SANTA CRUZ BIOTECHNOLOGY, INC.

HA-probe (F-7): sc-7392

SANTA CRUZ BIOTECHNOLOGY

The Power to Question

BACKGROUND

Plasmid vectors for the expression of coding regions of eukaryotic genes in bacterial, insect and mammalian hosts are in common usage; such expression vectors are frequently used to encode hybrid fusion proteins consisting of a eukaryotic target protein and a specialized region designed to aid in the purification and visualization of the target protein. For example, the pCDM8 expression vector and derivatives thereof encode fusions between the target protein and an eleven amino acid peptide derived from the influenza protein hemagglutinin (HA). The HA epitope tag is useful in Western blotting and immunohistochemical localization of expressed fusion proteins when examined with antibodies raised specifically against the HA-epitope tag.

SOURCE

HA-probe (F-7) is a mouse monoclonal antibody specific to epitope mapping within an internal region of the influenza hemagglutinin (HA) protein.

PRODUCT

Each vial contains 200 µg IgG_{2a} in 1.0 ml of PBS with < 0.1% sodium azide and 0.1% gelatin.

Blocking peptide available for competition studies, sc-7392 P, (100 µg peptide in 0.5 ml PBS containing < 0.1% sodium azide and 0.2% BSA).

Available as phycoerythrin (sc-7392 PE), PerCP (sc-7392 PerCP) or PerCP-Cy5.5 (sc-7392 PCPC5) conjugates for flow cytometry 100 tests.

Available as agarose conjugate for immunoprecipitation, sc-7392 AC, 500 µg/ 0.25 ml agarose in 1 ml.

Available as TransCruz reagent for ChIP application, sc-7392 X, 200 µg/0.1 ml.

Available as HRP conjugate for Western blotting, sc-7392 HRP, 200 µg/ml.

Available as fluorescein (sc-7392 FITC) or rhodamine (sc-7392 TRITC) conjugates for use in immunofluorescence, 200 µg/ml.

Available as Alexa Fluor® 405 (sc-7392 AF405), Alexa Fluor® 488 (sc-7392 AF488) or Alexa Fluor® 647 (sc-7392 AF647) conjugates for immunofluorescence; 100 µg/2 ml.

Alexa Fluor® is a trademark of Molecular Probes, Inc., Oregon, USA

APPLICATIONS

HA-probe (F-7) is recommended for detection of proteins containing the HA tag by Western Blotting (starting dilution 1:200, dilution range 1:100-1:1000), immunoprecipitation [1-2 µg per 100-500 µg of total protein (1 ml of cell lysate)], immunofluorescence (starting dilution 1:50, dilution range 1:50-1:500), flow cytometry (1 µg per 1 x 10^6 cells) and solid phase ELISA (starting dilution 1:30, dilution range 1:30-1:3000).

HA-probe (F-7) X TransCruz antibody is recommended for ChIP assays.

Positive Controls: HA fusion proteins produced by cells transfected with pCruz HA™ expression vector (sc-5045).

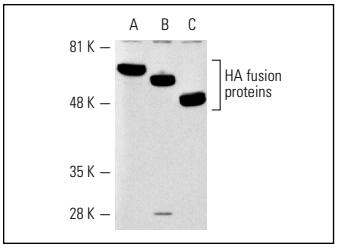
RESEARCH USE

For research use only, not for use in diagnostic procedures.

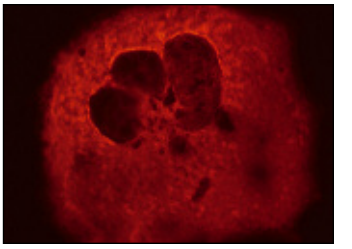
STORAGE

Store at 4° C, **DO NOT FREEZE**. Stable for one year from the date of shipment. Non-hazardous. No MSDS required.

DATA



HA-probe (F-7): sc-7392. Western blot analysis of HA-tagged fusion proteins showing N-terminal HA-tagged JNK2 (**A**) and JNK1 (**C**) and C-terminal HA-tagged Daxx (**B**).



HA-probe (F-7): sc-7392. Immunofluorescence staining of methanol-fixed COS cells transfected with HA fusion protein showing cytoplasmic staining.

SELECT PRODUCT CITATIONS

1. Runnels, L.W., et al. 2001. TRP-PLIK, a bifunctional protein with kinase and ion channel activities. Science 291: 1043-1047.
2. Bhunia, A.K., et al. 2002. PKD1 induces p21Waf1 and regulation of the cell cycle via direct activation of the JAK/Stat signaling pathway in a process requiring PKD2. Cell 109: 157-168.
3. Guipponi, M., et al. 2002. The transmembrane serine protease (TMPRSS3) mutated in deafness DFNB8/10 activates the epithelial sodium channel (ENaC) *in vitro*. Hum. Mol. Genet. 11: 2829-2836.
4. Kilstrup-Nielsen, C., et al. 2003. Pbx 1 nuclear export is regulated independently of PBX–MEINOX interaction by PKA phosphorylation of the PBC-B domain. EMBO J. 22: 89-99.
5. Soares, L., et al. 2004. Two isoforms of otubain 1 regulate T cell anergy via GRAIL. Nat. Immunol. 5: 45-54.
6. Nishio, M., et al. 2005. Human parainfluenza virus type 4 is incapable of evading the interferon-induced antiviral effect. J. Virol. 79: 14756-14768.
7. Chen, Z., et al. 2007. Proteolytic processing and deubiquitinating activity of papain-like proteases of human coronavirus NL63. J. Virol. 81: 6007-6018.
8. Zaug, A.J., et al. 2010. Functional interaction between telomere protein TPP1 and telomerase. Genes Dev. 24: 613-622.
9. Meng, H., et al. 2010. Low density lipoprotein receptor-related protein-1 (LRP1) regulates thrombospondin-2 (TSP2) enhancement of Notch3 signaling. J. Biol. Chem. 285: 23047-23055.
10. Kinchen, J.M., et al. 2010. Identification of two evolutionarily conserved genes regulating processing of engulfed apoptotic cells. Nature 464: 778-782.
11. Ko, J.C., et al. 2011. Modulation of Rad51, ERCC1, and thymidine phosphorylase by emodin result in synergistic cytotoxic effect in combination with capecitabine. Biochem. Pharmacol. 81: 680-690.