

# NMRlipids III: Lipid-cholesterol interactions in atomistic resolution molecular dynamics simulations

Fernando Favela-Rosales,<sup>1</sup> Peter Heftberger,<sup>2</sup> Matti Javanainen,<sup>3,4</sup> Josef Melcr,<sup>5</sup>  
Markus Miettinen,<sup>6</sup> O. H. Samuli Ollila,<sup>5,7,\*</sup> Georg Pabst,<sup>2</sup> and Thomas Piggot<sup>8</sup>

<sup>1</sup>*Departamento de Física, Centro de Investigación y de Estudios Avanzados del IPN, Apartado Postal 14-740, 07000 México D.F., México*

<sup>2</sup>*University of Graz*

<sup>3</sup>*Department of Physics, Tampere University of Technology, Tampere, Finland*

<sup>4</sup>*University of Helsinki*

<sup>5</sup>*Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic, Prague 6, Czech Republic*

<sup>6</sup>*MPI*

<sup>7</sup>*Institute of Biotechnology, University of Helsinki*

<sup>8</sup>*University of Southampton*

(Dated: May 8, 2019)

The quantitative quality of lipid-cholesterol interactions in atomistic resolution models will be determined against NMR and scattering data.

## INTRODUCTION

Details of intermolecular interactions determine the phase behaviour and details lateral organization of lipid and bilayers containing cholesterol [1]. **1. Fix Ole's name among the authors of this paper.** Formation of lateral heterogeneities [2], lipid rafts [3] and superlattices [4] in cellular membranes have been suggested to be driven by interactions between cholesterol and lipids. While detailed experimental information of these interactions is relatively sparse, atomistic resolution molecular dynamics simulations have been widely applied to give detailed explanations of lipid bilayer lateral organization and lipid-cholesterol interactions [5, 6]. However, the simulations must reproduce the measured experimental details, such as NMR order parameters and scattering form factors, to be useful in interpretation of molecular details in lipid bilayer mixtures.

Simulations qualitatively reproduce the cholesterol condensation effect, but quantitative comparison to robust experimental data has not been typically done. This is partly due to the lack of availability of systematic experimental data sets for the comparison and partly due to the lack of protocols for validating intermolecular interactions in lipid bilayer simulations against experiments. Scattering experiments are more difficult to interpret for mixed lipid bilayers than for single component bilayers [7–9], thus area per molecule values, one of the main quantity used to compare simulations to experiments, have not been available for lipid cholesterol mixtures. Systematic experimental data set for lipid C-H bond order parameters with different cholesterol concentrations in POPC bilayer has been published only relatively recently [10].

In this work we present the experimental scattering form factor data for POPC-cholesterol mixtures by systematically increasing the cholesterol concentration, scanning the same concentrations that were previously studied with ssNMR [10]. Our goal is to show that the combination of systematically measured C-H bond order parameter and scattering form factor data can be used to validate the quality of lipid-cholesterol intermolecular interactions in MD simulations. MD simula-

tions can be also potentially used to give structural interpretation for form factors measured from mixed systems, which is a major challenge for current methods [7–9]. The approach should be also applicable for mixed lipid bilayers with other than lipid-cholesterol mixtures.

Order parameters for C-H bond vectors in lipid bilayer systems, measured with <sup>13</sup>C or <sup>2</sup>H NMR techniques, give indirect information about the structural sampling of individual molecules [11]. The observed increase of acyl chain order parameters with added cholesterol in simulations and experiments can be explained by increased trans conformations in acyl chains [10], which is suggested to play critical role in the phase behaviour of PC-lipid-cholesterol mixtures [1]. Consequently, the correct cholesterol ordering effect is expected to be a necessary condition for a model used to understand lipid-cholesterol phase behaviour.

## METHODS

### X-ray scattering experiments

SAXS data on POPC multilamellar vesicles (MLVs) at various cholesterol concentrations has been measured. Data have been obtained at the EMBL BioSAXS beamline (Hamburg) using 20 keV photons, T = 27°C. Data were analyzed in terms of the SDP-GAP model described in Heftberger et al., J. Appl. Cryst. 2013 and Heftberger et al. Biophys. J. 2015. Data from MLVs are a convolute of structure factor (the crystalline lattice) and form factor. By fitting the scattered intensity data we obtain both contributions. Here we posted only form factors (ASCII format). For information on the quality of the fit we also give plots of the fitted intensity data. The electron density profile has been modelled in terms of the SDP model (see papers by Kucerka and coworkers), that is volume distribution functions are modelled by individual Gaussians or error functions. Cholesterol is also accounted for by two Gaussians. This model has been proposed by Jianjun Pan (USF, Tampa,

FL), but is to the best of our knowledge not published (see also PhD Thesis by Peter Heftberger). Additional figures show the volume distribution functions and the resulting electron density profiles.

Authors to consult and potentially include in publications using this data: Peter Heftberger (peter.heftberger@gmx.at), Georg Pabst (georg.pabst@uni-graz.at)

### **Molecular dynamics simulations**

Molecular dynamics simulation data was collected using the open collaboration method [12]. Simulated systems are listed in Table I and the full simulation details are given in references or in supplementary material.

TABLE I: Simulated lipid bilayers containing cholesterol. The simulation file data sets marked with \* include also part of the trajectory. <sup>a</sup> The number of lipid molecules <sup>b</sup> The number of cholesterol molecules <sup>c</sup> Cholesterol concentration (mol%) <sup>d</sup> The number of water molecules <sup>e</sup> Simulation temperature <sup>f</sup> The total simulation time <sup>g</sup> Time frames used in the analysis <sup>h</sup> Reference link for the downloadable simulation files <sup>i</sup> Reference for the full simulation details

Force field	lipid	<sup>a</sup> N <sub>l</sub>	<sup>b</sup> N <sub>chol</sub>	<sup>c</sup> C <sub>CHOL</sub>	<sup>d</sup> N <sub>w</sub>	<sup>e</sup> T (K)	<sup>f</sup> t <sub>sim</sub> (ns)	<sup>g</sup> t <sub>anal</sub> (ns)	<sup>h</sup> Files	<sup>i</sup> Details
Berger-POPC-07 [13]	POPC	128	0	0%	7290	298	270	240	[14]*	[15]
/Höltje-CHOL-13 [10, 16]										
	POPC	120	8	6%	7290	298	100	80	[17]*	[10]
	POPC	110	18	14%	8481	298	100	80	[18]*	[10]
	POPC	84	44	34%	6794	298	100	80	[19]*	[10]
	POPC	64	64	50%	10314	298	100	80	[20]*	[10]
	POPC	50	78	61%	5782	298	100	80	[21]*	[10]
CHARMM36[22, 23]	POPC	200	0	0%	9000	310	?	100	[24]*	SI
	POPC	200	22	10%	9000	310	?	100	[24]*	SI
	POPC	200	50	20%	9000	310	?	100	[24]*	SI
	POPC	200	86	30%	9000	310	?	100	[24]*	SI
	POPC	200	134	40%	15030	310	109	100	[25]*	SI
	POPC	200	200	50%	18000	310	109	100	[25]*	SI
Slipids[26–28]	POPC	200	0	0%	?	310	?	100	[29]*	SI
	POPC	512	0	0%	23943	310	170	100	[30]*	SI
	POPC	200	22	10%	?	310	?	100	[29]*	SI
	POPC	200	50	20%	?	310	?	100	[29]*	SI
	POPC	200	86	30%	?	310	?	100	[29]*	SI
	POPC	358	154	30%	21183	298	170	100	[31]*	SI
	POPC	200	134	40%	?	310	109	100	[32]*	SI
	POPC	200	200	50%	?	310	109	100	[32]*	SI
	POPC	256	256	50%	20334	298	170	100	[33]*	SI
MacRog[34]	POPC	128	0	0%	6400	310	400	200	[35]*	[12]
	POPC	114	14	11%	6400	310	400	200	[35]*	[12]
	POPC	72	56	44%	6400	310	400	200	[35]*	[12]
	POPC	64	64	50%	6400	310	400	200	[35]*	[12]
	POPC	56	72	56%	6400	310	400	200	[35]*	[12]

## RESULTS AND DISCUSSION

### Monitoring molecular interactions between POPC and cholesterol in lipid bilayer using NMR and MD simulations

Addition of cholesterol substantially increases the acyl chain C-H order parameters in PC lipid bilayers (Fig. 1), which can be explained by increased trans conformations in acyl chains [10? ]. The absolute values of order parameters of *sn*-1 chain from  $^{13}\text{C}$  NMR experiments [10] exhibit approximately linear increase with cholesterol up to equimolar mixture (Fig. S2), which is expected because phase separation is not observed for this mixture [10, 36].

Experimental acyl chain order parameters of pure POPC lipid bilayers are well reproduced by most force fields (Fig. 1), except for the  $\text{C}_2$  carbon in *sn*-1 chain, which is typically the case in the state of the art lipid force fields (for review see [11]). However, the order parameters in the beginning of *sn*-1 chain are underestimated in the MACROG simulation and CHARMM36 simulations slightly overestimates the order parameters, also when used with other than Gromacs simulation packages (section S1 in the supplementary information). The absolute values of *sn*-1 acyl chain order parameters exhibit approximately linear increase upon addition of cholesterol also in simulations below equimolar mixture (Figs. S3-S7). However, the slope of the increase (Fig. 2) and order parameters in equimolar mixtures (Fig. 1) are overestimated by all force fields. The overestimation of order parameters observed in CHARMM without cholesterol and in all force fields with cholesterol is smaller than the contribution by undulations in large simulations (section S3 in the supplementary information).

3.Also experimental cholesterol order parameters are available. Maybe these should be calculated from simulations as well.

### Lipid bilayer dimensions and density profiles as a function of cholesterol

Lipid bilayer dimensions can be experimentally accessed by measuring X-ray scattering form factor, which is related to the electron density along membrane normal via Fourier transformation [7–9, 11? ]. The form factor can be translated to density profiles, area per lipids and bilayer thickness using SDP model or its combination with MD simulations [7? –9]. Sophisticated approaches for single component lipid bilayers are available [? ], but multicomponent systems are more difficult to interpret [7? –9].

State of the art force fields give good agreement with experimental form factors for pure POPC lipid bilayers [? ], which is the case also here, except for the Berger simulation (Fig. 3). Upon addition of cholesterol, the third minima in form factor systemically decreases from  $\sim 0.42$  to  $\sim 0.32$ . This may be related to the reduced area per molecule upon addition of cholesterol (Fig. 4). The area per total amount of molecules decreases with cholesterol in all simulations, partly due to the

smaller area covered by the cholesterol than lipids, but also because lipids ordered by cholesterol require less space. Due to the latter effect, the area per PC headgroup does not essentially increase up to the addition of  $\sim 15$  mol% of cholesterol.

Visual inspection of form factors (Fig. 3) and quantitative quality estimation (Fig. 5) suggests that the Berge/Holtje force field gives the best agreement with experiments with large cholesterol concentration. This is surprising because the same model overestimated acyl chain order parameter increase upon addition of cholesterol more than CHARMM36 or Slipids simulations (Figs. 1 and 2). The Berger/Holtje model also exhibits the most pronounced decrease in the area per molecule upon addition of cholesterol (Fig. ??). On the other hand, the electron density profiles from Berger/Holtje simulations are closer to the result from the SDP model than other MD simulation models for both pure POPC bilayer and equimolar POPC/cholesterol mixture, especially in the middle of the bilayer where SDP model gives substantially lower density than MD simulations (Fig. 6). Therefore, the good quality of form factor Berger/Holtje model with large amounts of cholesterol may rather indicate the importance of this low electron density in the bilayer center, rather than better quality of lipid-cholesterol interactions in this model.

## PRESENTING THE QUALITY OF LIPID BILAYER MD SIMULATION WITH BINARY MIXTURES

### CONCLUSIONS

Cholesterol ordering effect is overestimated in Berger/Holtje and MacRog models. Slight overestimation is observed also in CHARMM36 and Slipid models, but more careful analysis is required to conclude if this is significant or not.

---

\* samuli.ollila@helsinki.fi

- [1] J. H. Ipsen, G. Karlström, O. Mourtsen, H. Wennerström, and M. Zuckermann, *Biochim. Biophys. Acta* **905**, 162 (1987).
- [2] P. K. Kinnunen, *Chemistry and Physics of Lipids* **57**, 375 (1991).
- [3] K. Simons and E. Ikonen, *Nature* **387**, 569 (1997).
- [4] P. Somerharju, J. A. Virtanen, K. H. Cheng, and M. Hermansson, *Biochim. Biophys. Acta - Biomembranes* **1788**, 12 (2009).
- [5] T. Róg and I. Vattulainen, *Chem. Phys. Lipids* **184**, 82 (2014).
- [6] A. J. Sodt, M. L. Sandar, K. Gawrisch, R. W. Pastor, and E. Lyman, *J. Am. Chem. Soc.* **136**, 725 (2014).
- [7] J. Pan, X. Cheng, F. A. Heberle, B. Mostofian, N. Kučerka, P. Drazba, and J. Katsaras, *J. Phys. Chem. B* **116**, 14829 (2012).
- [8] P. Heftberger, B. Kollmitzer, A. Rieder, H. Amenitsch, and G. Pabst, *Biophys. J.* **108**, 854 (2015), ISSN 0006-3495.
- [9] D. Marquardt, F. A. Heberle, J. D. Nickels, G. Pabst, and J. Katsaras, *Soft Matter* **11**, 9055 (2015).

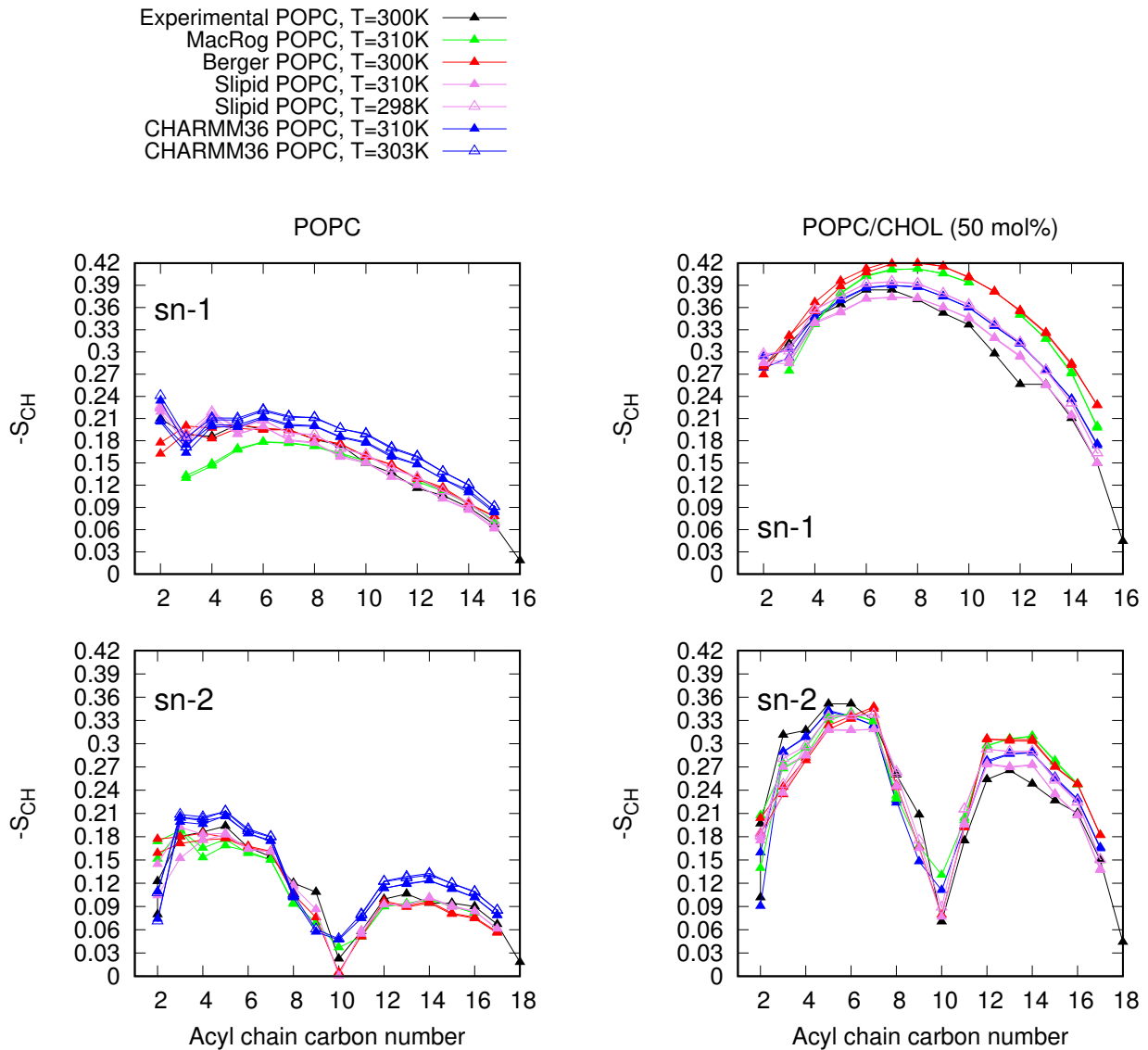


FIG. 1: Order parameters from simulations and experiments for acyl chains of 1-palmitoyl-2-oleoylphosphatidylcholine (POPC).

## 2.Lipid14 results?

- [10] T. M. Ferreira, F. Coreta-Gomes, O. H. S. Ollila, M. J. Moreno, W. L. C. Vaz, and D. Topgaard, *Phys. Chem. Chem. Phys.* **15**, 1976 (2013).
- [11] O. S. Ollila and G. Pabst, *Biochimica et Biophysica Acta (BBA) - Biomembranes* **1858**, 2512 (2016), biosimulations of lipid membranes coupled to experiments.
- [12] A. Botan, F. Favela-Rosales, P. F. J. Fuchs, M. Javanainen, M. Kanduć, W. Kulig, A. Lamberg, C. Loison, A. Lyubartsev, M. S. Miettinen, et al., *J. Phys. Chem. B* **119**, 15075 (2015).
- [13] S. Ollila, M. T. Hyvönen, and I. Vattulainen, *J. Phys. Chem. B* **111**, 3139 (2007).
- [14] O. H. S. Ollila, T. Ferreira, and D. Topgaard, *MD simulation trajectory and related files for POPC bilayer (Berger model delivered by Tieleman, Gromacs 4.5)* (2014), URL <http://dx.doi.org/10.5281/zenodo.13279>.
- [15] T. M. Ferreira, O. H. S. Ollila, R. Pigliapochi, A. P. Dabkowska, and D. Topgaard, *J. Chem. Phys.* **142**, 044905 (2015).
- [16] M. Höltje, T. Förster, B. Brandt, T. Engels, W. von Rybinski, and H.-D. Höltje, *Biochim. Biophys. Acta* **1511**, 156 (2001).
- [17] O. H. S. Ollila, T. Ferreira, and D. Topgaard, *MD simulation trajectory and related files for POPC/cholesterol (7 mol%) bilayer (Berger model delivered by Tieleman, modified Höltje, Gromacs 4.5)* (2014), URL <http://dx.doi.org/10.5281/zenodo.13282>.
- [18] O. H. S. Ollila, T. Ferreira, and D. Topgaard, *MD simulation trajectory and related files for POPC/cholesterol (15 mol%) bilayer (Berger model delivered by Tieleman, modified Höltje, Gromacs 4.5)* (2014), URL <http://dx.doi.org/10.5281/zenodo.13281>.
- [19] O. H. S. Ollila, T. Ferreira, and D. Topgaard, *MD simulation trajectory and related files for POPC/cholesterol (34 mol%) bilayer (Berger model delivered by Tieleman, modified Höltje, Gromacs 4.5)* (2014), URL <http://dx.doi.org/10.5281/zenodo.13283>.
- [20] O. H. S. Ollila, T. Ferreira, and D. Topgaard, *MD simulation trajectory and related files for POPC/cholesterol (50 mol%) bi-*

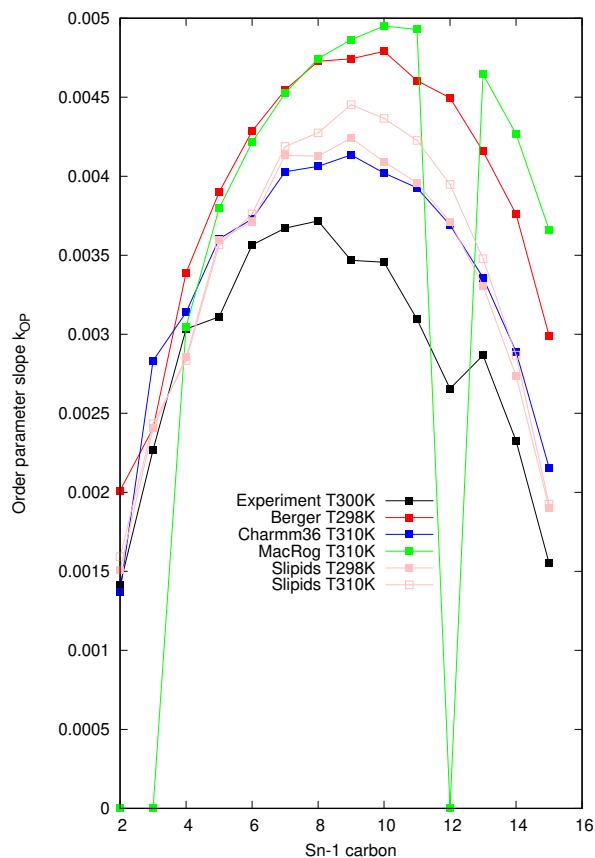


FIG. 2: Slopes of order parameters as a function of cholesterol.

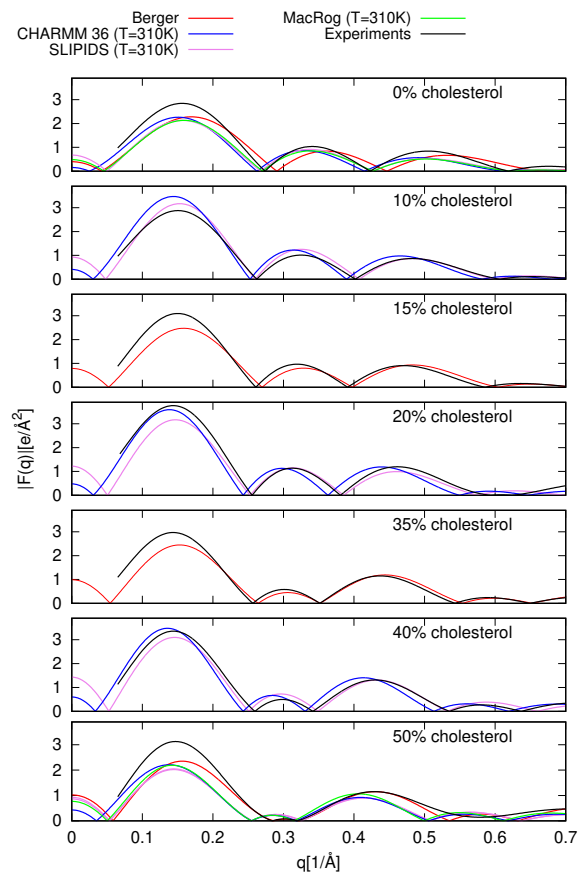


FIG. 3: Form factors from simulations and experiments.

- layer (Berger model delivered by Tieleman, modified Höltje, Gromacs 4.5) (2014), URL {<http://dx.doi.org/10.5281/zenodo.13285>}.
- [21] O. H. S. Ollila, T. Ferreira, and D. Topgaard, *MD simulation trajectory and related files for POPC/cholesterol (60 mol%) bi-layer (Berger model delivered by Tieleman, modified Höltje, Gromacs 4.5)* (2014), URL {<http://dx.doi.org/10.5281/zenodo.13286>}.
- [22] J. B. Klauda, R. M. Venable, J. A. Freites, J. W. O'Connor, D. J. Tobias, C. Mondragon-Ramirez, I. Vorobyov, A. D. MacKerell Jr, and R. W. Pastor, *J. Phys. Chem. B* **114**, 7830 (2010).
- [23] J. B. Lim, B. Rogaski, and J. B. Klauda, *J. Phys. Chem. B* **116**, 203 (2012).
- [24] M. Javanainen, *POPC with 0, 10, 20, and 30 mol-310 K. Charmm36 force field.* (2016), URL <https://doi.org/10.5281/zenodo.159759>.
- [25] M. Javanainen, *POPC with 40 and 50 mol-Slipids force field.* (2016), URL <https://doi.org/10.5281/zenodo.154346>.
- [26] J. P. M. Jämbeck and A. P. Lyubartsev, *J. Chem. Theory Comput.* **8**, 2938 (2012).
- [27] J. P. M. Jämbeck and A. P. Lyubartsev, *J. Phys. Chem. B* **116**, 3164 (2012).
- [28] J. P. M. Jämbeck and A. P. Lyubartsev, *Journal of Chemical Theory and Computation* **9**, 774 (2013).
- [29] M. Javanainen, *POPC with 0, 10, 20, and 30 mol-310 K. Slipids force field.* (2016), URL <https://doi.org/10.5281/zenodo.60607>.

- 4.Details about form factor calculation code is discussed in issues <https://github.com/NMRLipids/MATCH/issues/56> and <https://github.com/NMRLipids/MATCH/issues/50>. Once the code is finalized, we should recalculate and check the form factors. CURRENTLY, 10%-40% cholesterol concentrations are calculated with a code which gives possibly incorrect heights for the maxima.
- 5.The y-axis scale cannot be explicitly measured in experiments. Therefore, the y-axis of the experimental form factor is typically scaled to match with simulations. This currently not done, because we have several simulations which give inequal form factors, and therefore it is not clear against which simulation results we should scale the experimental results. I have created a issue for this discussion: <https://github.com/NMRLipids/NmrLipidsCholXray/issues/18>
- 6.Not all experimental and simulation data is here (maybe?).
- [30] F. Favela-Rosales, *MD simulation trajectory of a lipid bilayer: Pure POPC in water. SLIPIDS, Gromacs 4.6.3. 2016.* (2016), URL <https://doi.org/10.5281/zenodo.166034>.
- [31] F. Favela-Rosales, *MD simulation trajectory of a lipid bi-layer: 70/30 mol% POPC/Cholesterol . SLIPIDS, Gromacs 4.6.3. 2016.* (2016), URL <https://doi.org/10.5281/zenodo.62026>.

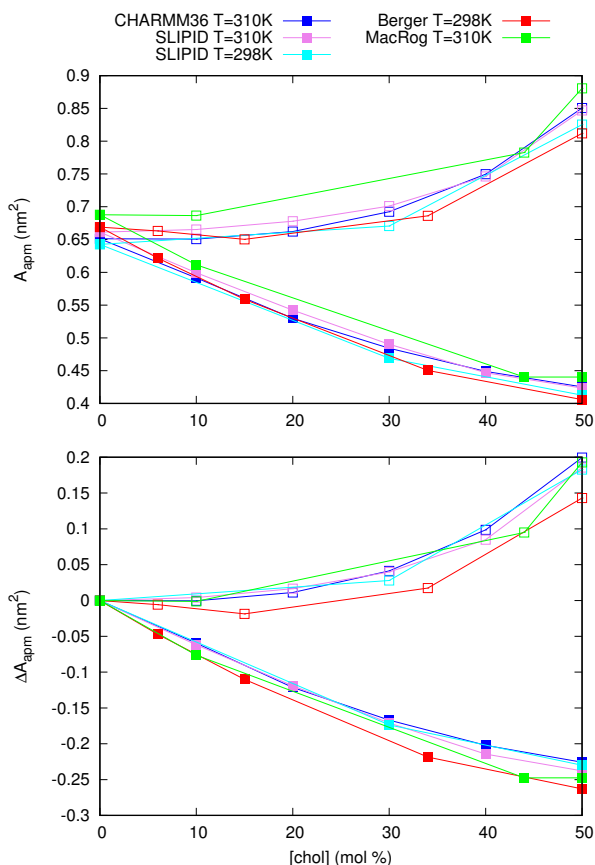
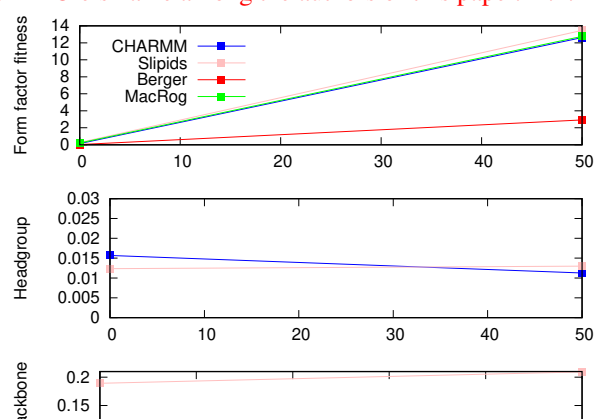


FIG. 4: Area per molecules calculated from different simulation models as a function of cholesterol concentration. The solid symbols are area per total amount of molecules (chol+PC) and the empty symbols area per PC headgroups. Top figure shows absolute values and bottom figure shows changes respect to pure lipid system.

- [32] M. Javanainen, *POPC with 40 and 50 mol-Slipids force field*. (2016), URL <https://doi.org/10.5281/zenodo.154346>.
- [33] F. Favela-Rosales, *MD simulation trajectory of a lipid bilayer: 50/50 mol% POPC/Cholesterol . SLIPIDS, Gromacs 4.6.3. 2016*. (2016), URL <https://doi.org/10.5281/zenodo.159434>.
- [34] W. Kulig, M. Pasenkiewicz-Gierula, and T. Róg, *Chem. Phys. Lipids* **195**, 12 (2016).
- [35] M. Javanainen, *POPC/Cholesterol @ 310K. 0, 10, 40, 50 and 60 mol-cholesterol. Model by Maciejewski and Róg* (2015), URL <http://dx.doi.org/10.5281/zenodo.13877>.
- [36] I. Ionova, V. Livshits, and D. Marsh, *Biophysical Journal* **102**, 1856 (2012).

## ToDo

1. Fix Ole's name among the authors of this paper. . . . . 1



3. Also experimental cholesterol order parameters are available. Maybe these should be calculated from simulations as well. . . . . 4

2. Lipid14 results? . . . . . 5

4. Details about form factor calculation code is discussed in issues <https://github.com/NMRLipids/MATCH/issues/56> and <https://github.com/NMRLipids/MATCH/issues/50>. Once the code is finalized, we should recalculate and check the form factors. CURRENTLY, 10%-40% cholesterol concentrations are calculated with a code which gives possibly incorrect heights for the maxima. 6

5. The y-axis scale cannot be explicitly measured in experiments. Therefore, the y-axis of the experimental form factor is typically scaled to match with simulations. This currently not done, because we have several simulations which give unequal form factors, and therefore it is not clear against which simulation results we should scale the experimental results. I have created a issue for this discussion: <https://github.com/NMRLipids/NmrLipidsCholXray/issues/18> 6

6. Not all experimental and simulation data is here (maybe?). . . . . 6

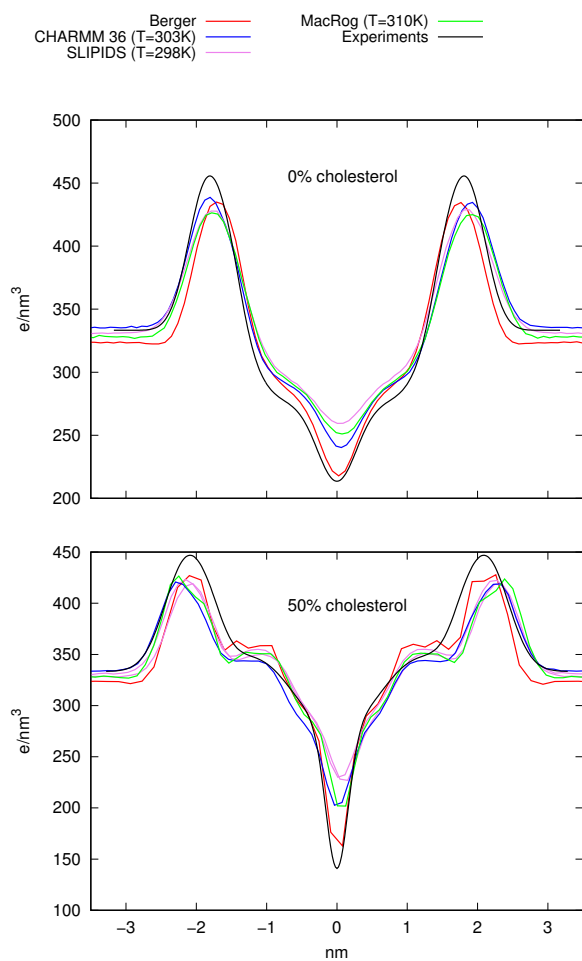


FIG. 6: Electron density profiles from MD simulations and SDP model.



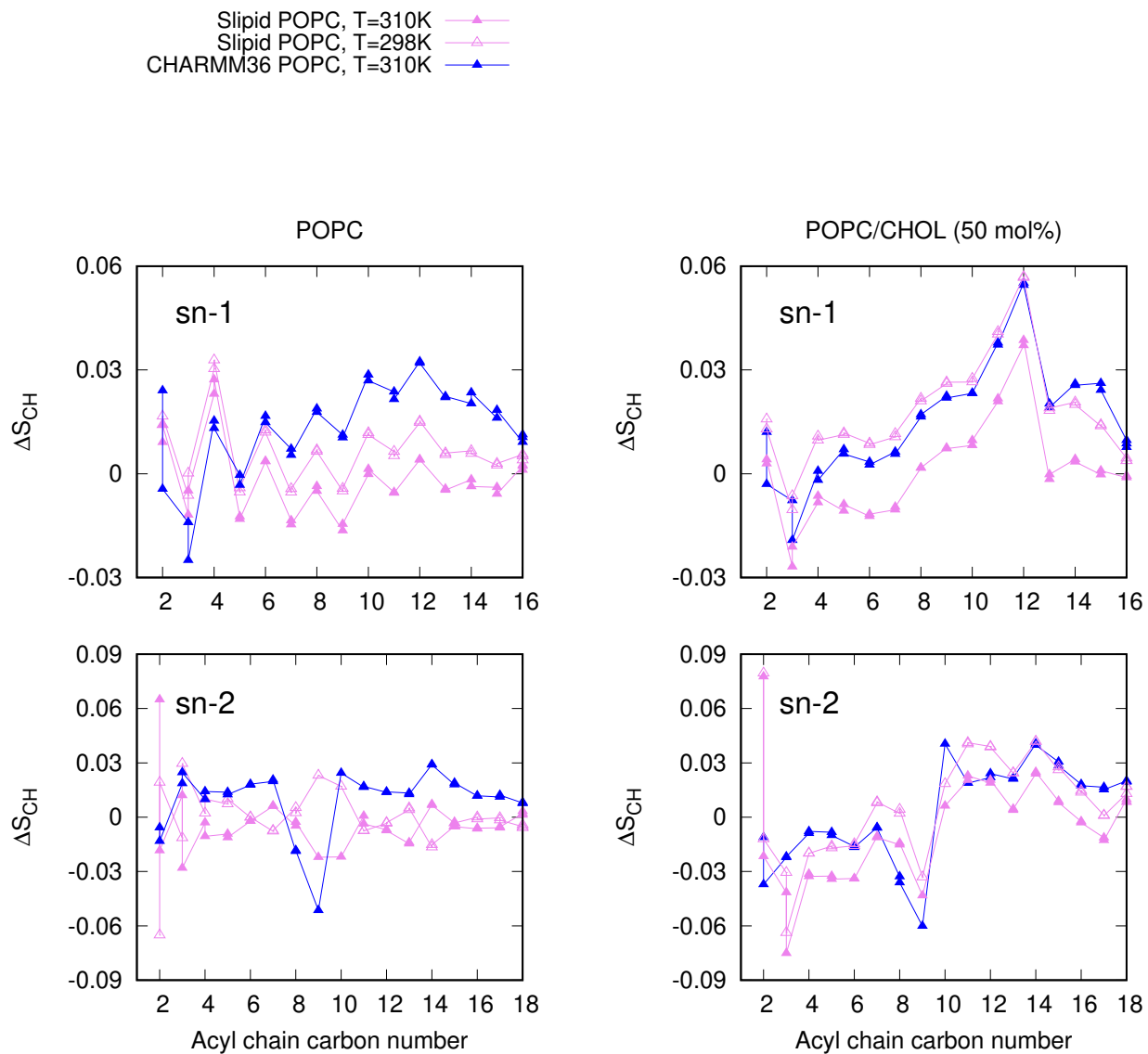


FIG. 7: Difference between order parameters from simulations and experiments for acyl chains of 1-palmitoyl-2-oleoylphosphatidylcholine (POPC).