

Introduction to metagenetics analysis with QIIME2





Websites

NEOF: <https://neof.org.uk/>

NERC: <https://nerc.ukri.org/>

CGR:

<https://www.liverpool.ac.uk/genomic-research/>



Twitter

NEOF: @NERC_EOF

NERC: @NERCscience

CGR: @CGR_UoL

Upcoming workshops

<https://neof.org.uk/training/>

- Metabarcoding for diet analysis and environmental DNA
 - 28th February & 2nd March 2023
- Microbial shotgun metagenomics
 - 21st & 23rd March 2023
- Eukaryote genome assembly
 - 18th & 20th April 2023
- More!

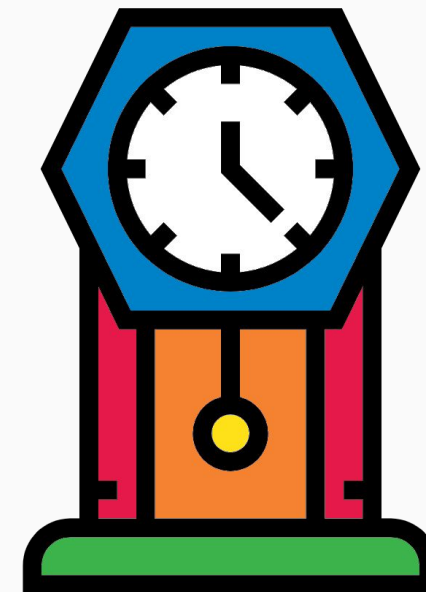




Format & Schedule

This intro
Bookdown
Theory
Practice
Exercises
MCQs
Optional materials

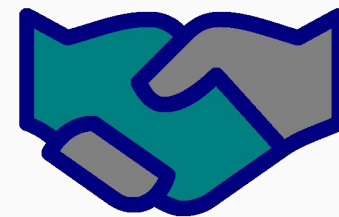
Work at your own
pace on your own
time





Introduction

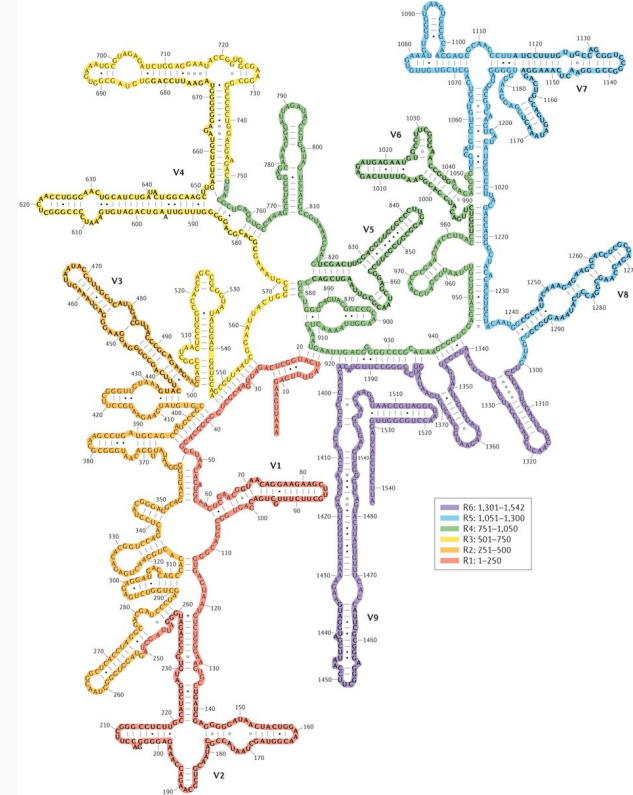
- Why 16S rRNA?
- What is QIIME2?
- QIIME2 Workflow
- Quality Control prior to QIIME2 analysis
- DADA2
- Sequence table and taxonomy classification
- Biodiversity
 - Alpha, Beta and Gamma
- Biomarker detection



16S rRNA



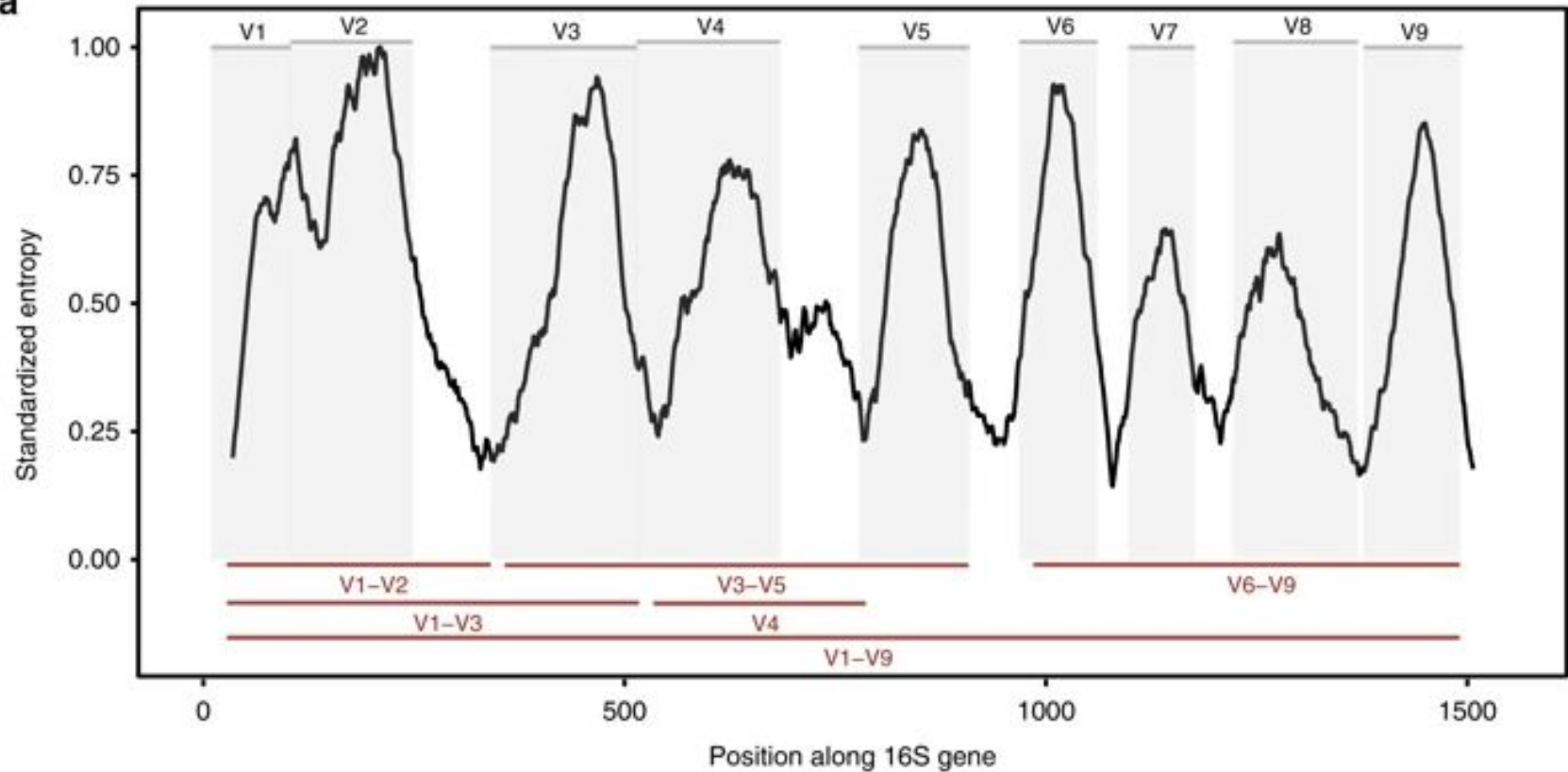
- 16S ribosomal RNA
- Prokaryotic ribosome
- Phylogeny reconstruction
- Slow rate of evolution
- Functional constancy
- ~1,500 bp long
- 9 variable regions
- Flanked by conserved regions



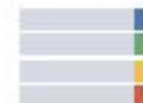
Nature Reviews | Microbiology

Yarza, Pablo, et al. "Uniting the classification of cultured and uncultured bacteria and archaea using 16S rRNA gene sequences." *Nature Reviews Microbiology* 12.9 (2014): 635-645.



a

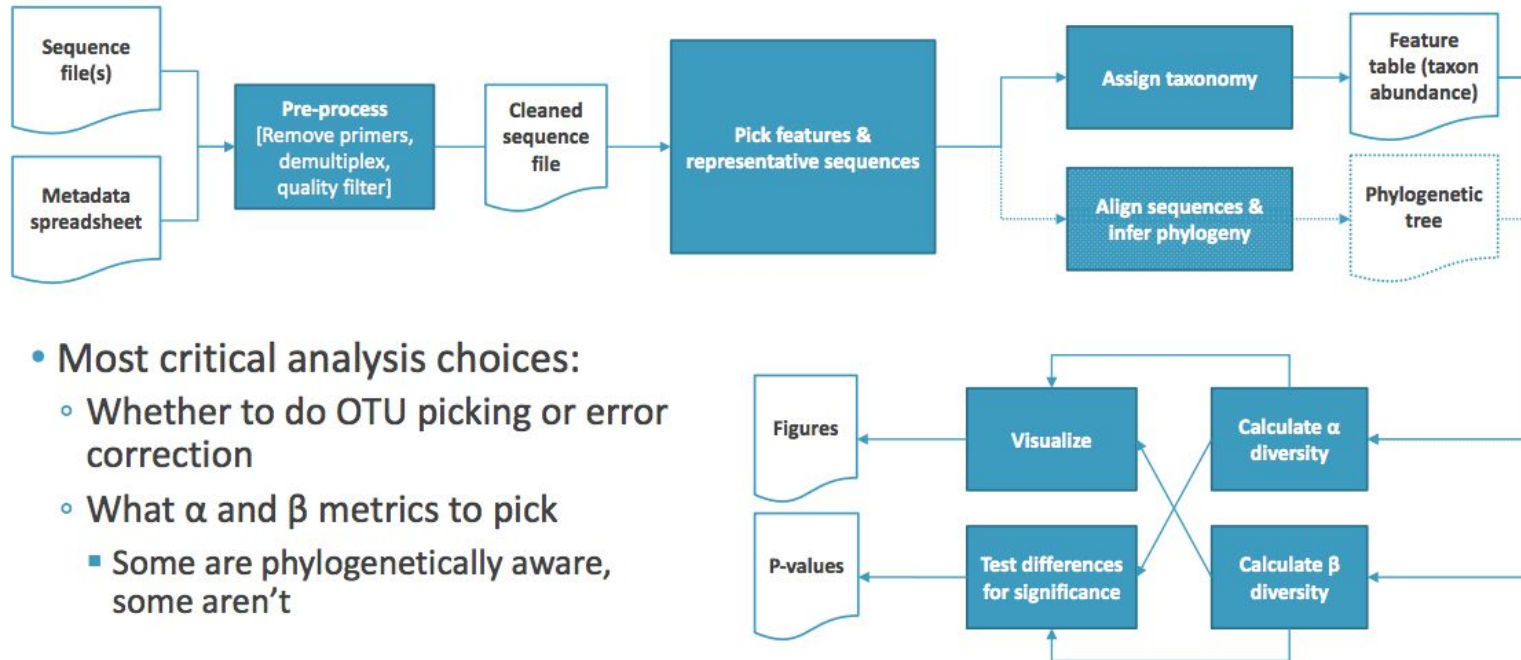
Johnson, Jethro S., et al. "Evaluation of 16S rRNA gene sequencing for species and strain-level microbiome analysis." *Nature communications* 10.1 (2019): 1-11.



What is qiime2?

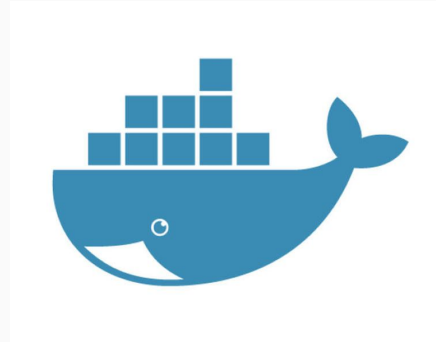
- Chiime
- Quantitative Insights Into Microbial Ecology
- Open-source pipeline
- Python
- Wraps Popular algorithms
- Comparison and analysis of microbial communities
- Next iteration of QIIME
 - Cited 24,228 times (07 APR 2021)
 - Publication: Caporaso, J. Gregory, et al. (2010)





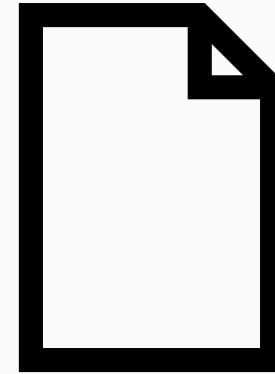
qime2 Installation

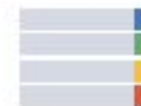
- ANACONDA
- VirtualBox
- Amazon Web Services
- Docker



supported input files

- Sequence files
 - Fastq
- Artifact files
 - .qza: Contains all data QIIME2 requires
 - .qzv: Files containing visualisation information





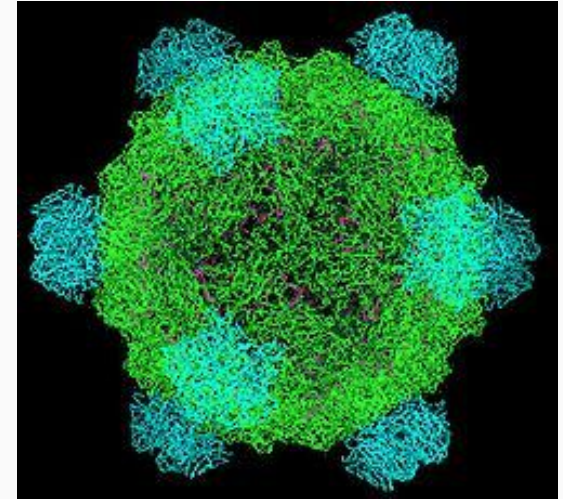
DADA2

- Divisive Amplicon Denoising Algorithm
- Models and corrects Illumina-sequenced amplicon errors
- Pipeline
 - Filtering
 - Dereplication
 - Chimera Identification
 - Merging paired-reads
- <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4927377/>



PhiX removal

- Remove sequences similar to PhiX
 - **BLAST**
- PhiX Quality control
 - cluster generation
 - Sequencing
 - alignment.
- PhiX virus





Chimera removal

DNA from two or more parent molecules

- PCR artifact
- Erroneous “novel” sequence

aborted amplification



next cycle's “primer”



chimeric sequence



Merging reads



- Align read pairs R1/R2 to each other
- Improves quality of reads
- Longer reads

R1 (30bp)	ACCGTACGTATGCGTAGCTGACGTAGCATG-----
R2 (30bp)	-----TGCGTAGCTGACGTAGCATGCGCGATTCTGA
Overlap (20bp)	-----TGCGTAGCTGACGTAGCATG-----
Stitched read (40bp)	ACCGTACGTATGCGTAGCTGACGTAGCATGCGCGATTCTGA

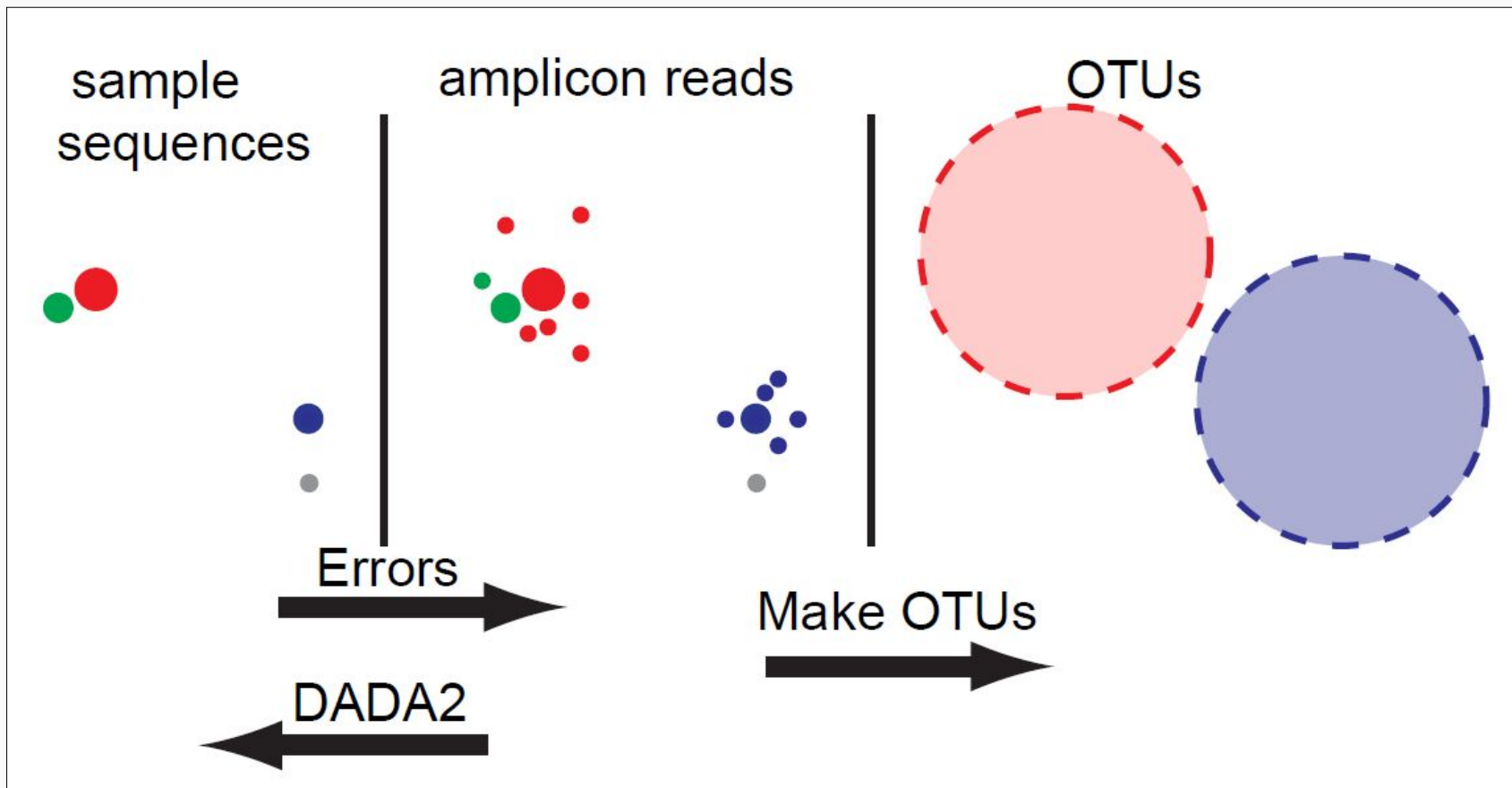


OTUs vs Amplicon Sequence Variants (ASV)

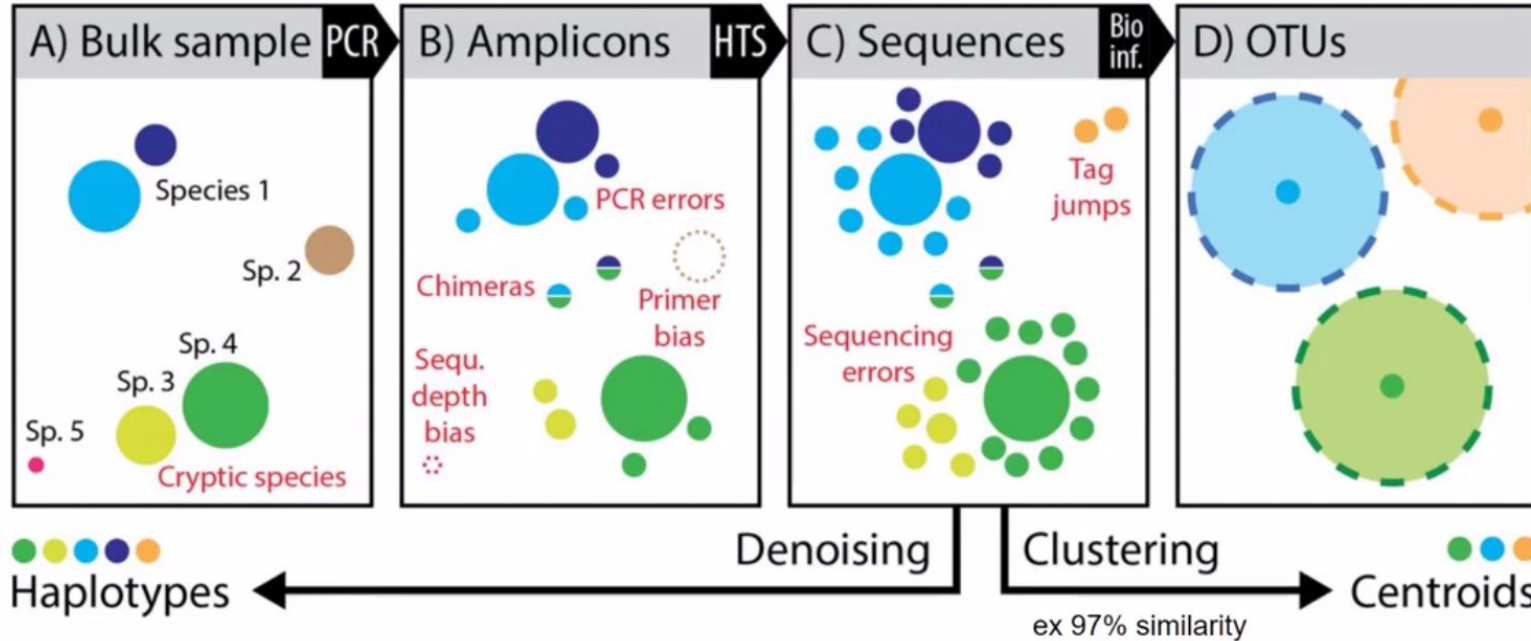


- Previous methods
 - OTUs (Operational taxonomic units)
 - Cluster sequences by identity e.g. 97% similarity for species
 - Can cause over clustering
 - 97% is chosen due to errors within Illumina data
- DADA2
 - Denoises and cleans reads so they represent real sequences
 - Much finer resolution on sequences
 - Can differentiate sequences that have only 1bp difference



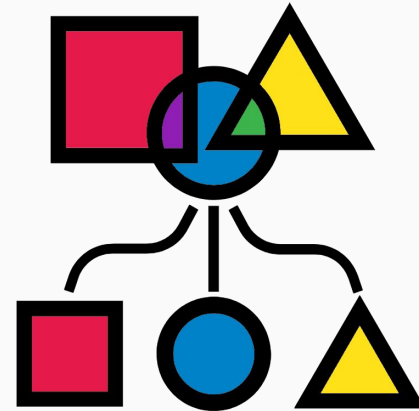


Errors and denoising recap

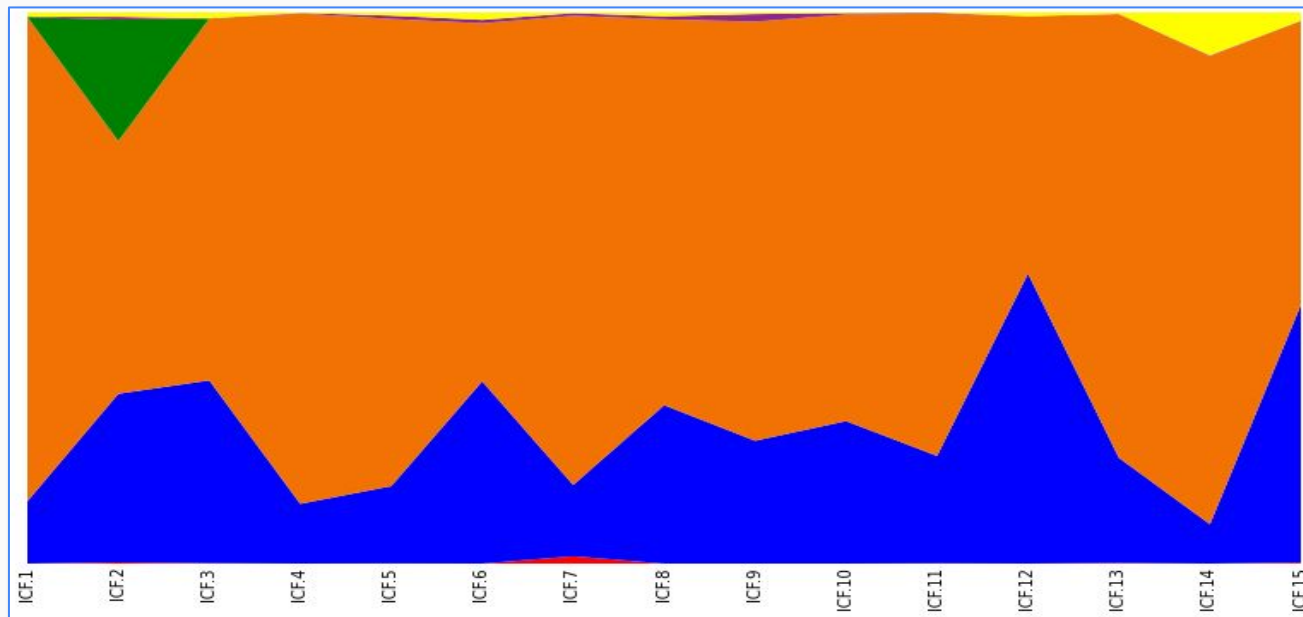


Taxa classification

- Assigns each ASV taxonomy based on a DB
- Reliant on:
 - Quality and completeness of DB
 - Tool used to search DB
- Two highly used DBs
 - Greengenes
 - Silva



Phylum level Taxonomy

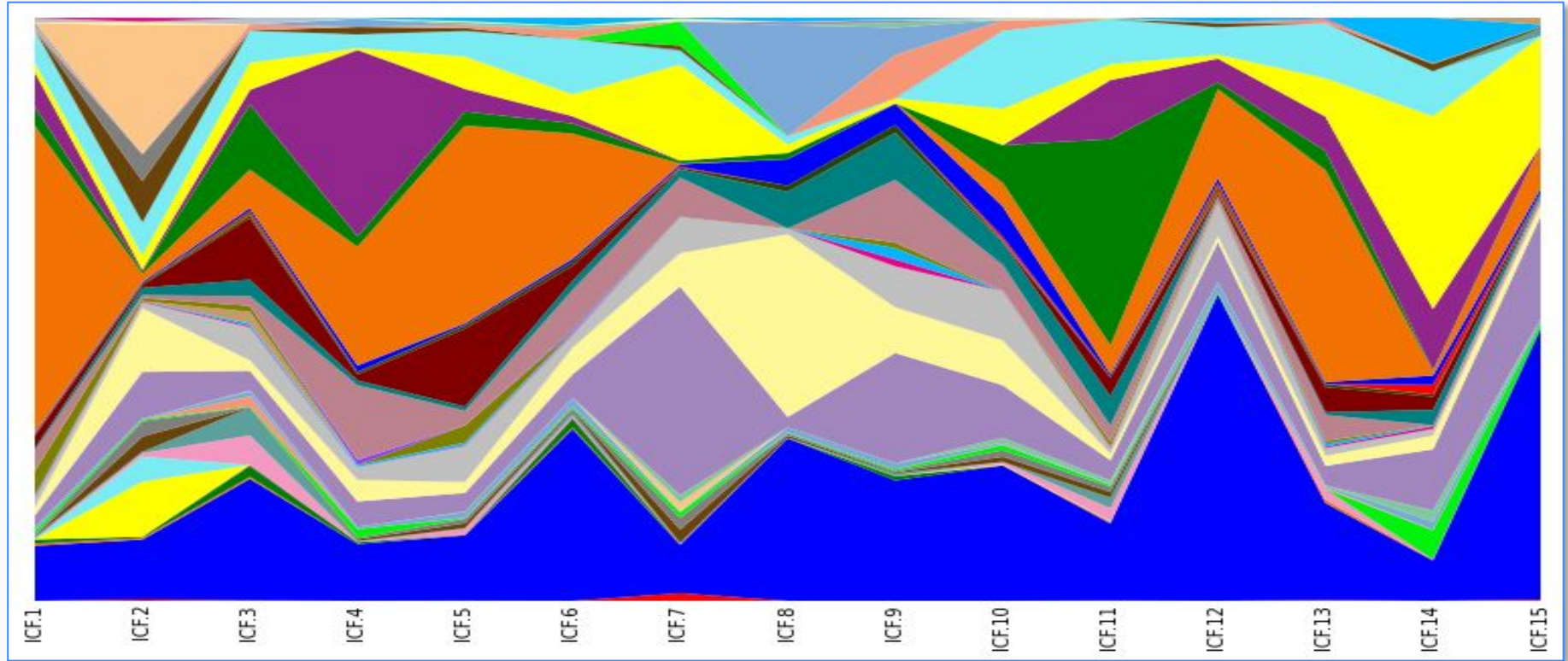


[View Table \(.txt\)](#)

Legend	Taxonomy	Total		ICF.1	ICF.2	ICF.3	ICF.4	ICF.5	ICF.6	ICF.7	ICF.8	ICF.9	ICF.10	ICF.11	ICF.12	ICF.13	ICF.14	ICF.15
		count	%	%	%	%	%	%	%	%	%	%	%	%	%	%	%	%
■	k_Bacteria;p_Actinobacteria	0	0.1%	0.0%	0.3%	0.1%	0.0%	0.0%	0.0%	0.0%	1.3%	0.0%	0.0%	0.0%	0.0%	0.1%	0.0%	0.2%
■	k_Bacteria;p_Bacteroidetes	4	24.5%	11.3%	30.6%	33.2%	10.8%	14.0%	33.1%	12.9%	28.7%	22.3%	25.9%	19.5%	52.6%	19.1%	7.1%	46.7%
■	k_Bacteria;p_Firmicutes	11	72.5%	87.8%	45.8%	65.6%	89.0%	84.8%	64.9%	85.1%	70.0%	76.0%	73.8%	80.2%	46.7%	80.5%	84.9%	51.5%
■	k_Bacteria;p_Fusobacteria	0	1.5%	0.0%	22.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
■	k_Bacteria;p_Proteobacteria	0	0.3%	0.0%	0.5%	0.0%	0.1%	0.6%	0.6%	0.5%	0.5%	1.4%	0.3%	0.2%	0.0%	0.0%	0.1%	0.0%
■	k_Bacteria;p_Tenericutes	0	1.1%	0.8%	0.8%	1.1%	0.1%	0.6%	1.4%	0.1%	0.7%	0.3%	0.1%	0.0%	0.7%	0.3%	7.8%	1.5%



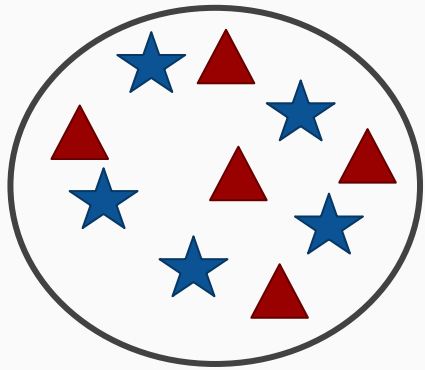
Genus level taxonomy



Biodiversity



- Richness
- Relative abundance
- Evenness



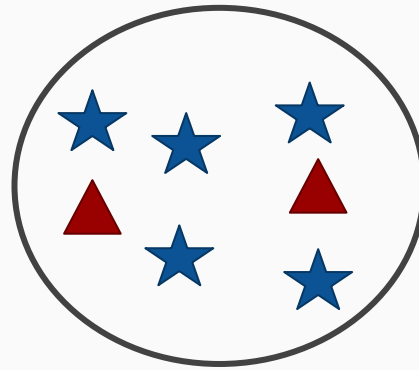
Richness: 2

Relative
abundance:

$$5/10 = 0.5$$

$$5/10 = 0.5$$

Even



Richness: 2

Relative
abundance:

$$2/7 = 0.29$$

$$5/7 = 0.71$$

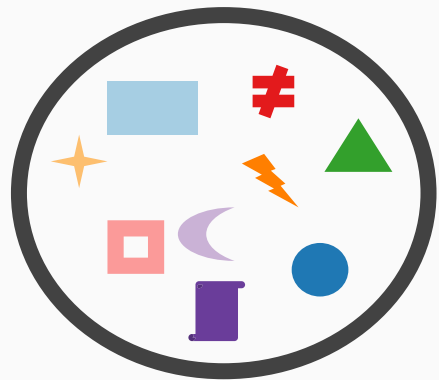
Uneven



Alpha diversity

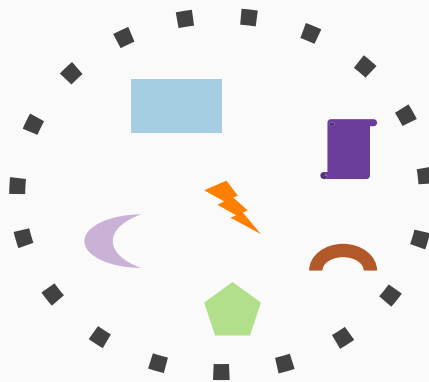


Richness example



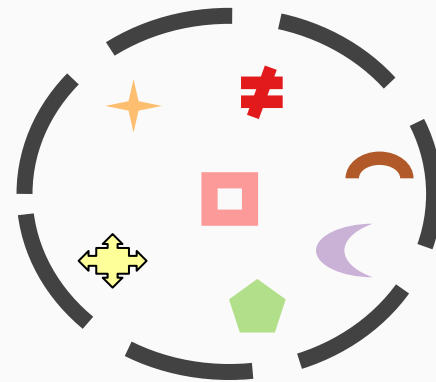
A

Richness: 9



B

Richness: 6



C

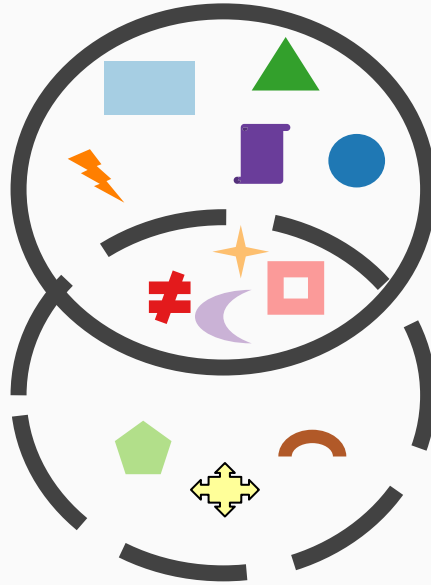
Richness: 7



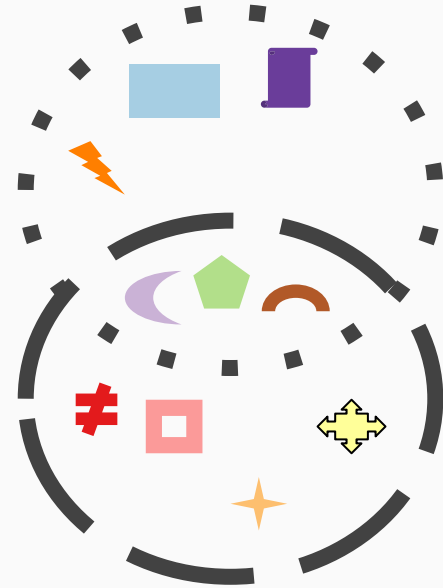
Richness example



A vs B



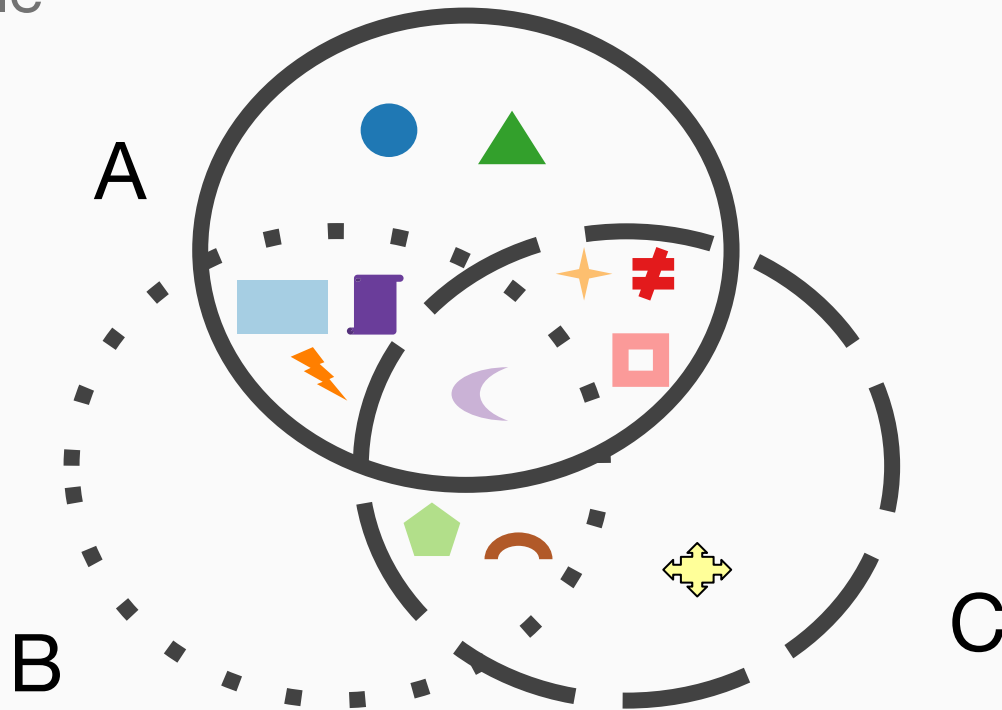
A vs C



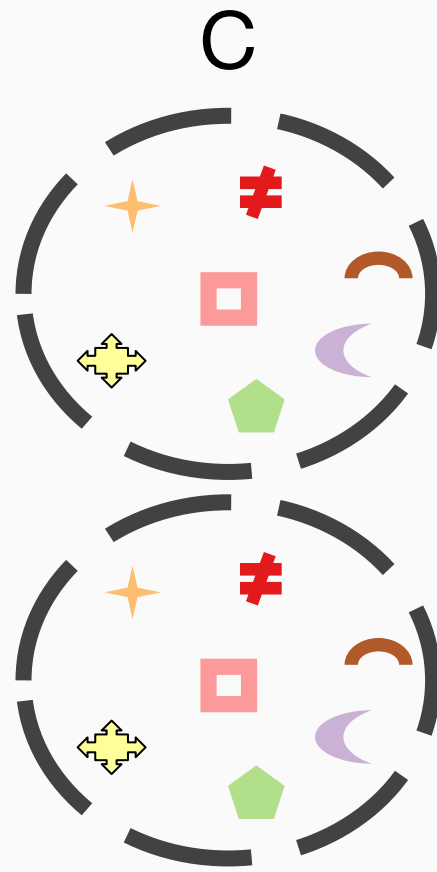
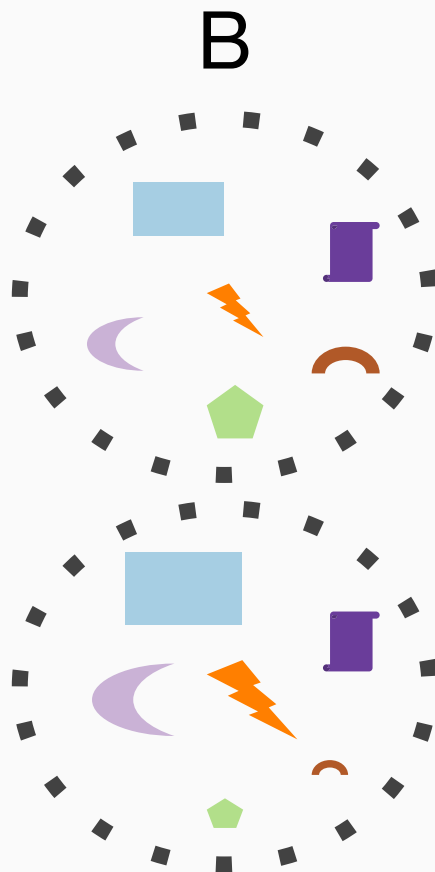
B vs C



Richness example



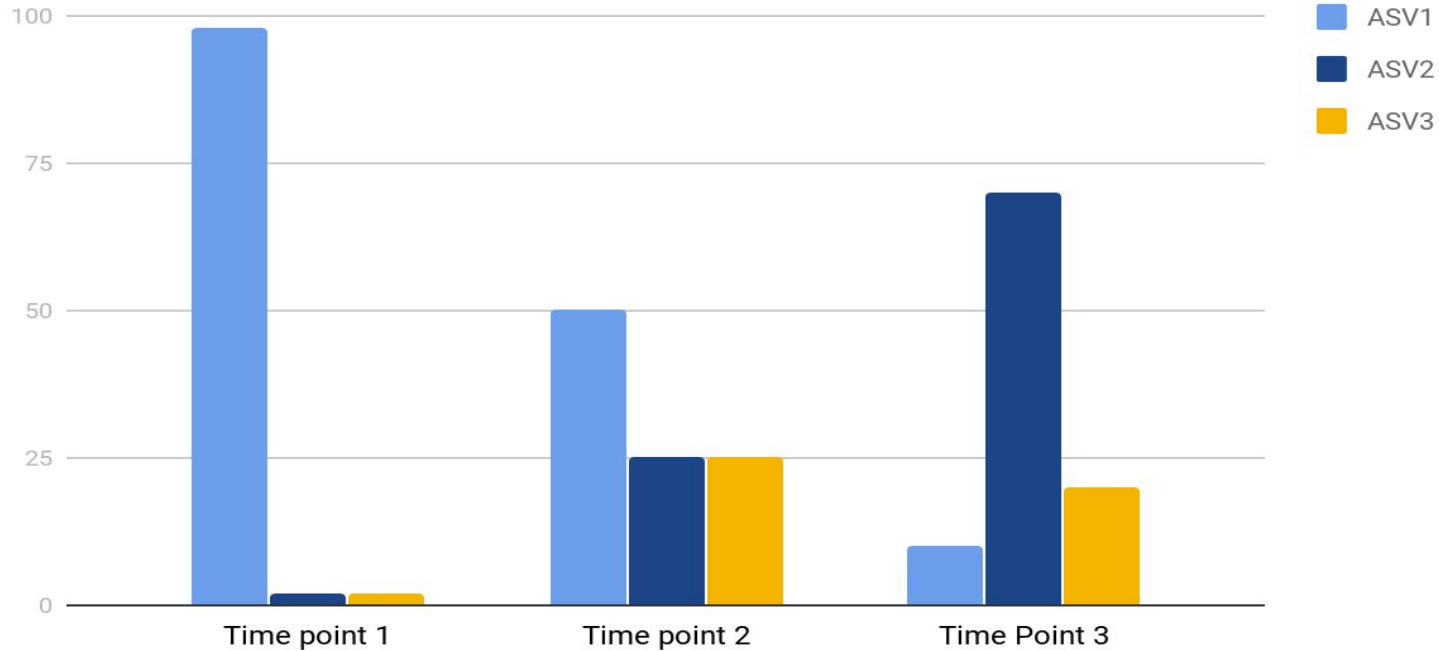
Relative abundance



Relative abundance



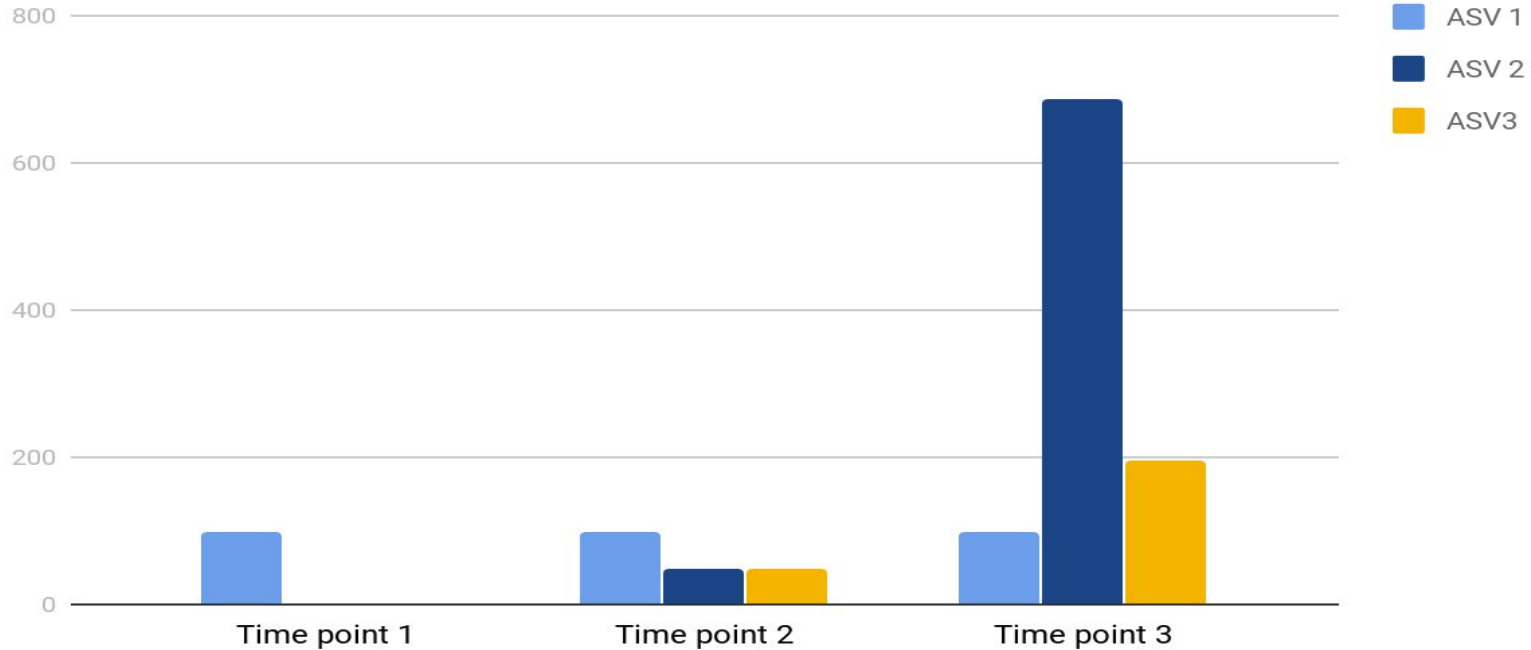
Relative abundance



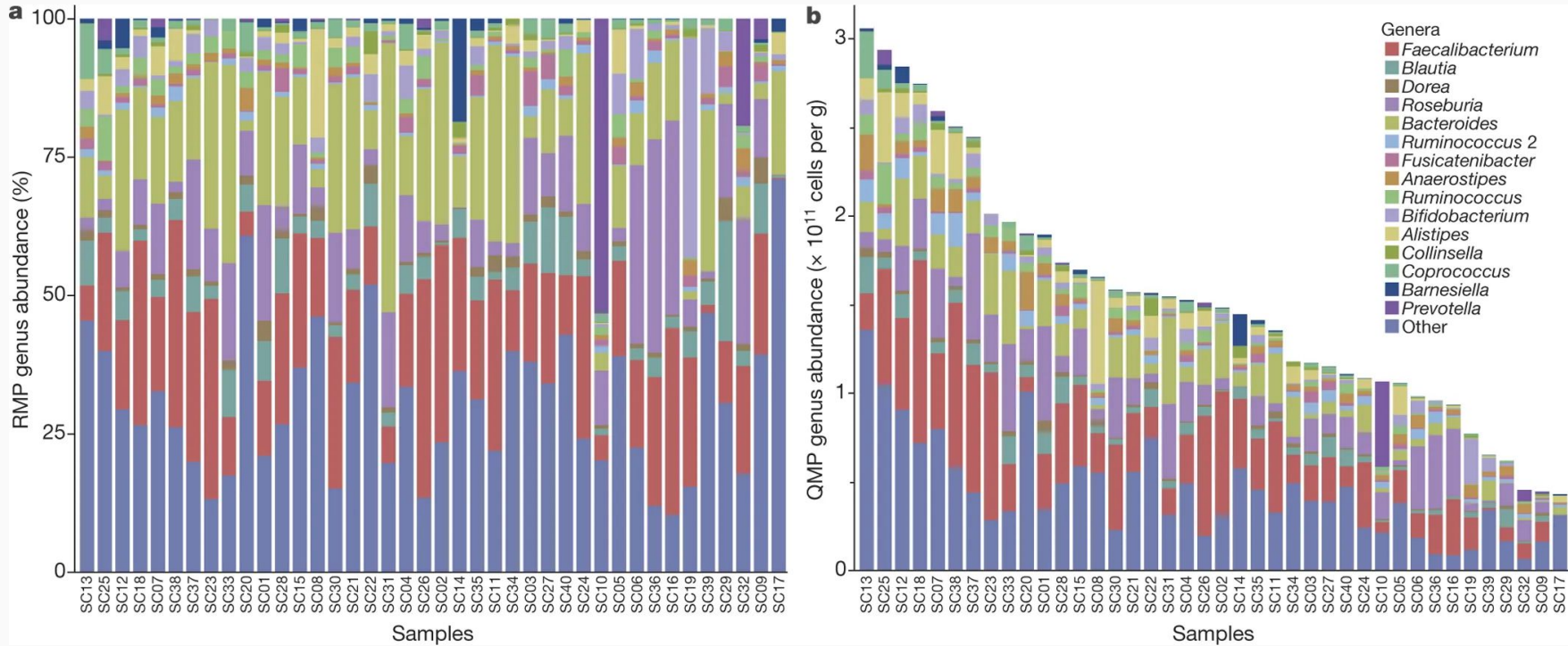
Total abundance



Total abundance



Relative versus quantitative



Vandeputte, Doris, et al. "Quantitative microbiome profiling links gut community variation to microbial load." *Nature* 551.7681 (2017): 507-511.



- Biomarker detection
- Both statistically heavy methods
- ANCOM
 - Analysis of composition of microbiomes: a novel method for studying microbial composition
 - Differential abundance analysis
 - ANCOM assumes that less than ~25% of features are changing between groups.
 - <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4450248/>
- gneiss
 - Differential abundance analysis
 - Attempts to account for using relative abundance through balances
 - In essence it looks at log ratios try to determine real changes





Online Resources

- QIIME2 documents
 - <https://docs.qiime2.org/2021.2/tutorials/>





Useful papers

- What is new and relevant for sequencing-based microbiome research?
A mini-review
 - Johnson, Jethro S., et al. "Evaluation of 16S rRNA gene sequencing for species and strain-level microbiome analysis." Nature communications 10.1 (2019): 1-11
- Evaluation of 16S rRNA gene sequencing for species and strain-level microbiome analysis
 - Vandeputte, Doris, et al. "Quantitative microbiome profiling links gut community variation to microbial load." Nature 551.7681 (2017): 507-511.





Reminders and Tips

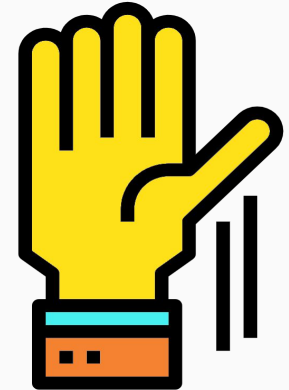
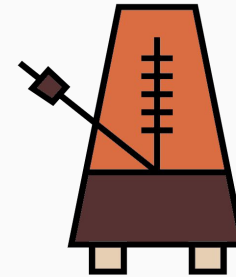
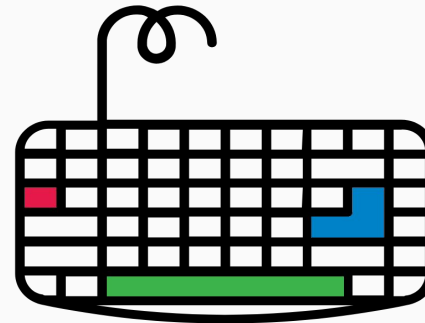
Work at your own pace

Typos

Ask questions

Breaks are important

Tab, space, and enter



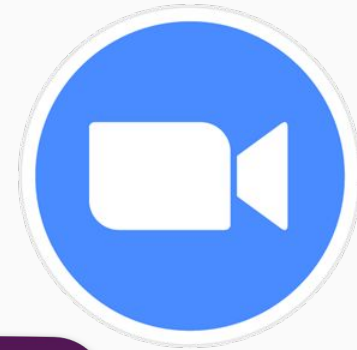


Online class info

Zoom - Ask via microphone if no question currently being asked/answered

Slack - Ask questions via the channel or ask to go into a zoom breakout room with one of us

WebVNC - We can connect to your webVNC to see and help with issues.

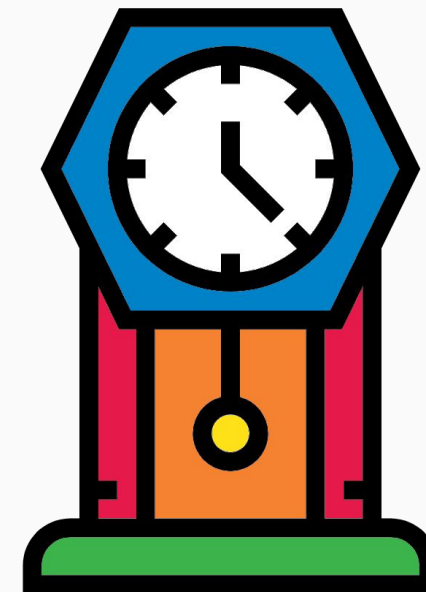




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Book

Online HTML book
Read through and follow
instructions

Link is in workshop
agenda
Active learning



16S metabarcoding

[1 Introduction](#)

[2 Background](#)

[3 Introduction to QIIME2](#)

[4 Cluster Introduction](#)

[5 Data](#)

[6 QIIME2 analysis workflow](#)

[Preprocessing of data](#)

[7 Sequence import](#)

[8 Trim the PCR primer sequences](#)

[ASVs, Taxonomy, and phylogeny](#)

[9 De-novo amplicon sequence varia...](#)

[10 ASV taxonomic assignment](#)

[11 Phylogenetic tree construction](#)

[Analysis](#)

[12 Sequencing depth evaluation](#)

[13 Diversity analysis](#)

[14 Alpha diversity statistical analysis](#)

[15 Beta diversity statistical analysis](#)

[16 Differential abundance analysis](#)

[17 Final consideration](#)

[Appendix](#)

[A Resources](#)

[B Software installation](#)

☰ 🔍 ↗ ⓘ

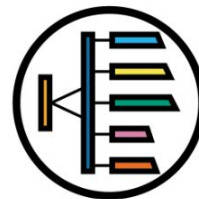
Bacterial 16S metabarcoding

Luca Lenzi and Matthew R. Gemmell

2022-08-22



Chapter 1 Introduction



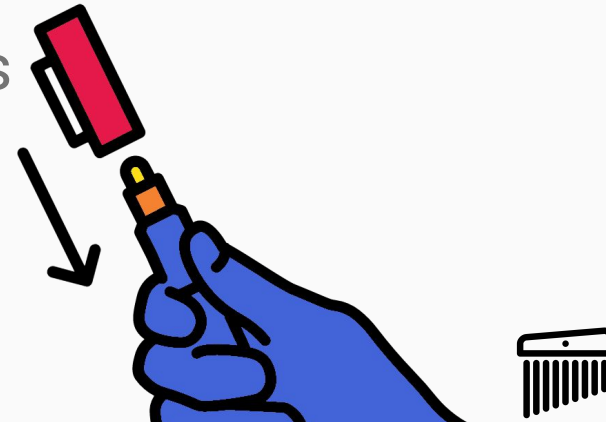
This practical session aims to introduce you to the analysis of bacterial 16S metabarcoding with QIIME2. The topics covered are:

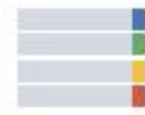
- [Background on the biology](#)
- [Introduction to QIIME2](#)
- [Cluster and webVNC information](#)
- [Information on initial data](#)
- [QIIME2 analysis workflow](#)



Recap

- Why 16S rRNA is used for microbial community analysis
- Qiime2 and its 16S rRNA workflow
- DADA2 quality control
- Alpha, Beta and Gamma Diversity analysis
- Relative abundance
- ANCOM & GNEISS





Segmentation fault





Thank you!

Questions?

