set.seed(1609) setwd('/home/nastasista/Metagenomics') Read files and data path <- '/home/nastasista/Metagenomics/data\_met/sequences'</pre> list.files(path) ## [1] "Abacumov-B-1\_S1\_L001\_R1\_001.fastq.gz" ## [2] "Abacumov-B-1\_S1\_L001\_R2\_001.fastq.gz" ## [3] "Abacumov-B-13\_S13\_L001\_R1\_001.fastq.gz" ## [4] "Abacumov-B-13\_S13\_L001\_R2\_001.fastq.gz" ## [5] "Abacumov-B-14\_S14\_L001\_R1\_001.fastq.gz" ## [6] "Abacumov-B-14\_S14\_L001\_R2\_001.fastq.gz" ## [7] "Abacumov-B-15\_S15\_L001\_R1\_001.fastq.gz" ## [8] "Abacumov-B-15\_S15\_L001\_R2\_001.fastq.gz" ## [9] "Abacumov-B-16\_S16\_L001\_R1\_001.fastq.gz" ## [10] "Abacumov-B-16\_S16\_L001\_R2\_001.fastq.gz" ## [11] "Abacumov-B-2\_S2\_L001\_R1\_001.fastq.gz" ## [12] "Abacumov-B-2\_S2\_L001\_R2\_001.fastq.gz" ## [13] "Abacumov-B-25\_S25\_L001\_R1\_001.fastq.gz" ## [14] "Abacumov-B-25\_S25\_L001\_R2\_001.fastq.gz" ## [15] "Abacumov-B-26\_S26\_L001\_R1\_001.fastq.gz" ## [16] "Abacumov-B-26\_S26\_L001\_R2\_001.fastq.gz" ## [17] "Abacumov-B-27\_S27\_L001\_R1\_001.fastq.gz" ## [18] "Abacumov-B-27\_S27\_L001\_R2\_001.fastq.gz" ## [19] "Abacumov-B-28\_S28\_L001\_R1\_001.fastq.gz" ## [20] "Abacumov-B-28\_S28\_L001\_R2\_001.fastq.gz" ## [21] "Abacumov-B-3\_S3\_L001\_R1\_001.fastq.gz" ## [22] "Abacumov-B-3\_S3\_L001\_R2\_001.fastq.gz" ## [23] "Abacumov-B-37\_S37\_L001\_R1\_001.fastq.gz" ## [24] "Abacumov-B-37\_S37\_L001\_R2\_001.fastq.gz" ## [25] "Abacumov-B-38\_S38\_L001\_R1\_001.fastq.gz" ## [26] "Abacumov-B-38\_S38\_L001\_R2\_001.fastq.gz" ## [27] "Abacumov-B-39\_S39\_L001\_R1\_001.fastq.gz" ## [28] "Abacumov-B-39\_S39\_L001\_R2\_001.fastq.gz" ## [29] "Abacumov-B-4\_S4\_L001\_R1\_001.fastq.gz" ## [30] "Abacumov-B-4\_S4\_L001\_R2\_001.fastq.gz" ## [31] "Abacumov-B-40\_S40\_L001\_R1\_001.fastq.gz" ## [32] "Abacumov-B-40\_S40\_L001\_R2\_001.fastq.gz" ## [33] "Abacumov-B-49\_S49\_L001\_R1\_001.fastq.gz" ## [34] "Abacumov-B-49\_S49\_L001\_R2\_001.fastq.gz" ## [35] "Abacumov-B-50\_S50\_L001\_R1\_001.fastq.gz" ## [36] "Abacumov-B-50\_S50\_L001\_R2\_001.fastq.gz" ## [37] "Abacumov-B-51\_S51\_L001\_R1\_001.fastg.gz" ## [38] "Abacumov-B-51\_S51\_L001\_R2\_001.fastq.gz" ## [39] "Abacumov-B-52\_S52\_L001\_R1\_001.fastq.gz" ## [40] "Abacumov-B-52\_S52\_L001\_R2\_001.fastq.gz" ## [41] "Abacumov-B-61\_S61\_L001\_R1\_001.fastq.gz" ## [42] "Abacumov-B-61\_S61\_L001\_R2\_001.fastg.gz" ## [43] "Abacumov-B-62\_S62\_L001\_R1\_001.fastq.gz" ## [44] "Abacumov-B-62\_S62\_L001\_R2\_001.fastq.gz" ## [45] "Abacumov-B-63\_S63\_L001\_R1\_001.fastq.gz" ## [46] "Abacumov-B-63\_S63\_L001\_R2\_001.fastq.gz" ## [47] "Abacumov-B-64\_S64\_L001\_R1\_001.fastq.gz" ## [48] "Abacumov-B-64\_S64\_L001\_R2\_001.fastq.gz" ## [49] "filtered" metadata <- read.csv('data\_met/map.csv')</pre> metadata\$SampleID <- paste(metadata\$Source, metadata\$Site, metadata\$Horizont, metadata\$Repeat, sep=".")</pre> metadata ## Filename Source Site Horizont Repeat ## 1 Abacumov-B-1 Self-growing Dumps B1 Abacumov-B-2 Self-growing Dumps B1 Abacumov-B-3 Self-growing Dumps B1 ## 4 Abacumov-B-4 Self-growing Dumps B1 ΑY ## 5 Abacumov-B-13 С Litostrat B2 ## 6 Abacumov-B-14 Litostrat B2 ## 7 Abacumov-B-15 Litostrat B2 ## 8 Abacumov-B-16 Litostrat B2 ## 9 Abacumov-B-25 Coal Mine Terricon B3 ## 10 Abacumov-B-26 Coal Mine Terricon B3 ## 11 Abacumov-B-27 Coal Mine Terricon B3 ## 12 Abacumov-B-28 Coal Mine Terricon B3 С ## 13 Abacumov-B-37 Local Reference B4 ## 14 Abacumov-B-38 Local Reference B4 ΑY ## 15 Abacumov-B-39 Local Reference B4 ## 16 Abacumov-B-40 Local Reference B4 ## 17 Abacumov-B-49 Embryo Sand B5 ΑY ## 18 Abacumov-B-50 Embryo Sand B5 ΑY ## 19 Abacumov-B-51 Embryo Sand B5 ## 20 Abacumov-B-52 Embryo Sand B5 ΑY ## 21 Abacumov-B-61 Regional Reference B6 1 ## 22 Abacumov-B-62 Regional Reference B6 2 ## 23 Abacumov-B-63 Regional Reference B6 ΑY 3 ## 24 Abacumov-B-64 Regional Reference B6 ## SampleID ## 1 Self-growing Dumps.B1.AY.1 ## 2 Self-growing Dumps.B1.AY.2 ## 3 Self-growing Dumps.B1.AY.3 ## 4 Self-growing Dumps.B1.AY.4 ## 5 Litostrat.B2.C.1 Litostrat.B2.C.2 ## 6 Litostrat.B2.C.3 ## 8 Litostrat.B2.C.4 ## 9 Coal Mine Terricon.B3.C.1 ## 10 Coal Mine Terricon.B3.C.2 ## 11 Coal Mine Terricon.B3.C.3 ## 12 Coal Mine Terricon.B3.C.4 Local Reference.B4.AY.1 Local Reference.B4.AY.2 ## 14 Local Reference.B4.AY.3 ## 15 Local Reference.B4.AY.4 ## 16 ## 17 Embryo Sand.B5.AY.1 Embryo Sand.B5.AY.2 ## 18 Embryo Sand.B5.AY.3 ## 19 Embryo Sand.B5.AY.4 ## 21 Regional Reference.B6.AY.1 ## 22 Regional Reference.B6.AY.2 ## 23 Regional Reference.B6.AY.3 ## 24 Regional Reference.B6.AY.4 Run DADA2 pipeline A realisation of a basic tutorial from https://benjjneb.github.io/dada2/tutorial.html # Forward and reverse fastq filenames have format: SAMPLENAME\_R1\_001.fastq and SAMPLENAME\_R2\_001.fastq fnFs <- sort(list.files(path, pattern="\_R1\_001.fastq", full.names = TRUE))</pre> fnRs <- sort(list.files(path, pattern="\_R2\_001.fastq", full.names = TRUE))</pre> # Extract sample names, assuming filenames have format: SAMPLENAME\_XXX.fastq sample.names <- sapply(strsplit(basename(fnFs), "\_"), `[`, 1)</pre> sample.names ## [1] "Abacumov-B-1" "Abacumov-B-13" "Abacumov-B-14" "Abacumov-B-15" ## [5] "Abacumov-B-16" "Abacumov-B-2" "Abacumov-B-25" "Abacumov-B-26" ## [9] "Abacumov-B-27" "Abacumov-B-28" "Abacumov-B-3" "Abacumov-B-37" ## [13] "Abacumov-B-38" "Abacumov-B-39" "Abacumov-B-4" "Abacumov-B-40" ## [17] "Abacumov-B-49" "Abacumov-B-50" "Abacumov-B-51" "Abacumov-B-52" ## [21] "Abacumov-B-61" "Abacumov-B-62" "Abacumov-B-63" "Abacumov-B-64" Quality plot plotQualityProfile(fnFs[1:2]) Reads: 43307 Reads: 41438 # !Long Operations plotQualityProfile(fnFs, aggregate = T) Total reads: 929445 Cycle plotQualityProfile(fnRs, aggregate = T) Cycle ### Filter and Trim filtFs <- file.path(path, "filtered", paste0(sample.names, "\_F\_filt.fastq.gz"))</pre> filtRs <- file.path(path, "filtered", paste0(sample.names, "\_R\_filt.fastq.gz"))</pre> names(filtFs) <- sample.names</pre> names(filtRs) <- sample.names</pre> out <- filterAndTrim(fnFs, filtFs, fnRs, filtRs, truncLen=c(240,180),</pre> maxN=0, maxEE=c(2,2), truncQ=2, rm.phix=TRUE, compress=TRUE, multithread=TRUE) out ## reads.in reads.out ## Abacumov-B-1\_S1\_L001\_R1\_001.fastq.gz 43307 36622 ## Abacumov-B-13\_S13\_L001\_R1\_001.fastq.gz 41438 34160 31002 ## Abacumov-B-14\_S14\_L001\_R1\_001.fastq.gz 38072 ## Abacumov-B-15\_S15\_L001\_R1\_001.fastq.gz 46395 37918 38962 ## Abacumov-B-16\_S16\_L001\_R1\_001.fastq.gz 47274 ## Abacumov-B-2\_S2\_L001\_R1\_001.fastq.gz 45972 39386 ## Abacumov-B-25\_S25\_L001\_R1\_001.fastq.gz 29444 22676 17052 ## Abacumov-B-26\_S26\_L001\_R1\_001.fastq.gz 21786 ## Abacumov-B-27\_S27\_L001\_R1\_001.fastq.gz 19659 25002 ## Abacumov-B-28\_S28\_L001\_R1\_001.fastq.gz 25214 19501 ## Abacumov-B-3\_S3\_L001\_R1\_001.fastg.gz 47256 54999 ## Abacumov-B-37\_S37\_L001\_R1\_001.fastq.gz 31738 26915 ## Abacumov-B-38\_S38\_L001\_R1\_001.fastg.gz 33322 27806 22885 ## Abacumov-B-39\_S39\_L001\_R1\_001.fastq.gz 27048 ## Abacumov-B-4\_S4\_L001\_R1\_001.fastq.gz 40115 34065 ## Abacumov-B-40\_S40\_L001\_R1\_001.fastq.gz 28630 24332 45257 ## Abacumov-B-49\_S49\_L001\_R1\_001.fastq.gz 54922 ## Abacumov-B-50\_S50\_L001\_R1\_001.fastq.gz 41565 34229 ## Abacumov-B-51\_S51\_L001\_R1\_001.fastq.gz 50422 42869 ## Abacumov-B-52\_S52\_L001\_R1\_001.fastq.gz 38603 31780 ## Abacumov-B-61\_S61\_L001\_R1\_001.fastq.gz 48250 41246 ## Abacumov-B-62\_S62\_L001\_R1\_001.fastq.gz 39777 33175 ## Abacumov-B-63\_S63\_L001\_R1\_001.fastq.gz 38230 32469 ## Abacumov-B-64\_S64\_L001\_R1\_001.fastq.gz 37920 32221 Trimmed quality plot # !Long Operations plotQualityProfile(filtFs, aggregate = T) 24 files (aggregated) Total reads: 773443 plotQualityProfile(filtRs, aggregate = T) 24 files (aggregated) Total reads: 773443 Reads are trimmed fairly, everything is OK, go to the next step Build a model and apply it #графики вероятности перехода # !Long Operation errF <- learnErrors(filtFs, multithread=TRUE)</pre> ## 101232000 total bases in 421800 reads from 14 samples will be used for learning the error rates. errR <- learnErrors(filtRs, multithread=TRUE)</pre> ## 100742940 total bases in 559683 reads from 18 samples will be used for learning the error rates. plotErrors(errF, nominalQ=TRUE) ## Warning: Transformation introduced infinite values in continuous y-axis ## Transformation introduced infinite values in continuous y-axis A2G 1e-01 1e-02 -1e-03 -1e-04 C2A C2C C2G C2T 1e+00 1e-02 -G2C G2T G2G 1e-03 -1e-04 -T2G T2T 1e+00 0 0 0 0 0 0 0 0 0 0 0 0 0 1e-01 -1e-03 dadaFs <- dada(filtFs, err=errF, multithread=TRUE)</pre> ## Sample 1 - 36622 reads in 28087 unique sequences. ## Sample 2 - 34160 reads in 18735 unique sequences. ## Sample 3 - 31002 reads in 18643 unique sequences. ## Sample 4 - 37918 reads in 19870 unique sequences. ## Sample 5 - 38962 reads in 20810 unique sequences. ## Sample 6 - 39386 reads in 30781 unique sequences. ## Sample 7 - 22676 reads in 6818 unique sequences. ## Sample 8 - 17052 reads in 5663 unique sequences. ## Sample 9 - 19659 reads in 6237 unique sequences. ## Sample 10 - 19501 reads in 5706 unique sequences. ## Sample 11 - 47256 reads in 33296 unique sequences. ## Sample 12 - 26915 reads in 19311 unique sequences. ## Sample 13 - 27806 reads in 20185 unique sequences. ## Sample 14 - 22885 reads in 17428 unique sequences. ## Sample 15 - 34065 reads in 26006 unique sequences. ## Sample 16 - 24332 reads in 18733 unique sequences. ## Sample 17 - 45257 reads in 27334 unique sequences. ## Sample 18 - 34229 reads in 21495 unique sequences. ## Sample 19 - 42869 reads in 27611 unique sequences. ## Sample 20 - 31780 reads in 20178 unique sequences. ## Sample 21 - 41246 reads in 30101 unique sequences. ## Sample 22 - 33175 reads in 23318 unique sequences. ## Sample 23 - 32469 reads in 22700 unique sequences. ## Sample 24 - 32221 reads in 23605 unique sequences. dadaRs <- dada(filtRs, err=errR, multithread=TRUE)</pre> ## Sample 1 - 36622 reads in 28652 unique sequences. ## Sample 2 - 34160 reads in 19562 unique sequences. ## Sample 3 - 31002 reads in 18627 unique sequences. ## Sample 4 - 37918 reads in 19876 unique sequences. ## Sample 5 - 38962 reads in 20775 unique sequences. ## Sample 6 - 39386 reads in 31028 unique sequences. ## Sample 7 - 22676 reads in 7053 unique sequences. ## Sample 8 - 17052 reads in 5659 unique sequences. ## Sample 9 - 19659 reads in 7019 unique sequences. ## Sample 10 - 19501 reads in 5888 unique sequences. ## Sample 11 - 47256 reads in 34628 unique sequences. ## Sample 12 - 26915 reads in 20489 unique sequences. ## Sample 13 - 27806 reads in 20955 unique sequences. ## Sample 14 - 22885 reads in 17871 unique sequences. ## Sample 15 - 34065 reads in 26746 unique sequences ## Sample 16 - 24332 reads in 19350 unique sequences. ## Sample 17 - 45257 reads in 28159 unique sequences. ## Sample 18 - 34229 reads in 22116 unique sequences. ## Sample 19 - 42869 reads in 28277 unique sequences. ## Sample 20 - 31780 reads in 21109 unique sequences. ## Sample 21 - 41246 reads in 31022 unique sequences. ## Sample 22 - 33175 reads in 24219 unique sequences. ## Sample 23 - 32469 reads in 23696 unique sequences. ## Sample 24 - 32221 reads in 24015 unique sequences. Merge reads and create table # !Long Operation mergers <- mergePairs(dadaFs, filtFs, dadaRs, filtRs, verbose=TRUE)</pre> ## 15729 paired-reads (in 416 unique pairings) successfully merged out of 27255 (in 4019 pairings) input. ## 22924 paired-reads (in 625 unique pairings) successfully merged out of 30302 (in 3396 pairings) input. ## 19335 paired-reads (in 524 unique pairings) successfully merged out of 27027 (in 3343 pairings) input. ## 26430 paired-reads (in 702 unique pairings) successfully merged out of 34157 (in 3582 pairings) input. ## 27267 paired-reads (in 730 unique pairings) successfully merged out of 34815 (in 3483 pairings) input. ## 16156 paired-reads (in 409 unique pairings) successfully merged out of 28750 (in 4321 pairings) input. ## 19847 paired-reads (in 475 unique pairings) successfully merged out of 21816 (in 1222 pairings) input. ## 14224 paired-reads (in 389 unique pairings) successfully merged out of 16302 (in 1077 pairings) input. ## 16700 paired-reads (in 416 unique pairings) successfully merged out of 18925 (in 1182 pairings) input. ## 17225 paired-reads (in 415 unique pairings) successfully merged out of 18993 (in 1045 pairings) input. ## 22615 paired-reads (in 620 unique pairings) successfully merged out of 36917 (in 5423 pairings) input. ## 11775 paired-reads (in 329 unique pairings) successfully merged out of 21544 (in 3228 pairings) input. ## 12074 paired-reads (in 349 unique pairings) successfully merged out of 22312 (in 3288 pairings) input. ## 9115 paired-reads (in 236 unique pairings) successfully merged out of 17755 (in 2532 pairings) input. ## 14492 paired-reads (in 393 unique pairings) successfully merged out of 25241 (in 3708 pairings) input. ## 9929 paired-reads (in 281 unique pairings) successfully merged out of 18440 (in 2615 pairings) input. ## 26537 paired-reads (in 764 unique pairings) successfully merged out of 38769 (in 4673 pairings) input. ## 19792 paired-reads (in 566 unique pairings) successfully merged out of 28803 (in 3524 pairings) input. ## 23682 paired-reads (in 611 unique pairings) successfully merged out of 36076 (in 4670 pairings) input. ## 17117 paired-reads (in 533 unique pairings) successfully merged out of 26351 (in 3344 pairings) input. ## 18224 paired-reads (in 443 unique pairings) successfully merged out of 33192 (in 5163 pairings) input. ## 14348 paired-reads (in 385 unique pairings) successfully merged out of 26975 (in 3871 pairings) input. ## 14376 paired-reads (in 386 unique pairings) successfully merged out of 26025 (in 3621 pairings) input. ## 13777 paired-reads (in 341 unique pairings) successfully merged out of 25769 (in 3546 pairings) input. seqtab <- makeSequenceTable(mergers)</pre> dim(seqtab) ## [1] 24 7004 Taxonomy annotation seqtab.nochim <- removeBimeraDenovo(seqtab, method="consensus", multithread=TRUE, verbose=TRUE)</pre> ## Identified 1948 bimeras out of 7004 input sequences. dim(seqtab.nochim) ## [1] 24 5056 sum(seqtab.nochim)/sum(seqtab) ## [1] 0.8053931 getN <- function(x) sum(getUniques(x))</pre> track <- cbind(out, sapply(dadaFs, getN), sapply(dadaRs, getN), sapply(mergers, getN), rowSums(seqtab.nochim))</pre> # If processing a single sample, remove the sapply calls: e.g. replace sapply(dadaFs, getN) with getN(dadaFs) colnames(track) <- c("input", "filtered", "denoisedF", "denoisedR", "merged", "nonchim")</pre> rownames(track) <- sample.names</pre> track input filtered denoisedF denoisedR merged nonchim ## Abacumov-B-1 43307 36622 29113 32777 15729 14462 ## Abacumov-B-13 41438 34160 31668 32193 22924 17630 ## Abacumov-B-14 38072 31002 28282 29053 19335 15500 ## Abacumov-B-15 46395 37918 35599 35901 26430 19483 ## Abacumov-B-16 47274 38962 36181 36939 27267 19850 30707 ## Abacumov-B-2 45972 39386 35254 16156 14720 ## Abacumov-B-25 29444 22676 22213 22195 19847 11714 ## Abacumov-B-26 21786 17052 16638 16612 14224 8413 ## Abacumov-B-27 25002 19659 19210 19247 16700 9839 ## Abacumov-B-28 25214 19501 19192 19214 17225 9896 ## Abacumov-B-3 54999 47256 39718 20250 42453 22615 ## Abacumov-B-37 31738 26915 22832 24440 11775 ## Abacumov-B-38 33322 27806 23564 25429 12074 10827 ## Abacumov-B-39 27048 22885 20676 9115 8157 30465 14492 ## Abacumov-B-4 40115 34065 26982 13500 ## Abacumov-B-40 28630 24332 19694 21767 9929 ## Abacumov-B-49 54922 45257 40839 41993 26537 21718 ## Abacumov-B-50 41565 34229 30510 31488 19792 17003 ## Abacumov-B-51 50422 42869 38023 39726 23682 19938 ## Abacumov-B-52 38603 31780 27870 29113 17117 14984 ## Abacumov-B-61 48250 41246 34823 38016 18224 16231 ## Abacumov-B-62 39777 33175 28334 30630 14348 12889 ## Abacumov-B-63 38230 32469 27559 29584 14376 12724 ## Abacumov-B-64 37920 32221 27196 29463 13777 12192 Merging leads to losses in reads. Re-run more relaxed filtering taxa <- assignTaxonomy(seqtab.nochim, "/home/nastasista/Metagenomics/silva\_nr\_v132\_train\_set.fa.gz", multithread= TRUE) taxa.print <- taxa # Removing sequence rownames for display only rownames(taxa.print) <- NULL</pre> head(taxa.print) Phylum Class Kingdom Family Genus ## [1,] "Bacteria" "Chloroflexi" "AD3" ## [5,] "Bacteria" "Cyanobacteria" "Oxyphotobacteria" "Nostocales" NA NA ## [6,] "Bacteria" "Cyanobacteria" "Oxyphotobacteria" "Nostocales" NA rownames(metadata) <- metadata\$Filename</pre> ps <- phyloseq(otu\_table(seqtab.nochim, taxa\_are\_rows=FALSE),</pre> sample\_data(metadata), tax\_table(taxa)) ## phyloseq-class experiment-level object ## otu\_table() OTU Table: [ 5056 taxa and 24 samples ] ## sample\_data() Sample Data: [ 24 samples by 6 sample variables ] ## tax\_table() Taxonomy Table: [ 5056 taxa by 6 taxonomic ranks ] sample\_names(ps) ## [1] "Abacumov-B-1" "Abacumov-B-13" "Abacumov-B-14" "Abacumov-B-15" ## [5] "Abacumov-B-16" "Abacumov-B-2" "Abacumov-B-25" "Abacumov-B-26" ## [9] "Abacumov-B-27" "Abacumov-B-28" "Abacumov-B-3" "Abacumov-B-37" ## [13] "Abacumov-B-38" "Abacumov-B-39" "Abacumov-B-4" "Abacumov-B-40" ## [17] "Abacumov-B-49" "Abacumov-B-50" "Abacumov-B-51" "Abacumov-B-52" ## [21] "Abacumov-B-61" "Abacumov-B-62" "Abacumov-B-63" "Abacumov-B-64" Rename phyloseq-object according to our needs metadata Filename Source Site Horizont Repeat ## Abacumov-B-1 Abacumov-B-1 Self-growing Dumps B1 ## Abacumov-B-2 Abacumov-B-2 Self-growing Dumps B1 AY ## Abacumov-B-3 Abacumov-B-3 Self-growing Dumps B1 AY 3 ## Abacumov-B-4 Abacumov-B-4 Self-growing Dumps B1 AY 4 ## Abacumov-B-13 Abacumov-B-13 Litostrat B2 C 1 ## Abacumov-B-14 Abacumov-B-14 Litostrat B2 C ## Abacumov-B-15 Abacumov-B-15 Litostrat B2 ## Abacumov-B-16 Abacumov-B-16 Litostrat B2 ## Abacumov-B-25 Abacumov-B-25 Coal Mine Terricon B3 ## Abacumov-B-26 Abacumov-B-26 Coal Mine Terricon B3 ## Abacumov-B-27 Abacumov-B-27 Coal Mine Terricon B3 ## Abacumov-B-28 Abacumov-B-28 Coal Mine Terricon B3 ## Abacumov-B-37 Abacumov-B-37 Local Reference B4 ## Abacumov-B-38 Abacumov-B-38 Local Reference B4 ## Abacumov-B-39 Abacumov-B-39 Local Reference B4 ΑY ## Abacumov-B-40 Abacumov-B-40 Local Reference B4 ΑY ## Abacumov-B-49 Abacumov-B-49 Embryo Sand B5 ΑY 1 ## Abacumov-B-50 Abacumov-B-50 Embryo Sand B5 ΑY ## Abacumov-B-51 Abacumov-B-51 Embryo Sand B5 ΑY ## Abacumov-B-52 Abacumov-B-52 Embryo Sand B5 ΑY AY 1 ## Abacumov-B-61 Abacumov-B-61 Regional Reference B6 ## Abacumov-B-62 Abacumov-B-62 Regional Reference B6 AY ## Abacumov-B-63 Abacumov-B-63 Regional Reference B6 ΑY ## Abacumov-B-64 Abacumov-B-64 Regional Reference B6 ## Abacumov-B-1 Self-growing Dumps.B1.AY.1 ## Abacumov-B-2 Self-growing Dumps.B1.AY.2 ## Abacumov-B-3 Self-growing Dumps.B1.AY.3 ## Abacumov-B-4 Self-growing Dumps.B1.AY.4 ## Abacumov-B-13 Litostrat.B2.C.1 ## Abacumov-B-14 Litostrat.B2.C.2 ## Abacumov-B-15 Litostrat.B2.C.3 ## Abacumov-B-16 Litostrat.B2.C.4 ## Abacumov-B-25 Coal Mine Terricon.B3.C.1 ## Abacumov-B-26 Coal Mine Terricon.B3.C.2 ## Abacumov-B-27 Coal Mine Terricon.B3.C.3 ## Abacumov-B-28 Coal Mine Terricon.B3.C.4 ## Abacumov-B-37 Local Reference.B4.AY.1 ## Abacumov-B-38 Local Reference.B4.AY.2 ## Abacumov-B-39 Local Reference.B4.AY.3 ## Abacumov-B-40 Local Reference.B4.AY.4 ## Abacumov-B-49 Embryo Sand.B5.AY.1 ## Abacumov-B-50 Embryo Sand.B5.AY.2 ## Abacumov-B-51 Embryo Sand.B5.AY.3 ## Abacumov-B-52 Embryo Sand.B5.AY.4 ## Abacumov-B-61 Regional Reference.B6.AY.1 ## Abacumov-B-62 Regional Reference.B6.AY.2 ## Abacumov-B-63 Regional Reference.B6.AY.3 ## Abacumov-B-64 Regional Reference.B6.AY.4 ## Rename Samples new.names <- ps@sam\_data %>% data.frame() %>% dplyr::select(Filename, SampleID) %>% arrange(Filename, levels = sample\_names(ps)) if (all(sample\_names(ps) == new.names\$Filename)) { sample\_names(ps) <- ps@sam\_data\$SampleID</pre> print("Renamed") ## [1] "Renamed" sample\_names(ps) ## [1] "Self-growing Dumps.B1.AY.1" "Litostrat.B2.C.1" ## [3] "Litostrat.B2.C.2" "Litostrat.B2.C.3" ## [5] "Litostrat.B2.C.4" "Self-growing Dumps.B1.AY.2" ## [7] "Coal Mine Terricon.B3.C.1" "Coal Mine Terricon.B3.C.2" ## [9] "Coal Mine Terricon.B3.C.3" "Coal Mine Terricon.B3.C.4" ## [11] "Self-growing Dumps.B1.AY.3" "Local Reference.B4.AY.1" ## [13] "Local Reference.B4.AY.2" "Local Reference.B4.AY.3" ## [15] "Self-growing Dumps.B1.AY.4" "Local Reference.B4.AY.4" ## [17] "Embryo Sand.B5.AY.1" "Embryo Sand.B5.AY.2" ## [19] "Embryo Sand.B5.AY.3" "Embryo Sand.B5.AY.4" ## [21] "Regional Reference.B6.AY.1" "Regional Reference.B6.AY.2" ## [23] "Regional Reference.B6.AY.3" "Regional Reference.B6.AY.4" dna <- Biostrings::DNAStringSet(taxa\_names(ps))</pre> names(dna) <- taxa\_names(ps)</pre> ps <- merge\_phyloseq(ps, dna)</pre> taxa\_names(ps) <- paste0("ASV", seq(ntaxa(ps)))</pre> Save phyloseq-object and aquire checksum saveRDS(ps, "ps.RData") ps <- readRDS("ps.RData")</pre>

tools::md5sum("ps.RData")

## "9767b2490c215ecc89058c1d259db9bd"

ps.RData

**Dada2 Processing** 

knitr::opts\_chunk\$set(fig.width=14, fig.height=8)

Anastasia Poluzerova

library('dada2')
library('phyloseq')
library('dplyr')

2023-06-07