-06	1.018e-06	0.1277	0.7222
Residuals	58	0.0004625	7.974e-06	NA	NA

```
check_model(mod930CR) # observe the diagnostics of the model
```

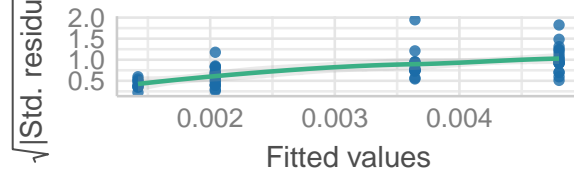
Linearity

Reference line should be flat and horizontal



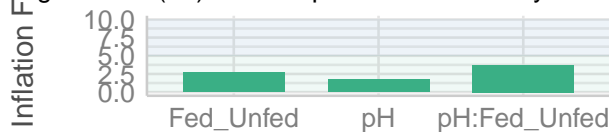
Homogeneity of Variance

Reference line should be flat and horizontal



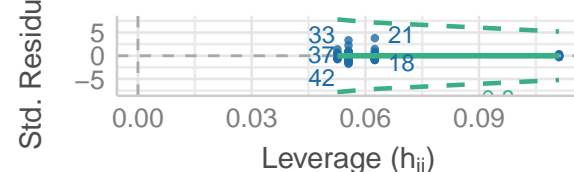
Collinearity

Higher bars (>5) indicate potential collinearity issues



Influential Observations

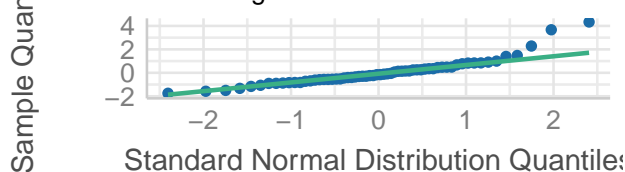
Points should be inside the contour lines



low (< 5) moderate (< 10) high (> 10)

Normality of Residuals

Points should fall along the line



Normality of Residuals

Distribution should be close to the normal curve



```
shapiro.test(residuals(mod930CR)) # non normal
```

```
##  
## Shapiro-Wilk normality test  
##  
## data: residuals(mod930CR)  
## W = 0.89661, p-value = 7.692e-05
```

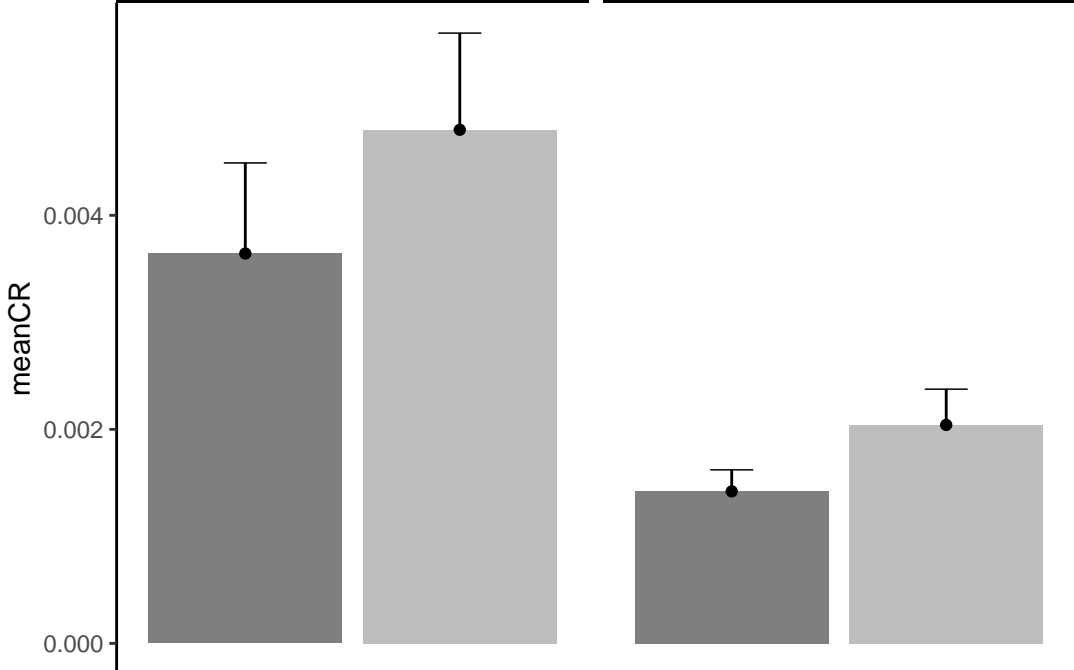
```
leveneTest(mod930CR) # good
```

```
## Levene's Test for Homogeneity of Variance (center = median)  
##      Df F value Pr(>F)  
## group 3  4.7854 0.00478 **  
##      58  
## ---  
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
# summary(lmer(ClearanceRate_L_hour_mm~ pH * Fed_Unfed + (1/Replicate), data = (df_total.930 %>% dplyr
```

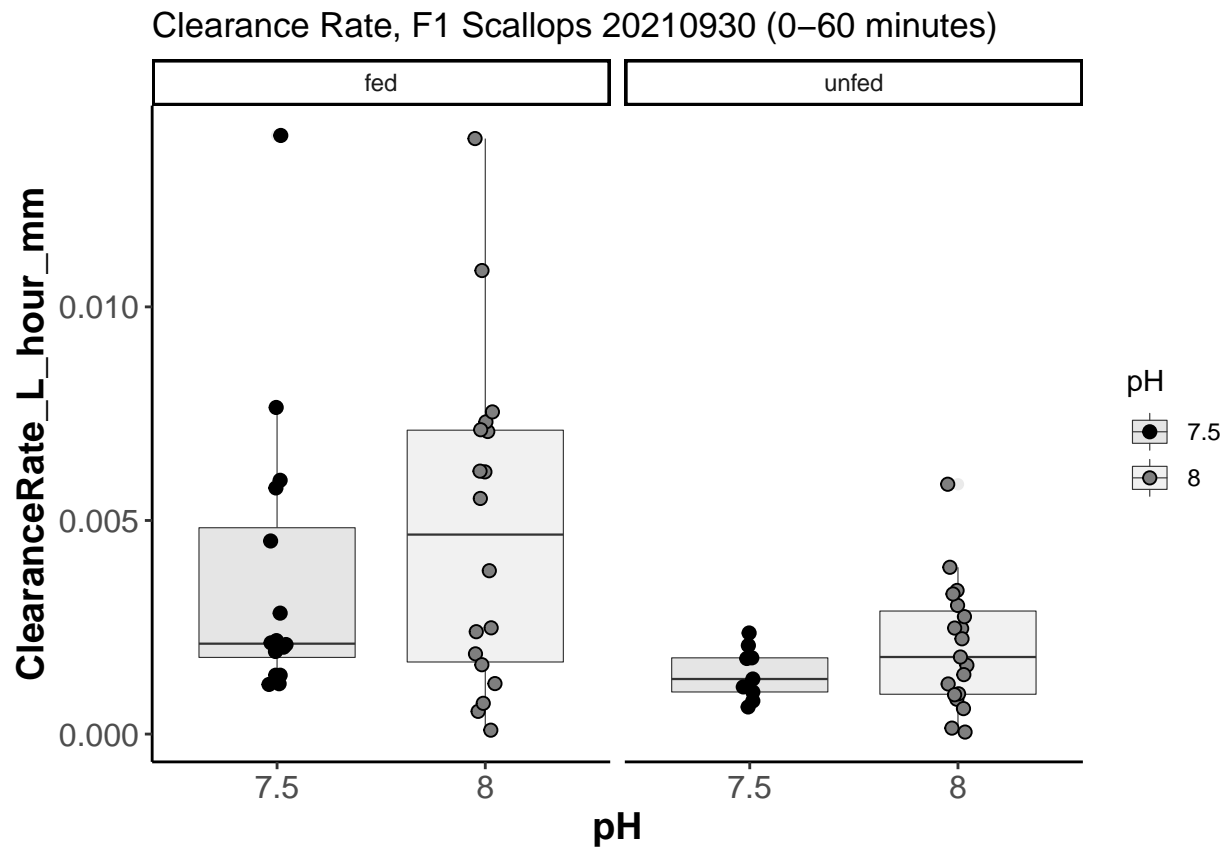
```
ClearRates_930_Means$pH_feed <- paste(ClearRates_930_Means$pH, ClearRates_930_Means$Fed_Unfed, sep='_')  
CR_930_boxplot <- ClearRates_930_Means %>%  
#dplyr::filter(!Time_period %in% c('40-50', '50-60')) %>%  
ggplot(aes(x=pH , y=meanCR, fill = pH)) +  
  geom_bar(position=position_dodge(), aes(y=meanCR), stat="identity", alpha=0.5) +  
  scale_fill_manual(values=c("black", "grey50")) +  
  geom_errorbar(position=position_dodge(width=0.9), aes(ymin=meanCR+seCR, ymax=meanCR+seCR), width=0.2,  
  geom_linerange(aes(ymin = meanCR, ymax = meanCR+seCR)) +  
  geom_point(position=position_dodge(width=0.9), aes(y=meanCR)) +  
  theme_classic() +  
  ggtitle("Clearance Rate, F1 Scallops 20210930 (0-60 minutes)") +  
  facet_wrap(~ Fed_Unfed)  
CR_930_boxplot
```

fed	unfed
-----	-------



```
CR_930_barplot <- df_totals.930 %>%
  dplyr::filter(!ClearanceRate_L_hour_mm < 0) %>%
  #dplyr::filter(!ClearanceRate_L_hour_mm %in% '-Inf') %>%
  #dplyr::filter(!Time_period %in% c('40-50', '50-60')) %>%
  ggplot(aes(pH , ClearanceRate_L_hour_mm , fill = pH)) +
  theme(panel.grid=element_blank()) +
  geom_boxplot(size=0.2, alpha=0.1, aes(fill=pH)) +
  scale_fill_manual(values=c("black", "grey50")) +
  geom_point(shape = 21, size = 2, position = position_jitterdodge(jitter.width = 0.1)) +
  theme_classic() +
  theme(axis.text=element_text(size=12),
        axis.title=element_text(size=14,face="bold")) +
  # labs(title = "F1 Scallops: Slope algae cells/time - feeding rate trials 20210930",
  #       y = expression(Slope~"="~absolute~value~"("~Live~algae~cells~mL^{-1}~hour^{-1}~")"),
  #       x = expression(italic(p)*CO[2]~Treatment~"("~mu*atm~")")) +
  ggtitle("Clearance Rate, F1 Scallops 20210930 (0-60 minutes)") +
  facet_wrap(~ Fed_Unfed)
```

CR_930_barplot



```
# Save plots ::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::::
pdf(paste0("C:/Users/samjg/Documents/Github_repositories/Airradiations_OA/RAnalysis/Output/FeedingRates/CL
ggarrange(CR_930_barplot, CR_930_boxplot,
  labels = c("A", "B"),
  ncol = 1, nrow = 2)
dev.off()
```

```
## pdf
## 2
```