**RNA isolation and quantitative RT-PCR**

RNA isolation, cDNA synthesis and quantitative real-time PCR (qRT-PCR) was performed as previously described in Albrecht et al. with minor modifications. Five-day-old seedlings grown on half MS 1 % sucrose pH 5.8 were transferred to liquid half MS 1 % sucrose pH 5.8 and grown for seven days in 24 well plates. Each sample, constituted of three individual seedlings, were blot-dried, transferred to 2 mL tubes containing 2 mm glass beads and flash-frozen in liquid nitrogen. Samples were grinded while frozen for 1.5 minute at 1.500 rpm using BioRad TissueLyser. RNA was extracted using TRIzol™ Reagent (ThermoFischer scientific). First-strand cDNA synthesis was performed using RevertAid first strand cDNA synthesis kit (ThermoFischer scientific) and oligo(dT)18 according to the manufacturer’s instructions. cDNA was amplified in triplicate by quantitative PCR by using PowerUp™ SYBR™ Green Master Mix (ThermoFischer scientific) and a 7500 Fast real-time PCR detection system. The relative transcript accumulation values were determined by using U-box as reference and the comparative Ct method (2-ΔΔCt). The following primers were used for cDNA amplification, CPD (AT5G05690): forward: 5’-CCCAAACCACTTCAAAGATGCT-3’ and reverse 5’-GGGCCTGTCGTTACCGAGTT-3’, DWF4 (AT3G50660): forward ‘5-CATAAAGCTCTTCAGTCACGA-3’ and reverse ‘5-CGTCTGTTCTTTGTTTCCTAA-3’ and U-BOX (AT5G15400): forward ‘5-TGCGCTGCCAGATAATACACTATT-3’ and reverse ‘5-TGCTGCCCAACATCAGGTT-3’