**Microbiota predict *Clostridium difficile* severity in ‘humanized’ germ-free mice**

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**Abstract**

*Clostridium difficile* causes diarrheal disease when it successfully colonizes a dysbiotic gut microbial community. Current mouse models to study *C. difficile* infection (CDI) rely on pre-treatment with antibiotics to disrupt the mouse microbiome prior to infection with the pathogen. This is an effective model for many studies but does not allow for accurate characterization of human microbial community members that may allow for or prevent *C. difficile* colonization and expansion. To study human-associated microbes in the context of CDI, we colonized germ-free C57BL/6 mice with one of 16 human fecal samples from diarrheal or healthy donors and challenged with *C. difficile* 14 days later. Five unique donor-mice combinations resulted in severe CDI while the remaining 11 only experienced mild disease. Within the five, both healthy and diarrheal donors were susceptible to the negative effects of CDI. To determine if the presence or absence of specific microbes could predict severity, we built a classification Random Forest machine learning model based on relative abundance data of the communities on day zero. The model identified a number of bacterial populations associated with the development of severe CDI, including *Bacilliales*, *Ruminococcaceae*, *Ruminococcus, Staphylococcus, Streptococcus* and *Bacteriodetes.* Additionally, a regression model accurately predicted colonization levels of *C. difficile* at one to ten days post-infection. This model explained 99% of the variance in the number of CFU isolated from mouse stool. Members of the X, Y and Z bacterial families were predictive of future *C. difficile* colonization levels. Finally, challenging these mice with different strains of *C. difficile* revealed that susceptible human-associated microbial communities were prone to severe disease independent of strain. Taken together these results suggest that human-associated microbial communities can be recapitulated in germ-free mice and used to characterize dynamics of CDI. Because both healthy and diarrheal patients were susceptible to severe CDI, machine-learning models are useful to identify bacterial populations that allow colonization and contribute to the development of *C. difficile* associated disease in humans.

**(Importance?)**

**Introduction**

* Cdiff is a diarrheal disease, healthcare costs
* Mouse models aren’t human models
* A need for understanding human microbe contribution to disease
* Here we (for the first time) blah blah blah

**Results**

1. Germ-free mice inoculated with human feces as model for *C. difficile* infection
2. *C. difficile* infection dynamics
3. Random Forest predicts CDI severity from day 0
4. Microbial community on day 0 predicts future *C. difficile* CFU
5. Propensity for severe CDI is community-dependent and strain-independent

**Discussion**

* Discuss results, caveats about mouse weights and differneces between donors
* No donors were colonization resistant, discuss donor differences
* Discuss prediction methods and outcomes
* Discuss potential mechanisms for interesting OTUs
* Discuss different strain results
* Future work blah blah

**Methods**

* Mice ULAM number
* Donor stool ERIN IRB shit
* Bacteria/plating
* Sequencing
* Data analysis
* Machine learning

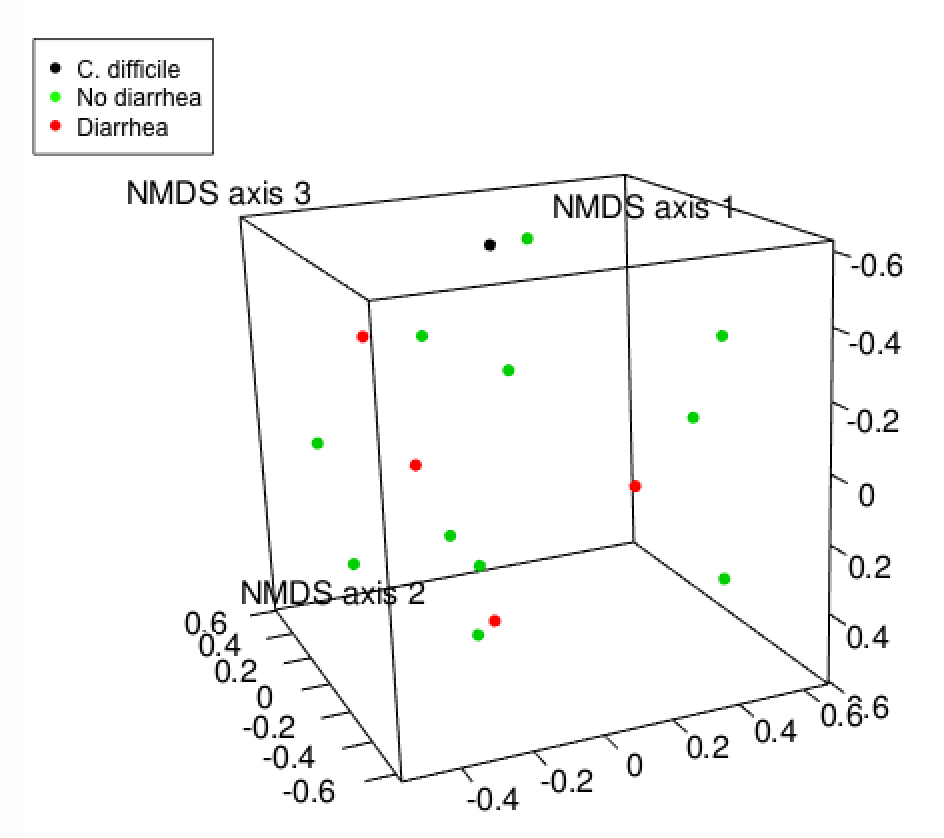
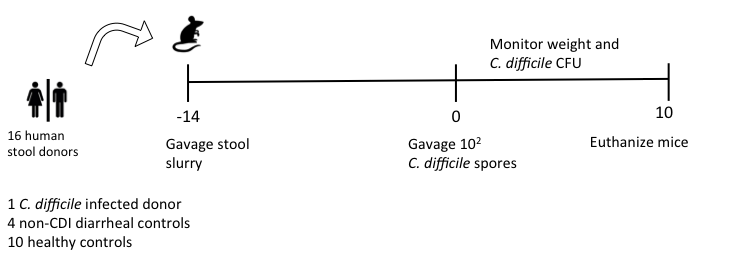
**Acknowledgments**

Lab, sequencing core, Jhansi

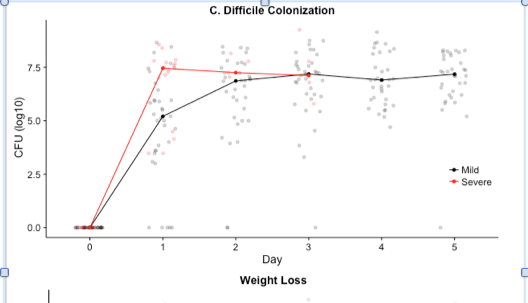
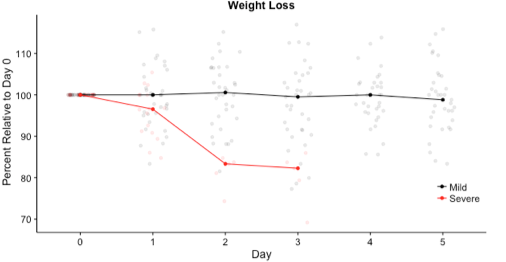
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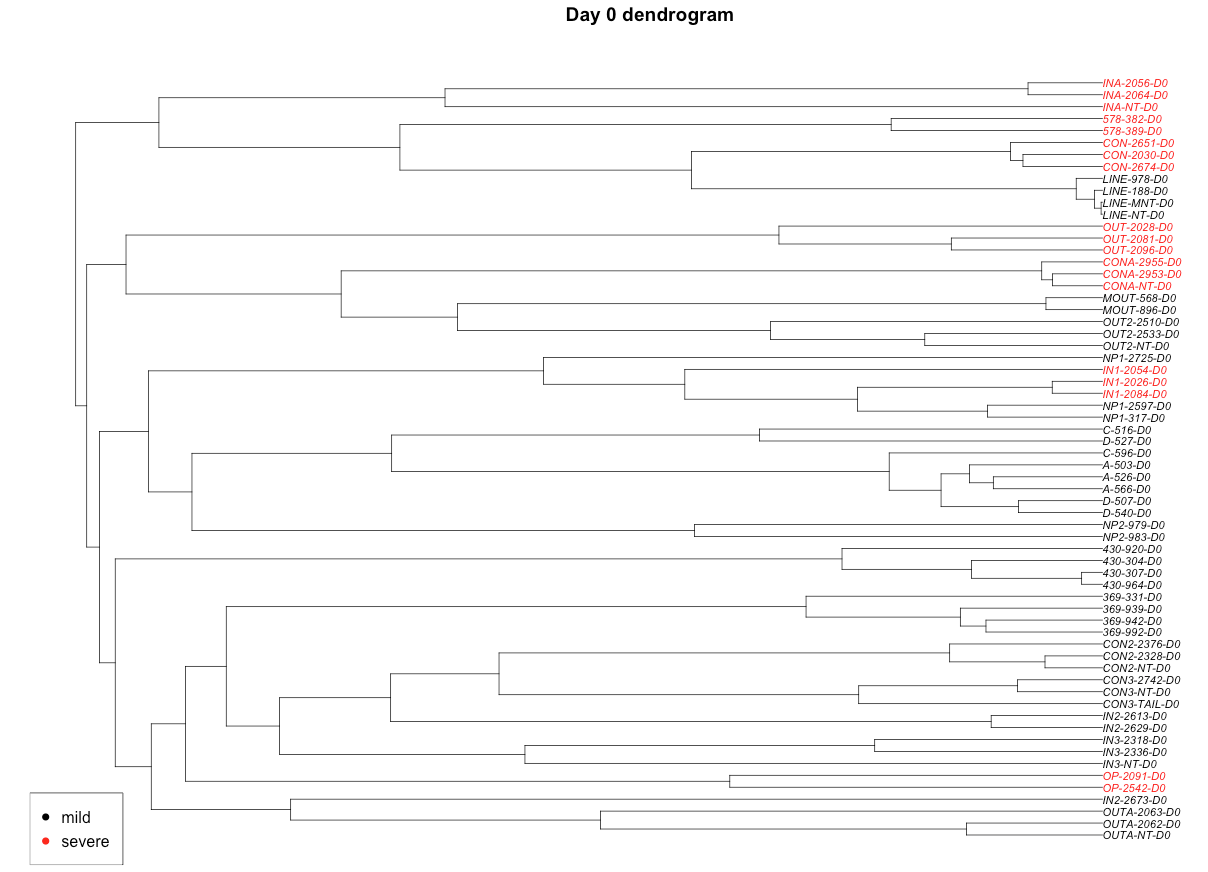
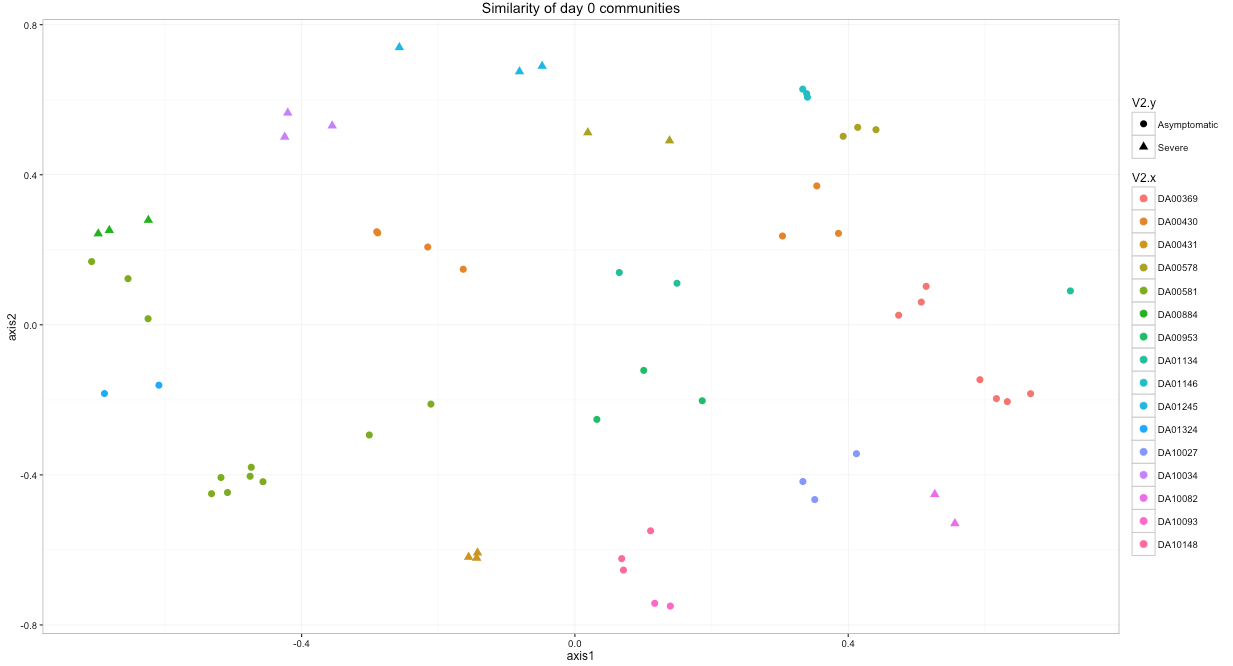
**Figure 1: Germ-free mice inoculated with human feces as a model for *C. difficile* infection**

A B

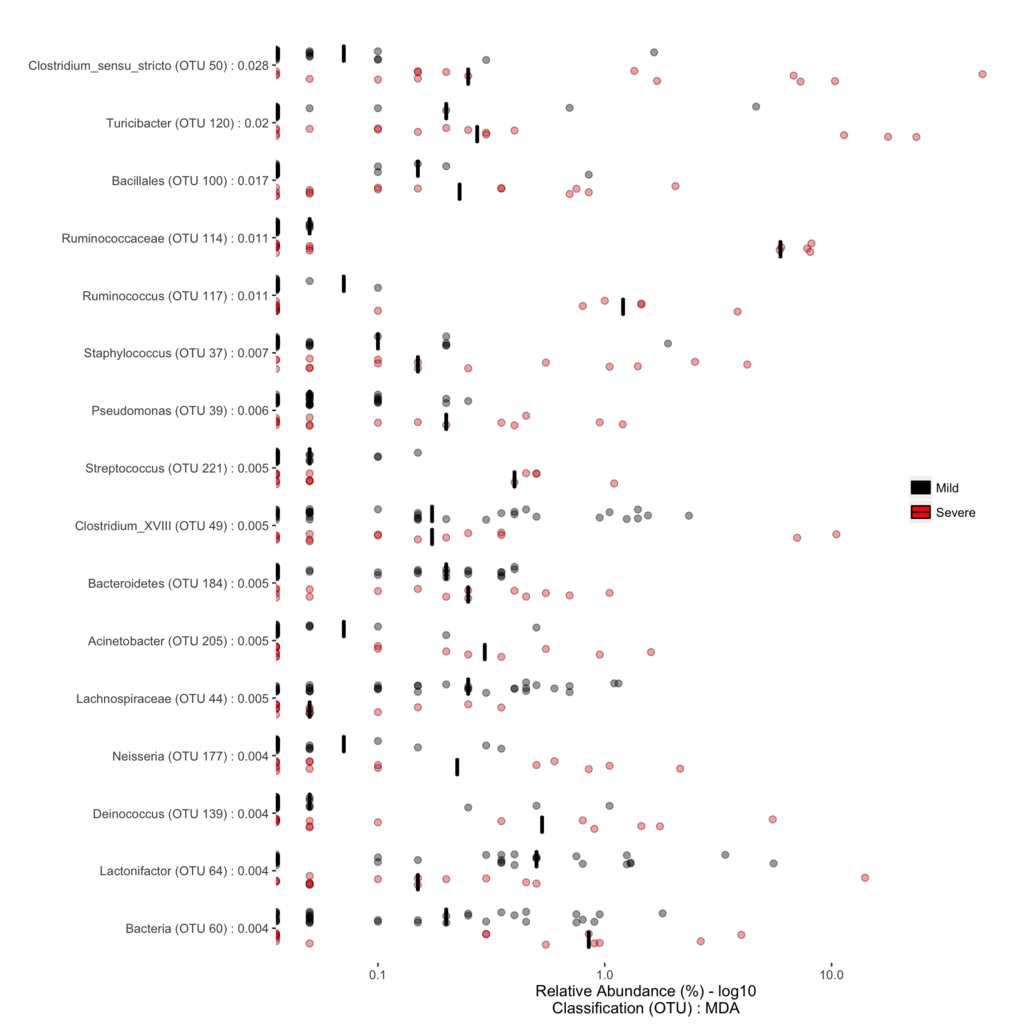
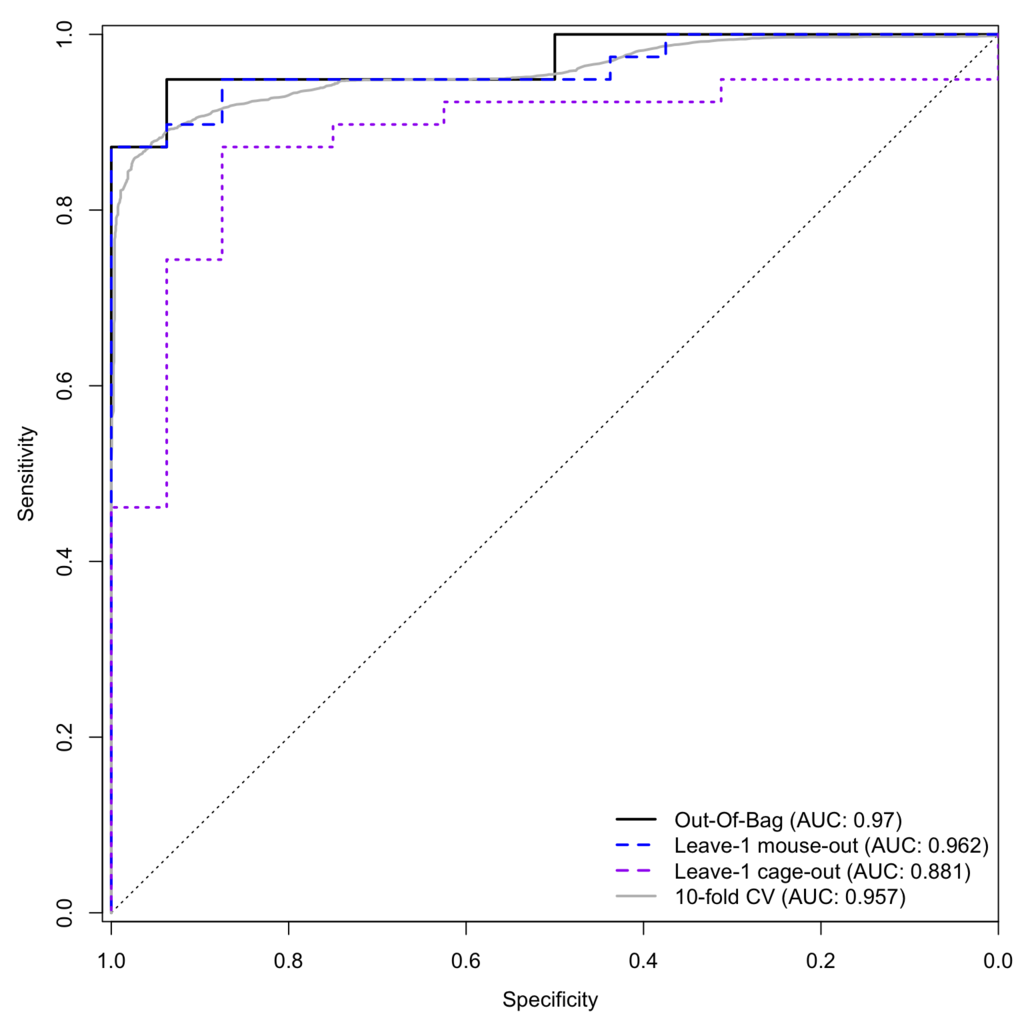


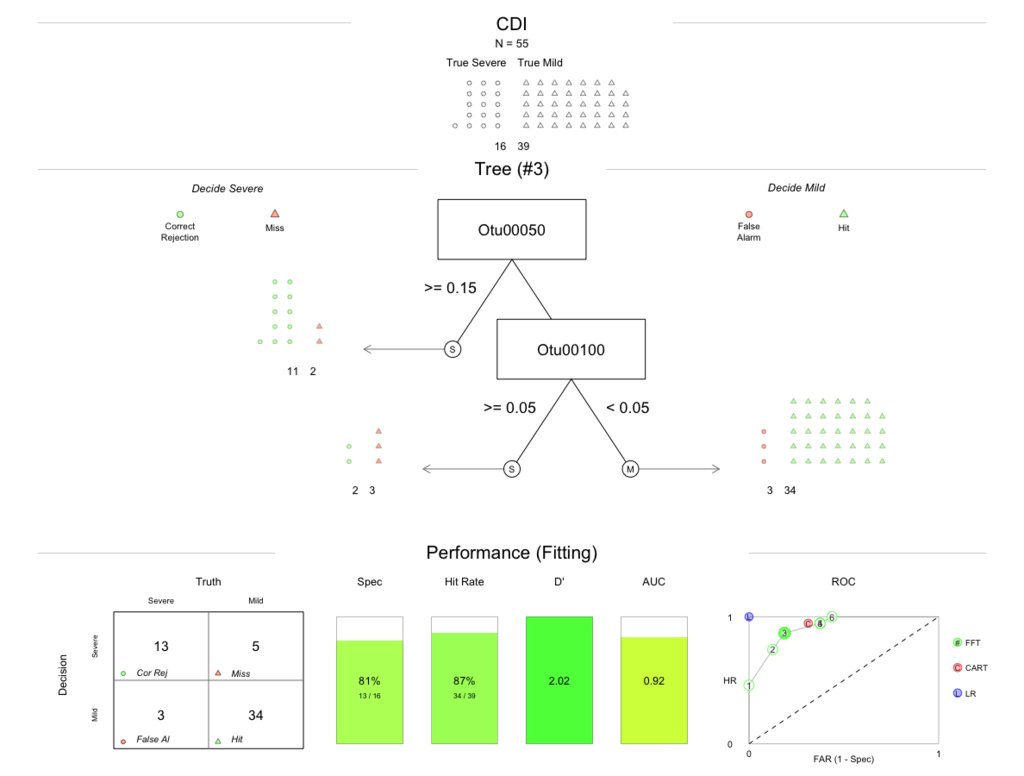
**Figure 2. *C. difficile* infection dynamics**

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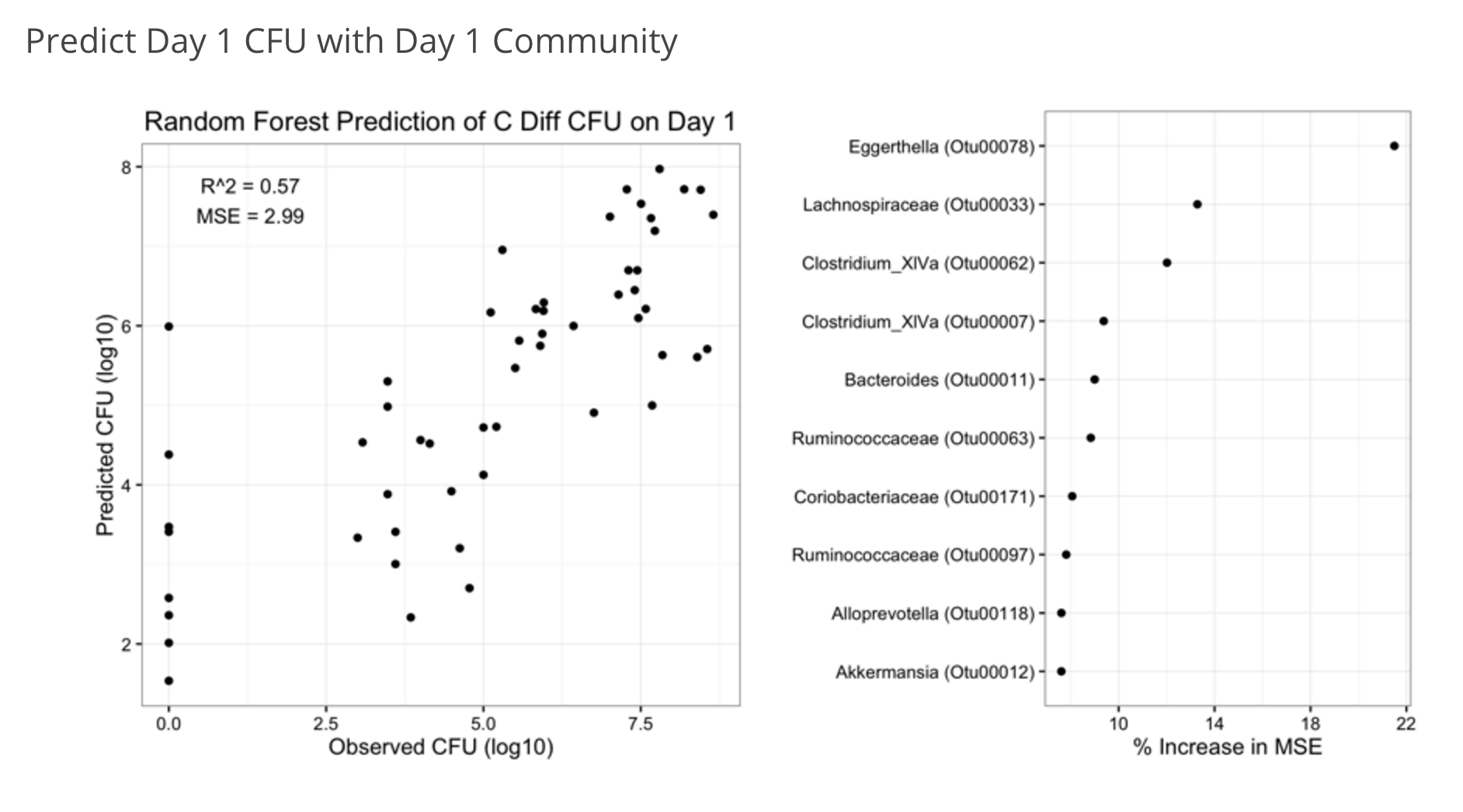
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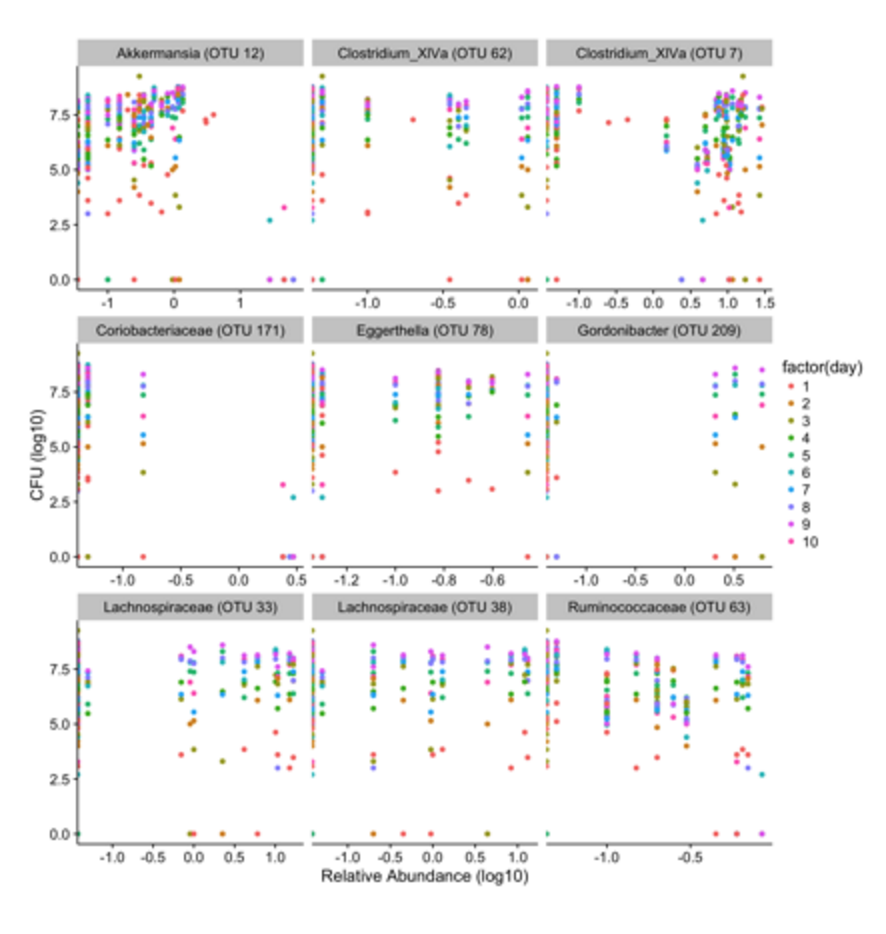
**Figure 3. Random forest predicts CDI severity from day 0 microbiome.**

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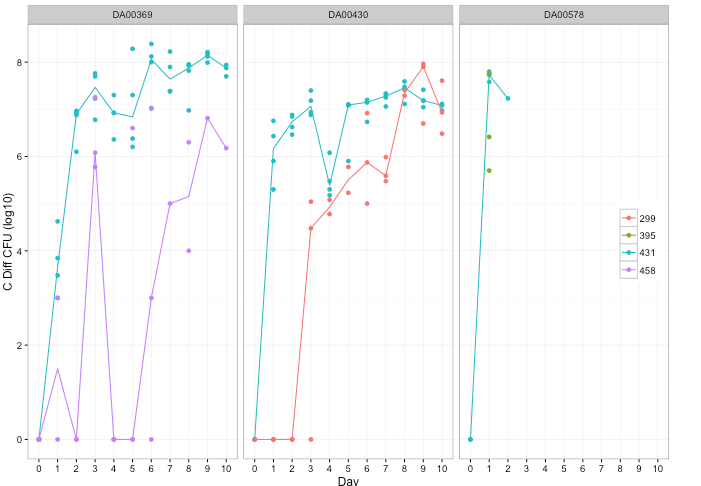
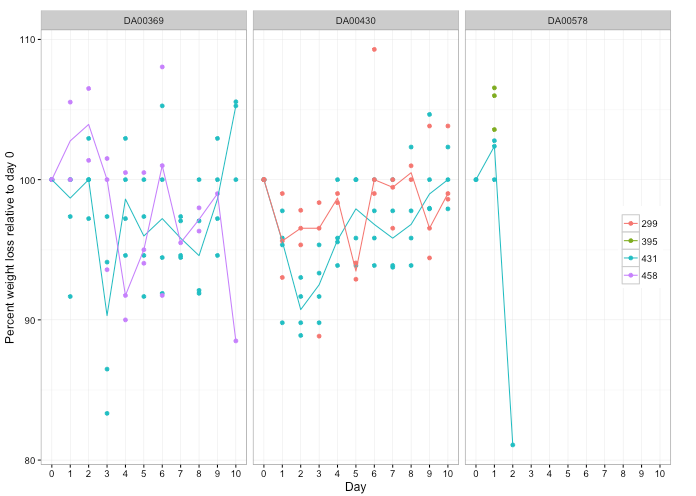
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**Figure 4. Microbial community on day 0 predicts future *C. difficile* CFU**

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**Figure 5. Propensity for severe CDI is community-dependent and strain-independent**

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