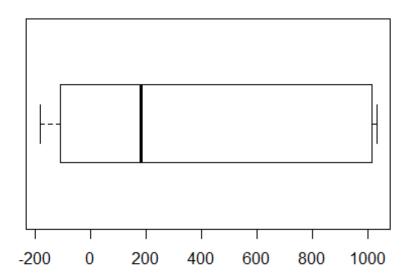
Mid_termQ1.R

```
source("lvalprogs.r")
library("aplpack")
## Loading required package: tcltk
time \leftarrow c(1,7,8,10,14,16,21,24,28,30,42,46,60,63,65)
wt2 <- c(143,-184,182,-110,1017,986,1010,1001,-111,-60,-151,-111,1024,1031,10
28)
data = matrix(data=c(time,wt2),byrow=FALSE,ncol=2,nrow=15)
colnames(data) <- c("time","wt2")</pre>
k <- summary(wt2)</pre>
print (k)
##
      Min. 1st Qu. Median
                             Mean 3rd Qu.
                                               Max.
   -184.0 -110.5
                     182.0
                             446.3 1014.0 1031.0
1 <- lval(wt2)
#d <- Lval.sub(wt2)
print (1)
##
     Depth Lower Upper
                           Mid Spread pseudo-s
## M
       8.0 182.0 182.0 182.0
                                    0
                                        0.0000
## F
       4.5 -110.5 1013.5 451.5 1124 833.2224
## E
       2.5 -131.0 1026.0 447.5 1157 502.8907
## D
       1.5 -167.5 1029.5 431.0 1197 390.1258
## C
       1.0 -184.0 1031.0 423.5 1215 326.1339
stem(wt2)
##
##
     The decimal point is 3 digit(s) to the right of the
##
##
     -0 | 221111
##
     0 | 12
##
     0
##
     1 | 0000000
stem.leaf(wt2)
## 1 | 2: represents 1200
## leaf unit: 100
##
               n: 15
           -0* | 111110
##
      6
     (2)
            0* | 11
##
##
             t
```



```
#2

0.4 + 0.007*5000

## [1] 35.4

#35.4

#3A)

#Here data is very small and its positively skewed, hence I suggest no transformation

#3B)

#All the five batch have different spreads, so I suggest some transformation which can result in comparable batches.

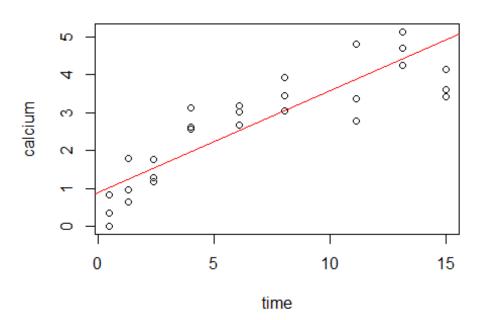
#May be a log transformation on y as log(f), where f is f-spread, x as median.
```

```
#3C)
#Data is having heavy tails . Transformation would affect the tails, Hence I
suggest no transformation
#3D)
#Data is positive skewed, We can transform the data.
#May be we can transform
\#x = (upperforth^2 + Lowerforth^2)/4*Median
#y = (upperforth - lowerforth)/2
#4
#The kernel estimate is a weighted average of the observations within the smo
othing window.
#two type of linear smothers are
#1) linear regression line and 2) Sample mean.
#linear smothers tend to be affected by outliers and are smooth over sharp fe
atures.
#nonlinear smoothers use median hence are flexible and are resistant.
#Disadvantage is the need to use iterative optimization to compute parameter
estimate.
```

```
source("E:/Study stuff/Subjects and courses/S 670 Exploratory Data Analysis/H
ome work/Mid term/midtermexam/rrline1.R")
#source("E:/Study stuff/Subjects and courses/S 670 Exploratory Data Analysis/
Home work/Mid term/midtermexam/run rrline.r")
source('E:/Study stuff/Subjects and courses/S 670 Exploratory Data Analysis/H
ome work/Assignment 4/run rrline.R')
#5)
#5a)
time \leftarrow c(0.45, 0.45, 0.45, 1.3, 1.3, 1.3, 1.3, 2.4, 2.4, 2.4, 4, 4, 4, 6.1, 6.1, 6.1, 8.05, 8.05,
8.05,
           11.15,11.15,11.15,13.15,13.15,13.15,15,15,15)
calcium \leftarrow c(0.342,0,0.825,1.78,0.954,0.641,1.751,1.275,1.173,3.123,2.61,2.57
4,3.179,
              3.008, 2.671, 3.06, 3.943, 3.437, 4.807, 3.356, 2.783, 5.138, 4.703, 4.257
,3.604,
             4.15, 3.425)
data <- matrix(data=c(time,calcium),byrow=TRUE,nrow=2,ncol=27)</pre>
row.names(data) <- c("time","calcium")</pre>
plot(data[2,]~data[1,],main="Time vs calcium plot",xlab="time",ylab="calcium"
```

```
rr <- rrline1(data[1,],data[2,])
#fit <- lm(calcium~time)
abline(rr$a,rr$b,col='red')</pre>
```

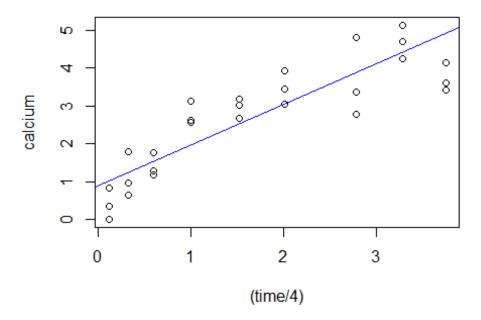
Time vs calcium plot



```
#slope of resistant regression after 1st iteration is 0.26970
#Intercept of resistant regression after 1st iteration is 0.8888776

#5b)
#I would like to apply transformation on above data.

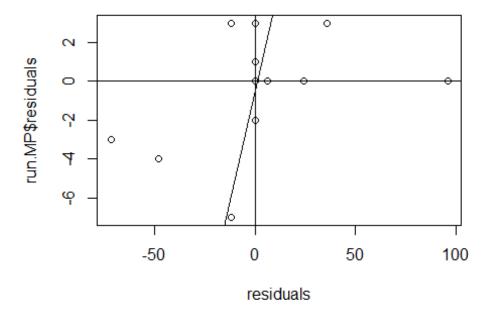
# 5C and D)I see that y - axis varaibles remained same where as x axis variab les were shrinked specifically 15 was shrinked to a number near 4.I believe t hat x coordinated were divided by 4.
plot((time/4),calcium)
rr2 <- rrline1((data[1,]/4),data[2,])
abline(rr2$a,rr2$b,col='blue')</pre>
```



```
#plot((time^1/4),calcium)
#slope of resistant regression after 1st iteration is 1.078819
#Intercept of resistant regression after 1st iteration is 0.8888776
#5E)The residual values are clustered in panel c where are residuals in panel
D are more scattered. Understanding panel D is more easy.
#5F)
rr2 <- run.rrline(time^(1/2),calcium)</pre>
##
                     b
                          |res|
## 1 -0.40474 1.28554 13.13373
## 2 0.35643 -0.12409 13.25724
## 3 -0.05537 0.03574 13.17437
## 4 0.01595 -0.01029 13.19824
## 5 -0.00459 0.00296 13.19137
##
     -0.09232 1.18986 13.19137
print (rr2)
## $a
## [1] -0.09232189
##
## $b
## [1] 1.189859
##
```

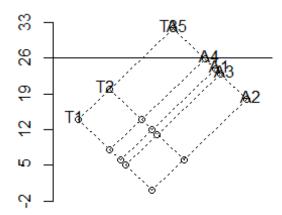
```
## $res
## [1] -0.36385990 -0.70585990 0.11914010 0.51567371 -0.31032629
## [6] -0.62332629   0.00000000 -0.47600000 -0.57800000   0.83560356
## [11] 0.32260356 0.28660356 0.33258654 0.16158654 -0.17541346
## [16] -0.22360863   0.65939137   0.15339137   0.92618989   -0.52481011
## [21] -1.09781011 0.91554408 0.48054408 0.03454408 -0.91198284
## [26] -0.36598284 -1.09098284
##
## $coef
##
                                  Iresl
                             b
                a
## 1 -0.404739546 1.285538623 13.13373
## 2 0.356432222 -0.124090740 13.25724
## 3 -0.055368655 0.035740313 13.17437
## 4 0.015947145 -0.010293838 13.19824
## 5 -0.004593058 0.002964806 13.19137
  -0.092321892 1.189859165 13.19137
rr3 <- run.rrline(time^(1/2),calcium)</pre>
                         |res|
##
                     b
## 1 -0.40474 1.28554 13.13373
## 2 0.35643 -0.12409 13.25724
## 3 -0.05537 0.03574 13.17437
## 4 0.01595 -0.01029 13.19824
## 5 -0.00459 0.00296 13.19137
## -0.09232 1.18986 13.19137
ynew <- calcium-(-0.124)*time^{(1/2)} - 0.256
rr4 <- rrline1(time^(1/2),ynew)</pre>
rr5 <- rrline1(time^(1/2), rr4$res)
print(rr5)
## $a
## [1] 0.4065408
##
## $b
## [1] -0.1399387
##
## $sumres
## [1] 13.26545
##
## $res
## [1] -0.3919284 -0.7339284 0.0910716 0.5026035 -0.3233965 -0.6363965
## [7]
       0.0000000 -0.4760000 -0.5780000 0.8500091 0.3370091 0.3010091
## [13] 0.3620052 0.1910052 -0.1459948 -0.1824486
                                                     0.7005514 0.1945514
## [19] 0.9833885 -0.4676115 -1.0406115 0.9819180 0.5469180 0.1009180
## [25] -0.8377259 -0.2917259 -1.0167259
#slope -0.1399387
#intercept 0.4065408
```

```
library("miscTools")
source('E:/Study stuff/Subjects and courses/S 670 Exploratory Data Analysis/H
ome work/Mid term/midtermexam/forgetitplot.R')
T1 \leftarrow c(5,6,3,11,10)
T2 \leftarrow c(14,10,6,12,21)
T3 \leftarrow c(16,24,15,26,32)
Area <- matrix(data=c(T1,T2,T3),nrow=3,ncol=5,byrow=TRUE)
Area1 <- Area
row.names(Area) <- c('T1','T2','T3')</pre>
colnames(Area) <- c('A1', 'A2', 'A3', 'A4', 'A5')</pre>
#6A)
run.MP <- medpolish(Area)</pre>
## 1: 28
## Final: 28
Area <- cbind(Area,run.MP$row)</pre>
Area <- rbind(Area,c(run.MP$col,0))</pre>
Area[4,6] <- run.MP$overall
print (Area)
      A1 A2 A3 A4 A5
##
## T1 5 6 3 11 10 -6
## T2 14 10 6 12 21 0
## T3 16 24 15 26 32 12
      -1 0 -6 2 8 12
##
#After applying Median polish each term in matrix Area can be obtained
#by adding Overall + roweffect + coloumn effect + residual.
AnalogR2 <- 1 - sum(abs(run.MP$residuals))/sum(abs(Area1 - run.MP$overall))</pre>
#Analog r2 is pretty good indicating its a good fit.
#6C)
#Diagnostic plot can used to understand how well the model fits the data.
residuals <- matrix(run.MP$row,ncol=1) %*% matrix(run.MP$col,nrow=1)</pre>
plot(residuals,run.MP$residuals)
fit=lm(run.MP$residuals~residuals)
abline(fit, v=0, h=0)
## Warning in abline(fit, v = 0, h = 0): only using the first two of 30
## regression coefficients
```



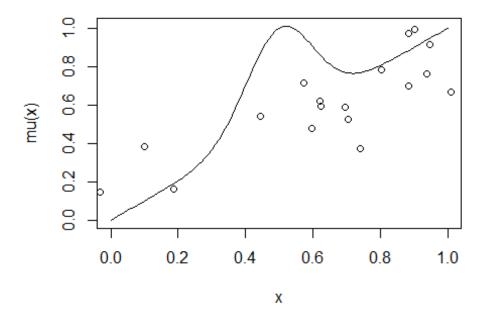
```
#Data does not reveal any treand.
```

```
#6D)
forgetitplot(run.MP)
abline(h=10)
```



```
#
x = seq(0.01,0.99,by=0.02)
y = c(-.0937, .0247, .1856, .1620, -.0316, .1442, .0993, .3823, -.0624, .3262
, .1271, -.4158, .0975, -.0836, .7410, .3749, .4446, .5432, .6946, .5869, .93
84, .7647, .9478, .9134, 1.2437, .9070, 1.2289, .9638, .8834, .6982, .5729, .
7160, 1.0083, .6681, .5964, .4759, .6217, .6221, .6244, .5918, .7047, .5234, .9022, .9930, .8045, .7858, 1.1939, .9272, .8832, .9751)
mat = matrix(y, ncol = 2, byrow = T)

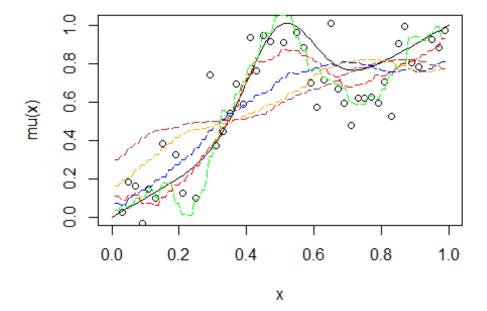
mu = function(t){
    t + 0.5 *exp(-50*(t-0.5)^2)
    }
curve(mu(x),0,1)
points(mat[,1],mat[,2])
```



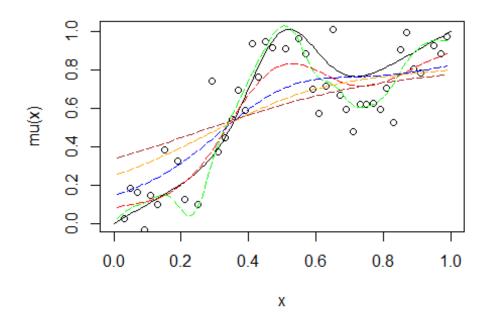
```
#7B)

plot_ksmooth = function(x, y, mu, method) {
    lam = c(0.1, 0.3, 0.5, 0.7, 0.9)
    col = c("green", "red", "blue", "orange", "brown")
    curve(mu(x), 0, 1)
    points(x, y)
    for(i in 1:length(lam)) {
        fit = ksmooth(x, y, kernel = method, bandwidth = lam[i])
        lines(fit$x, fit$y, lty = 5, col = col[i])
    }
}

plot_ksmooth(x,y,mu,"box")
```

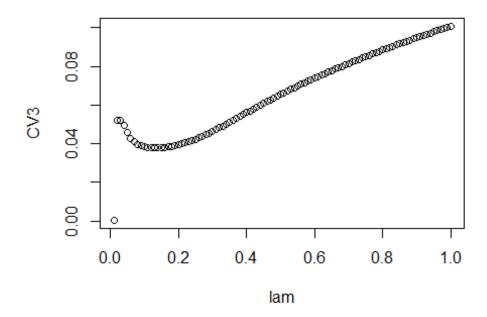


plot_ksmooth(x,y,mu,"normal")

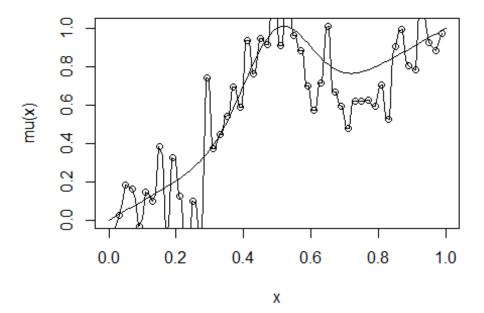


```
#7C)
CV <- function(x, y, lam, spline = TRUE) {</pre>
```

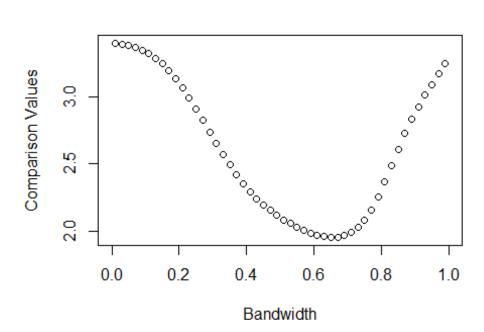
```
n \leftarrow length(x)
CV <-numeric(n)</pre>
for(i in 1:n) {
  if(spline == FALSE) {
    fit = ksmooth(x[-i], y[-i], kernel = "normal", bandwidth = lam, n.points
= n)
  } else {
    fit = smooth.spline(x[-i], y[-i], spar = lam)
  CV[i] = (y[i] - fit$y[i])^(2)
CV = sum(CV, na.rm = T)
return(CV/n)
CV_plot = function(x, y, spline = TRUE) {
  lam = seq(0.01, 1, by = 0.01)
  CV3 = c()
  for(i in 1:length(lam)) {
    CV2 <- CV(x, y, lam[i], spline = spline)
    CV3 = c(CV3, CV2)
  }
  plot(lam,CV3)
  lam_min = lam[which.min(CV3)]
  return(lam_min)
  }
minLamda = CV_plot(x, y, spline = FALSE)
```



```
curve(mu(x), 0, 1)
points(x, y)
fit = ksmooth(x, y, kernel = "normal", bandwidth = minLamda)
lines(fit$x, fit$y)
```

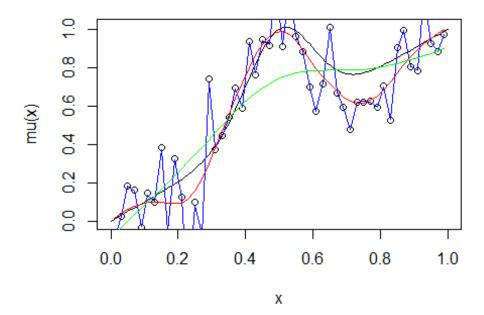


```
CV1 = function(bw,x,y){
    CV= numeric(length(x))
    for (i in 1:length(x)){
        fit = smooth.spline(x[-i],y[-i],spar=bw,cv=TRUE)
        hat_y=fit$y
        CV[i]=(y[i]-hat_y[i])^2
        }
    sum(CV,na.rm=T)
    }
Score_CV=c()
for (j in 1:length(x)){
    Score_CV <- c(Score_CV,CV1(x[j],x,y))
    }
plot(x, Score_CV, xlab="Bandwidth",ylab="Comparison Values",xlim=c(0,1))</pre>
```

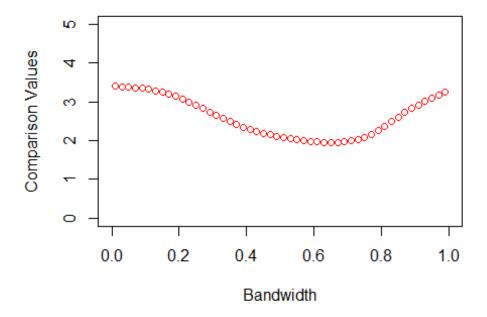


```
#7E)

curve(mu(x),0,1)
points(x,y)
low = smooth.spline(x, y, spar=0.05)
lines(low,col="blue")
def = smooth.spline(x, y)
lines(def,col="red")
high = smooth.spline(x, y,spar=0.9)
lines(high,col="green")
```



```
#7.f
CVFUNCs = function(bw,x,y){
n = length(x)
CV = numeric(n)
for (i in 1:n) {
  fit = smooth.spline(x[-i],y[-i],spar=bw,cv=TRUE)
  hat_y=fit$y
  CV[i]=(y[i]-hat_y[i])^2
sum(CV,na.rm=T)
1 < -seq(0.01, 1, by=0.02)
Score_CV<-c()
for (\bar{j} \text{ in } 1: length(1)){
  Score_CV<-c(Score_CV,CVFUNCs(1[j],x,y))</pre>
  }
plot(l, Score_CV, xlab="Bandwidth",ylab="Comparison Values",xlim=c(0,1),ylim=
c(0,5),col="red")
```



```
q7x<-x
q7y<-y
OptimumLamda <- 0.0003
OptimumSpar <- 0.648
Spar_sam \leftarrow seq(0.4, 0.9, by=0.001)
getCVandGCV <- function(x, y, spar) {</pre>
  Fit_CV <- smooth.spline(x,y,cv=TRUE,spar=s)</pre>
  Fit_GCV <- smooth.spline(x,y,cv=FALSE, spar=s)</pre>
  list(cv = Fit CV$cv.crit, gcv = Fit GCV$cv.crit, CV lambda = Fit CV$lambda,
GCV_lambda = Fit_GCV$lambda)
  }
CV_all <- c()
GCV_all <- c()
CVLambda_all <- c()
Lambda_allGCV <- c()
for(s in Spar_sam) {
  fit <- getCVandGCV(q7x, q7y, s)</pre>
  CV_all <- c(CV_all, fit$cv)
  GCV_all <- c(GCV_all, fit$gcv)</pre>
  CVLambda all <- c(CVLambda all, fit$CV lambda)</pre>
  Lambda_allGCV = c(Lambda_allGCV, fit$GCV_lambda)
  }
plot(Spar_sam, CV_all, col="green", main="GCV ,CV vs Lamda")
points(Spar_sam, GCV_all, col="blue")
```

GCV ,CV vs Lamda

