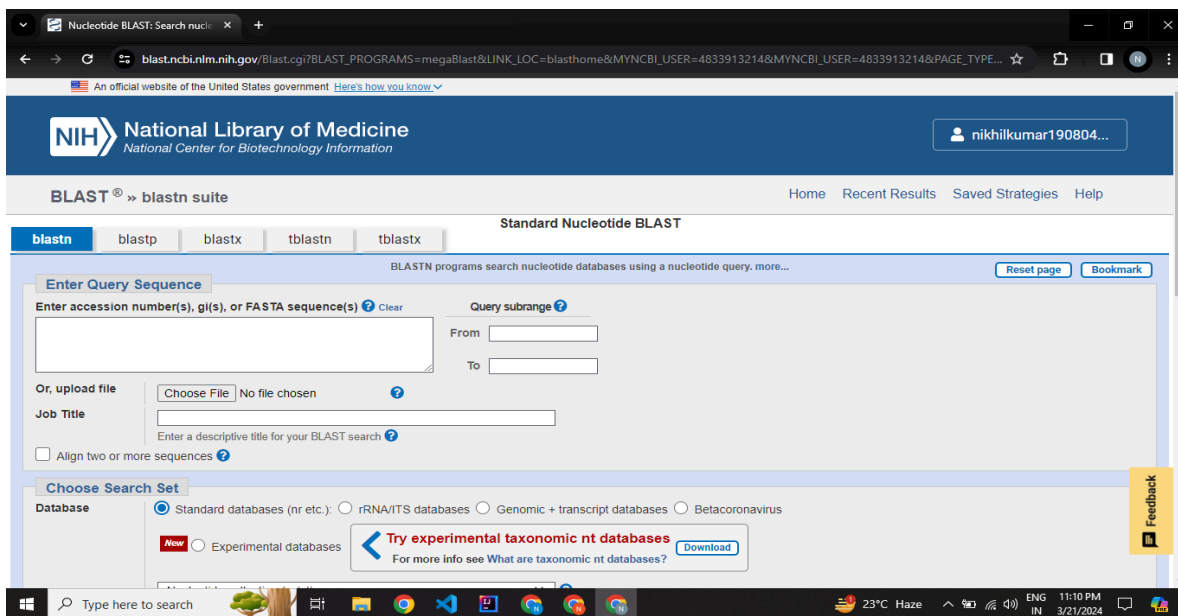


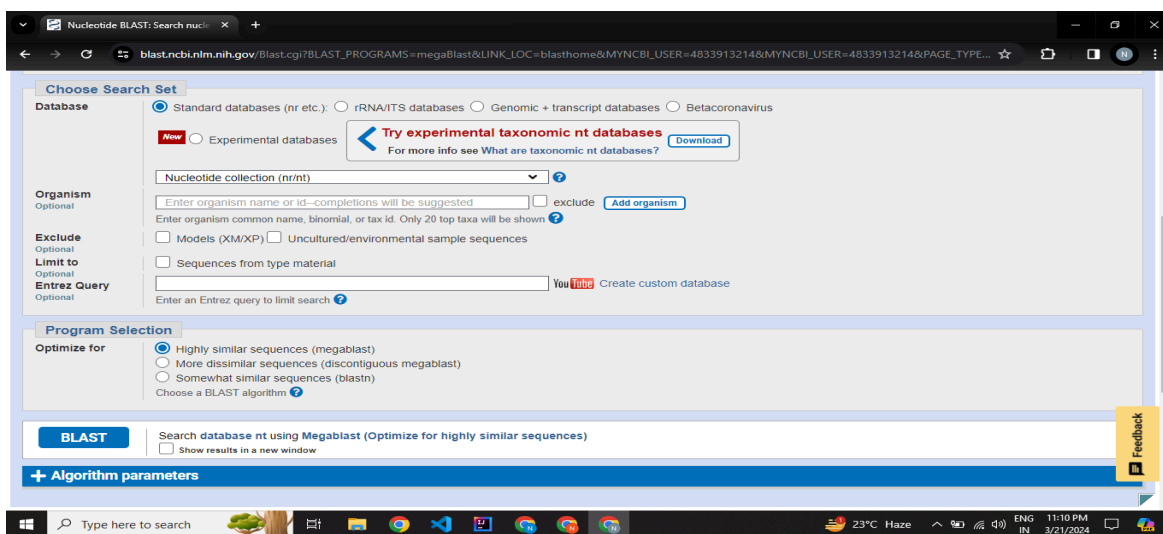
# Using BLAST To Compare the PDC gene sequence across different Saccharomyces cerevisiae strains



The screenshot shows the NCBI BLASTn suite interface. The top navigation bar includes the NIH logo, the text "National Library of Medicine National Center for Biotechnology Information", and a user profile "nikhilkumar190804...". Below this is the "BLAST® » blastn suite" header with links for "Home", "Recent Results", "Saved Strategies", and "Help". The main section is titled "Standard Nucleotide BLAST" and contains three tabs: "blastn", "blastp", and "tblastx". The "blastn" tab is active. The "Enter Query Sequence" section has a text input field for "Enter accession number(s), gi(s), or FASTA sequence(s)", a "Query subrange" section with "From" and "To" fields, and an "Or, upload file" section with a "Choose File" button. Below this is a "Job Title" field and a checkbox for "Align two or more sequences". The "Choose Search Set" section has a "Database" dropdown set to "Standard databases (nr etc.)", with options for "rRNA/ITS databases", "Genomic + transcript databases", and "Betacoronavirus". There is a "Try experimental taxonomic nt databases" button and a "Download" button. The bottom of the interface shows a Windows taskbar with various icons and system information.

This is the standard view of the BLASTn. It is divided into three sections, namely: “Enter Query Sequence,” “Choose Search Set”, and” Program Selection.”

In the section “Enter Query Sequence,” we have to Enter the input query i.e.- the sequence for which we want to perform the BLAST. Here we can either paste the sequence directly or give the accession number of that sequence and provide a subrange for that sequence.

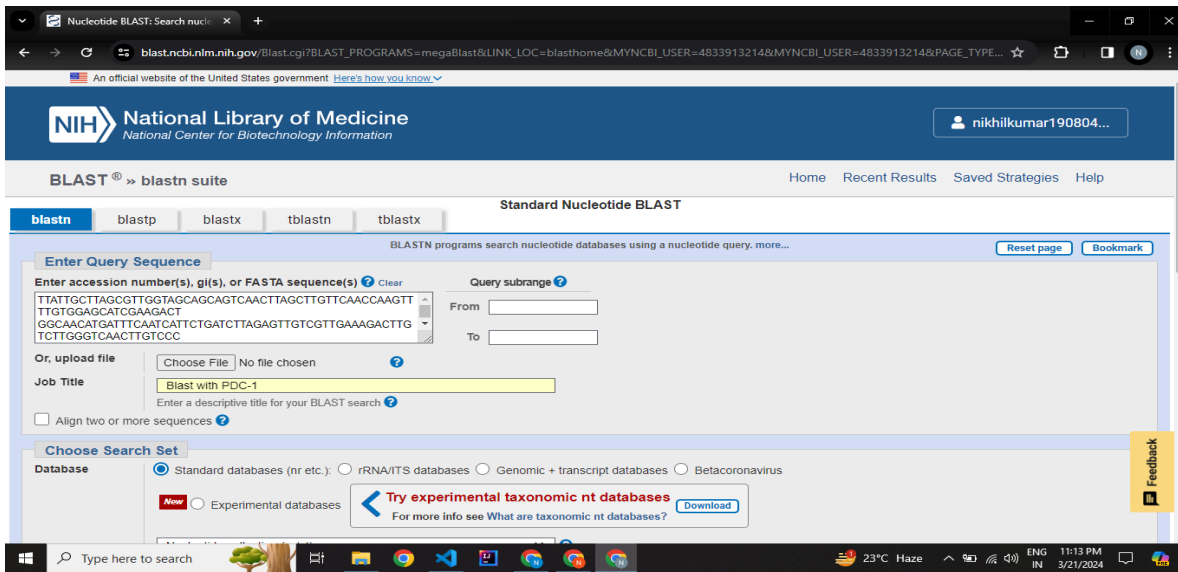


The screenshot shows the NCBI BLASTn suite interface, focusing on the "Choose Search Set" and "Program Selection" sections. The "Choose Search Set" section has a "Database" dropdown set to "Standard databases (nr etc.)", with options for "rRNA/ITS databases", "Genomic + transcript databases", and "Betacoronavirus". There is a "Try experimental taxonomic nt databases" button and a "Download" button. Below this is a "Nucleotide collection (nr/nt)" dropdown, an "Organism" field with a search box, and an "Exclude" section with checkboxes for "Models (XM/XP)", "Uncultured/environmental sample sequences", and "Sequences from type material". There is also an "Entrez Query" field. The "Program Selection" section has an "Optimize for" dropdown set to "Highly similar sequences (megablast)", with options for "More dissimilar sequences (discontiguous megablast)" and "Somewhat similar sequences (blastn)". There is a "Choose a BLAST algorithm" button. At the bottom, there is a "BLAST" button and a "Search database nt using Megablast (Optimize for highly similar sequences)" checkbox. The bottom of the interface shows a Windows taskbar with various icons and system information.

Now, in the section “Choose Search Set” we can select the database we want to use for BLAST. By default, it is selected as “Nucleotide” and standard database (non-redundant). Also here, we have the option to add an organism or to exclude an organism from the database. And we also have the option to filter our database using an Entrez Query.

And in the last section, “Program Selection” we have some optimization options to enhance our searches. And lastly we can change some algorithm parameters according to our needs.

# BLAST With PDC-1



Nucleotide BLAST: Search nucleotide sequences

blastn | blastp | blastx | tblastn | tblastx

Standard Nucleotide BLAST

BLASTn programs search nucleotide databases using a nucleotide query, more...

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [Clear](#)

Query subrange

From

To

Or, upload file  No file chosen

Job Title

Enter a descriptive title for your BLAST search

☐ Align two or more sequences

Choose Search Set

Database

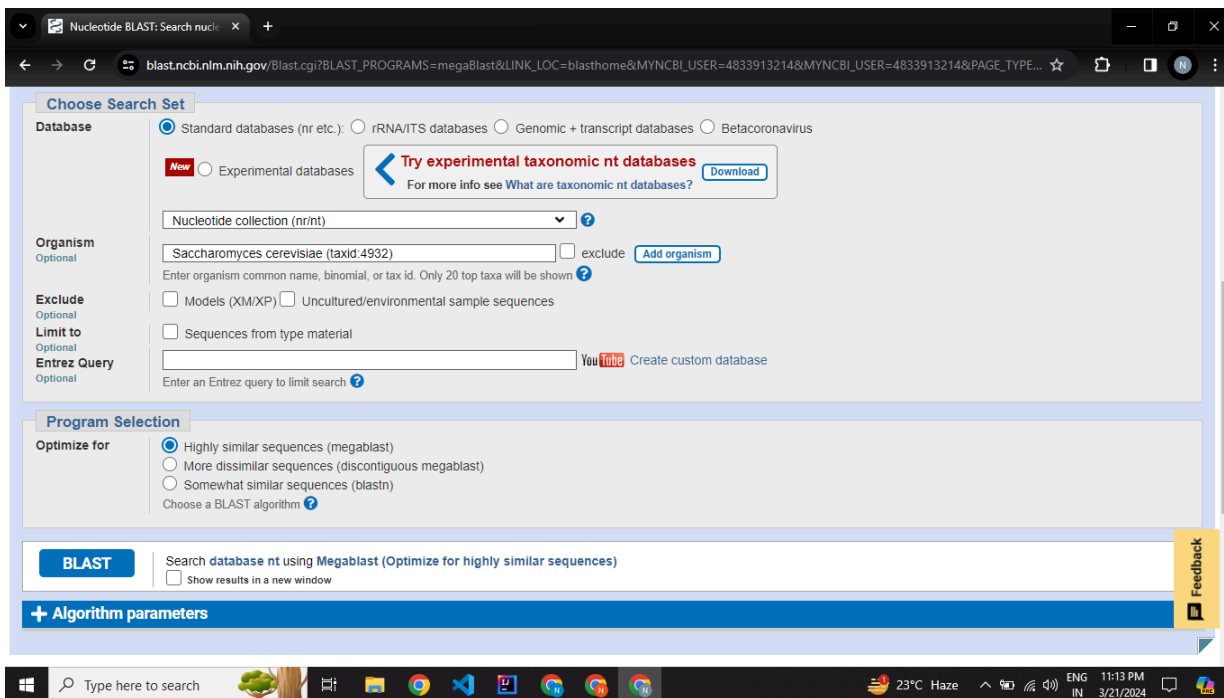
☒ Standard databases (nr etc.) ☐ rRNA/ITS databases ☐ Genomic + transcript databases ☐ Betacoronavirus

☐ Experimental databases

[Try experimental taxonomic nt databases](#) [Download](#)

For more info see [What are taxonomic nt databases?](#)

Now, here i am performing BLAST with input as the PDC-1 gene. I pasted the sequence of the PDC-1 gene, which I downloaded using Python, and gave a suitable Job Title to this. The accession number for PDC-1 is NC\_001144.5 and the sequence starts from 234081 to 232390. It is in reverse complement form.



Nucleotide BLAST: Search nucleotide sequences

blastn | blastp | blastx | tblastn | tblastx

Standard Nucleotide BLAST

BLASTn programs search nucleotide databases using a nucleotide query, more...

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [Clear](#)

Query subrange

From

To

Or, upload file  No file chosen

Job Title

Enter a descriptive title for your BLAST search

☐ Align two or more sequences

Choose Search Set

Database

☒ Standard databases (nr etc.) ☐ rRNA/ITS databases ☐ Genomic + transcript databases ☐ Betacoronavirus

☐ Experimental databases

[Try experimental taxonomic nt databases](#) [Download](#)

For more info see [What are taxonomic nt databases?](#)

Nucleotide collection (nr/nt)

Organism

☐ exclude [Add organism](#)

Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown

Exclude

☐ Models (XM/XP) ☐ Uncultured/environmental sample sequences

Limit to

☐ Sequences from type material

Entrez Query

[YouTube](#) [Create custom database](#)

Enter an Entrez query to limit search

Program Selection

Optimize for

☒ Highly similar sequences (megablast)

☐ More dissimilar sequences (discontiguous megablast)

☐ Somewhat similar sequences (blastn)

Choose a BLAST algorithm

[BLAST](#) Search database nt using Megablast (Optimize for highly similar sequences)

☐ Show results in a new window

+ Algorithm parameters

Here I selected the database as Standard database, which is a by default option, and selected the Nucleotide collection (nr/nt) which again was by default. After that i added the Organism "Saccharomyces cerevisiae," and I selected the top among them with taxid as 4932. After that, in section "Program Selection" I kept the options by default, and in algorithm parameters I kept everything by default only. And finally, i clicked on the button for BLAST. It took some time to complete this request, and after some time, the result page was shown.

# Result of BLAST

The screenshot shows the NCBI BLAST results page for a search of PDC-1. The page is titled "BLAST® » blastn suite » results for RID-ZSWF0251013". On the left, there is a "Job Title" section with details: Job Title: Blast with PDC-1, RID: ZSWF0251013, Program: BLASTN, Database: nt, Query ID: lcl|Query\_7474245, Description: None, Molecule type: dna, Query Length: 1692. On the right, there is a "Filter Results" panel with options to filter by Organism, Percent Identity, E value, and Query Coverage. The Organism filter is currently set to "only top 20 will appear".

This is the result page of the BLAST. On the left side, it provides the details related to the search like Job Title, a unique ID for every search, the BLAST program that was selected (blastn.blastp, tblastx etc.), the database which was selected, and the query length which was 1692 bases long for the PDC-1. On the right side, it gives the option to filter out the results based on some parameters like percent identity and E value or by using an organism name.

The screenshot shows the "Sequences producing significant alignments" section of the BLAST results page. It displays a table with 10 columns: Description, Scientific Name, Max Score, Total Score, Query Cover, E value, Per. Ident, Acc. Len, and Accession. The table lists 10 sequences, all of which are from *Saccharomyces cerevisiae*. The sequences are sorted by E value, with the lowest E value (0.0) at the top. The table is titled "Sequences producing significant alignments" and includes a "Download" button and a "Select columns" dropdown. The table is also titled "Sequences producing significant alignments" and includes a "Download" button and a "Select columns" dropdown.

Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession
<a href="#">Saccharomyces cerevisiae strain YSR128 chromosome XII complete sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5087	100%	0.0	100.00%	1076801	<a href="#">CP036478.1</a>
<a href="#">Saccharomyces cerevisiae strain SY14 chromosome I complete sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5903	100%	0.0	100.00%	11848804	<a href="#">CP029160.1</a>
<a href="#">Saccharomyces cerevisiae strain RY4742 chromosome XII complete sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5087	100%	0.0	100.00%	1104511	<a href="#">CP026300.1</a>
<a href="#">Saccharomyces cerevisiae strain DBVPG6765 chromosome XII sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5093	100%	0.0	100.00%	1022186	<a href="#">CP020168.1</a>
<a href="#">Saccharomyces cerevisiae strain S288c chromosome XII sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5087	100%	0.0	100.00%	1075542	<a href="#">CP020134.1</a>
<a href="#">Saccharomyces cerevisiae strain HB_S_GIMBLETTROAD_14 chromosome XII sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5093	100%	0.0	100.00%	1077529	<a href="#">CP008264.1</a>
<a href="#">Saccharomyces cerevisiae strain HB_C_TUKITUKI1_16 chromosome XII sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5093	100%	0.0	100.00%	1077438	<a href="#">CP008400.1</a>
<a href="#">Saccharomyces cerevisiae strain HB_C_TUKITUKI2_10 chromosome XII sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5093	100%	0.0	100.00%	1077494	<a href="#">CP008383.1</a>
<a href="#">Saccharomyces cerevisiae strain W1_S_JASA_5 chromosome XII sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5093	100%	0.0	100.00%	1076962	<a href="#">CP008349.1</a>
<a href="#">Saccharomyces cerevisiae strain T52 chromosome XII sequence</a>	<a href="#">Saccharomyces cerevisiae</a>	3125	5089	100%	0.0	100.00%	1077515	<a href="#">CP008502.1</a>

Now, here we can clearly see the BLAST of PDC-1 against various *Saccharomyces cerevisiae* strains. Along with this we also have their alignments with PDC-1 and with percent identity, E value, scores, etc. There were more than 600 strains displayed on keeping the number of records to a maximum allowed value in NCBI BLAST. Hence, this shows the BLAST of *Saccharomyces cerevisiae* strains with the PDC-1 gene. A similar approach can be used to perform the BLAST with every PDC gene (I performed this in my code).

Now for further SNP analysis, I selected some of the *Saccharomyces cerevisiae* strains and wrote a Python code to find the SNPs between the PDC gene and strain sequence.

I selected these two strains: *Saccharomyces cerevisiae* YJM996 and *Saccharomyces cerevisiae* YJM1199. The script I wrote prints the position of SNP change along with the base being changed.