machine learning(732A99) lab
2 block 2

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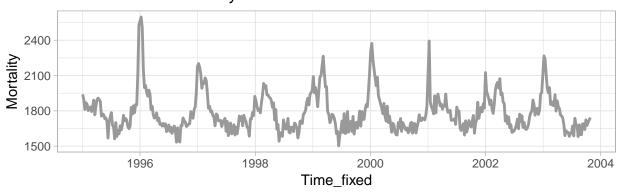
Assignment 1

Loading The Libraries

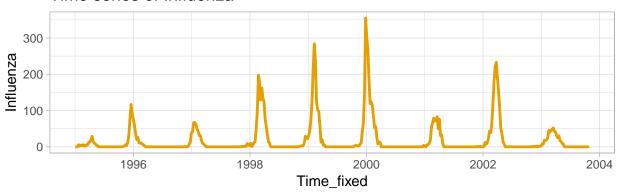
1. Use time series plots to visually inspect how the mortality and influenza number vary with time (use Time as X axis). By using this plot, comment how the amounts of influenza cases are related to mortality rates.

```
set.seed(12345)
# Importing data
flu_data = read.xlsx("influenza.xlsx", sheetName = "Raw data")
flu_data$Time_fixed <- as.Date(paste(flu_data$Year, flu_data$Week, 1, sep="-"), "%Y-%U-%u")
flu_data$influ_perc <- (flu_data$Influenza/flu_data$Mortality) * 100</pre>
# Plot
p1 <- ggplot(flu_data, aes(x=Time_fixed, y = Mortality)) +</pre>
  geom_line(color = "#999999", size = 1) +
    scale_fill_brewer() +
      theme_light() +
  ggtitle("Time series of Mortality")
p2 <- ggplot(flu_data, aes(x=Time_fixed, y = Influenza)) +</pre>
  geom_line(color = "#E69F00", size = 1) +
      scale_fill_brewer() +
      theme light() +
  ggtitle("Time series of Influenza")
p3 <- ggplot(flu_data, aes(x=Time_fixed, y = influ_perc)) +
  geom_line(color = "#56B4E9", size = 1) +
      scale_fill_brewer() +
      theme_light() +
  ggtitle("Time series of % Mortalitiy due to Influenza")
gridExtra::grid.arrange(p1, p2, ncol=1)
```

Time series of Mortality

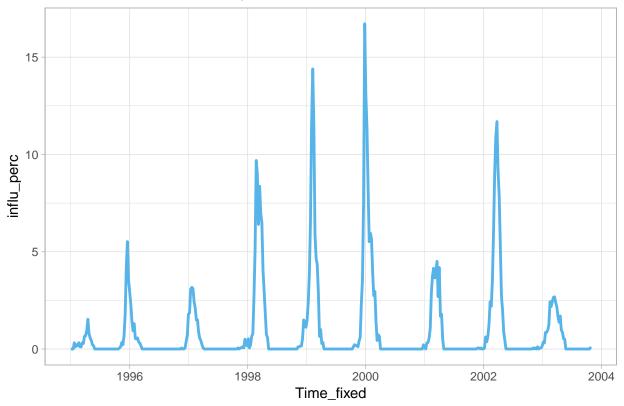


Time series of Influenza



рЗ





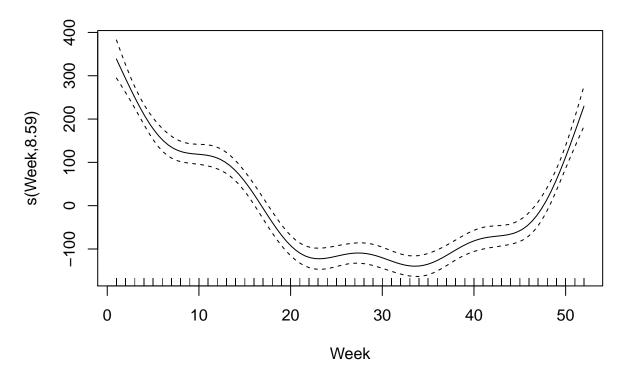
Analsis: From the plots is we can defintely see that Influenza and Mortality in the given dataset are in sync, everytime Mortality peaks so does influenza, however the magnitude of peaking is not in sync, that is the highest cases of mortality were observed in '1996' while for influenza its in year '2000'.

From the third plot, we can see the percentage of mortality due to influenza, here also the peaks match with the other plots, suggests that these two events are closely correleated.

2. Use gam() function from mgcv package to fit a GAM model in which Mortality is normally distributed and modelled as a linear function of Year and spline function of Week, and make sure that the model parameters are selected by the generalized cross-validation. Report the underlying probabilistic model.

```
gam_model <- mgcv::gam(data = flu_data, Mortality~Year+s(Week), method = "GCV.Cp")</pre>
summary(gam_model)
##
## Family: gaussian
## Link function: identity
##
## Formula:
## Mortality ~ Year + s(Week)
##
## Parametric coefficients:
##
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) -652.060
                           3448.379
                                    -0.189
                                                 0.85
                  1.219
                              1.725
                                      0.706
                                                 0.48
## Year
```

Plot of GAM fit on Flu Data



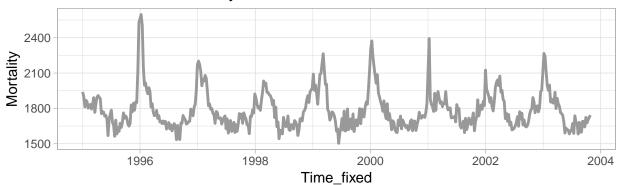
3. Plot predicted and observed mortality against time for the fitted model and comment on the quality of the fit. Investigate the output of the GAM model and report which terms appear to be significant in the model. Is there a trend in mortality change from one year to another? Plot the spline component and interpret the plot.

```
temp <- flu_data
temp$Fitted_Mortality <- gam_model$fitted.values

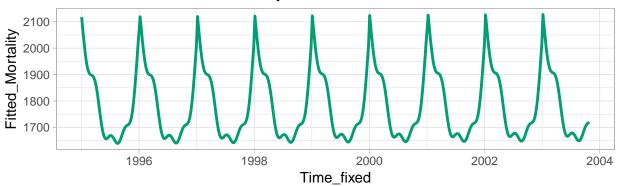
p5 <- ggplot(data=temp, aes(x = Time_fixed, y = Fitted_Mortality)) +
    geom_line(color = "#009E73", size = 1) +</pre>
```

```
scale_fill_brewer() +
    theme_light() +
ggtitle("Time series of Fitted Mortality")
grid.arrange(p1, p5, nrow = 2)
```

Time series of Mortality



Time series of Fitted Mortality

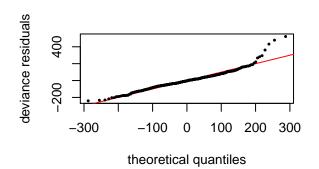


summary(gam_model)

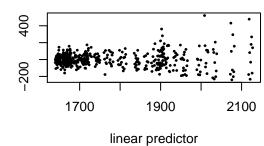
```
## Family: gaussian
## Link function: identity
##
## Formula:
## Mortality ~ Year + s(Week)
##
## Parametric coefficients:
            Estimate Std. Error t value Pr(>|t|)
## (Intercept) -652.060
                      3448.379 -0.189
                                       0.85
##
## Approximate significance of smooth terms:
           edf Ref.df
                       F
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
```

```
## R-sq.(adj) = 0.661 Deviance explained = 66.8%
## GCV = 9014.6 Scale est. = 8806.7 n = 459
gam.check(gam_model,pch=19,cex=.3)
```

esiduals

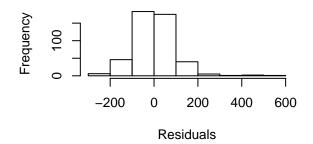


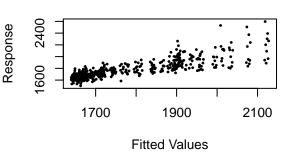
Resids vs. linear pred.



Histogram of residuals

Response vs. Fitted Values





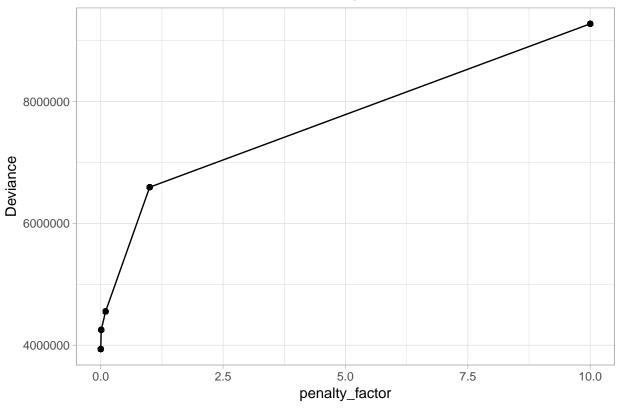
```
##
## Method: GCV
                 Optimizer: magic
## Smoothing parameter selection converged after 7 iterations.
\mbox{\tt \#\#} The RMS GCV score gradient at convergence was 0.114505 .
## The Hessian was positive definite.
## Model rank = 11 / 11
##
## Basis dimension (k) checking results. Low p-value (k-index<1) may
## indicate that k is too low, especially if edf is close to k'.
##
##
                edf k-index p-value
## s(Week) 9.00 8.59
                         1.04
                                 0.74
s=interp(temp$Year,temp$Week, fitted(gam_model))
persp3d(s$x, s$y, s$z, col="red")
```

Analysis: From the plot of residuals we can see that the resisuals are normally distributed. Thus this is a good fit.

4. Examine how the penalty factor of the spline function in the GAM model from step 2 influences the estimated deviance of the model. Make plots of the predicted and observed mortality against time for cases of very high and very low penalty factors. What is the relation of the penalty factor to the degrees of freedom? Do your results confirm this relationship?

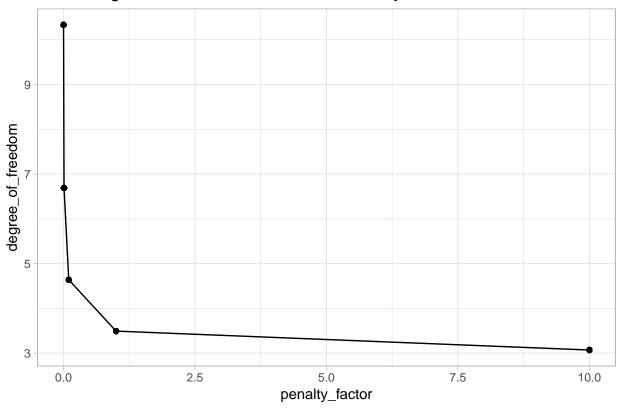
```
model deviance <- NULL
for(sp in c(0.001, 0.01, 0.1, 1, 10))
  k=length(unique(flu_data$Week))
gam_model <- mgcv::gam(data = flu_data, Mortality~Year+s(Week, k=k, sp=sp), method = "GCV.Cp")</pre>
temp <- cbind(gam_model$deviance, gam_model$fitted.values, gam_model$y, flu_data$Time_fixed,
              sp, sum(influence(gam_model)))
model_deviance <- rbind(temp, model_deviance)</pre>
}
model_deviance <- as.data.frame(model_deviance)</pre>
colnames(model_deviance) <- c("Deviance", "Predicted_Mortality", "Mortality", "Time",</pre>
                               "penalty_factor", "degree_of_freedom")
model_deviance$Time <- as.Date(model_deviance$Time, origin = '1970-01-01')</pre>
# plot of deviance
p6 <- ggplot(data=model_deviance, aes(x = penalty_factor, y = Deviance)) +</pre>
geom_point() +
  geom_line() +
      theme_light() +
ggtitle("Plot of Deviance of Model vs. Penalty Factor")
р6
```

Plot of Deviance of Model vs. Penalty Factor



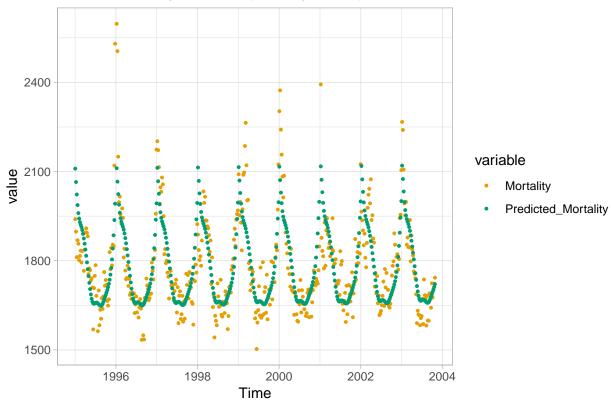
```
# plot of degree of freedom
p7 <- ggplot(data=model_deviance, aes(x = penalty_factor, y = degree_of_freedom)) +
geom_point() +
    geom_line() +
    theme_light() +
ggtitle("Plot of degree_of_freedom of Model vs. Penalty Factor")
p7</pre>
```

Plot of degree_of_freedom of Model vs. Penalty Factor

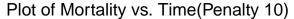


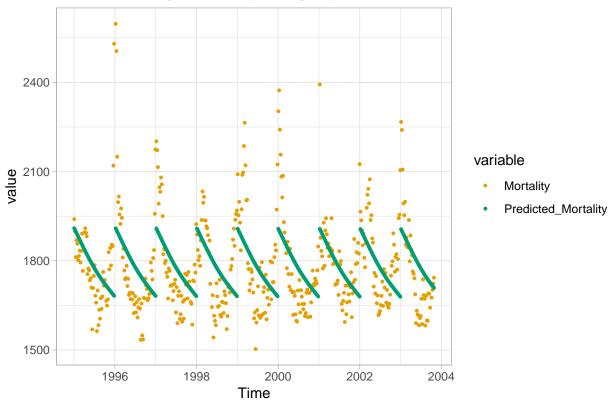
```
model_deviance_wide <- melt(model_deviance[,c("Time", "penalty_factor",</pre>
                                               "Mortality", "Predicted_Mortality")],
                            id.vars = c("Time", "penalty_factor"))
# plot of predicted vs. observed mortality
p8 <- ggplot(data=model_deviance_wide[model_deviance_wide$penalty_factor == 0.001,],
             aes(x= Time, y = value)) +
  geom_point(aes(color = variable), size=0.7) +
  scale_color_manual(values=c("#E69F00", "#009E73")) +
  theme_light() +
  ggtitle("Plot of Mortality vs. Time(Penalty 0.001)")
p9 <- ggplot(data=model_deviance_wide[model_deviance_wide$penalty_factor == 10,],
             aes(x= Time, y = value)) +
  geom_point(aes(color = variable), size=0.7) +
  scale_color_manual(values=c("#E69F00", "#009E73")) +
    theme_light() +
  ggtitle("Plot of Mortality vs. Time(Penalty 10)")
р8
```

Plot of Mortality vs. Time(Penalty 0.001)



р9





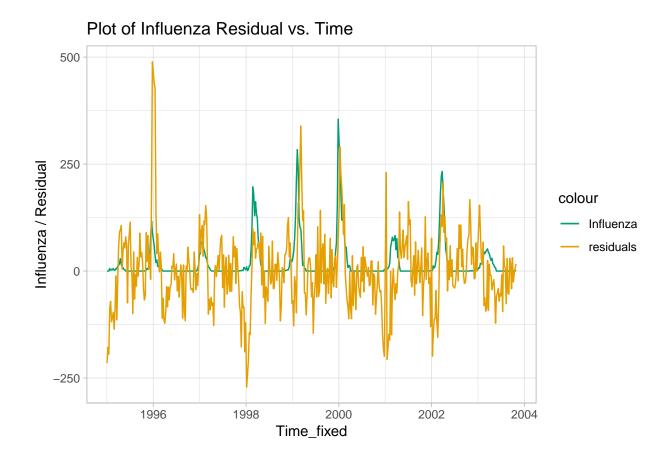
Analysis: theortical maximum degree of freedom is k-1.

5. Use the model obtained in step 2 and plot the residuals and the influenza values against time (in one plot). Is the temporal pattern in the residuals correlated to the outbreaks of influenza?

```
k=length(unique(flu_data$Week))
gam_model <- mgcv::gam(data = flu_data, Mortality~Year+s(Week, k=k), method = "GCV.Cp")

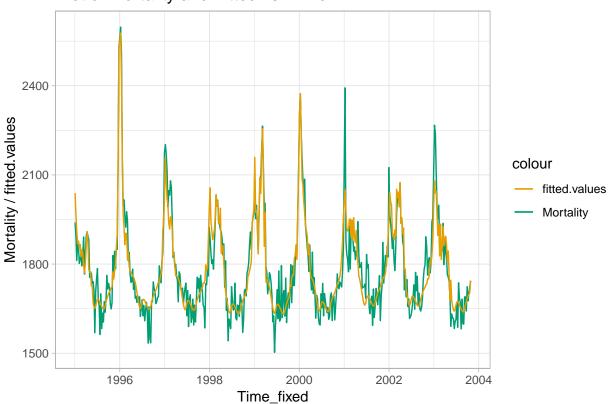
temp <- flu_data
temp <- cbind(temp, residuals = gam_model$residuals)

p10 <- ggplot(data = temp, aes(x = Time_fixed)) +
    geom_line(aes( y = Influenza, color = "Influenza")) +
    geom_line(aes(y = residuals, color = "residuals")) +
        theme_light() +
    scale_color_manual(values=c(Influenza = "#009E73", residuals = "#E69F00")) +
    labs(y = "Influenza / Residual") +
    ggtitle("Plot of Influenza Residual vs. Time")</pre>
```



6. Fit a GAM model in R in which mortality is be modelled as an additive function of the spline functions of year, week, and the number of confirmed cases of influenza. Use the output of this GAM function to conclude whether or not the mortality is influenced by the outbreaks of influenza. Provide the plot of the original and fitted Mortality against Time and comment whether the model seems to be better than the previous GAM models.

Plot of Mortality and Fitted vs. Time



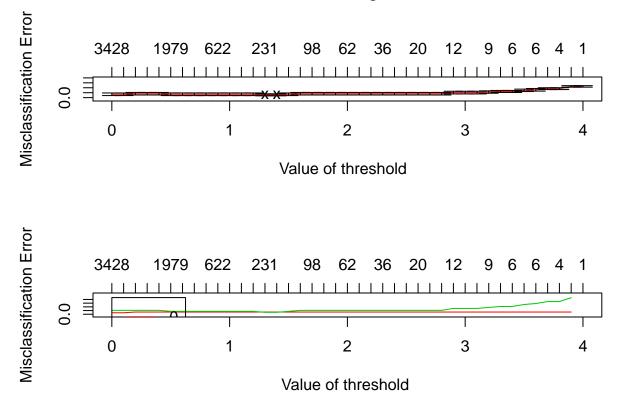
Assignment 2

1. Divide data into training and test sets (70/30) without scaling. Perform nearest shrunken centroid classification of training data in which the threshold is chosen by cross-validation. Provide a centroid plot and interpret it. How many features were selected by the method? List the names of the 10 most contributing features and comment whether it is reasonable that they have strong effect on the discrimination between the conference mails and other mails? Report the test error.

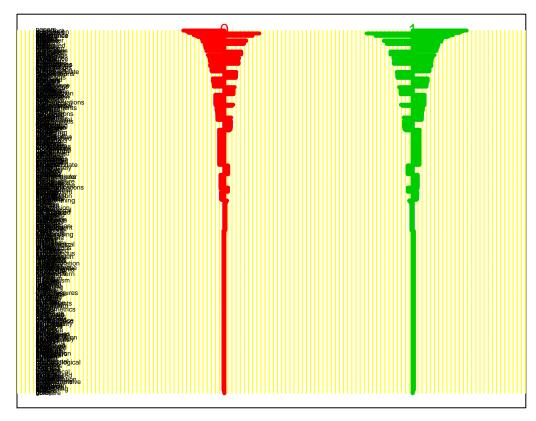
```
rm(list=ls())
gc()
```

```
used (Mb) gc trigger (Mb) max used (Mb)
## Ncells 2906655 155.3 4746755 253.6 4746755 253.6
## Vcells 4769045 36.4 10146329 77.5 8388608 64.0
data <- read.csv(file = "data.csv", sep = ";", header = TRUE)</pre>
n=NROW(data)
data$Conference <- as.factor(data$Conference)</pre>
set.seed(12345)
id=sample(1:n, floor(n*0.7))
train=data[id,]
test = data[-id,]
rownames(train)=1:nrow(train)
x=t(train[,-4703])
y=train[[4703]]
rownames(test)=1:nrow(test)
x_{\text{test=t}}(\text{test}[,-4703])
y_test=test[[4703]]
mydata = list(x=x,y=as.factor(y),geneid=as.character(1:nrow(x)), genenames=rownames(x))
mydata_test = list(x=x_test,y=as.factor(y_test),geneid=as.character(1:nrow(x)), genenames=rownames(x))
model=pamr.train(mydata,threshold=seq(0,4, 0.1))
cvmodel=pamr.cv(model, mydata)
important_gen <- as.data.frame(pamr.listgenes(model, mydata, threshold = 0.9))</pre>
predicted_scc_test <- pamr.predict(model, newx = x_test, threshold = 0.9)</pre>
pamr.plotcv(cvmodel)
```

Number of genes



pamr.plotcen(model, mydata, threshold = 0.9)



```
conf_scc <- table(y_test, predicted_scc_test)</pre>
names(dimnames(conf_scc)) <- c("Actual Test", "Predicted Srunken Centroid Test")</pre>
result_scc <- caret::confusionMatrix(conf_scc)</pre>
caret::confusionMatrix(conf_scc)
## Confusion Matrix and Statistics
##
##
              Predicted Srunken Centroid Test
## Actual Test 0 1
##
             0 10 0
             1 2 8
##
##
##
                  Accuracy: 0.9
##
                    95% CI: (0.683, 0.9877)
       No Information Rate: 0.6
##
##
       P-Value [Acc > NIR] : 0.003611
##
##
                     Kappa : 0.8
    Mcnemar's Test P-Value : 0.479500
##
##
               Sensitivity: 0.8333
##
               Specificity: 1.0000
##
            Pos Pred Value : 1.0000
##
##
            Neg Pred Value: 0.8000
##
                Prevalence: 0.6000
            Detection Rate: 0.5000
##
```

```
## Detection Prevalence : 0.5000
## Balanced Accuracy : 0.9167
##
## 'Positive' Class : 0
##
```

2. Compute the test error and the number of the contributing features for the following methods fitted to the training data: a. Elastic net with the binomial response and alpha = 0.5 in which penalty is selected by the cross-validation. b. Support vector machine with "vanilladot" kernel. Compare the results of these models with the results of the nearest shrunken centroids (make a comparative table). Which model would you prefer and why?

```
x = train[,-4703] %>% as.matrix()
y = train[,4703]
x_{test} = test[,-4703] \% as.matrix()
y_{test} = test[,4703]
cvfit = cv.glmnet(x=x, y=y, alpha = 0.5, family = "binomial")
predicted_elastic_test <- predict.cv.glmnet(cvfit, newx = x_test, s = "lambda.min", type = "class")</pre>
conf_elastic_net <- table(y_test, predicted_elastic_test)</pre>
names(dimnames(conf_elastic_net)) <- c("Actual Test", "Predicted ElasticNet Test")</pre>
result_elastic_net <- caret::confusionMatrix(conf_elastic_net)</pre>
caret::confusionMatrix(conf_elastic_net)
## Confusion Matrix and Statistics
##
##
              Predicted ElasticNet Test
## Actual Test 0 1
##
             0 10 0
             1 2 8
##
##
##
                  Accuracy: 0.9
##
                    95% CI: (0.683, 0.9877)
       No Information Rate: 0.6
##
       P-Value [Acc > NIR] : 0.003611
##
##
##
                     Kappa : 0.8
##
    Mcnemar's Test P-Value: 0.479500
##
##
               Sensitivity: 0.8333
##
               Specificity: 1.0000
##
            Pos Pred Value: 1.0000
##
            Neg Pred Value: 0.8000
##
                Prevalence: 0.6000
##
            Detection Rate: 0.5000
##
      Detection Prevalence: 0.5000
##
         Balanced Accuracy: 0.9167
##
##
          'Positive' Class: 0
```

```
##
# svm
svm_fit <- kernlab::ksvm(x, y, kernel="vanilladot", scale = FALSE, type = "C-svc")</pre>
## Setting default kernel parameters
predicted_svm_test <- predict(svm_fit, x_test, type="response")</pre>
conf_svm_tree <- table(y_test, predicted_svm_test)</pre>
names(dimnames(conf_svm_tree)) <- c("Actual Test", "Predicted SVM Test")</pre>
result_svm <- caret::confusionMatrix(conf_svm_tree)</pre>
caret::confusionMatrix(conf_svm_tree)
## Confusion Matrix and Statistics
##
              Predicted SVM Test
## Actual Test 0 1
             0 10 0
##
##
             1 1 9
##
##
                  Accuracy: 0.95
##
                     95% CI: (0.7513, 0.9987)
##
       No Information Rate: 0.55
##
       P-Value [Acc > NIR] : 0.0001114
##
##
                      Kappa : 0.9
##
   Mcnemar's Test P-Value: 1.0000000
##
##
               Sensitivity: 0.9091
               Specificity: 1.0000
##
##
            Pos Pred Value: 1.0000
            Neg Pred Value: 0.9000
##
                Prevalence: 0.5500
##
##
            Detection Rate: 0.5000
##
      Detection Prevalence: 0.5000
##
         Balanced Accuracy: 0.9545
##
##
          'Positive' Class: 0
##
# creating table
final_result <- cbind(result_scc$overall[[1]]*100,</pre>
                      result_elastic_net$overall[[1]]*100,
                      result_svm$overall[[1]] *100) %>% as.data.frame()
colnames(final_result) <- c("Accuracy of Nearest Shrunken Centroid Model",</pre>
                                    "Accuracy of ElasticNet",
                                    "Accuracy SVM Model")
knitr::kable(final_result, caption = "Accuracy of Model on Test dataset")
```

Table 1: Accuracy of Model on Test dataset

Accuracy of Nearest Shrunken Centroid Model	Accuracy of ElasticNet	Accuracy SVM Model
90	90	95

3. Implement Benjamini-Hochberg method for the original data, and use t.test() for computing p-values. Which features correspond to the rejected hypotheses? Interpret the result.

Appendix

```
knitr::opts chunk$set(echo = TRUE)
if (!require("pacman")) install.packages("pacman")
pacman::p_load(xlsx, ggplot2, tidyr, dplyr, reshape2, gridExtra,
               mgcv, rgl, akima, pamr, caret, glmnet, kernlab)
set.seed(12345)
options("jtools-digits" = 2, scipen = 999)
# colours (colour blind friendly)
cbPalette <- c("#999999", "#E69F00", "#56B4E9", "#009E73", "#F0E442", "#0072B2",
               "#D55E00", "#CC79A7")
## Making title in the center
theme_update(plot.title = element_text(hjust = 0.5))
set.seed(12345)
# Importing data
flu data = read.xlsx("influenza.xlsx", sheetName = "Raw data")
flu_data$Time_fixed <- as.Date(paste(flu_data$Year, flu_data$Week, 1, sep="-"), "%Y-%U-%u")
flu_data$influ_perc <- (flu_data$Influenza/flu_data$Mortality) * 100</pre>
# Plot
p1 <- ggplot(flu_data, aes(x=Time_fixed, y = Mortality)) +
  geom_line(color = "#999999", size = 1) +
   scale_fill_brewer() +
      theme_light() +
  ggtitle("Time series of Mortality")
p2 <- ggplot(flu data, aes(x=Time fixed, y = Influenza)) +
  geom line(color = "#E69F00", size = 1) +
      scale_fill_brewer() +
      theme_light() +
  ggtitle("Time series of Influenza")
p3 <- ggplot(flu_data, aes(x=Time_fixed, y = influ_perc)) +
  geom_line(color = "#56B4E9", size = 1) +
      scale_fill_brewer() +
      theme_light() +
  ggtitle("Time series of % Mortalitiy due to Influenza")
gridExtra::grid.arrange(p1, p2, ncol=1)
```

```
рЗ
gam model <- mgcv::gam(data = flu data, Mortality~Year+s(Week), method = "GCV.Cp")</pre>
summary(gam model)
#plot the fit
p4 <- plot(gam_model, main= "Plot of GAM fit on Flu Data")
temp <- flu_data
temp$Fitted_Mortality <- gam_model$fitted.values</pre>
p5 <- ggplot(data=temp, aes(x = Time_fixed, y = Fitted_Mortality)) +
   geom_line(color = "#009E73", size = 1) +
    scale_fill_brewer() +
      theme_light() +
  ggtitle("Time series of Fitted Mortality")
grid.arrange(p1, p5, nrow = 2)
summary(gam_model)
gam.check(gam_model,pch=19,cex=.3)
s=interp(temp$Year,temp$Week, fitted(gam_model))
persp3d(s$x, s$y, s$z, col="red")
model_deviance <- NULL</pre>
for(sp in c(0.001, 0.01, 0.1, 1, 10))
  k=length(unique(flu_data$Week))
gam_model <- mgcv::gam(data = flu_data, Mortality~Year+s(Week, k=k, sp=sp), method = "GCV.Cp")</pre>
temp <- cbind(gam_model$deviance, gam_model$fitted.values, gam_model$y, flu_data$Time_fixed,
              sp, sum(influence(gam_model)))
model_deviance <- rbind(temp, model_deviance)</pre>
model deviance <- as.data.frame(model deviance)</pre>
colnames(model_deviance) <- c("Deviance", "Predicted_Mortality", "Mortality", "Time",</pre>
                                "penalty_factor", "degree_of_freedom")
model_deviance$Time <- as.Date(model_deviance$Time, origin = '1970-01-01')</pre>
# plot of deviance
p6 <- ggplot(data=model_deviance, aes(x = penalty_factor, y = Deviance)) +</pre>
geom_point() +
  geom_line() +
      theme_light() +
ggtitle("Plot of Deviance of Model vs. Penalty Factor")
p6
# plot of degree of freedom
p7 <- ggplot(data=model_deviance, aes(x = penalty_factor, y = degree_of_freedom)) +
geom_point() +
  geom_line() +
      theme_light() +
```

```
ggtitle("Plot of degree_of_freedom of Model vs. Penalty Factor")
р7
model_deviance_wide <- melt(model_deviance[,c("Time", "penalty_factor",</pre>
                                                "Mortality", "Predicted_Mortality")],
                             id.vars = c("Time", "penalty_factor"))
# plot of predicted vs. observed mortality
p8 <- ggplot(data=model_deviance_wide[model_deviance_wide$penalty_factor == 0.001,],
             aes(x= Time, y = value)) +
  geom_point(aes(color = variable), size=0.7) +
  scale_color_manual(values=c("#E69F00", "#009E73")) +
  theme_light() +
  ggtitle("Plot of Mortality vs. Time(Penalty 0.001)")
p9 <- ggplot(data=model_deviance_wide[model_deviance_wide$penalty_factor == 10,],
             aes(x= Time, y = value)) +
  geom_point(aes(color = variable), size=0.7) +
  scale_color_manual(values=c("#E69F00", "#009E73")) +
    theme_light() +
  ggtitle("Plot of Mortality vs. Time(Penalty 10)")
8q
p9
k=length(unique(flu_data$Week))
gam_model <- mgcv::gam(data = flu_data, Mortality~Year+s(Week, k=k), method = "GCV.Cp")</pre>
temp <- flu_data</pre>
temp <- cbind(temp, residuals = gam_model$residuals)</pre>
p10 <- ggplot(data = temp, aes(x = Time_fixed)) +
  geom_line(aes( y = Influenza, color = "Influenza")) +
  geom_line(aes(y = residuals, color = "residuals")) +
      theme_light() +
  scale_color_manual(values=c(Influenza = "#009E73", residuals = "#E69F00")) +
  labs(y = "Influenza / Residual") +
  ggtitle("Plot of Influenza Residual vs. Time")
p10
\#gam\_model\_additive \leftarrow mgcv::gam(data = flu\_data, Mortality \sim s(Year) + s(Week), method = "GCV.Cp")
k1 = length(unique(flu_data$Year))
k2 = length(unique(flu_data$Week))
k3 = length(unique(flu_data$Influenza))
gam_model_additive <- gam(Mortality ~ s(Year, k=k1) +</pre>
                                      s(Week, k=k2) +
                                     s(Influenza, k=k3),
                           data = flu_data)
```

```
flu_data$fitted.values = gam_model_additive$fitted.values
p11 <- ggplot(data = flu_data, aes(x = Time_fixed)) +
  geom_line(aes( y = Mortality, color = "Mortality")) +
  geom_line(aes(y = fitted.values, color = "fitted.values")) +
      theme_light() +
  scale color manual(values=c(Mortality = "#009E73", fitted.values = "#E69F00")) +
  labs(y = "Mortality / fitted.values") +
  ggtitle("Plot of Mortality and Fitted vs. Time")
p11
rm(list=ls())
gc()
data <- read.csv(file = "data.csv", sep = ";", header = TRUE)</pre>
n=NROW(data)
data$Conference <- as.factor(data$Conference)</pre>
set.seed(12345)
id=sample(1:n, floor(n*0.7))
train=data[id,]
test = data[-id,]
rownames(train)=1:nrow(train)
x=t(train[,-4703])
y=train[[4703]]
rownames(test)=1:nrow(test)
x_test=t(test[,-4703])
y_test=test[[4703]]
mydata = list(x=x,y=as.factor(y),geneid=as.character(1:nrow(x)), genenames=rownames(x))
mydata_test = list(x=x_test,y=as.factor(y_test),geneid=as.character(1:nrow(x)), genenames=rownames(x))
model=pamr.train(mydata,threshold=seq(0,4, 0.1))
cvmodel=pamr.cv(model, mydata)
important gen <- as.data.frame(pamr.listgenes(model, mydata, threshold = 0.9))
predicted_scc_test <- pamr.predict(model, newx = x_test, threshold = 0.9)</pre>
pamr.plotcv(cvmodel)
pamr.plotcen(model, mydata, threshold = 0.9)
conf_scc <- table(y_test, predicted_scc_test)</pre>
names(dimnames(conf_scc)) <- c("Actual Test", "Predicted Srunken Centroid Test")</pre>
result_scc <- caret::confusionMatrix(conf_scc)</pre>
caret::confusionMatrix(conf_scc)
x = train[,-4703] %>% as.matrix()
y = train[,4703]
x_{test} = test[,-4703] \% as.matrix()
y_{test} = test[,4703]
cvfit = cv.glmnet(x=x, y=y, alpha = 0.5, family = "binomial")
```

```
predicted_elastic_test <- predict.cv.glmnet(cvfit, newx = x_test, s = "lambda.min", type = "class")</pre>
conf_elastic_net <- table(y_test, predicted_elastic_test)</pre>
names(dimnames(conf_elastic_net)) <- c("Actual Test", "Predicted ElasticNet Test")</pre>
result_elastic_net <- caret::confusionMatrix(conf_elastic_net)</pre>
caret::confusionMatrix(conf_elastic_net)
# svm
svm_fit <- kernlab::ksvm(x, y, kernel="vanilladot", scale = FALSE, type = "C-svc")</pre>
predicted_svm_test <- predict(svm_fit, x_test, type="response")</pre>
conf_svm_tree <- table(y_test, predicted_svm_test)</pre>
names(dimnames(conf_svm_tree)) <- c("Actual Test", "Predicted SVM Test")</pre>
result_svm <- caret::confusionMatrix(conf_svm_tree)</pre>
caret::confusionMatrix(conf_svm_tree)
# creating table
final_result <- cbind(result_scc$overall[[1]]*100,</pre>
                       result_elastic_net$overall[[1]]*100,
                       result_svm$overall[[1]] *100) %>% as.data.frame()
colnames(final_result) <- c("Accuracy of Nearest Shrunken Centroid Model",</pre>
                                     "Accuracy of ElasticNet",
                                     "Accuracy SVM Model")
knitr::kable(final_result, caption = "Accuracy of Model on Test dataset")
```