Supporting Information:

NMRlipids IV: Headgroup & glycerol backbone structures, and cation binding in bilayers with PE and PG lipids

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S1 R-PDLF and SDROSS experiments



Figure S1: (A) Chemical structure of POPE with the labeling of headgroup and glycerol backbone carbons. (B) INEPT spectra from POPE sample with the headgroup and glycerol backbone peaks labeled. (C) 2D R-PDLF spectra (D) Dipolar sliced from the 2D R-PDLF spectra with the resulting order parameters on top of figures. (E) Experimetal S-DROSS curves giving signs of the order parameters.

1.A, B etc. labels to be put in the figure.



Figure S2: (A) Chemical structure of POPG with the labeling of headgroup and glycerol backbone carbons. (B) INEPT spectra from POPG sample with the headgroup and glycerol backbone peaks labeled. (C) 2D R-PDLF spectra (D) Dipolar sliced from the 2D R-PDLF spectra with the resulting order parameters on top of figures. (E) Experimetal S-DROSS curves giving signs of the order parameters.



Figure S3: Simpson simulaton of S-DROSS curve of β -carbon of POPG.

S2 Comparison of headgroup order parameters from different force fields to experiments

The quality of PE and PG headgroup conformational ensembles in different simulations against NMR experiments is evaluated in figures S4 and S5 using C-H bond order parameters as in our previous studies for PC and PS lipids. ^{1,2} Conclusions are the same for all lipids: None of the force fields correctly captures the lipid headgroup conformational ensembles, but CHARMM36 gives results closest to experiments.

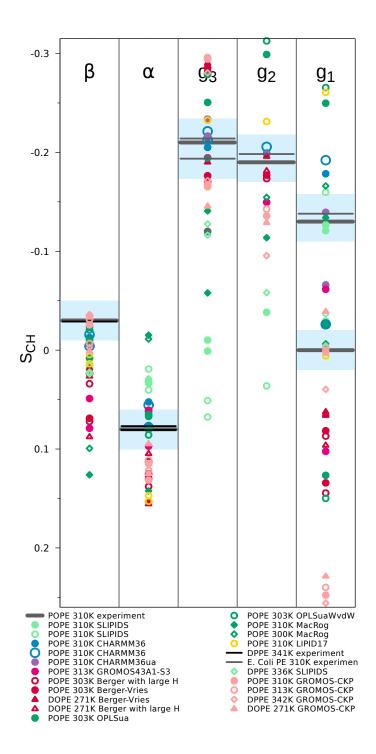


Figure S4: The headgroup and glycerol backbone order parameters of PE lipids from experiments (POPE and signs this work, DPPE from Ref. 3 and E.coliPE from Ref. 4) and simulations with different force fields.

2. This should be clarified as in NMRlipidsI and error bars should be added. Probably larger error bars for united atom models based on the report by Fuchs et al.



Figure S5: The headgroup and glycerol backbone order parameters of PG lipids from experiments (POPG and signs from this work and from Ref. 5, DPPG with 100mM NaCl from Ref. 6,and E.Coli PG results from Ref. 4). and simulations with different force fields.

S3 PC headgroup in mixtures with PE or PG lipids

Headgroup order parameters of PC lipids are unchanged upon addition of zwitterionic lipids or cholesterol in experiments, but increase upon addition of negatively charged PG or PS lipids because headgroup dipole tilts more parallel to the membrane plane after incorporation of negative charges into the membrane. ^{7,10,11} The response of PC headgroup order parameters to the addition of PE or PG lipids from different simulations is compared with experiments in figure S6. None of the simulations reproduce neither the experimentally observed increase in PC headgroup order parameters with increasing amount of PG nor the related tilting of the headgroup more parallel with the membrane. Similar observations in our previous work for PS lipids were explained by the overestimated counterion binding affinity that neturalizes the effect of added negative charge.² All simulations except Berger-OPLS predict tilting of P-N headgroup outwards from the membrane and decrease of PC headgroup order parameters upon addition of PE lipids. These results are not in line with experiments where the PC headgroup order parameters are not affected by zwitterionic lipids. The good performance of Berger-OPLS simulations in here is surprising because headgroup conformational enemble is not very close to experiments in this model and the response of headgroup order parameters to cholesterol was significantly overestimated by the Berger/Höltje force field in our previous work.1

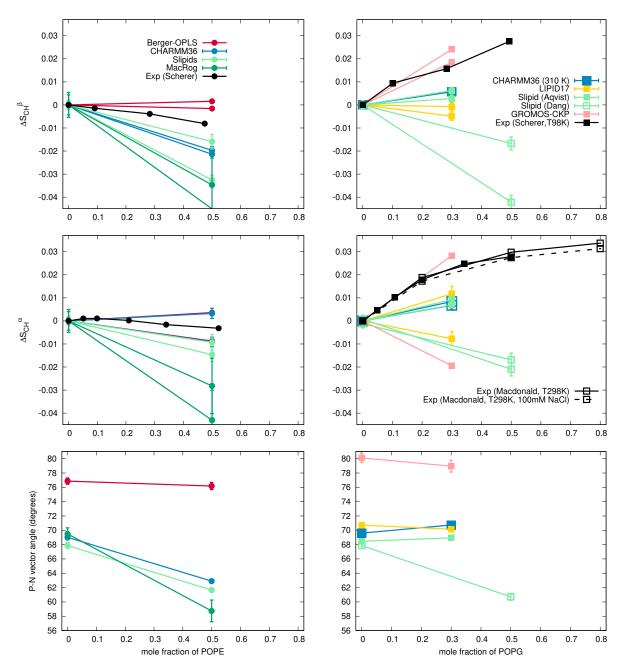


Figure S6: Modulation of POPC headgroup order parameters with increasing amount of POPE (left) and POPG (right) in bilayer from experiments 7,8 and simulations with different force fields. Signs are determined as discussed in. 1,9

S4 PG headgroup in mixtures with PC lipids

Changes in other than PC lipid headgroup with changing membrane composition are less extensively characterized in the literature. The β -carbon order parameter in PG headgroup increases mildly⁸ or is unchanged⁵ upon increasing amount of PC lipids (Fig. S7), but experimental data from α -carbon is not available. Also the tested force fields predict very small changes for the β -carbon order parameter, while the P-N vector tilt and its response to the increased amount of PC varies significantly between force fields in figure S7. Therefore, more experimental data and more accurate force fields are still required to resolve the PG conformational ensembles in mixtures with other lipids.

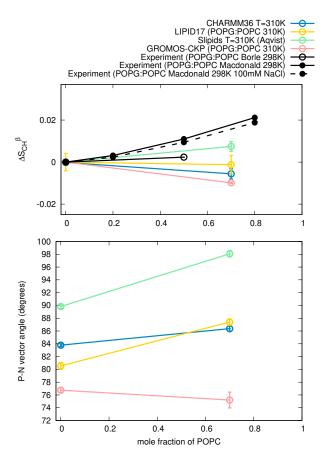


Figure S7: Modulation of PG lipid headgroup order parameters with the increasing amount of PC in lipid bilayer from experiments^{5,8} and simulations with different force fields.

S5 Calcium binding to POPC:POPG mixtures

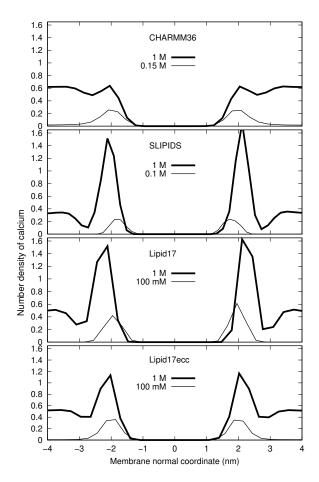


Figure S8: Calcium ion density profiles along membrane normal from simulations of POPC:POPG (1:1) mixtures with different force fields. The changes in the order parameters upon addition of $CaCl_2$ are compared with experiments in figure 10 in the main text.

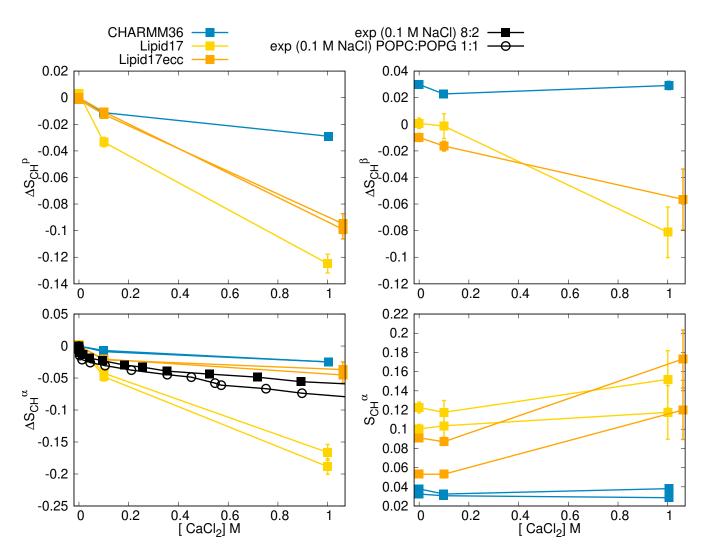


Figure S9: Modulation of headgroup order parameters of POPC (*left*) and POPG (*right*) in POPC:POPG (4:1) mixture upon addition of $CaCl_2$ in 298 K temperature from experiments⁸ and simulations. The changes with respect to the systems without $CaCl_2$ are shown for other data than for the α -carbon of POPG for which experimental order parameter is not available.

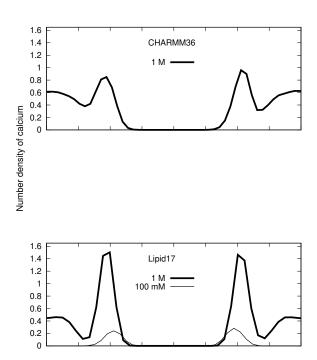


Figure S10: Calcium ion density profiles along membrane normal from simulations of POPC:POPG (4:1) mixtures with different force fields.

S6 Changes in headgroup conformations upon addition of $CaCl_2$

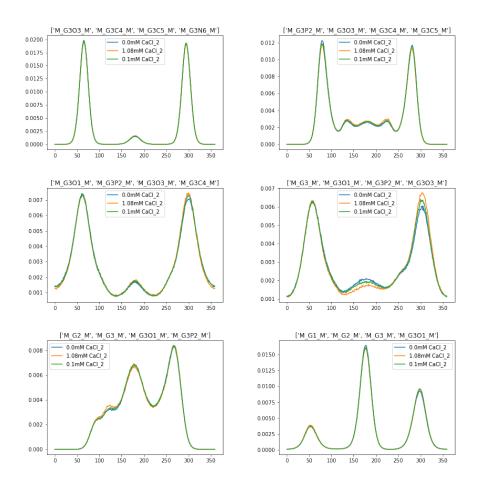


Figure S11: Changes in POPC CHARMM36 dihedrals with increasing amount of CaCl₂.

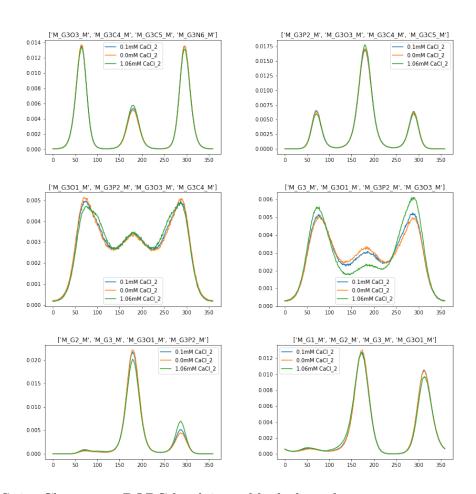


Figure S12: Changes in POPC lipid17ecc dihedrals with increasing amount of CaCl₂.

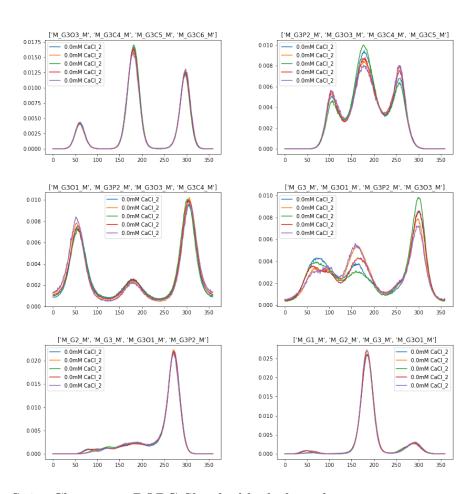


Figure S13: Changes in POPG Slipids dihedrals with increasing amount of CaCl₂.

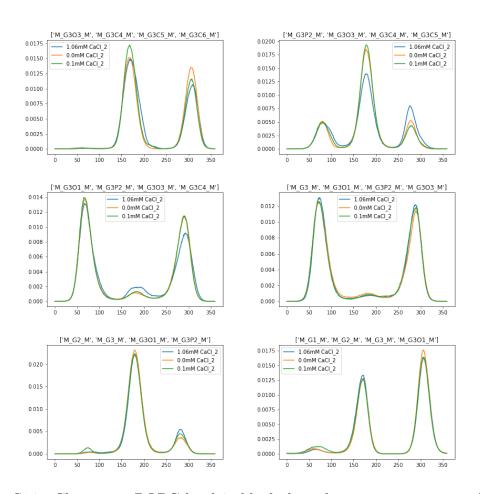


Figure S14: Changes in POPG lipid17 dihedrals with increasing amount of CaCl₂.

S7 Simulated systems

S7.1 CHARMM36

POPE 25. Simulation details by M. Javanainen.

POPE with additional NaCl 26. Simulation details by A. Peon.

POPG 27.Simulation details by Ollila.

POPG with additional NaCl 28.Simulation details by A. Peon.

POPC:POPE mixtures Data is available at.^{62,63} 300 K with v-rescale (tau=0.1 ps), 1 bar with PR semiisotropic (tau=4 ps, compressibility=4.5e-5 bar⁻¹), PME order 4 and space 0.12, recoulomb and rvdw 1.0, 128 lipids per leaflet, no ion 29.Full simulation details by Fuchs et al. POPC:POPG mixture with additional calcium 30.Simulation details by J. Madsen.

POPC:POPG mixture with additional NaCl 31.Simulation details by A. Peon.

S7.2 CHARMM36ua

POPE Data is available at. 15 32. Simulation details by T. Piggot.

S7.3 Slipids

POPE Data is available at. 18 33. Simulation details by T. Piggot.

POPE with additional NaCl 34. Simulation details by A. Peon. I have assumed that ion parameters are default Slipids, i.e., Aqvist, please correct if this is not true.

DPPE Data is available at. 17 35. Simulation details by F. Favela.

POPG Data is available at. 44 36. Simulation details by F. Favela. I have assumed that ion parameters are default Slipids, i.e., Åqvist, please correct if this is not true.

POPG with additional NaCl 37. Simulation details by A. Peon. I have assumed that ion parameters are default Slipids, i.e., Aqvist, please correct if this is not true.

DPPG Data in 298 K is available at ⁴⁶ and in 314 K at. ⁴⁵ 38. Simulation details by F. Favela. I have assumed that ion parameters are default Slipids, i.e., Åqvist, please correct if this is not true.

Table S1: List of MD simulations with PE lipids.

SO					I									I					ſ					
g	12	13	14	15	17	18	19	21	22	23	24	25	26	27	28	29	31	32	33	34	35	36	38	39
$f_{\rm t_{anal}}$ (ns)	400	100	100	2×100	100	2×100	100	100	2×400	2×400	100	100	2×400	2×100	2×100	2×100	350	300	2×100	2×100	2×100	2×100	100	100
$^{e}\mathrm{t}_{\mathrm{sim}}\mathrm{(ns)}$	200	200	200	2×200	200	2×200	200	200	2×500	2×500	200	200	2×500	2×200	2×200	2×200	200	200	2×200	2×200	2×200	2×300	200	200
$^{d}\mathrm{T}\left(\mathrm{K}\right)$	310	310	310	310	336	310	310	310	342	313	310	310	271	313	303	303	310	300	303	303	271	271	310	310
$^c\mathrm{N}_\mathrm{c}$	0	0	20	0	0	0	0	20	0	0	0	20	0	0	0	0	0	0	0	0	0	0	20	20
$^b\mathrm{N}_\mathrm{w}$	2260	25000	25000	15254	9386	<i>د</i> ٠	25000	25000	3655	3552	25000	25000	4789	3552	3328	3328	2760	5120	3552	3552	4789	4789	25000	25000
$^a\mathrm{N}_1$	144	500	500	336	288	336	500	500	128	128	500	500	128	128	128	128	144	128	128	128	128	128	200	200
NaCl (M)	0	0	0.11	0	0	0	0	0.11	0	0	0	0.11	0	0	0	0	0	0	0	0	0	0	0	0.11
force field for lipids / ions	CHARMM367	CHARMM36?	CHARMM36?	$ m CHARMM36ua^{2}$	Slipids ¹⁶	$\mathrm{Slipids}^{16}$	$\mathrm{Slipids}^{16}$	Slipids / Åqvist 16,20	GROMOS-CKP?	GROMOS-CKP?	GROMOS-CKP?	GROMOS-CKP?	GROMOS-CKP?	GROMOS 43A1-S3?	OPLS-UA vdW on H?	OPLS-UA?	$OPLS-MacRog^{30}$	${ m OPLS-MacRog}^{30}$	Berger-Vries?	Berger-largeH?	Berger-Vries?	$\operatorname{Berger-largeH}^?$	LIPID_{17}^{37}	$\text{LIPID}_{17^{37}}$
lipid/counter-ions	POPE	POPE	POPE	POPE	DPPE	POPE	POPE	POPE	DPPE	POPE	POPE	POPE	DOPE	POPE	POPE	POPE	POPE	POPE	POPE	POPE	DOPE	DOPE	POPE	POPE

 $^{^{}a}$ Number of lipid molecules with largest mole fraction

9.Citations for Berger-* simulations?

10.LIPID17 simulations with correct dihedrals still coming

8. Citation for OPLS-UA models?

 $^{^{}b}$ Number of water molecules

cNumber of additional cations

 $^{^{}d}$ Simulation temperature

 $[^]eT$ otal simulation time fT ime used for analysis

 $[^]g\mathrm{Reference}$ for simulation files

^{3.}Simulations with added NaCl are not currently used here, maybe should be removed from the table? 7. Citation for GROMOS 43A1-S3? 4. Citation for CHARMM36 PE? 5. Which ion model is used in ¹⁴? 6. Citation for GROMOS-CKP?

Table S2: List of MD simulations with PG lipids.

g	40	41	42	44	45	46	47	48	50	51	52	53
$f_{ m tanal} (m ns)$	100	100	100	100		, 100	100	, 100				100
$^{e}\mathrm{t_{sim}(ns)}$	100	200	200	250	200	400	200	200	200	200	200	200
$^{d}\mathrm{T}$ (K)		310	310	298	314	298	310	310	310	310	310	310
$^c{ m N}_{ m c}$	0	49	0	0	0	0	0	49	0	49	0	49
$^{b}{ m N_w}$	4110	25000	25000	10664	11232	11232	25000	25000	25000	25000	25000	25000
$^a\mathrm{N}_1$	118	200	500	288	288	288	200	200	200	500	200	200
NaCl (M)	0	0.11	0	0	0	0	0	0.11	0	0.11	0	0.11
lipid/counter-ions force field for lipids / ions	CHARMM36? 11.	CHARMM36?	CHARMM36?	_	_	_	Slipids / Åqvist 20,43		$\frac{\text{LIPID17}}{\text{Dang}^{37,49}}$	$LIPID17^{?}$	GROMOS-CKP?	GROMOS-CKP?
lipid/counter-ions	$POPG/K^+$	POPG	POPG	$POPG/Na^{+}$	$DPPG/Na^{+}$	$\mathrm{DPPG/Na^+}$	POPG	POPG	POPG	POPG	POPG	POPG

 $[^]a\mathrm{Number}$ of lipid molecules with largest mole fraction

13.Citations and ion model for CHARMM36?

14.Lipid17 simulation with ions with correct dihedral potentials still coming?

15. Citation and ion model for GROMOS-CKP?

 $[^]b\mathrm{Number}$ of water molecules

 $[^]c\mathrm{Number}$ of additional cations

 $[^]d\mathbf{Simulation\ temperature}$

 $[^]f$ Time used for analysis e Total simulation time

 $[^]g$ Reference for simulation files

^{12.}Simulations with added NaCl are not currently used here, maybe should be removed from the table?

Table S3: List of MD simulations with PE and PG lipids mixed with PC.

	lipid/counter-ions	force field for lipids $/$ ions	NaCl (M)	$NaCl(M) CaCl_2(M)$	$^a\mathrm{N}_1$	$^{ m w}{ m N}_q$	$^c\mathrm{N}_\mathrm{c}$	d T (K)	$^{e}\mathrm{t}_{\mathrm{sim}}(\mathrm{ns})$	$f_{\rm t_{anal}}$ (ns)	
	POPC	CHARMM36?	0	0	200	25000	0	310	200	100	54
	POPC:POPG (7:3)	${ m CHARMM36}^{?}$	0	0	350	25000	0	310	200	100	55
	POPC:POPG (1:1)	${ m CHARMM36}^{?}$	0	0	150:150	31500	0	298	200	400	56
	POPC:POPG (1:1)	${ m CHARMM36}^{?}$	0	0.1	150:150	31329	22	298	400	300	22
	POPC:POPG (1:1)	${ m CHARMM36}^{?}$	0	1.08	150:150	29766	578	298	200	400	28
	POPC:POPG (4:1)	${ m CHARMM36}^{?}$	0	0	350:88	26280	0	298	200	400	59
	POPC:POPG (4:1)	${ m CHARMM36}^{?}$	0	0.1	350.88	26280	47	298	200	400	09
	POPC:POPG (4:1)	${ m CHARMM36}^{?}$	0	1.0	350:88	24927	451	298	200	400	61
	POPC	CHARMM36?	0	0	256	8704	0	300	300	250	62
	POPC:POPE (1:1)	${ m CHARMM36}^{?}$	0	0	128	8704	0	300	300	250	63
	POPC	OPLS-MacRog ³⁰	0	0	128	5120	0	300	200	300	64
	POPC:POPE (1:1)	${ m OPLS-MacRog}^{30}$	0	0	128	5120	0	300	200	300	65
•	POPC	$Slipid^{16}$	0	0	512	23943	0	298	170	100	99
	POPC:POPE (1:1)	$ m Slipid^{16}$	0	0	128	5120	0	298	200	300	29
•	POPC	GROMOS-CKP / ??? ?	0	0	200	25000	0	310	200	100	89
	POPC:POPG (7:3)	GROMOS-CKP / $??$?	0	0	350:150	25000	0	310	200	100	69
•	POPC	Slipid ¹⁶	0	0	200	25000	0	310	200	100	70
	POPC:POPG (7:3)	Slipid / Åqvist 16,20	0	0	350:150	25000	0	310	200	100	71
	POPC:POPG (1:1)	Slipid / $Dang^{16,49,72,73}$	0	0	128:128	12800	0	298	200	400	74
	POPC:POPG (1:1)	Slipid / $Dang^{16,49,72,73}$	0	0.1	128:128	12800	23	298	200	400	74
	POPC:POPG (1:1)	Slipid / $Dang^{16,49,72,73}$	0	0.2	128:128	12800	46	298	1500	200	74
	POPC:POPG (1:1)	Slipid / $Dang^{16,49,72,73}$	0	0.5	128:128	12800	115	298	1500	200	74
	POPC:POPG (1:1)	Slipid / $Dang^{16,49,72,73}$	0	1.0	128:128	12800	230	298	1500	200	74
1											

 $[^]a\mathrm{Number}$ of lipid molecules with largest mole fraction $^b\mathrm{Number}$ of water molecules

16. Citation and ion model for GROMOS-CKP?

17. Upcoming Lipid17ecc with POPC:POPS (4:1) mixture simulations to be added.

 $[^]c\mathrm{Number}$ of additional cations

 $[^]d {\bf Simulation\ temperature} \\ ^e {\bf Total\ simulation\ time}$

 $[^]f$ Time used for analysis

 $[^]g$ Reference for simulation files

Table S4: List of MD simulations with PE and PG lipids mixed with PC.

gfiles	75	92	2.2	78	79	80	84	82	98	87	88	68	06	91	92	93	94
$f_{ m t_{anal}} (m ns)$	350	250	200	200	198	200	300	300	300	333	300	400	200	200	200	200	200
$^{e}\mathrm{t_{sim}(ns)}$	400	400	1200	320	718	720	400	400	400	347.8	400	009	300	300	300	300	300
$^{d}\mathrm{T}\left(\mathrm{K}\right)$	298	298	298	298	298	298	298	298	298	298	298	298	300	300	300	300	300
$^c\mathrm{N}_\mathrm{c}$	0	47	475	0	22	569	0	47	475	0	54	269	0	0	0	0	0
$^{ m b}_{ m N}$	26265	26124	24840	31572	31401	29865	26265	26124	24840	31572	29865	29865	10240	11008	10240	11008	11008
$^a\mathrm{N}_1$	350:88	350:88	350:88	150:150	150:150	150:150	350:88	350:88	350:88	150:150	150:150	150:150	256	128	128	256	128
$CaCl_{2}(M)$	0	0.1	1.0	0	0.1	1.0	0	0.1	1.0	0	0.1	1.0	0	0	0	0	0
NaCl (M)	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
force field for lipids / ions	$Lipid17 / Dang^{37,49,73}$	${ m Lipid17} / { m Dang}^{37,49,73}$	$Lipid17 / Dang^{37,49,73}$	$Lipid17 / Dang^{37,49,73}$	$Lipid17 / Dang^{37,49,73}$	$Lipid17 / Dang^{37,49,73}$	$Lipid17ecc / ECC-ions^{81-83}$	Berger 7 18.	Berger? 19.	Berger [?] 20.	Berger? 21.	Berger [?] 22.					
lipid/counter-ions	POPC:POPG (4:1)	POPC:POPG (4:1)	POPC:POPG (4:1)	POPC:POPG (1:1)	POPC:POPG (1:1)	POPC:POPG (1:1)	POPC:POPG (4:1)	POPC:POPG (4:1)	POPC:POPG (4:1)	POPC:POPG (1:1)	POPC:POPG (1:1)	POPC:POPG (1:1)	POPC	POPC:POPE (1:1)	POPC:DOPE (1:1)	DOPC	DOPC:DOPE (1:1)

 $[^]a\mathrm{Number}$ of lipid molecules with largest mole fraction

23. Citation and description for "Berger" model?

24. Upcoming Lipid17ecc with POPC:POPS (4:1) mixture simulations to be added.

 $[^]b\mathrm{Number}$ of water molecules

 $[^]c\mathrm{Number}$ of additional cations

 $[^]d\mathbf{Simulation\ temperature}$

 $[^]e\mathrm{Total}$ simulation time

 $[^]f\! \mathrm{Time}$ used for analysis $^g\! \mathrm{Reference}$ for simulation files

POPC:POPG mixture with additional NaCl 39. Simulation details by A. Peon. I have assumed that ion parameters are default Slipids, i.e., Aqvist, please correct if this is not true.

S7.4 Berger

POPE Data is available at. 33,34 40. Simulation details by T. Piggot.

DOPE Data is available at. 35,36 41. Simulation details by T. Piggot.

POPC:POPE, POPC:DOPE and DOPC:DOPE mixtures Data is available at.^{90,91} 300 K with v-rescale (tau=0.1 ps), 1 bar with PR semiisotropic (tau=4 ps, compressibility=4.5e-5 bar⁻¹), PME order 4 and space 0.12, recoulomb and rvdw 1.0, 128 lipids per leaflet, no ion 42.Simulation details by Fuchs et al.

S7.5 GROMOS 43A1-S3

POPE Data is available at. 27 43. Simulation details by T. Piggot.

S7.6 OPLS-UA

POPE Data is available at. 29 44. Simulation details by T. Piggot.

POPE with vdW interaction in H Data is available at. 28 45. Simulation details by T. Piggot.

S7.7 GROMOS-CKP and GROMOS-CKPM

POPE Data is available at. 23 46. Simulation details by T. Piggot.

DOPE Data is available at. 26 47. Simulation details by T. Piggot.

DPPE Data is available at. 22 48. Simulation details by T. Piggot.

S7.8 Lipid17

S7.9 ECC-LIPID POPG

In ECC-lipid models, electronic continuum correction (ECC) is applied to implicitly include the missing electronic polarizability into the force field description. 95,96 In practise, this is implemented by scaling the charges and Lennard-Jones σ s of headgroup, glycerol backbone, and carbonyl regions of Amber Lipid14/17 models are scaled by constant factors. Here, we follow the approach that previously improved ion binding to bilayers containing negatively charged PS lipids: 96 ECC-POPC parameters (scaling factors f_q =0.8 and f_σ =0.89 applied to Lipid14 POPC parameters) 95 were used for POPC and scaling factors of f_q =0.75 and f_σ =0.89 were applied to the charges and Lennard-Jones σ s of headgroup, glycerol backbone, and carbonyl regions of Amber Lipid17 POPG parameters. The Lipid17 parameters (described above) and initial configurations were taken from Ref. 50 with the correct dihedral type, and the resulting parameters are available from Ref. ? . ECC-ion parameters with the scaled charges, $^{81-83}$ downloaded from bitbucket.org/hseara/ions/src/master/, were used in these simulations.

References

- (1) Botan, A.; Favela-Rosales, F.; Fuchs, P. F. J.; Javanainen, M.; Kanduč, M.; Kulig, W.; Lamberg, A.; Loison, C.; Lyubartsev, A.; Miettinen, M. S. et al. Toward Atomistic Resolution Structure of Phosphatidylcholine Headgroup and Glycerol Backbone at Different Ambient Conditions. J. Phys. Chem. B 2015, 119, 15075–15088.
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- (3) Seelig, J.; Gally, H. U. Investigation of phosphatidylethanolamine bilayers by deuterium and phosphorus-31 nuclear magnetic resonance. *Biochemistry* **1976**, *15*, 5199–5204.
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