

ICE PBPK workflow

Description

The workflow allows the flexibility to select from three different rat and/or human PK models from the EPA's httk package: solve_pbtok, solve_gas_pbtok, and solve_fetal_pbtok. The workflow is to predict plasma and tissue concentration profiles across time following a dose of a substance. Exposure routes include oral ingestion (solve_pbtok, solve_fetal_pbtok), intravenous (solve_pbtok, solve_fetal_pbtok), and inhalation (solve_gas_pbtok). For inhalation exposures using the dose is provided as an air concentration, uM or ppmv.

An example input file is included: ChemicalData_Rnotebook.txt

Load libraries

```
#load libraries
library(tidyverse)
library(rlang)
library(deSolve)
library(doParallel)
library(httk) #this is needed for models: solve_pbtok, solve_gas_pbtok, solve_fetal_pbtok.
               #The code is compatible with httk_2.2.2
library(openxlsx) #for writing the excel file that is user output
library(readxl)   #for read_excel() #2025
library(data.table)
```

Define custom functions

```
#function to parse time series list into second output table format.
#The input for this function is a list object (x) that contains an
#entry with the pbpk outputs from the httk functions along with the
#plasma CSS and CASRN for the chemical

## assemble.table_* function is to assemble direct outputs from different models to a format that is us
## Because the number of output columns are different for each model, so the assemble.table_* is modifi
#nOutParam - total number of time column and predicted tissue concentration column in the output table
#nStartCol - start column number, including time column
#nStartConCol - start column number of tissue concentration , excluding time column
#nTotalCol - number of total columns in the output table
#assemble.table <- function(x, nOutParam=10, nStartCol=4, nStartConCol=5, nTotalCol=13){
assemble.table_pbtok <- function(x, nOutParam=NULL, nStartCol=NULL, nStartConCol=NULL, nTotalCol=NULL){
  CASRN <- rep(x$CASRN[1], nOutParam) #x is "output" data frame; 9 rows, 8
  compound <- rep(chem.physical_and_invitro.data[which(
```

```

    chem.physical_and_invitro.data$CAS==x$CASRN[1]),1], nOutParam) #extract the "compound" information
    dtxsid <- rep(chem.physical_and_invitro.data[which(
      chem.physical_and_invitro.data$CAS==x$CASRN[1]),4], nOutParam) #extract the "DTXSID" information fo
    half_life_h <- rep(x$half_life_h[1], nOutParam)
    maxAUC.h <- rep(x$maxAUC.h[1], nOutParam)
    CSS_plasma <- rep(x$CSS_plasma[1], nOutParam)
    Cmax <- c(NA, sapply(as.data.frame(x[nStartConCol:nTotalCol]), function(y) max(as.numeric(y)))) #from
    Unit <- rep(ConcentrationUnit, nOutParam)
    compartment <- names(x)[nStartCol:nTotalCol] #nStartCol - start column number, in
    concs <- t(x[nStartCol:nTotalCol]) #nTotalCol - number of total columns

    out <- as.data.frame(cbind(CASRN, compound, dtxsid, half_life_h, maxAUC.h, CSS_plasma, Cmax, Unit, co
    rownames(out) <- NULL
    return(out)
  }

assemble.table_gas <- function(x, nOutParam=NULL, nStartCol=NULL, nStartConCol=NULL, nTotalCol=NULL){
  CASRN <- rep(x$CASRN[1], nOutParam) #x is "output" data frame; 9 rows, 8
  compound <- rep(chem.physical_and_invitro.data[which(
    chem.physical_and_invitro.data$CAS==x$CASRN[1]),1], nOutParam) #extract the "compound" information
  dtxsid <- rep(chem.physical_and_invitro.data[which(
    chem.physical_and_invitro.data$CAS==x$CASRN[1]),4], nOutParam) #extract the "DTXSID" information fo
  half_life_h <- rep(x$half_life_h[1], nOutParam)
  maxAUC.h <- rep(x$maxAUC.h[1], nOutParam)
  #CSS_plasma <- rep(x$CSS_plasma[1], nOutParam)
  Cmax <- c(NA, sapply(as.data.frame(x[nStartConCol:nTotalCol]), function(y) max(as.numeric(y)))) #from
  Unit <- rep(ConcentrationUnit, nOutParam)
  compartment <- names(x)[nStartCol:nTotalCol] #nStartCol - start column number, in
  concs <- t(x[nStartCol:nTotalCol]) #nTotalCol - number of total columns

  out <- as.data.frame(cbind(CASRN, compound, dtxsid, half_life_h, maxAUC.h, Cmax, Unit, compartment, co
  rownames(out) <- NULL
  return(out)
}

assemble.table_fetal <- function(x, nOutParam=NULL, nStartCol=NULL, nStartConCol=NULL, nTotalCol=NULL){
  CASRN <- rep(x$CASRN[1], nOutParam) #x is "output" data frame; 21 rows,
  compound <- rep(chem.physical_and_invitro.data[which(
    chem.physical_and_invitro.data$CAS==x$CASRN[1]),1], nOutParam) #extract the "compound" information
  dtxsid <- rep(chem.physical_and_invitro.data[which(
    chem.physical_and_invitro.data$CAS==x$CASRN[1]),4], nOutParam) #extract the "DTXSID" information fo
  half_life_h <- rep(x$half_life_h[1], nOutParam)
  maxAUC.h <- rep(x$maxAUC.h[1], nOutParam)
  maxFetalAUC.h <- rep(x$maxfAUC.h[1], nOutParam)
  Cmax <- c(NA, sapply(as.data.frame(x[nStartConCol:nTotalCol]), function(y) max(as.numeric(y)))) #from
  Unit <- rep(ConcentrationUnit, nOutParam)
  compartment <- names(x)[nStartCol:nTotalCol] #nStartCol - start column number, in
  concs <- t(x[nStartCol:nTotalCol]) #nTotalCol - number of total columns

  out <- as.data.frame(cbind(CASRN, compound, dtxsid, half_life_h, maxAUC.h, maxFetalAUC.h, Cmax, Unit,
  rownames(out) <- NULL
  return(out)
}

```

Input data and output file path

There are several input variables needed to run the code. Some variables are model specific, as detailed. Adjust the file paths to point to the file in your directory.

chemFile: The file containing chemical data from ICE. this has the CASRN as the first field

```
chemFile <- "ChemicalData_Rnotebook.txt"      # chemicals data from ICE, include CASRN field as identifier
#chemFile <- "TemplatePhysChemFile.txt"      # used for comparing results from running R scripts
Userout <- "User_Results.xlsx"                # file name (and path) for the output
                                              # file for the tissue concentrations
output_image <- "User_plot.pdf"               # the file for the plot of the results
```

Model variables

Details about what model, route, and dose need to be specified.

What model?

Currently, species in ICE is limited to human or rat. Using this notebook one can expand the species with minor editing of the code, provided the parameters are available.

```
species <- "human"                          # "human" or "rat", the solve_fetal_pbt model is only for human
modelType <- "solve_pbt"                    # "solve_pbt", "solve_fetal_pbt", or "solve_gas_pbt"
```

For the PBPK models some additional parameters can be modified. An inhalation exposure has additional parameters. The route determines where the chemical will enter the system. Exposure route, interval, and days are needed for all PBPK models.

```
route <- "oral"      # options are "inhalation", "iv", "oral"; "solve_gas_pbt" uses the inhalation (gas)
bWeight <- 70        # unit is kg; the body weight of simulated subject, default is 70 for human, and 0.25 for rat
interv <- 24         # dosing interval, hours
ndays <- 3          # number of days dosing is done
expDose <- 1         # current calculations assume 1mg/kg/dose for solve_pbt, solve_fetal_pbt model
expConc <- 1
ConcentrationUnit <- "uM" # options are uM and mg/L
```

Inhalation specific parameters

For an inhalation route of exposure, the “expDose” exposure route is currently not available. Instead, the “expConc” exposure route, which is more typical of a gas exposure, can be used. This models an inhaled concentration over a duration of time, set by “expLength”.

Note that “expDose” is currently not supported.

```
gasDosing <- "expConc" # "expConc" or "expDose"
#This is only needed if gasDosing == "expConc" bc otherwise default to expDose
if(gasDosing == "expConc" & (!exists("expConc")) || is.null(expConc)){
  expConc <- 1 #current calculations assume 1mg/kg/dose
}
gasInputUnit <- "ppmv" # Input concentration unit for gas model (httkv2.1.0 and later), "uM" or "ppmv"
expLength <- 0.25 #length of the gas exposure, hours
```

Gestation model (solve_fetal_pbtk) specific parameters

```
gestationDays <- 91 #gestational days when dose starts, range is 91 days (14 weeks) - 280 days (40 weeks)
```

Load data

Notice that the chemical input has the source of the FU and Clint data. ICE now has 2 different sources of these values so the source is important for tracking this information.

```
chemical <- as.data.frame(read_delim(chemFile, delim = "\t")) #there are single quotes  
                                     #in chemical names that need to be addressed, tidyR handles this well  
#chemical[1:2,]      #to check the format of input data
```

Preparing data

Minor prep work is done on the data that comes from ICE. Partitioning coefficients are obtained by internal function from the htk package using the provided phys chem parameters. Compare with the example input file for naming. Note that a few checks are done - these will generate flags that can be used to follow up as needed.

```
##column names should be labeled correctly. RegEx have been used to help, see the example file  
  
#cleaning the names for taking from an ICE Query-stricter matching is used to handel including the ADME  
colnames(chemical)<-gsub(".*Name.*", "ChemicalName", colnames(chemical))  
colnames(chemical)<-gsub(".*Parameters fu.*", "fu", colnames(chemical), ignore.case =TRUE)  
colnames(chemical)<-gsub(".*funbound.*", "fu", colnames(chemical), ignore.case =TRUE)  
colnames(chemical)<-gsub(".*fu Source.*", "fu Source", colnames(chemical), ignore.case =TRUE)  
colnames(chemical)<-gsub(".*MW.*", "MW", colnames(chemical))  
colnames(chemical)<-gsub(".*HL.*", "HL", colnames(chemical))  
colnames(chemical)<-gsub(".*Parameters Clint.*", "Clint", colnames(chemical), ignore.case = TRUE)  
colnames(chemical)<-gsub(".*Clint Source.*", "Clint Source", colnames(chemical), ignore.case = TRUE)  
colnames(chemical)<-gsub(".*LogP.*", "LogP", colnames(chemical))  
colnames(chemical)<-gsub(".*pKa, Basic.*", "pka_Accept", colnames(chemical))  
colnames(chemical)<-gsub(".*pKa, Acidic.*", "pka_Donor", colnames(chemical))  
  
#Converting units for internsic clearance values  
#chemical$Clint<-10^chemical$Clint #this moves from log10 ul/ml/10^6 cells  
                                     #to just ul/ml/10^6 cells  
chemical$logPwa<--1*(chemical$HL+1.61) #this gets the water-air partition coefficient, as the unit for  
#Add the flag here as a warning if model not appropriate or other issues related to generating prediction  
chemical$Flag <-""  
#catch for low Fu  
chemical$Flag[chemical$fu<=1e-10]<-"Fu is zero,likely due to rounding. Setting to 1e-5"  
chemical$fu[chemical$fu<=1e-10]<-1e-5  
  
#setting up additional defaults:  
if(!exists("expDose") || is.null(expDose)){  
  expDose <- 1 #current calculations assume 1mg/kg/dose  
}
```

PBPK models

Currently, the PBPK analysis in ICE supports the “solve_pbt”, “solve_gas_pbpk” and “solve_fetal_pbt” model types. This is wrapper code to format the inputs and parameters needed to run the models specified by the httk package.

Parameter processing for the PBPK models

The httk package needs some additional parameters including the correct capitalization of the species and ensuring that the adding of the user-specified data to the chem table is properly integrated.

```
#library(httk)
##preprocessing variables:
if (tolower(species) == "rat") {
  species_1 <- "Rat"
}
if (tolower(species) == "human") {
  species_1 <- "Human"
}
options(stringsAsFactors = FALSE)
#add chemical info to the table. Using variable coming from ICE
#to deal with mapping issues
chemical$DTXSID2<-paste0(chemical$DTXSID, "_n")
if(species_1=="Human"){
  chem.physical_and_invitro.data <- add_chemtable(chemical, current.table = chem.physical_and_invitro.d
}else{
  chem.physical_and_invitro.data <- add_chemtable(chemical, current.table = chem.physical_and_invitro.d
  chem.physical_and_invitro.data <- add_chemtable(chemical, current.table = chem.physical_and_invitro.d
}
chem.physical_and_invitro.data["50-00-0", "Compound"] <- 'Formaldehyde' #2025

if(route != "iv"){
  iv.dose = FALSE
}else{iv.dose =TRUE}

dpd<-24/interv #calculating the doses per day
```

to allow the user to change the bodyweight for rat and human

```
if(species_1=="Human"){
  physiology.data[4,6] <- bWeight
}else{
  physiology.data[4,4] <- bWeight #the rat body weight
}
```

initialize outputs and counter

```
output_list <- list()
p.out_all <- NULL #2025
count <- 0
```

solve_pbtck model

The solve_pbtck model is a PBPK model that has gut lumen, gut, liver, kidneys, veins, arteries, lungs, and the rest of the body compartments.

```
# note that dose=0 per mark to stop the initial dosing of the compartments
if(modelType == "solve_pbtck"){
  for(this.cas in chemical[, "CASRN"]) {
    set.seed(1234) #2025
    Kkidney2pu <- chemical$Kkidney2pu[chemical$CASRN == this.cas] #2025June, place it a
    Kliver2pu <- chemical$Kliver2pu[chemical$CASRN == this.cas] #2025June, place it a
    Krest2pu <- chemical$Krest2pu[chemical$CASRN == this.cas] #2025June, place it a
    Kgut2pu <- chemical$Kgut2pu[chemical$CASRN == this.cas] #2025June, place it a
    print(this.cas)
    half_life <- calc_half_life(chem.cas = this.cas, species = species_1) #Half life calculated

    param <- parameterize_pbtck(chem.cas = this.cas, species = species_1, default.to.human = TRUE)
    print(param$BW) # to make sure that the body weight had been changed.
    param$Kkidney2pu <- ifelse(!is.na(Kkidney2pu), Kkidney2pu, param$Kkidney2pu) #2025June
    param$Kkidney2pu_Source <- ifelse(!is.na(Kkidney2pu), "CUSTOM", "httkCalculated") #2025June
    param$Kliver2pu <- ifelse(!is.na(Kliver2pu), Kliver2pu, param$Kliver2pu) #2025June
    param$Kliver2pu_Source <- ifelse(!is.na(Kliver2pu), "CUSTOM", "httkCalculated") #2025June
    param$Krest2pu <- ifelse(!is.na(Krest2pu), Krest2pu, param$Krest2pu) #2025June
    param$Krest2pu_Source <- ifelse(!is.na(Krest2pu), "CUSTOM", "httkCalculated") #2025June
    param$Kgut2pu <- ifelse(!is.na(Kgut2pu), Kgut2pu, param$Kgut2pu) #2025June
    param$Kgut2pu_Source <- ifelse(!is.na(Kgut2pu), "CUSTOM", "httkCalculated") #2025June

    p.out0 <- as.data.frame(param) #automatically resets row names #for output #2025
    colnames(p.out0) <- paste0("ModelParam_", colnames(p.out0)) #for output #2025
    rownames(p.out0) <- this.cas #for output #2025
    p.out <- rownames_to_column(p.out0, var = "CASRN") #for output #2025

    output0 <- solve_pbtck(chem.cas = this.cas, parameters = param, doses.per.day = dpd, days = ndays, t.
    output0 <- as.data.frame(output0)
    maxAUC.h <- max(output0$AUC)*24
    #steady state concentration as a result of infusion dosing for the solve_pbtck model
    CSS0 <- calc_analytic_css(chem.cas = this.cas, parameters = param, daily.dose = expDose*dpd, doses.p
    concs <- output0 %>% select(c("time", "Cgut", "Cliver", "Cven", "Clung", "Cart", "Crest", "Ckidney"

    if(route == "iv"){
      #ignore computational errors associated with the solver
      to_ignore <- 2 #<- time point 2 will always be anomalous
      n_to_ignore <- max(1, floor((24*ndays)/interv))
      if((24/interv) == 1){
        steps_between_doses <- 97
      }else if(24/interv > 1){
```

```

      steps_between_doses <- floor(97 / (24/interv))+1
    }else if(24/interv < 1){
      steps_between_doses <- floor(97 / (24/interv))
    }
    for(i in 1:n_to_ignore){
      to_ignore[i+1] = to_ignore[i] + steps_between_doses
    }
    concs <- concs[-to_ignore,]
  }

  #convert units and store output
  output_time.h <- sapply(concs[,1], function(x) x*24) #days to hours
  output_vals <- concs %>% select(c("Cgut", "Cliver", "Cven", "Clung", "Cart", "Crest", "Ckidney", "C"))
  output <- as.data.frame(cbind('CASRN'=rep(this.cas, length(output_time.h)),
                                'half_life_h' = rep(half_life, length(output_time.h)),
                                'maxAUC.h' = rep(maxAUC.h, length(output_time.h)),
                                'CSS_plasma'=rep(CSS0, length(output_time.h)),
                                output_time.h,
                                output_vals))

  count <- count+1
  output_list[[count]] <- as.data.frame(output)
  p.out_all <- rbind(p.out_all, p.out) #2025
}
}

```

solve_gas_pbtok model

The solve_gas_pbtok is a PBPK model similar to the “solve_pbtok” model but is uses an inhalation route of exposure assuming the chemical is volatile (gas). As a result, it has some additional checks to see if the assumption of gas exposure is reasonable but will still proceed with a flag warning.

```

if(modelType == "solve_gas_pbtok"){

  #check the assumption of volatility
  chemical$Flag[chemical$HL <= -7.80388 & chemical$Flag!=""]<-"Fu is zero,likely due to rounding. Setting
    Chemical likely nonvolatile, consider appropriateness of model"
  chemical$Flag[chemical$HL <= -7.80388 & chemical$Flag==""]<-"Chemical likely nonvolatile, consider appropriate"
  # --- expDose option has currently been disabled due to a bug in the htk package ---
  # if(gasDosing == "expDose"){
  #   ConcentrationUnit <- "uM" # output unit only has one option of "uM" for solve_gas_pbtok model
  #   concMax <- max(solve_gas_pbtok(chem.cas = this.cas, parameters = NULL, doses.per.day = dpd, days = days))
  # }

  for(this.cas in chemical[, "CASRN"]) {
    set.seed(1234)
    Kkidney2pu <- chemical$Kkidney2pu[chemical$CASRN == this.cas] #2025June, place it a
    Kliver2pu <- chemical$Kliver2pu[chemical$CASRN == this.cas] #2025June, place it a
    Krest2pu <- chemical$Krest2pu[chemical$CASRN == this.cas] #2025June, place it a
    Kgut2pu <- chemical$Kgut2pu[chemical$CASRN == this.cas] #2025June, place it a
    print(this.cas)
    half_life <- calc_half_life(chem.cas = this.cas, species = species_1) #Half life calculated
  }
}

```

```

if(gasDosing == "expConc"){
  param <- parameterize_gas_pbtck(chem.cas = this.cas, species = species_1, default.to.human = TRUE) #
  param$Kkidney2pu <- ifelse(!is.na(Kkidney2pu), Kkidney2pu, param$Kkidney2pu) #2025June
  param$Kkidney2pu_Source <- ifelse(!is.na(Kkidney2pu), "CUSTOM", "httkCalculated") #2025June
  param$Kliver2pu <- ifelse(!is.na(Kliver2pu), Kliver2pu, param$Kliver2pu) #2025June
  param$Kliver2pu_Source <- ifelse(!is.na(Kliver2pu), "CUSTOM", "httkCalculated") #2025June
  param$Krest2pu <- ifelse(!is.na(Krest2pu), Krest2pu, param$Krest2pu) #2025June
  param$Krest2pu_Source <- ifelse(!is.na(Krest2pu), "CUSTOM", "httkCalculated") #2025June
  param$Kgut2pu <- ifelse(!is.na(Kgut2pu), Kgut2pu, param$Kgut2pu) #2025June
  param$Kgut2pu_Source <- ifelse(!is.na(Kgut2pu), "CUSTOM", "httkCalculated") #2025June

  p.out0 <- as.data.frame(param) #automatically resets row names #for output #2025
  colnames(p.out0) <- paste0("ModelParam_", colnames(p.out0)) #for output #2025
  rownames(p.out0) <- this.cas #for output #2025
  p.out <- rownames_to_column(p.out0, var = "CASRN") #for output #2025

  output0 <- solve_gas_pbtck(chem.cas = this.cas, parameters = param, doses.per.day = NULL, days = nday)
  output0 <- as.data.frame(output0)
  maxAUC.h <- max(output0$AUC)*24
  concs <- output0 %>% select(c("time", "Cgut", "Cliver", "Cven", "Clung", "Cart", "Crest", "Ckidney", "Cplacenta"))
}
#steady state concentration
#CSS0 <- calc_analytic_css(chem.cas = this.cas, parameters = NULL, daily.dose = expDose*dpd, doses.per.day = nday)

#convert units and store output
output_time.h <- sapply(concs[,1], function(x) x*24) #days to hours
output_vals <- concs %>% select(c("Cgut", "Cliver", "Cven", "Clung", "Cart", "Crest", "Ckidney", "Cplacenta"))
output <- as.data.frame(cbind('CASRN'=rep(this.cas, length(output_time.h)),
                             'half_life_h' = rep(half_life, length(output_time.h)),
                             'maxAUC.h' = rep(maxAUC.h, length(output_time.h)),
                             #'CSS_plasma'=rep(CSS0, length(output_time.h)),
                             output_time.h,
                             output_vals))

count <- count+1
output_list[[count]] <- as.data.frame(output)
p.out_all <- rbind(p.out_all, p.out) #2025
}
}

```

solve_fetal_pbtck model

The solve_fetal_pbtck is a human PBPK model that includes both maternal and fetal compartments and a placenta modeled as a joint organ shared by mother and fetus. It simulated gestation day of 91-280 days. A flag warning will show if the gestation day is out of this range. The exposure routes for this model are oral and IV.

```

if(modelType == "solve_fetal_pbtck"){

  if(species_1 == "Rat"){
    stop("Gestation model is for human only")
  }
}

```

```

if(gestationDays < 91 || gestationDays > 280) {
  print("Warning: the gestation day is the day when dosing starts, which shall be within the range of
}

for(this.cas in chemical[, "CASRN"]) {
  set.seed(1234)
  Kkidney2pu <- chemical$Kkidney2pu[chemical$CASRN == this.cas] #2025June, place it a
  Kliver2pu <- chemical$Kliver2pu[chemical$CASRN == this.cas] #2025June, place it a
  Krest2pu <- chemical$Krest2pu[chemical$CASRN == this.cas] #2025June, place it a
  Kgut2pu <- chemical$Kgut2pu[chemical$CASRN == this.cas] #2025June, place it a
  print(this.cas)
  half_life <- calc_half_life(chem.cas = this.cas, species = species_1) #Half life calculated fr

  param <- parameterize_fetal_pbtk(chem.cas = this.cas, species = species_1, default.to.human = TRUE)
  param$pre_pregnant_BW <- bWeight #Default 61.1 kg, not tied to physiology.data #2025Sept
  print(param$pre_pregnant_BW) #2025Sept
  #print(param$BW) # to make sure that the body weight had been changed.
  param$Kkidney2pu <- ifelse(!is.na(Kkidney2pu), Kkidney2pu, param$Kkidney2pu) #2025June
  param$Kkidney2pu_Source <- ifelse(!is.na(Kkidney2pu), "CUSTOM", "httkCalculated") #2025June
  param$Kliver2pu <- ifelse(!is.na(Kliver2pu), Kliver2pu, param$Kliver2pu) #2025June
  param$Kliver2pu_Source <- ifelse(!is.na(Kliver2pu), "CUSTOM", "httkCalculated") #2025June
  param$Krest2pu <- ifelse(!is.na(Krest2pu), Krest2pu, param$Krest2pu) #2025June
  param$Krest2pu_Source <- ifelse(!is.na(Krest2pu), "CUSTOM", "httkCalculated") #2025June
  param$Kgut2pu <- ifelse(!is.na(Kgut2pu), Kgut2pu, param$Kgut2pu) #2025June
  param$Kgut2pu_Source <- ifelse(!is.na(Kgut2pu), "CUSTOM", "httkCalculated") #2025June

  p.out0 <- as.data.frame(param) #automatically resets row names #for output #2025
  colnames(p.out0) <- paste0("ModelParam_", colnames(p.out0)) #for output #2025
  rownames(p.out0) <- this.cas #for output #2025
  p.out <- rownames_to_column(p.out0, var = "CASRN") #for output #2025

  output_f <- solve_fetal_pbtk(chem.cas = this.cas, parameters = param, times=seq(gestationDays,gesta
  output0 <- as.data.frame(output_f)
  maxAUC.h <- max(output0$AUC)*24 #convert days to hours
  maxfAUC.h <- max(output0$fAUC)*24 #convert days to hours
  concs <- output0 %>% select(c("time", "Cgut", "Cliver", "Cven", "Clung", "Cart", "Cadipose", "Crest

  if(route == "iv"){
    #ignore computational errors associated with the solver
    to_ignore <- 2 #<- time point 2 will always be anomalous
    n_to_ignore <- max(1, floor((24*ndays)/interv))
    if((24/interv) == 1){
      steps_between_doses <- 41
    }else if(24/interv > 1){
      steps_between_doses <- floor(41 / (24/interv))+1
    }else if(24/interv < 1){
      steps_between_doses <- floor(41 / (24/interv))
    }
    for(i in 1:n_to_ignore){
      to_ignore[i+1] = to_ignore[i] + steps_between_doses
    }
    concs <- concs[-to_ignore,]
  }
}

```

```

#convert units and store output
output_time.h <- sapply(concs[,1], function(x) x*24) #days to hours
output_vals <- concs %>% select(c("Cgut", "Cliver", "Cven", "Clung", "Cart", "Cadipose", "Crest", "C
output <- as.data.frame(cbind('CASRN'=rep(this.cas, length(output_time.h)),
                             'half_life.h' = rep(half_life, length(output_time.h)),
                             'maxAUC.h' = rep(maxAUC.h, length(output_time.h)),
                             'maxfAUC.h' = rep(maxfAUC.h, length(output_time.h)),
                             output_time.h,
                             output_vals))

count <- count+1
output_list[[count]] <- as.data.frame(output)
p.out_all <- rbind(p.out_all, p.out) #2025
}
}

```

Save output files

Table

An Excel output file will be created with two tabs, 1 for the PBPK parameters and one for the analysis output.

```

#build output table
#formatted_list <- lapply(output_list, assemble.table)
names(output_list) <- chemical[, "CASRN"] # <- chemical[, "CASRN"][1] for testing just one CASRN

if(modelType == "solve_pbtk"){
  formatted_list <- lapply(output_list, function(x) assemble.table_pbtk(x, nOutParam=9, nStartCol=5, nS
})
if(modelType == "solve_gas_pbtk"){
  formatted_list <- lapply(output_list, function(x) assemble.table_gas(x, nOutParam=9, nStartCol=4, nSt
})
if(modelType == "solve_fetal_pbtk"){
  formatted_list <- lapply(output_list, function(x) assemble.table_fetal(x, nOutParam=21, nStartCol=5, n
})

#for single chemical
if(length(chemical[, "CASRN"]) == 1){
  output_table <- formatted_list[[1]]
}

#for multiple chemicals
if(length(chemical[, "CASRN"]) > 1 && modelType != "solve_gas_pbtk"){
  #output_table <- rbindlist(formatted_list)
  output_table <- rbindlist(formatted_list, fill=TRUE) #to fill missing columns use fill=TRUE
  #trim redundant time rows
  to_remove <- which(output_table$compartment == 'output_time.h')[-1]
  output_table <- output_table[-to_remove]
  output_table[1, c(1:8)] <- NA
  #for solve_pbtk model, removed values for the 1st row for columns before "compartment", which are "CA

```

```

#for solve_fetal_pbt model, removed values for the 1st row for columns before "compartment", which are
colnames(output_table)[10:ncol(output_table)] <- paste('t', 1:length(output_time.h)) # "10" is where t
}

if(length(chemical[, "CASRN"]) > 1 && modelType == "solve_gas_pbt"){
  #output_table <- rbindlist(formatted_list)
  output_table <- rbindlist(formatted_list, fill=TRUE) #to fill missing columns use fill=TRUE
  to_remove <- which(output_table$compartment == 'output_time.h')[-1] #trim redundant time rows
  output_table <- output_table[-to_remove]
  output_table[1, c(1:7)] <- NA
  #removed values for the 1st row for columns before "compartment", which are "CASRN, compound, dtxsi
  colnames(output_table)[9:ncol(output_table)] <- paste('t', 1:length(output_time.h)) # "9" is where t
}

```

Preparing the chemical parameter file for writing. This will have the modeling conditions

```

param_table <- chemical
param_table$Species <- species; param_table$Model <- modelType;
param_table$Dose, mg/kg <- expDose;
if(route == "gas" & gasDosing == "expConc"){
  param_table$Exposure Concentration <- expConc #for if inhalation
  param_table$Exposure Concentration Unit <- gasInputUnit #for if inhalation
  param_table$Exposure Length, hr <- expLength #for if gas
  param_table$Exposure Period, hr <- interv #for if gas
  param_table <- param_table[, setdiff(colnames(param_table), c("Dose, mg/kg"))]
}

param_table$Route <- route
param_table$Dose Interval, hrs <- interv
param_table$Length of Dosing, Days <- ndays
#reformatting columns to match the other models:
param_table <- param_table %>% select(any_of(c("CASRN", "ChemicalName", "DTXSID", "Flag", "Species", "Model",
  "Route", "Dose, mg/kg", "Exposure Concentration", "Exposure Concentration Unit",
  "Length of Dosing, Days", "Clint", "Clint Source", "Clint_InputSource", "Clint_InputUnit",
  "LogP", "LogP_InputSource", "HL", "HL_InputSource", "pka_Accept", "pka_Acceptor", "pka_Donor", "pka_Donor",
  "pka_Accept", "pka_Acceptor", "pka_Donor", "pka_Donor")))
#colnames(param_table) <- gsub("pka_Accept", "pka Acceptor", colnames(param_table)) #2025Jun
#colnames(param_table) <- gsub("pka_Donor", "pka Donor", colnames(param_table)) #2025Jun
param_table <- left_join(param_table, p.out_all, by = "CASRN") #2025

```

#The output file is an excel workbook. This has 2 tabs, one for the PBPK parameters and one for the PBPK results

```

wb <- createWorkbook()
addWorksheet(wb, "PBPKresults"); writeData(wb, "PBPKresults", output_table)
addWorksheet(wb, "PBPKparameters"); writeData(wb, "PBPKparameters", param_table)
saveWorkbook(wb, file=Userout, overwrite = TRUE)

```

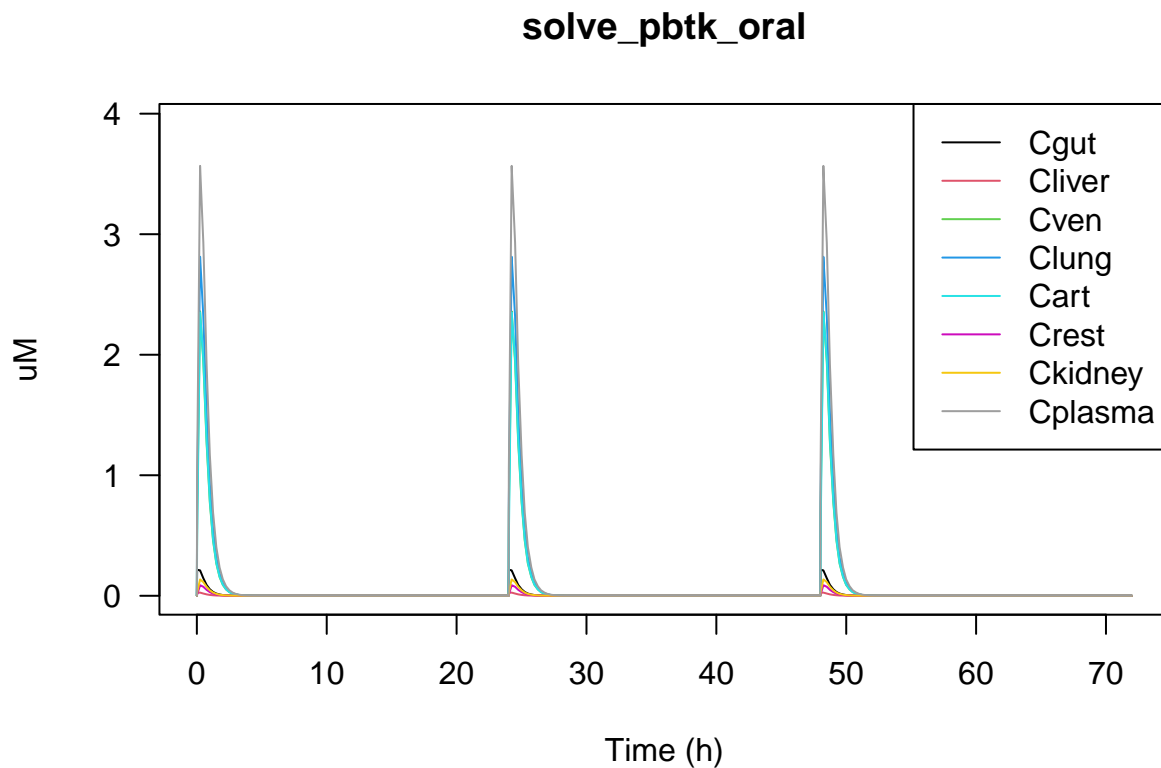
Figure

An example figure showing time series outputs for each compartment is provided. This figure shows the concentration profile in each compartment using the first CASRN provided in the chemical list.

```
example_cas_index <- which(output_table$CASRN == chemical$CASRN[1])

points_plotted <- unlist(output_table[example_cas_index, 8:ncol(output_table)])
ymax <- max(as.numeric(points_plotted),na.rm = T)*1.1

par(las=1)
plot(x=as.numeric(output_table[1,8:ncol(output_table)]),
     y=as.numeric(output_table[example_cas_index[1],8:ncol(output_table)]), type='l', ylim=c(0,ymax), ylab='uM')
for(i in 2:length(example_cas_index)){
  lines(x=as.numeric(output_table[1,8:ncol(output_table)]), y=as.numeric(output_table[example_cas_index[i],8:ncol(output_table)]), lty=1, col=c(1:length(example_cas_index)))
}
legend('topright', legend = output_table$compartment[example_cas_index], lty=1, col=c(1:length(example_cas_index)))
```



Information on the session

```
sessionInfo()
```

```
## R version 4.4.3 (2025-02-28 ucrt)
```

```

## Platform: x86_64-w64-mingw32/x64
## Running under: Windows 11 x64 (build 26100)
##
## Matrix products: default
##
##
## locale:
## [1] LC_COLLATE=English_United States.utf8
## [2] LC_CTYPE=English_United States.utf8
## [3] LC_MONETARY=English_United States.utf8
## [4] LC_NUMERIC=C
## [5] LC_TIME=English_United States.utf8
##
## time zone: America/New_York
## tzcode source: internal
##
## attached base packages:
## [1] parallel stats graphics grDevices utils datasets methods
## [8] base
##
## other attached packages:
## [1] data.table_1.17.0 readxl_1.4.5 openxlsx_4.2.8 httpk_2.2.2
## [5] doParallel_1.0.17 iterators_1.0.14 foreach_1.5.2 deSolve_1.40
## [9] rlang_1.1.5 lubridate_1.9.4 forcats_1.0.0 stringr_1.5.1
## [13] dplyr_1.1.4 purrr_1.0.4 readr_2.1.5 tidyr_1.3.1
## [17] tibble_3.2.1 ggplot2_3.5.2 tidyverse_2.0.0 knitr_1.50
##
## loaded via a namespace (and not attached):
## [1] gtable_0.3.6 xfun_0.52 lattice_0.22-6 tzdb_0.5.0
## [5] vctrs_0.6.5 tools_4.4.3 Rdpack_2.6.3 generics_0.1.3
## [9] pkgconfig_2.0.3 Matrix_1.7-2 lifecycle_1.0.4 compiler_4.4.3
## [13] munsell_0.5.1 mitools_2.4 codetools_0.2-20 survey_4.4-2
## [17] htmltools_0.5.8.1 yaml_2.3.10 pillar_1.10.1 crayon_1.5.3
## [21] tidyselect_1.2.1 zip_2.3.2 digest_0.6.37 mvtnorm_1.3-3
## [25] stringi_1.8.7 splines_4.4.3 fastmap_1.2.0 grid_4.4.3
## [29] colorspace_2.1-1 expm_1.0-0 cli_3.6.4 magrittr_2.0.3
## [33] survival_3.8-3 withr_3.0.2 scales_1.3.0 bit64_4.6.0-1
## [37] timechange_0.3.0 rmarkdown_2.29 bit_4.6.0 cellranger_1.1.0
## [41] hms_1.1.3 msm_1.8.2 evaluate_1.0.3 rbibutils_2.3
## [45] Rcpp_1.0.14 glue_1.8.0 DBI_1.2.3 rstudioapi_0.17.1
## [49] vroom_1.6.5 R6_2.6.1

```