



# SUPPLEMENTARY INFORMATION FOR

## NMRlipids III: Lipid–Cholesterol Interactions in

### Atomistic Resolution Molecular Dynamics

### Simulations

Fernando Favela-Rosales,<sup>†</sup> Peter Heftberger,<sup>‡</sup> Matti Javanainen,<sup>\*,¶,§</sup> Jesper J. Madsen,<sup>||,⊥</sup> Josef Melcr,<sup>¶</sup> Markus Miettinen,<sup>#,@</sup> O. H. Samuli Ollila,<sup>\*,¶,§</sup> Georg Pabst,<sup>‡,△</sup> and Thomas Piggot<sup>∇,††</sup>

<sup>†</sup>*Departamento de Física, Centro de Investigación y de Estudios Avanzados del IPN,  
Apartado Postal 14-740, 07000 México D.F., México*

<sup>‡</sup>*Institute of Molecular Biosciences, Biophysics Division, NAWI Graz, University of Graz,  
Graz 8010, Austria*

<sup>¶</sup>*Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech  
Republic, Prague 6, Czech Republic*

<sup>§</sup>*Institute of Biotechnology, University of Helsinki*

<sup>||</sup>*Department of Global Health, College of Public Health*

<sup>⊥</sup>*University of South Florida*

<sup>#</sup>*Department of Chemistry, University of Bergen, Norway*

<sup>@</sup>*Computational Biology Unit, Department of Informatics, University of Bergen, Norway*

<sup>△</sup>*BioTechMed-Graz, Graz 8010, Austria*

<sup>∇</sup>*School of Chemistry, University of Southampton, Southampton SO17 1BJ, United  
Kingdom*

<sup>††</sup>*Chemical Biological and Radiological Sciences, Defence Science and Technology Laboratory,  
Porton Down, Salisbury, Wiltshire SP4 0JQ, United Kingdom*

E-mail: matti.javanainen@helsinki.fi; samuli.ollila@helsinki.fi

# Contents

<b>S1 Simulation Details</b>	<b>S3</b>
<b>S2 Additional Results</b>	<b>S6</b>
S2.1 Scattering Intensities . . . . .	S6
S2.2 Form Factors & Electron Density Profiles . . . . .	S8
S2.3 Order Parameters . . . . .	S10
S2.4 Dynamic Properties . . . . .	S13
S2.5 Finite-Size Effects . . . . .	S15
<b>References</b>	<b>S17</b>

## S1 Simulation Details

The force field specific simulation parameters used for each force field are listed in Table S1. The parameter input files (`.mdp`) are provided in the Zenodo portal (links in the main text). For all simulations, we used an integration time step of 2 fs and the leap-frog integrator of GROMACS. The isothermal–isobaric (NPT) ensemble was used with a temperature of 298 K and a pressure of 1 bar. All simulations were 1  $\mu$ s long, and trajectories were written every 1 ns. The P-LINCS constraint algorithm<sup>1,2</sup> was used for the bonds noted in Table S1. NMRlipids databank<sup>3</sup> ID numbers of simulations are given in table S2.

Table S1: Simulation parameters used for different force fields.

Parameter	CHARMM36	Slipids	Lipid17	MacRog
Neighbour list type	Verlet <sup>4</sup>	Verlet <sup>4</sup>	Verlet <sup>4</sup>	Verlet <sup>4</sup>
Long-range electrostatics	PME <sup>5,6</sup>	PME <sup>5,6</sup>	PME <sup>5,6</sup>	PME <sup>5,6</sup>
LJ cut-off	1.2 nm	1.4 nm	0.9 nm	1.0 nm
LJ modifier	Force switch 1.0–1.2 nm	–	–	–
Dispersion correction	–	Energy & pressure <sup>7</sup>	Energy & pressure <sup>7</sup>	Energy & pressure <sup>7</sup>
Thermostat	Nosé–Hoover <sup>8,9</sup>	Stochastic rescaling <sup>10</sup>	Nosé–Hoover <sup>8,9</sup>	Stochastic rescaling <sup>10</sup>
Time constant ( $T$ )	1 ps	0.5 ps	1 ps	0.1 ps
Coupling groups	Lipids & water	Lipids & water	Lipids & water	Lipids & water
Barostat	Parrinello–Rahman <sup>11</sup>	Berendsen <sup>12</sup>	Parrinello–Rahman <sup>11</sup>	Parrinello–Rahman <sup>11</sup>
Coupling type ( $P$ )	semi-isotropic	semi-isotropic	semi-isotropic	semi-isotropic
Time constant ( $P$ )	5 ps	10 ps	5 ps	4 ps
Compressibility	$4.5 \cdot 10^{-5}$ 1/bar	$4.5 \cdot 10^{-5}$ 1/bar	$4.5 \cdot 10^{-5}$ 1/bar	$4.5 \cdot 10^{-5}$ 1/bar
Constraints	Bonds with H	All bonds	Bonds with H	All bonds
Water model	TIP3P <sup>13</sup>	TIP3P <sup>14</sup>	TIP3P <sup>14</sup>	TIP3P <sup>14</sup>
FF Source	CHARMM-GUI	CHARMM-GUI	CHARMM-GUI	Refs. 15 & 16
Parameter source	CHARMM-GUI	Slipids website	CHARMM-GUI	Refs. 15 & 16

**Table S2: NMRlipids databank IDs of performed simulations.**

CHARMM36		Slipids	
System	ID	System	ID
1024POPC_128CHOL_57600SOL_298K	426	Slipids_POPC_CHOL20_L	691
64POPC_3200SOL_298K	546	Slipids_POPC_CHOL47_M	712
1024POPC_896CHOL_96000SOL_298K	543	Slipids_POPC_CHOL29_S	693
64POPC_40CHOL_5200SOL_298K	620	Slipids_POPC_CHOL20_S	692
64POPC_56CHOL_6000SOL_298K	91	Slipids_POPC_CHOL20_M	687
1024POPC_416CHOL_72000SOL_298K	412	Slipids_POPC_CHOL11_S	672
1024POPC_256CHOL_64000SOL_298K	88	Slipids_POPC_CHOL11_M	681
256POPC_64CHOL_16000SOL_298K	72	Slipids_POPC_L	696
64POPC_8CHOL_3600SOL_298K	109	Slipids_POPC_CHOL38_S	661
256POPC_104CHOL_18000SOL_298K	298	Slipids_POPC_M	708
64POPC_16CHOL_4000SOL_298K	525	Slipids_POPC_CHOL38_L	670
64POPC_26CHOL_4500SOL_298K	393	Slipids_POPC_S	664
1024POPC_640CHOL_83200SOL_298K	550	Slipids_POPC_CHOL29_L	709
256POPC_160CHOL_20800SOL_298K	275	Slipids_POPC_CHOL29_M	659
256POPC_224CHOL_24000SOL_298K	166	Slipids_POPC_CHOL11_L	697
256POPC_32CHOL_14400SOL_298K	119	Slipids_POPC_CHOL38_M	668
		Slipids_POPC_CHOL47_S	703
		Slipids_POPC_CHOL47_L	682
Lipid17		MacRog	
System	ID	System	ID
Lipid17_POPC_CHOL29_M	700	MacRog_POPC_CHOL38_S	716
Lipid17_POPC_CHOL47_L	717	MacRog_POPC_CHOL29_L	705
Lipid17_POPC_CHOL11_L	663	MacRog_POPC_CHOL47_M	699
Lipid17_POPC_CHOL29_L	683	MacRog_POPC_CHOL47_L	671
Lipid17_POPC_CHOL20_S	662	MacRog_POPC_CHOL20_L	677
Lipid17_POPC_CHOL47_M	714	MacRog_POPC_S	674
Lipid17_POPC_CHOL38_S	688	MacRog_POPC_CHOL20_M	706
Lipid17_POPC_L	684	MacRog_POPC_CHOL38_L	698
Lipid17_POPC_S	715	MacRog_POPC_CHOL38_M	679
Lipid17_POPC_CHOL20_M	666	MacRog_POPC_M	675
Lipid17_POPC_M	657	MacRog_POPC_L	658
Lipid17_POPC_CHOL29_S	686	MacRog_POPC_CHOL29_M	655
Lipid17_POPC_CHOL38_L	673	MacRog_POPC_CHOL11_L	660
Lipid17_POPC_CHOL47_S	707	MacRog_POPC_CHOL11_S	665
Lipid17_POPC_CHOL11_S	680	MacRog_POPC_CHOL29_S	690
Lipid17_POPC_CHOL20_L	667	MacRog_POPC_CHOL11_M	695
Lipid17_POPC_CHOL38_M	694	MacRog_POPC_CHOL47_S	704
Lipid17_POPC_CHOL11_M	689	MacRog_POPC_CHOL20_S	702

## S2 Additional Results

### S2.1 Scattering Intensities

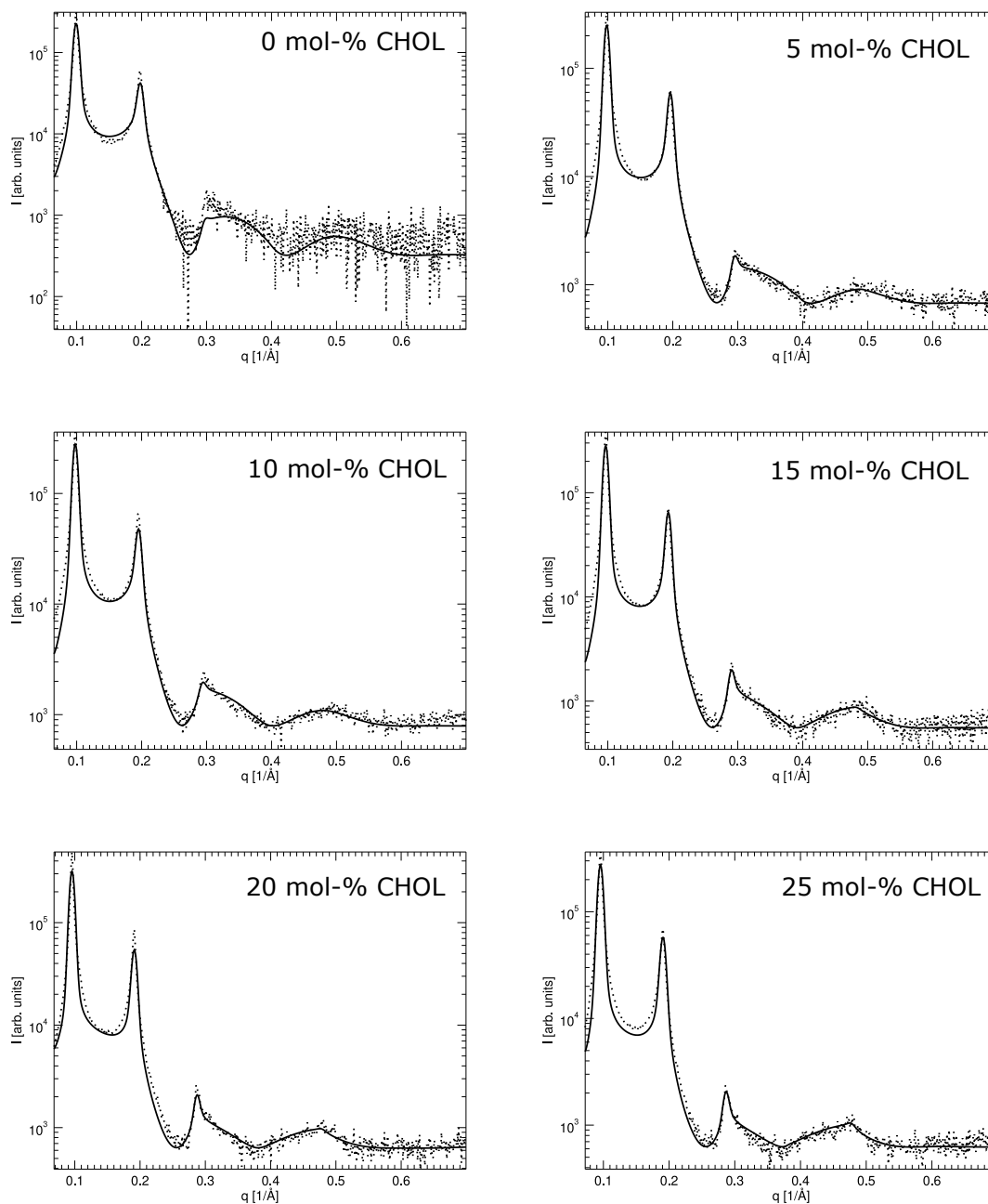


Figure S1: Scattering intensities from X-ray scattering experiments with various concentrations of cholesterol. More data are shown in the next figure.

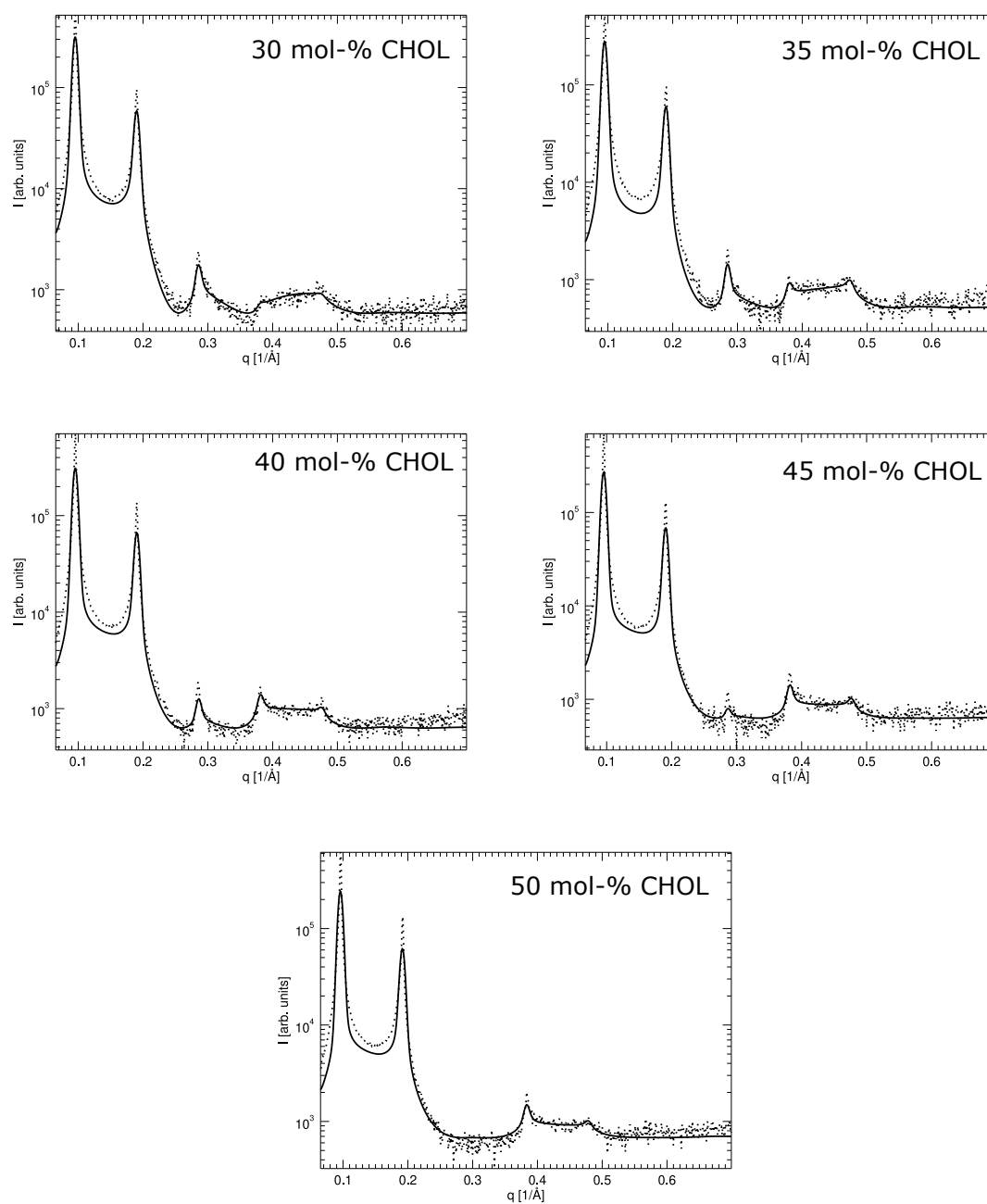


Figure S2: Scattering intensities from X-ray scattering experiments with various concentrations of cholesterol. More data are shown in the previous figure.

## S2.2 Form Factors & Electron Density Profiles

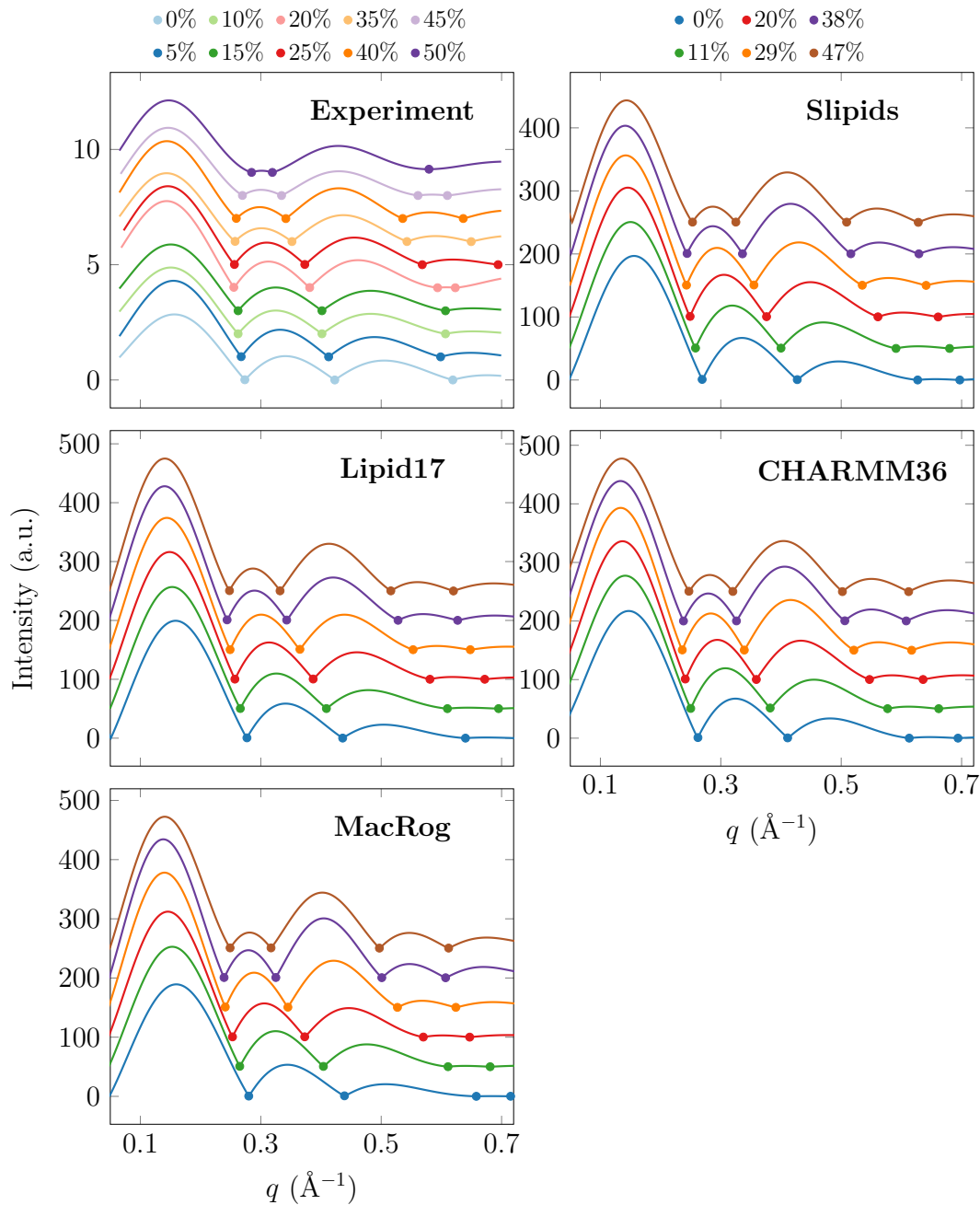


Figure S3: **Scattering intensities as a function of scattering vector from experiments and simulations.** Each of the profiles is shifted vertically with respect to the previous one, by 1 for the experimental profiles and by 50 for the computational ones. The minima are marked by filled circles to guide the eye.



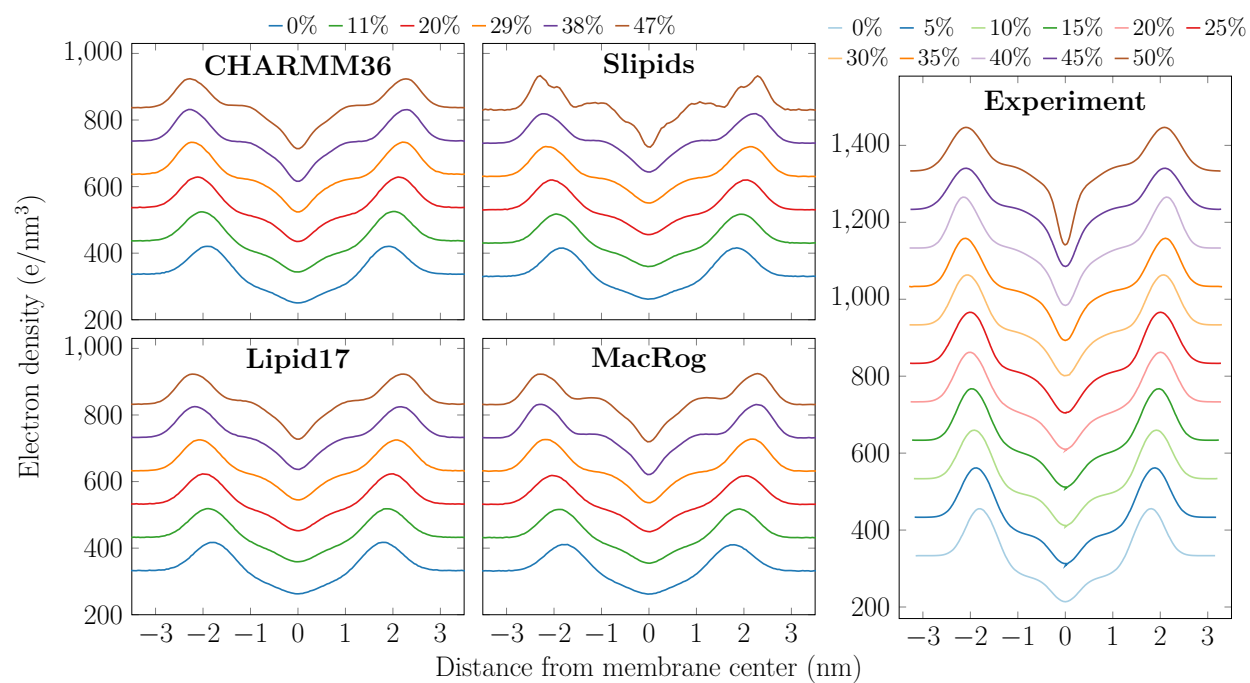


Figure S4: **Electron density profiles.** Each of the profiles is shifted vertically with respect to the previous one by 100 units.

## S2.3 Order Parameters

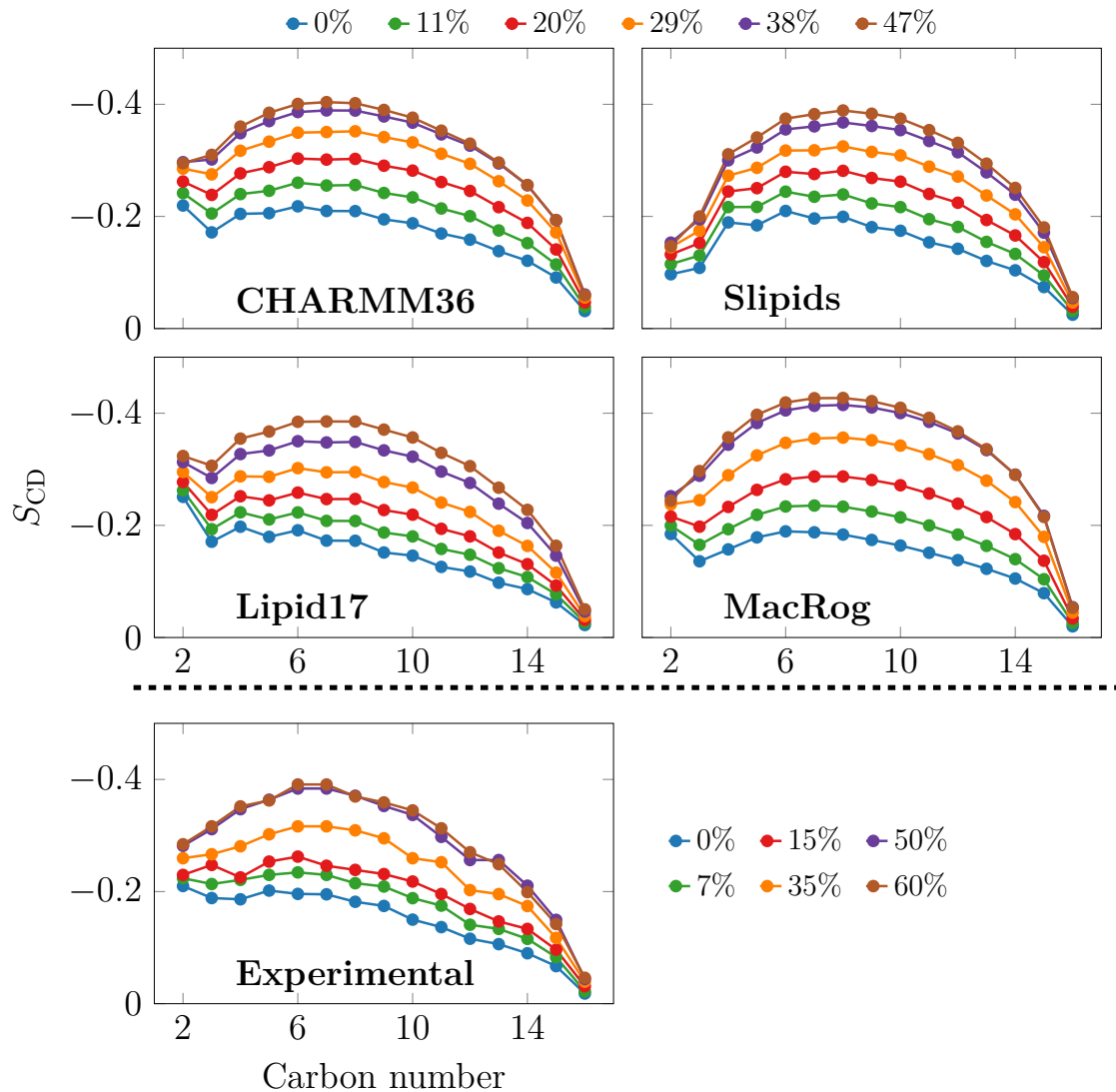


Figure S5: **Effect of cholesterol on the acyl chain order parameters of the POPC *sn*-1 (palmitate) chain.** The legend at the top corresponds to all simulations, and the one at the bottom to the experiments.

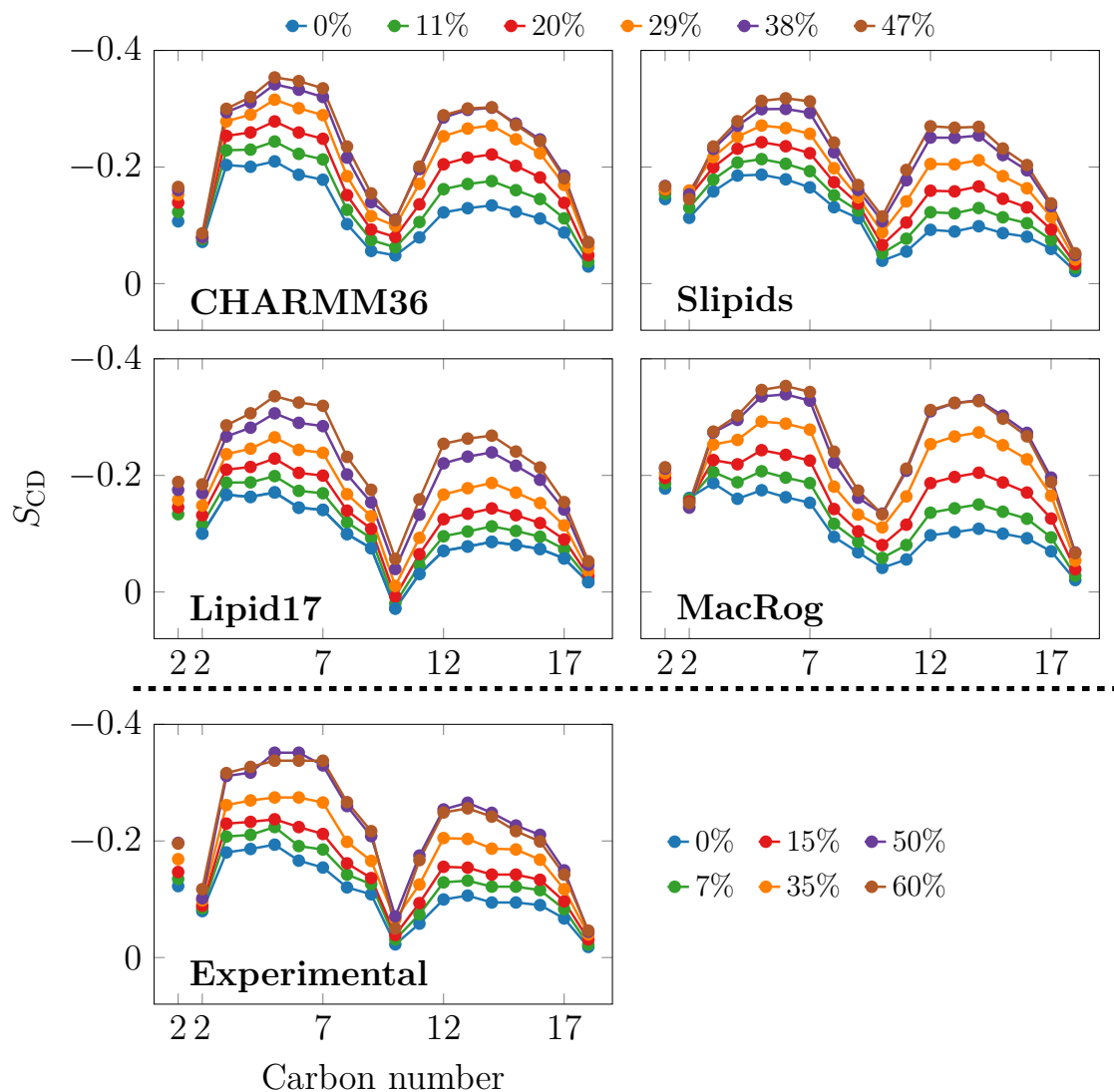


Figure S6: **Effect of cholesterol on the acyl chain order parameters of the POPC *sn*-2 (oleate) chain.** The legend at the top corresponds to all simulations, and the one at the bottom to the experiments. Since the order parameters measured for the two hydrogens bound to the C2 carbon differ, they are both shown in the plots.

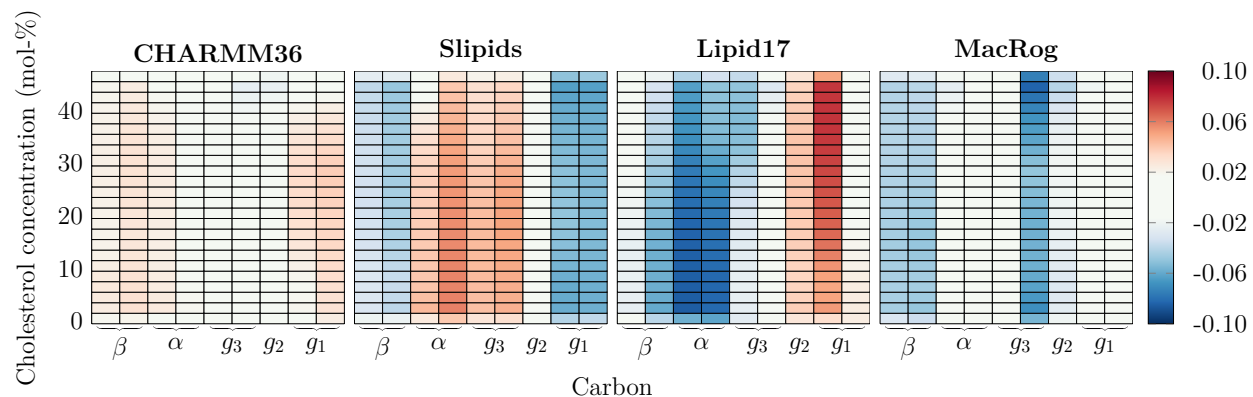


Figure S7: The deviation of POPC head group parameters from experimental values as a function of CHOL concentration.

## S2.4 Dynamic Properties

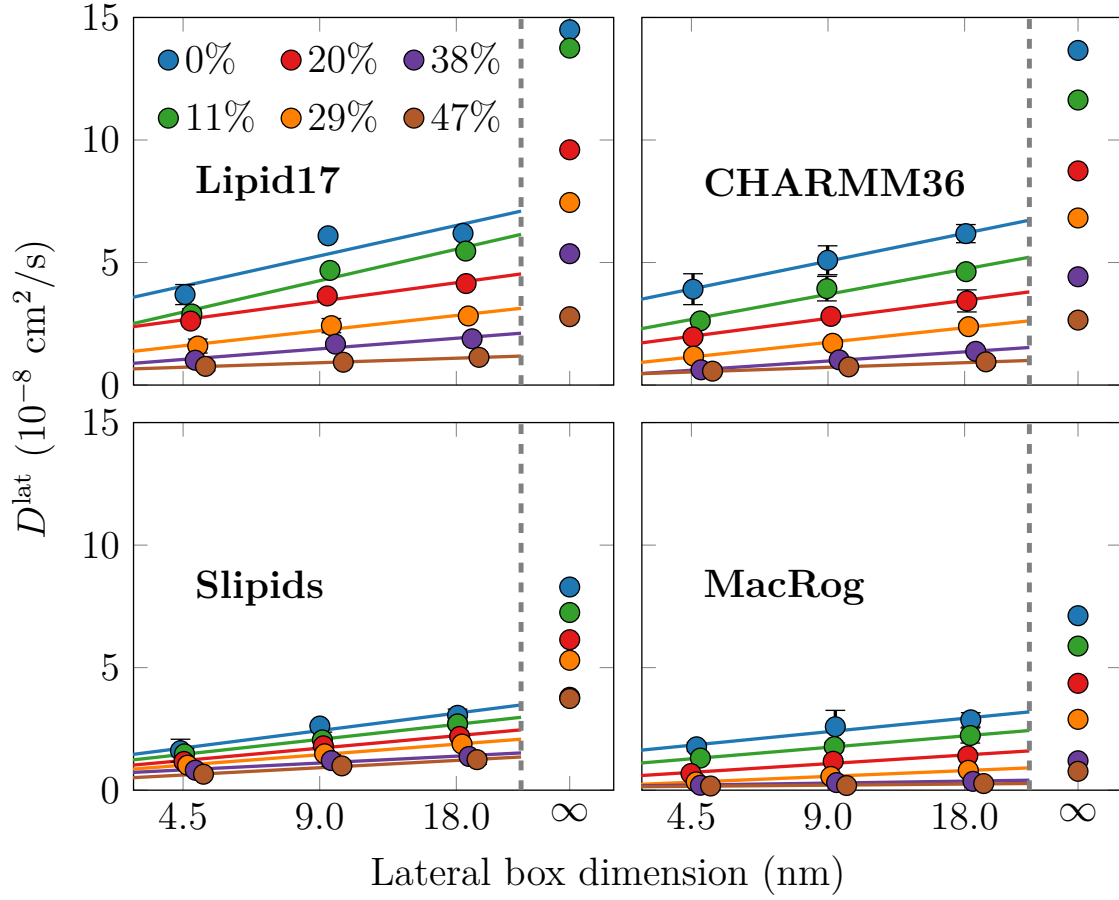


Figure S8: **Dependence of lateral diffusion coefficients on simulation box size.** The values calculated for the lipid centres of mass with `gmx msd` after eliminating leaflet drift. The values for the three system sizes are shown as markers together with fits of Eq. (2). The values extrapolated to infinite system sizes are also shown in the separate column.

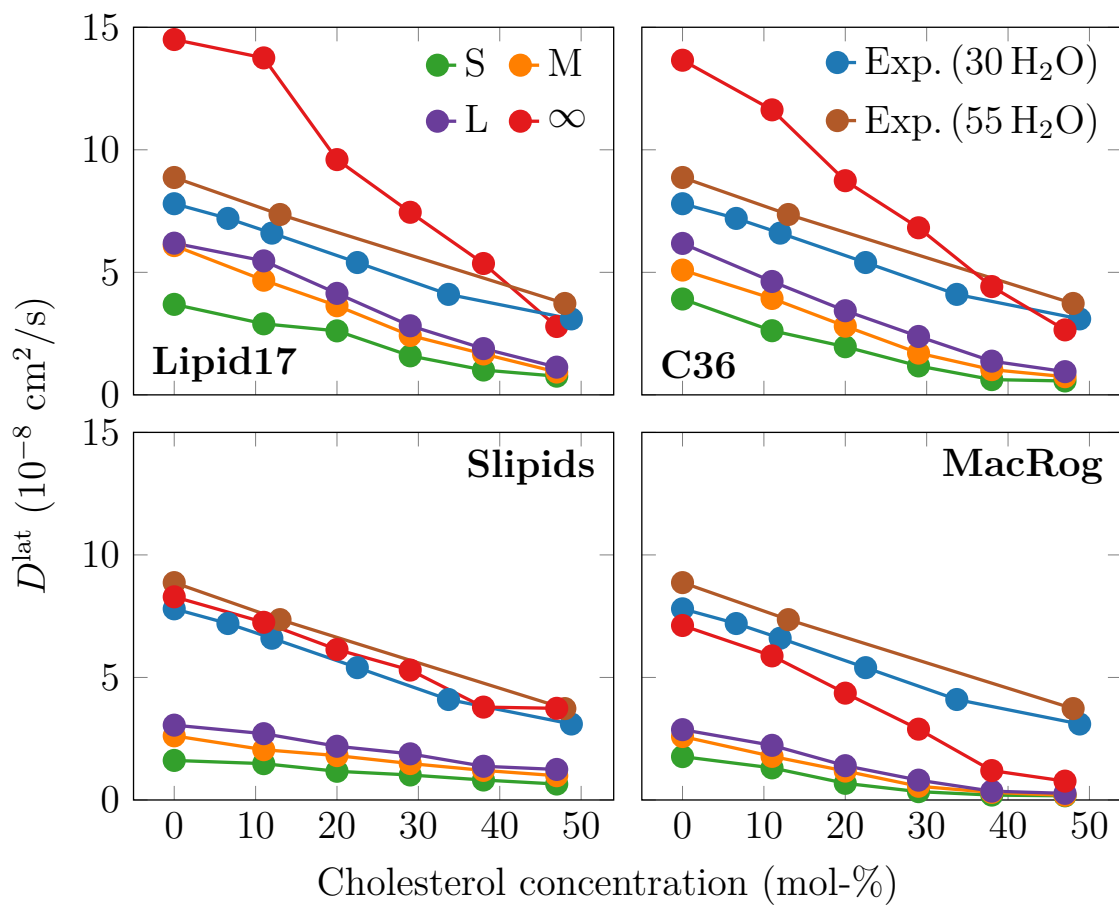


Figure S9: **Dependence of lateral diffusion coefficients on cholesterol concentration.** Data are shown for all system sizes; small (S), medium (M), and large (L). The values extrapolated to infinity are shown as well ( $\infty$ ). Experimental measured at two hydration levels, 30 m-% and 55-% of water.<sup>17,18</sup>

## S2.5 Finite-Size Effects

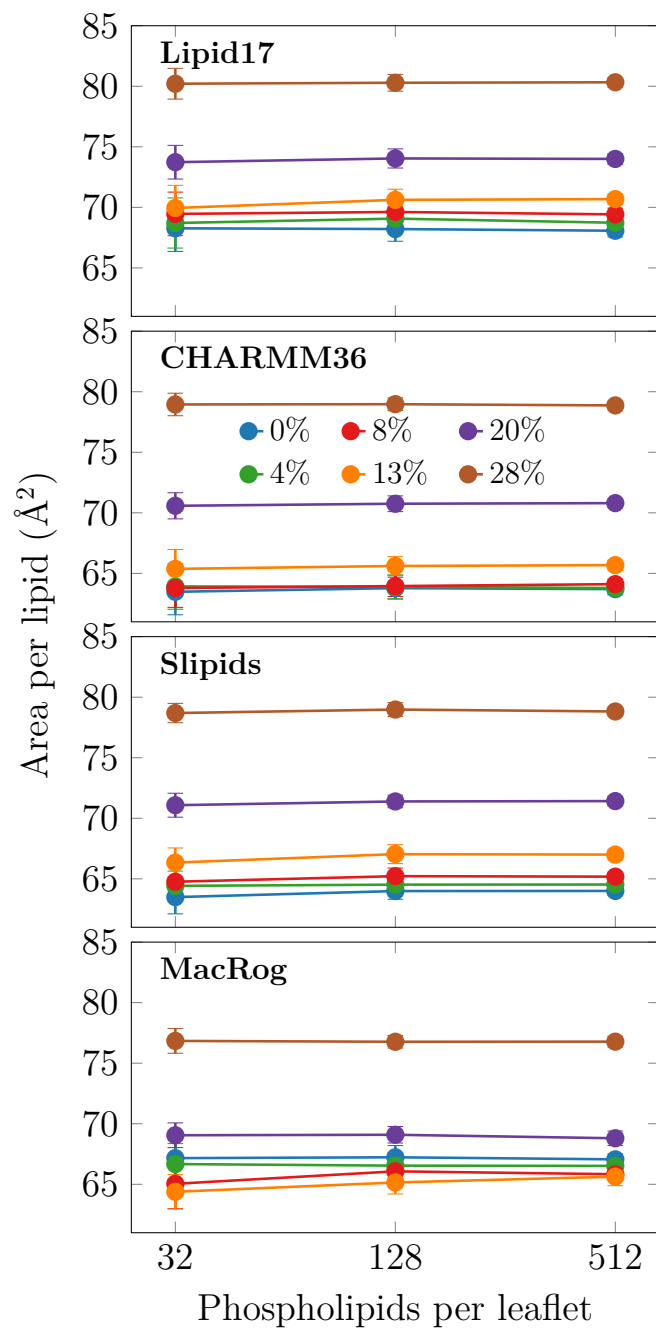


Figure S10: **Dependence of area per lipid on simulation box size.** Area per lipid is calculated by dividing the membrane area by the number of lipids in one leaflet. Error bars show standard error extracted using block averaging in `gmx analyze`.

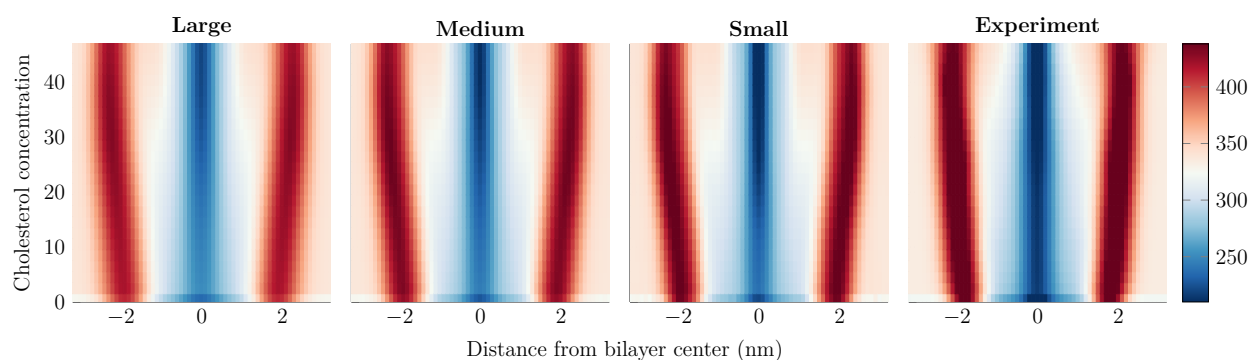


Figure S11: **Effect of system size on the density profiles.** Membrane undulations are larger in the larger systems, which leads to the smearing of the electron density profiles. Here, data are shown for CHARMM36 in the large (1024 POPC in total), medium (256 POPC in total), or small (64 POPC in total) systems. The experimental electron density profile is shown for comparison.



## References

- (1) Hess, B.; Bekker, H.; Berendsen, H. J. C.; Fraaije, J. G. E. M. LINCS: A Linear Constraint Solver for Molecular Dynamics Simulations. *J. Comput. Chem.* **1997**, *18*, 1463–1472.
- (2) Hess, B. P-LINCS: A Parallel Linear Constraint Solver for Molecular Simulation. *J. Chem. Theory Comput.* **2008**, *4*, 116–122.
- (3) Kiirikki, A. M.; Antila, H. S.; Bort, L.; Buslaev, P.; Favela, F.; Ferreira, T. M.; Fuchs, P. F.; Garcia-Fandino, R.; Gushchin, I.; Kav, B. et al. NMRlipids Databank: Making Data-Driven Analyses of Membrane Properties Accessible for All. 2023; <https://doi.org/10.26434/chemrxiv-2023-jrpwm>.
- (4) Páll, S.; Hess, B. A Flexible Algorithm for Calculating Pair Interactions on SIMD Architectures. *Comput. Phys. Commun.* **2013**, *184*, 2641–2650.
- (5) Essman, U. L.; Perera, M. L.; Berkowitz, M. L.; Larden, T.; Lee, H.; Pedersen, L. G. A Smooth Particle Mesh Ewald Potential. *J. Chem. Phys.* **1995**, *103*, 8577–8592.
- (6) Darden, T.; York, D.; Pedersen, L. Particle Mesh Ewald: An N·log(N) Method for Ewald Sums in Large Systems. *J. Chem. Phys.* **1993**, *98*, 10089–10092.
- (7) Shirts, M. R.; Mobley, D. L.; Chodera, J. D.; Pande, V. S. Accurate and Efficient Corrections for Missing Dispersion Interactions in Molecular Simulations. *J. Phys. Chem. B* **2007**, *111*, 13052–13063.
- (8) Hoover, W. G. Canonical Dynamics: Equilibrium Phase-Space Distributions. *Phys. Rev. A* **1985**, *31*, 1695–1697.
- (9) Nose, S. A Molecular Dynamics Method for Simulations in the Canonical Ensemble. *Mol. Phys.* **1984**, *52*, 255–268.

- (10) Bussi, G.; Donadio, D.; Parrinello, M. Canonical Sampling Through Velocity Rescaling. *J. Chem. Phys.* **2007**, *126*.
- (11) Parrinello, M.; Rahman, A. Polymorphic Transitions in Single Crystals: A New Molecular Dynamics Method. *J. Appl. Phys.* **1981**, *52*, 7182–7190.
- (12) Berendsen, H. J.; Postma, J. v.; Van Gunsteren, W. F.; DiNola, A.; Haak, J. R. Molecular Dynamics With Coupling to an External Bath. *J. Chem. Phys.* **1984**, *81*, 3684–3690.
- (13) Durell, S. R.; Brooks, B. R.; Ben-Naim, A. Solvent-Induced Forces Between Two Hydrophilic Groups. *J. Phys. Chem.* **1994**, *98*, 2198–2202.
- (14) Jorgensen, W. L.; Chandrasekhar, J.; Madura, J. D.; Impey, R. W.; Klein, M. L. Comparison of Simple Potential Functions for Simulating Liquid Water. *J. Chem. Phys.* **1983**, *79*, 926–935.
- (15) Kulig, W.; Pasenkiewicz-Gierula, M.; Róg, T. *Cis* and *Trans* Unsaturated Phosphatidylcholine Bilayers: A Molecular Dynamics Simulation Study. *Chem. Phys. Lipids* **2016**, *195*, 12–20.
- (16) Milan Rodriguez, P.; Fuchs, P. F. MacRog Pure POPC MD Simulation (300 K - 500ns - 1 bar). 2020; <https://doi.org/10.5281/zenodo.3741793>.
- (17) Filippov, A.; Orädd, G.; Lindblom, G. The Effect of Cholesterol on the Lateral Diffusion of Phospholipids in Oriented Bilayers. *Biophys. J.* **2003**, *84*, 3079–3086.
- (18) Filippov, A.; Orädd, G.; Lindblom, G. Influence of Cholesterol and Water Content on Phospholipid Lateral Diffusion in Bilayers. *Langmuir* **2003**, *19*, 6397–6400.