CS 109A/Final Project: Data Processing and EDA

Group 3: Ning Shen, Lars von Buchholtz

Alzheimer's disease (AD) is characterized by 2 major diagnostic features: pathological changes in the brain namely beta-amyloid placques and deterioration of the mental state leading to cognitive impairment and ultimately dementia. Neither of these features is sufficient but both are necessary for a definitive AD diagnosis. While beta-amyloid placques are present very early during disease progression, cognitive impairment only manifests in the latest stages of the disease when a treatment is deemed impossible. In this project, we will focus on these 2 features separately and 1) explore if gene expression profile taken from patients blood can predict beta-amyloid level from CSF; and 2) try to predict the mental state from brain imaging data. These 2 parts are inspired by subchallenges 2 and 3 of the ADNI Big Data DREAM Challenge.

Part 1: EDA on gene expression data combined with AD Dream Challenge Subchallenge 2

Predicting CSF beta-amyloid level is one subchallenge in the 2014 DREAM challenge (https://www.synapse.org/#!Synapse:syn2290704/wiki/60828

(https://www.synapse.org/#!Synapse:syn2290704/wiki/60828)). However, all participating groups for this subchallenged failed in predicting beta-amyloid levels given the data provided. However, the ADNI database provides more information than what's included in the subchallenge training data. After reviewing the data available in ADNI database, we decided to include microarray gene expression data to predict beta-amyloid levels. The gene expression data are taken from blood samples of the sample patients, and available for download at

(https://ida.loni.usc.edu/pages/access/geneticData.isp

(https://ida.loni.usc.edu/pages/access/geneticData.jsp)). Predicting beta-amyloid levels in the cerebrospinal fluid (CSF) from gene expression in blood samples would provide a much less invasive way to diagnose amyloid placques. Furthermore, the identified signature genes could be used as clinical biomarkers.

```
In [1]: # import required modules
        import warnings
        warnings.filterwarnings('ignore')
        import numpy as np
        import pandas as pd
        import matplotlib
        import matplotlib.pyplot as plt
        import sklearn.metrics as metrics
        from scipy import stats
        from sklearn.model selection import cross val score
        from sklearn.model selection import GridSearchCV
        from sklearn.model_selection import train_test_split
        import statsmodels.api as sm
        from statsmodels.api import OLS
        from sklearn.model selection import KFold
        from sklearn.decomposition import PCA
        from sklearn.linear model import LinearRegression
        from sklearn.linear model import Ridge
        from sklearn.linear_model import RidgeCV
        from sklearn.linear model import ElasticNetCV
        from sklearn.linear model import ElasticNet
        from sklearn.utils import resample
        from sklearn.model_selection import cross_val_score
        from sklearn.metrics import accuracy score
        from sklearn.metrics import r2 score
        from sklearn.ensemble import GradientBoostingRegressor
        from sklearn.ensemble import RandomForestRegressor
        from scipy.stats import pearsonr
        from scipy.stats import ttest 1samp
        from sklearn.linear_model import LogisticRegression
        from sklearn.linear model import LogisticRegressionCV
        import seaborn.apionly as sns
        sns.set context("poster")
        from IPython.display import display
        matplotlib.style.use('ggplot')
        %matplotlib inline
```

1. Cleaning gene expression data and merge with Dream data

The gene expression data provided in ADNI contains information about gene locus, ~ 49,000 gene expression levels, gene annotation, phase, visit, year of collection etc. This dataset provides rich information but is not well formated. In order to add the gene expression data to the original DREAM challenge data, the gene expression dataframe had to be cleaned and transposed first. After the cleaning of gene expression data, the converted table merged with the Dream challenge table, generated a well formated table for following EDA.

In []: import zipfile
 zip = zipfile.ZipFile(r'ADNI_Gene_Expression_Profile.zip')
 zip.extractall(r'.')

In [2]: # Read in the microarray expression data
 file_expression = pd.read_csv("ADNI_Gene_Expression_Profile.csv", header=None
 file_expression.describe()

Out[2]:

	1	2	3	4	5	6	7	8	
count	43789	48158	49394.000	49394.000	49394.00	49394.000	49394.000	49394.000	49394.00
unique	18725	20093	10122.000	10181.000	10157.00	10123.000	10128.000	10099.000	10154.00
top	LOC4763	NF1	2.269	2.129	2.27	2.288	2.293	2.377	2.19
freq	16	19	111.000	123.000	84.00	93.000	88.000	120.000	142.00

4 rows × 747 columns

In [3]: # Original data format
 file expression.head(10)

1

2

3

Out[3]:

0							
Phase	NaN	NaN	ADNIGO	ADNI2	ADNI2	ADNIGO	ADN
Visit	NaN	NaN	m48	v03	v03	m48	v(
SubjectID	NaN	NaN	116_S_1249	037_S_4410	006_S_4153	116_S_1232	099_S_420
260/280	NaN	NaN	2.05	2.07	2.04	2.03	2.(
260/230	NaN	NaN	0.55	1.54	2.1	1.52	1
RIN	NaN	NaN	7.7	7.6	7.2	6.8	7
Affy Plate	NaN	NaN	7	3	6	7	
YearofCollection	NaN	NaN	2011	2012	2011	2011	201
ProbeSet	LocusLink	Symbol	NaN	NaN	NaN	NaN	Na
11715100_at							
	LOC8355	HIST1H3G	2.237	2.294	2.14	2.062	2.0

5

6

7

10 rows × 747 columns

```
In [4]: ## Set column names as subjectID
    col_names = file_expression.loc['SubjectID']
    col_names[0:2]=['LocusLink','Symbol']
    file_expression.columns = list(col_names)
    file_expression.head()
```

Out[4]:

LocusLink Symbol 116_S_1249 037_S_4410 006_S_4153 116_S_1232 099_S_4205 007_

0

Phase	NaN	NaN	ADNIGO	ADNI2	ADNI2	ADNIGO	ADNI2	
Visit	NaN	NaN	m48	v03	v03	m48	v03	
SubjectID	LocusLink	Symbol	116_S_1249	037_S_4410	006_S_4153	116_S_1232	099_S_4205	007_
260/280	NaN	NaN	2.05	2.07	2.04	2.03	2.01	
260/230	NaN	NaN	0.55	1.54	2.1	1.52	1.6	

5 rows × 747 columns

Out[5]:

	Phase	Visit	PTID	260/280	260/230	RIN	Affy Plate	YearofCollection	ProbeSet	
116_S_1249	ADNIGO	m48	116_S_1249	2.05	0.55	7.7	7	2011	NaN	
037_S_4410	ADNI2	v03	037_S_4410	2.07	1.54	7.6	3	2012	NaN	
006_S_4153	ADNI2	v03	006_S_4153	2.04	2.1	7.2	6	2011	NaN	
116_S_1232	ADNIGO	m48	116_S_1232	2.03	1.52	6.8	7	2011	NaN	
099_S_4205	ADNI2	v03	099_S_4205	2.01	1.6	7.9	9	2011	NaN	

5 rows × 49395 columns

In [6]: transformed_table.describe()

Out[6]:

	Phase	Visit	PTID	260/280	260/230	RIN	Affy Plate	YearofCollection	ProbeSet	117151
count	744	744	744	744	744	744	744	744	0.0	_
unique	2	12	744	40	173	38	9	4	0.0	
top	ADNI2	v03	041_S_4427	2.03	1.65	7.1	3	2011	NaN	
freq	449	357	1	73	13	61	88	383	NaN	

4 rows × 49395 columns

Out[7]:

	RID	PTID	AGE	PTGENDER	PTEDUCAT	APOE4	MMSE	ABETA	SAGE.Q2	APOE Genotype
0	5	011_S_0005	73.7	Male	16	0	29	115.0	1	3,3
1	19	067_S_0019	73.1	Female	18	0	29	260.0	0	2,3
2	31	023_S_0031	77.7	Female	18	0	30	240.0	0	3,3
3	43	018_S_0043	76.2	Male	16	0	29	175.0	1	2,3
4	47	100_S_0047	84.7	Male	20	0	30	252.0	0	2,3

In [8]: dream_data.describe()

Out[8]:

	RID	AGE	PTEDUCAT	APOE4	MMSE	ABETA	SAGE.Q2
count	176.000000	176.000000	176.000000	176.000000	176.000000	176.000000	176.000000
mean	2543.375000	75.309091	16.255682	0.272727	29.028409	195.500000	0.460227
std	1874.399901	5.364692	2.600543	0.517737	1.239488	53.370559	0.499838
min	5.000000	62.000000	8.000000	0.000000	24.000000	75.000000	0.000000
25%	558.000000	71.775000	14.000000	0.000000	29.000000	149.950000	0.000000
50%	4032.500000	74.900000	16.000000	0.000000	29.000000	202.000000	0.000000
75%	4281.250000	78.400000	18.000000	0.000000	30.000000	240.000000	1.000000
max	4516.000000	89.600000	20.000000	2.000000	30.000000	302.800000	1.000000

Out[9]:

	RID	PTID	AGE	PTGENDER	PTEDUCAT	APOE4	MMSE	ABETA	SAGE.Q2	APOE Genotype
40	984	021_S_0984	76.6	Male	14	1	30	75.0	1	3,4
77	4179	033_S_4179	83.0	Male	20	2	30	82.7	1	4,4
95	4339	082_S_4339	84.3	Male	17	2	29	90.7	1	4,4
124	4474	031_S_4474	85.6	Male	18	0	28	92.5	1	3,3
94	4335	021_S_4335	71.7	Female	15	0	30	95.4	1	3,3

5 rows × 49404 columns

```
In [10]: data_common.shape
Out[10]: (130, 49404)
In [11]: data_common.columns[0:30]
```

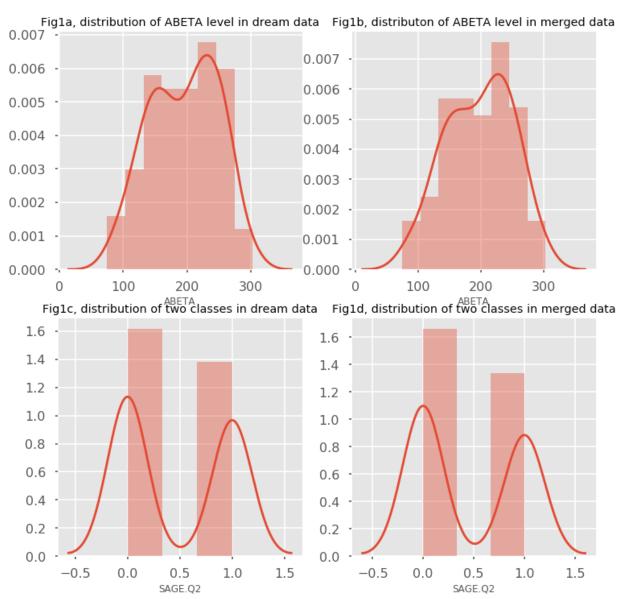
2. EDA

1) After data cleaning and merging, the first question we ask is: is amyloid beta level distribution bimodal in the data we work with? In addition, are the two classes that we are going to classify balanced?

Fig1 addresses this question, showing bimodal distribution in both the original dream data and the merged table and well-balanced data.

In [12]: # Check distribution of amyloid-beta 42 level (ABETA) and class (SAGE.Q2)
fig, axes = plt.subplots(ncols=2, nrows=2, figsize=(12,12))
sns.distplot(dream_data['ABETA'], ax=axes[0,0])
axes[0,0].set_title("Figla, distribution of ABETA level in dream data")
sns.distplot(data_common['ABETA'], ax=axes[0,1])
axes[0,1].set_title("Figlb, distribution of ABETA level in merged data")
sns.distplot(dream_data['SAGE.Q2'], ax=axes[1,0])
axes[1,0].set_title("Figlc, distribution of two classes in dream data")
sns.distplot(data_common['SAGE.Q2'], ax=axes[1,1])
axes[1,1].set_title("Figld, distribution of two classes in merged data")

Out[12]: <matplotlib.text.Text at 0x10d7d3400>

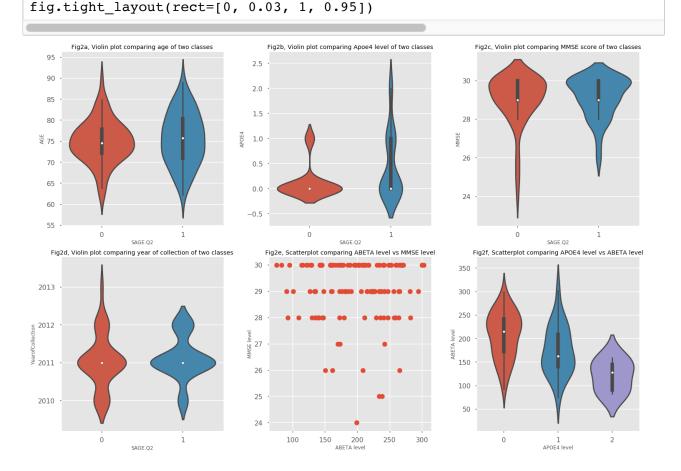


2) Is there any correlation/difference in the two classes with regard to age, apo4 level, MMSE score, and year of collection?

Fig2 addresses these questions. The Age of patients in class 0 are more concentrated around 75 compared to class 1 (Fig2a). The year of collection of patient samples in class 0 are more spread out than in class 1 (Fig2d). The Apoe4 level in class 0 is much more concentrated near 0, whereas class 1 shows more variation in both directions (Fig2b). When comparing Apoe4 level with ABETA level,

there is a clear negative correlation between the two (Fig2f). The MMSE score distribution in class 0 is lower than in class 1 (Fig2c). The scatterplot comparing ABETA level vs MMSE score (Fig2e) showed similar trend, implying high levels of beta-amyloid might correlate with lower cognition.

```
# Check distribution of amyloid-beta 42 level (ABETA) and class (SAGE.Q2)
In [13]:
         fig, axes = plt.subplots(ncols=3, nrows=2, figsize=(21,15))
         sns.violinplot(data_common['SAGE.Q2'], data_common['AGE'], ax=axes[0,0])
         axes[0,0].set_title("Fig2a, Violin plot comparing age of two classes")
         sns.violinplot(data common['SAGE.Q2'], data common['APOE4'], ax=axes[0,1])
         axes[0,1].set_title("Fig2b, Violin plot comparing Apoe4 level of two classes
         sns.violinplot(data_common['SAGE.Q2'], data_common['MMSE'], ax=axes[0,2])
         axes[0,2].set title("Fig2c, Violin plot comparing MMSE score of two classes'
         sns.violinplot(data common['SAGE.Q2'], data common.YearofCollection.convert
         axes[1,0].set_title("Fig2d, Violin plot comparing year of collection of two
         axes[1,1].scatter(data common['ABETA'],data common['MMSE'])
         axes[1,1].set_xlabel('ABETA level')
         axes[1,1].set ylabel('MMSE level')
         axes[1,1].set title("Fig2e, Scatterplot comparing ABETA level vs MMSE level"
         sns.violinplot(data_common['APOE4'], data_common['ABETA'], ax=axes[1,2])
         axes[1,2].set_ylabel('ABETA level')
         axes[1,2].set_xlabel('APOE4 level')
         axes[1,2].set_title("Fig2f, Scatterplot comparing APOE4 level vs ABETA level
```



3) How does high level look of gene expression profile differ between the two classes?

To address this question, we made a heatmap plot with each row correspond to a patient sample with class label on the side, each column correspond to a gene (We included 100 random genes in the plot for illustration purposes). If the gene expression profile of the two classes are distinctly different, we would expect the heatmap clustering on the y-axis (class bar on the left side of the heatmap) be very well separated between the two classes (red vs blue). The heatmap suggests the gene expression profiles between the two classes are not distinctly different based on this limited subset of genes. However, some of the samples do cluster close, showing thick red/blue class bars on the left, suggesting there might be potential that gene expression profile can help separate the two classes.

```
In [14]: ## Check gene expression profile difference by heatmap visualization

select_data = data_common[data_common.columns[19:100]]

select_data = select_data.convert_objects(convert_numeric=True)

select_data['class'] = data_common['SAGE.Q2']

exp_table = select_data.pop('class')

exp = dict(zip(exp_table.unique(), 'rbg'))

row_colors = exp_table.map(exp)

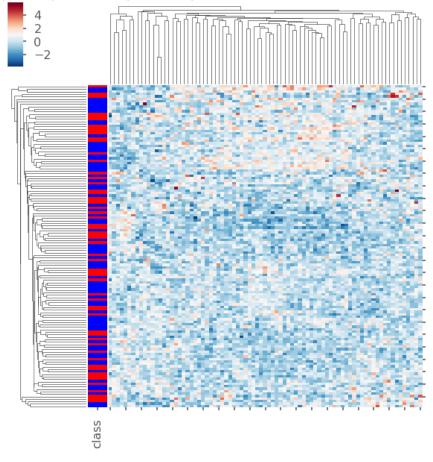
g = sns.clustermap(select_data, row_colors=row_colors, z_score=1, cmap="RdBuplt.setp(g.ax_heatmap.set_yticklabels(''));

plt.setp(g.ax_heatmap.set_xticklabels(''));

plt.title("Fig3. Heatmap comparing gene expression profile across patient sa
```

Out[14]: <matplotlib.text.Text at 0x1150fd278>





The basic data cleaning and EDA suggested gene expression profile might be able to help classifying the ABETA groups. In order to confirm this, further feature selection and dimension reduction like PCA need to be applied for modeling. For the modeling part, this can be pursued as either classification or regression. However, due the limitation of time and effort, we decide to focus the modeling the part 2 of the project - predicting mental scores from MRI image data.

PART 2: Predicting mental state (MMSE) from brain imaging data (MRI)

In the second part of the project, we focus on the question if we can predict the mental state of a patient from brain imaging data. In the ADNI dataset, the mental state is represented by the Mini-Mental State Exam (MMSE) score which is the variable that we are trying to predict in this part of the project. Since the psychological pathology develops relatively late in the disease progression, it would be advantageous to be able to predict the mental state from MRI brain imaging data which is routinely acquired in clinical settings (e.g. to exclude other types of dementia). In our analysis, we follow closely the the guidelines given in the AD Big Data Dream Challenge Subchallenge 3 (https://www.synapse.org/#!Synapse:syn2290704/wiki/64635)).

We downloaded the data for this challenge from https://ida.loni.usc.edu/pages/access/studyData.jsp?categoryId=43&subCategoryId=94 (password-protected login).

Initial Data Exploration: The data in the baseline_data.csv file consists of 628 rows corresponding to unique observations of 628 patients. It contains 2150 measurements of brain geometry derived from 3D MRI images, e.g. area, thickness, curvature, etc. of different brain regions. It can be assumed that many of these variables are correlated with each other because of geometrical necessity. The spreadsheeet also contains demographic data such as education, ethnicity, gender, race and age as well as some diagnostic data: MMSE score, Diagnosis, Apoe4 genotype. The Apoe4 allele is a polymorphism of the Apo E gene that is associated with AD susceptibility. In addition, there is an indicator variable for Apoe4 imputation.

```
In [15]: # we define a function to calculate the Concordance Correlation Coefficient
def ccc_function(y,y_hat):
    pcc, _ = pearsonr(y_hat,y)
    ccc = 2 * pcc * y.std() * y_hat.std()/(y.var() + y_hat.var() + (y.mean() return ccc

In [16]: # function to evaluate our quantitative models and store the result in a sur def evaluate_and_store(y_train,y_hat_train,y_test,y_hat_test,name,summary):
    # input:
```

```
In [16]: # function to evaluate our quantitative models and store the result in a sur
def evaluate_and_store(y_train,y_hat_train,y_test,y_hat_test,name,summary):
    # input:
    # y_train,y_hat_train,y_test,y_hat_test
    # summary: dictionary with fields model name, R2train, R2test, PCC and (
    pcc, _ = pearsonr(y_test,y_hat_test)
    ccc = ccc_function(y_test,y_hat_test)
    r2train = r2_score(y_train,y_hat_train)
    r2test = r2_score(y_test,y_hat_test)
    summary['model name'].append(name)
    summary['R2train'].append(r2train)
    summary['R2test'].append(r2test)
    summary['PCC'].append(pcc)
    summary['CCC'].append(ccc)
    return r2train,r2test,pcc,ccc,summary

# set up a summary dictionary
summary = {'model name':[],'R2train':[],'R2test':[],"PCC":[],'CCC':[]}
```

Data Cleaning and EDA

We load the data and have a look at the imaging part and the demographic part.

```
In [17]: #read data from file
    data = pd.read_csv('baseline_data.csv')
    print(data.shape)
# a lot more predictors than observations, lots of potential for overfitting

print(len(data.RID.unique()))
# no doubles in patient ID, so we can keep rows as patients/observations

#split data into image data and demographic data
    image_data = data.iloc[:,:-13].copy()
    demographic = data.iloc[:,-13:].copy()
    mmse = data['MMSE']
# for diagnosis: CN = 0, LMCI = 1, AD = 2
    dx = (data['DX.bl'] == 'LMCI') * 1. + (data['DX.bl'] == 'AD') * 2.

display(image_data.head())
    display(demographic.head())
```

(628, 2163) 628

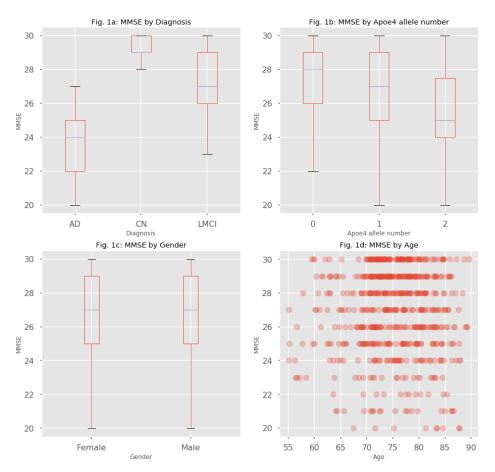
	area.1002	area.1003	area.1005	area.1006	area.1007	area.1008	area.1009	a
0	612.577638	2514.366512	1652.050796	731.718725	3794.131041	5012.960558	4951.994756	5666
1	735.292087	2435.629408	1947.966106	721.230657	4101.035394	4469.814924	4002.936490	5427
2	1080.976588	2190.801306	1613.620315	636.078912	5146.969073	6192.609394	4640.889149	685₄
3	840.850798	2293.601605	1599.807666	729.344575	3351.924971	4231.417941	3991.795466	5047
4	592.882184	1827.195664	1479.821407	535.558408	3459.934118	5063.103074	3583.954659	4120

5 rows × 2150 columns

	directory.id	Subject	RID	Image.Data.ID	DX.bl	AGE	PTGENDER	Р
0	178eeac87ff2460568a8709ba32f9b1e	002_S_0295	295	45108	CN	84.8	Male	_
1	4d953ce78fb484052e8b735e5493770a	002_S_0413	413	45117	CN	76.3	Female	
2	da022a5df80d136d3aa26bfa06702278	002_S_0619	619	48617	AD	77.5	Male	
3	7d700a43e372c9dfabd6c2dfdc0edcf8	002_S_0685	685	40683	CN	89.6	Female	
4	c033f6026da1179ab7b5f7e4f5559f59	002_S_0729	729	40708	LMCI	65.1	Female	

```
In [18]: fig1 = plt.figure(figsize = [15,15])
         plt.subplot(2,2,1)
         ax = plt.gca()
         data.boxplot('MMSE',by = ['DX.bl'], ax = ax)
         plt.title('Fig. 1a: MMSE by Diagnosis')
         plt.xlabel('Diagnosis')
         plt.ylabel('MMSE')
         plt.subplot(2,2,2)
         ax = plt.gca()
         data.boxplot('MMSE',by = ['APOE4'], ax = ax)
         plt.title('Fig. 1b: MMSE by Apoe4 allele number')
         plt.xlabel('Apoe4 allele number')
         plt.ylabel('MMSE')
         plt.subplot(2,2,3)
         ax = plt.gca()
         data.boxplot('MMSE',by = ['PTGENDER'], ax = ax)
         plt.title('Fig. 1c: MMSE by Gender')
         plt.xlabel('Gender')
         plt.ylabel('MMSE')
         plt.subplot(2,2,4)
         ax = plt.gca()
         plt.scatter(data['AGE'], mmse, alpha=0.3)
         plt.title('Fig. 1d: MMSE by Age')
         plt.xlabel('Age')
         plt.ylabel('MMSE');
         fig1.suptitle('Fig. 1: EDA of MMSE Scores with Respect to Diagnosis and Demo
```

Fig. 1: EDA of MMSE Scores with Respect to Diagnosis and Demographic Factors



There is a clear correlation of MMSE score with the diagnosis of the patient. Healthy control patients (CN) have an MMSE score near the maximum of 30, whereas patients with light cognitive impairment (LMCI) have a lower median MMSE and Alzheimer's disease patients (AD) have the lowest median MMSE score. This indicates that MMSE can be used to predict Alzheimer's disease or dementia in general.

The MMSE score is also strongly correlated with the Apoe4 genotype. In particular, individuals that are homozygous for the Apoe4 allele, i.e. they have 2 copies of it, display a lower MMSE score. On the other hand, other demographic predictors like gender or age do not appear to be strongly correlated with the MMSE score.

We now try to find the features in the imaging data that are most correlated with the MMSE score:

```
In [19]: # find the image data columns that are most correlated with mmse

corr_coefs = []
pvals = []

for column in image_data.columns.values:
        coef,pval = pearsonr(image_data[column],mmse)
        corr_coefs.append(coef)
        pvals.append(pval)

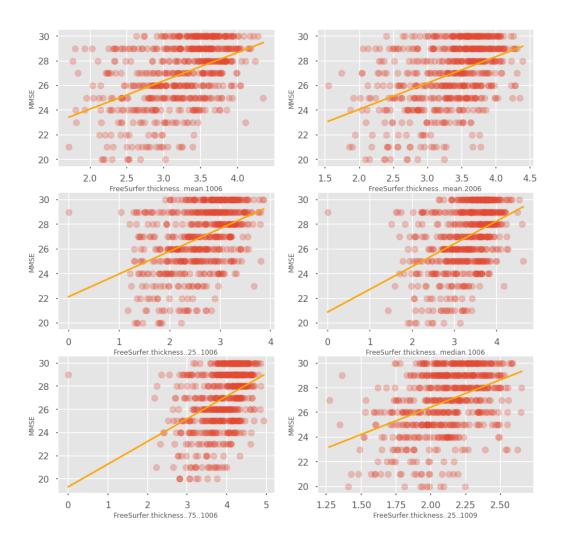
print("image measurements most closely correlated with mmse score:")
best6 = image_data.columns.values[np.argsort(pvals)[:6]]
print(best6)
```

```
image measurements most closely correlated with mmse score:
['FreeSurfer.thickness..mean.1006' 'FreeSurfer.thickness..mean.2006'
'FreeSurfer.thickness..25..1006' 'FreeSurfer.thickness..median.1006'
'FreeSurfer.thickness..75..1006' 'FreeSurfer.thickness..25..1009']
```

```
In [20]: # plot mmse vs most closely correlated image measurements
fig2 = plt.figure(figsize= [15,15])

for i,feature in enumerate(best6):
    plt.subplot(3,2,i+1)
    plt.scatter(image_data[feature],mmse,alpha=0.3)
    plt.xlabel(feature)
    plt.ylabel('MMSE')
    betal,beta0 = np.polyfit(image_data[feature],mmse,deg=1)
    f = lambda x: beta1 * x + beta0
    x = np.array([np.min(image_data[feature]), np.max(image_data[feature])])
    plt.plot(x,f(x),c='orange')
    sns.despine()
fig2.suptitle('Fig. 2: EDA of Image Features most closely correlated with MM
```

Fig. 2: EDA of Image Features most closely correlated with MMSE score



Some of the features in the imaging data appear to be strongly correlated with the MMSE score which makes the analysis/prediction promising. We also notice that we now have 2159 predictors which is a lot more than the 628 observations we have in our complete dataset. This causes a big danger of overfitting any training data.

We next clean up the demographic variables and split our data into training and test sets. We group all the predictors (demographic + MRI imaging) together and extract 2 outcome variables: 1.) the

MMSE score which we want to predict, and 2.) the diagnosis which we will use to evaluate the usefulness of our MMSE prediction in the last part.

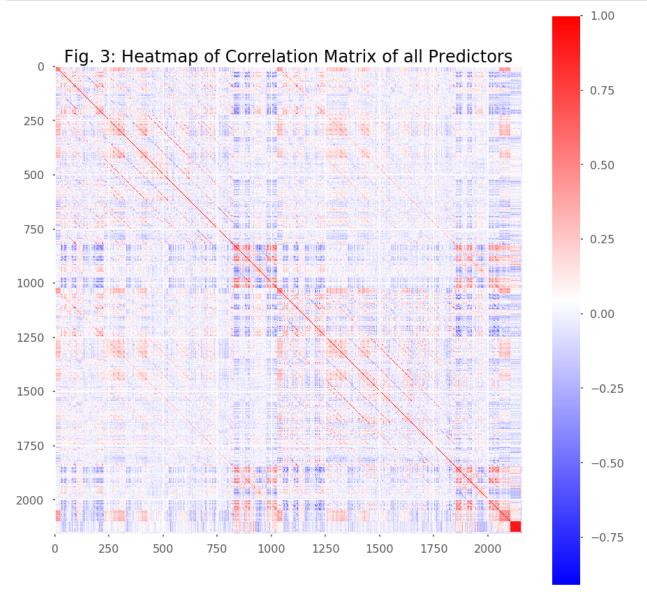
Then we will check if we have multi-colinearity in the imaging predictors.

```
In [21]: # clean up demographic variables
         # impute Unknown as Not Hisp/Latino and turn into binary with Hisp/Latino as
         demographic['PTETHCAT'] = (demographic.PTETHCAT == 'Hisp/Latino') * 1.
          # race and number of Apoe4 alleles are turned into dummy variables
          demographic = pd.get_dummies(demographic,columns=['PTRACCAT','APOE4'],drop_f
          # turn gender into a binary variable with Female = 1.0
          demographic['PTGENDER'] = (demographic.PTGENDER == 'Female') * 1.
          # change imputed genotype from boolean to 0.0/1.0
         demographic['imputed_genotype'] = (demographic.imputed_genotype == True)* 1.
         # get X and y ; we have 2 different outcome variables: y = MMSE score for the
         # y dx = diagnosis for evaluating our predicted MMSE in the last part
         y = mmse
         y_dx = dx
         image_columns = image_data.columns.values
          \# we discard the ID columns and the diagnosis column which would give away \pi
         demographic columns = ['AGE', 'PTGENDER', 'PTEDUCAT', 'PTETHCAT', 'imputed genot
         # and concatenate the image and remaining demographic data
         X = pd.concat([image data,demographic[demographic columns]],axis=1)
         print('number of observations: ',X.shape[0])
         print('number of predictors: ',X.shape[1])
         #check that there is no NaN's in the data
         print('number of NULL values: ',np.sum(np.sum(X.isnull())))
         # no NaN's
         # train test split: we have 2 different outcome variables: y = MMSE score for
          # y dx = diagnosis for evaluating our predicted MMSE in the last part
         X train, X test, y train, y test, y dx train, y dx test = train test split()
          # standardize training and test
         mean = X train.mean(axis=0)
         std = X train.std(axis=0)
         X train = (X train - mean)/std
         X \text{ test} = (X \text{ test} - \text{mean})/\text{std}
```

```
number of observations: 628
number of predictors: 2159
number of NULL values: 0
```

Checking Multi-Colinearity in the Imaging Data

We calculate a correlation matrix between the predictors and display it as a heatmap.



There is a lot of colinearity between the predictors in the imaging data. Colinear variables appear to be organized in ordered blocks. In order to get a working prediction model, we will have to reduce the dimensionality and co-linearity to reduce the danger of overfitting to the training set.

Baseline Model: Simple Linear Regression

We first try a simple linear regression model with all predictors and MMSE as the outcome.

```
In [23]:
    # try a super-simple linear regression model on all predictors
    simple_est = LinearRegression(fit_intercept = True)
    simple_est.fit(X_train,y_train)
    y_hat_train = simple_est.predict(X_train)
    y_hat_test = simple_est.predict(X_test)

    name = 'linear regression with all predictors'
    r2train, r2test, pcc, ccc, summary = evaluate_and_store(y_train,y_hat_train,
    print(name,': ')
    print('R2 score on training set: ',r2train)
    print('R2 score on validation set: ',r2test)
    print("Pearson's correlation coefficient between estimate and ground truth(t)
    print("Concordance correlation coefficient between estimate and ground truth)
```

```
linear regression with all predictors:
R2 score on training set: 1.0
R2 score on validation set: -0.575703582103
Pearson's correlation coefficient between estimate and ground truth(test set) 0.292637073937
Concordance correlation coefficient between estimate and ground truth(test set) 0.29124810158
```

Because of the large number of predictors and the multi-colinearity of the predictors, the model with all predictors suffers from high variance. Overall, variance appears to be the biggest problem in predicting this dataset. We will use principal components analysis, stepwise feature selection and ensemble strategies to reduce the dimensionality and eliminate colinearity. In addition, we will use regularization to further reduce variance problems.

Principal Component Analysis and Regularized Linear Regression

First, we reduce the dimensionality by principal component analysis and keep enough principal components that capture 90% of the variance in the data.

```
In [24]: # PCA transformation with PCs that account for 90% of variance in training s
    fullpca = PCA(n_components = 0.9,svd_solver = 'full')
    X_train_pca = fullpca.fit_transform(X_train)
    X_test_pca = fullpca.transform(X_test)
    total_pcs = X_train_pca.shape[1]
    print("Number of PCs accounting for 90% of variance: ", total_pcs)
```

Number of PCs accounting for 90% of variance: 197

We still have 197 predictors left after principal component analysis. Let's try a simple linear regression model:

```
In [25]: # try a simple linear regression model without regularization on all 197 PCs
    pca_lin_est = LinearRegression(fit_intercept = True)
    pca_lin_est.fit(X_train_pca,y_train)
    y_hat_train = pca_lin_est.predict(X_train_pca)
    y_hat_test = pca_lin_est.predict(X_test_pca)

name = 'linear regression on {} PCs'.format(total_pcs)
    r2train, r2test, pcc, ccc, summary = evaluate_and_store(y_train,y_hat_train,
    print(name,': ')
    print('R2 score on training set: ',r2train)
    print('R2 score on validation set: ',r2test)
    print("Pearson's correlation coefficient between estimate and ground truth(total_print("Concordance correlation coefficient between estimate and ground truth(total_print("Concordance correlation coefficient between estimate and ground truth(total_print("Concordance correlation coefficient between estimate and ground truth()
```

```
linear regression on 197 PCs:
R2 score on training set: 0.607144507434
R2 score on validation set: 0.146129866785
Pearson's correlation coefficient between estimate and ground truth(test set) 0.462698583976
Concordance correlation coefficient between estimate and ground truth(test set) 0.434770218566
```

We still have a relatively high discrepancy between performance on the training and test set which is indicative of variance problems. Even though we have eliminated co-linearity between predictors through PCA, this does not mean that each of our predictors is correlated with our outcome variable MMSE. We will try different types of regularization to reduce variance and improve our bias/variance trade-off.

We first try ridge regularization with cross-validation to identify the ideal regularization weight hyperparameter, then train on the whole training set with the optimal parameter.

```
In [26]:
         # try ridge regularization
         alphas = [100000, 50000, 10000, 5000, 1000, 500, 100, 50, 10, 5, 1, 0.5, 0.1, 0.05, 0.01]
         ridge pca est = RidgeCV(alphas=alphas,fit_intercept = True,cv=5)
         ridge pca est.fit(X train pca,y train)
         best_alpha = ridge_pca_est.alpha_
         # now train on whole training set with best alpha
         ridge pca est = Ridge(alpha=best alpha,fit intercept = True)
         ridge pca est.fit(X train pca,y train)
         y hat train = ridge pca est.predict(X train pca)
         y hat test = ridge pca est.predict(X test pca)
         name = 'ridge regression with {} PCs'.format(total pcs)
         r2train, r2test, pcc, ccc, summary = evaluate and store(y_train,y_hat_train,
         print(name,': ')
         print('Regularization parameter: ', best_alpha)
         print('R2 score on training set: ',r2train)
         print('R2 score on validation set: ',r2test)
         print("Pearson's correlation coefficient between estimate and ground truth(t
         print("Concordance correlation coefficient between estimate and ground truth
```

```
ridge regression with 197 PCs:
Regularization parameter: 5000
R2 score on training set: 0.471061556011
R2 score on validation set: 0.265014380849
Pearson's correlation coefficient between estimate and ground truth(test set) 0.518424013095
Concordance correlation coefficient between estimate and ground truth(test set) 0.419587774173
```

This has improved our R2 score on the test set quite dramatically. We try another type of regularization, Elastic Net regularization. This method combines L1 (Lasso) and L2 (Ridge) regularization. We cross-validate for regularization strength and the ratio between L1 and L2.

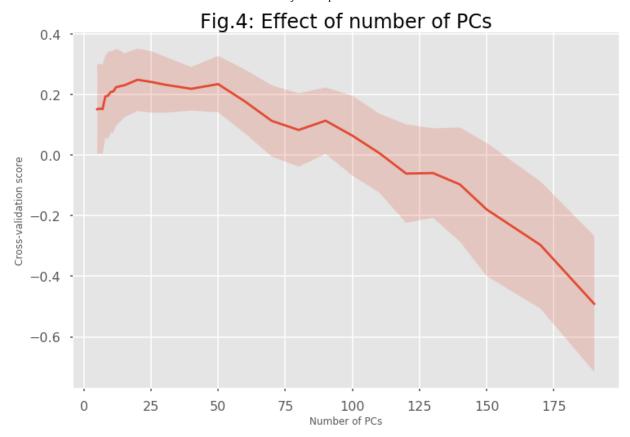
```
In [27]:
         # elastic net regularization
         ratios = [.001, .01, .1, .5, .7, .9, .95, .99, 1]
         elnet_est = ElasticNetCV(l1_ratio = ratios, alphas=alphas,fit_intercept = T)
         elnet_est.fit(X_train_pca,y_train)
         best_alpha = elnet_est.alpha_
         best_ratio = elnet_est.ll_ratio_
         elnet est = ElasticNet(l1 ratio = best ratio, alpha=best alpha,fit intercept
         elnet est.fit(X train pca,y train)
         y hat_train = elnet est.predict(X_train pca)
         y hat_test = elnet_est.predict(X_test_pca)
         name = 'Elastic Net regression with {} PCs'.format(total pcs)
         r2train, r2test, pcc, ccc, summary = evaluate and store(y_train,y_hat_train,
         print(name,': ')
         print('Regularization parameter: ', best_alpha)
         print('L1 ratio: ',best_ratio)
         print('R2 score on training set: ',r2train)
         print('R2 score on validation set: ',r2test)
         print("Pearson's correlation coefficient between estimate and ground truth(t
         print("Concordance correlation coefficient between estimate and ground truth
```

```
Elastic Net regression with 197 PCs:
Regularization parameter: 10.0
L1 ratio: 0.01
R2 score on training set: 0.430632422622
R2 score on validation set: 0.267113344982
Pearson's correlation coefficient between estimate and ground truth(test set) 0.52064312814
Concordance correlation coefficient between estimate and ground truth(test set) 0.41525211834
```

The Elastic Net regularization has not improved our model compared to ridge regularization. Moreover, the optimal ratio between L1 and L2 regularization is very close to pure L2 regularization, further confirming ridge regularization as the optimal decision in this scenario

```
In [28]: # find optimal number of PCs to include in model by determining the 5fold call
         # in a linear regression model
         scorelist = []
         #pcs = range(1,total pcs+1)
         pcs = [5,6,7,8,9,10,11,12,13,14,15,20,25,30,40,50,60,70,80,90,100,110,120,13]
         for use pcs in pcs:
             scores = cross val score(pca lin est, X train pca[:,:use pcs], y train, cv=
             scorelist.append(scores)
         #plot crossvalidation score means and std as a function of number of PCs
         score_array = np.vstack(scorelist)
         valid means = score array.mean(axis=1)
         valid stds = score array.std(axis=1)
         fig4 = plt.figure(figsize = [12,8])
         plt.title('Fig.4: Effect of number of PCs', fontsize = 24)
         ax = plt.gca()
         plt.plot(pcs, valid means)
         ax.fill between(pcs,valid means+valid stds,valid means-valid stds,alpha = 0.
         ax.set ylabel('Cross-validation score')
         ax.set_xlabel('Number of PCs')
         sns.despine()
         # determine optimal number of PCs based on crossvalidation score and fit a
         optimal_pcs = pcs[np.argmax(valid_means)]
         print("\nOptimal number of principal components: ",optimal pcs)
         # fit to complete training set with best set of predictors (PCs)
         pca lin est.fit(X train pca[:,:optimal pcs],y train)
         # get stats
         y hat train = pca lin est.predict(X train pca[:,:optimal pcs])
         y_hat_test = pca_lin_est.predict(X_test_pca[:,:optimal_pcs])
         name = 'linear regression with {} PCs'.format(optimal pcs)
         r2train, r2test, pcc, ccc, summary = evaluate_and_store(y_train,y_hat_train,
         print(name,': ')
         print('R2 score on training set: ',r2train)
         print('R2 score on validation set: ',r2test)
         print("Pearson's correlation coefficient between estimate and ground truth(t
         print("Concordance correlation coefficient between estimate and ground truth
```

```
Optimal number of principal components: 20 linear regression with 20 PCs:
R2 score on training set: 0.321961145795
R2 score on validation set: 0.226915996482
Pearson's correlation coefficient between estimate and ground truth(test set) 0.486073194638
Concordance correlation coefficient between estimate and ground truth(test set) 0.41508971522
```



The test R2 score of this unregularized model with 20 PCs is comparable to the Ridge regression model on all 197 PCs. Further regularization does not improve this model (data not shown).

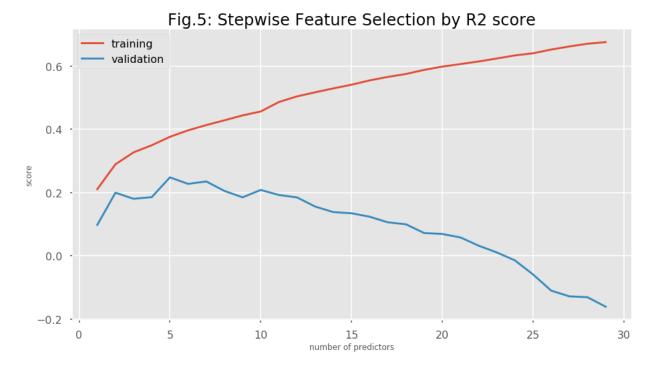
Step-wise feature selection based on cross-validation score

In lieu of PCA, we also try dimensionality reduction by step-wise feature selection. We first split our training set into a smaller training set to stepwise select predictors and a validation set to select the number of predictors to keep. We use a simple linear regression model for this purpose.

```
In [29]: # we split off another validation set to choose the number of features
         X train sm, X valid, y train sm, y valid = train test split(X train, y train, te
         # now we select predictors stepwise by crossvalidation
         all_predictors = X_train.columns.values
         predictors = []
         # add predictors step-wise
         max predictors = 30
         for k in range(1, max predictors):
             if k == 1:
                 used predictors = []
             else:
                 used_predictors = predictors[-1]
             # get currently unused predictors
             unused predictors = list(set(all predictors) - set(used predictors))
             # add each of the unused predictors one at a time and store the mean of
             xvalscores = []
             for predictor in unused predictors:
                 # cross validate with chosen predictors on small training set
                 k predictors = used predictors + [predictor]
                 X train k = X train sm[k predictors]
                 linreg_est = LinearRegression(fit_intercept = True)
                 kf = KFold(n splits=3, shuffle=True, random state=7)
                 score = cross_val_score(linreg_est,X_train_k,y_train_sm,cv=kf).mean(
                 xvalscores.append(score)
             # then choose the predictor that gives the best cross-validation score &
             best k = used predictors + [unused predictors[np.argmax(xvalscores)]]
             predictors.append(best k)
```

```
In [30]: # we now choose the number of predictors based on the R2 score on the held-
         train scores = []
         valid scores = []
         # loop through sets of predictors
         for p in predictors:
             #fit a linear regression model with chosen set of predictors on small to
             linreg est.fit(X train sm[p],y train sm)
             # get R2 score for small training set and validation set
             y hat train sm = linreg est.predict(X train sm[p])
             y hat valid = linreg est.predict(X valid[p])
             train scores.append(r2 score(y train sm,y hat train sm))
             valid scores.append(r2 score(y valid,y hat valid))
         # plot train and validation set scores as a function of number of predictors
         fig5 = plt.figure(figsize= [15,8])
         plt.plot(range(1, max predictors), train scores)
         plt.plot(range(1, max predictors), valid scores)
         plt.title('Fig.5: Stepwise Feature Selection by R2 score', fontsize=24)
         plt.xlabel('number of predictors')
         plt.ylabel('score')
         plt.legend(['training','validation'])
         # choose the model with the best test score
         best predictors = predictors[np.argmax(valid_scores)]
         # extract best predictors to new dataframe
         X train feat = X train[best predictors]
         X test feat = X test[best predictors]
         # fit linear regression model to complete (not small) training set and evalu
         linreg est = LinearRegression()
         linreg est.fit(X train feat,y train)
         y_hat_train = linreg_est.predict(X_train_feat)
         y hat test = linreg est.predict(X test feat)
         name = 'linear regression with {} predictors'.format(len(best predictors))
         r2train, r2test, pcc, ccc, summary = evaluate_and_store(y_train,y_hat_train,
         print(name,': ')
         print('optimal number of predictors: ',len(best_predictors))
         print('predictors: ',best predictors)
         print('R2 score on training set: ',r2train)
         print('R2 score on validation set: ',r2test)
         print("Pearson's correlation coefficient between estimate and ground truth(t
         print("Concordance correlation coefficient between estimate and ground truth
         linear regression with 5 predictors :
         optimal number of predictors: 5
         predictors: ['FreeSurfer.thickness..mean.1009', 'FreeSurfer.thickness..m
         ean.2006', 'mean.curvature..kurtosis.1007', 'geodesic.depth..SD.1011', 'P
         TEDUCAT']
         R2 score on training set: 0.347888052415
         R2 score on validation set: 0.263384791347
         Pearson's correlation coefficient between estimate and ground truth(test
         set) 0.522998265342
```

Concordance correlation coefficient between estimate and ground truth(tes t set) 0.457283947517



This simple linear regression model with only 5 predictors a similar performance as the PCA models with an R2 score of 0.26.

```
In [31]:
         # test if regularization improves the model
         ridge feat est = RidgeCV(alphas=alphas, fit intercept = True, cv=5)
         ridge_feat_est.fit(X_train_feat,y_train)
         best_alpha = ridge_feat_est.alpha_
         # now train on whole training set with best alpha
         ridge_feat_est.fit(X_train_feat,y_train)
         y hat train = ridge feat est.predict(X train feat)
         y hat test = ridge feat est.predict(X test feat)
         name = 'linear regression model with Ridge regularization on {} PCA predicted
         r2train, r2test, pcc, ccc, summary = evaluate and store(y_train,y_hat_train,
         print(name,': ')
         print("Regularization parameter for Ridge regularization: ",best alpha)
         print('R2 score on training set: ',r2train)
         print('R2 score on validation set: ',r2test)
         print("Pearson's correlation coefficient between estimate and ground truth(t
         print("Concordance correlation coefficient between estimate and ground truth
```

linear regression model with Ridge regularization on 197 PCA predictors:

```
Regularization parameter for Ridge regularization: 50
R2 score on training set: 0.346354646594
R2 score on validation set: 0.267638593243
Pearson's correlation coefficient between estimate and ground truth(test set) 0.522653783263
Concordance correlation coefficient between estimate and ground truth(test set) 0.441699871133
```

Regularization does not appear to improve the plain linear regression model further.

Ensemble Methods

Ensemble methods like Random Forest Regression or Gradient Boosting are another way to reduce the number of features being considered in the model. We first try Random Forest Regression and crossvalidate to optimize the maximum depth of the trees on a relatively small number of trees (64), we fix the number of features to the square root of all features (46). We then increase the number of trees to 512 to acchieve a slightly better model. Because of the long time it takes to compute these Random Forests we did not perform an exhaustive grid search of all hyper-parameters.

```
In [32]: # because the random forest regression takes to train a long time, we just of
         # we set the max features to 'sqrt' and the number of trees to 64
         # rf = RandomForestRegressor(n estimators = 64, max features = 'sqrt', random
         rf = RandomForestRegressor(n_estimators = 64, max_features = 'sqrt', random_st
         parameters = {'max depth':[i for i in range(3,15)]}
         gs_rf = GridSearchCV(rf,parameters)
         gs rf.fit(X train,y train)
         print(gs_rf.best_estimator_)
         print('optimal max depth: ',gs_rf.best_estimator_.max depth)
         RandomForestRegressor(bootstrap=True, criterion='mse', max_depth=6,
                    max features='sqrt', max leaf nodes=None,
                    min_impurity_decrease=0.0, min_impurity_split=None,
                    min_samples_leaf=1, min_samples_split=2,
                    min weight fraction leaf=0.0, n estimators=64, n jobs=1,
                    oob score=False, random state=7, verbose=0, warm start=False)
         optimal max depth: 6
In [33]: # we then increase the number of trees to improve our model
         rf_est = RandomForestRegressor(n_estimators = 512, max_features = 'sqrt', max_
         rf_est.fit(X_train,y_train)
         y_hat_train = rf_est.predict(X_train)
         y_hat_test = rf_est.predict(X_test)
         name = 'Random Forest Regressor with all predictors'
         r2train, r2test, pcc, ccc, summary = evaluate_and_store(y_train,y_hat_train,
         print(name,': ')
         print('R2 score on training set: ',r2train)
         print('R2 score on validation set: ',r2test)
         print("Pearson's correlation coefficient between estimate and ground truth(t
         print("Concordance correlation coefficient between estimate and ground trutk
```

```
Random Forest Regressor with all predictors:
R2 score on training set: 0.797614336929
R2 score on validation set: 0.249720758769
Pearson's correlation coefficient between estimate and ground truth(test set) 0.507697810406
Concordance correlation coefficient between estimate and ground truth(test set) 0.361389204613
```

At least for the limited set of hyper-parameters we tested, the random forest model does not perform as well as our previous models.

We then tried a Gradient Boosting Regressor for our prediction using crossvalidation to determine the optimal depth of the individual estimators as well as the number of estimators. We then retrain on the complete training set and acchieve the best model performance so far.

```
In [34]: # try Gradient Boosting
         gbr = GradientBoostingRegressor()
         parameters = {'max_depth':[i for i in range(1,5)],'n_estimators':[i*50 for i
         gs_gbr = GridSearchCV(gbr,parameters)
         gs_gbr.fit(X_train,y_train)
         print(gs_gbr.best_estimator_)
         GradientBoostingRegressor(alpha=0.9, criterion='friedman_mse', init=None,
                      learning_rate=0.1, loss='ls', max_depth=3, max_features=Non
         e,
                      max leaf nodes=None, min impurity decrease=0.0,
                      min_impurity_split=None, min_samples_leaf=1,
                      min samples split=2, min weight fraction leaf=0.0,
                      n_estimators=50, presort='auto', random_state=None,
                      subsample=1.0, verbose=0, warm_start=False)
In [35]: # retrain on the complete training set with optimal parameters
         gbr_est = GradientBoostingRegressor(max_depth=4,n_estimators=100)
         gbr_est.fit(X_train,y_train)
         y_hat_train = gbr_est.predict(X_train)
         y_hat_test = gbr_est.predict(X_test)
         name = 'gradient boosting regressor with all predictors'
         r2train, r2test, pcc, ccc, summary = evaluate and store(y_train,y_hat_train,
         #display(pd.DataFrame(np.array([name,r2train, r2test, pcc, ccc]).reshape(1,
                                                                            "Pearson's
         #
                                                                           "Concordance
         print(name,': ')
         print('R2 score on training set: ',r2train)
         print('R2 score on validation set: ',r2test)
         print("Pearson's correlation coefficient between estimate and ground truth(t
         print("Concordance correlation coefficient between estimate and ground truth
```

```
gradient boosting regressor with all predictors:
R2 score on training set: 0.996273889194
R2 score on validation set: 0.285321655084
Pearson's correlation coefficient between estimate and ground truth(test set) 0.535376303013
Concordance correlation coefficient between estimate and ground truth(test set) 0.459441767335
```

Model Performance Comparison

To evaluate our models, we used 3 different metrics: R2 score, Pearson's correlation coefficient and Concordance correlation coefficient, all on the test set. The latter is the metric used in the AD DREAM challenge to score the leaderboard.

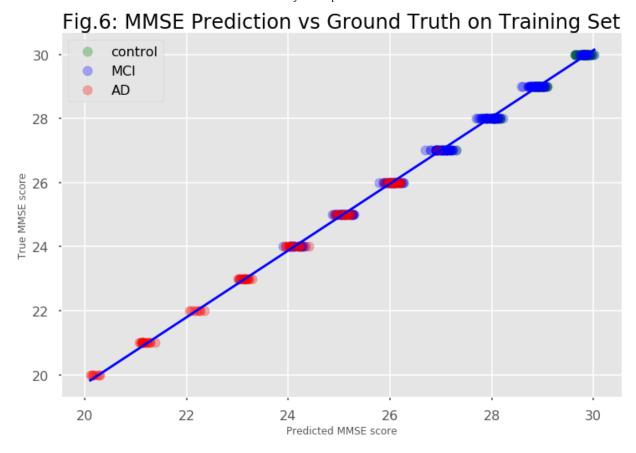
In [36]: display(pd.DataFrame(summary)[['model name', 'R2train', 'R2test', 'PCC', 'CCC']]

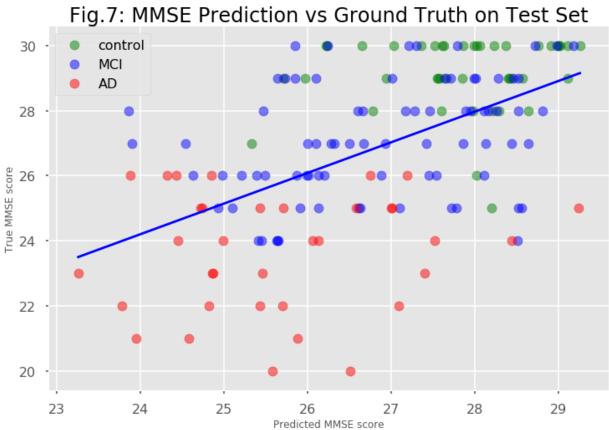
	model name	R2train	R2test	PCC	ccc
0	linear regression with all predictors	1.000000	-0.575704	0.292637	0.291248
1	linear regression on 197 PCs	0.607145	0.146130	0.462699	0.434770
2	ridge regression with 197 PCs	0.471062	0.265014	0.518424	0.419588
3	Elastic Net regression with 197 PCs	0.430632	0.267113	0.520643	0.415252
4	linear regression with 20 PCs	0.321961	0.226916	0.486073	0.415090
5	linear regression with 5 predictors	0.347888	0.263385	0.522998	0.457284
6	linear regression model with Ridge regularizat	0.346355	0.267639	0.522654	0.441700
7	Random Forest Regressor with all predictors	0.797614	0.249721	0.507698	0.361389
8	gradient boosting regressor with all predictors	0.996274	0.285322	0.535376	0.459442

The best model in all 3 evaluation metrics (R2 score, Pearson's correlation coefficient, Concordance correlation coefficient, all on test set) is the gradient boosting regressor. Assuming our test set is representative of the test set used for the AD DREAM challenge, our Concordance correlation coefficient of 0.47 would place us on rank 8 of the final leaderboard (https://www.synapse.org/#!Synapse:syn2290704/wiki/68513

(https://www.synapse.org/#!Synapse:syn2290704/wiki/68513)). The Pearson's correlation coefficient would place us even higher. We use our optimal model to visualize how our predicted MMSE scores compare to the true MMSE scores, both on the training set and test set. We also visualize the different diagnoses classes (CN = control, MCI = mild cognitive impairment and AD = Alzheimer's disease) in these diagrams.

```
In [37]: # visualization of the best model
         y_hat_train = gbr_est.predict(X_train)
         y_hat_test = gbr_est.predict(X_test)
         # indices for the 3 diagnoses
         idx1 = y_dx_train == 0
         idx2 = y dx train == 1
         idx3 = y_dx_train == 2
         # plot of prediction vs truth on training set
         fig6 = plt.figure(figsize = [12,8])
         plt.scatter(y hat train[idx1],y train[idx1],alpha=0.3,c='green')
         plt.scatter(y hat train[idx2],y train[idx2],alpha=0.3,c='blue')
         plt.scatter(y_hat_train[idx3],y_train[idx3],alpha=0.3,c='red')
         plt.xlabel('Predicted MMSE score')
         plt.ylabel('True MMSE score')
         plt.legend(['control','MCI','AD'])
         # plot regression line
         beta1,beta0 = np.polyfit(y_hat_train,y_train,deg=1)
         f = lambda x: beta1 * x + beta0
         x = np.array([np.min(y hat train), np.max(y hat train)])
         plt.plot(x,f(x),c='blue')
         plt.title('Fig.6: MMSE Prediction vs Ground Truth on Training Set', fontsize
         sns.despine()
         # plot of prediction vs truth on test set
         idx1 = y dx test == 0
         idx2 = y dx test == 1
         idx3 = y dx test == 2
         fig7 = plt.figure(figsize = [12,8])
         plt.scatter(y_hat_test[idx1],y_test[idx1],alpha=0.5,c='green')
         plt.scatter(y hat test[idx2],y test[idx2],alpha=0.5,c='blue')
         plt.scatter(y_hat_test[idx3],y_test[idx3],alpha=0.5,c='red')
         plt.xlabel('Predicted MMSE score')
         plt.ylabel('True MMSE score')
         plt.legend(['control','MCI','AD'])
         plt.title('Fig.7: MMSE Prediction vs Ground Truth on Test Set', fontsize=24)
         #plot regression line
         beta1,beta0 = np.polyfit(y hat test,y test,deg=1)
         f = lambda x: beta1 * x + beta0
         x = np.array([np.min(y hat test), np.max(y hat test)])
         plt.plot(x,f(x),c='blue')
         sns.despine()
```





Based on the distribution of different diagnosis classes in this graph, it should be helpful to use the predicted MMSE in prediction of diagnoses.

Influence of estimated MMSE on diagnosis prediction

We are first comparing three simple logistic regression classifiers: One that uses only the demographic data for predicting diagnosis, one that uses the actual MMSE score and one that uses our predicted MMSE score. The first achieves only 46% accuracy on the test set, whereas the second achieves 69%. The model with our predicted MMSE score is halfway between these 2 extremes with 56% accuracy.

```
In [38]: # try to predict diagnosis based on demographic predictors only (logistic re
         X_est_train = X_train[demographic_columns].copy()
         X_est_test = X_test[demographic_columns].copy()
         Cs = [100000, 50000, 10000, 5000, 1000, 500, 100, 50, 10, 5, 1, 0.5, 0.1, 0.05, 0.01, 0.005]
         logreg_clf = LogisticRegressionCV(Cs=Cs)
         logreg_clf.fit(X_est_train,y_dx_train)
         y hatdx train = logreg clf.predict(X est train)
         y_hatdx_test = logreg_clf.predict(X_est_test)
         train acc = np.sum(y hatdx train == y dx train)/len(y dx train)
         test_acc = np.sum(y_hatdx_test == y_dx_test)/len(y_dx_test)
         print('model with demographic factors only:')
         print('training accuracy: ',train_acc)
         print('test accuracy: ', test_acc)
         model with demographic factors only:
         training accuracy: 0.511677282378
         test accuracy: 0.464968152866
In [39]: # new model based on demographic predictors + true MMSE score
         X_est_train['MMSE'] = y_train
         X_est_test['MMSE'] = y_test
         Cs = [100000, 50000, 10000, 5000, 1000, 500, 100, 50, 10, 5, 1, 0.5, 0.1, 0.05, 0.01, 0.005]
         logreg_clf = LogisticRegressionCV(Cs=Cs)
         logreg clf.fit(X est train,y dx train)
         y hatdx train = logreg clf.predict(X est train)
         y_hatdx_test = logreg_clf.predict(X est test)
         train acc = np.sum(y hatdx train == y dx train)/len(y dx train)
         test_acc = np.sum(y_hatdx_test == y_dx_test)/len(y_dx_test)
         print('model with demographic factors + true MMSE score:')
         print('training accuracy: ',train acc)
         print('test accuracy: ', test acc)
         model with demographic factors + true MMSE score:
         training accuracy: 0.713375796178
         test accuracy: 0.694267515924
```

```
In [40]: # new model based on demographic predictors + our predicted MMSE score
    X_est_train['MMSE'] = y_hat_train
    X_est_test['MMSE'] = y_hat_test

Cs = [100000,50000,10000,5000,1000,500,100,50,10,5,1,0.5,0.1,0.05,0.01,0.005
logreg_clf = LogisticRegressionCV(Cs=Cs)
logreg_clf.fit(X_est_train,y_dx_train)
    y_hatdx_train = logreg_clf.predict(X_est_train)
    y_hatdx_test = logreg_clf.predict(X_est_test)

train_acc = np.sum(y_hatdx_train == y_dx_train)/len(y_dx_train)
    test_acc = np.sum(y_hatdx_test == y_dx_test)/len(y_dx_test)

print('model with demographic factors + our estimated MMSE score:')
    print('training accuracy: ', train_acc)
    print('test accuracy: ', test_acc)
```

```
model with demographic factors + our estimated MMSE score: training accuracy: 0.72186836518 test accuracy: 0.573248407643
```

In order to getter a better estimate and a significance value for the improvement, we perform bootstrapping by resampling our training set with replacement, use the remaining observations as test set and determine the difference in test accuracies between the base model and our (putatively) improved model.

```
In [41]: # get p-values for the effect of estimated mmse score using a paired t-test
         num iterations = 100
         train_size = X_est_train.shape[0]
         data = pd.concat([pd.concat([X_est_train,X_est_test]),pd.concat([y_dx_train,
         test acc diffs = []
         for i in range(num_iterations):
             # prepare train and test sets
             train = resample(data, n_samples=train_size)
             test = np.array([x for x in data if x.tolist() not in train.tolist()])
             X_train_bs = train[:,:-1]
             y_train_bs = train[:,-1]
             X \text{ test bs} = \text{test}[:,:-1]
             y_test_bs = test[:,-1]
             # fit base model without mmse
             logreg clf = LogisticRegressionCV(Cs=Cs)
             logreg clf.fit(X train bs[:,:-1],y train bs)
             y hat base = logreg clf.predict(X test bs[:,:-1])
             acc base = np.sum(y_hat_base == y_test_bs)/len(y_test_bs)
             # fit mmse model
             logreg clf.fit(X train bs,y train bs)
             y hat mmse = logreg clf.predict(X test bs)
             acc mmse = np.sum(y hat mmse == y test bs)/len(y test bs)
             test_acc_diffs.append(acc_mmse - acc_base)
         fig8 = plt.figure(figsize = [12,8])
         plt.hist(test acc diffs)
         plt.xlabel('improvement in accuracies')
         plt.ylabel('number of bootstraps')
         plt.title('Fig.8: Histogram of model improvement in bootstraps',fontsize=24)
```



Fig.8: Histogram of model improvement in bootstraps

In [42]:

,pval = ttest 1samp(test acc diffs,popmean=0) print('mean improvement by inclusion of predicted MMSE score: ',np.mean(test print('Null Hypothesis: Including predicted MMSE has no effect on accuracy print('p-value: ', pval)

mean improvement by inclusion of predicted MMSE score: 0.171083428693 Null Hypothesis: Including predicted MMSE has no effect on accuracy of cl assifier

p-value: 9.00462879262e-76

The improvement in test accuracies is roughly normally distributed and has a mean of 17%. Based on a paired t-test, these effects are highly significant.

Discussion

We have shown that our best models can compete with other submissions to the AD DREAM challenge and even outperform 2/3 of the submissions. We have also shown that the predicted MMSE scores from our best model are good enough to significantly improve Alzheimer's Disease/Dementia diagnosis. However, given more time, there might be ways to further improve our model: We currently use preprocessed geometrical data to approximate brain morphology as our predictors; the raw MRI data might contain information that is not captured by this data. Training a deep neural net on raw MRI images might improve our prediction.

It is remarkable that a number of our models is performing relatively well. Combining these models by stacking might further improve our prediction.

In []: