**Co-immunoprecipitation**

Twenty to thirty seedlings per plate were grown in wells of a 6-well plate for 2 weeks, transferred to 2 mM MES-KOH, pH 5.8, and incubated overnight. The next day, BL (final concentration 1 μM) and/or RALF23 (final concentration 1 μM) were added and incubated for 90 min. Seedlings were then frozen in liquid nitrogen and subjected to protein extraction. To analyze BRI1-GFP/BAK1 association, proteins were isolated in 50 mM Tris-HCl pH 7.5, 150 mM NaCl, 10 % glycerol, 5 mM dithiothreitol, 1 % protease inhibitor cocktail (Sigma-Aldrich), 2 mM Na2MoO4, 2.5 mM NaF, 1.5 mM activated Na3VO4, 1 mM phenylmethanesulfonyl fluoride, and 0.5% IGEPAL. For immunoprecipitations, GFP-Trap agarose beads (ChromoTek) were used and incubated with the crude extract for 3 to 4 hours at 4°C. Subsequently, beads were washed three times with wash buffer (50 mM Tris-HCl pH 7.5, 150 mM NaCl, 1 mM phenylmethanesulfonyl fluoride, 0,1 % IGEPAL) before adding Laemmli sample buffer and incubating for 10 minutes at 95°C. Analysis was carried out by SDS-PAGE and immunoblotting.

**Immunoblotting**

Protein samples were separated in 8-10 % bisacrylamide gels at 150 V for approximately 2 hours and transferred into activated PVDF membranes at 100 V for 90 min. Immunoblotting was performed with antibodies diluted in blocking solution (5 % fat-free milk in TBS with 0.1 % [v/v] Tween-20). Antibodies used in this study were α-BAK1 (1:5000, or Agrisera AS12 1858), α-BES1 (1:1000,), α-GFP-HRP (1:5000, sc-9996-HRP, Santa Cruz), α-GFP (1:5000, sc-9996, Santa Cruz). Blots were developed with Pierce ECL/ECL Femto Western Blotting Substrate (Thermo Scientific). The following secondary antibody was used: anti-rabbit IgG (whole molecule)–HRP (A0545, Sigma, dilution 1:10,000).