

HBP D11 CLBJ 2023 Sorting Subsample Experiment

Courtney Meier

2023-10-03

Goal

To determine the efficacy of subsampling various proportions of HBP samples at the D11 CLBJ site to reduce long sort times associated with removing OSD from current-year clipped biomass. **Subsampling is only evaluated in the context of clip harvests that do not require sorting to functional group** (i.e., non-peak biomass clips), because the subsample to total mass ratios will not apply to individual herbGroups.

Experimental Setup and Analyses

- Select $n=10$ plots (all plots for OAES), resulting in a maximum of $n=20$ clipID to test, due to both **exclosure = Y and N** for each plot. Random spatially-balanced plot locations, and locations of clipIDs within plots, will provide an unbiased estimate of biomass throughout the Tower airshed.
- For each clipID harvested in the field, test subsampling efficiency at various levels of sorting by creating subsamples (current-year + OSD) with the following percentages of the total **freshMass**:
 - 10%
 - 15%
 - 25%
 - 50%

The sum of all the subsamples = 100%; that is, the fresh mass of the entire clip strip.

- When subsampling is employed, calculate dryMass as follows: $dM = fM * (ssDM/ssFM)$, where:
 - dM = dryMass of current-year biomass in the clipID (no OSD)
 - fM = total freshMass in the clipID (current-year + OSD)
 - $ssDM$ = subsampleDryMass of current-year biomass in the subsample (no OSD)
 - $ssFM$ = fresh mass of all biomass in the subsample (current-year + OSD)
- Compare **dryMass** results calculated via subsampling with **dryMass** obtained with no subsampling, and use mixed effects models to analyze results.

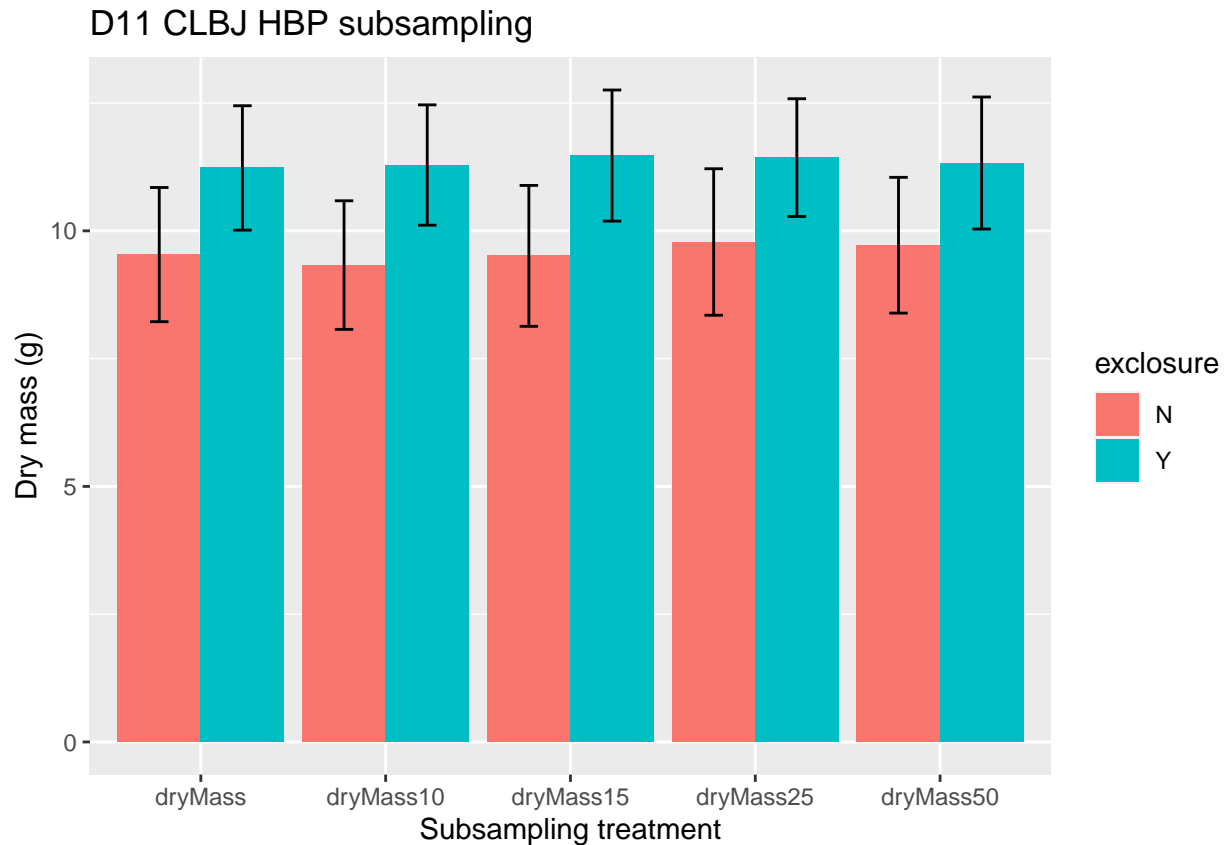
Procedure

1. Perform clip harvest in the field as normal, and bring clipped biomass back to the laboratory in cold storage as normal.
2. Identify up to $n=20$ clipIDs ($n=10$ for **exclosure = Y** and $n=10$ for **exclosure = N**), originating from each of the 10 plotIDs.
3. Thoroughly mix biomass from each clipID to homogenize as thoroughly as possible.
 - a. For large amounts of biomass, and when there is more than one bag of biomass for a given clipID, use a large bag, box, tray or equivalent vessel to mix the biomass.
4. For each clipID, weigh and record to 0.01 g:

- a. **freshMass** = total fresh mass in the clipID (current-year + OSD).
 - b. It is important that **freshMass** for the entire clipID and subsample fresh masses (below) are collected for a given clipID as close to each other in time as possible. That is, avoid weighing **freshMass** for a given clipID hours apart from the subsample fresh masses as water loss will affect the experimental results.
5. Based on the **freshMass**, calculate the desired subsample fresh masses for testing. For example, assuming **freshMass** = 100 g, the target subsample fresh masses are:
 - a. 10% subsample → 10 g
 - b. 15% subsample → 15 g
 - c. 25% subsample → 25 g
 - d. 50% subsample → 50 g
6. Label a coin envelope for each subsample above with the information below.
 - a. **subsampleTest**: 10%, 15%, 25% or 50%
 - b. **clipID**
 - c. **collectDate**
 - d. **exclosure**: Y/N
7. Weigh each fresh subsample created above (current-year + OSD), and record the information below.
 - a. **subsampleTest**: as above
 - b. **clipID**
 - c. **collectDate**
 - d. **exclosure**: Y/N
 - e. **subsampleFreshMass**: To the nearest 0.01 g; for subsamples < 0.5 g total mass, weigh to the nearest 0.0001 g
8. Sort current-year biomass from OSD for each subsample, and place sorted, current-year biomass into the corresponding labeled coin envelope.
 - a. Sorted OSD may be discarded at this point.
9. Dry subsamples until dry; minimum of 48 h @ 65 °C, track drying progress as normal.
10. Remove dry samples from the oven one at a time, and immediately weigh and record:
 - a. **subsampleDryMass**: To the nearest 0.01 g; for masses < 0.5 g, weigh to the nearest 0.0001 g.

Data summary

```
## `summarise()` has grouped output by 'treatment'. You can override using the
## `.groups` argument.
```



Results: Mixed-Effects model analysis

1. Models accounting for subsampling effect on **dryMass** and using **clipID** as a random effect (no need to use **enclosure** as a fixed effect since random effect accounts for variation across enclosure treatment).

Model: Data from eventID = "HBP.2023.CLBJ.18" only

```
## Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's
## method [lmerModLmerTest]
## Formula: estDryMass ~ treatment + (1 | clipID)
## Data: longDF %>% dplyr::filter(eventID == "HBP.2023.CLBJ.18")
##
##      AIC      BIC    logLik deviance df.resid
##    370.5    387.6   -178.3    356.5      78
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -3.5230 -0.3012 -0.0772  0.3370  2.3320
##
## Random effects:
##  Groups   Name                Variance Std.Dev.
## clipID   (Intercept)    32.466     5.698
## Residual                  1.523     1.234
## Number of obs: 85, groups: clipID, 17
##
## Fixed effects:
##              Estimate Std. Error    df t value Pr(>|t|)
## (Intercept)    9.32412    1.41398 18.28119   6.594 3.16e-06 ***
```

```

## treatmentdryMass10 -0.57647    0.42328 68.00000 -1.362    0.178
## treatmentdryMass15 -0.05882    0.42328 68.00000 -0.139    0.890
## treatmentdryMass25  0.24118    0.42328 68.00000  0.570    0.571
## treatmentdryMass50  0.19824    0.42328 68.00000  0.468    0.641
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##           (Intr) trtM10 trtM15 trtM25
## trtmntdrM10 -0.150
## trtmntdrM15 -0.150  0.500
## trtmntdrM25 -0.150  0.500  0.500
## trtmntdrM50 -0.150  0.500  0.500  0.500

## Type III Analysis of Variance Table with Satterthwaite's method
##           Sum Sq Mean Sq NumDF DenDF F value Pr(>F)
## treatment  7.2347  1.8087     4    68  1.1877 0.3241

Model: Data from eventID = "HBP.2023.CLBJ.34" only

## Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's
## method [lmerModLmerTest]
## Formula: estDryMass ~ treatment + (1 | clipID)
## Data: longDF %>% dplyr::filter(eventID == "HBP.2023.CLBJ.34")
##
##           AIC          BIC    logLik deviance df.resid
##          262.0          279.1   -124.0    248.0         78
##
## Scaled residuals:
##           Min           1Q       Median           3Q          Max
## -2.95982 -0.46198  0.00238  0.46201  2.41570
##
## Random effects:
## Groups Name Variance Std.Dev.
## clipID (Intercept) 18.2862  4.2762
## Residual          0.3568  0.5973
## Number of obs: 85, groups: clipID, 17
##
## Fixed effects:
##           Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)  11.34176    1.04721 17.53165  10.830 3.42e-09 ***
## treatmentdryMass10  0.41294    0.20487 68.00000   2.016  0.0478 *
## treatmentdryMass15  0.26000    0.20487 68.00000   1.269  0.2087
## treatmentdryMass25  0.21000    0.20487 68.00000   1.025  0.3090
## treatmentdryMass50  0.08647    0.20487 68.00000   0.422  0.6743
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##           (Intr) trtM10 trtM15 trtM25
## trtmntdrM10 -0.098
## trtmntdrM15 -0.098  0.500
## trtmntdrM25 -0.098  0.500  0.500
## trtmntdrM50 -0.098  0.500  0.500  0.500

## Type III Analysis of Variance Table with Satterthwaite's method

```

```

##          Sum Sq Mean Sq NumDF DenDF F value Pr(>F)
## treatment 1.7297 0.43242      4    68  1.2121 0.3138

Model: Data from both eventIDs

## Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's
## method [lmerModLmerTest]
## Formula: estDryMass ~ treatment * eventID + (1 | clipID)
## Data: longDF
##
##      AIC      BIC    logLik deviance df.resid
##    662.9    700.5   -319.5    638.9     158
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -4.4974 -0.3549 -0.0658  0.3285  2.9534
##
## Random effects:
## Groups Name Variance Std.Dev.
## clipID (Intercept) 25.3761  5.0375
## Residual          0.9398  0.9694
## Number of obs: 170, groups: clipID, 34
##
## Fixed effects:
##
##              Estimate Std. Error      df
## (Intercept)      9.32412    1.24418  36.02155
## treatmentdryMass10 -0.57647    0.33252 136.00000
## treatmentdryMass15 -0.05882    0.33252 136.00000
## treatmentdryMass25  0.24118    0.33252 136.00000
## treatmentdryMass50  0.19824    0.33252 136.00000
## eventIDHBP.2023.CLB.J.34 2.01765    1.75954  36.02155
## treatmentdryMass10:eventIDHBP.2023.CLB.J.34 0.98941    0.47025 136.00000
## treatmentdryMass15:eventIDHBP.2023.CLB.J.34 0.31882    0.47025 136.00000
## treatmentdryMass25:eventIDHBP.2023.CLB.J.34 -0.03118    0.47025 136.00000
## treatmentdryMass50:eventIDHBP.2023.CLB.J.34 -0.11176    0.47025 136.00000
##
##              t value Pr(>|t|)
## (Intercept)      7.494 7.43e-09 ***
## treatmentdryMass10 -1.734  0.0852 .
## treatmentdryMass15 -0.177  0.8598
## treatmentdryMass25  0.725  0.4695
## treatmentdryMass50  0.596  0.5521
## eventIDHBP.2023.CLB.J.34 1.147  0.2591
## treatmentdryMass10:eventIDHBP.2023.CLB.J.34 2.104  0.0372 *
## treatmentdryMass15:eventIDHBP.2023.CLB.J.34 0.678  0.4989
## treatmentdryMass25:eventIDHBP.2023.CLB.J.34 -0.066  0.9472
## treatmentdryMass50:eventIDHBP.2023.CLB.J.34 -0.238  0.8125
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) trtM10 trtM15 trtM25 trtM50 eIDHBP tM10:I tM15:I tM25:I
## trtmntdrM10 -0.134
## trtmntdrM15 -0.134  0.500
## trtmntdrM25 -0.134  0.500  0.500
## trtmntdrM50 -0.134  0.500  0.500  0.500

```

```

## eIDHBP.2023 -0.707  0.094  0.094  0.094  0.094
## tM10:IDHBP.  0.094 -0.707 -0.354 -0.354 -0.354 -0.134
## tM15:IDHBP.  0.094 -0.354 -0.707 -0.354 -0.354 -0.134  0.500
## tM25:IDHBP.  0.094 -0.354 -0.354 -0.707 -0.354 -0.134  0.500  0.500
## tM50:IDHBP.  0.094 -0.354 -0.354 -0.354 -0.707 -0.134  0.500  0.500  0.500

## Type III Analysis of Variance Table with Satterthwaite's method
##               Sum Sq Mean Sq NumDF DenDF F value Pr(>F)
## treatment      1.9734  0.49334     4   136   0.5249  0.7176
## eventID         1.5830  1.58297     1    34   1.6843  0.2031
## treatment:eventID 6.9910  1.74774     4   136   1.8597  0.1211

```

2. Null model, and using **clipID** as a random effect. Theoretically, if all subsampling results in the same dryMass as the full sort, the null model should be no different than the “treatment” model.

```
## Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's
## method [lmerModLmerTest]
## Formula: estDryMass ~ 1 + (1 | clipID)
## Data: longDF
##
##      AIC      BIC    logLik deviance df.resid
##    655.8    665.2   -324.9    649.8      167
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -4.8908 -0.2938 -0.0651  0.2928  3.1272
##
## Random effects:
##  Groups   Name                Variance Std.Dev.
## clipID   (Intercept)  26.629     5.160
## Residual                    1.006     1.003
## Number of obs: 170, groups: clipID, 34
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)  10.4103      0.8883  34.0000   11.72 1.73e-13 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

## Data: longDF
## Models:
## m4ME: estDryMass ~ 1 + (1 | clipID)
## m3ME: estDryMass ~ treatment * eventID + (1 | clipID)
##      npar    AIC    BIC  logLik deviance Chisq Df Pr(>Chisq)
## m4ME     3 655.78 665.19 -324.89   649.78
## m3ME    12 662.92 700.55 -319.46   638.92 10.863  9    0.2852
```

3. Linear model, using treatment*exclosure, and no random effect.

```
##
## Call:
## lm(formula = estDryMass ~ exclosure * treatment + eventID, data = longDF)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -8.7129 -4.0036 -0.4638  3.3652 14.4429
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      8.40965     1.29592    6.489 1.04e-09 ***
## exclosureY        1.69562     1.79640    0.944  0.34665
## treatmentdryMass10 -0.20500     1.74276   -0.118  0.90651
## treatmentdryMass15 -0.02500     1.74276   -0.014  0.98857
## treatmentdryMass25  0.24667     1.74276    0.142  0.88762
## treatmentdryMass50  0.18333     1.74276    0.105  0.91635
## eventIDHBP.2023.CLBJ.34 2.25071     0.80198    2.806  0.00564 **
## exclosureY:treatmentdryMass10 0.26188     2.54049    0.103  0.91803
## exclosureY:treatmentdryMass15 0.26688     2.54049    0.105  0.91647
## exclosureY:treatmentdryMass25 -0.04479     2.54049   -0.018  0.98596
## exclosureY:treatmentdryMass50 -0.08708     2.54049   -0.034  0.97270
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 5.228 on 159 degrees of freedom
## Multiple R-squared:  0.07486,    Adjusted R-squared:  0.01668
## F-statistic: 1.287 on 10 and 159 DF,  p-value: 0.2421
##
## Analysis of Variance Table
##
## Response: estDryMass
##              Df Sum Sq Mean Sq F value    Pr(>F)
## exclosure      1  133.4  133.438   4.8816 0.028574 *
## treatment      4    2.0    0.493   0.0180 0.999356
## eventID        1  215.3  215.291   7.8760 0.005636 **
## exclosure:treatment  4    1.0    0.250   0.0091 0.999833
## Residuals     159 4346.3   27.335
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```


Summary of subsampling results

Table 1: Difference in estimated dryMass to total dryMass for subsampling (%)

eventID	ss10	ss15	ss25	ss50
HBP.2023.CLBJ.18	-6.2	-0.6	2.6	2.10
HBP.2023.CLBJ.34	3.6	2.3	1.9	0.76

Conclusions

Summary of D11 CLBJ results:

1. Random effect associated with **clipID** explains considerable variability in the data. See summary outputs for all mixed-effects models.
2. Summary output for Mixed Effects model indicates that D11 CLBJ Clip Strips can be subsampled at the 15% level for all bouts with **herbGroup** = ALL.
 - Comparing results from two separate bouts, estimates of dryMass using a 10% subsample are more variable relative to total dryMass from the full sort compared to other subsampling levels.
 - Estimated dryMass from the 10% subsample is significantly different than the full sort for eventID = "HBP.2023.CLBJ.34" ($p = 0.05$)
 - When eventIDs are analyzed separately (m1ME, m2ME), other subsamples are not significantly different than the entire sorted subsample ($p > 0.22$) \rightarrow go with 15% subsample.
 - With both eventIDs analyzed together, there are no differences between the full sort dryMass and estimated dryMass from subsampling ($p = 0.74$).
3. Effect of **exclosure*treatment**, as evaluated in standard linear model (m1LM):
 - Effect of **exclosure** is significant ($p < 0.05$).
 - Effect of **treatment** not significant ($p = 0.99$).
 - No detection of **exclosure:treatment** interaction effect ($p = 0.99$).