

Brief overview

Biological Rationale

Resources & Timeline

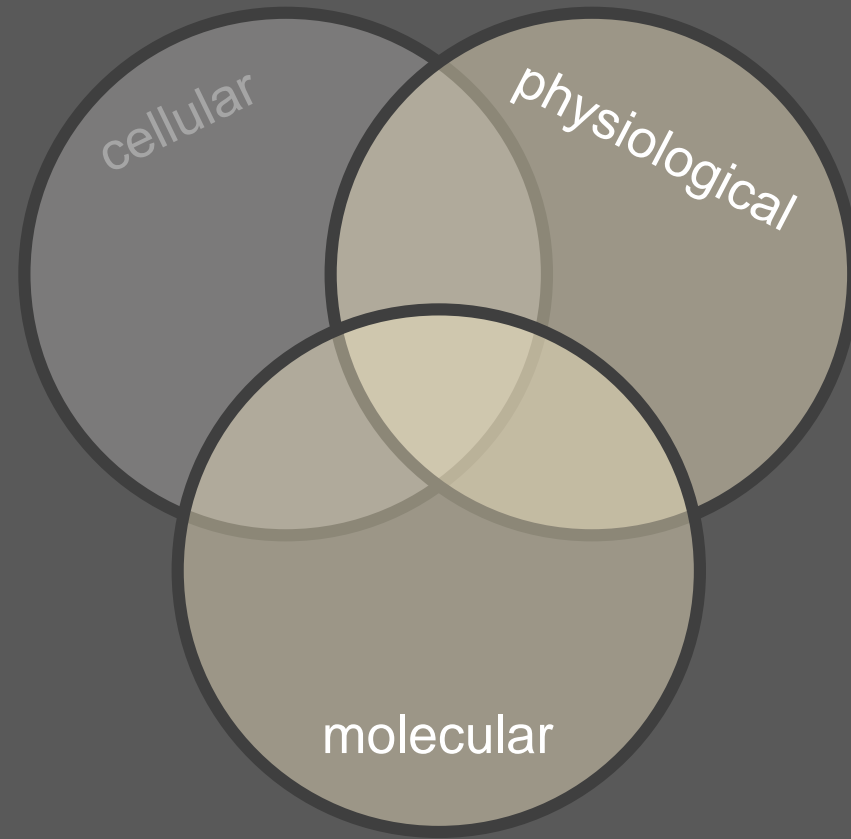
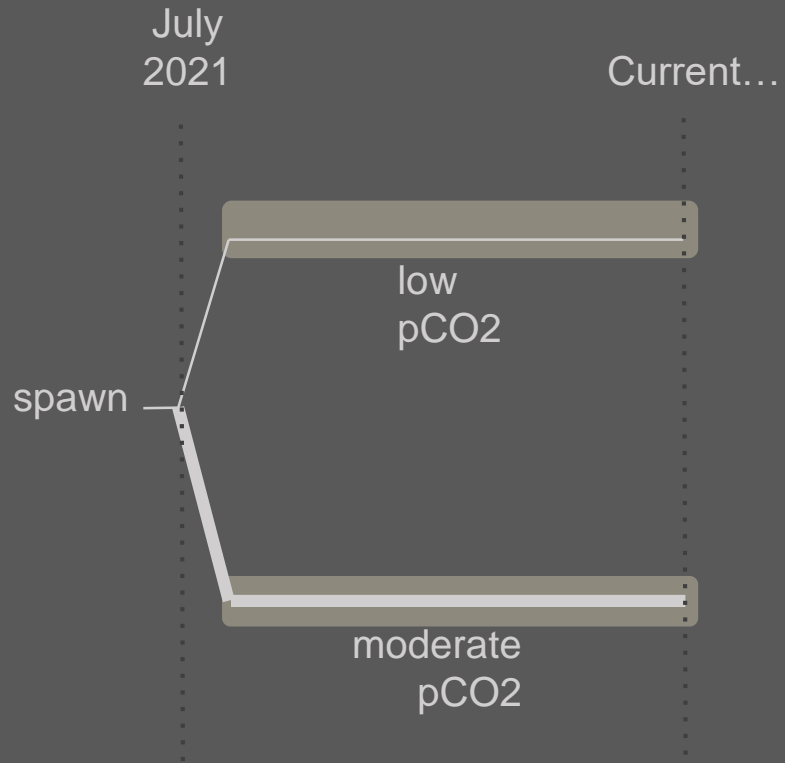
by Sam Gurr

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Our ongoing scallop experiment...



Suite of physiological measurements

- growth/survivorship
- respiration rate
- clearance rate
- biodepositon
- excretion

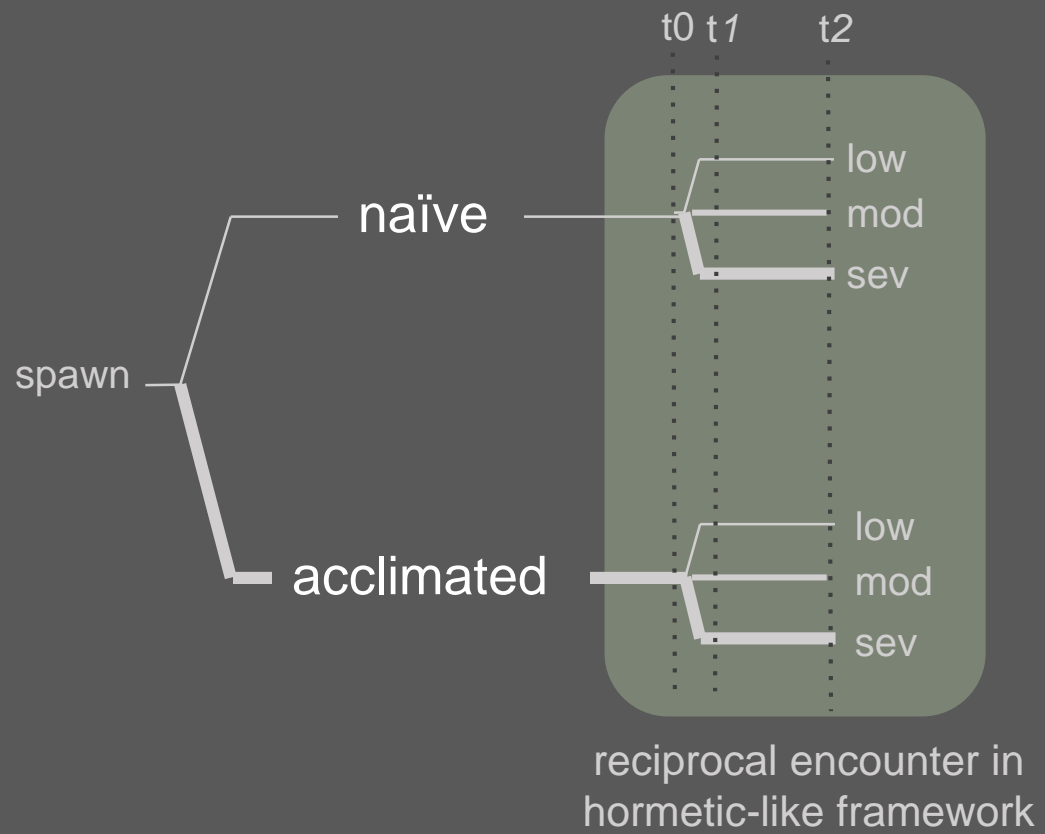
genetic variation

- allele frequency
- putatively adaptive loci

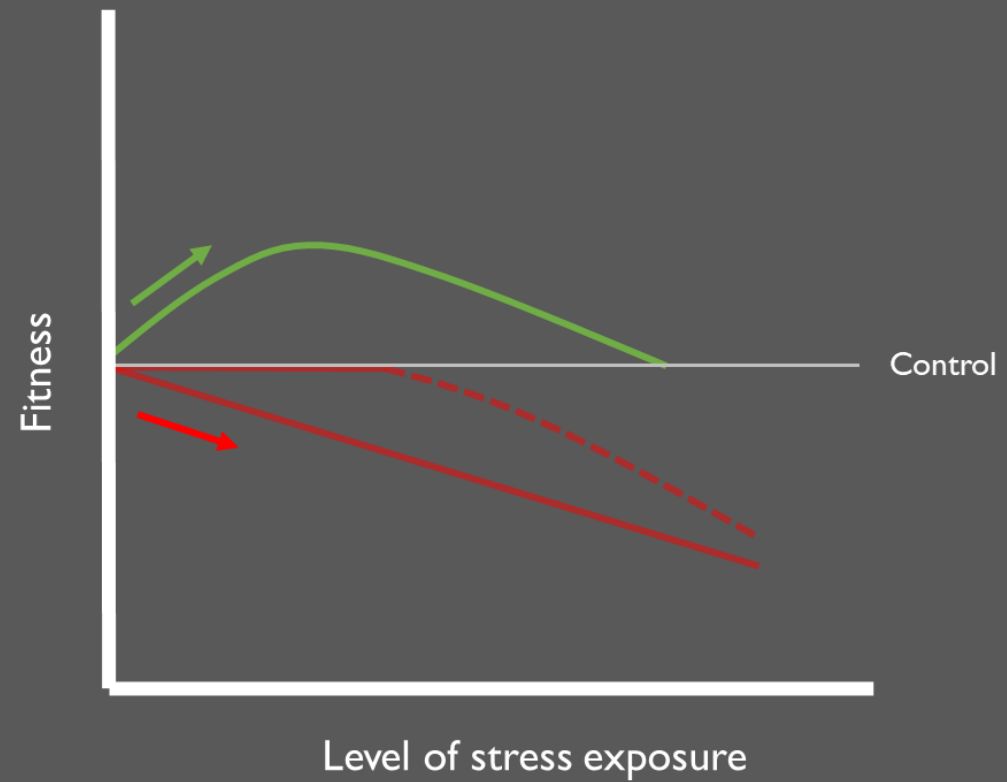
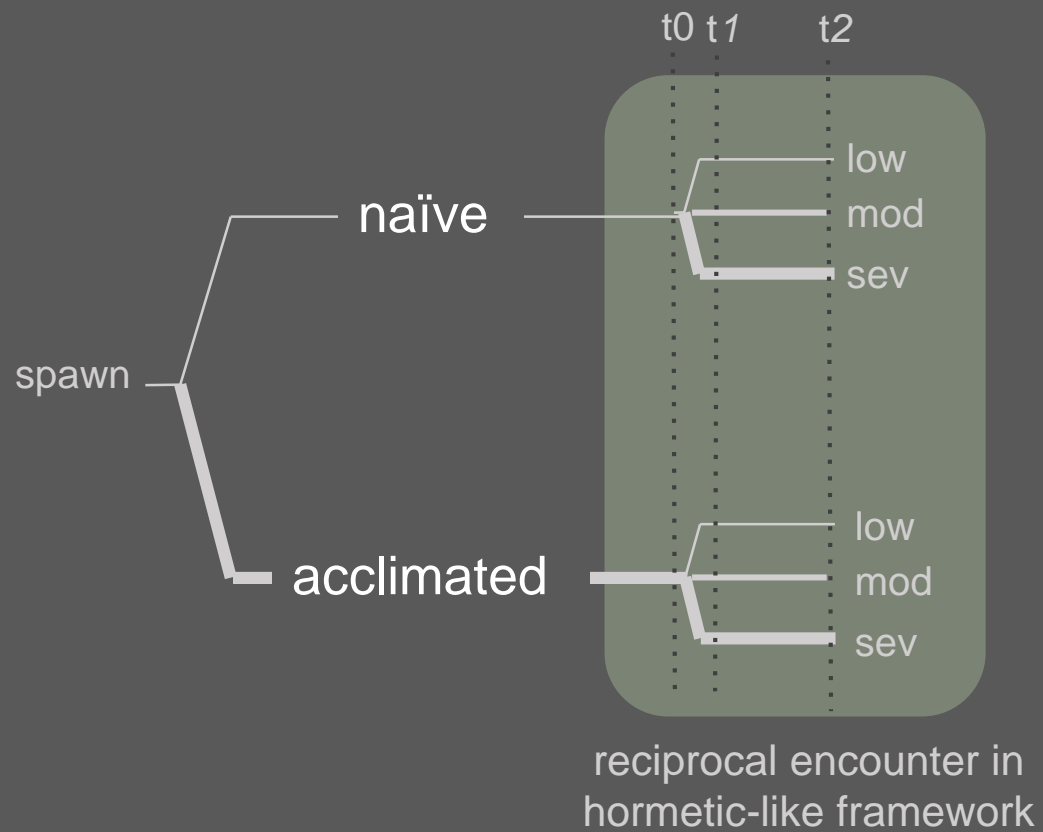
gene expression

- enriched functions/pathways
- differentially-expressed genes

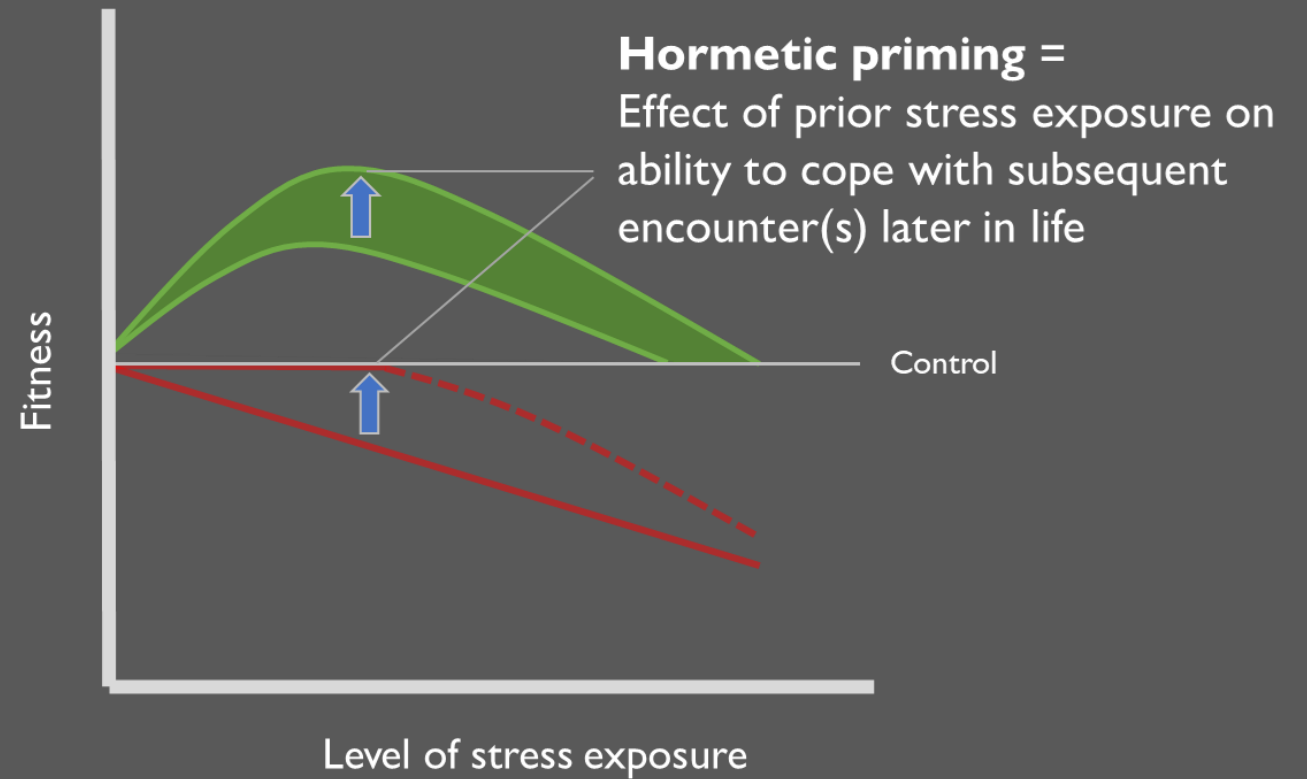
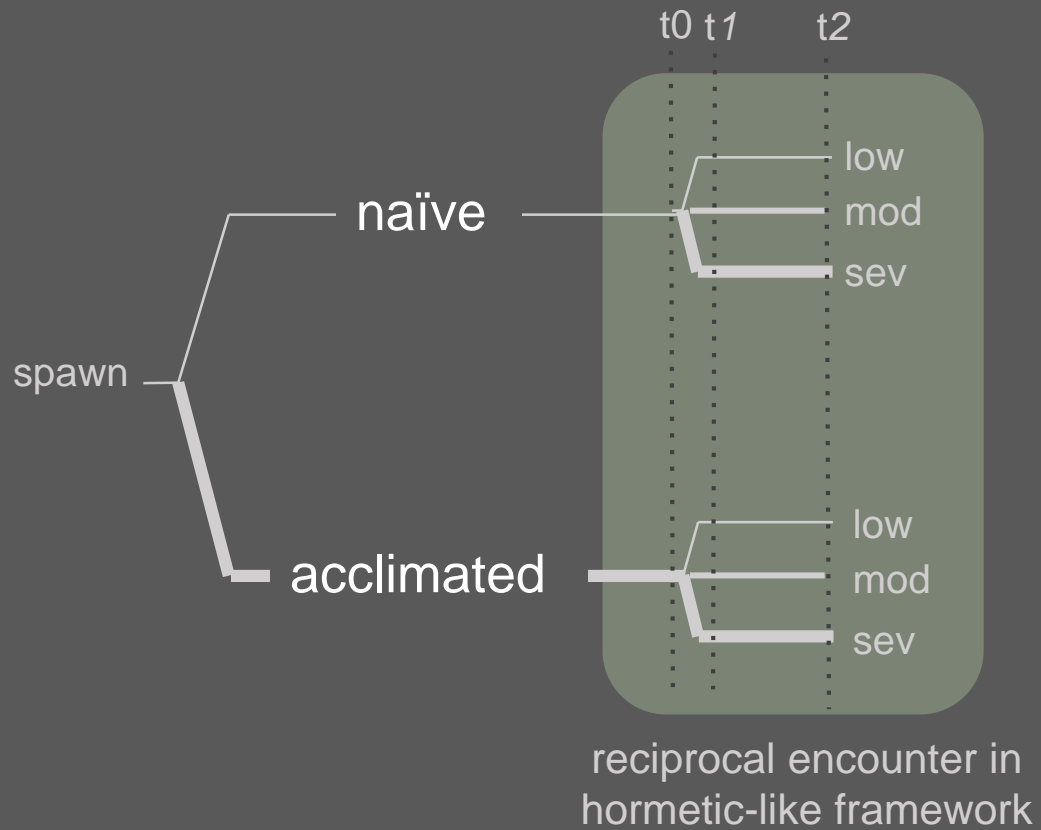
Understand cellular mechanisms of OA response



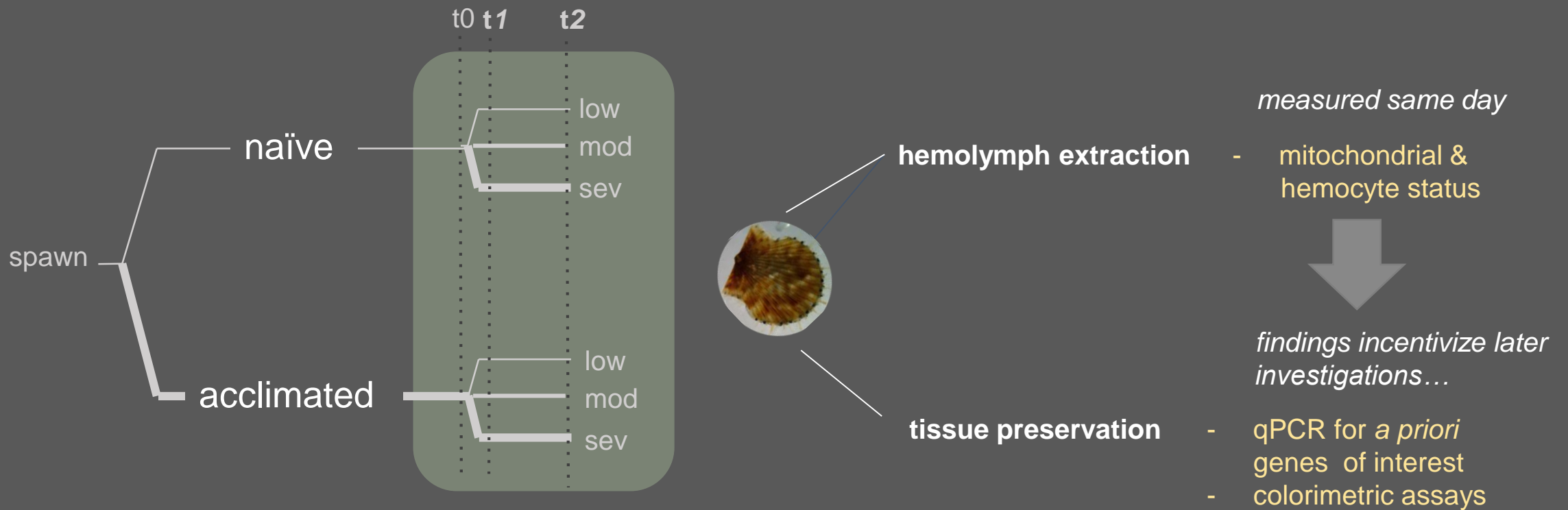
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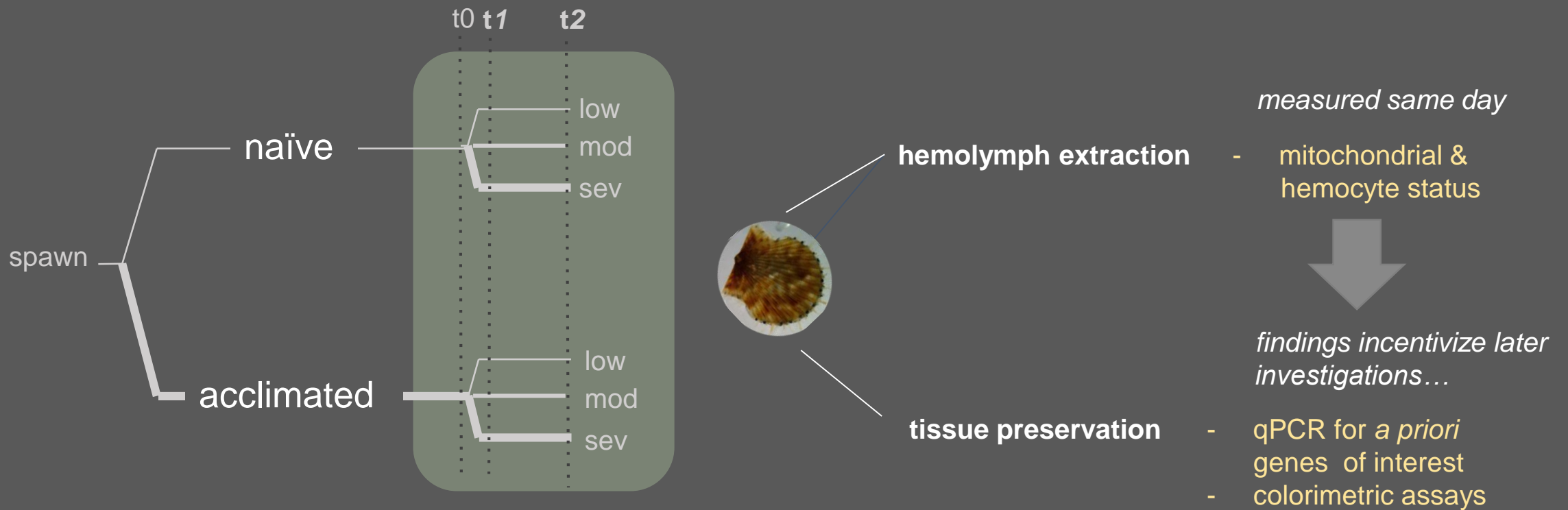
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Environmental sensor

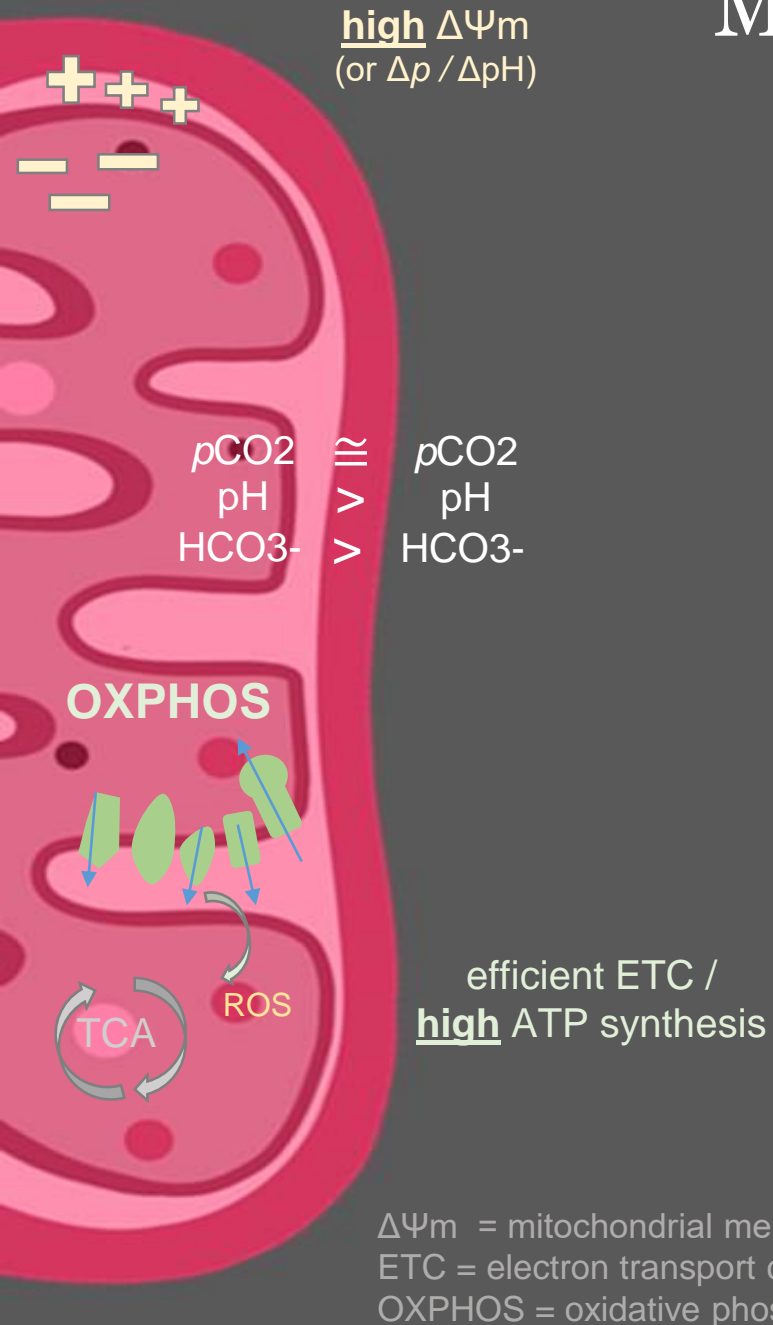
Mitochondrial dysfunction & alternative pathways

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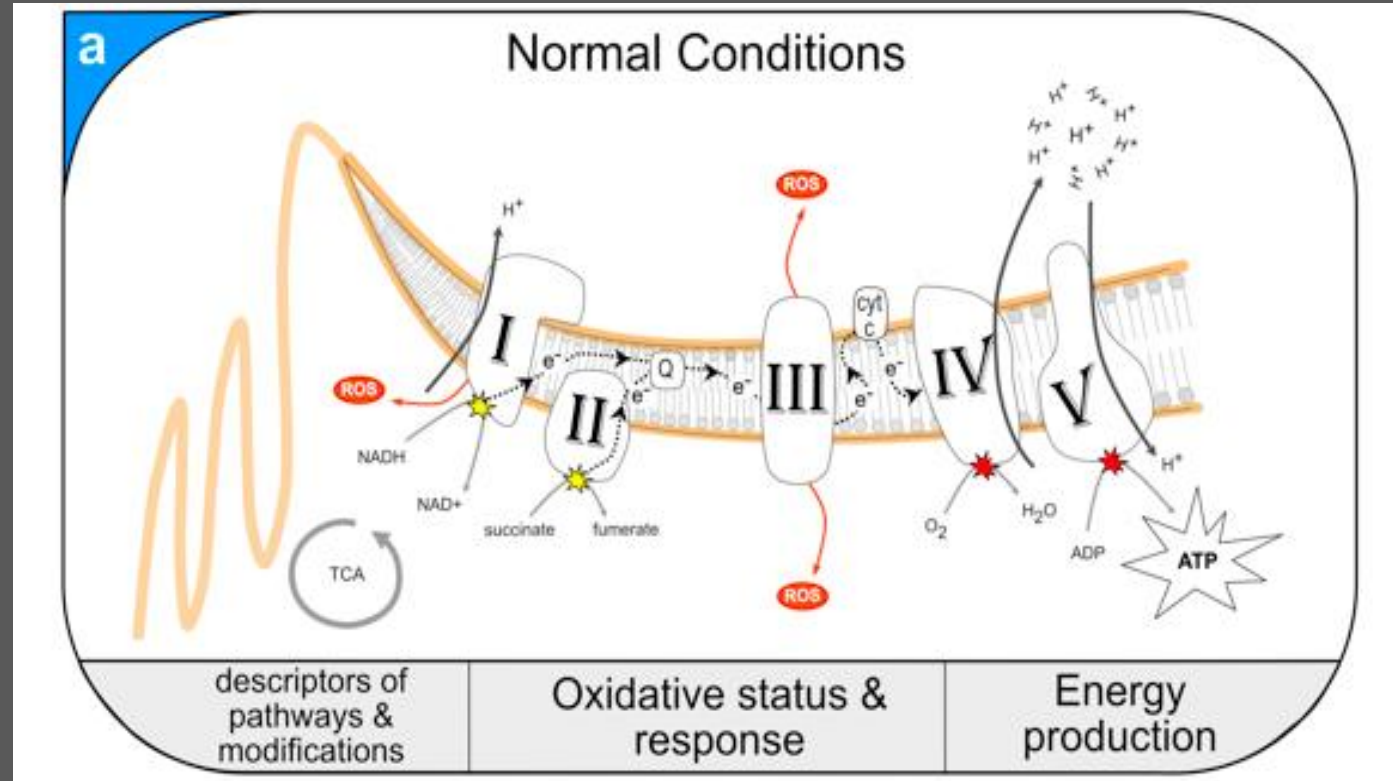
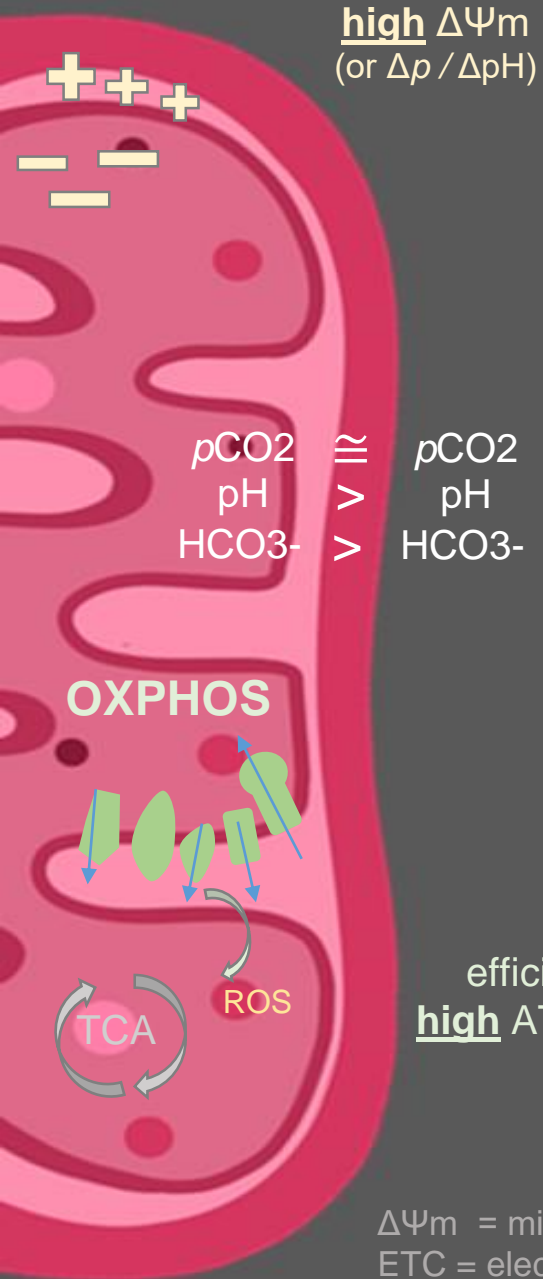
Mitochondria under normal/optimal conditions



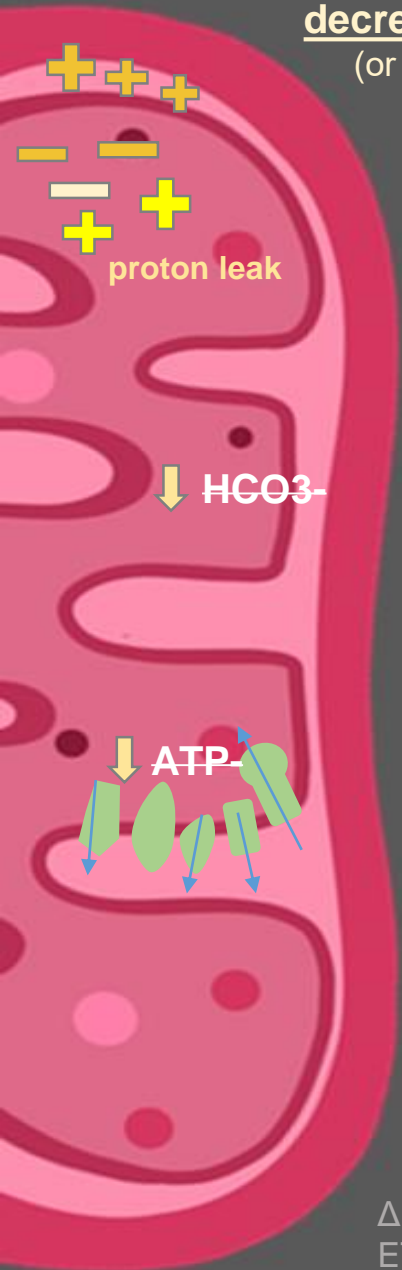
‘Power house’ of the cell

- electropositive gradient drives ATP synthesis (+) intermembrane space vs (-) inner matrix
- main source of metabolically-produced CO_2
 - high HCO_3^- in mitochondria acts as a buffer, inner mitochondrial pH typically $>$ cytosol

Mitochondria under normal/optimal conditions



Mitochondrial dysfunction under OA conditions



hypercapnia/acidosis

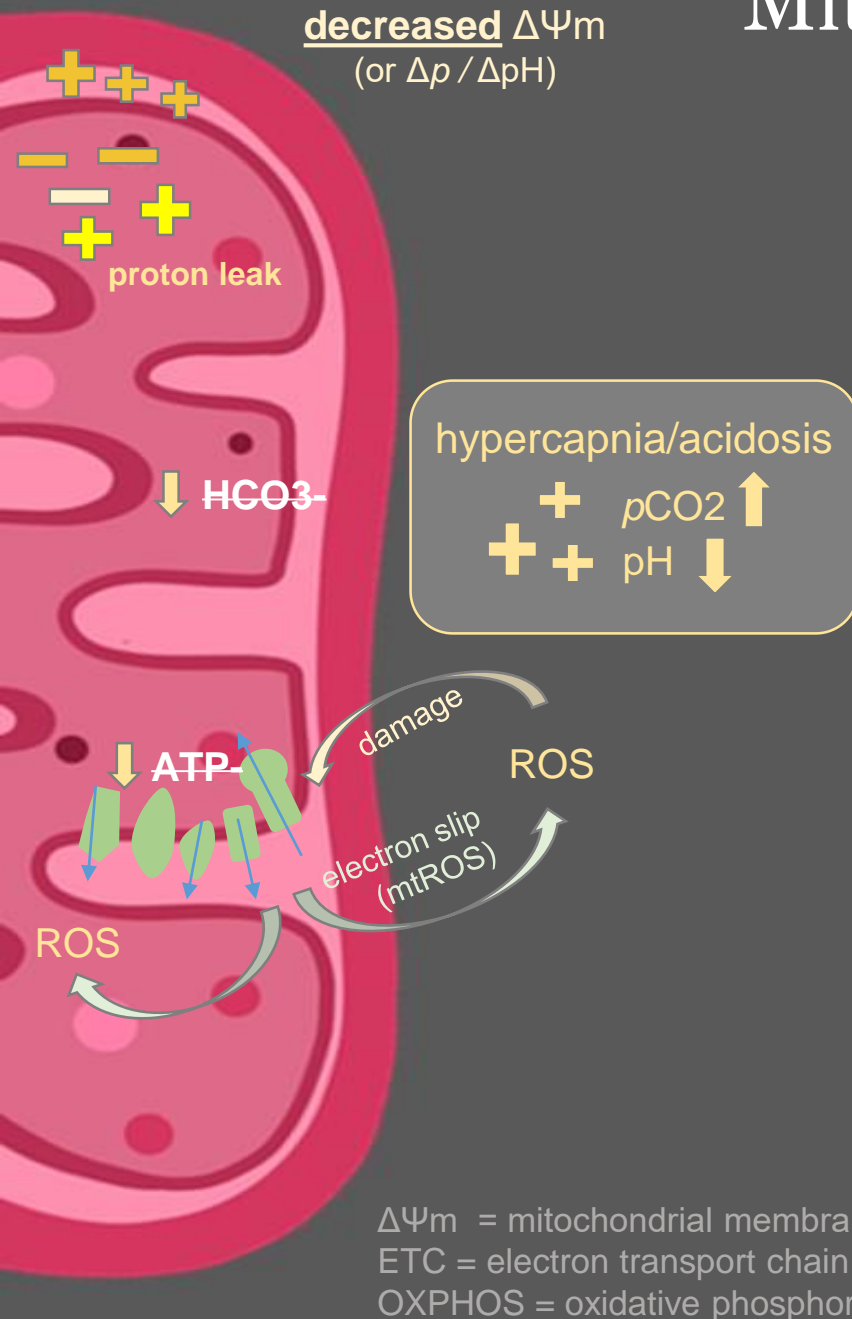
+ pCO_2 ↑
+ pH ↓

Several routes to decrease ATP

- overloads bicarbonate buffer in matrix
 - proton leak
 - lower membrane potential ($\Delta\Psi_m$)
 - decreased efficiency of OXPHOS

$\Delta\Psi_m$ = mitochondrial membrane potential
ETC = electron transport chain
OXPHOS = oxidative phosphorylation

Mitochondrial dysfunction under OA conditions



Several routes to decrease ATP

- overloads bicarbonate buffer in matrix
 - proton leak
 - lower membrane potential ($\Delta\Psi_m$)
 - decreased efficiency of OXPHOS
- increased production of free radicals
 - damage to mitochondrial membrane proteins
 - electron slip, superoxide byproduct (O_2^-)

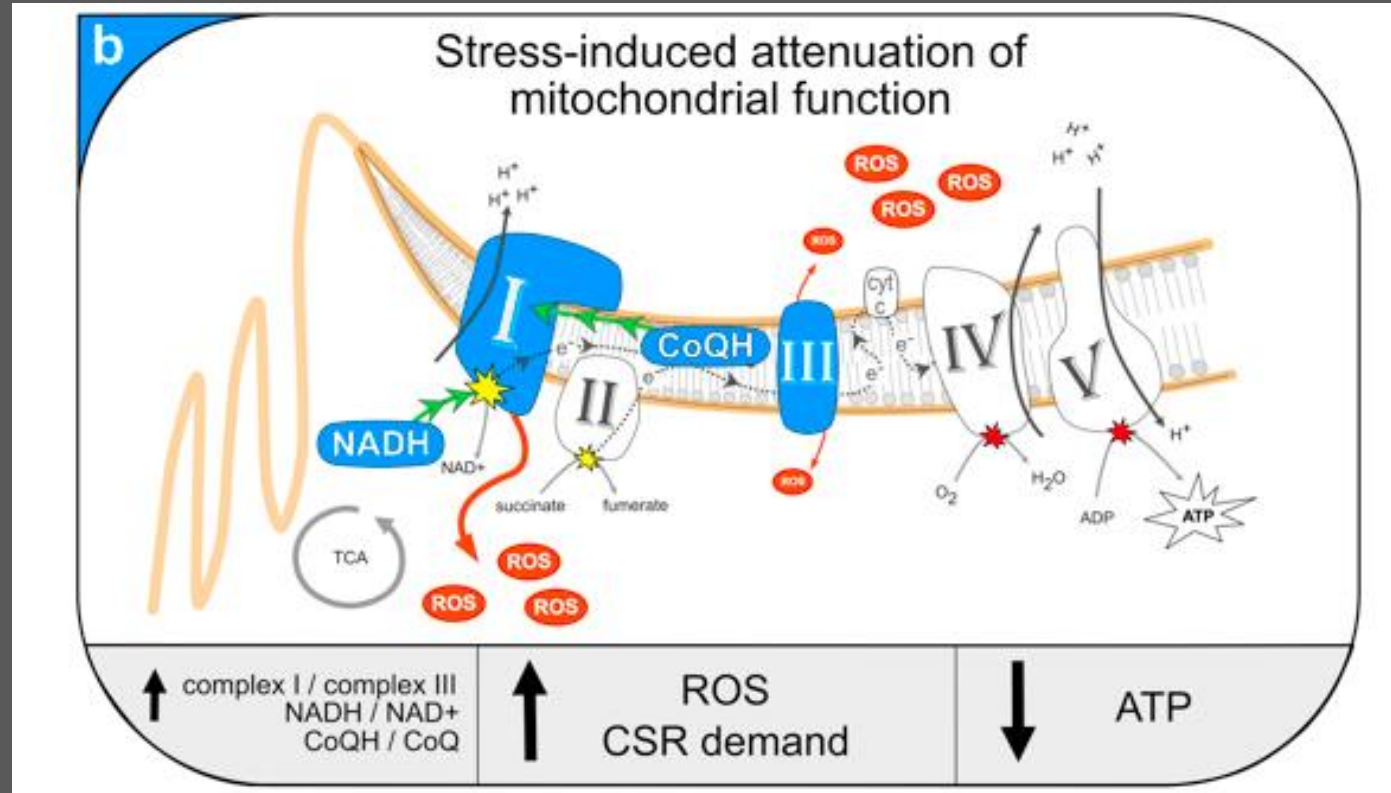
Mitochondrial dysfunction under OA conditions

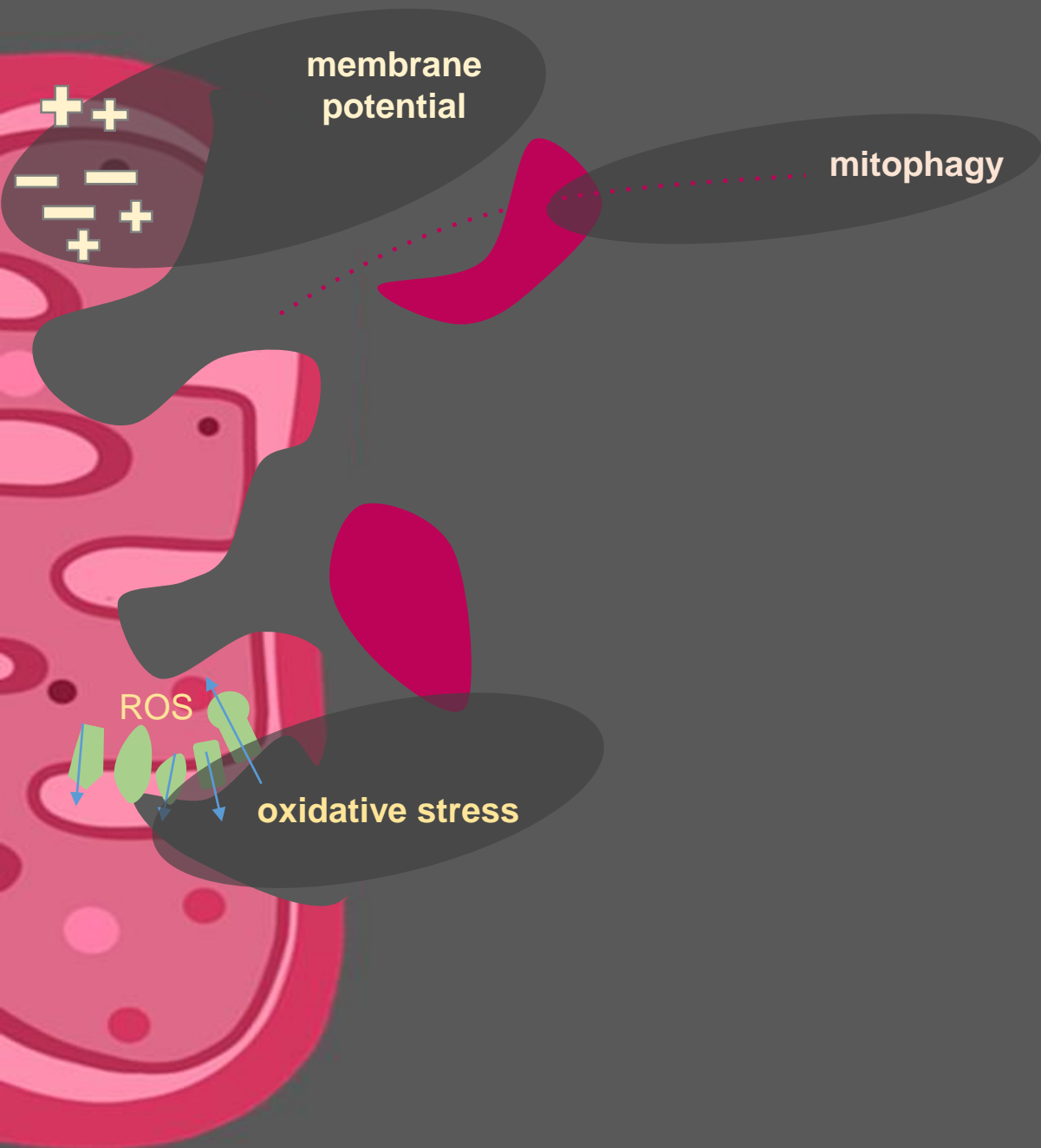


Several routes to decrease ATP

- overloads bicarbonate buffer in matrix
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 - lower membrane potential ($\Delta\Psi_m$)
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 - damage to mitochondrial membrane proteins
 - electron slip, superoxide byproduct (O_2^-)
- mitochondrial recycling and potential cell death
 - dysfunction signals for mitophagy, can be precursor to apoptosis

Mitochondrial dysfunction under OA conditions





...targets for flow cytometry!

mitochondrial **membrane potential** ($\Delta\Psi_m$)

- probe = JC-10

mitochondrial production of **free radicals**

- probe = DCHF-DA

mitophagy & lysosomal content

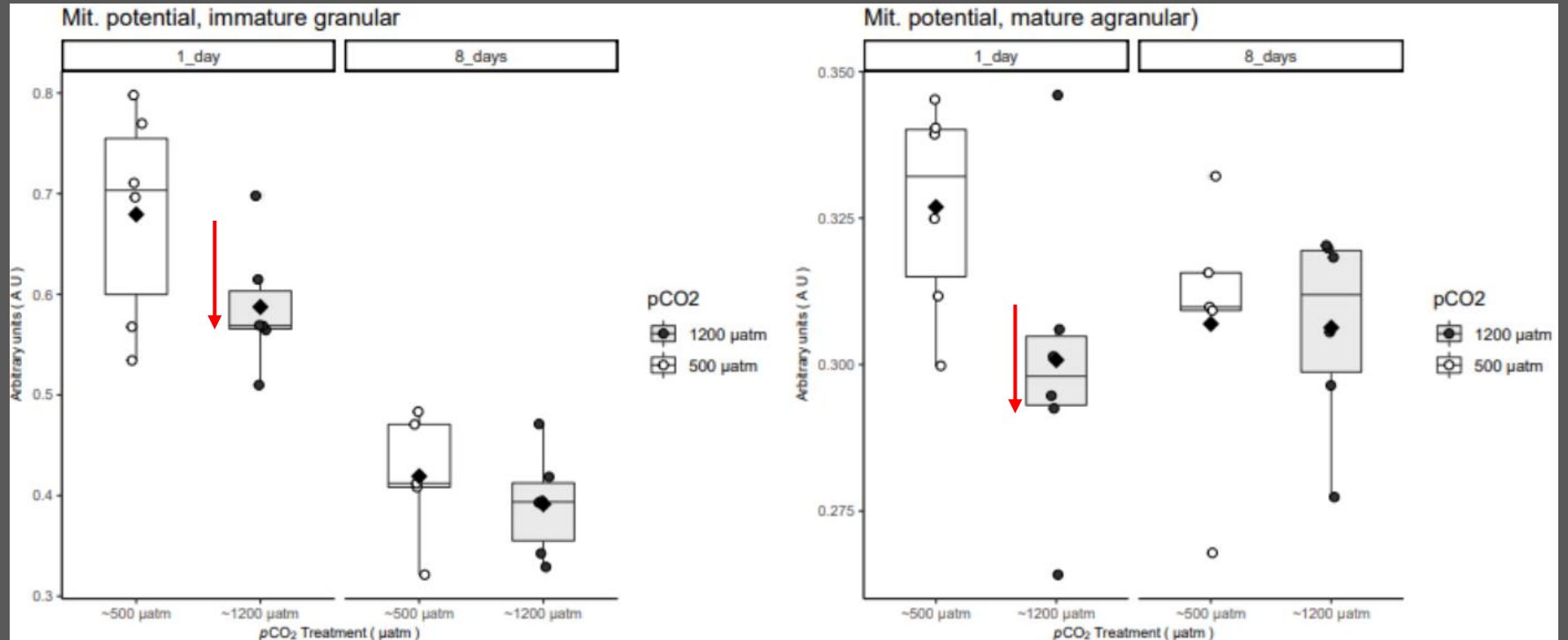
- probe = MitoTracker (Green or Deep Red)
- probe = LysoTracker Red DND-99

hemocyte status / **mortality** / cytomorphology

- probe = syber green and propidium iodide

membrane
potential

preliminary results in adult oysters...



significant decrease in membrane potential for immature granular and mature agranular cells, 'recovery' after 8 days of exposure

- hypothesized effect persisted albeit a low sample size!
- animals were also starved throughout this trial, confounding timepoint comparison

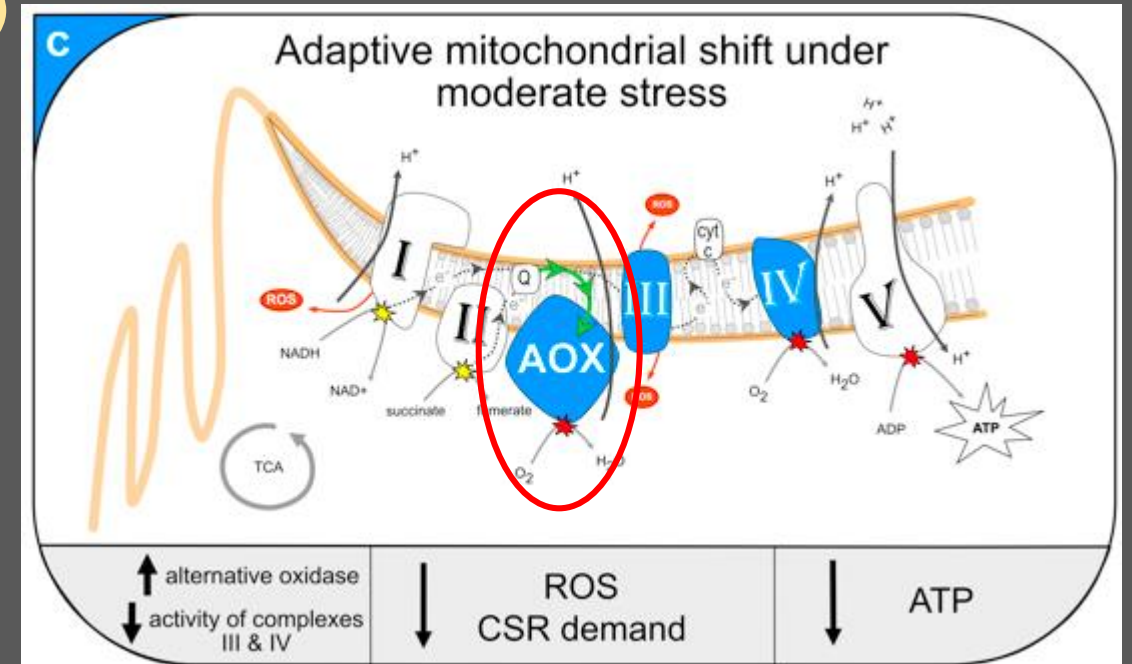
NOAA DFO grant

Colorimetric assays

- NAD/NADH ratio
 - marker of reverse electron transport (dysfunction)
 - optimum level mitochondrial respiration
- Lipid peroxidation
 - a damage signal necessary to infer oxidative stress (coupled with DCFH-DA)

a priori genes of interest (qPCR or TagSeq)

- mitochondrial complexes I and III
 - major components that change activity/concentration under dysfunction
- uncoupling proteins
 - regulate the proton gradient and cell resp.
- alternative oxidase (AOX, view figure →)
 - of growing interest as an adaptive response
- mitophagy pathway
 - FOXO, pink-1 parkin, etc.
- carbonic anhydrase etc.
 - acid-base regulatory proteins



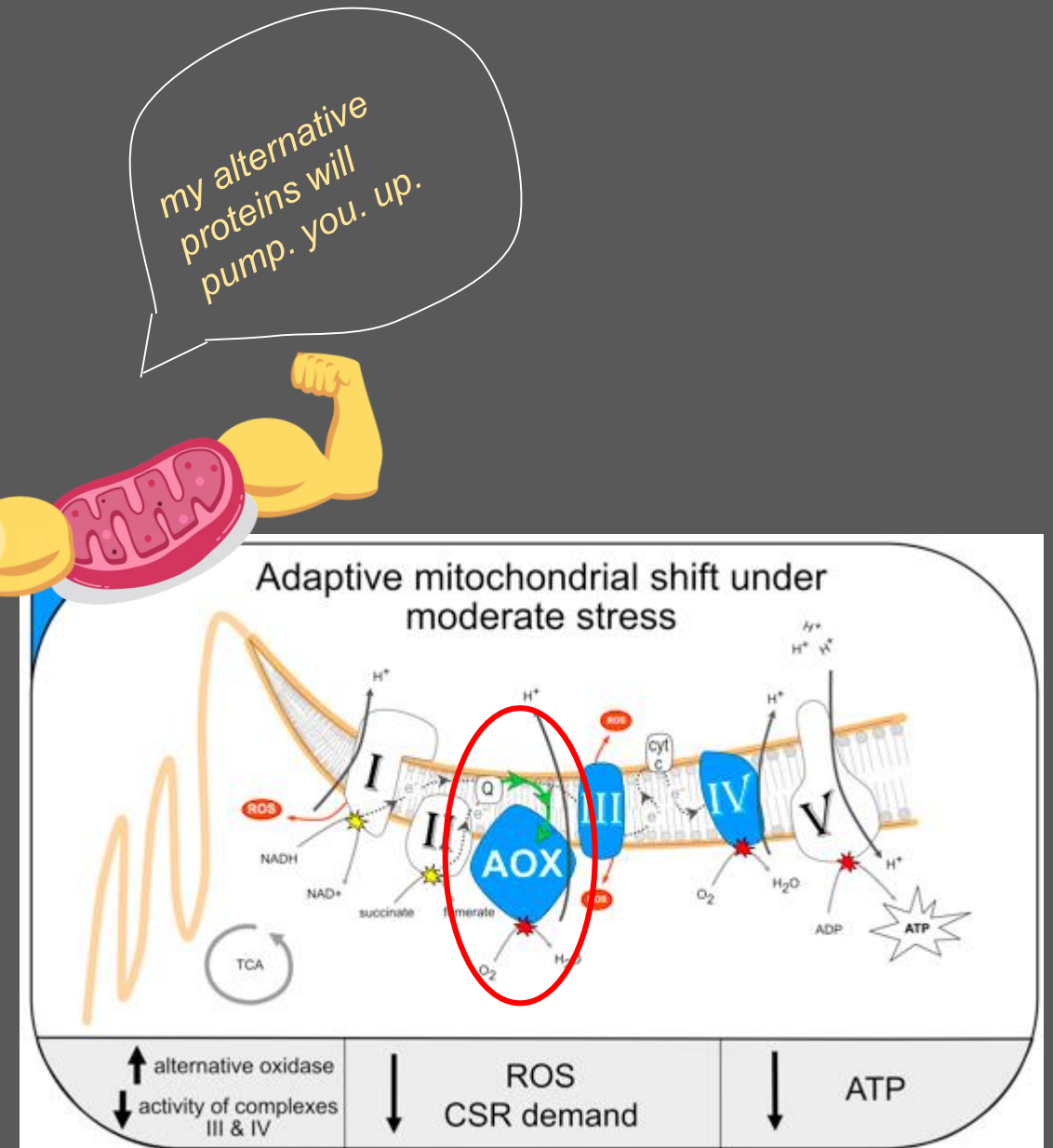
Future directions (with preserved tissues!)

Colorimetric assays

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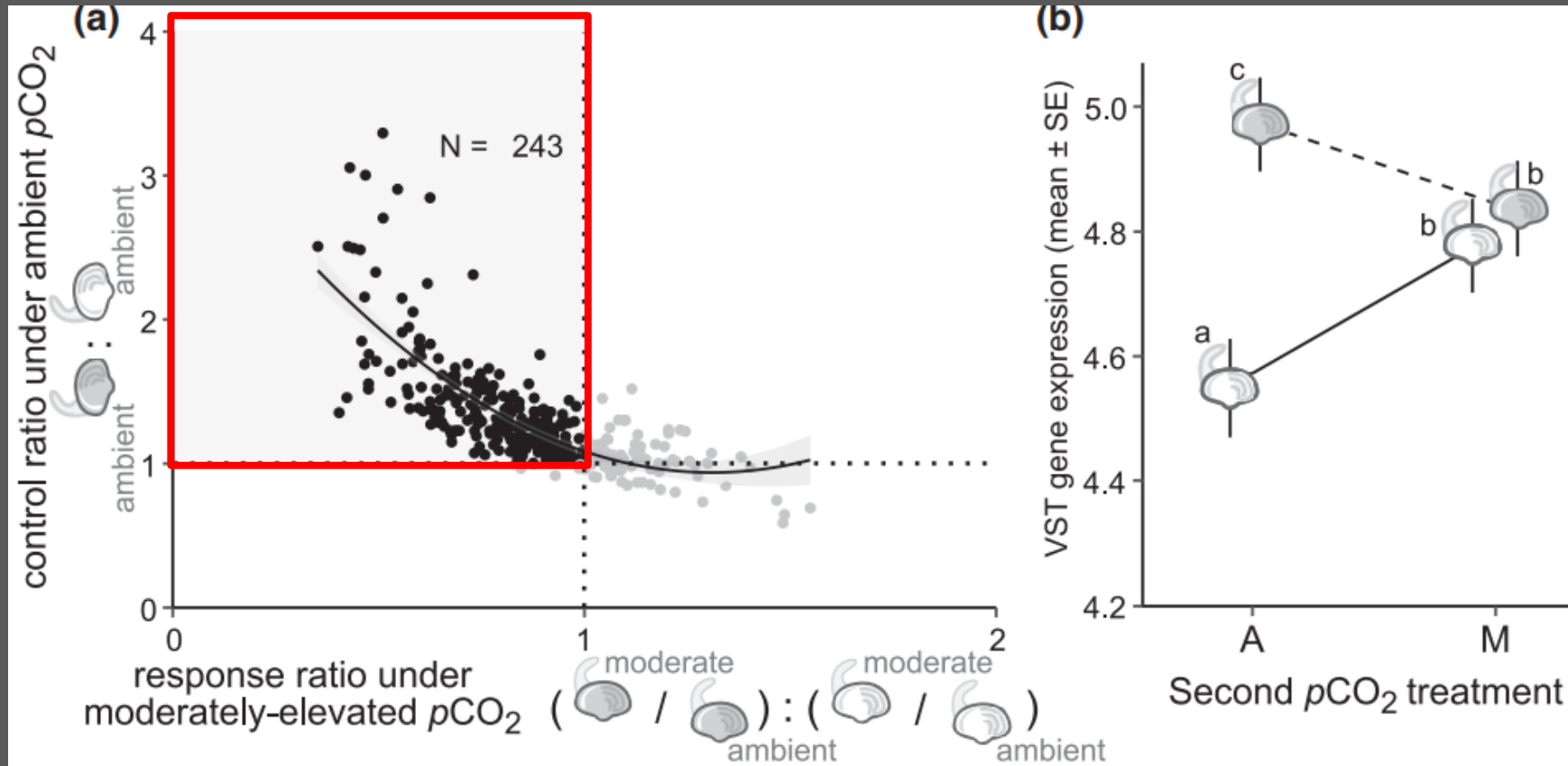
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Future directions (with preserved tissues!)

constitutive gene frontloading of our acclimatized cohort

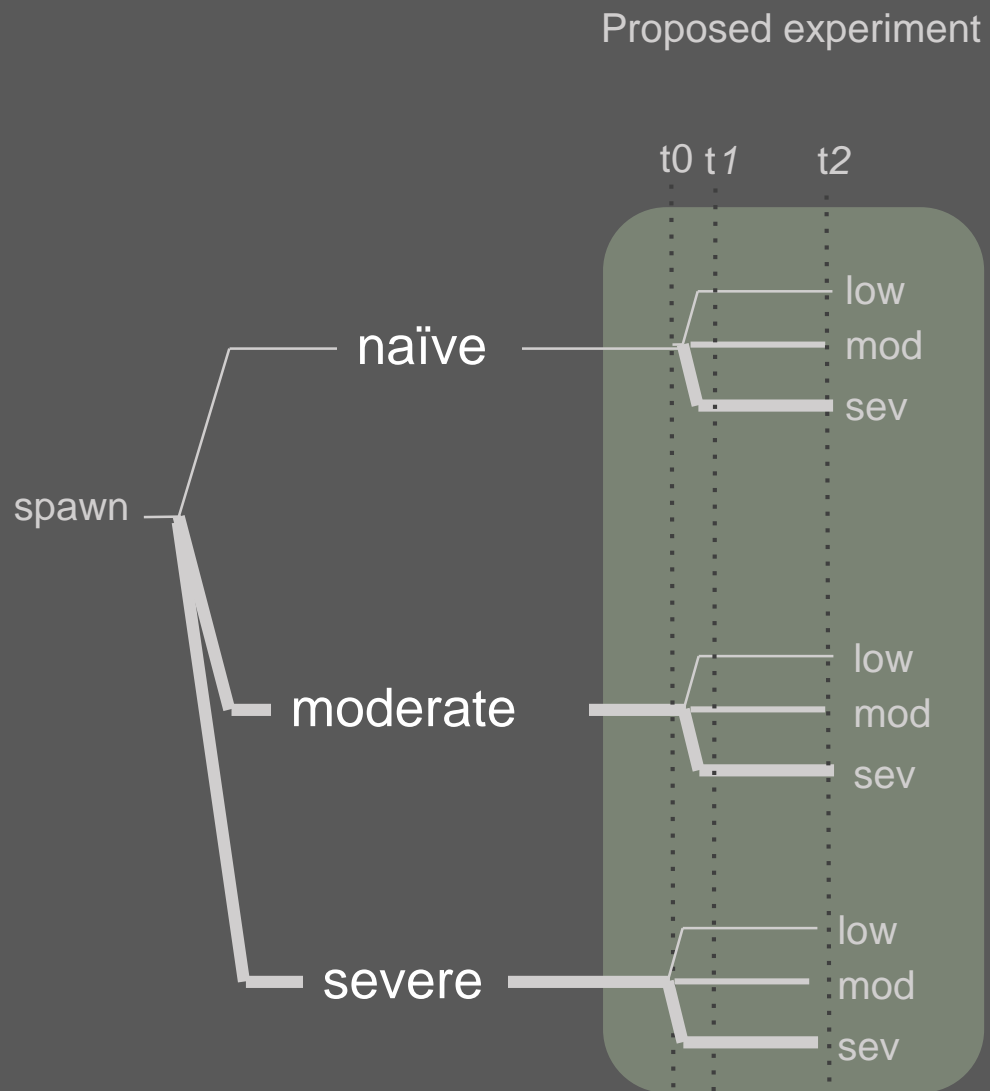


Gurr et al. 2022

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About treatments

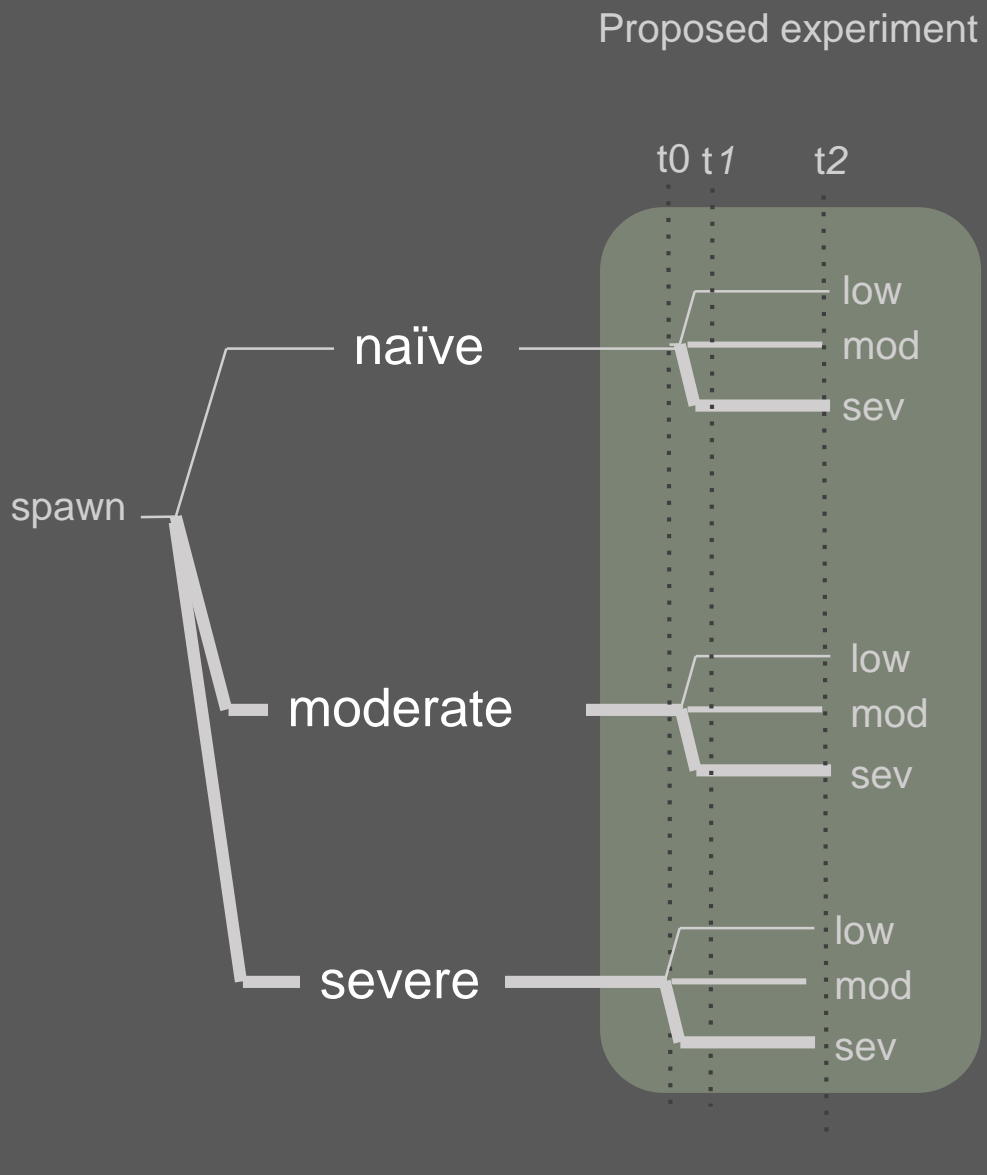
$p\text{CO}_{2\text{HISTORY}}: N = 3$
 $p\text{CO}_{2\text{EXPOSURE}}: N = 3$

$p\text{CO}_{2\text{HISTORY}} \times p\text{CO}_{2\text{EXPOSURE}}: N = 9$

tank replication: $N = 3$

total tanks: $N = 9$





Number of F2s

Low pCO₂

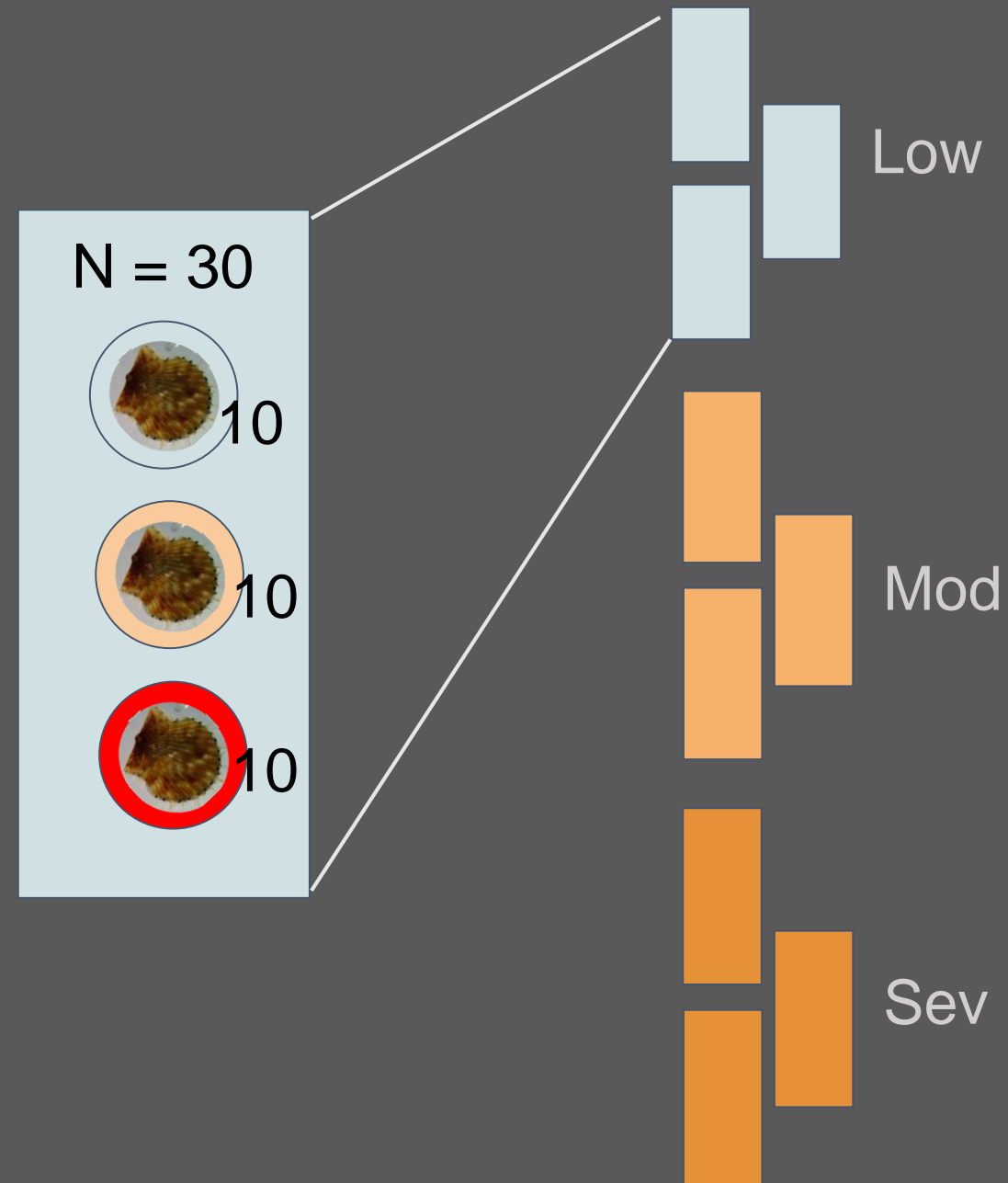
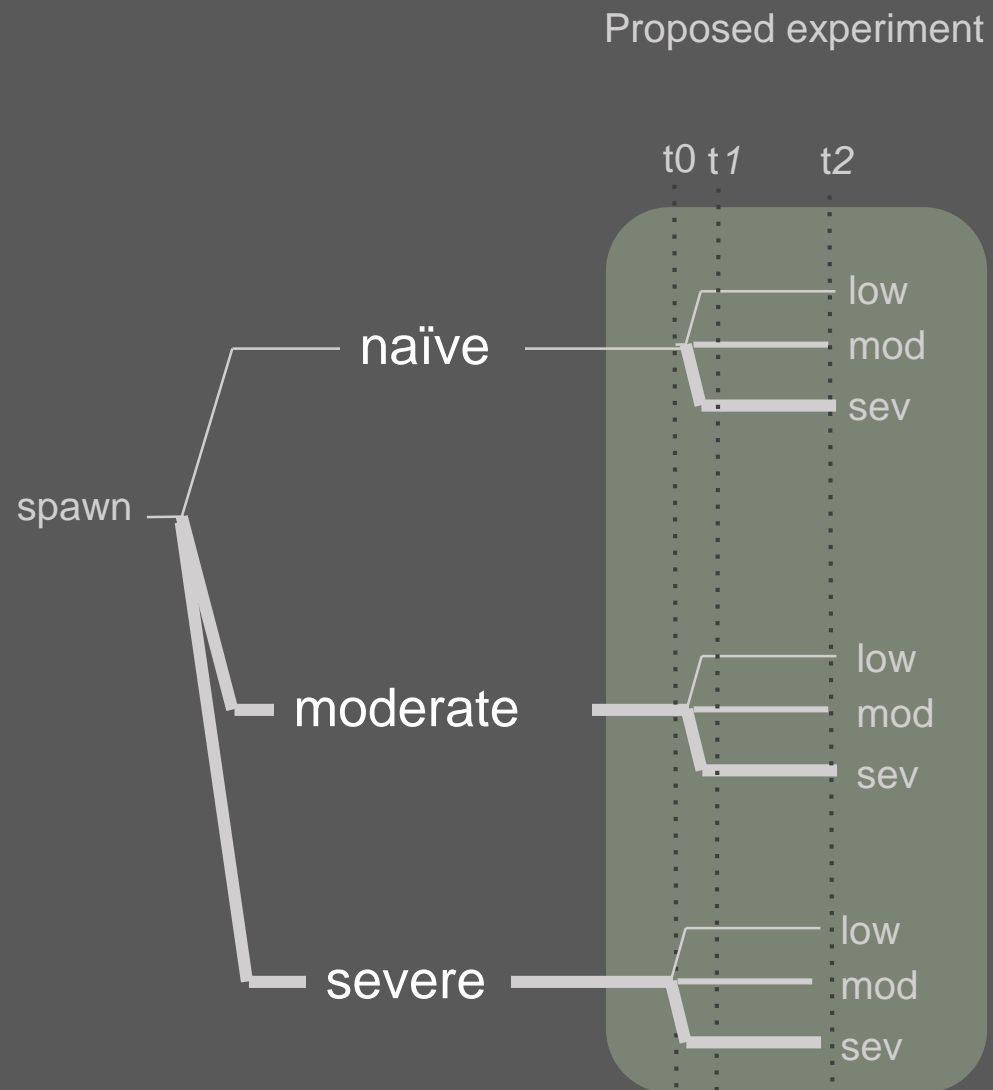


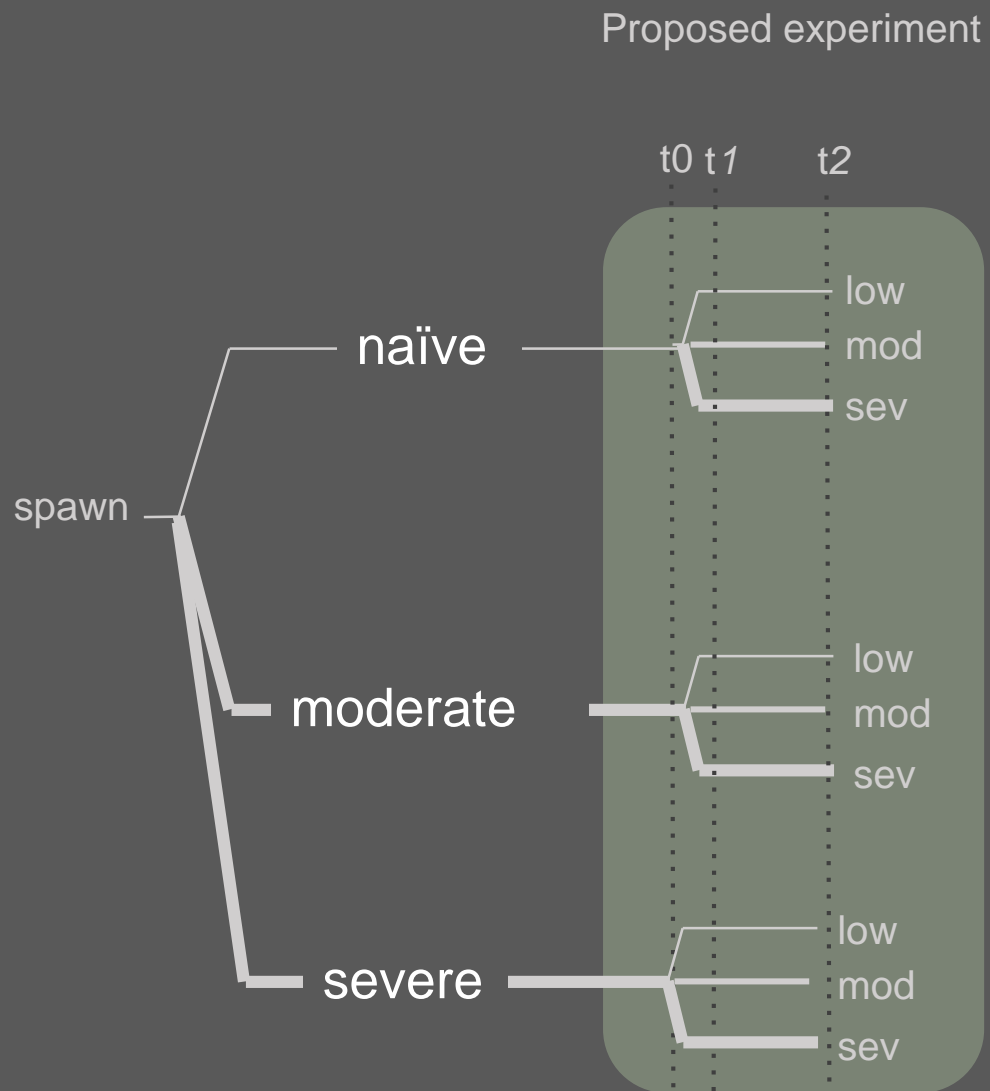
'Moderate' pCO₂



'Severe' pCO₂







N = 10

3/4

3/4


3/4


each tank add to 10 each
with either 3 or 4 indivs
represented by pCO2 history


The diagram shows three tanks, each containing 10 individuals (N = 10). Each tank is labeled with 3/4, indicating that 3 or 4 individuals are represented by pCO2 history. The tanks are arranged vertically, and the text 'each tank add to 10 each with either 3 or 4 indivs represented by pCO2 history' is written below them.



Timepoint 1: April 4th

x Low x Mod x Sev
 5 per treatment; total = 15

x Low x Mod x Sev
 5 per treatment; total = 15

x Low x Mod x Sev
 5 per treatment; total = 15

N = 45



Timepoint 2: April 18th

x Low x Mod x Sev



5 per treatment; total = 15

x Low x Mod x Sev



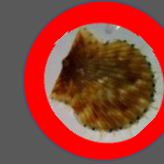
5 per treatment; total = 15

x Low x Mod x Sev



5 per treatment; total = 15

N = 45..
again!





5 per treatment; total = 15



5 per treatment; total = 15



5 per treatment; total = 15



45 animals at each time point (90 total)



same day:
hemolymph



future analysis:
gill tissue x2 per individual!

Hemolymph probes

- **SYBR green** = live cells
- **JC-10 + CCCP** = mitochondrial membrane potential with and without depolarizing (control)
- **MitoSox Green** = mitochondrial free radicals

same day:
hemolymph

45 animals at each time point (90 total)

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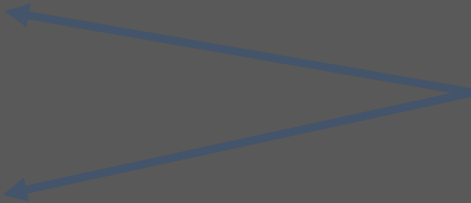
Future analysis of gill tissue!

Gene frontloading

- TagSeq (N=64 samples max)

Colorimetric assays with tissue homogenate

- NAD/NADH Quantification
- Lipid peroxidation (MDA)
- ADP/ATP
- corrected for Total protein



future analysis:
gill x2