***T. brucei* DAPI nucleic acid staining for FACS**

1. Sample préparation :

* collect 106 cells in a 15 ml tube,
* pellet by centrifugation 10min at 1800g and discard the supernatant,
* tap the tube to resuspend the pellet in the residual liquid and add 1ml of PBS,
* add 4ml of absolute ethanol at -20°C, pipet the cell suspension slowly (do not vortex) and leave them 30min at -20°C,
* pellet by centrifugation 10min at 1800g and discard the supernatant,
* tap the tube to loosen the pellet and add 5ml of PBS, allow the cells to rehydrate for 15min at room temperature.

1. Staining :

* dilute the DAPI stock (100µg/ml) for a final concentration of 1µg/ml in staining buffer,
* centrifuge the cells 10min at 1800g and discard the supernatant,
* tap to loosen the pellet and add 1ml of DAPI diluted in staining buffer,
* incubate for 15min at room temperature,
* analyze by flow cytometry or stock à 4°C maximum overnight.

Staining buffer :

* 100mM Tris, pH7,4
* 150mM NaCl,
* 1mM CaCl2
* 0,5mM MgCl2
* 0,1% NP-40