# **Supporting Information:**

# NMRlipids IV: Headgroup & glycerol backbone structures, and cation binding in bilayers with PE and PG lipids

Pavel Buslaev,<sup>†</sup> Fernando Favela-Rosales,<sup>‡</sup> Patrick Fuchs,<sup>¶</sup> Matti Javanainen,<sup>§</sup> Jesper J. Madsen,<sup>∥,⊥</sup> Josef Melcr,<sup>§,#</sup> Markus S. Miettinen,<sup>@</sup> O. H. Samuli Ollila,\*,<sup>△</sup> Chris G. Papadopoulos,<sup>▽</sup> Antonio Peón,<sup>††</sup> Thomas J. Piggot,<sup>‡‡</sup> and Pierre Poulain<sup>¶</sup>

† University of Jyväskylä

‡Departamento de Investigación, Tecnológico Nacional de México, Campus Zacatecas
Occidente, México

¶Paris, France

§Institute of Organic Chemistry and Biochemistry of the Czech Academy of Sciences, Flemingovo nám. 542/2, CZ-16610 Prague 6, Czech Republic

 $\parallel Department\ of\ Chemistry,\ The\ University\ of\ Chicago,\ Chicago,\ Illinois,\ United\ States\ of\ America$ 

⊥Department of Global Health, College of Public Health, University of South Florida, Tampa,
Florida, United States of America

#Groningen Biomolecular Sciences and Biotechnology Institute and The Zernike Institute for Advanced Materials, University of Groningen, 9747 AG Groningen, The Netherlands @Department of Theory and Bio-Systems, Max Planck Institute of Colloids and Interfaces, 14424 Potsdam, Germany

 $\triangle Institute of Biotechnology, University of Helsinki$ 

 $\nabla I2BC$  - University Paris Sud S2  $\dagger\dagger Spain$ 

‡‡Chemistry, University of Southampton, Highfield, Southampton SO17 1BJ, United

#### S1 Simulated systems

#### **S1.1 CHARMM36**

POPE Simulation details by M. Javanainen.

POPE with additional NaCl Simulation details by A. Peon.

POPG Simulation details by Ollila.

POPG with additional NaCl Simulation details by A. Peon.

POPC:POPE mixtures Data is available at.<sup>1,2</sup> 300 K with v-rescale (tau=0.1 ps), 1 bar with PR semiisotropic (tau=4 ps, compressibility=4.5e-5 bar<sup>-1</sup>), PME order 4 and space 0.12, recoulomb and rvdw 1.0, 128 lipids per leaflet, no ion Full simulation details by Fuchs et al. POPC:POPG mixture with additional calcium Simulation details by J. Madsen.

POPC:POPG mixture with additional NaCl Simulation details by A. Peon.

#### S1.2 CHARMM36ua

POPE Data is available at. 3 Simulation details by T. Piggot.

#### S1.3 Slipids

POPE Data is available at. 4 Simulation details by T. Piggot.

POPE with additional NaCl Simulation details by A. Peon. I have assumed that ion parameters are default Slipids, i.e., Åqvist, please correct if this is not true.

DPPE Data is available at. <sup>5</sup> Simulation details by F. Favela.

*POPG* Data is available at.<sup>6</sup> Simulation details by F. Favela. I have assumed that ion parameters are default Slipids, i.e., Åqvist, please correct if this is not true.

POPG with additional NaCl Simulation details by A. Peon. I have assumed that ion parameters are default Slipids, i.e., Åqvist, please correct if this is not true.

DPPG Data in 298 K is available at<sup>7</sup> and in 314 K at.<sup>8</sup> Simulation details by F. Favela. I have assumed that ion parameters are default Slipids, i.e., Åqvist, please correct if this is not

true.

POPC:POPG mixture with additional NaCl Simulation details by A. Peon. I have assumed that ion parameters are default Slipids, i.e., Åqvist, please correct if this is not true.

#### S1.4 Berger

POPE Data is available at. 9,10 Simulation details by T. Piggot.

DOPE Data is available at.  $^{11,12}$  Simulation details by T. Piggot.

POPC:POPE, POPC:DOPE and DOPC:DOPE mixtures Data is available at.<sup>13,14</sup> 300 K with v-rescale (tau=0.1 ps), 1 bar with PR semiisotropic (tau=4 ps, compressibility=4.5e-5 bar<sup>-1</sup>), PME order 4 and space 0.12, recoulomb and rvdw 1.0, 128 lipids per leaflet, no ion Simulation details by Fuchs et al.

#### S1.5 GROMOS 43A1-S3

POPE Data is available at. 15 Simulation details by T. Piggot.

#### S1.6 OPLS-UA

*POPE* Data is available at. <sup>16</sup> Simulation details by T. Piggot.

POPE with vdW interaction in H Data is available at. <sup>17</sup> Simulation details by T. Piggot.

#### S1.7 GROMOS-CKP and GROMOS-CKPM

POPE Data is available at. 18 Simulation details by T. Piggot.

DOPE Data is available at. 19 Simulation details by T. Piggot.

DPPE Data is available at. 20 Simulation details by T. Piggot.

#### S1.8 Lipid17

#### S1.9 ECC-LIPID POPG

In ECC-lipid models, electronic continuum correction (ECC) is applied to implicitly include the missing electronic polarizability into the force field description.  $^{21,22}$  In practise, this is implemented by scaling the charges and Lennard-Jones  $\sigma$ s of headgroup, glycerol backbone, and carbonyl regions of Amber Lipid14/17 models are scaled by constant factors. Here, we follow the approach that previously improved ion binding to bilayers containing negatively charged PS lipids:  $^{22}$  ECC-POPC parameters (scaling factors  $f_q$ =0.8 and  $f_\sigma$ =0.89 applied to Lipid14 POPC parameters) $^{21}$  were used for POPC and scaling factors of  $f_q$ =0.75 and  $f_\sigma$ =0.89 were applied to the charges and Lennard-Jones  $\sigma$ s of headgroup, glycerol backbone, and carbonyl regions of Amber Lipid17 POPG parameters. The Lipid17 parameters (described above) were taken from Ref.  $^{23}$  with the correct dihedral type, and the resulting parameters are available from Ref.  $^{2}$  ECC-ion parameters with the scaled charges  $^{24-26}$  were used in these simulations.

### S2 R-PDLF and SDROSS experiments

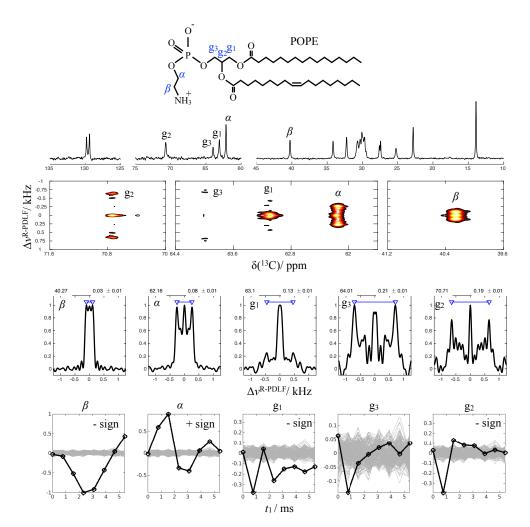


Figure S1: (A) Chemical structure of POPE with the labeling of headgroup and glycerol backbone carbons. (B) INEPT spectra from POPE sample with the headgroup and glycerol backbone peaks labeled. (C) 2D R-PDLF spectra (D) Dipolar sliced from the 2D R-PDLF spectra with the resulting order parameters on top of figures. (E) Experimetal S-DROSS curves giving signs of the order parameters.

A, B etc. labels to be put in the figure.

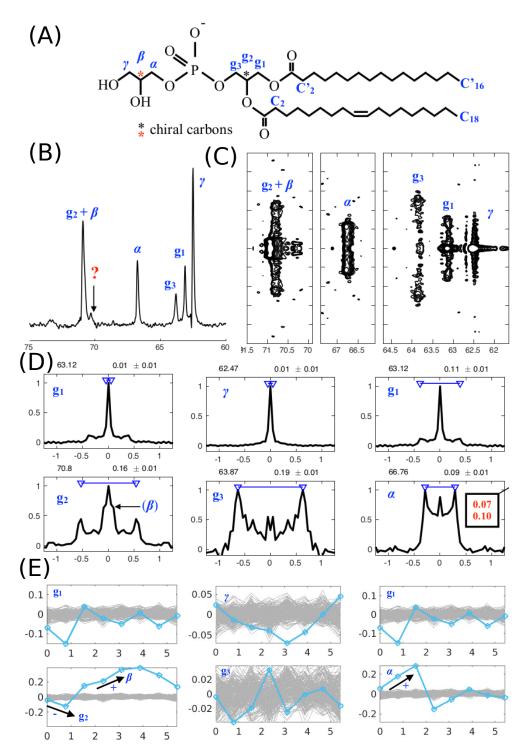


Figure S2: (A) Chemical structure of POPG with the labeling of headgroup and glycerol backbone carbons. (B) INEPT spectra from POPG sample with the headgroup and glycerol backbone peaks labeled. (C) 2D R-PDLF spectra (D) Dipolar sliced from the 2D R-PDLF spectra with the resulting order parameters on top of figures. (E) Experimetal S-DROSS curves giving signs of the order parameters.

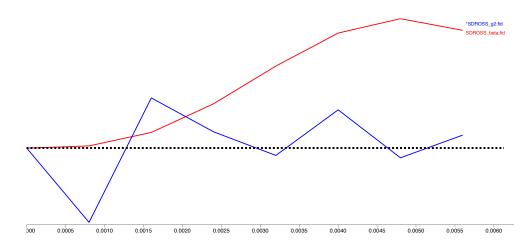


Figure S3: Simpson simulaton of S-DROSS curve of  $\beta$ -carbon of POPG.

# S3 Changes of PG headroup order parameters upon addition of PC

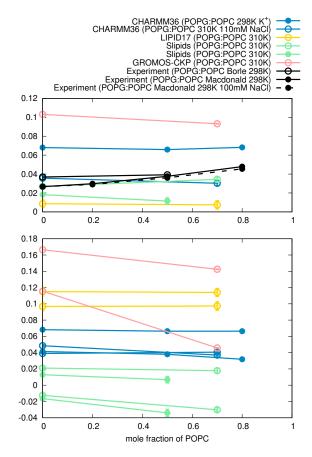


Figure S4: Modulation of PG lipid headgroup order parameters with the increasing amount of PC in lipid bilayer from experiments <sup>27,28</sup> and simulations with different force fields.

# S4 Sodium binding to POPC simulations

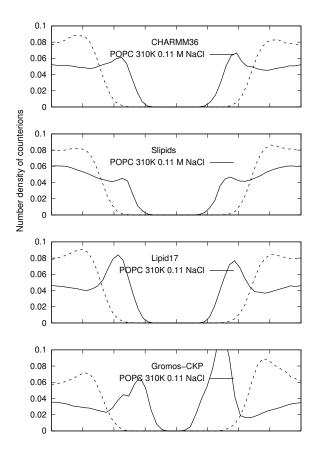


Figure S5: Sodium (solid line) and choride ion density profiles along membrane normal from different simulations with PC lipids.

Discussion about differences to the NMR lipids II to be discussed once we have the details on ions models.

## S5 Calcium binding to POPC:POPG (4:1) mixtures

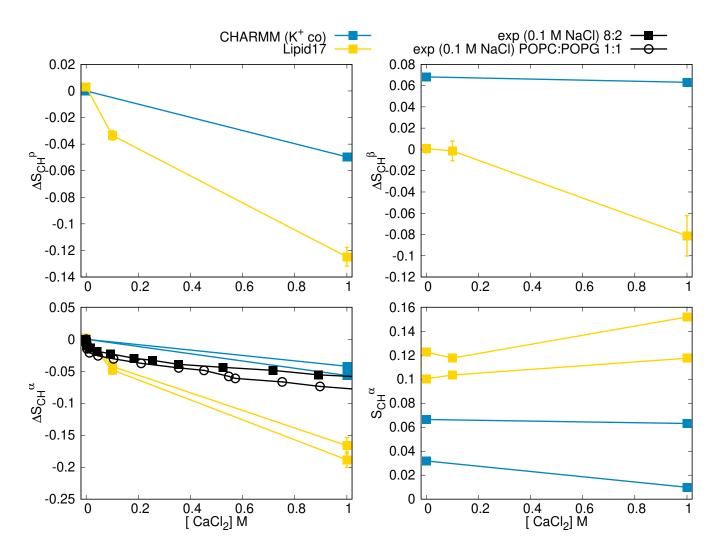


Figure S6: Modulation of headgroup order parameters of POPC (left) and POPG (right) in POPC:POPG (4:1) mixture upon addition of CaCl<sub>2</sub> in 298 K temperature from experiments <sup>28</sup> and simulations. The changes with respect to the systems without CaCl<sub>2</sub> are shown for other data than for the  $\alpha$ -carbon of POPG for which experimental order parameter is not available. CHARMM36 simulations should be longer and with Na couterions.

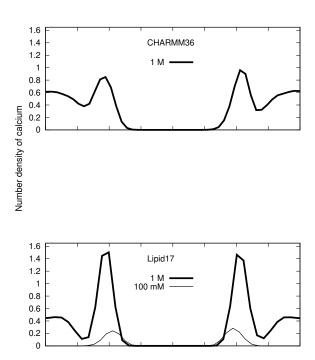


Figure S7: Calcium ion density profiles along membrane normal from simulations of POPC:POPG (4:1) mixtures with different force fields.

#### References

- (1) Papadopoulos, C.; Fuchs, P. F. CHARMM36 pure POPC MD simulation (300 K 300ns 1 bar). 2018; https://doi.org/10.5281/zenodo.1306800.
- (2) Papadopoulos, C.; Fuchs, P. F. CHARMM36 POPC/POPE (50%-50%) MD simulation (300 K 300ns 1 bar). 2018; https://doi.org/10.5281/zenodo.1306821.
- (3) Piggot, T. CHARMM36-UA POPE Simulations (versions 1 and 2) 310 K (NOTE: hexagonal membrane and POPE is called PEUA). 2018; https://doi.org/10.5281/zenodo.1293774.
- (4) Piggot, T. Slipids POPE Simulations (versions 1 and 2) 310 K (NOTE: hexagonal membrane). 2018; https://doi.org/10.5281/zenodo.1293813.
- (5) Favela-Rosales, F. MD simulation trajectory of a fully hydrated DPPE bilayer: SLIPIDS, Gromacs 5.0.4. 2017. 2017; https://doi.org/10.5281/zenodo.495247.
- (6) Favela-Rosales, F. MD simulation trajectory of a fully hydrated POPG bilayer: SLIPIDS, Gromacs 5.0.4. 2017. 2017; https://doi.org/10.5281/zenodo.546133.
- (7) Favela-Rosales, F. MD simulation trajectory of a fully hydrated DPPG bilayer @298K: SLIPIDS, Gromacs 5.0.4. 2017. 2017; https://doi.org/10.5281/zenodo.546135.
- (8) Favela-Rosales, F. MD simulation trajectory of a fully hydrated DPPG bilayer @314K: SLIPIDS, Gromacs 5.0.4. 2017. 2017; https://doi.org/10.5281/zenodo.546136.
- (9) Piggot, T. Berger POPE Simulations (versions 1 and 2) 303 K de Vries repulsive H. 2018; https://doi.org/10.5281/zenodo.1293889.
- (10) Piggot, T. Berger POPE Simulations (versions 1 and 2) 303 K larger repulsive H. 2018; https://doi.org/10.5281/zenodo.1293891.

- (11) Piggot, T. Berger DOPE Simulations (versions 1 and 2) 271 K de Vries repulsive H. 2018; https://doi.org/10.5281/zenodo.1293928.
- (12) Piggot, T. Berger DOPE Simulations (versions 1 and 2) 271 K larger repulsive H. 2018; https://doi.org/10.5281/zenodo.1293905.
- (13) AmÃllie, B.; F.J., F. P. Berger pure POPC MD simulation (300 K 300ns 1 bar). 2018; https://doi.org/10.5281/zenodo.1402417.
- (14) AmÃllie, B.; F.J., F. P. Berger POPC/POPE (50:50 ratio) MD simulation (300 K 400ns 1 bar). 2018; https://doi.org/10.5281/zenodo.1402449.
- (15) Piggot, T. GROMOS 43A1-S3 POPE Simulations (versions 1 and 2) 313 K (NOTE: anisotropic pressure coupling). 2018; https://doi.org/10.5281/zenodo.1293762.
- (16) Piggot, T. OPLS-UA POPE Simulations (versions 1 and 2) 303 K. 2018; https://doi.org/10.5281/zenodo.1293855.
- (17) Piggot, T. OPLS-UA POPE Simulations (versions 1 and 2) 303 K with vdW on H atoms. 2018; https://doi.org/10.5281/zenodo.1293853.
- (18) Piggot, T. GROMOS-CKP POPE Simulations (versions 1 and 2) 313 K. 2018; https://doi.org/10.5281/zenodo.1293932.
- (19) Piggot, T. GROMOS-CKP DOPE Simulations (versions 1 and 2) 271 K. 2018; https://doi.org/10.5281/zenodo.1293941.
- (20) Piggot, T. GROMOS-CKP DPPE Simulations (versions 1 and 2) 342 K. 2018; https://doi.org/10.5281/zenodo.1293957.
- (21) Melcr, J.; Martinez-Seara, H.; Nencini, R.; Kolafa, J.; Jungwirth, P.; Ollila, O. H. S. Accurate Binding of Sodium and Calcium to a POPC Bilayer by Effective Inclusion of Electronic Polarization. *The Journal of Physical Chemistry B* **2018**, *122*, 4546–4557.

- (22) Melcr, J.; Ferreira, T.; Jungwirth, P.; Ollila, O. H. S. Improved Cation Binding to Lipid Bilayer with Negatively Charged POPS by Effective Inclusion of Electronic Polarization. https://github.com/ohs0llila/ecc\_lipids/blob/master/Manuscript/manuscript.pdf, Submitted.
- (23) PEON, A. LIPID17 POPG Bilayer Simulation (Last 100 ns, 310 K). 2019; https://doi.org/10.5281/zenodo.3247659.
- (24) Pluhařová, E.; Fischer, H. E.; Mason, P. E.; Jungwirth, P. Hydration of the chloride ion in concentrated aqueous solutions using neutron scattering and molecular dynamics. *Mol. Phys.* 2014, 112, 1230–1240.
- (25) Kohagen, M.; Mason, P. E.; Jungwirth, P. Accounting for Electronic Polarization Effects in Aqueous Sodium Chloride via Molecular Dynamics Aided by Neutron Scattering. J. Phys. Chem. B 2016, 120, 1454–1460.
- (26) Martínek, T.; Duboué-Dijon, E.; Timr, Š.; Mason, P. E.; Baxová, K.; Fischer, H. E.; Schmidt, B.; Pluhařová, E.; Jungwirth, P. Calcium ions in aqueous solutions: Accurate force field description aided by ab initio molecular dynamics and neutron scattering. J. Chem. Phys. 2018, 148, 222813.
- (27) Borle, F.; Seelig, J. Ca2+ binding to phosphatidylglycerol bilayers as studied by differential scanning calorimetry and 2H- and 31P-nuclear magnetic resonance. *Chemistry and Physics of Lipids* **1985**, *36*, 263 283.
- (28) Macdonald, P. M.; Seelig, J. Calcium binding to mixed phosphatidylglycerol-phosphatidylcholine bilayers as studied by deuterium nuclear magnetic resonance.

  Biochemistry 1987, 26, 1231–1240.