



# Missing Values in Clinical Research (EP16)

## Multiple Imputation

**Nicole Erler**

Department of Biostatistics, Erasmus MC  
[n.erler@erasmusmc.nl](mailto:n.erler@erasmusmc.nl)

14 - 18 May, 2018

**Erasmus MC**  
University Medical Center Rotterdam



## Part I: Multiple Imputation

How does multiple imputation work?

- The ideas behind MI
- Understanding sources of uncertainty
- Implementation of MI and MICE

## Part II: Multiple Imputation Workflow

How to perform MI with the **mice** package in R, from getting to know the data to the final results.

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**Practical:** Imputation with **mice**

# Outline (cont.)

## Part III: When MICE might fail

### Introduction to

- settings where standard use of **mice** is problematic
- alternative imputation approaches
- alternative R packages

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**Practical:** Imputation in complex settings

## Part IV: Multiple Imputation Strategies

Some tips & tricks

# Part I

## Multiple Imputation

# Outline of Part I

1. What is Multiple Imputation?
  - 1.1 History & Ideas
  - 1.2 Three steps
2. Imputation step
  - 2.1 Univariate missing data
  - 2.2 Multivariate missing data
  - 2.3 FCS/MICE
  - 2.4 Checking convergence
3. Analysis step
4. Pooling
  - 4.1 Why pooling?
  - 4.2 Rubin's Rules
5. A closer look at the imputation step
  - 5.1 Bayesian multiple imputation
  - 5.2 Bootstrap multiple imputation
  - 5.3 Semi-parametric imputation
  - 5.4 What is implemented in software?

# 1. What is Multiple Imputation?

## 1.1. History & Ideas

- Developed by **Donald B. Rubin** in the 1970s,
- to handle missing values in **public use databases**,  
e.g., census data provided by the government,
- motivated by the **increase in missing values**, and
- increased **availability of computers**.

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- increased **availability of computers**.

Such data should be usable by [11]

- a **large number of analysts**, who commonly have to rely on
- standard **software that can only handle complete data**, and usually
- are **not experts in handling incomplete data**.

# 1. What is Multiple Imputation?

## 1.1. History & Ideas

### Rubin's thoughts: [12]

One imputed value can not be correct in general.

➡ We need to represent missing values by a **number of imputations**.

To find **sensible values** to fill in, we need some kind of **model**.



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This **distribution depends on assumptions** that have been made about the model.



What we want to impute is the **'predictive distribution'** of the missing values given the observed values.

# 1. What is Multiple Imputation?

## 1.1. History & Ideas

### **How to obtain that predictive distribution?**

Rubin suggests to

- fit a model to the observed data (“respondents”), and to
  - obtain for each “nonrespondent” the conditional distribution of the missing data (given the observed data) as if he/she was a respondent.
- ➡ We assume nonrespondents are just like respondents, and obtain the predictive distribution from the model of the respondents data.

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### How to obtain that predictive distribution?

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#### **Example:** survey about age, gender and height

Boys aged 10 – 12 years old answered (on average) that they are 1.45m tall.

- ➡ We assume that boys aged 10 to 12 who did not report their height are also around 1.45m tall.

# 1. What is Multiple Imputation?

## 1.1. History & Ideas

### How to represent the multiple imputed values?

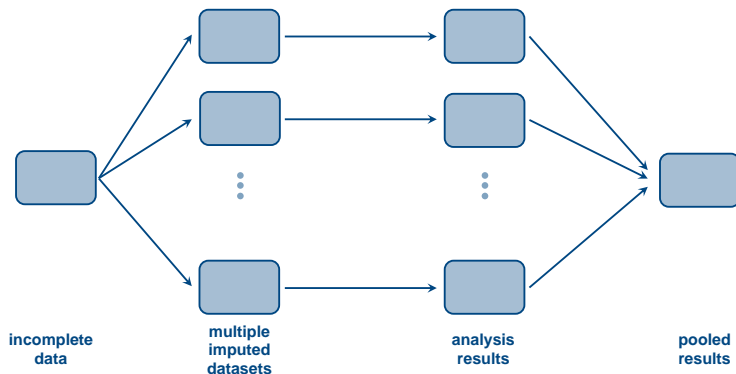
For each missing value, we now have multiple imputed values.

- For each set of imputed values, create a dataset (those datasets agree in the observed values but imputed values differ).
- Analyse each dataset, and
- take the results from each analysis.

➡ We can describe how (much) the **results vary between the imputed datasets**, and calculate summary measures.

# 1. What is Multiple Imputation?

## 1.2. Three steps



### In summary:

1. **Imputation:** impute multiple times ➡ multiple completed datasets
2. **Analysis:** analyse each of the datasets
3. **Pooling:** combine results, taking into account additional uncertainty

## 2. Imputation step

### 2.1. Univariate missing data

#### How can we actually get imputed values?

For now: assume only one continuous variable has missing values (**univariate missing data**)

$X_1$	$X_2$	$X_3$	$X_4$
✓	NA	✓	✓
✓	✓	✓	✓
✓	NA	✓	✓
⋮	⋮	⋮	⋮

## 2. Imputation step

### 2.1. Univariate missing data

#### How can we actually get imputed values?

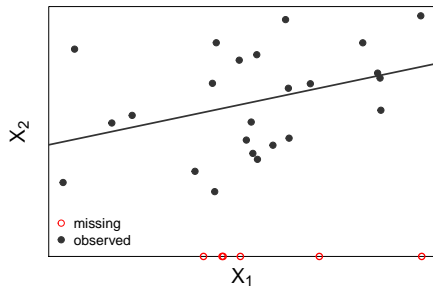
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**Idea:** Predict values

Model:

$$x_{i2} = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i3} + \beta_3 x_{i4} + \varepsilon_i$$





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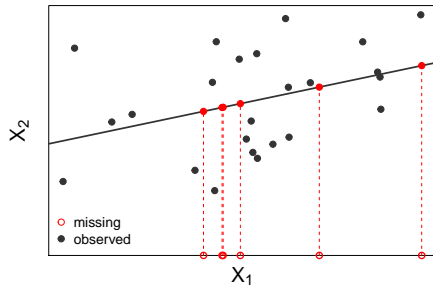
**Idea:** Predict values

Model:

$$x_{i2} = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i3} + \beta_3 x_{i4} + \varepsilon_i$$

Imputed/predicted value:

$$\hat{x}_{i2} = \hat{\beta}_0 + \hat{\beta}_1 x_{i1} + \hat{\beta}_2 x_{i3} + \hat{\beta}_3 x_{i4}$$



## 2. Imputation step

### 2.1. Univariate missing data

#### Problem:

- We can obtain **only one imputed value** per missing value, but we wanted a whole distribution.
- The predicted values do not take into account the added **uncertainty** due to the missing values.

## 2. Imputation step

### 2.1. Univariate missing data

#### Problem:

- We can obtain **only one imputed value** per missing value, but we wanted a whole distribution.
  - The predicted values do not take into account the added **uncertainty** due to the missing values.
- ➔ We need to take into account **two sources of uncertainty**:
- The **parameters** are estimated with **uncertainty** (represented by the std. error).
  - There is **random variation / prediction error** (variation of the residuals).

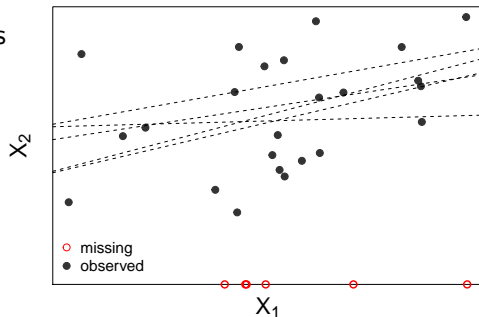
## 2. Imputation step

### 2.1. Univariate missing data

#### Taking into account uncertainty about the parameters $\beta$ :

We assume that  $\beta$  has a **distribution**, and we can sample realizations of  $\beta$  from that distribution.

When plugging the different realizations of  $\beta$  into the predictive model, we obtain **slightly different regression lines**.



## 2. Imputation step

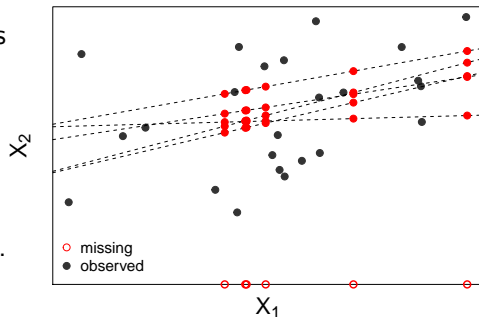
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We assume that  $\beta$  **has a distribution**, and we can sample realizations of  $\beta$  from that distribution.

When plugging the different realizations of  $\beta$  into the predictive model, we obtain **slightly different regression lines**.

With each set of coefficients, we also get slightly **different predicted values**.



## 2. Imputation step

### 2.1. Univariate missing data

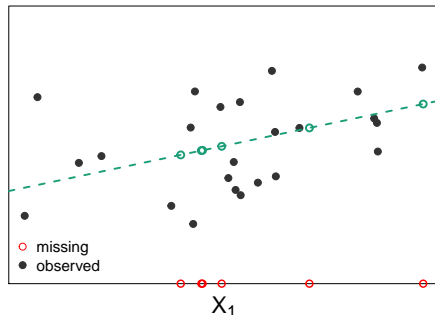
#### Taking into account the prediction error:

The model does not fit the data perfectly: observations are scattered around the regression lines.

We assume that the **data have a distribution**, where

- the **mean** for each value is given by the **predictive model**, and

$X_2$



## 2. Imputation step

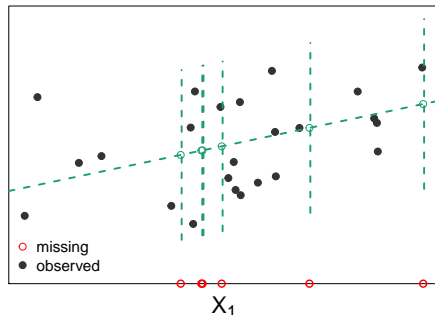
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We assume that the **data have a distribution**, where

- the **mean** for each value is given by the **predictive model**, and
- the **variance** is determined by the variance of the residuals  $\epsilon$ .



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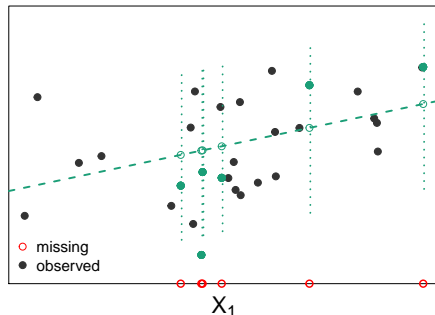
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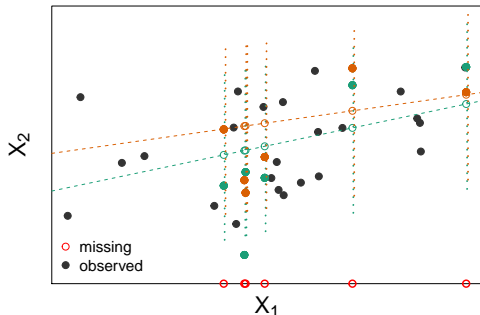
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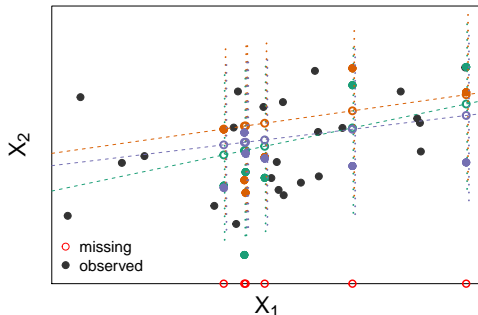
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## 2. Imputation step

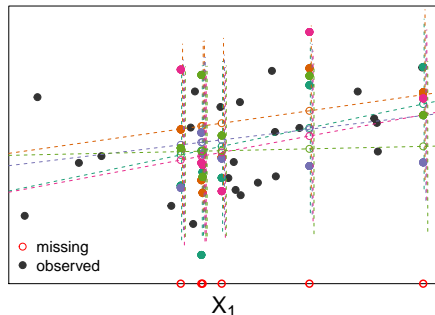
### 2.1. Univariate missing data

#### Taking into account the prediction error:

The model does not fit the data perfectly: observations are scattered around the regression lines.

We assume that the **data have a distribution**, where

- the **mean** for each value is given by the **predictive model**, and
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In the end, we obtain one imputed dataset for each color.

## 2. Imputation step

### 2.2. Multivariate missing data

#### **Multivariate missing data:**

What if we have **missing values in more than one variable?**

## 2. Imputation step

### 2.2. Multivariate missing data

#### Multivariate missing data:

What if we have **missing values in more than one variable**?

In case of **monotone missing values** we can use the technique for univariate missing data in a chain:

impute  $x_4$  given  $x_1$

impute  $x_3$  given  $x_1$  and  $x_4$

impute  $x_2$  given  $x_1$ ,  $x_4$  and  $x_3$

$X_1$	$X_2$	$X_3$	$X_4$
✓	NA	✓	✓
✓	NA	NA	✓
✓	NA	NA	NA
⋮	⋮	⋮	⋮

## 2. Imputation step

### 2.2. Multivariate missing data

#### Multivariate missing data:

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$X_1$	$X_2$	$X_3$	$X_4$
✓	NA	✓	✓
✓	NA	NA	✓
✓	NA	NA	NA
⋮	⋮	⋮	⋮

When we have **non-monotone missing data** there is no sequence without conditioning on unobserved values.

$X_1$	$X_2$	$X_3$	$X_4$
✓	NA	✓	✓
NA	✓	NA	NA
✓	NA	✓	NA
⋮	⋮	⋮	⋮

## 2. Imputation step

### 2.2. Multivariate missing data

There are **two popular approaches** for the imputation step in **multivariate non-monotone** missing data:

#### Fully conditional specification

- Multiple Imputation using Chained Equations (**MICE**)
- sometimes also: sequential regression
- Implemented in SPSS, R, Stata, SAS, ...
- our focus here

## 2. Imputation step

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#### Joint model imputation

(more details later)



## 2. Imputation step

### 2.2. Multivariate missing data

#### Markov Chain Monte Carlo

is a technique to **draw samples from a complex probability distribution** by creating a chain of random variables (a Markov Chain). The distribution each element in the chain is sampled from depends on the value of the previous element. When certain conditions are met, the chain eventually stabilizes and by continuing to sample elements of the chain a sample from the complex distribution of interest can be obtained.

## 2. Imputation step

### 2.2. Multivariate missing data

#### Markov Chain Monte Carlo

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#### Gibbs sampling

is an MCMC method where a **sample from a multivariate distribution** is obtained by repeatedly drawing from each of the univariate full conditional distributions instead.

## 2. Imputation step

### 2.3. FCS/MICE

**MICE** (**M**ultiple **I**mputation using **C**hained **E**quations) or  
**FCS** (multiple imputation using **F**ully **C**onditional **S**pecification)

extends univariable imputation to the setting with multivariate non-monotone missingness:

MICE/FCS

- imputes multivariate missing data on a variable-by-variable basis,
- using the technique for univariable missing data.

## 2. Imputation step

### 2.3. FCS/MICE

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extends univariable imputation to the setting with multivariate non-monotone missingness:

#### MICE/FCS

- imputes multivariate missing data on a variable-by-variable basis,
- using the technique for univariable missing data.

Moreover, MICE/FCS is

- an iterative procedure, specifically
- a Markov Chain Monte Carlo (MCMC) method,
- uses the idea of the Gibbs sampler, and
- is a Gibbs sampler if the conditional distributions are compatible (we will come back to this)

## 2. Imputation step

### 2.3. FCS/MICE

#### Notation

- $X$ :  $n \times p$  data matrix with  $n$  rows and  $p$  variables  $x_1, \dots, x_p$
- $R$ :  $n \times p$  missing indicator matrix containing 0 (missing) or 1 (observed)

$$\mathbf{X} = \begin{array}{c|ccc} & X_{-2} & X_2 & X_{-2} \\ \hline x_{1,1} & x_{1,2} & \dots & x_{1,p} \\ x_{2,1} & x_{2,2} & \dots & x_{2,p} \\ \vdots & \vdots & \ddots & \vdots \\ x_{n,1} & x_{n,2} & \dots & x_{n,p} \end{array}$$

$$\mathbf{R} = \begin{array}{cccc} R_{1,1} & R_{1,2} & \dots & R_{1,p} \\ R_{2,1} & R_{2,2} & \dots & R_{2,p} \\ \vdots & \vdots & \ddots & \vdots \\ R_{n,1} & R_{n,2} & \dots & R_{n,p} \end{array}$$

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For example:

$$X = \begin{array}{c|cccc} & X_1 & X_2 & X_3 & X_4 \\ \hline \checkmark & \text{NA} & \checkmark & \checkmark \\ \checkmark & \checkmark & \text{NA} & \text{NA} \\ \checkmark & \text{NA} & \checkmark & \text{NA} \end{array}$$

$$\Rightarrow R = \begin{array}{c|cccc} & 1 & 0 & 1 & 1 \\ \hline 1 & 1 & 0 & 0 \\ 1 & 0 & 1 & 0 \end{array}$$

## 2. Imputation step

### 2.3. FCS/MICE

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**Algorithm 1** MICE algorithm [17] for **one** imputed dataset

---

- 1: **for**  $j$  in  $1, \dots, p$ : ▷ Setup
- 2:     Specify imputation model for variable  $X_j$   
       $p(X_j^{mis} \mid X_j^{obs}, X_{-j}, R)$
- 3:     Fill in starting imputations  $\dot{X}_j^0$  by random draws from  $X_j^{obs}$ .
- 4: **end for**

## 2. Imputation step

### 2.3. FCS/MICE

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5: for  $t$  in  $1, \dots, T$ : ▷ loop through iterations
6:   for  $j$  in  $1, \dots, p$ : ▷ loop through variables

10:   end for
11: end for
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---



## 2. Imputation step

### 2.3. FCS/MICE

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5: for  $t$  in  $1, \dots, T$ : ▷ loop through iterations
6:   for  $j$  in  $1, \dots, p$ : ▷ loop through variables
7:     Define currently complete data except  $X_j$ 
      $\dot{X}_{-j}^t = (\dot{X}_1^t, \dots, \dot{X}_{j-1}^t, \dot{X}_{j+1}^{t-1}, \dots, \dot{X}_p^{t-1})$ .

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## 2. Imputation step

### 2.3. FCS/MICE

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8:     Draw parameters  $\dot{\theta}_j^t \sim p(\theta_j^t \mid X_j^{obs}, \dot{X}_{-j}^t, R)$ .

10:   end for
11: end for
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## 2. Imputation step

### 2.3. FCS/MICE

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4: end for

5: for  $t = 1$ : ▷ loop through iterations
6:   for  $j = 1$ : ▷ loop through variables
7:     Define currently complete data except  $X_1$ 
      $\dot{X}_{-1}^1 = (\dot{X}_2^0, \dot{X}_3^0, \dot{X}_4^0)$ .
8:     Draw parameters  $\dot{\theta}_1^1 \sim p(\theta_1^1 \mid X_1^{obs}, \dot{X}_{-1}^1, R)$ .
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6:   for  $j = 2$ : ▷ loop through variables
7:     Define currently complete data except  $X_2$ 
      $\dot{X}_{-2}^1 = (\dot{X}_1^1, \dot{X}_3^0, \dot{X}_4^0)$ .
8:     Draw parameters  $\dot{\theta}_2^1 \sim p(\theta_2^1 \mid X_2^{obs}, \dot{X}_{-2}^1, R)$ .
9:     Draw imputations  $\dot{X}_2^1 \sim P(X_2^{mis} \mid \dot{X}_{-2}^1, R, \dot{\theta}_2^1)$ .
10:   end for
11: end for
```

---

## 2. Imputation step

### 2.3. FCS/MICE

---

**Algorithm 1** MICE algorithm [17] for **one** imputed dataset

---

```
1: for  $j$  in  $1, \dots, p$ : ▷ Setup
2:   Specify imputation model for variable  $X_j$ 
    $p(X_j^{mis} \mid X_j^{obs}, X_{-j}, R)$ 
3:   Fill in starting imputations  $\dot{X}_j^0$  by random draws from  $X_j^{obs}$ .
4: end for

5: for  $t = 1$ : ▷ loop through iterations
6:   for  $j = 3$ : ▷ loop through variables
7:     Define currently complete data except  $X_3$ 
      $\dot{X}_{-3}^1 = (\dot{X}_1^1, \dot{X}_2^1, \dot{X}_4^0)$ .
8:     Draw parameters  $\dot{\theta}_3^1 \sim p(\theta_3^1 \mid X_3^{obs}, \dot{X}_{-3}^1, R)$ .
9:     Draw imputations  $\dot{X}_3^1 \sim P(X_3^{mis} \mid \dot{X}_{-3}^1, R, \dot{\theta}_3^1)$ .
10:   end for
11: end for
```

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## 2. Imputation step

### 2.3. FCS/MICE

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**Algorithm 1** MICE algorithm [17] for **one** imputed dataset

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4: end for

5: for  $t = 1$ : ▷ loop through iterations
6:   for  $j = 4$ : ▷ loop through variables
7:     Define currently complete data except  $X_4$ 
      $\dot{X}_{-4}^1 = (\dot{X}_1^1, \dot{X}_2^1, \dot{X}_3^1)$ .
8:     Draw parameters  $\dot{\theta}_4^1 \sim p(\theta_4^1 \mid X_4^{obs}, \dot{X}_{-4}^1, R)$ .
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10:   end for
11: end for
```

---

## 2. Imputation step

### 2.3. FCS/MICE

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**Algorithm 1** MICE algorithm [17] for **one** imputed dataset

---

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1: for  $j$  in  $1, \dots, p$ : ▷ Setup
2:   Specify imputation model for variable  $X_j$ 
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3:   Fill in starting imputations  $\dot{X}_j^0$  by random draws from  $X_j^{obs}$ .
4: end for

5: for  $t = 2$ : ▷ loop through iterations
6:   for  $j = 1$ : ▷ loop through variables
7:     Define currently complete data except  $X_1$ 
      $\dot{X}_{-1}^2 = (\dot{X}_2^1, \dot{X}_3^1, \dot{X}_4^1)$ .
8:     Draw parameters  $\dot{\theta}_1^2 \sim p(\theta_1^2 \mid X_1^{obs}, \dot{X}_{-1}^2, R)$ .
9:     Draw imputations  $\dot{X}_1^2 \sim P(X_1^{mis} \mid \dot{X}_{-1}^2, R, \dot{\theta}_1^2)$ .
10:   end for
11: end for
```

---



## 2. Imputation step

### 2.3. FCS/MICE

---

**Algorithm 1** MICE algorithm [17] for **one** imputed dataset

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```
1: for  $j$  in  $1, \dots, p$ : ▷ Setup
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10:   end for
11: end for
```

---

## 2. Imputation step

### 2.3. FCS/MICE

The imputed values from the **last iteration**,

$$\left(\dot{X}_1^T, \dots, \dot{X}_p^T\right),$$

are then used to replace the missing values in the original data.

One run through the algorithm ➡ one imputed dataset.

## 2. Imputation step

### 2.3. FCS/MICE

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➡ To obtain  $m$  imputed datasets: **repeat  $m$  times**

## 2. Imputation step

### 2.3. FCS/MICE

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are then used to replace the missing values in the original data.

One run through the algorithm ➡ one imputed dataset.

➡ To obtain  $m$  imputed datasets: **repeat  $m$  times**

We refer to the **sequence of imputations** for one missing value, from starting value to final iteration, as a **chain**. Each run through the MICE algorithm produces one chain per missing value.

## 2. Imputation step

### 2.3. FCS/MICE

#### Why iterations?

- Imputed values in one variable depend on the imputed values of the other variables (Gibbs sampling).
- If the starting values (random draws) are far from the actual distribution, imputed values from the first few iterations are not draws from the distribution of interest.

#### How many iterations?

Until **convergence**

= when the sampling distribution does not change any more

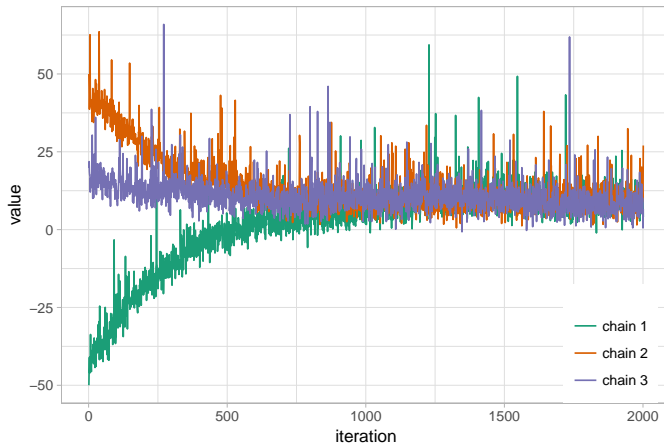
(Note: the imputed value will still vary between iterations.)

#### How to evaluate convergence?

The **traceplot** (x-axis: iteration number, y-axis: imputed value) should show a horizontal band

## 2. Imputation step

### 2.4. Checking convergence



Each chain is the sequence of imputed values (from starting value to final imputed value) for the same missing value.

## 2. Imputation step

### 2.4. Checking convergence

In imputation we have

- several variables with missing values (e.g.,  $p$ )
  - several missing values in each of these variables
  - $m$  chains for each missing value
- ➔ possibly a large number of chains

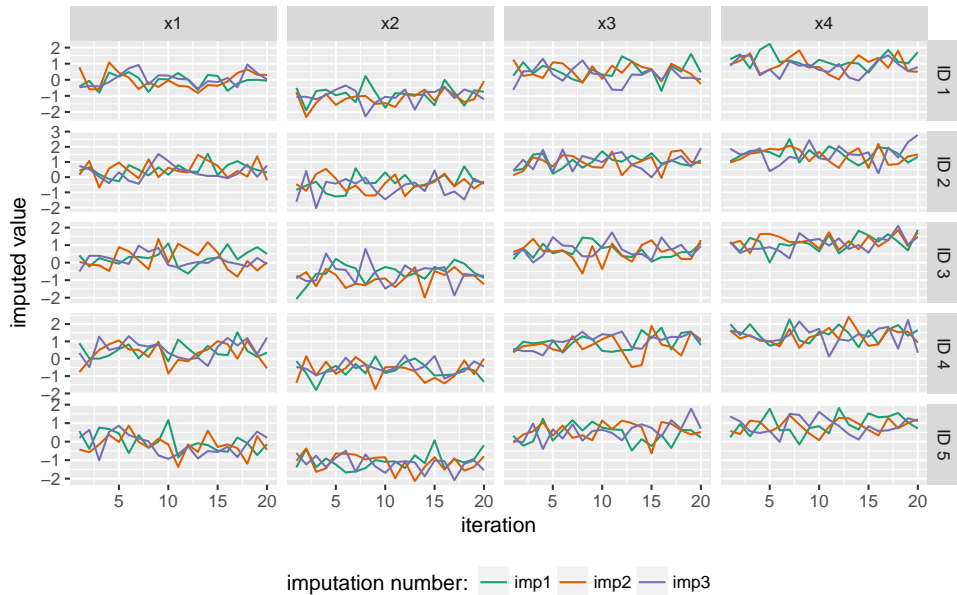
To check all chains separately could be very time consuming in large datasets (and storing all iterations from all imputed values is inefficient).

**Alternative:** Calculate and plot a summary (e.g., the mean) of the imputed values over all subjects, separately per chain and variable

➔ only  $m \times p$  chains to check

## 2. Imputation step

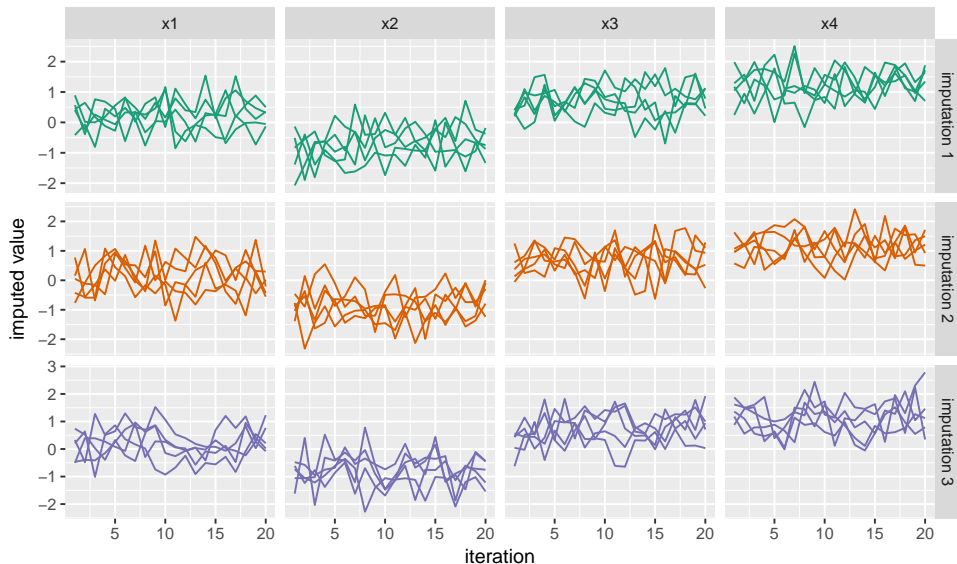
### 2.4. Checking convergence





## 2. Imputation step

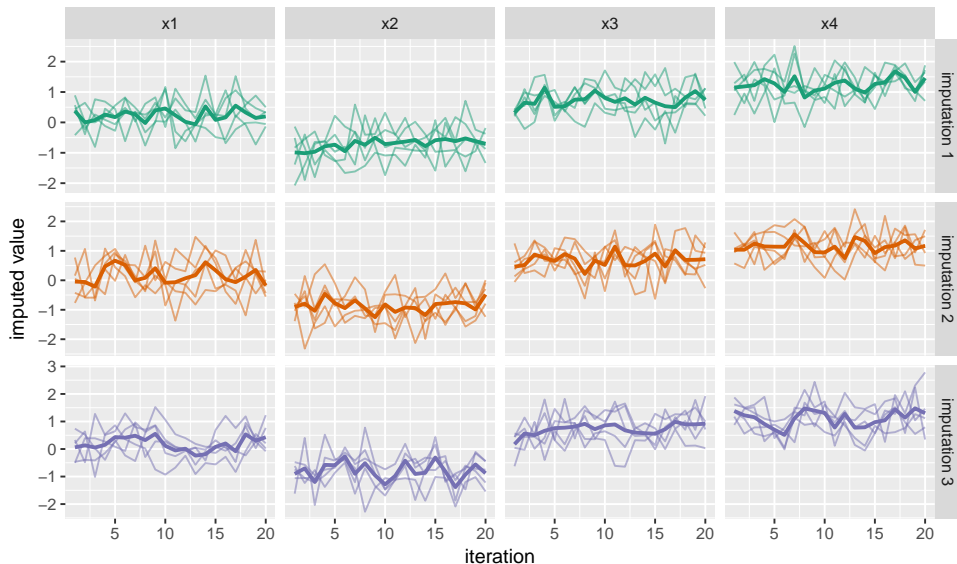
### 2.4. Checking convergence



imputation number: — 1 — 2 — 3

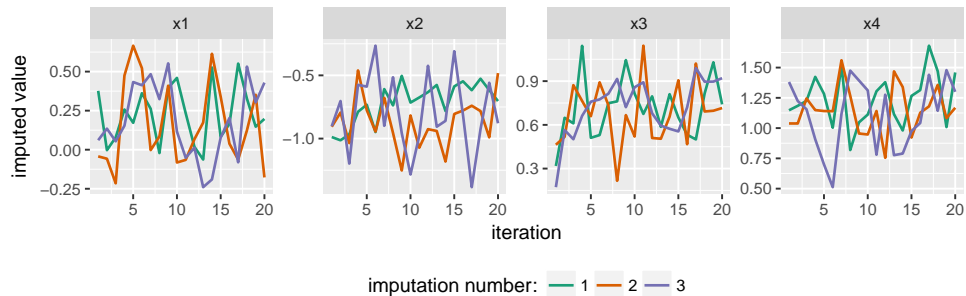
## 2. Imputation step

### 2.4. Checking convergence



## 2. Imputation step

### 2.4. Checking convergence



### 3. Analysis step

Multiple imputed datasets:

$X_1$	$X_2$	$X_3$	$X_4$
1.4	9.2	1.8	2.0
0.5	12.4	2.3	0.1
-0.5	10.7	2.6	-1.6
$\vdots$	$\vdots$	$\vdots$	$\vdots$

$X_1$	$X_2$	$X_3$	$X_4$
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$X_1$	$X_2$	$X_3$	$X_4$
1.4	10.0	1.8	2.0
0.5	12.4	2.2	-1.4
-0.5	8.6	2.6	-1.0
$\vdots$	$\vdots$	$\vdots$	$\vdots$

### 3. Analysis step

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$\vdots$	$\vdots$	$\vdots$	$\vdots$

Analysis model of interest, e.g.,

$$x_1 = \beta_0 + \beta_1 x_2 + \beta_2 x_3 + \beta_3 x_4$$

### 3. Analysis step

Multiple imputed datasets:

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Analysis model of interest, e.g.,

$$x_1 = \beta_0 + \beta_1 x_2 + \beta_2 x_3 + \beta_3 x_4$$

Multiple sets of results:

	est.	se
$\beta_0$	0.35	0.21
$\beta_1$	0.14	0.02
$\beta_2$	-0.64	0.03
$\beta_3$	0.18	0.03

	est.	se
$\beta_0$	0.44	0.23
$\beta_1$	0.12	0.01
$\beta_2$	-0.61	0.03
$\beta_3$	0.22	0.03

	est.	se
$\beta_0$	0.19	0.21
$\beta_1$	0.14	0.01
$\beta_2$	-0.59	0.03
$\beta_3$	0.2	0.03

## 4. Pooling

### 4.1. Why pooling?

Recall from slide 6:

We need to represent missing values by a **number of imputations**.

➡  $m$  imputed datasets

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## 4. Pooling

### 4.1. Why pooling?

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➡  $m$  imputed datasets

From the different imputed datasets we get **different sets of parameter estimates**, each of them with a standard error, representing the uncertainty about the estimate.

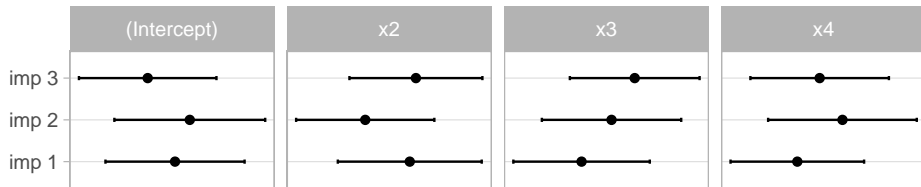
We want to **summarize** the results and describe **how (much) the results vary** between the imputed datasets.

## 4. Pooling

### 4.1. Why pooling?

In the results from multiply imputed data there are **two types of variation/uncertainty**:

- within imputation (represented by the confidence intervals)
- between imputation (horizontal shift between imputations)

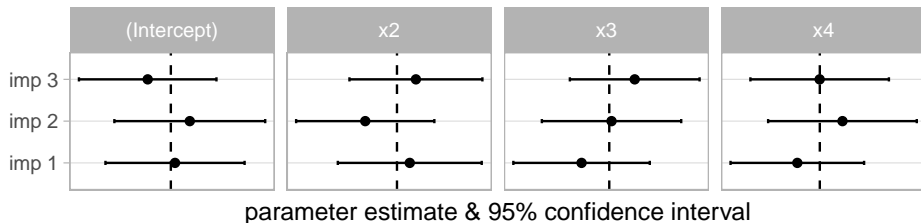


parameter estimate & 95% confidence interval

## 4. Pooling

### 4.1. Why pooling?

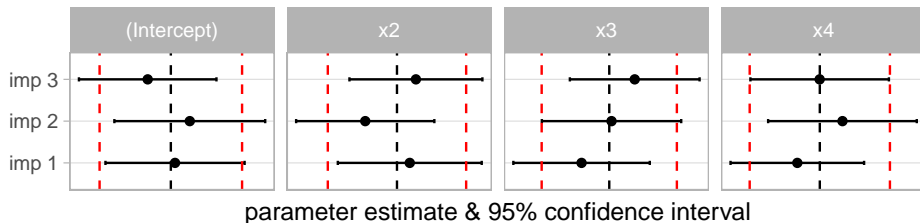
To summarize the results, we can take the mean of the results from the separate analyses. This is the **pooled point estimate**.



## 4. Pooling

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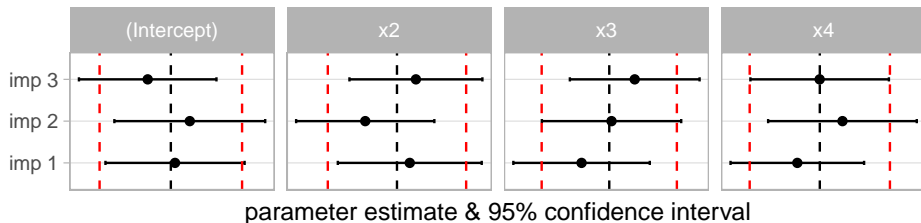


But does the same work for the std. error (or bounds of the CIs)?

## 4. Pooling

### 4.1. Why pooling?

To summarize the results, we can take the mean of the results from the separate analyses. This is the **pooled point estimate**.



But does the same work for the std. error (or bounds of the CIs)?

The averaged CI's (marked in red) seem to underestimate the total variation (within + between).

## 4. Pooling

### 4.2. Rubin's Rules

The most commonly used method to pool results from analyses of multiply imputed data was introduced by Rubin [10], hence **Rubin's Rules**.

#### **Notation:**

$m$ : number of imputed datasets

$Q_\ell$ : quantity of interest (e.g., regr. parameter  $\beta$ ) from  $\ell$ -th imputation

$U_\ell$ : variance of  $Q_\ell$  (e.g.,  $\text{var}(\beta) = \text{se}(\beta)^2$ )

#### **Pooled parameter estimate:**

$$\bar{Q} = \frac{1}{m} \sum_{\ell=1}^m \hat{Q}_\ell$$

## 4. Pooling

### 4.2. Rubin's Rules

The **variance** of the pooled parameter estimate is calculated from the **within and between imputation variance**.

**Average within imputation variance:**

$$\bar{U} = \frac{1}{m} \sum_{\ell=1}^m \bar{U}_{\ell}$$

**Between imputation variance:**

$$B = \frac{1}{m-1} \sum_{\ell=1}^m \left( \hat{Q}_{\ell} - \bar{Q} \right)^T \left( \hat{Q}_{\ell} - \bar{Q} \right)$$

**Total variance:**

$$T = \bar{U} + B + B/m$$

## 4. Pooling

### 4.2. Rubin's Rules

**Confidence intervals** for pooled estimates can be obtained using the **pooled standard error**  $\sqrt{T}$  and a **reference  $t$  distribution** with degrees of freedom

$$\nu = (m - 1) (1 + r_m^{-1})^2,$$

where  $r_m = \frac{(B+B/m)}{\bar{U}}$  is the relative increase in variance that is due to the missing values.

The  $(1 - \alpha)$  **100% confidence interval** is then

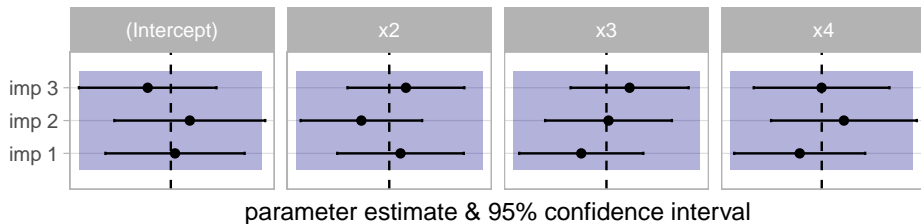
$$\bar{Q} \pm t_{\nu}(\alpha/2)\sqrt{T},$$

where  $t_{\nu}$  is the  $\alpha/2$  quantile of the  $t$  distribution with  $\nu$  degrees of freedom.



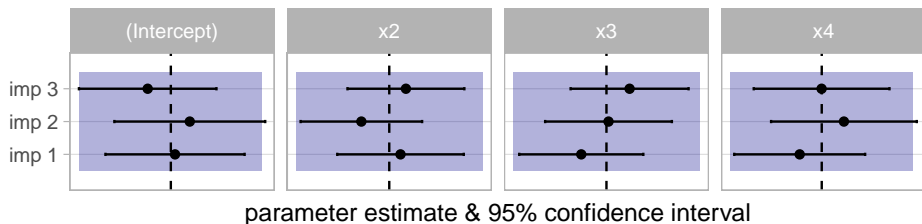
## 4. Pooling

### 4.2. Rubin's Rules



## 4. Pooling

### 4.2. Rubin's Rules



The corresponding **p-value** is the probability

$$Pr \left\{ F_{1,\nu} > (Q_0 - \bar{Q}_m)^2 / T \right\},$$

where  $F_{1,\nu}$  is a random variable that has an F distribution with 1 and  $\nu$  degrees of freedom, and  $Q_0$  is the null hypothesis value (typically zero).

## Quiz

To reiterate the content of the above sections, you can take the corresponding quiz. An interactive version can be found at

[https://emcbiostatistics.shinyapps.io/MICourse\\_Quiz\\_PartI](https://emcbiostatistics.shinyapps.io/MICourse_Quiz_PartI)

or you can download an html version from Canvas (Practicals/Quiz\_PartI\_static.html). [\[update links\]](#).

## 5. A closer look at the imputation step

### 5.1. Bayesian multiple imputation

The imputation step consists itself of two (or three) steps:

0. Specification of the imputation model,
1. **estimation** or sampling **of the parameters**, and
2. **drawing imputed values** from the predictive distribution.

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0. Specification of the imputation model,
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2. **drawing imputed values** from the predictive distribution.

#### Notation:

Let  $\mathbf{y}$  be the incomplete covariate to be imputed, and  $\mathbf{X}$  the design matrix of other (complete or imputed) variables.

$$\mathbf{y} = \left\{ \begin{array}{l} \mathbf{y}_{obs} \\ \mathbf{y}_{mis} \end{array} \right\} \left[ \begin{array}{c} y_1 \\ \vdots \\ y_q \\ NA \\ \vdots \\ NA \end{array} \right] \quad \mathbf{X} = \left\{ \begin{array}{l} \mathbf{X}_{obs} \\ \mathbf{X}_{mis} \end{array} \right\} \left[ \begin{array}{cccc} 1 & x_{11} & \dots & x_{1p} \\ \vdots & \vdots & \dots & \vdots \\ 1 & x_{q1} & \dots & x_{qp} \\ 1 & x_{q+1,1} & \dots & x_{q+1,p} \\ \vdots & \vdots & \dots & \vdots \\ 1 & x_{n1} & \dots & x_{np} \end{array} \right]$$

## 5. A closer look at the imputation step

### 5.1. Bayesian multiple imputation

In the **Bayesian framework**, **everything unknown** or unobserved is considered as a **random variable**. Here, this includes for example regression coefficients  $\beta$ , residual variance  $\sigma^2$  and missing values  $\mathbf{y}_{mis}$  and  $\mathbf{X}_{mis}$ .

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Random variables have a **probability distribution**. The **expectation** of that distribution quantifies where which **values** of the random variable are **most likely**, the **variance** is a measure of the **uncertainty** about the values.

## 5. A closer look at the imputation step

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Random variables have a **probability distribution**. The **expectation** of that distribution quantifies where which **values** of the random variable are **most likely**, the **variance** is a measure of the **uncertainty** about the values.

In **Bayesian imputation**, the **information obtained from the observed data** is used to **estimate the probability distributions** for the missing values and unknown parameters, and values are **imputed by draws** from that posterior (= after having seen the data) distribution.



## 5. A closer look at the imputation step

### 5.1. Bayesian multiple imputation

To determine the **expectation** of the posterior distribution of the missing values, usually a **regression model** is used, that depends on the unknown coefficients  $\beta$ .

$$\mathbb{E}(\mathbf{y}_{mis} \mid \mathbf{X}, \beta) = f(\mathbf{X}_{mis}\beta)$$

The posterior distribution of  $\beta$  and  $\sigma$ ,  $p(\beta, \sigma \mid \mathbf{y}_{obs} \mid \mathbf{y}_{obs}, \mathbf{X}_{obs})$ , is estimated from the corresponding regression model on the observed data.

## 5. A closer look at the imputation step

### 5.1. Bayesian multiple imputation

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To **impute** missing values, while **taking into account the uncertainty about  $\beta$  and  $\sigma$** , the estimated posterior distributions of the missing values and parameters are multiplied

$$p(\mathbf{y}_{mis} \mid \mathbf{X}_{mis}, \beta, \sigma) p(\beta, \sigma \mid \mathbf{y}_{obs}, \mathbf{X}_{obs})$$

In practice, this can be implemented by first making a draw from the posterior distributions of  $\beta$  and  $\sigma$ , and plugging the values into the distribution of  $\mathbf{y}_{obs}$ .

## 5. A closer look at the imputation step

### 5.1. Bayesian multiple imputation

**Example:** We assume that  $\mathbf{y}$  given  $\mathbf{X}$  is approximately normal.

Then  $p(\mathbf{y}_{mis} \mid \mathbf{X}_{mis}, \boldsymbol{\beta}, \sigma)$  is a normal distribution and we can

- draw  $\tilde{\boldsymbol{\beta}}$  from  $p(\boldsymbol{\beta} \mid \mathbf{y}_{obs}, \mathbf{X}_{obs})$ ,
- draw  $\tilde{\sigma}$  from  $p(\sigma \mid \mathbf{y}_{obs}, \mathbf{X}_{obs})$ ,
- draw  $\tilde{\mathbf{y}}_{mis}$  from a normal distribution with mean (= expectation)  $\mathbf{X}\tilde{\boldsymbol{\beta}}$  and variance  $\tilde{\sigma}^2$ .

This is actually the approach we have seen previously on Slides 12/13 and 19.

## 5. A closer look at the imputation step

### 5.2. Bootstrap multiple imputation

An alternative approach is to capture the uncertainty with **bootstrap** sampling.

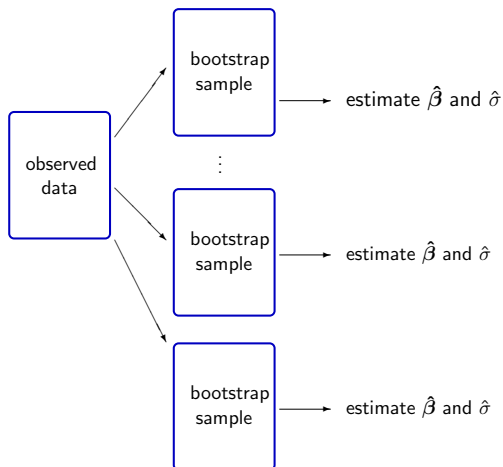
In empirical **Bootstrap**, (many) replications of the data are created by repeatedly drawing values from the original data.

## 5. A closer look at the imputation step

### 5.2. Bootstrap multiple imputation

An alternative approach is to capture the uncertainty with **bootstrap** sampling.

In empirical **Bootstrap**, (many) replications of the data are created by repeatedly drawing values from the original data.



Bootstrap samples can contain some **observations multiple times** and some **observations not at all**.

The statistic of interest is then calculated on each of the bootstrap samples.

## 5. A closer look at the imputation step

### 5.2. Bootstrap multiple imputation

In **bootstrap multiple imputation**,

- **one bootstrap sample** of the **observed data** is created per imputation,
- the **least-squares estimates** of the parameters are calculated from

$$\mathbf{y}_{obs} = \mathbf{X}_{obs} \underset{\downarrow \hat{\beta}}{\beta} + \underset{\downarrow \hat{\sigma}}{\varepsilon_{obs}} \quad (\text{step 1}).$$

- Imputed values are sampled from  $p(\mathbf{y}_{mis} \mid \mathbf{X}_{mis}, \hat{\beta}, \hat{\sigma})$  (step 2).

## 5. A closer look at the imputation step

### 5.2. Bootstrap multiple imputation

In **bootstrap multiple imputation**,

- **one bootstrap sample** of the **observed data** is created per imputation,
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$$\mathbf{y}_{obs} = \underset{\downarrow \hat{\boldsymbol{\beta}}}{\mathbf{X}_{obs}} \underset{\downarrow \hat{\sigma}}{\boldsymbol{\beta}} + \varepsilon_{obs} \quad (\text{step 1}).$$

- Imputed values are sampled from  $p(\mathbf{y}_{mis} \mid \mathbf{X}_{mis}, \hat{\boldsymbol{\beta}}, \hat{\sigma})$  (step 2).

Analogous to Bayesian multiple imputation, for a normal imputation model,  $p()$  is the normal distribution and

$$\tilde{\mathbf{y}}_{mis} = \mathbf{X}_{mis} \hat{\boldsymbol{\beta}} + \tilde{\boldsymbol{\varepsilon}}$$

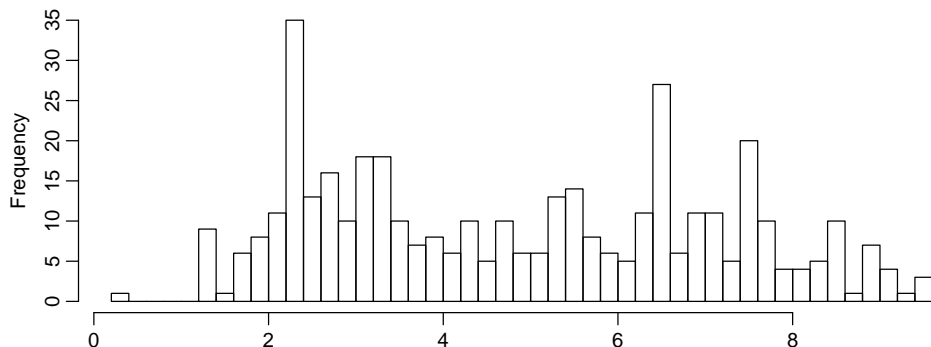
where  $\tilde{\boldsymbol{\varepsilon}}$  is drawn independently from  $N(0, \hat{\sigma}^2)$ .

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

Both Bayesian and bootstrap multiple imputation sample imputed values from a distribution  $p()$  in step 2.

Sometimes, the empirical distribution can not be adequately approximated by a known probability distribution.





## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

**Predictive Mean Matching (PMM)** was developed to provide a semi-parametric approach to imputation for settings where the normal distribution is not a good choice for the predictive distribution.[8, 9]

The idea is to **find cases in the observed data that are similar to the cases with missing values** and to fill in the missing value with the observed value from one of those cases.

To find similar cases, the predicted values of complete and incomplete cases are compared.

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

#### The steps in PMM:

1. Obtain parameter estimates for  $\hat{\beta}$  and  $\hat{\sigma}$  (see later)
2. Calculate the predicted values for the observed data

$$\hat{y}_{obs} = \mathbf{X}_{obs}\hat{\beta}$$

3. Calculate the predicted value for the incomplete data

$$\hat{y}_{mis} = \mathbf{X}_{mis}\hat{\beta}$$

4. For each missing value, find  $d$  donor candidates that fulfill a given criterium (details on the next slide).
5. Randomly select one of the donors.

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

Several **criteria to select donors** have been proposed:

1. The donor is the **(one) case with the smallest absolute difference**  
 $|\hat{y}_{mis,i} - \hat{y}_{obs,j}|, j = 1, \dots, q.$
2. Donor candidates are the  **$d$  cases with the smallest absolute difference**  
 $|\hat{y}_{mis,i} - \hat{y}_{obs,j}|, j = 1, \dots, q.$  The donor is selected randomly from the candidates.
3. Donor candidates are those cases for which the **absolute difference is smaller than some limit  $\eta$** :  $|\hat{y}_{mis,i} - \hat{y}_{obs,j}| < \eta, j = 1, \dots, q.$  The donor is selected randomly from the candidates.
4. Select candidates like in 2. or 3., but select the donor from the candidates with probability that depends on  $|\hat{y}_{mis,i} - \hat{y}_{obs,j}|$ . [16]

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

#### Potential issues with donor selection

- Selection criteria 2. - 4., **require the number of candidates  $d$**  (or maximal difference  $\eta$ ) to be specified. Common choices for  $d$  are 3, 5 or 10.

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

#### Potential issues with donor selection

- Selection criteria 2. - 4., **require the number of candidates  $d$**  (or maximal difference  $\eta$ ) to be specified. Common choices for  $d$  are 3, 5 or 10.
- If the same donor is chosen in many/all imputations (e.g., because only a few similar observed cases are available), the **uncertainty about the missing values will be underestimated**.

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

#### Potential issues with donor selection

- Selection criteria 2. - 4., **require the number of candidates  $d$**  (or maximal difference  $\eta$ ) to be specified. Common choices for  $d$  are 3, 5 or 10.
  - If the same donor is chosen in many/all imputations (e.g., because only a few similar observed cases are available), the **uncertainty about the missing values will be underestimated**.
- ➔ PMM may be **problematic** when
- the **dataset is very small**,
  - the **proportion of missing values is large**, or
  - one/some **predictor variable(s) are strongly related to the missingness**.

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

#### Potential issues with donor selection

- Selection criteria 2. - 4., **require the number of candidates  $d$**  (or maximal difference  $\eta$ ) to be specified. Common choices for  $d$  are 3, 5 or 10.
- If the same donor is chosen in many/all imputations (e.g., because only a few similar observed cases are available), the **uncertainty about the missing values will be underestimated**.
  - ➔ PMM may be **problematic** when
    - the **dataset is very small**,
    - the **proportion of missing values is large**, or
    - one/some **predictor variable(s) are strongly related to the missingness**.
- Therefore, using  $d = 1$  (selection criterion 1.) is not a good idea. On the other hand, using too many candidates can lead to bad matches.

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

#### Potential issues with donor selection

- Selection criteria 2. - 4., **require the number of candidates  $d$**  (or maximal difference  $\eta$ ) to be specified. Common choices for  $d$  are 3, 5 or 10.
- If the same donor is chosen in many/all imputations (e.g., because only a few similar observed cases are available), the **uncertainty about the missing values will be underestimated**.
  - ➔ PMM may be **problematic** when
    - the **dataset is very small**,
    - the **proportion of missing values is large**, or
    - one/some **predictor variable(s) are strongly related to the missingness**.
- Therefore, using  $d = 1$  (selection criterion 1.) is not a good idea. On the other hand, using too many candidates can lead to bad matches.
- Schenker and Taylor [15] proposed an adaptive procedure to select  $d$ , but it is not used much in practice.



## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

For the **sampling of the parameters** (step 1 on slide 43), different approaches have been introduced in the literature:

- Type-0 point estimates  $\hat{\beta}$  are used in both prediction models (least squares or maximum likelihood)
- Type-I  $\hat{\beta}$  to predict  $\hat{y}_{obs}$ ;  $\tilde{\beta}$  to predict  $\hat{y}_{mis}$  is sampled from the posterior distribution of  $\beta$  (Bayesian) or bootstrapped
- Type-II  $\tilde{\beta}$  to predict  $\hat{y}_{obs}$  as well as  $\hat{y}_{mis}$
- Type-III different draws  $\tilde{\beta}^{(1)}$  and  $\tilde{\beta}^{(2)}$  to predict  $\hat{y}_{obs}$  and  $\hat{y}_{mis}$ , respectively

The use of point estimates (Type-0 and Type-I matching) **underestimates the uncertainty** about the regression parameters.

## 5. A closer look at the imputation step

### 5.3. Semi-parametric imputation

Another point of consideration is the **choice of the set of data used to train the prediction models**.

In the version presented on slide 43, the same set of data (all cases with observed  $y$ ) is used to train the model and to produce predicted values of  $y_{obs}$ .

The predictive model will likely fit the observed cases better than the missing cases, and, hence, **variation will be underestimated**.

As an alternative, the **model could be trained on the whole data** (using previously imputed values) or to use a **leave-one-out approach** on the observed data.

## 5. A closer look at the imputation step

### 5.4. What is implemented in software?

#### **mice** (in R):

- **PMM** via `mice.impute.pmm()`
  - specification of number of donors  $d$  (same for all variables)
  - Type-0, Type-I, Type-II matching
- **PMM** via `mice.impute.midastouch()`
  - allows leave-one-out estimation of the parameters
  - distance based donor selection
  - Type-0, Type-I, Type-II matching
- **bootstrap** linear regression via `mice.impute.norm.boot()`
- **bootstrap** logistic regression via `mice.impute.logreg.boot()`
- **Bayesian** linear regression via `mice.impute.norm()`
- ...

## 1. What is Multiple Imputation?

- Rubin's two ideas:
  - Missing values need to be represented by multiple imputed values.
  - A model is necessary to obtain good imputations.
- Imputed values are obtained from the predictive distribution of the missing data, given the observed data.
- Multiple completed datasets are created from the multiple imputed values.
- Multiple imputation has three steps: Imputation, analysis, pooling

# Summary of Part I (cont.)

## 2. Imputation step

- Two sources of variation need to be taken into account
  - parameter uncertainty
  - random variation
- For two main approaches to MI for imputation of non-monotone multivariate missing data
  - MICE/FCS
  - Joint model imputation
- The MICE algorithm re-uses univariate imputation models by iterating through all incomplete variables, multiple times (iterations)
- Multiple runs through the algorithm are necessary to create multiple imputed dataset
- The convergence of the chains needs to be checked.

# Summary of Part I (cont.)

## 3. Analysis step

- Analyse each imputed dataset the way you would analyse a complete dataset

## 4. Pooling

- Results from analyses of multiple imputed datasets can be summarized by taking the average of the regression coefficients
- For the total variance, two sources of variation need to be considered:
  - within imputation variance
  - between imputation variance

# Summary of Part I (cont.)

## 5. A closer look at the imputation step

- Two parametric approaches for imputation:
  - Bayesian (sample from posterior distribution of parameters)
  - Bootstrap (uses bootstrap samples of the data to estimate parameters)
- Predictive mean matching is a semi-parametric alternative (it matches observed and missing cases based on their predicted values).
- In PMM we need to consider
  - donor selection
  - matching type (how parameters are sampled/estimated),
  - the set of data used to calculate/estimate the parameters.
- Bayesian and bootstrap imputation take into account the variation, while many choices in PMM lead to underestimation of the variation.

## Part II

# Multiple Imputation Workflow



# Outline of Part II

## 6. Know your data

- 6.1 Missing data patterns
- 6.2 Data distributions
- 6.3 Correlations & patterns
- 6.4 Why are values missing?
- 6.5 Auxiliary variables

## 7. Imputation with `mice()`

- 7.1 Main function arguments
- 7.2 Imputation methods
- 7.3 Predictor matrix
- 7.4 Passive imputation
- 7.5 Post processing
- 7.6 Visit sequence
- 7.7 Good to know

## 8. Convergence & Diagnostics

- 8.1 Logged events
- 8.2 Convergence
- 8.3 Diagnostics

## 9. Analyse & pool the imputed data

- 9.1 Analyzing imputed data
- 9.2 Pooling results
- 9.3 Functions for pooled results

## 10. Additional functions in `mice()`

- 10.1 Extracting and exporting imputed data
- 10.2 Combining `mids` objects
- 10.3 Adding variables to `mids` objects

## 11. Multiple Imputation in SPSS

- 11.1 Where to get help
- 11.2 Multiple Imputation Features

## 6. Know your data

### 6.1. Missing data patterns

There are several packages in R that provide functions to create and **plot the missing data pattern**.

Examples are:

**mice**, **VIM**, **Amelia**, **visdat**, **nanian**, ...

To create the plots on the following slides, we need to load the example data

```
datadir <-      # fill in path to folder containing the data  
load(file.path(datadir, "NHANES.RData"))
```

## 6. Know your data

### 6.1. Missing data patterns

```
mdp <- mice::md.pattern(NHANES)
head(mdp[, -c(7:14)]) # omit some columns to fit it on the slide
```

```
##      age gender race DM educ smoke hypchol creat albu uricacid bili alc HyperMed
## 572    1      1    1  1    1    1      1    1    1      1    1    1      1  0
## 1      1      1    1  1  1    1    0      1    1    1      1    1    1      1  1
## 141    1      1    1  1  1    1    1      1    1    1      1    1    0      1  1
## 17     1      1    1  1  1    1    1      1    1    1      1    1    1      1  1
## 1063   1      1    1  1  1    1    1      1    1    1      1    1    1      0  1
## 18     1      1    1  1  1    1    1      1    1    1      1    1    1      1  1
```

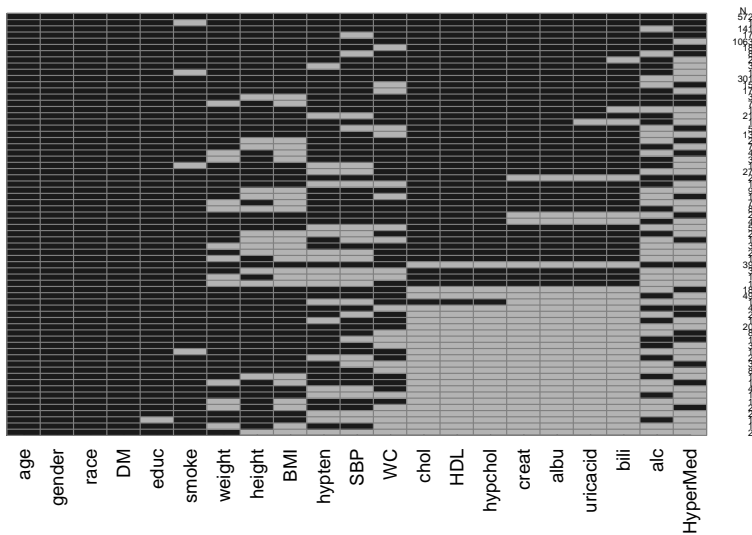
```
tail(mdp[, -c(7:14)])
```

```
##      age gender race DM educ smoke hypchol creat albu uricacid bili alc HyperMed
## 2      1      1    1  1  1    1    0      0    0      0    0    0      1  11
## 2      1      1    1  1  1    1    0      0    0      0    0    0      0  12
## 1      1      1    1  1  1    0    1      0    0    0      0    0    1      0  12
## 1      1      1    1  1  1    1    1      0    0    0      0    0    0      0  12
## 2      1      1    1  1  1    1    1      0    0    0      0    0    0      0  14
##      0      0    0  0  1    4    175    184    184      185    188 627      1606 3975
```

## 6. Know your data

### 6.1. Missing data patterns

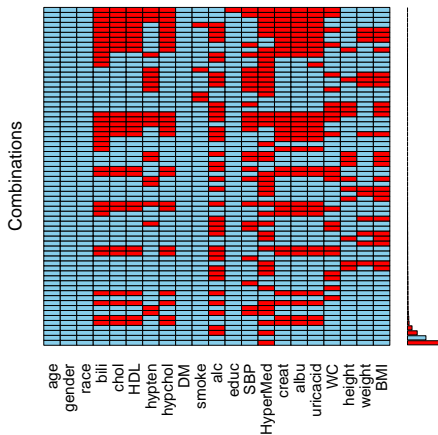
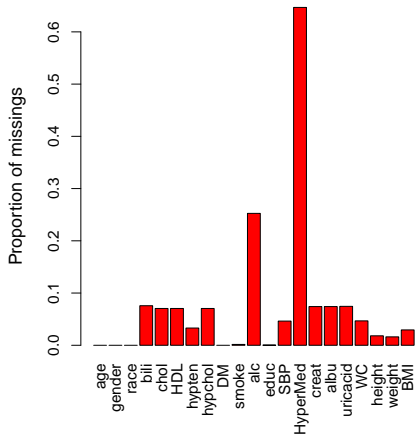
```
par(mar = c(5, 0.5, 1, 3), mgp = c(2, 0.6, 0))  
JointAI::md_pattern(NHANES, print = FALSE, yaxis_pars = list(cex.axis = 0.5))
```



## 6. Know your data

### 6.1. Missing data patterns

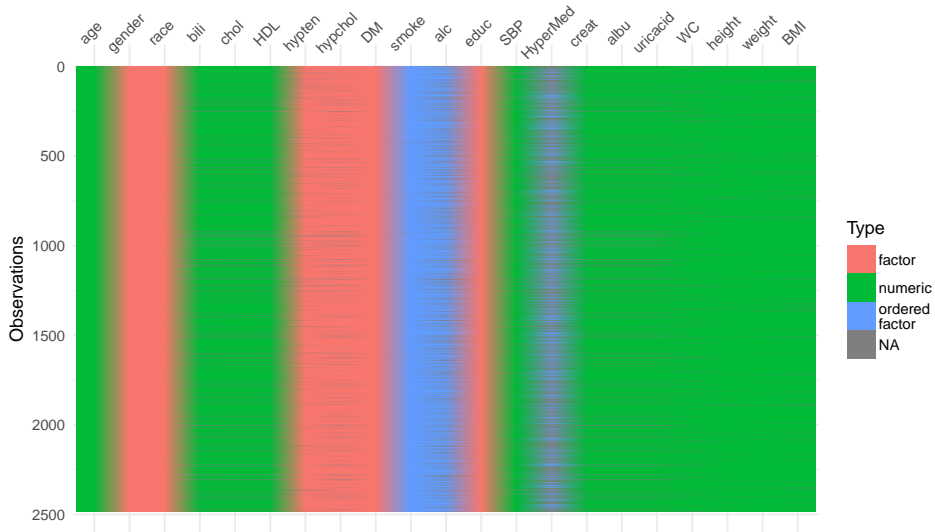
```
par(mar = c(6, 3, 2, 1))  
VIM::aggr(NHANES, prop = T, numbers = FALSE)
```



## 6. Know your data

### 6.1. Missing data patterns

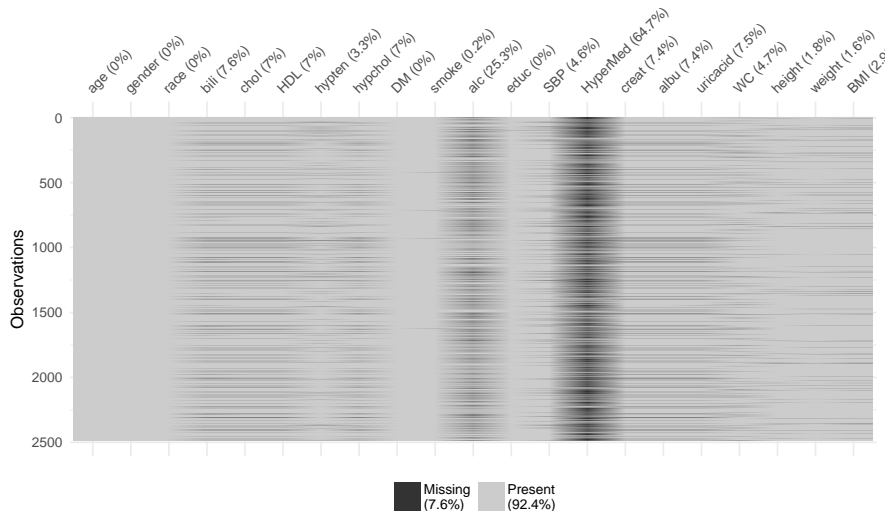
```
visdat::vis_dat(NHANES, sort_type = FALSE)
```



## 6. Know your data

### 6.1. Missing data patterns

```
visdat::vis_miss(NHANES)
```



## 6. Know your data

### 6.1. Missing data patterns

```
# number and proportion of complete cases
Ncc <- cbind(
  "#" = table(complete.cases(NHANES)),
  "%" = round(100 * table(complete.cases(NHANES))/nrow(NHANES), 2)
)
rownames(Ncc) <- c("incompl.", "complete")
```

```
# number and proportion of missing values per variable
nmis <- cbind("# NA" = sort(colSums(is.na(NHANES))),
              "% NA" = round(sort(colMeans(is.na(NHANES))) * 100, 2))
```



## 6. Know your data

### 6.1. Missing data patterns

#### Number and proportion of (in)complete cases

```
##          #      %  
## incompl. 1911 76.96  
## complete  572 23.04
```

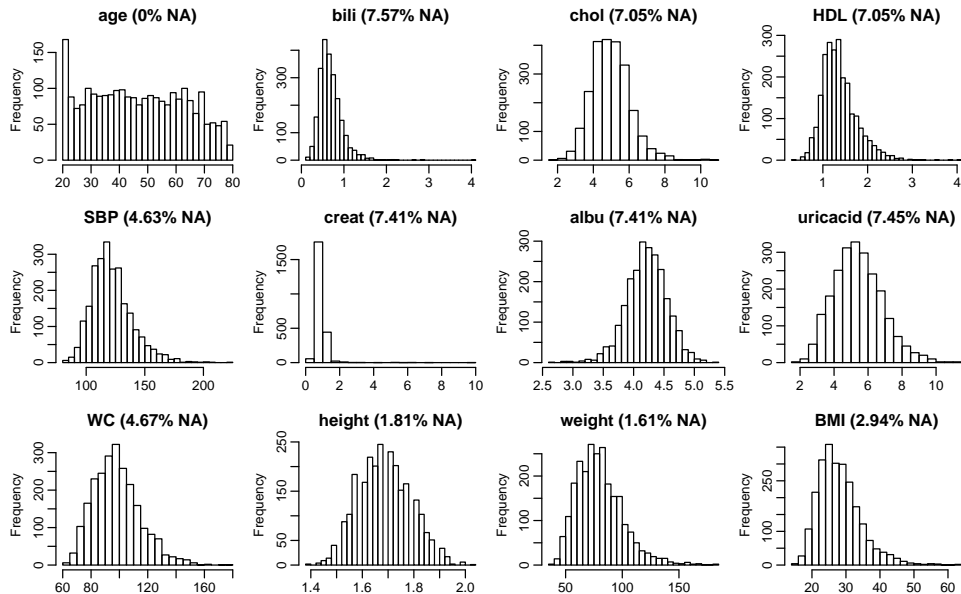
#### Number and proportion of missing values per variable

```
##      # NA % NA  
## age      0 0.00  
## gender    0 0.00  
## race      0 0.00  
## DM        0 0.00  
## educ      1 0.04  
## smoke     4 0.16  
## weight    40 1.61  
## height    45 1.81  
## BMI       73 2.94  
## hypten    82 3.30  
## SBP      115 4.63
```

```
##      # NA % NA  
## WC      116 4.67  
## chol     175 7.05  
## HDL      175 7.05  
## hypchol  175 7.05  
## creat    184 7.41  
## albu     184 7.41  
## uricacid 185 7.45  
## bili     188 7.57  
## alc      627 25.25  
## HyperMed 1606 64.68
```

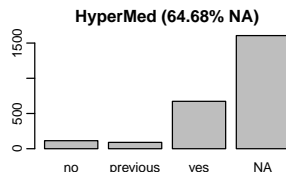
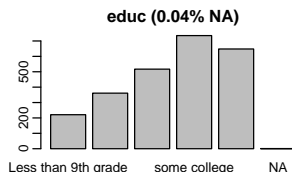
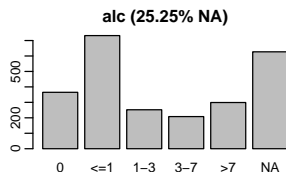
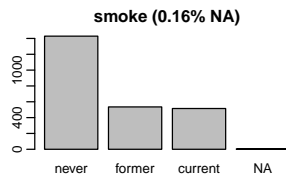
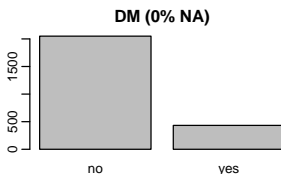
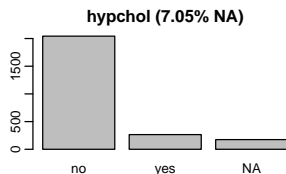
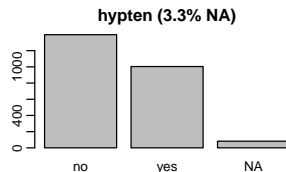
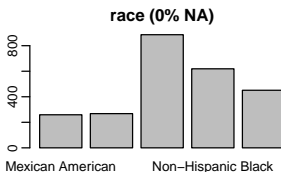
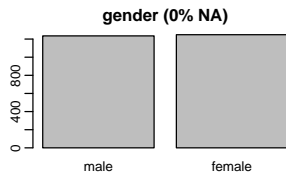
# 6. Know your data

## 6.2. Data distributions



# 6. Know your data

## 6.2. Data distributions



## 6. Know your data

### 6.2. Data distributions

```
# syntax for continuous variables
NHANESnum <- NHANES[, sapply(NHANES, is.numeric)]
par(mfrow = c(3, 4), mar = c(3, 3.2, 0.5, 0.5), mgp = c(2, 0.6, 0))
for (i in 1:ncol(NHANESnum)) {
  hist(NHANESnum[, i], nclass = 30, xlab = names(NHANESnum)[i], main = "")
  legend("topright", bty = "n",
        legend = paste0(round(mean(is.na(NHANESnum[, i]))*100, 2), "% NA"))
}

# syntax for factors
NHANESfac <- NHANES[, sapply(NHANES, is.factor)]
par(mfrow = c(3, 5), mar = c(3, 3.2, 2.5, 0.5), mgp = c(2, 0.6, 0))
for (i in 1:ncol(NHANESfac)) {
  tab <- table(NHANESfac[, i], exclude = NULL)
  names(tab)[is.na(names(tab))] <- "NA"
  barplot(tab, main = paste0(names(NHANESfac)[i], " (",
                             round(mean(is.na(NHANESfac[, i]))*100,
                                     2), "% NA)"))
}
```

## 6. Know your data

### 6.3. Correlations & patterns

A quick (and dirty) way to check for strong correlations between variables is:

```
# re-code all variables as numeric and calculate spearman correlation
Corr <- cor(sapply(NHANES, as.numeric),
            use = "pairwise.complete.obs", method = "spearman")

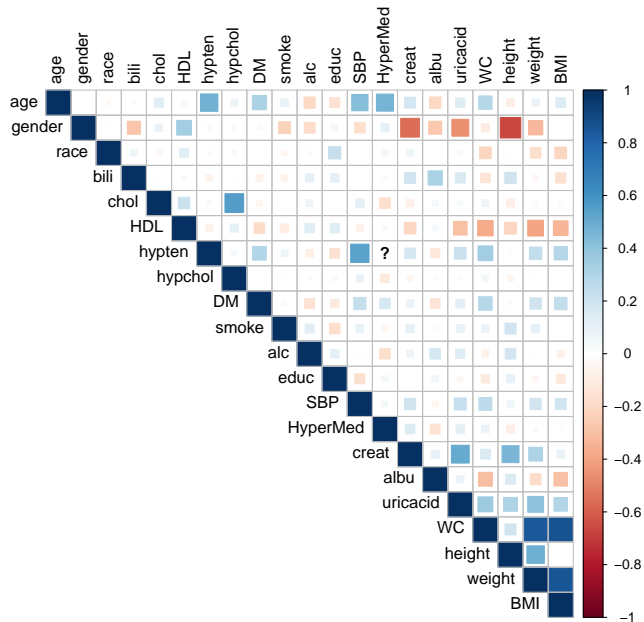
## Warning in cor(sapply(NHANES, as.numeric), use = "pairwise.complete.obs",
: the standard deviation is zero
```

```
corrplot::corrplot(Corr, method = "square", type = "upper", tl.col = "black")
```

**Note:** We only use the correlation coefficient for binary variables in this visualization, not as a statistical result!

## 6. Know your data

### 6.3. Correlations & patterns



## 6. Know your data

### 6.3. Correlations & patterns

Check out what the problem is with `hypertension` and `HyperMed`:

```
table(hypertension = NHANES$hypten,  
      HyperMed = NHANES$HyperMed, exclude = NULL)
```

```
##           HyperMed  
## hypertension  no previous  yes <NA>  
##           no      0         0    0 1397  
##           yes   114        90  673  127  
##           <NA>    0         0    0   82
```

## 6. Know your data

### 6.4. Why are values missing?

Knowing your data also means to be able to answer these questions:

- Do missing values in multiple variables always **occur together**?  
(e.g. blood measurements)
- Are there **structural missing values**? (e.g. pregnancy status in men)
- Are there **patterns** in the missing values?  
(e.g. only patients with hypertension have observations of [HyperMed](#))
- Are values **missing by design**?
- Is the **assumption of ignorable missingness** (MAR or MCAR) justifiable?



## 6. Know your data

### 6.5. Auxiliary variables

**Auxiliary variables** are variables that are not part of the analysis but **can help during imputation**.

Good auxiliary variables

- are **related to the probability of missingness** in a variable, or
- are **related to the incomplete variable** itself,
- do **not have many missing values** themselves and
- are (mostly) **observed** when the incomplete variable of interest is missing.

## 7. Imputation with `mice()`

### 7.1. Main function arguments

The main arguments needed to impute data with `mice()` are:

- `data`: the dataset
- `m`: number of imputed datasets (default is 5)
- `maxit`: number of iterations (default is 5)
- `method`: vector of imputation methods
- `defaultMethod`: vector of default imputation methods for numerical, binary, unordered and ordered factors with  $> 2$  levels (default is `c("pmm", "logreg", "polyreg", "polr")`)
- `predictorMatrix`: matrix specifying roles of variables

# 7. Imputation with mice()

## 7.2. Imputation methods

**mice** has implemented many **imputation methods**, the most commonly used ones are:

- **pmm**: predictive mean matching (any)
- **norm**: Bayesian linear regression (numeric)
- **logreg**: binary logistic regression (binary)
- **polr**: proportional odds model (ordered factors)
- **polyreg**: polytomous logistic regression (unordered factors)

## 7. Imputation with mice()

### 7.2. Imputation methods

#### Change the default imputation method:

E.g.: to use `norm` instead of `pmm` for all continuous incomplete variables, use:

```
mice(NHANES, defaultMethod = c("norm", "logreg", "polyreg", "polr"))
```

## 7. Imputation with mice()

### 7.2. Imputation methods

#### Change the default imputation method:

E.g.: to use `norm` instead of `pmm` for all continuous incomplete variables, use:

```
mice(NHANES, defaultMethod = c("norm", "logreg", "polyreg", "polr"))
```

#### Change imputation method for a single variable:

To change the imputation method for single variables (but also for changes in other arguments) it is convenient to **do a setup run** of `mice()` without iterations (`maxit = 0`) and to extract and modify the parameters from there.

## 7. Imputation with mice()

### 7.2. Imputation methods

#### Change the default imputation method:

E.g.: to use `norm` instead of `pmm` for all continuous incomplete variables, use:

```
mice(NHANES, defaultMethod = c("norm", "logreg", "polyreg", "polr"))
```

#### Change imputation method for a single variable:

To change the imputation method for single variables (but also for changes in other arguments) it is convenient to **do a setup run** of `mice()` without iterations (`maxit = 0`) and to extract and modify the parameters from there.

#### Exclude variable from imputation:

When a variable that has missing values should not be imputed, the method needs to be set to `"`.

## 7. Imputation with mice()

### 7.2. Imputation methods

```
library(mice)
imp0 <- mice(NHANES, maxit = 0)
meth <- imp0$method
meth

##      age      gender      race      bili      chol      HDL      hypten
##      ""         ""         ""      "pmm"      "pmm"      "pmm"      "logreg"
##  hypchol      DM      smoke      alc      educ      SBP      HyperMed
## "logreg"         ""      "polr"      "polr" "polyreg"      "pmm"      "polr"
##      creat      albu  uricacid      WC      height      weight      BMI
##      "pmm"      "pmm"      "pmm"      "pmm"      "pmm"      "pmm"      "pmm"

meth["albu"] <- "norm"
meth["HyperMed"] <- ""
# imp <- mice(NHANES, method = meth)
```

## 7. Imputation with mice()

### 7.3. Predictor matrix

The `predictorMatrix` is a matrix that specifies **which variables are used as predictors** in which imputation model.

Each row represents the model for the variable given in the rowname.

```
head(imp0$predictorMatrix)[, 1:11]
```

##	age	gender	race	bili	chol	HDL	hypten	hypchol	DM	smoke	alc
## age	0	0	0	0	0	0	0	0	0	0	0
## gender	0	0	0	0	0	0	0	0	0	0	0
## race	0	0	0	0	0	0	0	0	0	0	0
## bili	1	1	1	0	1	1	1	1	1	1	1
## chol	1	1	1	1	0	1	1	1	1	1	1
## HDL	1	1	1	1	1	0	1	1	1	1	1

Variables **not used as predictor** are (or have to be set to) **zero**.

By **default, all variables** (except the variable itself) **are used** as predictor.  
For complete variables all entries are 0.



## 7. Imputation with mice()

### 7.3. Predictor matrix

#### Important:

A variable that has **missing values needs to be imputed** in order to be used as predictor for other imputation models!!!

#### Note:

By default, **ALL** variables with missing values are imputed and **ALL** variables are used as predictor variables.

- ➡ Make sure to adjust the `predictorMatrix` and `method` to avoid using ID variables or other columns of the data that should not be part of the imputation.
- ➡ Make sure all **variables are coded correctly**, so that the chosen imputation models are appropriate (e.g., ordered factors).

## 7. Imputation with mice()

### 7.3. Predictor matrix

```
library(mice)
imp0 <- mice(NHANES, maxit = 0,
             defaultMethod = c("norm", "logreg", "polyreg", "polr"))
meth <- imp0$method
meth["educ"] <- "polr"
meth["HyperMed"] <- ""

pred <- imp0$predictorMatrix
pred[, "HyperMed"] <- 0
imp <- mice(NHANES, method = meth, predictorMatrix = pred, printFlag = F)
```

## 7. Imputation with mice()

### 7.4. Passive imputation

In some cases, variables are **functions of other variables**, e.g.,  $BMI = \frac{weight}{height^2}$ .

If we impute **BMI** directly, its values may be **inconsistent** with the (imputed) values of **height** and **weight**.

```
DF1 <- complete(imp, 1) # select the first imputed dataset
round(cbind("wgt/hgt^2" = DF1$weight/DF1$height^2,
           BMI = DF1$BMI)[is.na(NHANES$BMI), ], 2)[1:5, ]

##      wgt/hgt^2   BMI
## [1,]    27.25 28.77
## [2,]    23.80 22.94
## [3,]    25.77 24.06
## [4,]    27.56 27.50
## [5,]    23.75 24.07
```

The imputed values of **BMI** are impossible given the corresponding values of **height** and **weight**.

## 7. Imputation with mice()

### 7.4. Passive imputation

Moreover, if some components of a variable are observed we want to use that **information to reduce uncertainty**.

```
table(weight_missing = is.na(NHANES$weight),  
       height_missing = is.na(NHANES$height))
```

```
##           height_missing  
## weight_missing FALSE TRUE  
##           FALSE  2410   33  
##           TRUE    28   12
```

Here we have  $33 + 28 = 61$  cases in which either `height` or `weight` is observed.

We would like to impute `height` and `weight` separately and calculate `BMI` from the (imputed) values of the two variables.

## 7. Imputation with mice()

### 7.4. Passive imputation

If BMI is not a relevant predictor in any of the other imputation models, we could just exclude BMI from the imputation and **re-calculate it afterwards**.

To use BMI as predictor in the imputation, it has to be **calculated in each iteration** of the algorithm. In **mice** this is possible with **passive imputation**.

Instead of using a standard imputation **method**, we can specify a formula to calculate BMI:

```
meth["BMI"] <- "~I(weight/height^2)"      # specify formula to impute BMI
pred[c("weight", "height"), "BMI"] <- 0  # prevent feedback
```

To **prevent feedback** from BMI in the imputation of **height** and **weight** the **predictorMatrix** needs to be modified.

## 7. Imputation with mice()

### 7.4. Passive imputation

Since BMI depends on weight, and the two variables are highly correlated ( $\rho=0.87$ ) it may be beneficial **not to use them simultaneously** as predictors in the other imputation models.

Which one to use may differ between imputation models.

**Passive imputation** can also be useful in settings where

- imputation models include an **interaction terms** between incomplete variables (see [17, p. 133] for an example), or when
- a number of covariates is used to form a **sum score**. The sum score, instead of all single elements, can then be used as predictor in other imputation models.

## 7. Imputation with mice()

### 7.5. Post processing

`mice()` has an argument `post` that can be used to specify functions that modify imputed values.

Helpful functions are

- `squeeze()` to censor variables at given boundaries
- ~~`ifdo()` for conditional manipulation~~ (not yet implemented)

#### Example:

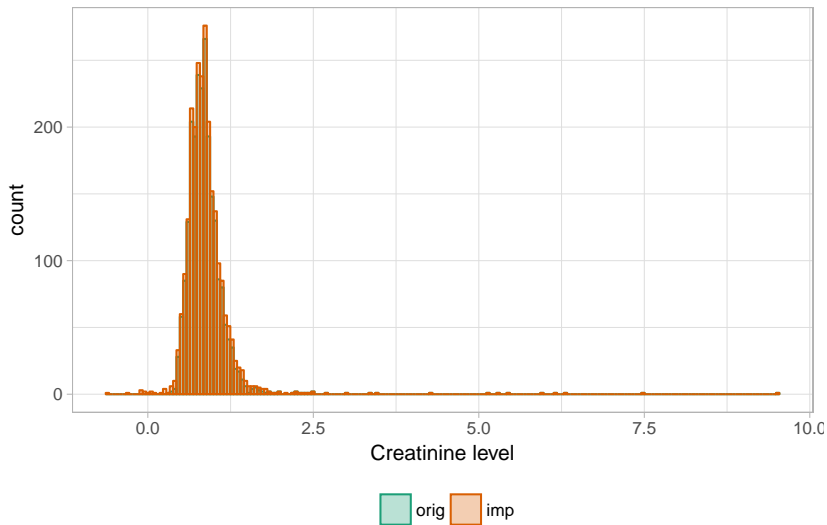
When inspecting the imputed values from `imp`, we find that some imputed values in `creat` are negative.

```
summary(DF1$creat) # DF1 is the first imputed dataset we extracted earlier

##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
## -0.6155  0.7000  0.8300  0.8853  0.9900  9.5100
```

## 7. Imputation with mice()

### 7.5. Post processing





## 7. Imputation with mice()

### 7.5. Post processing

With the following syntax all imputed values of `creat` that are outside the interval `c(0, 100)` will be **set to those limiting values**.

```
post <- imp$post
post["creat"] <- "imp[[j]][,i] <- squeeze(imp[[j]][,i], c(0, 100))"
imp2 <- update(imp, post = post, maxit = 20, seed = 123)
```

#### Note:

When many observations are outside the limits it may be better to **change the imputation model** since the implied **assumption of the imputation model** apparently **does not fit the** (assumption about the) **complete data distribution**.

## 7. Imputation with mice()

### 7.5. Post processing

This **post-processing** of imputed values allows for many **more data manipulations** and is not restricted to `squeeze()` (and `ifdo()`). Any strings of R commands provided will be evaluated after the corresponding variable is imputed, within each iteration.

For example, if subjects with  $SBP > 140$  should be classified as hypertensive:

```
post["hypten"] <- "imp[[j]][p$data[where[, j], 'SBP'] > 140, i] <- 'yes'"
```

This also allows for (some) **MNAR scenarios**, for example, by multiplying or adding a constant to the imputed values or to re-impute values, depending on their current value.

## 7. Imputation with `mice()`

### 7.6. Visit sequence

When the **post-processed or passively imputed values** of a variable depend on other variables, the **sequence in which the variables are imputed** may be important to obtain **consistent values**.

#### Example:

If `BMI` is passively imputed (calculated) before the new imputations for `height` and `weight` are drawn, the resulting values of `BMI`, will match `height` and `weight` from the **previous iteration**, but not the iteration given in the imputed dataset.

In `mice()` the argument `visitSequence` specifies in which order the columns of the data are imputed. By default `mice()` imputes in the order of the columns in data.

## 7. Imputation with mice()

### 7.6. Visit sequence

```
visitSeq <- imp2$visitSequence  
visitSeq
```

```
##      bili      chol      HDL      hypten      hypchol      smoke      alc      educ  
##         4         5         6         7         8         10        11        12  
##      SBP HyperMed      creat      albu uricacid      WC      height      weight  
##        13        14        15        16        17        18        19        20  
##      BMI  
##        21
```

Currently, `hypten` is imputed before `SBP`, but the imputed values of `hypten` are post-processed depending on the current value of `SBP`. To get consistent values of these two variables, we need to change the `visitSequence`.

## 7. Imputation with mice()

### 7.6. Visit sequence

```
visitSeq <- c(visitSeq[-which(names(visitSeq) == "hypten")],  
             visitSeq["hypten"])
```

```
visitSeq
```

##	bili	chol	HDL	hypchol	smoke	alc	educ	SBP
##	4	5	6	8	10	11	12	13
##	HyperMed	creat	albu	uricacid	WC	height	weight	BMI
##	14	15	16	17	18	19	20	21
##	hypten							
##	7							

The `visitSequence` may specify that a column is visited multiple times during one iteration. All incomplete variables must be visited at least once.

`visitSequence` can also be specified using one of the keywords `"roman"` (left to right), `"arabic"` (right to left), `"monotone"` (sorted in increasing amount of missingness), `"revmonotone"` (reverse of monotone)

## 7. Imputation with `mice()`

### 7.7. Good to know

`mice()` performs some **pre-processing** and **removes**

- incomplete variables that are not imputed but are specified as predictor,
- constant variables, and
- collinear variables.

In each iteration

- linearly dependent variables are removed and
- `polr` imputation models that do not converge are replaced by `polyreg`.

### **Why?**

To avoid problems in the imputation models.

## 7. Imputation with `mice()`

### 7.7. Good to know

As a **consequence**

- imputation models may differ from what the user has specified or assumes is happening, or
- variables that should be imputed are not.

- ➔ Know your data
- ➔ Make sure `method` and `predictorMatrix` are specified appropriately
- ➔ Check the output and log of these automatic actions carefully

## 7. Imputation with mice()

A note

*“Please realize that these choices are always needed. Imputation software needs to make default choices. These choices are intended to be useful across a wide range of applications. However, the **default choices are not necessarily the best for the data at hand. There is simply no magical setting that always works**, so often some tailoring is needed.” [17, p. 124]*



## 8. Convergence & Diagnostics

### 8.1. Logged events

The log of the automatic changes (slide 89) is returned as part of the `mids` object:

```
head(imp2$loggedEvents)
```

```
##   it im co   dep    meth out
## 1  1  1 10 smoke multinom
## 2  1  1 11  alc multinom
## 3  1  1 12 educ multinom
## 4  1  2 10 smoke multinom
## 5  1  2 11  alc multinom
## 6  1  2 12 educ multinom
```

With columns

<code>it</code>	iteration number
<code>im</code>	imputation number
<code>co</code>	column number in the data
<code>dep</code>	dependent variable
<code>meth</code>	imputation method used
<code>out</code>	names of altered or removed predictors

## 8. Convergence & Diagnostics

### 8.2. Convergence

Recall from slides 19 and 23:

**mice** uses an **iterative algorithm** and imputations from the first few iterations may not be samples from the “correct” distributions.

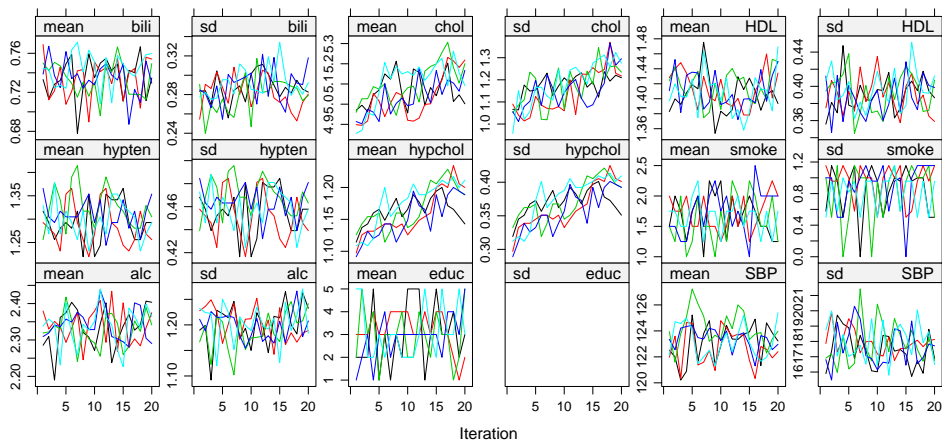
**Traceplots** can be used to visually assess **convergence**.

In **mice** the function `plot()` produces traceplots of the mean and standard deviation (across subjects) per incomplete variable (see slide 25).

# 8. Convergence & Diagnostics

## 8.2. Convergence

```
plot(imp2, layout = c(6, 3))
```



## 8. Convergence & Diagnostics

### 8.2. Convergence

The traceplots show that the imputations for `chol` and `hypchol` have an upward trend.

**Strong trends** and traces that show **correlation** between variables indicate **problems of feedback**. This needs to be investigated and resolved in the specification of the `predictorMatrix`.

**Weak trends** may be artefacts that often disappear when the imputation is performed with more iterations.

## 8. Convergence & Diagnostics

### 8.3. Diagnostics

When MCMC chains have converged, the **distributions of the imputed and observed values** can be compared to investigate differences between observed and imputed data.

**Note:**

Plots usually show the **marginal** distributions of observed and imputed values, which do not have to be identical under MAR.

**Recall:** the **conditional** distributions (given all the other variables in the imputation model) of the imputed values are assumed to be the same as the conditional distributions of the observed data.

## 8. Convergence & Diagnostics

### 8.3. Diagnostics

**mice** provides several functions for visual diagnosis of imputed values:

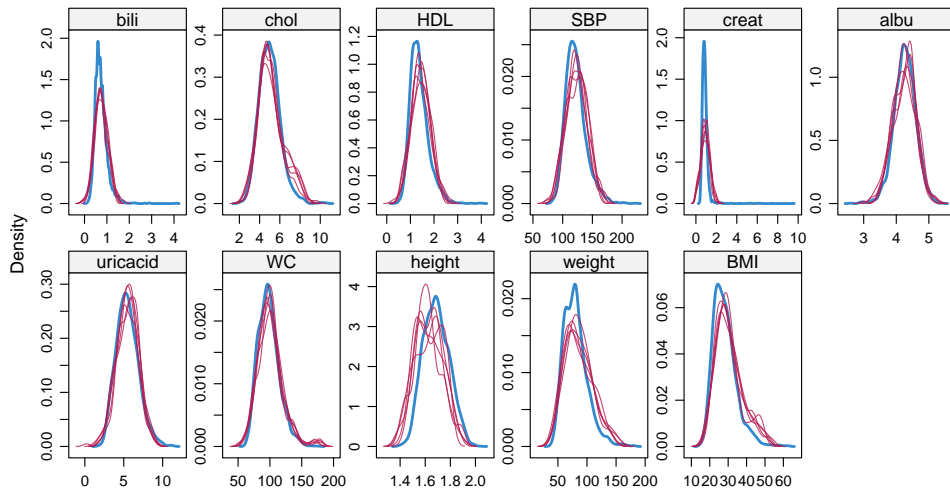
- `densityplot()` (for large datasets and variables with many NAs)
- `stripplot()` (for smaller datasets and/or variables with few NAs)
- `bwplot()`
- `xyplot()`

These functions create `lattice graphics`, which can be modified analogous to their parent functions from the **lattice** package.

## 8. Convergence & Diagnostics

### 8.3. Diagnostics

```
densityplot(imp2)
```



## 8. Convergence & Diagnostics

### 8.3. Diagnostics

The `densityplot()` shows that the distribution of imputed values of `creat` is wider than the distribution of the observed values and that imputed values of `height` are smaller than the observed values.

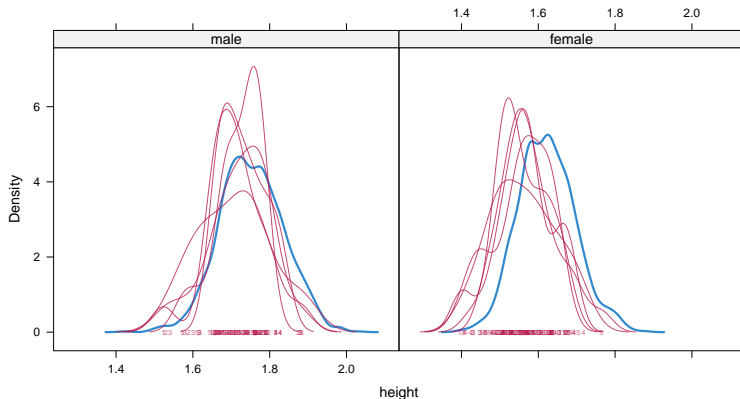


## 8. Convergence & Diagnostics

### 8.3. Diagnostics

In some cases differences in distributions can be explained by strata in the data, however, here, **gender** does not explain the difference in observed and imputed values.

```
densityplot(imp2, ~height|gender, plot.points = T)
```



## 8. Convergence & Diagnostics

### 8.3. Diagnostics

As an alternative, we might consider `race` to explain the differences

```
densityplot(imp2, ~height|race)

## Error in density.default(x = c(NA, NA, NA, NA, NA, NA, NA, NA, NA, NA, :
## need at least 2 points to select a bandwidth automatically
```

However, there are not enough missing values of `height` per categories of `race` to estimate densities.

```
with(NHANES, table(race = race, "height missing" = is.na(height)))
```

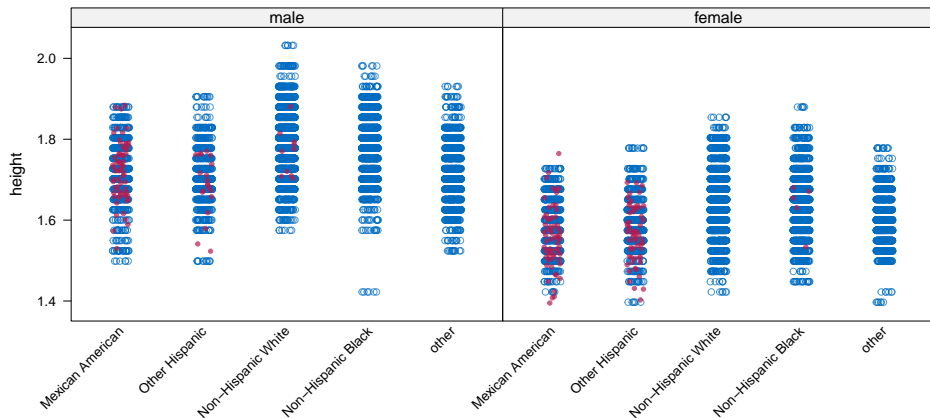
##		height missing	
## race		FALSE	TRUE
## Mexican American		233	26
## Other Hispanic		252	16
## Non-Hispanic White		884	2
## Non-Hispanic Black		618	1
## other		451	0

## 8. Convergence & Diagnostics

### 8.3. Diagnostics

In that case, a `stripplot()` may be better suited. Here we can also split the data for `gender` and `race`.

```
stripplot(imp2, height ~ race|gender, pch = c(1, 20),  
          scales = list(x = list(rot = 45)))
```



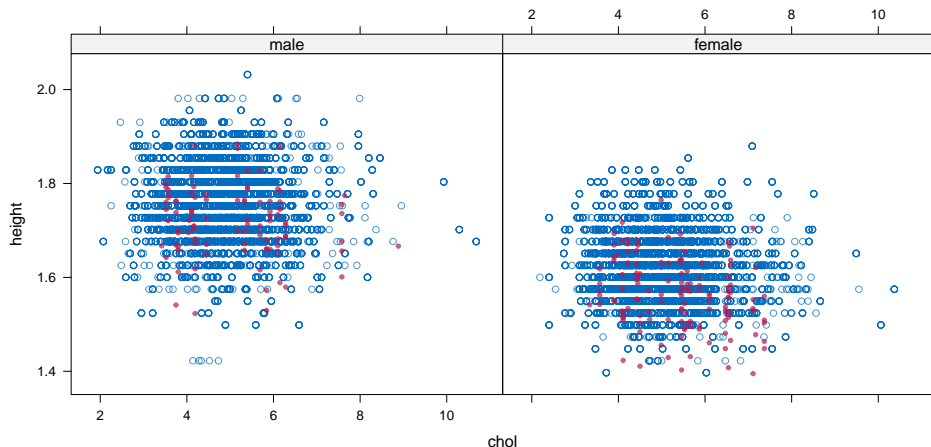


## 8. Convergence & Diagnostics

### 8.3. Diagnostics

The function `xyplot()` allows multivariate investigation of the imputed versus observed values.

```
xyplot(imp2, height ~ chol|gender, pch = c(1,20))
```



## 8. Convergence & Diagnostics

### 8.3. Diagnostics

All of the above graphs displayed only continuous imputed variables. For categorical variables we can compare the proportion of values in each category.

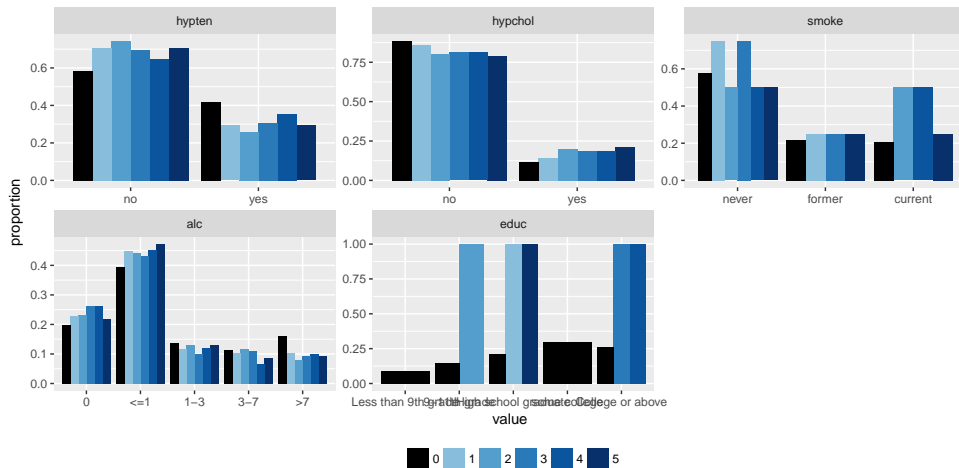
**mice** does not provide a function to do this, but we can write one ourselves, as for instance the function `probplot()`, for which the syntax can be found [here](#). The function can be loaded into R using:

```
devtools::source_gist("0d00375da460dd33839b98faeee2fdab",  
                      filename = "probplot.R")
```

# 8. Convergence & Diagnostics

## 8.3. Diagnostics

```
probplot(imp2)
```



## 8. Convergence & Diagnostics

### 8.3. Diagnostics

`smoke` and `educ` have very few missing values (4 and 1, respectively), so we do not need to worry about differences between observed and imputed data for those variables.

For `alc`, missing values are imputed by the lower consumption categories more often than we would expect from the observed data, `hyp ten` is less frequent and `hyp chol` a bit more frequent, in the imputed data compared to the observed.

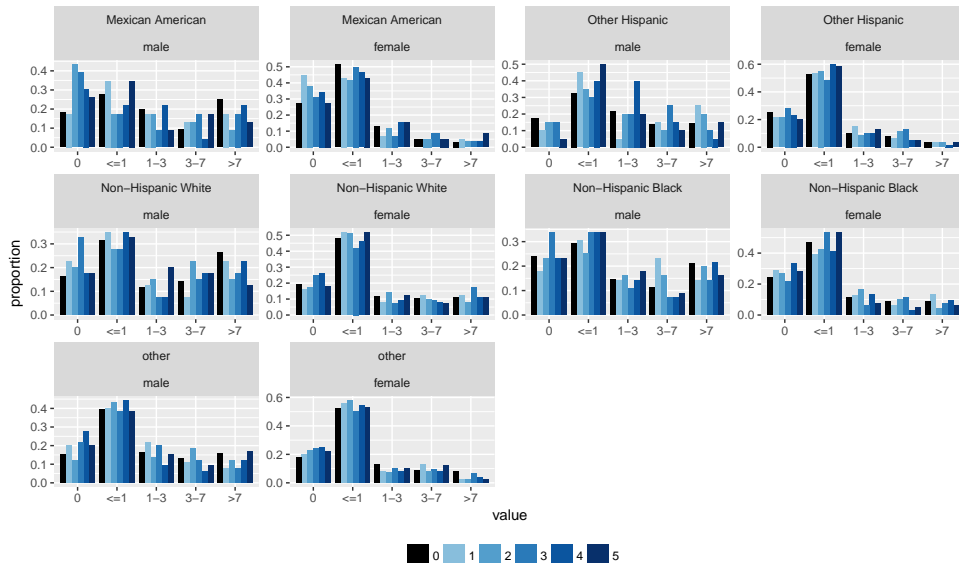
If we expect that `gender` and `race` might explain the differences for `hyp ten`, we can include those factors into the plot.



# 8. Convergence & Diagnostics

## 8.3. Diagnostics

```
probplot(imp2, formula = alc ~ race + gender)
```



## 8. Convergence & Diagnostics

### 8.3. Diagnostics

Since hypertension is more common in older individuals, we may want to investigate if `age` can explain the differences in imputed values of `hypten`.

```
round(sapply(split(NHANES[, "age"], addNA(NHANES$hypten)), summary), 1)
```

```
##           no  yes <NA>
## Min.      20.0 20.0 20.0
## 1st Qu.    28.0 47.0 30.0
## Median     38.0 59.0 38.5
## Mean       40.7 56.9 41.5
## 3rd Qu.    51.0 68.0 50.8
## Max.       79.0 79.0 78.0
```

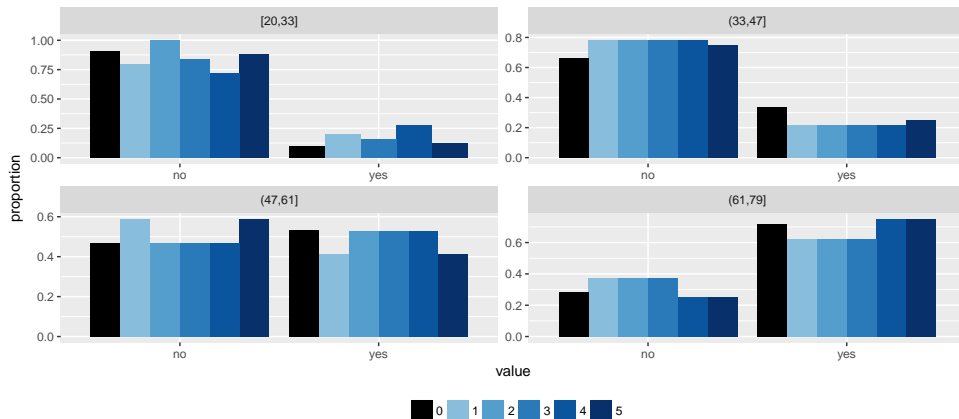
The table shows that the distribution of `age` in participants with missing `hypten` is very similar to the distribution of `age` in participants without `hypten`.

## 8. Convergence & Diagnostics

### 8.3. Diagnostics

Plotting the proportions of observed and imputed `hypten` separately per quartile of `age`:

```
probplot(imp2, formula = hypten ~ cut(age, quantile(age), include.lowest = T))
```



## 9. Analyse & pool the imputed data

### 9.1. Analyzing imputed data

Once we have confirmed that our imputation was successful, we can move on to the analysis of the imputed data.

For example, we might be interested in the following logistic regression model:

```
glm(DM ~ age + gender + hypchol + BMI + smoke + alc,  
    family = "binomial")
```

To fit the model on each of the imputed datasets, we do not need to extract the data from the `mids` object, but can use `with()`.

```
mod1 <- with(imp2, glm(DM ~ age + gender + hypchol + BMI + smoke + alc,  
                      family = "binomial"))
```

## 9. Analyse & pool the imputed data

### 9.1. Analyzing imputed data

`mod1` is an object of class `mira` and has elements

```
names(mod1)
## [1] "call"      "call1"     "nmis"      "analyses"
```

where `mod1$analyses` is a list containing the fitted model for each imputed dataset.

## 9. Analyse & pool the imputed data

### 9.2. Pooling results

Pooled results can be obtained using `pool()` and its summary.

```
options(width = 90)
res1 <- summary(pool(mod1))
round(res1, 3)
```

##	est	se	t	df	Pr(> t )	lo 95	hi 95	nmis	fmi	lambda
## (Intercept)	-7.133	0.429	-16.616	2240.573	0.000	-7.975	-6.291	NA	0.013	0.012
## age	0.056	0.004	12.952	2468.059	0.000	0.048	0.065	0	0.001	0.001
## gender2	-0.422	0.128	-3.304	1749.968	0.001	-0.673	-0.172	NA	0.026	0.025
## hypchol2	-0.064	0.188	-0.342	403.591	0.732	-0.434	0.305	NA	0.095	0.090
## BMI	0.106	0.009	11.576	2265.730	0.000	0.088	0.123	73	0.012	0.011
## smoke2	0.129	0.144	0.896	2432.312	0.370	-0.153	0.411	NA	0.005	0.004
## smoke3	0.080	0.166	0.479	1953.715	0.632	-0.246	0.405	NA	0.021	0.020
## alc2	-0.277	0.150	-1.845	475.882	0.066	-0.573	0.018	NA	0.085	0.082
## alc3	-0.570	0.220	-2.585	1192.205	0.010	-1.002	-0.137	NA	0.042	0.041
## alc4	-0.466	0.246	-1.894	154.639	0.060	-0.952	0.020	NA	0.165	0.155
## alc5	-0.741	0.220	-3.375	747.163	0.001	-1.172	-0.310	NA	0.063	0.060

## 9. Analyse & pool the imputed data

### 9.2. Pooling results

Pooling with `mice::pool()` is available for most types of models.

Generally, it works for models for which the functions `coef()` and `vcov()` can extract the (fixed effects) coefficients and variance-covariance matrix of these coefficients.

An alternative is offered by the package **mitools** and the function `MIcombine()`.

## 9. Analyse & pool the imputed data

### 9.3. Functions for pooled results

**mice** currently has two functions available for evaluating model fit / model comparison

For **linear** regression models the pooled  $R^2$  can be calculated using `pool.r.squared()`

```
mod2 <- with(imp2, lm(SBP ~ DM + age + hypten))
pool.r.squared(mod2, adjusted = TRUE)

##               est         lo 95         hi 95         fmi
## adj R^2 0.3243655 0.2940335 0.3547556 0.006885114
```

The argument `adjusted` specifies whether the adjusted  $R^2$  or the standard  $R^2$  is returned.



## 9. Analyse & pool the imputed data

### 9.3. Functions for pooled results

The function `pool.compare()` allows to compare **nested models** (i.e., models where one is a special case of the other, with some parameters fixed to zero) using a **Wald test**.

**Example:** To test if `smoke` has a relevant contribution to the model for `DM` from above we re-fit the model without `smoke` and compare the two models:

```
mod3 <- with(imp2, glm(DM ~ age + gender + hypchol + BMI + alc,
                      family = "binomial"))

# Wald test
pool.compare(mod1, mod3)$pvalue

##           [,1]
## [1,] 0.6577086
```

## 9. Analyse & pool the imputed data

### 9.3. Functions for pooled results

The package **miceadds** extends **mice**, for example with the following functionality:

**Combine  $\chi^2$  or F statistics from multiply imputed data:**

```
miceadds::micombine.chisquare(dk, df, display = TRUE, version = 1)  
miceadds::micombine.F(values, df1, display = TRUE, version = 1)
```

These functions take vectors of statistics computed on each imputed dataset and pool them.

## 9. Analyse & pool the imputed data

### 9.3. Functions for pooled results

The package **miceadds** extends **mice**, for example with the following functionality:

#### Combine $\chi^2$ or F statistics from multiply imputed data:

```
miceadds::micombine.chisquare(dk, df, display = TRUE, version = 1)
miceadds::micombine.F(values, df1, display = TRUE, version = 1)
```

These functions take vectors of statistics computed on each imputed dataset and pool them.

#### Calculate correlation or covariance of imputed data:

```
miceadds::micombine.cor(mi.res, variables = NULL, conf.level = 0.95,
                        method = "pearson", nested = FALSE, partial = NULL)
miceadds::micombine.cov(mi.res, variables = NULL, conf.level = 0.95,
                        nested = FALSE)
```

These functions take `mids` objects as input.

## 10. Additional functions in `mice()`

### 10.1. Extracting and exporting imputed data

The function `complete()` allows **extraction of the imputed data** from a `mids` object:

```
mice::complete(x, action = 1, include = FALSE)
```

- `x`: the `mids` object
- `action`:
  - `1, ..., m` (single imputed dataset)
  - `"long"`: long format (imputed data stacked vertically)
  - `"broad"`: wide format (imputed data combined horizontally; ordered by imputation)
  - `"repeated"`: (like `"broad"`, but ordered by variable)
- `include`: include the original data?  
(if `action` is `"long"`, `"broad"` or `"repeated"`)

## 10. Additional functions in mice()

### 10.1. Extracting and exporting imputed data

The function `mids2spss()` allows the **export of imputed data** (`mids` objects) to SPSS.

```
mids2spss(imp2,  
  filedat = "datafile.txt", # the file containing the data  
  filesps = "importsyntax.sps", # syntax to convert .txt to .sav  
  silent = TRUE  
)
```

Data from `mids` objects can also be exported to MPLUS using `mids2mplus()`.

## 10. Additional functions in mice()

### 10.2. Combining `mids` objects

To **increase the number of imputed datasets** without re-doing the initial  $m$  imputations, a second set of imputations can be done and the two `mids` objects combined using `ibind()`.

```
# same syntax as before, but different seed
imp2b <- update(imp2, post = post, maxit = 20, seed = 456)
imp2combi <- ibind(imp2, imp2b)
imp2combi$m
```

```
## [1] 10
```

## 10. Additional functions in mice()

### 10.3. Adding variables to mids objects

The function `cbind.mids()` allows to **add columns** to a `mids` object. The extra columns can either be a `data.frame`, `matrix`, `vector` or `factor` or another `mids` object.

For example data columns that should be part of the imputed data for completeness, but are not needed in the imputation.

```
extravar <- rnorm(nrow(NHANES))  
impextra <- mice::cbind.mids(x = imp2, extravar = extravar)
```

**Note:** `cbind()` just adds columns to the data, you need to make sure they are **sorted correctly** so that the rows of the new data are from the same subjects as the corresponding rows in the impute data.

# 11. Multiple Imputation in SPSS

## 11.1. Where to get help

A walk-through how to do multiple imputation in SPSS can be found

- **for older versions of SPSS**

- > Help
  - > Case Studies
    - > Missing Values Option
      - > Multiple Imputation
        - > Using Multiple Imputation to Complete and Analyze a Dataset

- **for newer versions online**

[https://www.ibm.com/support/knowledgecenter/en/SSLVMB\\_24.0.0/spss/tutorials/mi\\_table.html](https://www.ibm.com/support/knowledgecenter/en/SSLVMB_24.0.0/spss/tutorials/mi_table.html)



# 11. Multiple Imputation in SPSS

## 11.1. Where to get help

A walk-through how to do multiple imputation in SPSS can be found

- **for older versions of SPSS**

- > Help
  - > Case Studies
    - > Missing Values Option
      - > Multiple Imputation
        - > Using Multiple Imputation to Complete and Analyze a Dataset

- **for newer versions online**

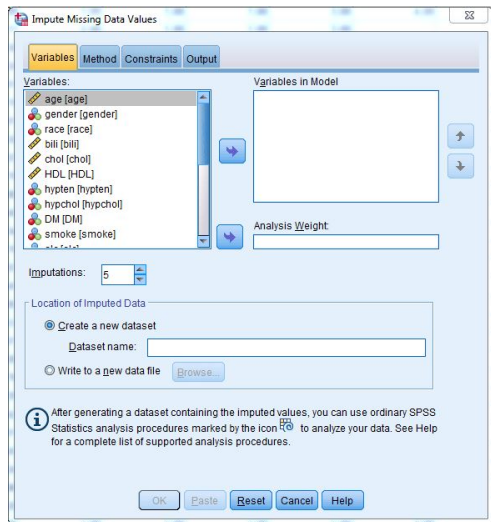
[https://www.ibm.com/support/knowledgecenter/en/SSLVMB\\_24.0.0/spss/tutorials/mi\\_table.html](https://www.ibm.com/support/knowledgecenter/en/SSLVMB_24.0.0/spss/tutorials/mi_table.html)

The **procedure** itself is located in the menu

- > Analyze
  - > Multiple Imputation
    - > Impute Missing Data Values

# 11. Multiple Imputation in SPSS

## 11.2. Multiple Imputation Features

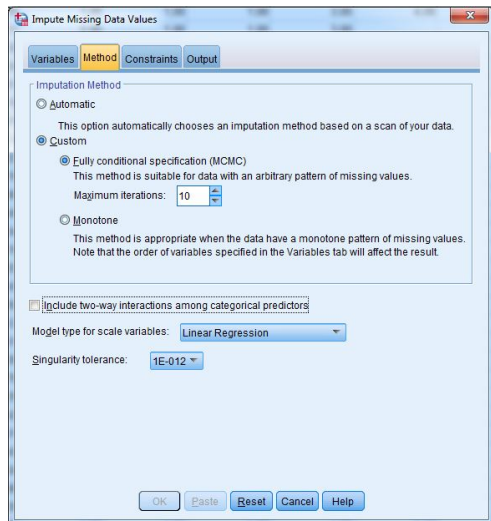


SPSS lets you

- specify number of imputations

# 11. Multiple Imputation in SPSS

## 11.2. Multiple Imputation Features

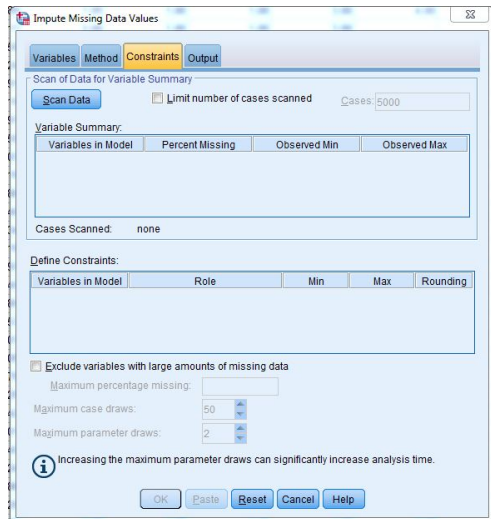


SPSS lets you

- specify number of imputations
- specify number of iterations
- include interactions
- choose between lin. regression and pmm for continuous variables

# 11. Multiple Imputation in SPSS

## 11.2. Multiple Imputation Features

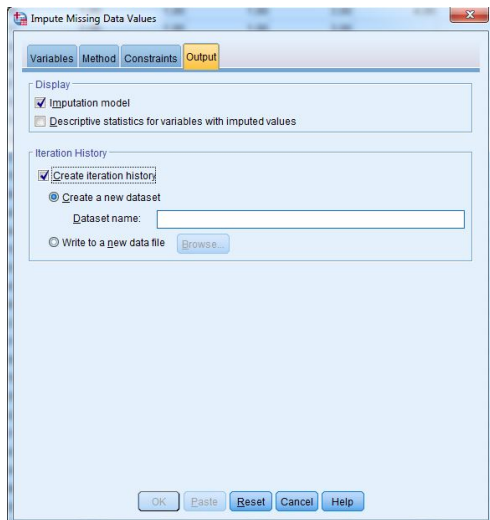


SPSS lets you

- specify number of imputations
- specify number of iterations
- include interactions
- chose between lin. regression and pmm for continuous variables
- restrict variables to certain values
- select which variables to impute
- select which variables are used as predictors

# 11. Multiple Imputation in SPSS

## 11.2. Multiple Imputation Features



SPSS lets you

- specify number of imputations
- specify number of iterations
- include interactions
- chose between lin. regression and pmm for continuous variables
- restrict variables to certain values
- select which variables to impute
- select which variables are used as predictors
- save the iteration history

# 11. Multiple Imputation in SPSS

## 11.2. Multiple Imputation Features

SPSS does not let you

- select between linear regression imputation and pmm **per** variable (only jointly for all variables)
- uses **only one donor** in predictive mean matching
- uses logistic regression for any categorical variable
- chose per imputation model which variables should be used as predictors
- re-calculate variables during the iterations
- ...

# 11. Multiple Imputation in SPSS

## 11.2. Multiple Imputation Features

In SPSS the **list of models that can be pooled** is available in the help under

- > Help
  - > Missing Values Option
    - > Multiple Imputation
      - > Analyzing Multiple Imputation Data

[https://www.ibm.com/support/knowledgecenter/en/SSLVMB\\_24.0.0/spss/mva/mi\\_analysis.html](https://www.ibm.com/support/knowledgecenter/en/SSLVMB_24.0.0/spss/mva/mi_analysis.html)

## Part III

### When MICE might fail



# Outline of Part III

## 12. Settings where MICE may have problems

- 12.1 Example: Quadratic effect

- 12.2 Example: Interaction effect

- 12.3 Example: Longitudinal outcome

- 12.4 Example: Survival data

## 13. Requirements for MICE to work (well)

- 13.1 Joint and conditional distributions

- 13.2 Some conditions and definitions

- 13.3 Why imputation with MICE can go wrong

## 14. Alternatives to MICE

- 14.1 Joint model imputation

- 14.2 Multivariate Normal Model

- 14.3 Sequential Factorization

# Outline of Part III (cont.)

## 15. Imputation with non-linear functional forms

- 15.1 R package mice
- 15.2 R package JointAI
- 15.3 R package smcfcs
- 15.4 R package jomo
- 15.5 Comparison of results

## 16. Imputation of longitudinal data

- 16.1 R package mice
- 16.2 R package JointAI
- 16.3 R package jomo
- 16.4 Comparison of results

# Outline of Part III (cont.)

## 17. Imputation of survival data

- 17.1 Results from literature

- 17.2 R package mice

- 17.3 R package smcfcs

- 17.4 R package jomo

- 17.5 Comparison of results

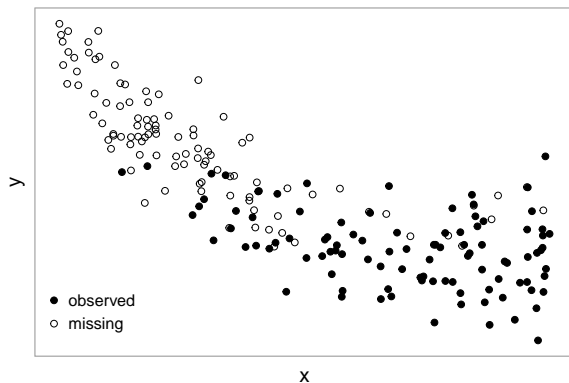
## 12. Settings where MICE may have problems

### 12.1. Example: Quadratic effect

Consider the case where the **analysis model** (which we assume to be true) is

$$y = \beta_0 + \beta_1 x + \beta_2 x^2 + \dots,$$

i.e.,  $y$  has a **quadratic relationship** with  $x$ , and  $x$  is incomplete.



The original data show a curved pattern.

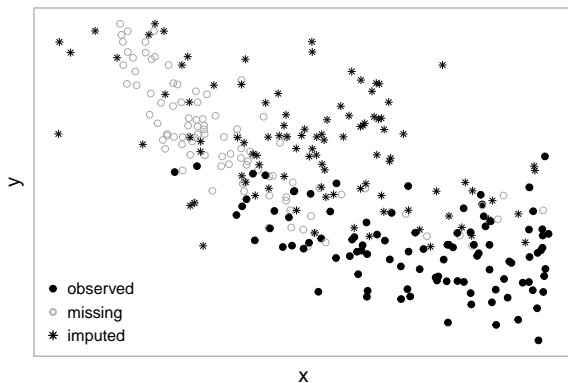
## 12. Settings where MICE may have problems

### 12.1. Example: Quadratic effect

The model used to **impute**  $x$  when using MICE (naively) is

$$x = \theta_{10} + \theta_{11}y + \dots,$$

i.e., a **linear relation** between  $x$  and  $y$  is assumed.

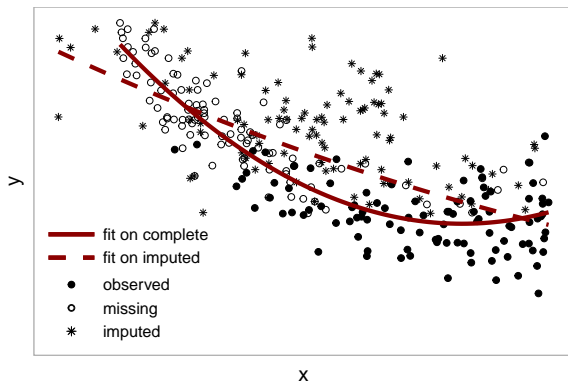


The imputed values **distort the curved pattern** of the original data.

## 12. Settings where MICE may have problems

### 12.1. Example: Quadratic effect

The model fitted on the imputed data gives **severely biased results**; the non-linear shape of the curve has almost completely disappeared.



	$\beta$	95% CI
<b>Original</b>		
Intercept	-0.99	[-1.04, -0.95]
$x$	-0.61	[-0.66, -0.56]
$x^2$	0.52	[0.43, 0.62]
<b>Imputed</b>		
Intercept	-0.73	[-0.79, -0.66]
$x$	-0.53	[-0.62, -0.44]
$x^2$	0.07	[-0.07, 0.22]

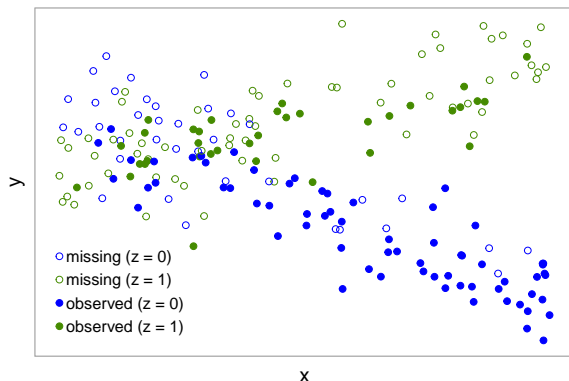
## 12. Settings where MICE may have problems

### 12.2. Example: Interaction effect

Another example occurs when the analysis model (again, assumed to be true) is

$$y = \beta_0 + \beta_x x + \beta_z z + \beta_{xz} xz + \dots,$$

i.e.,  $y$  has a **non-linear relationship** with  $x$  due to the **interaction term**.



The original data shows a “<” shaped pattern.

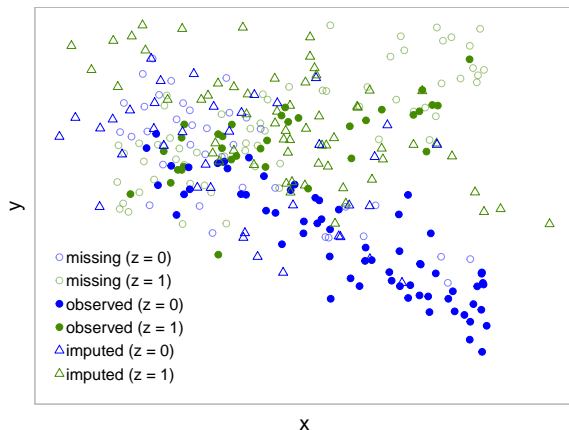
## 12. Settings where MICE may have problems

### 12.2. Example: Interaction effect

The model used to impute  $x$  when using MICE (naively) is

$$x = \theta_{10} + \theta_{11}y + \theta_{12}z + \dots,$$

i.e., a linear relation between  $x$  and  $y$  is assumed.



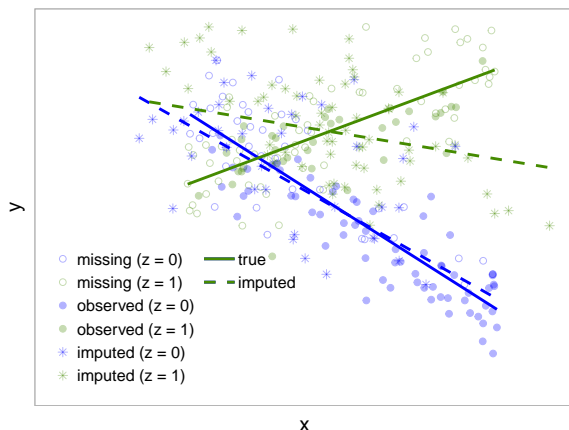
The “<” shaped pattern of the true data is **distorted by the imputed values**.



## 12. Settings where MICE may have problems

### 12.2. Example: Interaction effect

And the analysis on these naively imputed values leads to **severely biased estimates**.

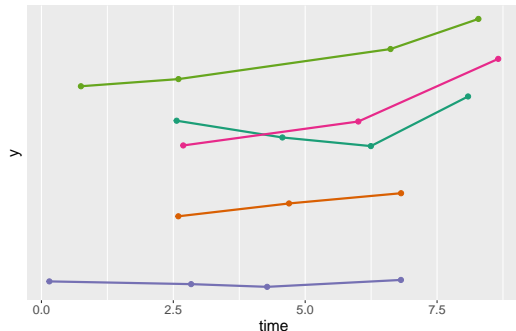


	$\beta$	95% CI
<b>Original</b>		
Intercept	-0.96	[-1.00, -0.92]
x	-0.59	[-0.65, -0.53]
z	0.5	[0.45, 0.56]
x:z	0.94	[0.85, 1.03]
<b>Imputed</b>		
Intercept	-0.96	[-1.01, -0.91]
x	-0.52	[-0.61, -0.44]
z	0.46	[0.39, 0.54]
x:z	0.37	[0.24, 0.51]

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

Another setting where imputation with MICE is not straightforward is when the **outcome variable is longitudinal**.



ID	y	x <sub>1</sub>	x <sub>2</sub>	x <sub>3</sub>	x <sub>4</sub>	time
5	✓	✓	NA	✓	✓	2.56
5	✓	✓	NA	✓	✓	4.57
5	✓	✓	NA	✓	✓	6.25
5	✓	✓	NA	✓	✓	8.09
6	✓	✓	NA	NA	✓	2.60
6	✓	✓	NA	NA	✓	4.69
6	✓	✓	NA	NA	✓	6.82
8	✓	✓	✓	✓	NA	2.69
8	✓	✓	✓	✓	NA	6.01
8	✓	✓	✓	✓	NA	8.66
18	✓	✓	NA	✓	✓	0.75
18	✓	✓	NA	✓	✓	2.60
18	✓	✓	NA	✓	✓	6.62
18	✓	✓	NA	✓	✓	8.28
⋮	⋮	⋮	⋮	⋮	⋮	⋮

Here,  $x_1, \dots, x_4$  are baseline covariates, i.e., not measured repeatedly.

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

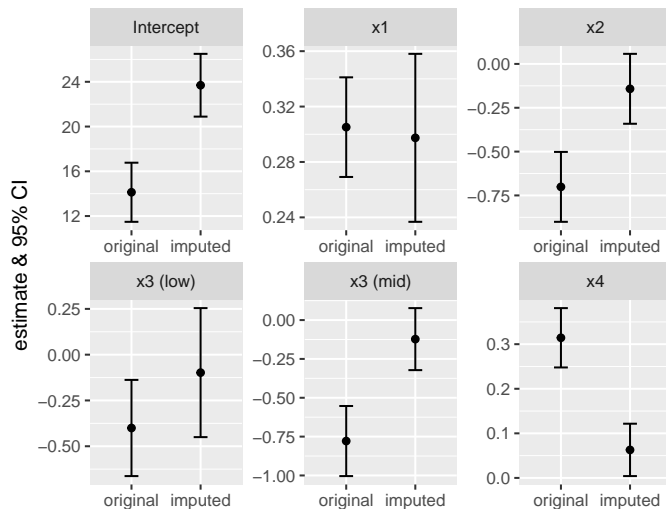
If we use MICE in the data in this (long) format, each row would be regarded as independent, which may cause bias and **inconsistent imputations**.

Imputed values of baseline covariates are imputed with different values, creating data that could not have been observed.

ID	y	x <sub>1</sub>	x <sub>2</sub>	x <sub>3</sub>	x <sub>4</sub>	time
5	✓	✓	boy	✓	✓	2.56
5	✓	✓	girl	✓	✓	4.57
5	✓	✓	girl	✓	✓	6.25
5	✓	✓	girl	✓	✓	8.09
6	✓	✓	girl	high	✓	2.60
6	✓	✓	boy	mid	✓	4.69
6	✓	✓	girl	high	✓	6.82
8	✓	✓	✓	✓	38.27	2.69
8	✓	✓	✓	✓	38.45	6.01
8	✓	✓	✓	✓	40.71	8.66
18	✓	✓	boy	✓	✓	0.75
18	✓	✓	boy	✓	✓	2.60
18	✓	✓	boy	✓	✓	6.62
18	✓	✓	boy	✓	✓	8.28
⋮	⋮	⋮	⋮	⋮	⋮	⋮

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

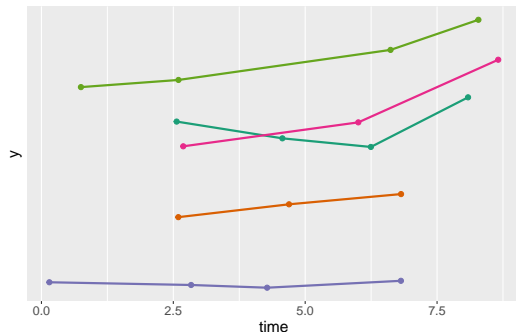


Estimates can be severely biased.

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

In some settings **imputation in wide format** may be possible.

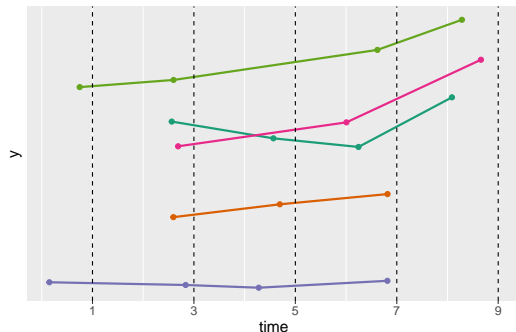


ID	y	x <sub>1</sub>	x <sub>2</sub>	x <sub>3</sub>	x <sub>4</sub>	time
5	✓	✓	NA	✓	✓	2.56
5	✓	✓	NA	✓	✓	4.57
5	✓	✓	NA	✓	✓	6.25
5	✓	✓	NA	✓	✓	8.09
6	✓	✓	NA	NA	✓	2.60
6	✓	✓	NA	NA	✓	4.69
6	✓	✓	NA	NA	✓	6.82
8	✓	✓	✓	✓	NA	2.69
8	✓	✓	✓	✓	NA	6.01
8	✓	✓	✓	✓	NA	8.66
18	✓	✓	NA	✓	✓	0.75
18	✓	✓	NA	✓	✓	2.60
18	✓	✓	NA	✓	✓	6.62
18	✓	✓	NA	✓	✓	8.28
⋮	⋮	⋮	⋮	⋮	⋮	⋮

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

In some settings **imputation in wide format** may be possible.



ID	y	x <sub>1</sub>	x <sub>2</sub>	x <sub>3</sub>	x <sub>4</sub>	time
5	✓	✓	NA	✓	✓	2.56
5	✓	✓	NA	✓	✓	4.57
5	✓	✓	NA	✓	✓	6.25
5	✓	✓	NA	✓	✓	8.09
6	✓	✓	NA	NA	✓	2.60
6	✓	✓	NA	NA	✓	4.69
6	✓	✓	NA	NA	✓	6.82
8	✓	✓	✓	✓	NA	2.69
8	✓	✓	✓	✓	NA	6.01
8	✓	✓	✓	✓	NA	8.66
18	✓	✓	NA	✓	✓	0.75
18	✓	✓	NA	✓	✓	2.60
18	✓	✓	NA	✓	✓	6.62
18	✓	✓	NA	✓	✓	8.28
⋮	⋮	⋮	⋮	⋮	⋮	⋮

## 12. Settings where MICE may have problems

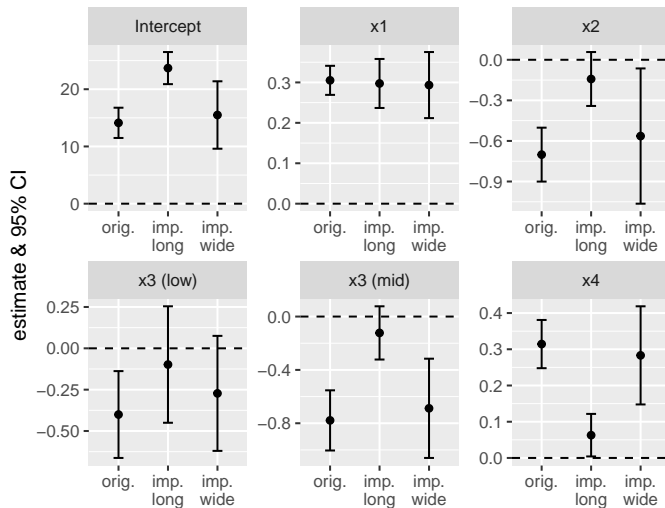
### 12.3. Example: Longitudinal outcome

id	y.1	y.3	y.5	y.7	y.9	time.1	time.3	time.5	time.7	time.9	...
5	NA	35.19	34.94	34.81	35.56	NA	2.56	4.57	6.25	8.09	...
6	NA	33.74	33.94	34.09	NA	NA	2.6	4.69	6.82	NA	...
8	NA	34.82	NA	35.18	36.13	NA	2.69	NA	6.01	8.66	...
18	35.71	35.82	NA	36.28	36.73	0.75	2.6	NA	6.62	8.28	...
⋮	⋮	⋮	⋮	⋮	⋮	⋮	⋮	⋮	⋮	⋮	⋮

In this **wide format data** frame, missing values in outcome and measurement times need to be imputed (to be able to use them as predictors to impute covariates), even though we would not need to impute them for the analysis (mixed model valid when outcome measurements are M(C)AR).

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

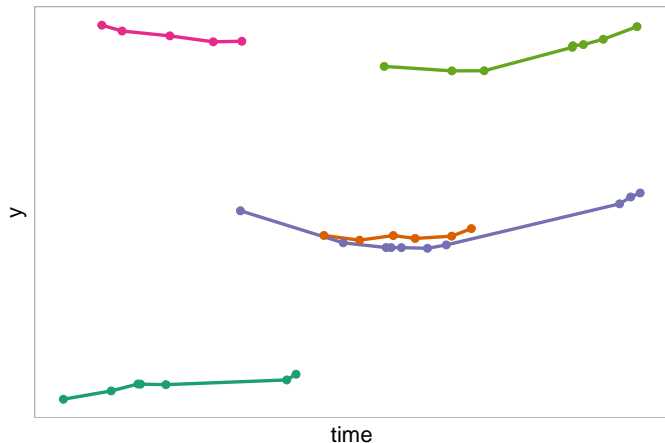


Better, but very large confidence intervals.



## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

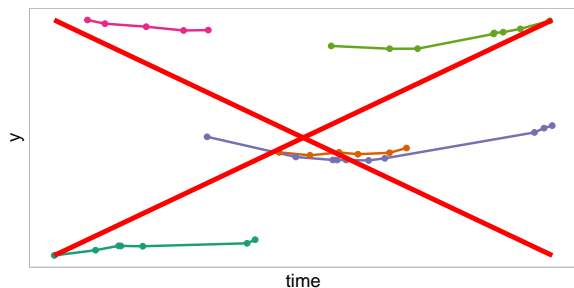


When the data is very **unbalanced**, i.e., there are no clear cut-offs in time, transformation to wide format is not possible.

(Or at least transformation to wide format leads to variables with high proportions of missing values.)

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

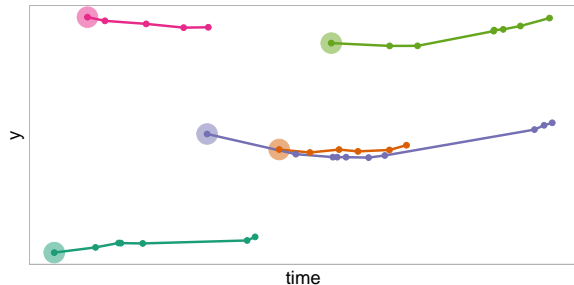
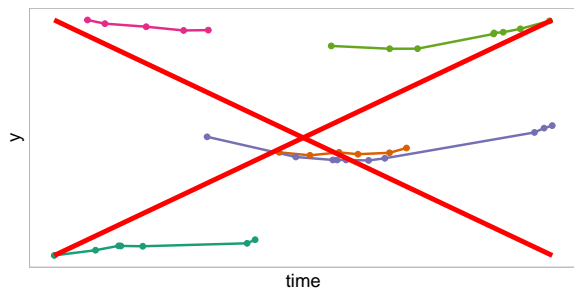


Naive approaches that are sometimes used are to

- **ignore the outcome** in the imputation

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome

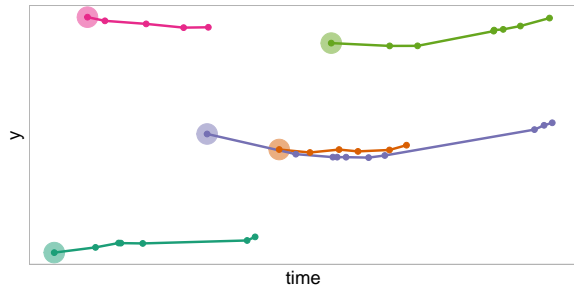
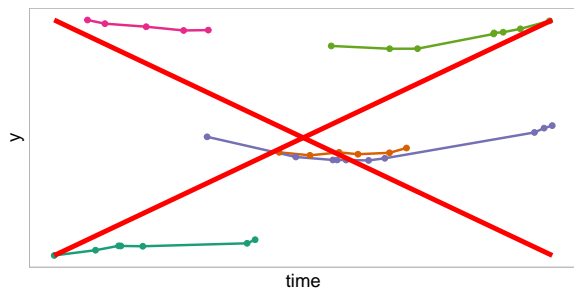


Naive approaches that are sometimes used are to

- **ignore the outcome** in the imputation, or to
- use only the **first/baseline outcome**

## 12. Settings where MICE may have problems

### 12.3. Example: Longitudinal outcome



Naive approaches that are sometimes used are to

- **ignore the outcome** in the imputation, or to
- use only the **first/baseline outcome**

However, **important information may be lost**, resulting in invalid imputations and biased results.

## 12. Settings where MICE may have problems

### 12.4. Example: Survival data

In **survival analysis**, the aim is to estimate the effect of covariates on the **time until an event** of interest happens.

## 12. Settings where MICE may have problems

### 12.4. Example: Survival data

In **survival analysis**, the aim is to estimate the effect of covariates on the **time until an event** of interest happens.

In the commonly used method: **Cox proportional hazards model**

$$h(t) = h_0(t) \exp(x\beta_x + z\beta_z),$$

- $h(t)$ : hazard = the instantaneous risk of an event at time  $t$ , given that the event has not occurred until time  $t$
- $h_0(t)$ : unspecified baseline hazard
- $x$  and  $z$ : **incomplete** and **complete** covariates, respectively

## 12. Settings where MICE may have problems

### 12.4. Example: Survival data

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- $h(t)$ : hazard = the instantaneous risk of an event at time  $t$ , given that the event has not occurred until time  $t$
- $h_0(t)$ : unspecified baseline hazard
- $x$  and  $z$ : **incomplete** and **complete** covariates, respectively

**Survival outcomes** are usually represented by the **observed event time**  $T$  and the **event indicator**  $D$  ( $D = 1$ : event,  $D = 0$ : censored).

## 12. Settings where MICE may have problems

### 12.4. Example: Survival data

**Naive use of MICE** treats the columns in the data set containing  $T$  and  $D$  just like any other variable, and the resulting imputation model for  $X$  would have the form

$$p(x \mid T, D, \mathbf{z}) = \theta_0 + \theta_1 T + \theta_2 D + \theta_3 z + \dots$$



## 12. Settings where MICE may have problems

### 12.4. Example: Survival data

**Naive use of MICE** treats the columns in the data set containing  $T$  and  $D$  just like any other variable, and the resulting imputation model for  $X$  would have the form

$$p(x \mid T, D, z) = \theta_0 + \theta_1 T + \theta_2 D + \theta_3 z + \dots$$

The correct conditional distribution of  $x$  given the other variables is, however,

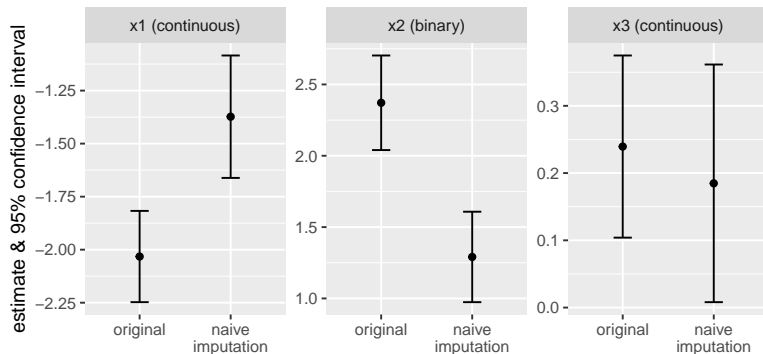
$$\log p(x \mid T, D, z) = \log p(x \mid z) + D(\beta_x x + \beta_z z) - \\ H_0(T) \exp(\beta_x x + \beta_z z) + \text{const.},$$

where  $H_0(T)$  is the cumulative baseline hazard.[20]

## 12. Settings where MICE may have problems

### 12.4. Example: Survival data

Using the naively assumed imputation model can lead to **severe bias**:



(Results from MICE imputation with two incomplete normal and one incomplete binary covariate.)

## 13. Requirements for MICE to work (well)

### 13.1. Joint and conditional distributions

**Recall:** The MICE algorithm is based on the idea of Gibbs sampling.

Gibbs sampling exploits the fact that a joint distribution is fully determined by its full conditional distributions.

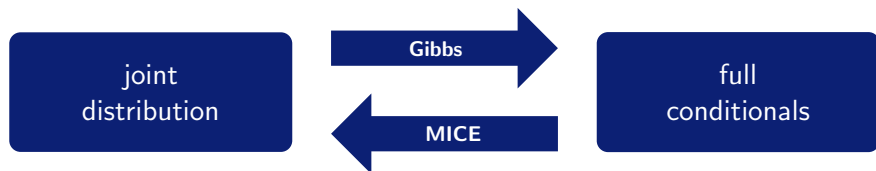


## 13. Requirements for MICE to work (well)

### 13.1. Joint and conditional distributions

**Recall:** The MICE algorithm is based on the idea of Gibbs sampling.

Gibbs sampling exploits the fact that a joint distribution is fully determined by its full conditional distributions.



In MICE, the full conditionals are not derived from the joint distribution: we directly specify the full conditionals and hope a joint distribution exists.

## 13. Requirements for MICE to work (well)

### 13.1. Joint and conditional distributions

The **uncertainty about whether a joint distribution exists** for the specified set of imputation models is often considered to be mainly a theoretical problem.

In practice, violations only have little impact on results in many applications.

However, as we have seen in the examples on the previous slides, there are **settings where the direct specification** of the full conditionals/imputation models **may lead to problems**, causing biased results.

# 13. Requirements for MICE to work (well)

## 13.2. Some conditions and definitions

Two important definitions:

### **Compatibility:**

*A joint distribution exists, that has the full conditionals (imputation models) as its conditional distributions.*

### **Congeniality:**

*The imputation model is compatible with the analysis model.*

# 13. Requirements for MICE to work (well)

## 13.2. Some conditions and definitions

**Important requirements** for MICE to work well include:

- Compatibility
- Congeniality
- MAR or MCAR (in the standard implementations)
- **all relevant variables** need to be included (omission might result in MNAR)
- **The outcome needs to be included** as predictor variable (but we usually do not impute missing outcome values)
- the imputation models (and analysis model) need to be **correctly specified** (which is a requirement in any standard analysis)

## 13. Requirements for MICE to work (well)

### 13.3. Why imputation with MICE can go wrong

#### What went wrong in our previous examples?

When incomplete variables have **non-linear associations** with the outcome, or with each other, the requirement(s) of **compatibility and/or congeniality are violated**.

**Omission, or inadequate inclusion, of the outcome** may result in **MNAR** missing mechanisms. The same is the case when other relevant predictor variables are not used as predictor variables in the imputation.

Furthermore, **omission of variables** may lead to **mis-specified models**, however, models may also be mis-specified when all relevant covariates are included, but **distributional assumptions** or the specified **form of associations** are incorrect.



# 14. Alternatives to MICE

## 14.1. Joint model imputation

To **avoid incompatible and uncongenial imputation models**, we need to

- specify the joint distribution
- and derive full conditionals / imputation models from this joint distribution

instead of specifying them directly.

## 14. Alternatives to MICE

### 14.1. Joint model imputation

To **avoid incompatible and uncongenial imputation models**, we need to

- specify the joint distribution
- and derive full conditionals / imputation models from this joint distribution

instead of specifying them directly.

#### **Problem:**

Especially in settings with several **variables of mixed type**, the joint distribution is usually not of any known form:

$$\begin{matrix} x_1 \sim N(\mu_1, \sigma_1^2) \\ x_2 \sim N(\mu_2, \sigma_2^2) \end{matrix} \Rightarrow \begin{pmatrix} x_1 \\ x_2 \end{pmatrix} \sim N \left( \begin{bmatrix} \mu_1 \\ \mu_2 \end{bmatrix}, \begin{bmatrix} \sigma_1^2 & \sigma_{12} \\ \sigma_{12} & \sigma_2^2 \end{bmatrix} \right)$$

**but** 
$$\begin{matrix} x_1 \sim N(\mu_1, \sigma_1^2) \\ x_2 \sim \text{Bin}(\mu_2) \end{matrix} \Rightarrow \begin{pmatrix} x_1 \\ x_2 \end{pmatrix} \sim ???$$

## 14. Alternatives to MICE

### 14.1. Joint model imputation

#### **Approach 1: Multivariate Normal Model**

Approximate the joint distribution by a known multivariate distribution (usually the normal distribution; this is the approach mentioned in Part I on slide 15)

#### **Approach 2: Sequential Factorization**

Factorize the joint distribution into a (sequence of) conditional and a marginal distributions

# 14. Alternatives to MICE

## 14.2. Multivariate Normal Model

### Assumption:

The outcome and incomplete variables follow a **joint multivariate normal distribution**, conditional on the completely observed covariates  $\mathbf{X}_c$ , parameters  $\boldsymbol{\theta}$  and, possibly, random effects,  $\mathbf{b}$ :

$$p(\mathbf{y}, \mathbf{x}_1, \dots, \mathbf{x}_p \mid \mathbf{X}_c, \boldsymbol{\theta}, \mathbf{b}) \sim N(\boldsymbol{\mu}, \boldsymbol{\Sigma})$$

# 14. Alternatives to MICE

## 14.2. Multivariate Normal Model

### Assumption:

The outcome and incomplete variables follow a **joint multivariate normal distribution**, conditional on the completely observed covariates  $\mathbf{X}_c$ , parameters  $\theta$  and, possibly, random effects,  $\mathbf{b}$ :

$$p(\mathbf{y}, \mathbf{x}_1, \dots, \mathbf{x}_p \mid \mathbf{X}_c, \theta, \mathbf{b}) \sim N(\mu, \Sigma)$$

### How do we get that multivariate normal distribution?

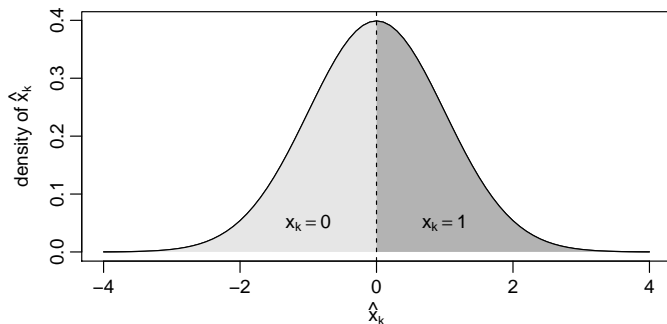
1. Assume **all** incomplete variables and the outcome are **(latent) normal**.
2. Specify linear (mixed) **models based on observed covariates**.
3. **Connect** using multivariate normal for **random effects & error terms**.

# 14. Alternatives to MICE

## 14.2. Multivariate Normal Model

### 1. Latent normal assumption:

e.g.:  $\mathbf{x}_k$  binary  $\rightarrow$  latent  $\hat{\mathbf{x}}_k$  is standard normal: 
$$\begin{cases} \mathbf{x}_k = 1 & \text{if } \hat{\mathbf{x}}_k \geq 0 \\ \mathbf{x}_k = 0 & \text{if } \hat{\mathbf{x}}_k < 0 \end{cases}$$



# 14. Alternatives to MICE

## 14.2. Multivariate Normal Model

### 2. Specify models:

$$\mathbf{y} = \mathbf{X}_c \boldsymbol{\beta}_y + \mathbf{Z}_y \mathbf{b}_y + \boldsymbol{\varepsilon}_y$$

$$\mathbf{w} = \mathbf{X}_c \boldsymbol{\beta}_w + \mathbf{Z}_w \mathbf{b}_w + \boldsymbol{\varepsilon}_w$$

$$\begin{array}{lcl} \hat{x}_1 & = & \mathbf{X}_c \boldsymbol{\beta}_{x_1} + \boldsymbol{\varepsilon}_{x_1} \\ & \vdots & \vdots \end{array}$$

$$\hat{x}_p = \mathbf{X}_c \boldsymbol{\beta}_{x_p} + \boldsymbol{\varepsilon}_{x_p}$$

# 14. Alternatives to MICE

## 14.2. Multivariate Normal Model

### 2. Specify models / 3. Connect random effects & error terms:

$$\begin{aligned} y &= \mathbf{X}_c \beta_y + \mathbf{Z}_y \mathbf{b}_y + \epsilon_y \\ w &= \mathbf{X}_c \beta_w + \mathbf{Z}_w \mathbf{b}_w + \epsilon_w \\ \hat{x}_1 &= \mathbf{X}_c \beta_{x_1} + \epsilon_{x_1} \\ &\vdots \\ \hat{x}_p &= \mathbf{X}_c \beta_{x_p} + \epsilon_{x_p} \end{aligned}$$

multivariate normal (optional, but suggested)

multivariate normal



# 14. Alternatives to MICE

## 14.2. Multivariate Normal Model

### Advantages:

- Easy to specify
- Relatively easy to implement
- Relatively easy to sample from
- Works for longitudinal outcomes

### Disadvantages:

- Assumes linear associations

Imputation with **non-linear associations** or **survival data** is possible with **extensions** of the multivariate normal approach.

# 14. Alternatives to MICE

## 14.3. Sequential Factorization

The **joint distribution** of two variables  $y$  and  $x$  can be written as the product of a conditional and a marginal distribution:

$$p(y, x) = p(y \mid x) p(x)$$

(or alternatively  $p(y, x) = p(x \mid y) p(y)$ )

## 14. Alternatives to MICE

### 14.3. Sequential Factorization

The **joint distribution** of two variables  $y$  and  $x$  can be written as the product of a conditional and a marginal distribution:

$$p(y, x) = p(y \mid x) p(x)$$

(or alternatively  $p(y, x) = p(x \mid y) p(y)$ )

This can easily be **extended for more variables**:

$$p(y, x_1, \dots, x_p, X_c) = \underbrace{p(y \mid x_1, \dots, x_p, X_c)}_{\text{analysis model}} p(x_1 \mid x_2, \dots, x_p, X_c) \dots p(x_p \mid X_c)$$

where  $x_1, \dots, x_p$  denote incomplete covariates and  $X_c$  contains all completely observed covariates.

# 14. Alternatives to MICE

## 14.3. Sequential Factorization

That the analysis model is part of the specification of the joint distribution has several advantages:

- The outcome is **automatically included in the imputation** procedure.
- The outcome does not appear in any of the predictors of the imputation models:
  - **no need to approximate** complex outcomes,
  - **no need to summarize** complex outcomes.
- The parameters of interest are obtained directly
  - ➔ imputation and analysis in one step
- **Non-linear associations** or interactions involving incomplete covariates are specified in the analysis model and thereby **automatically taken into account**

Since the joint distribution usually does not have a known form, Gibbs sampling is used to estimate parameters and sample imputed values.

# 14. Alternatives to MICE

## 14.3. Sequential Factorization

### Advantages:

- flexible with regards to outcome type
- univariate conditional distributions of incomplete covariates can be chosen according to type of variable
- non-linear associations and interactions can be taken into account
- assures congeniality and compatible imputation models

### Disadvantages:

- separate models need to be specified per incomplete variable: takes more time and consideration
- the joint distribution is of unknown form and sampling may be more computationally intensive

## 15. Imputation with non-linear functional forms

In the following we will not only consider the R package **mice**, but also three additional packages, **JointAI**, **smcfcs** and **jomo**, that provide alternatives to **mice**.

These three packages use Bayesian methodology to impute values, but once imputed datasets are obtained, standard complete data methods can be used.

**jomo** and **smcfcs** perform multiple imputation and create imputed datasets that can then be analysed the same way data imputed by **mice** would be analysed.

**JointAI** works fully Bayesian and performs the analysis and imputation simultaneously, so that the results from the analysis model of interest are obtained directly.

# 15. Imputation with non-linear functional forms

## 15.1. R package mice

There is no strategy for MICE that can guarantee valid imputations when non-linear functional forms and/or interactions are involved, but some settings in **mice** may help to reduce bias in the resulting estimates.

For imputation of variables that have non-linear associations

- `pmm` often works better than `norm`,
- Just Another Variable approach can reduce bias in interactions,
- `quadratic` can help to impute variables with quadratic association.

## 15. Imputation with non-linear functional forms

### 15.1. R package mice

In the **Just Another Variable (JAV)** approach the non-linear form (or interaction term) is calculated in the incomplete data, added as a column to the dataset and imputed as if it was just another variable.

`quadratic` provides imputation of covariates that have a quadratic association with the outcome, using the “polynomial combination” method.[17, pp. 139–141], [19].

This is to ensure the imputed values for  $x$  and  $x^2$  are consistent, and to reduce bias in the subsequent analysis that uses  $x$  and  $x^2$ .

In my experience, using `quadratic` can lead to numerical problems.



# 15. Imputation with non-linear functional forms

## 15.1. R package mice

To demonstrate the approaches, we use a simulated example dataset `DFnonlin`, with

- continuous outcome  $y$
- continuous (normal) covariate  $x$  (50% missing values MCAR)
- quadratic effect of  $x$  on  $y$
- binary covariate  $z$  (complete)
- interaction between  $x$  and  $z$

# 15. Imputation with non-linear functional forms

## 15.1. R package mice

To demonstrate the approaches, we use a simulated example dataset `DFnonlin`, with

- continuous outcome  $y$
- continuous (normal) covariate  $x$  (50% missing values MCAR)
- quadratic effect of  $x$  on  $y$
- binary covariate  $z$  (complete)
- interaction between  $x$  and  $z$

In the naive approach, we leave all settings to the defaults.

```
# naive imputation, using only y, x, z  
impnaive <- mice(DF_nonlin, printFlag = F)
```

# 15. Imputation with non-linear functional forms

## 15.1. R package mice

We use two different JAV approaches:

**JAV:** calculating the quadratic and interaction term before imputation

```
# add quadratic term and interaction to data
DF2 <- DF_nonlin
DF2$xx <- DF2$x^2
DF2$xz <- DF2$x * DF2$z

# JAV imputation
impJAV <- mice(DF2, printFlag = F, maxit = 20)
```

**JAV2:** additionally using an interaction between z and y

```
# add interaction between y and z to data
DF3 <- DF2
DF3$yz <- DF3$y * DF3$z

# JAV imputation with additional interaction
impJAV2 <- mice(DF3, printFlag = F, maxit = 20)
```

# 15. Imputation with non-linear functional forms

## 15.1. R package mice

We also try using imputation method `quadratic`.

```
# adapt the imputation method for quadratic imputation
methqdr <- impJAV$meth
methqdr[c("x", "xx", "xz")] <- c("quadratic", "~I(x^2)", "~I(x*z)")

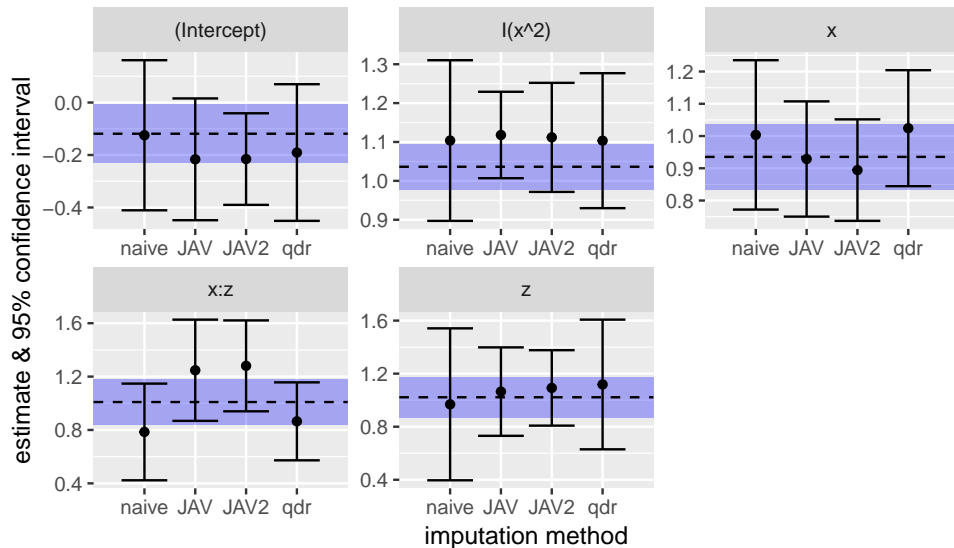
# adapt the predictor matrix
predqdr <- impJAV$pred
predqdr[, "xx"] <- 0

impqdr <- mice(DF2, meth = methqdr, pred = predqdr,
               printFlag = F, maxit = 10)
```

Note: there were warning messages about numerical issues for this approach.

# 15. Imputation with non-linear functional forms

## 15.1. R package mice



For this example, **none of the approaches provided satisfying results.**

# 15. Imputation with non-linear functional forms

## 15.2. R package JointAI

The package **JointAI** uses the **sequential factorization approach** to perform simultaneous analysis and imputation.[4, 3]

**JointAI** (version 0.1.0) can handle

- linear regression
- generalized linear regression
- linear mixed models

while assuring compatibility between analysis model and imputation models when non-linear functions or interactions are included.

The necessary Gibbs sampling is performed using JAGS (an external program), which is free, but needs to be installed from

<https://sourceforge.net/projects/mcmc-jags/files/>.

# 15. Imputation with non-linear functional forms

## 15.2. R package JointAI

**JointAI** can be installed from CRAN

```
install.packages("JointAI")
```

The development version (containing bug fixes and other improvements) can be installed from **GitHub**

```
install.packages("devtools")  
devtools::install_github(repo = "JointAI", username = "NErler")
```

A detailed explanation of the functionality is given in the help files of the package, and a vignette with an in-depth example analysis will be available soon.

# 15. Imputation with non-linear functional forms

## 15.2. R package JointAI

The syntax we use to analyse and impute the current example using **JointAI** is similar to the specification of a standard linear model using `lm()`.

```
library(JointAI)
JointAI_nonlin <- lm_imp(y ~ x*z + I(x^2), data = DF_nonlin, n.iter = 2500)
```

Convergence of the Gibbs sampler can be checked using a traceplot.

```
traceplot(JointAI_nonlin)
```

Results (no separate analysis & pooling is necessary) can be obtained with the `summary()` function:

```
res_JointAI_nonlin <- summary(JointAI_nonlin)
```



# 15. Imputation with non-linear functional forms

## 15.3. R package smcfcs

The package **smcfcs** performs multiple imputation using *substantive model compatible fully conditional specification*, a **hybrid approach between FCS and sequential factorization**.<sup>[1]</sup>

**smcfcs** (version 1.3.0) can handle

- linear regression,
- logistic regression,
- poisson regression,
- Cox proportional hazard models, and
- competing risk survival models,

while ensuring compatibility between analysis model and imputation models.

For more information see the help files and the [vignette](#).

# 15. Imputation with non-linear functional forms

## 15.3. R package smcfcs

The syntax to impute the data in the current example using the package **smcfcs** is:

```
library(smcfcs)
smcfcs_nonlin <- smcfcs(originaldata = DF_nonlin, smtype = "lm",
                        smformula = "y~x*z + I(x^2)",
                        method = c("", "norm", ""),
                        rjlimit = 3000, numit = 20)
```

The convergence of the procedure should be checked, for example with the following syntax:

```
par(mfrow = c(2,3), mar = c(2, 2, 0.5, 0.5), mgp = c(2, 0.6, 0))
for(i in 1:dim(smcfcs_nonlin$smCoefIter)[2]) {
  matplot(t(smcfcs_nonlin$smCoefIter[, i, ]), type = 'l', ylab = '')
}
```

## 15. Imputation with non-linear functional forms

### 15.3. R package smcfcs

To be able to use the convenient pooling function from the **mice** package we first need to convert the imputed data (which is a list) to a `mids` object.

This can be done with the function `datalist2mids()` from the **miceadds** package.

```
library(miceadds)
impobj_smcfcs_nonlin <- datalist2mids(smcfcs_nonlin$impDatasets)
```

The `mids` object can then be pooled and summarized as we have seen before with `mids` objects created by `mice()`.

```
models_smcfcs_nonlin <- with(impobj_smcfcs_nonlin, lm(y ~ x*z + I(x^2)))
res_smcfcs_nonlin <- summary(pool(models_smcfcs_nonlin))
```

# 15. Imputation with non-linear functional forms

## 15.4. R package jomo

The package **jomo** performs **joint model imputation** using the multivariate normal approach, with **extensions to assure compatibility** between analysis and imputation models.[2]

**jomo** (version 2.6-1) can handle

- linear regression,
- generalized linear regression,
- linear mixed models,
- generalized linear mixed models, and
- Cox proportional hazards models.

# 15. Imputation with non-linear functional forms

## 15.4. R package jomo

Using **jomo** we can impute the data in the current example as follows:

```
library(jomo)
jomo_nonlin <- jomo.lm(y ~ x*z + I(x^2), data = DF_nonlin)
```

To check the convergence of the model, the corresponding function with ending **.MCMCchain()** as to be used.

```
jomo_nonlinMCMC <- jomo.lm.MCMCchain(y ~ x*z + I(x^2), data = DF_nonlin)

par(mfcol = c(2, 3), mar = c(3, 2.5, 0.5, 0.5), mgp = c(2, 0.6, 0))
apply(jomo_nonlinMCMC$collectbeta[1, ], 1, plot, type = "l",
      xlab = 'iteration', ylab = '')
for (k in 1:dim(jomo_nonlinMCMC$collectomega)[1]) {
  apply(jomo_nonlinMCMC$collectomega[k, ], 1, plot, type = "l",
        xlab = 'iteration', ylab = '')
}

apply(jomo_nonlinMCMC$collectbetaY[1, ], 1, plot, type = "l",
      xlab = 'iteration', ylab = '')
plot(jomo_nonlinMCMC$collectvarY, type = 'l')
```

## 15. Imputation with non-linear functional forms

### 15.4. R package jomo

Again, we need to convert the output to a `mids` object using `datalist2mids()`. However, `jomo.lm()` returns a data frame, in which the original data and all imputed datasets are stacked onto each other.

`split()` splits the dataset by imputation number into a list of datasets, from which we need to exclude the first element (the original/incomplete data).

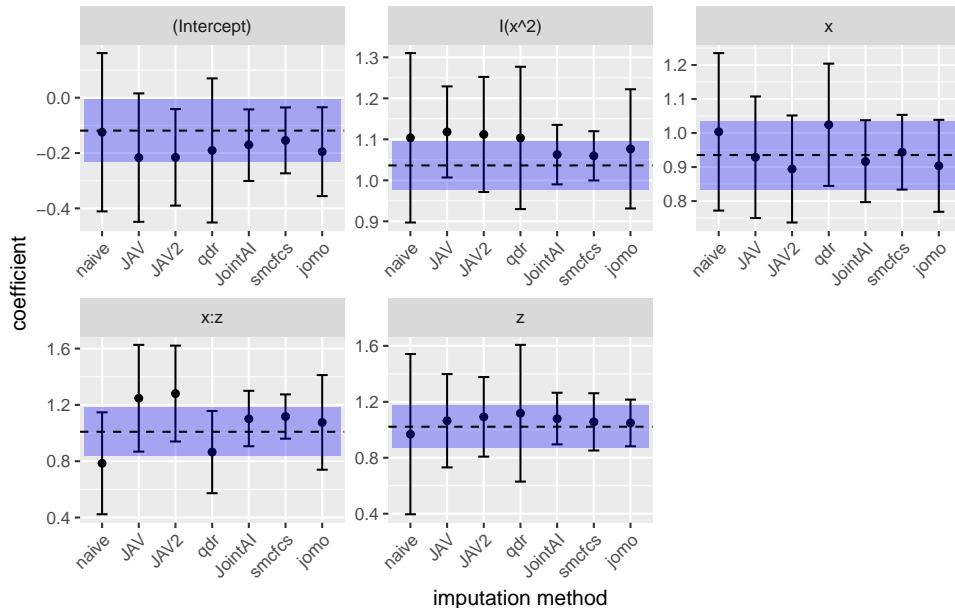
```
impobj_jomo_nonlin <- datalist2mids(split(jomo_nonlin,  
                                         jomo_nonlin$Imputation)[-1])
```

With the resulting `mids` object, analysis of the imputed data and pooling of the results works as in the above examples.

```
models_jomo_nonlin <- with(impobj_jomo_nonlin, lm(y ~ x*z + I(x^2)))  
res_jomo_nonlin <- summary(pool(models_jomo_nonlin))
```

# 15. Imputation with non-linear functional forms

## 15.5. Comparison of results



# 16. Imputation of longitudinal data

## 16.1. R package mice

**mice** has functions to allow imputation of longitudinal (2-level) data.

- **Level 1:**  
repeated measurements (within subjects) or subjects (within classes)
- **Level 2:**  
time-constant/baseline covariates, between subjects effects, variables on the group level

Imputation methods for **level-1** variables:

- `2l.pan`
- `2l.norm`
- `2l.lmer`

Imputation methods for **level-2** variables:

- `2lonly.norm`
- `2lonly.pmm`
- `2lonly.mean`



## 16. Imputation of longitudinal data

### 16.1. R package mice

`2l.pan` uses a linear two-level model with **homogeneous within group variances** using Gibbs sampling [14]. It needs the package **pan** to be installed.

`2l.pan` allows for different roles of predictor variables, that can be specified as different values in the `predictorMatrix`:

- grouping/ID variable: -2
- random effects (also included as fixed effects): 2
- fixed effects of group means: 3
- fixed effects of group means & random effects: 4

```
# random effects of x in model for y  
pred["y","x"] <- 2  
# fixed effects of x and group mean of x  
pred["y","x"] <- 3  
# random effects of x and group mean of x  
pred["y","x"] <- 4
```

## 16. Imputation of longitudinal data

### 16.1. R package mice

`2l.norm` implements a (Bayesian) linear two-level model with **heterogenous** group variances.

In the current implementation all predictors should be specified as random effects (set to 2 in the `predictorMatrix`, because the algorithm does not handle predictors that are specified as fixed effects).

## 16. Imputation of longitudinal data

### 16.1. R package mice

`2l.norm` implements a (Bayesian) linear two-level model with **heterogenous** group variances.

In the current implementation all predictors should be specified as random effects (set to 2 in the `predictorMatrix`, because the algorithm does not handle predictors that are specified as fixed effects).

`2l.lmer` imputes univariate systematically and sporadically missing data using a two-level normal model using `lmer()` from package **lme4** (developed in the context of individual patient meta analysis. [7, 6])

## 16. Imputation of longitudinal data

### 16.1. R package mice

`2l.norm` implements a (Bayesian) linear two-level model with **heterogenous** group variances.

In the current implementation all predictors should be specified as random effects (set to 2 in the `predictorMatrix`, because the algorithm does not handle predictors that are specified as fixed effects).

`2l.lmer` imputes univariate systematically and sporadically missing data using a two-level normal model using `lmer()` from package **lme4** (developed in the context of individual patient meta analysis. [7, 6])

`2lonly.norm` and `2lonly.pmm` can be used to impute level-2 variables (in combination with `2l.pan` for level-1 variables).

In all case, the group identifier ("id" variable") needs to be set to -2 in the `predictorMatrix`.

## 16. Imputation of longitudinal data

### 16.1. R package mice

`2lonly.mean` imputes values with the mean of the observed values per class. This method should only be used to fill in values that are known to be constant per class and have some values observed in each class.

**Example:** In a multi-center trial the type of some medical equipment is known to be the same for all patients treated in the same hospital, but not filled in for some patients.

## 16. Imputation of longitudinal data

### 16.1. R package mice

As an example, we will impute the second (unbalanced) longitudinal data example from above. The data contain

- $x_1$  (complete)
- $x_2$  (binary, 30% missing values)
- $x_3$  (3 categories, 30% missing values)
- $x_4$  (continuous/normal, 30% missing values)
- $y$  (longitudinal outcome)
- *time* (time variable with quadratic effect)
- *id* (id variable)

Since there is no 2-level method for categorical data, we use `2lonly.pmm` to impute  $x_2$  and  $x_3$ .

# 16. Imputation of longitudinal data

## 16.1. R package mice

As usual, we start with the setup run of `mice()`

```
imp0 <- mice(DFexlong2, maxit = 0)
meth <- imp0$method
pred <- imp0$predictorMatrix
```

and adjust the imputation `method` and `predictorMatrix`

```
meth[c("x2", "x3")] <- "2lonly.pmm"
meth[c("x4")] <- "2lonly.norm"

pred[, "id"] <- -2 # identify id variable
pred[, "ti"] <- 0 # don't use time-point indicator
```

We can then perform the imputation.

```
imp <- mice(DFexlong2, maxit = 10, meth = meth, pred = pred, printFlag = F)
```

## 16. Imputation of longitudinal data

### 16.1. R package mice

The imputed data can be analysed using either `lmer()` from the package **lme4**, or `lme()` from **nlme**. Here we use the former.

```
library(lme4)
models <- with(imp, lmer(y ~ x1 + x2 + x3 + x4 + time + I(time^2) +
                        (time|id)))
mice_longimp <- summary(pool(models))
```



## 16. Imputation of longitudinal data

### 16.1. R package mice

Currently, there is only limited documentation and examples available that show how to use these functions in **mice**.

Technical details can be obtained from the methodological references given in the help files of the R functions.

A [vignette](#) on multi-level imputation with **mice** is available. It gives a more elaborate example of how to analyse such data.

## 16. Imputation of longitudinal data

### 16.2. R package JointAI

Linear mixed models with incomplete covariates can also be analysed using the package **JointAI**.

The syntax is analogous the syntax used in `lme()` of the package **nlme**.

```
library(JointAI)
JointAI_long <- lme_imp(y ~ x1 + x3 + x2 + x4 + time + I(time^2),
                      random = ~time|id, data = DFexlong2,
                      n.iter = 5000)
```

Again, convergence of the Gibbs sampler should be checked using a traceplot,

```
traceplot(JointAI_long)
```

before obtaining the results:

```
res_JointAI_long <- summary(JointAI_long)
```

Contrary to the two-level imputation of **mice**, non-linear associations are appropriately handled.

## 16. Imputation of longitudinal data

### 16.3. R package jomo

In **jomo**, the functions `jomo.lmer()` and `jomo.glmer()` can be used to impute longitudinal data with normal or non-normal outcomes.

In the multi-level setting, the level of each variable needs to be specified (1: repeated measurements, 2: baseline covariates), and ordered the same way the variables occur in the data frame.

```
library(jomo)
# specify the level of each variable
lvl <- c("id" = 1, y = 1, x1 = 2, x2 = 2, x3 = 2, x4 = 2, time = 1)

jomo_long <- jomo.lmer(formula = y ~ x1 + x2 + x3 + x4 +
                        time + I(time^2) + (time|id),
                        data = DFexlong2[, names(lvl)], level = lvl)
```

Like in the example with non-linear effects, convergence of the imputation needs to be checked.

## 16. Imputation of longitudinal data

### 16.3. R package jomo

Again, the stacked dataframe returned by `jomo.lmer()` needs to be split by imputation number and the original data excluded, before fitting the model and pooling the results.

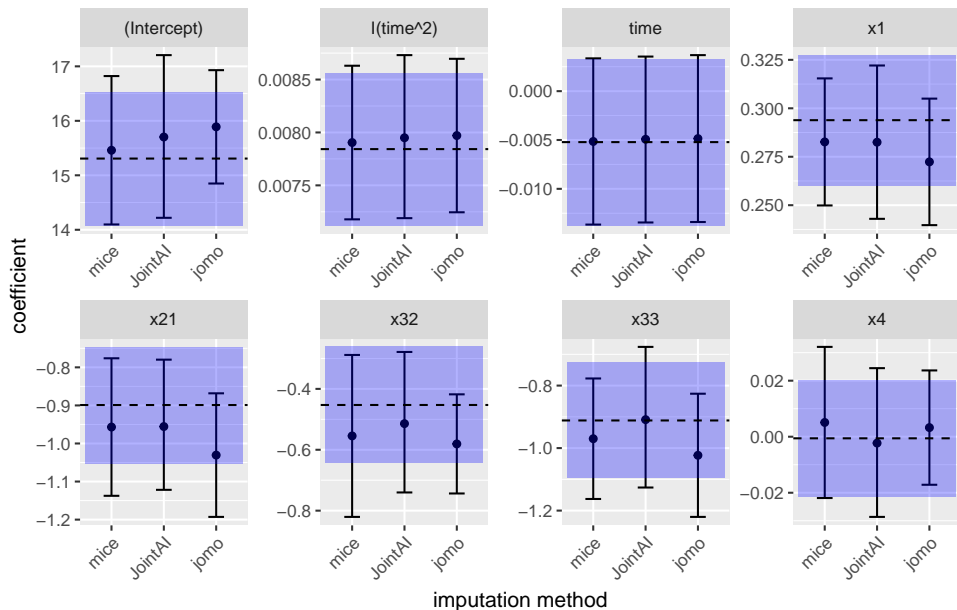
```
library(miceadds)
impobj_jomo_long <- datalist2mids(split(jomo_long,
                                       jomo_long$Imputation)[-1])
models_jomo_long <- with(impobj_jomo_long,
                        lmer(y ~ x1 + x3 + x2 + x4 + time + I(time^2) +
                            (time|clus)))
res_jomo_long <- summary(pool(models_jomo_long))
```

(Note: `jomo.lmer()` re-names the grouping variable to `clus`).

As in the examples for non-linear functional forms, congeniality of imputation models is maintained.

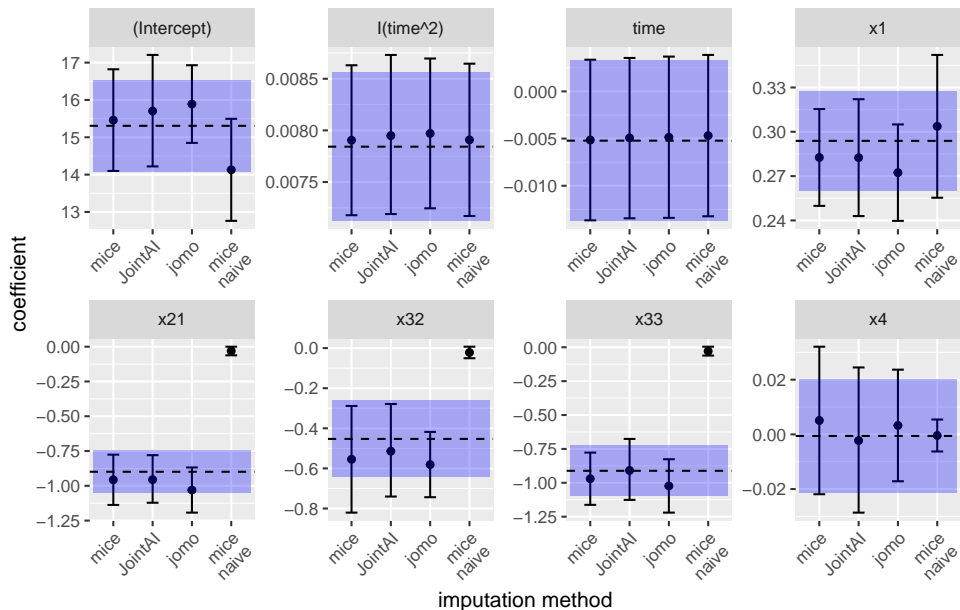
# 16. Imputation of longitudinal data

## 16.4. Comparison of results



# 16. Imputation of longitudinal data

## 16.4. Comparison of results



## 17. Imputation of survival data

### 17.1. Results from literature

On slide 145 we have seen the rather complex formula for imputation of an incomplete covariate in survival data.

White et al. [20] derived versions of this model for different settings (binary or continuous incomplete covariate  $X$ , and continuous, categorical or no complete covariate  $Z$ ) and investigated how to best approximate it.

They found that when **covariate effects and cumulative incidences are rather small**, using  $Z$ ,  $D$  and  $H_0(T)$ , and possibly an interaction term, as predictor variables in the imputation for  $X$  in MICE may work satisfactorily.

However, in practice  $H_0(T)$  is unspecified.

# 17. Imputation of survival data

## 17.1. Results from literature

Two main ideas:

- If covariate effects  $\beta_x$  and  $\beta_z$  are small,  $H_0(t) \approx H(t)$ , which can be approximated by the **Nelson-Aalen estimator**.
- **Estimate  $H_0(T)$  in an additional step** inside the MICE procedure by fitting a Cox model on the imputed data.

Neither of these approaches takes into account uncertainty about  $H_0(t)$  (but the impact is likely to be small).



## 17. Imputation of survival data

### 17.1. Results from literature

Two main ideas:

- If covariate effects  $\beta_x$  and  $\beta_z$  are small,  $H_0(t) \approx H(t)$ , which can be approximated by the **Nelson-Aalen estimator**.
- **Estimate  $H_0(T)$  in an additional step** inside the MICE procedure by fitting a Cox model on the imputed data.

Neither of these approaches takes into account uncertainty about  $H_0(t)$  (but the impact is likely to be small).

Based on results from their simulation study, White et al. conclude that **using  $Z$ ,  $D$  and the Nelson-Aalen estimator  $\hat{H}(T)$**  as predictors for the imputation of  $X$  worked best.

However, some **bias towards the null** should be expected when covariates have large effects.

# 17. Imputation of survival data

## 17.2. R package mice

In **mice**, `nelsonaalen()` can be used to **calculate the Nelson-Aalen estimator**, to use it as covariate in the imputation.

```
survdat$H0 <- nelsonaalen(survdat, timevar = Time, statusvar = event)
```

Then, we can prepare the imputation using the same steps as in previous examples:

```
# setup run
imp0 <- mice(survdat, maxit = 0)
meth <- imp0$method
pred <- imp0$predictorMatrix

# specify normal imputation for continuous covariates
meth[c("x1", "x3")] <- "norm"

# remove event time from predictor (high correlation with H0)
pred[, "Time"] <- 0
```

# 17. Imputation of survival data

## 17.2. R package mice

With the modified arguments `method` and `predictorMatrix` we run the imputation:

```
survimp <- mice(survdat, maxit = 10, method = meth, predictorMatrix = pred,  
               printFlag = F)
```

To obtain the pooled results, we first fit the model of interest

```
cox_mice <- with(survimp, coxph(Surv(Time, event) ~ x1 + x2 + x3))
```

and pool and summarize the results.

```
res_mice_surv <- summary(pool(cox_mice))  
  
## Warning in mice.df(m, lambda, dfcom, method): Large sample assumed.
```

The warning message refers to the way the degrees of freedom for the formulas we saw in part I (slide 32) are calculated and can be ignored.

# 17. Imputation of survival data

## 17.3. R package smcfcs

Using the package **smcfcs**, the same data can be imputed with the following syntax:

```
library(smcfcs)
smcfcs_surv <- smcfcs(originaldata = survdat, smtype = "coxph",
                      smformula = "Surv(Time, event) ~ x1 + x2 + x3",
                      method = c("", "", "logreg", "norm", "norm", ""),
                      numit = 20, rjlimit = 1500)
```

Convergence of the procedure should be checked, analogously to the previous example (see slide 172).

After the resulting object is converted to a `mids` object, fitting the model and pooling the results is identical to what was done with the data imputed by **mice**.

```
impobj_smcfcs_surv <- datalist2mids(smcfcs_surv$impDatasets)
models_smcfcs_surv <- with(impobj_smcfcs_surv,
                          coxph(Surv(Time, event) ~ x1 + x2 + x3))
res_smcfcs_surv <- summary(pool(models_smcfcs_surv))
```

# 17. Imputation of survival data

## 17.4. R package jomo

In the package **jomo**, the function `jomo.coxph()` can be used to impute our example survival data:

```
library(jomo)
jomo_surv <- jomo.coxph(formula = Surv(Time, event) ~ x1 + x2 + x3,
                        data = survdat)
```

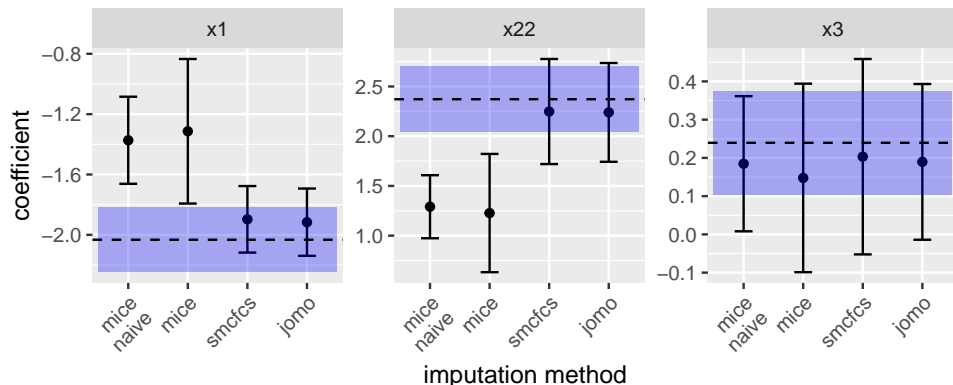
Note that the convergence of the procedure should be checked using `jomo.coxph.MCMCchain()` (see the previous examples using **jomo**).

To analyse & pool the imputed data the steps are identical to the other examples:

```
impobj_jomo_surv <- datalist2mids(split(jomo_surv, jomo_surv$Imputation)[-1])
models_jomo_surv <- with(impobj_jomo_surv,
                        coxph(Surv(Time, event) ~ x1 + x2 + x3))
res_jomo_surv <- summary(pool(models_jomo_surv))
```

# 17. Imputation of survival data

## 17.5. Comparison of results



The naive mice approach, and mice using the Nelson-Aalen estimator give very biased results for the effects of x1 and x2, but performed acceptably well for x3.

Note that the **true effects** (log HR) of x1 and x2 are **very large** (-2 and 2.5, respectively), and represent the setting where the approximation by the Nelson-Aalen estimate is **expected to be biased**.

## Summary & Conclusion of Part III

- MICE requires congenial & compatible imputation models to work well.
- When this is not the case, (naive) use of MICE can lead to biased results.
- Common settings that require special attention are
  - non-linear functional forms & interaction terms
  - longitudinal data
  - survival data
- When using the package **mice**, there are choices that can reduce bias
  - `pmm` tends to be less biased than `norm` for interactions or non-linear associations
  - JAV approach reduces bias in settings with interactions or non-linear associations
  - special 2-level imputation methods are available for longitudinal data
  - The Nelson-Aalen estimator can be used instead of the time variable for imputing survival data when effects are not too large.

## Summary & Conclusion of Part III (cont.)

- Generally, problems are more severe when
  - proportions of missing values are large,
  - effect sizes are large,
  - little other covariate information is available.

(Note that in the examples we had all of the above.)

- In settings where MICE may not provide valid imputations, alternative approaches are available and should be considered.
- R packages that provide such alternative approaches are for example:
  - JointAI (non-linear & longitudinal)
  - smcfcs (non-linear & survival)
  - jomo (non-linear, longitudinal & survival)
- These packages are very young.
  - Hence, they may still have some problems.
    - ➔ Use them carefully! (and email the maintainer about problems)
  - They are under active development, so resolutions of bugs and features are frequently added.



## Part IV

# Multiple Imputation Strategies

# Outline of Part IV

## 18. Strategies for using MICE

- 18.1 Imputation methods

- 18.2 Tips & Tricks

- 18.3 Number of imputed datasets

- 18.4 What to do with large datasets?

- 18.5 How much missing is too much?

- 18.6 Imputation of outcomes

- 18.7 Notes of caution & things to keep in mind

## 19. MICE and MI in the bigger picture

- 19.1 Other R packages that do imputation

- 19.2 Imputation in other software

- 19.3 Other approaches to handle missing values

# 18. Strategies for using MICE

## 18.1. Imputation methods

We have focussed on a few imputation methods that cover the most common types of data but there are many more methods implemented.

Imputation methods implemented in the **mice** package:

---

<code>mice.impute.2l.lmer</code>	<code>mice.impute.logreg</code>	<code>mice.impute.passive</code>
<code>mice.impute.2l.norm</code>	<code>mice.impute.logreg.boot</code>	<code>mice.impute.pmm</code>
<code>mice.impute.2l.pan</code>	<code>mice.impute.mean</code>	<code>mice.impute.polr</code>
<code>mice.impute.2lonly.mean</code>	<code>mice.impute.midastouch</code>	<code>mice.impute.polyreg</code>
<code>mice.impute.2lonly.norm</code>	<code>mice.impute.norm</code>	<code>mice.impute.quadratic</code>
<code>mice.impute.2lonly.pmm</code>	<code>mice.impute.norm.boot</code>	<code>mice.impute.rf</code>
<code>mice.impute.cart</code>	<code>mice.impute.norm.nob</code>	<code>mice.impute.ri</code>
<code>mice.impute.lda</code>	<code>mice.impute.norm.predict</code>	<code>mice.impute.sample</code>

---

**Note:** Just because a method is implemented does not mean you need to / should use it.

# 18. Strategies for using MICE

## 18.1. Imputation methods

Imputation methods implemented in the **miceadds** package:

---

<code>mice.impute.2l.binary</code>	<code>mice.impute.bygroup</code>
<code>mice.impute.2l.contextual.norm</code>	<code>mice.impute.eap</code>
<code>mice.impute.2l.contextual.pmm</code>	<code>mice.impute.grouped</code>
<code>mice.impute.2l.continuous</code>	<code>mice.impute.hotDeck</code>
<code>mice.impute.2l.eap</code>	<code>mice.impute.ml.lmer</code>
<code>mice.impute.2l.groupmean</code>	<code>mice.impute.plausible.values</code>
<code>mice.impute.2l.groupmean.elim</code>	<code>mice.impute.pls</code>
<code>mice.impute.2l.latentgroupmean.mcmc</code>	<code>mice.impute.pmm3</code>
<code>mice.impute.2l.latentgroupmean.ml</code>	<code>mice.impute.pmm4</code>
<code>mice.impute.2l.plausible.values</code>	<code>mice.impute.pmm5</code>
<code>mice.impute.2l.pls</code>	<code>mice.impute.pmm6</code>
<code>mice.impute.2l.pls2</code>	<code>mice.impute.tricube.pmm</code>
<code>mice.impute.2l.pmm</code>	<code>mice.impute.tricube.pmm2</code>
<code>mice.impute.2lonly.function</code>	<code>mice.impute.weighted.norm</code>
<code>mice.impute.2lonly.norm2</code>	<code>mice.impute.weighted.pmm</code>
<code>mice.impute.2lonly.pmm2</code>	

---

# 18. Strategies for using MICE

## 18.2. Tips & Tricks

In complex settings, variables may need to be **re-calculated** or **re-coded** after imputation:

- Use `complete()` to convert the imputed data from a `mids` object to a `data.frame`.
- Perform the necessary calculations.
- Convert the changed `data.frame` back to a `mids` object using the functions from the flow-diagram in the second practical (e.g., `as.mids()`, `datalist2mids()`, `imputationList()`, ...)

Not just in imputation: Set a **seed value** to create reproducible results.

- in R: `set.seed()`
- in `mice()`: argument `seed`

## 18. Strategies for using MICE

### 18.3. Number of imputed datasets

**Early publications** on multiple imputation suggested that 3 – 5 imputations are sufficient and this is still a common assumption in practice.[12]

The reasoning behind using a small number of imputed datasets was that **storage of imputed data was “expensive”** (which is no longer the case) and a larger number of imputations would only have little advantage.[13]

More **recent work** from various authors [21, 17, 5] considers the efficiency of the pooled estimates, reproducibility of the results, statistical power of tests or the width of the resulting confidence intervals compared to the width of the true confidence intervals.

## 18. Strategies for using MICE

### 18.3. Number of imputed datasets

A **suggested rule of thumb** is that the **number of imputed datasets** should be similar to the **percentage of incomplete cases**.<sup>[21]</sup> Since this percentage depends on the size of the dataset, the **average percentage of missing values** per variable could be used as an alternative.<sup>[17]</sup>

Generally, using **more imputed datasets should be preferred**, especially in settings where the computational burden allows for it. Even though results are unlikely to change with a larger number of imputations, it can increase the efficiency and reproducibility of the results.

## 18. Strategies for using MICE

### 18.4. What to do with large datasets?

In imputation, generally the **advice is to include as much information as possible** in the imputation models. Using a large number of predictor variables makes the **MAR assumption more plausible** (and, hence, reduces bias due to MNAR missingness) and can **reduce uncertainty** about the missing values.

This can **work well in small or medium sized datasets** (20 – 30 separate variables, i.e. without interactions, variables derived from others, ...) however, **in large datasets** (contain hundreds or thousands of variables) this is **not feasible**.<sup>[17]</sup>



## 18. Strategies for using MICE

### 18.4. What to do with large datasets?

For large datasets a possible strategy is to

- Include all **variables used in the analysis model(s)** (including the outcome!).
- Include auxiliary variables if they are **strong predictors of missingness**.
- Include auxiliary variables if they have **strong associations with the incomplete variables**.
- Use **auxiliary variables only if they do not have too many missing values** themselves (and are observed for most of the incomplete cases of the variable of interest).
- Use **auxiliary variables** only in those imputation models for which they are **relevant** (and exclude them for others using the predictor matrix).
- Calculate **summary scores** from multiple items referring to the same concept and use the summary score as predictor variable.

## 18. Strategies for using MICE

### 18.5. How much missing is too much?

There is **no clear cut-off** for the proportion of missing values that can be handled adequately using MICE (or any other imputation method).

The amount of missingness that can be handled **depends on the information that is available** to impute it.

- Are there **strong predictor variables** available & observed?
- Are there **sufficient observed cases** to get reliable estimates for the predictive distribution?

## 18. Strategies for using MICE

### 18.5. How much missing is too much?

There is **no clear cut-off** for the proportion of missing values that can be handled adequately using MICE (or any other imputation method).

The amount of missingness that can be handled **depends on the information that is available** to impute it.

- Are there **strong predictor variables** available & observed?
- Are there **sufficient observed cases** to get reliable estimates for the predictive distribution?

#### Example:

- In a set of  $N = 50$  cases, 50% missing values leave 25 cases to estimate the parameters of the predictive distribution.
- In a large set of  $N = 5000$  subjects, 50% missing cases leave 2500 observed cases to estimate parameters.

# 18. Strategies for using MICE

## 18.6. Imputation of outcomes

Usually, **missing outcome values are not imputed**.

### Why?

When there are no auxiliary variables, imputation and analysis model are equal.

- Parameters of the imputation model are estimated on observed cases of the outcome.
- Imputed values will fit the assumed model perfectly.
- Including imputed cases in the analysis does not add any information.

### Exception:

- When very strong auxiliary variables are available.
- Outcomes may be imputed when one imputation is performed for several analysis models, because not imputing the outcome(s) would mean
  - excluding cases with missing outcome(s) from the imputation, or
  - excluding the outcome variable(s) as predictor(s).

# 18. Strategies for using MICE

## 18.7. Notes of caution & things to keep in mind

Multiple imputation is not a quick and easy solution for missing data. It requires care and knowledge about

- the data to be imputed (and the context of the data),
- the statistical method used for imputation, and
- the software implementation used.

Moreover

- Never accept default settings of software blindly.
- Question the plausibility of the MAR assumption. If it is doubtful, use sensitivity analysis.

# 18. Strategies for using MICE

## 18.7. Notes of caution & things to keep in mind

### Remember:

- Use as much information as possible
  - include all covariates **and the outcome**
  - use auxiliary information
  - use the most detailed version of variables if possible
- Avoid feedback from derived variables to their originals.
- **Imputation models must fit the data**  
(correct assumption of error distribution and functional forms and possible interactions of predictor variables).
- Think carefully how to handle variables that are derived from other variables.
- Consider the impact the visit sequence may have.
- Choose an appropriate number of imputations.
- Make sure the imputation algorithm has converged.
- Use common sense when evaluating if the imputed values are plausible.

## 19. MICE and MI in the bigger picture

### 19.1. Other R packages that do imputation

In Part III of this course (and the second practical) we have worked with some R packages that perform imputation or provide functionality for missing data other than **mice**.

Currently, there are **218 packages** available on CRAN that **use the word “missing”** in either the title or description of the package, **132** that **use either “impute” or “imputation”** and **47** that **use the word “incomplete”**.

Not all of these packages perform imputation or are useful for our purposes, but even if we excluded those packages, the number of useful packages for dealing with missing data would still be too large to mention them all.

➡ **The mice package is often a good option, but certainly not the only option to perform imputation!**

# 19. MICE and MI in the bigger picture

## 19.2. Imputation in other software

In this second half of the course, we have focused on (multiple) imputation using R.

Naturally, R is not the only statistical software that can perform multiple imputation.

- **Stata, SAS and MPLUS** provide packages/functions to perform multiple imputation and pool the results.
- There are macros and additional packages available, e.g.e, **smcfcs** is implemented for **Stata** as well
- **SPSS** provides some functionality to perform MI



## 19. MICE and MI in the bigger picture

### 19.3. Other approaches to handle missing values

Finally, we should not forget that **MICE is not the only method to handle missing values**.

Besides MICE, **multiple imputation** can be performed in a **joint model approach** (as for instance implemented in the R package **jomo**).

Furthermore,

- **direct likelihood methods**,
- **fully Bayesian methods**(as implemented in **JointAI**), or
- **weighted estimating equations**

are valid alternative approaches when data are incomplete.

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# Erasmus MC

University Medical Center Rotterdam



n.erler@erasmusmc.nl



N\_Erler



NErler

Dep. Biostatistics: [www.erasmusmc.nl/biostatistiek](http://www.erasmusmc.nl/biostatistiek)