

HBP D11 OAES 2023 Sorting Subsample Experiment

Courtney Meier

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Goal

To determine the efficacy of subsampling various proportions of HBP samples at the D11 OAES site to reduce long sort times associated with removing OSD from current-year clipped biomass. **Subsampling is only evaluated in the context of clip harvests that do not require sorting to functional group** (i.e., non-peak biomass clips), because the subsample to total mass ratios will not apply to individual herbGroups.

Experimental Setup and Analyses

- Select n=10 plots (all plots for OAES), resulting in a maximum of n=20 clipID to test, due to both **exclosure = Y and N** for each plot. Random spatially-balanced plot locations, and locations of clipIDs within plots, will provide an unbiased estimate of biomass throughout the Tower airshed.
- For each clipID harvested in the field, test subsampling efficiency at various levels of sorting by creating subsamples (current-year + OSD) with the following percentages of the total **freshMass**:
 - 10%
 - 15%
 - 25%
 - 50%

The sum of all the subsamples = 100%; that is, the fresh mass of the entire clip strip.

- When subsampling is employed, calculate dryMass as follows: $dM = fM * (ssDM/ssFM)$, where:
 - dM = dryMass of current-year biomass in the clipID (no OSD)
 - fM = total freshMass in the clipID (current-year + OSD)
 - $ssDM$ = subsampleDryMass of current-year biomass in the subsample (no OSD)
 - $ssFM$ = fresh mass of all biomass in the subsample (current-year + OSD)
- Compare **dryMass** results calculated via subsampling with **dryMass** obtained with no subsampling, and use mixed effects models to analyze results.

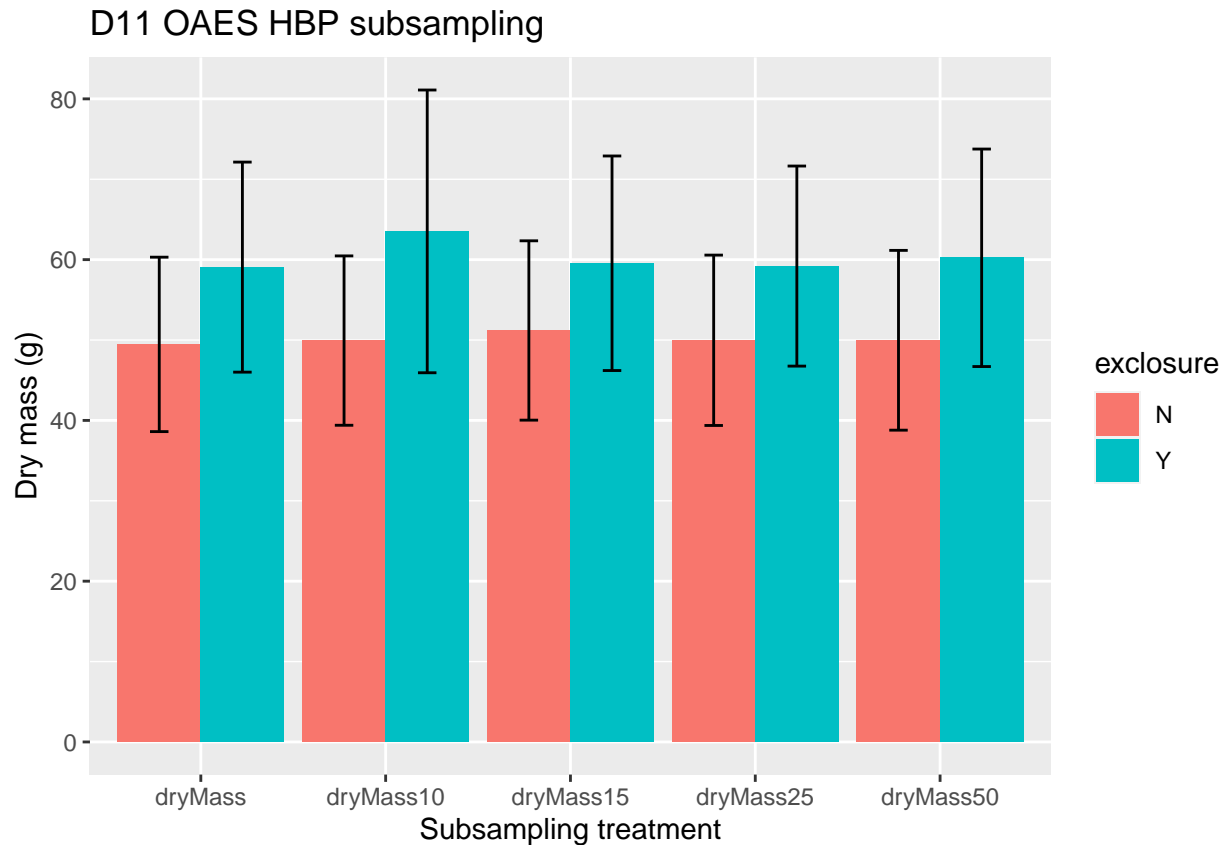
Procedure

1. Perform clip harvest in the field as normal, and bring clipped biomass back to the laboratory in cold storage as normal.
2. Identify up to n=20 clipIDs (n=10 for **exclosure = Y** and n=10 for **exclosure = N**), originating from each of the 10 plotIDs.
3. Thoroughly mix biomass from each clipID to homogenize as thoroughly as possible.
 - a. For large amounts of biomass, and when there is more than one bag of biomass for a given clipID, use a large bag, box, tray or equivalent vessel to mix the biomass.
4. For each clipID, weigh and record to 0.01 g:

- a. **freshMass** = total fresh mass in the clipID (current-year + OSD).
 - b. It is important that **freshMass** for the entire clipID and subsample fresh masses (below) are collected for a given clipID as close to each other in time as possible. That is, avoid weighing **freshMass** for a given clipID hours apart from the subsample fresh masses as water loss will affect the experimental results.
5. Based on the **freshMass**, calculate the desired subsample fresh masses for testing. For example, assuming **freshMass** = 100 g, the target subsample fresh masses are:
 - a. 10% subsample → 10 g
 - b. 15% subsample → 15 g
 - c. 25% subsample → 25 g
 - d. 50% subsample → 50 g
6. Label a coin envelope for each subsample above with the information below.
 - a. **subsampleTest**: 10%, 15%, 25% or 50%
 - b. **clipID**
 - c. **collectDate**
 - d. **exclosure**: Y/N
7. Weigh each fresh subsample created above (current-year + OSD), and record the information below.
 - a. **subsampleTest**: as above
 - b. **clipID**
 - c. **collectDate**
 - d. **exclosure**: Y/N
 - e. **subsampleFreshMass**: To the nearest 0.01 g; for subsamples < 0.5 g total mass, weigh to the nearest 0.0001 g
8. Sort current-year biomass from OSD for each subsample, and place sorted, current-year biomass into the corresponding labeled coin envelope.
 - a. Sorted OSD may be discarded at this point.
9. Dry subsamples until dry; minimum of 48 h @ 65 °C, track drying progress as normal.
10. Remove dry samples from the oven one at a time, and immediately weigh and record:
 - a. **subsampleDryMass**: To the nearest 0.01 g; for masses < 0.5 g, weigh to the nearest 0.0001 g.

Data summary

```
## `summarise()` has grouped output by 'treatment'. You can override using the
## `.groups` argument.
```



Results: Mixed-Effects model analysis

1. Models accounting for subsampling effect on **dryMass** and using **clipID** as a random effect (no need to use **enclosure** as a fixed effect since random effect accounts for variation across enclosure treatment).

Model: Data from eventID = "HBP.2023.OAES.24" only

```
## Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's
## method [lmerModLmerTest]
## Formula: estDryMass ~ treatment + (1 | clipID)
## Data: longDF %>% dplyr::filter(eventID == "HBP.2023.OAES.24")
##
##      AIC      BIC    logLik deviance df.resid
##    672.6    690.9   -329.3    658.6      93
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -4.9356 -0.3435 -0.0927  0.4176  3.8149
##
## Random effects:
##  Groups   Name      Variance Std.Dev.
## clipID   (Intercept) 199.77   14.134
## Residual                19.19    4.381
## Number of obs: 100, groups: clipID, 20
##
## Fixed effects:
##              Estimate Std. Error    df t value Pr(>|t|)
## (Intercept)    32.3385     3.3088 23.0967   9.774 1.13e-09 ***
```

```

## treatmentdryMass10 -0.0150      1.3852 80.0000 -0.011      0.991
## treatmentdryMass15  0.2005      1.3852 80.0000  0.145      0.885
## treatmentdryMass25  1.4835      1.3852 80.0000  1.071      0.287
## treatmentdryMass50  0.4175      1.3852 80.0000  0.301      0.764
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##           (Intr) trtM10 trtM15 trtM25
## trtmntdrM10 -0.209
## trtmntdrM15 -0.209  0.500
## trtmntdrM25 -0.209  0.500  0.500
## trtmntdrM50 -0.209  0.500  0.500  0.500

## Type III Analysis of Variance Table with Satterthwaite's method
##           Sum Sq Mean Sq NumDF DenDF F value Pr(>F)
## treatment 30.896   7.724     4     80  0.4025 0.8063

Model: Data from eventID = "HBP.2023.OAES.36" only

## Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's
## method [lmerModLmerTest]
## Formula: estDryMass ~ treatment + (1 | clipID)
## Data: longDF %>% dplyr::filter(eventID == "HBP.2023.OAES.36")
##
##           AIC          BIC    logLik deviance df.resid
##          840.2          858.0    -413.1    826.2         88
##
## Scaled residuals:
##           Min           1Q       Median           3Q            Max
## -2.8516 -0.2981  0.0556  0.1988  7.0689
##
## Random effects:
## Groups Name Variance Std.Dev.
## clipID (Intercept) 4707.1  68.61
## Residual          122.2  11.05
## Number of obs: 95, groups: clipID, 19
##
## Fixed effects:
##           Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)   77.5984    15.9427  19.7910   4.867 9.59e-05 ***
## treatmentdryMass10    5.1689     3.5862  76.0000   1.441   0.154
## treatmentdryMass15    2.0226     3.5862  76.0000   0.564   0.574
## treatmentdryMass25   -0.9163     3.5862  76.0000  -0.256   0.799
## treatmentdryMass50    1.2926     3.5862  76.0000   0.360   0.720
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##           (Intr) trtM10 trtM15 trtM25
## trtmntdrM10 -0.112
## trtmntdrM15 -0.112  0.500
## trtmntdrM25 -0.112  0.500  0.500
## trtmntdrM50 -0.112  0.500  0.500  0.500

## Type III Analysis of Variance Table with Satterthwaite's method

```

```

##          Sum Sq Mean Sq NumDF DenDF F value Pr(>F)
## treatment 415.43  103.86     4    76  0.8501  0.498

Model: Data from both eventIDs

## Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's
## method [lmerModLmerTest]
## Formula: estDryMass ~ treatment * eventID + (1 | clipID)
## Data: longDF
##
##      AIC      BIC    logLik deviance df.resid
## 1605.2   1644.5   -790.6   1581.2     183
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -3.7691 -0.2097  0.0132  0.2436  9.3973
##
## Random effects:
## Groups Name Variance Std.Dev.
## clipID (Intercept) 2395.63  48.945
## Residual          69.36   8.328
## Number of obs: 195, groups: clipID, 39
##
## Fixed effects:
##
##              Estimate Std. Error      df t value
## (Intercept)      32.3385     11.1018  40.8116   2.913
## treatmentdryMass10      -0.0150     2.6337 156.0000  -0.006
## treatmentdryMass15       0.2005     2.6337 156.0000   0.076
## treatmentdryMass25       1.4835     2.6337 156.0000   0.563
## treatmentdryMass50       0.4175     2.6337 156.0000   0.159
## eventIDHBP.2023.OAES.36    45.2599    15.9055  40.8116   2.846
## treatmentdryMass10:eventIDHBP.2023.OAES.36    5.1840     3.7733 156.0000   1.374
## treatmentdryMass15:eventIDHBP.2023.OAES.36    1.8221     3.7733 156.0000   0.483
## treatmentdryMass25:eventIDHBP.2023.OAES.36   -2.3998     3.7733 156.0000  -0.636
## treatmentdryMass50:eventIDHBP.2023.OAES.36    0.8751     3.7733 156.0000   0.232
##
##              Pr(>|t|)
## (Intercept)      0.00578 **
## treatmentdryMass10      0.99546
## treatmentdryMass15      0.93941
## treatmentdryMass25      0.57405
## treatmentdryMass50      0.87425
## eventIDHBP.2023.OAES.36    0.00690 **
## treatmentdryMass10:eventIDHBP.2023.OAES.36    0.17146
## treatmentdryMass15:eventIDHBP.2023.OAES.36    0.62984
## treatmentdryMass25:eventIDHBP.2023.OAES.36    0.52571
## treatmentdryMass50:eventIDHBP.2023.OAES.36    0.81690
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) trtM10 trtM15 trtM25 trtM50 eIDHBP tM10:I tM15:I tM25:I
## trtmntdrM10 -0.119
## trtmntdrM15 -0.119  0.500
## trtmntdrM25 -0.119  0.500  0.500
## trtmntdrM50 -0.119  0.500  0.500  0.500

```

```

## eIDHBP.2023 -0.698  0.083  0.083  0.083  0.083
## tM10:IDHBP.  0.083 -0.698 -0.349 -0.349 -0.349 -0.119
## tM15:IDHBP.  0.083 -0.349 -0.698 -0.349 -0.349 -0.119  0.500
## tM25:IDHBP.  0.083 -0.349 -0.349 -0.698 -0.349 -0.119  0.500  0.500
## tM50:IDHBP.  0.083 -0.349 -0.349 -0.349 -0.698 -0.119  0.500  0.500  0.500

## Type III Analysis of Variance Table with Satterthwaite's method
##               Sum Sq Mean Sq NumDF DenDF F value   Pr(>F)
## treatment      156.97   39.24     4    156  0.5658 0.687851
## eventID        602.74  602.74     1     39  8.6897 0.005381 **
## treatment:eventID 299.22   74.80     4    156  1.0785 0.369134
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

2. Null model, and using **clipID** as a random effect. Theoretically, if all subsampling results in the same dryMass as the full sort, the null model should be no different than the “treatment” model.

```
## Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's
## method [lmerModLmerTest]
## Formula: estDryMass ~ 1 + (1 | clipID)
## Data: longDF
##
##      AIC      BIC   logLik deviance df.resid
##  1601.3   1611.2   -797.7   1595.3      192
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -3.9879 -0.1959 -0.0171  0.1825  9.6311
##
## Random effects:
##  Groups   Name                Variance Std.Dev.
## clipID   (Intercept)  2931.93    54.147
## Residual                  72.22     8.498
## Number of obs: 195, groups: clipID, 39
##
## Fixed effects:
##              Estimate Std. Error    df t value Pr(>|t|)
## (Intercept)    55.340      8.692 39.000   6.367 1.6e-07 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

## Data: longDF
## Models:
## m4ME: estDryMass ~ 1 + (1 | clipID)
## m3ME: estDryMass ~ treatment * eventID + (1 | clipID)
##      npar    AIC    BIC  logLik deviance  Chisq Df Pr(>Chisq)
## m4ME     3 1601.3 1611.2 -797.67   1595.3
## m3ME    12 1605.2 1644.5 -790.60   1581.2 14.151  9    0.1171
```

3. Linear model, using treatment*exclosure, and no random effect.

```
##
## Call:
## lm(formula = estDryMass ~ exclosure * treatment + eventID, data = longDF)
##
## Residuals:
```

	Min	1Q	Median	3Q	Max
	-79.356	-20.643	-5.911	8.659	292.004

```
##
## Coefficients:
```

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	27.6124	12.1799	2.267	0.0246 *
exclosureY	8.3994	16.3104	0.515	0.6072
treatmentdryMass10	0.4747	16.5170	0.029	0.9771
treatmentdryMass15	1.7300	16.5170	0.105	0.9167
treatmentdryMass25	0.5095	16.5170	0.031	0.9754
treatmentdryMass50	0.5121	16.5170	0.031	0.9753
eventIDHBP.2023.OAES.36	46.1195	7.2963	6.321	1.91e-09 ***
exclosureY:treatmentdryMass10	3.9698	23.0648	0.172	0.8635
exclosureY:treatmentdryMass15	-1.2515	23.0648	-0.054	0.9568
exclosureY:treatmentdryMass25	-0.3805	23.0648	-0.016	0.9869
exclosureY:treatmentdryMass50	0.6469	23.0648	0.028	0.9777

```
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 50.91 on 184 degrees of freedom
## Multiple R-squared:  0.186, Adjusted R-squared:  0.1417
## F-statistic: 4.203 on 10 and 184 DF, p-value: 2.911e-05
## Analysis of Variance Table
##
## Response: estDryMass
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
exclosure	1	5079	5079	1.9595	0.1632
treatment	4	147	37	0.0142	0.9996
eventID	1	103552	103552	39.9548	1.912e-09 ***
exclosure:treatment	4	157	39	0.0151	0.9995
Residuals	184	476876	2592		

```
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```


Summary of subsampling results

Table 1: Difference in estimated dryMass to total dryMass for subsampling (%)

eventID	ss10	ss15	ss25	ss50
HBP.2023.OAES.24	0.0	0.6	4.6	1.3
HBP.2023.OAES.36	6.7	2.6	-1.2	1.7

Conclusions

Summary of D11 OAES results:

1. Random effect associated with **clipID** explains considerable variability in the data. See summary outputs for all mixed-effects models.
2. Summary output for Mixed Effects model indicates that D11 OAES Clip Strips can be subsampled at the 15% level for all bouts with **herbGroup** = ALL.
 - Comparing results from two separate bouts, estimates of dryMass using a 10% subsample are more variable relative to total dryMass from the full sort compared to other subsampling levels, even though the difference is not significant.
 - When eventIDs are analyzed separately (m1ME, m2ME), other subsamples are not significantly different than the entire sorted subsample ($p > 0.29$) \rightarrow go with 15% subsample.
 - With both eventIDs analyzed together, there are no differences between the full sort dryMass and estimated dryMass from subsampling ($p = 0.68$).
3. Effect of **exclosure*treatment**, as evaluated in standard linear model (m1LM):
 - Effect of **exclosure** is not significant ($p = 0.16$).
 - Effect of **treatment** not significant ($p = 0.99$).
 - No detection of **exclosure:treatment** interaction effect ($p = 0.99$).
 - Effect of **eventID** highly significant ($p < 0.0001$)