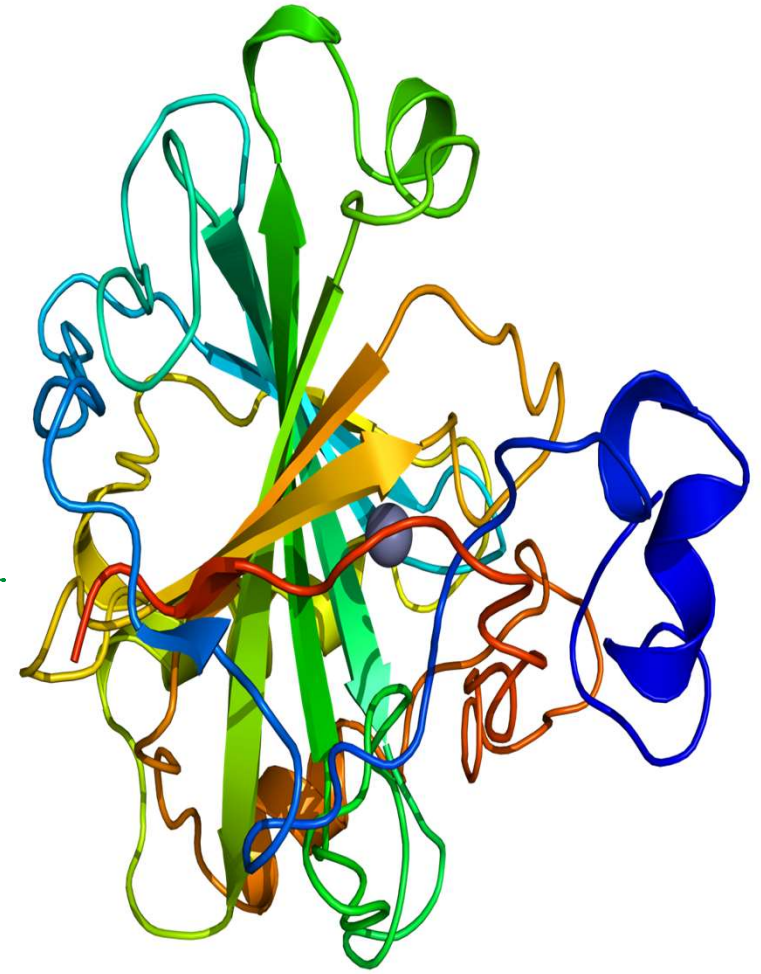


# Enzymes

Suma Somasekharan

# What are Enzymes?

- Enzymes are **biological catalysts** – they accelerate the rate of a reaction
- They are themselves not altered or consumed in the process.
- Functionally they are the largest group of **Proteins**
- The exception is **Ribozymes** which are RNA based enzymes
- Catalyze innumerable reactions- e.g. breakdown of food to generate, energy, DNA synthesis



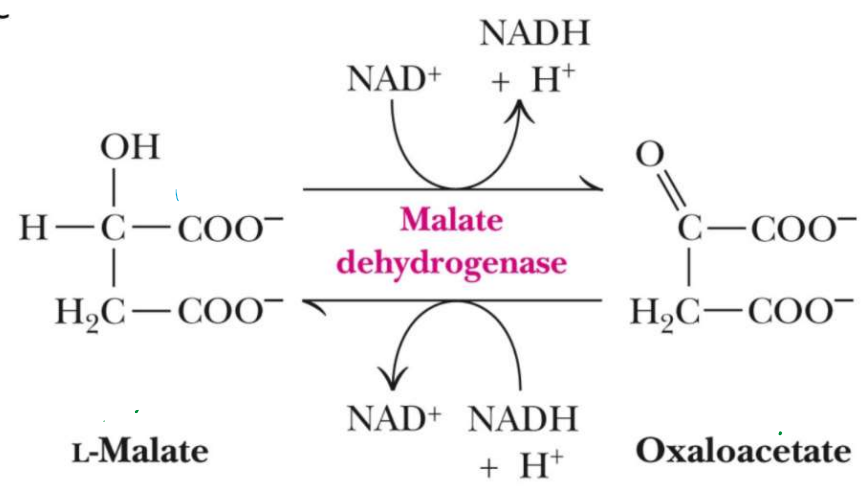
Structure of Carbonic anhydrase

Björn Sjöblom et al. PNAS 2009;106:10609-10613

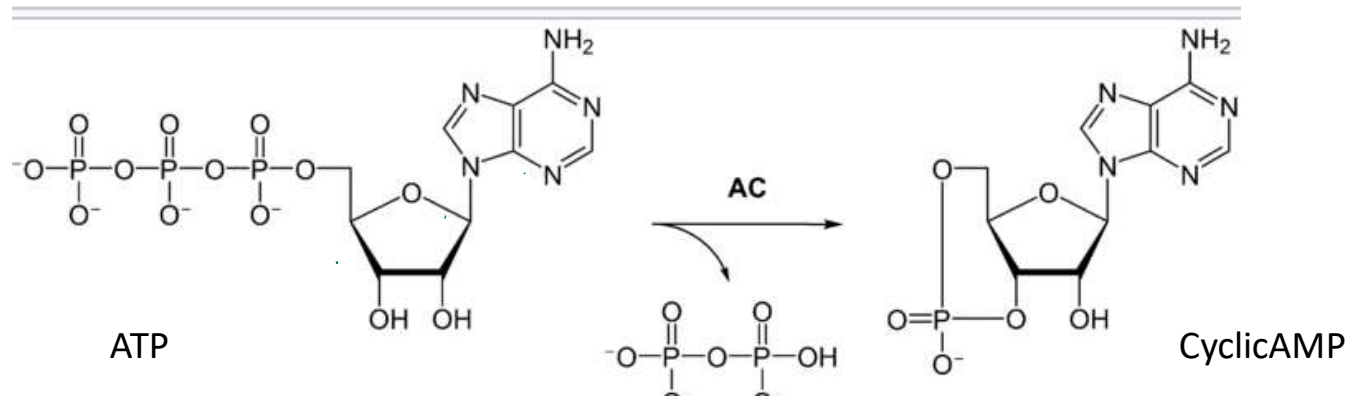
# Enzyme classes-Types of reactions catalyzed by Enzymes

Enzymes	Types of reactions	Examples
Oxidoreductases	Oxidation - Reduction	Pyruvate dehydrogenase, Carbonic anhydrase
Transferases	Group transfers	Aminotransferase, Kinases
Hydrolases	Hydrolysis	Chymotrypsin, Carboxypeptidase
Lyases	Group elimination or elimination of double bond	Adenyl cyclase, Decarboxylases
Isomerases	Isomerization	Isomerases
Ligases	Bond formation coupled with ATP	DNA ligase

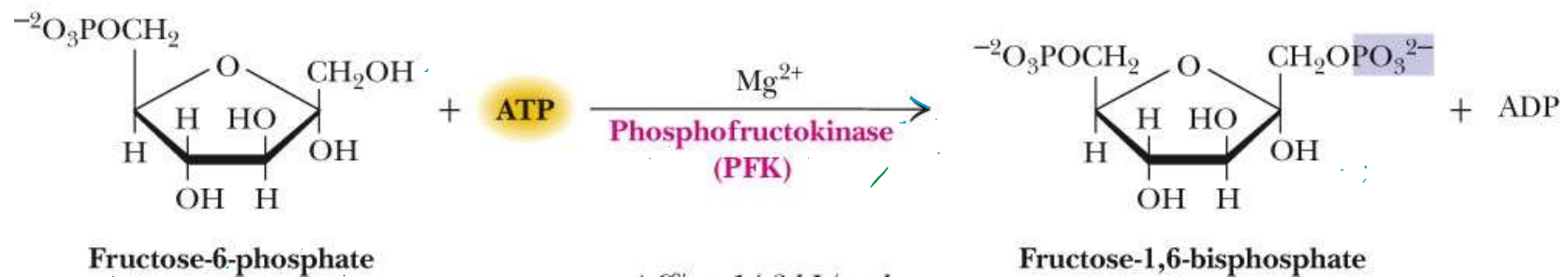
Oxidoreductases



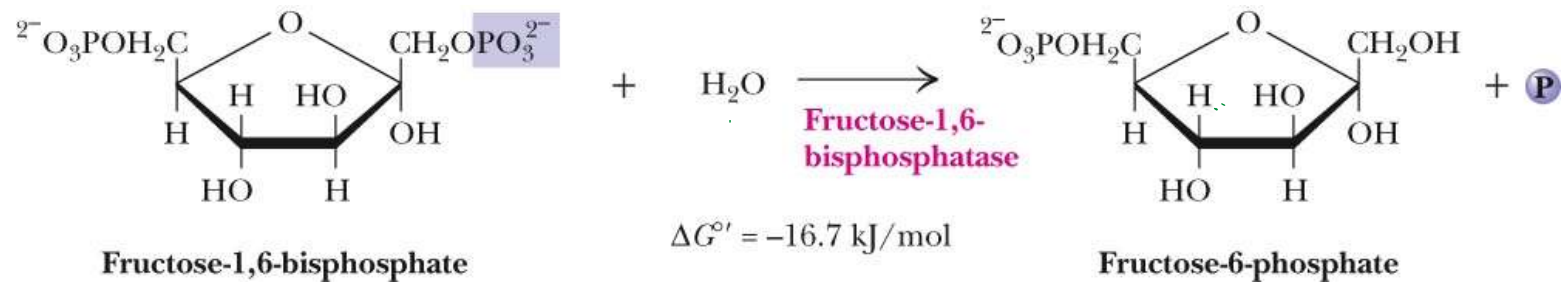
Lyases



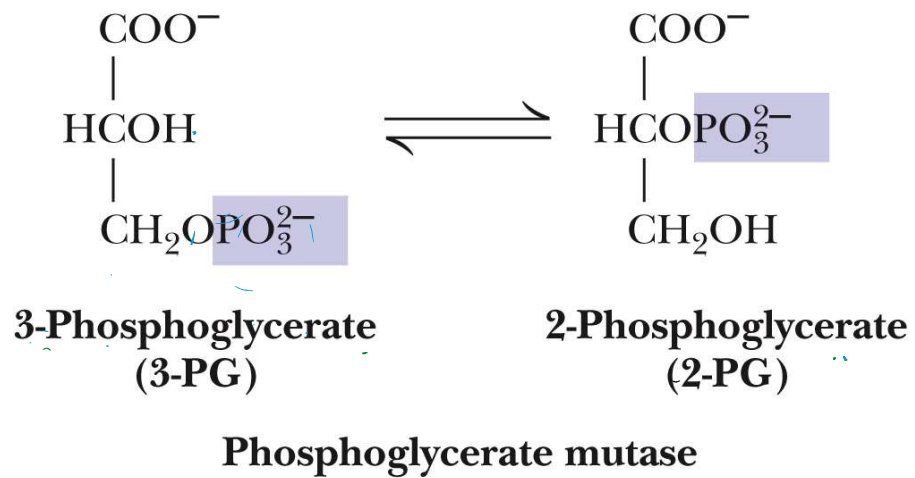
## Transferases



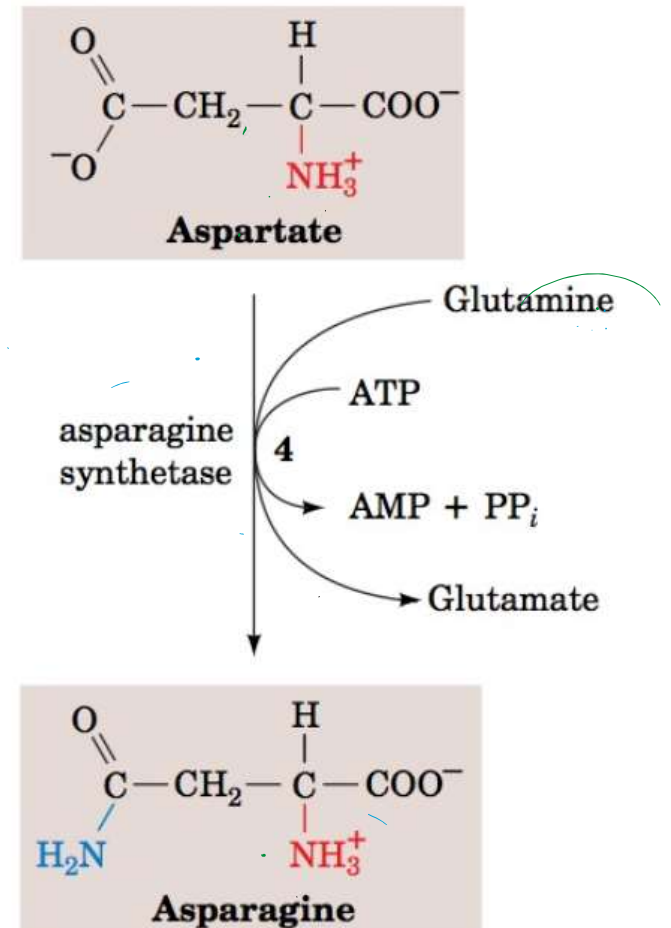
## Hydrolases



## Isomerases



## Ligases



# Catalytic Efficiency

Enzymes bind **one more substrates** and catalyze their **rapid conversion** to **products**.

The rate of an enzyme catalyzed reaction is many orders of magnitude greater than an uncatalyzed reaction

**Turnover rate:** The number of molecules of substrate converted to product per enzyme molecule per second



The enzyme **Carbonic anhydrase**.

It catalyzes this reaction at a rate **million times** faster than the uncatalyzed reaction



The enzyme in saliva- **Alpha amylase**  
Catalyzed reaction is **million times** faster than the uncatalyzed

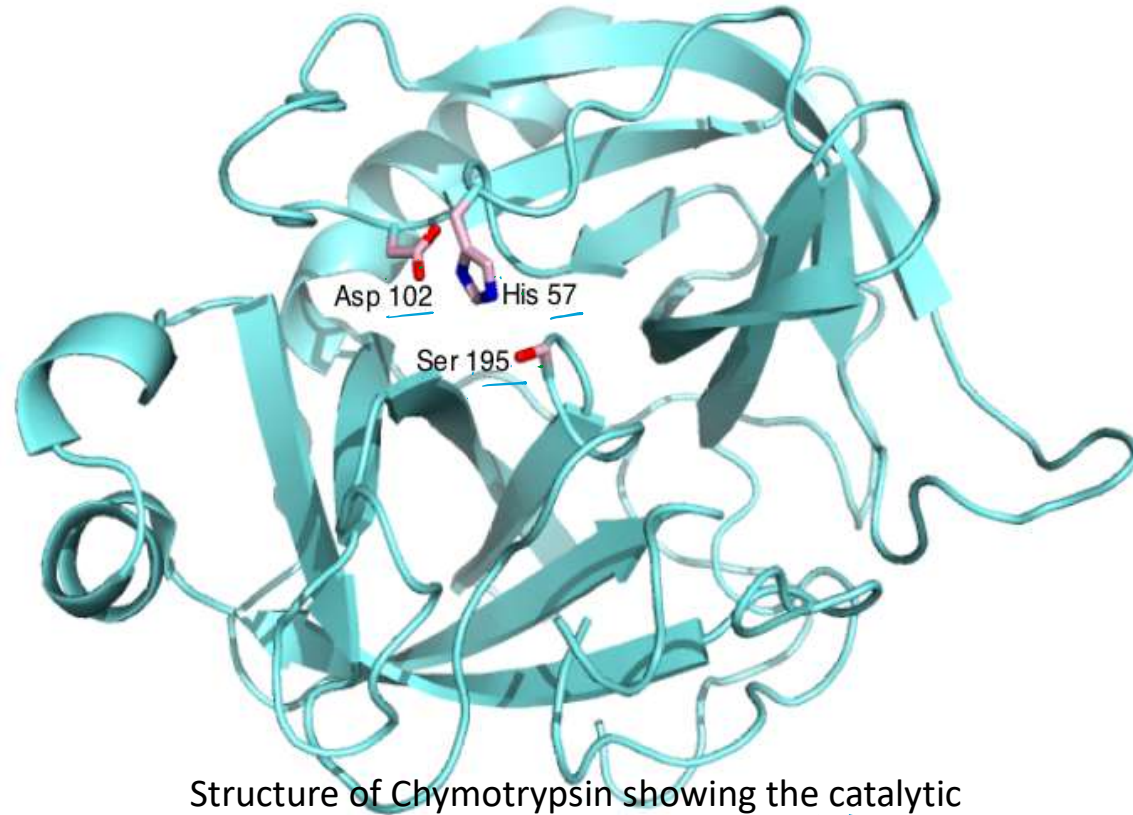
# Properties of Enzymes

- Enzymes show great specificity for one or more substrates and kinds of reactions they catalyze.
- Enzymes catalyze reactions under fairly **mild conditions of temperature and pH**
- Most enzymes have optimal activity at **37 degrees**
- Most enzymes have optimal activity in the pH range of 5 to 9
- Enzyme activity can be **regulated**-
  - by **limiting expression** of the enzyme
  - by **another molecule** other than substrate which binds the enzyme often at a site other than the active site-**Allosteric regulation**



# Structure of Enzymes and their Active site

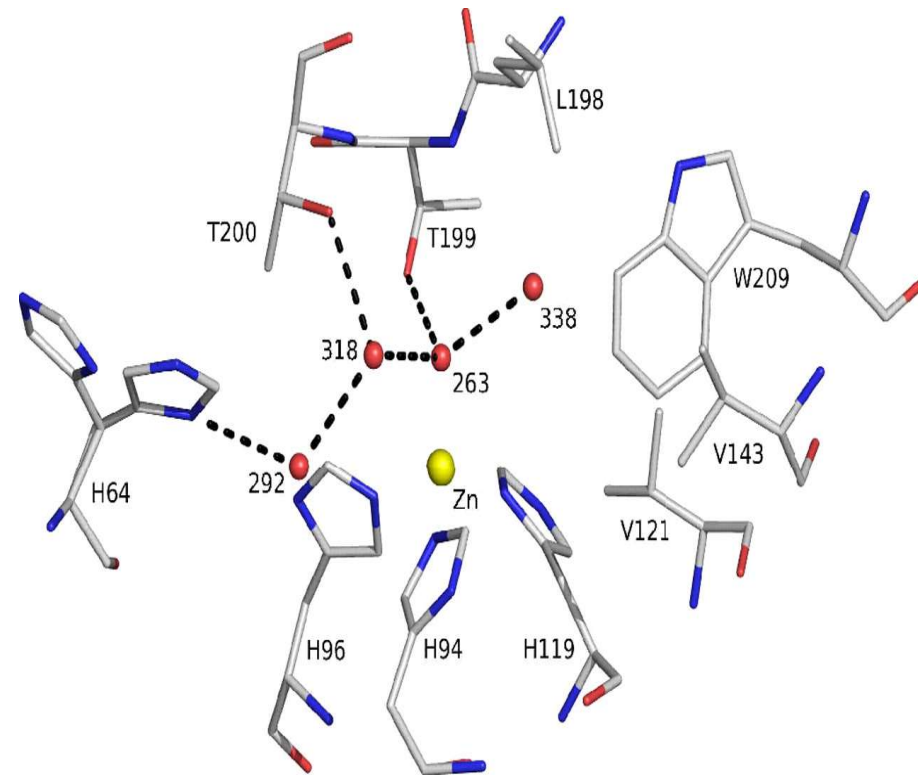
- The three dimensional structure of enzymes –large globular protein with a small **Active site**
- Active site- **small pocket** in the protein which binds substrate
- The active site brings together key residues that are far apart in the primary structure
- The **residues are essential in the catalytic process**-e.g. the catalytic triad in Chymotrypsin



Structure of Chymotrypsin showing the catalytic triad in the active site

# Active site

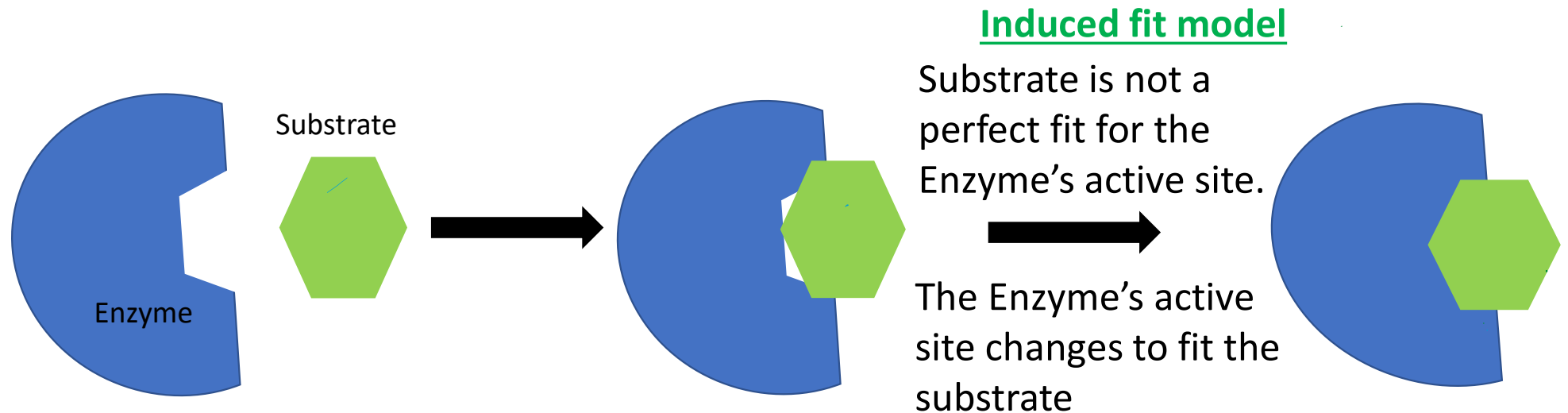
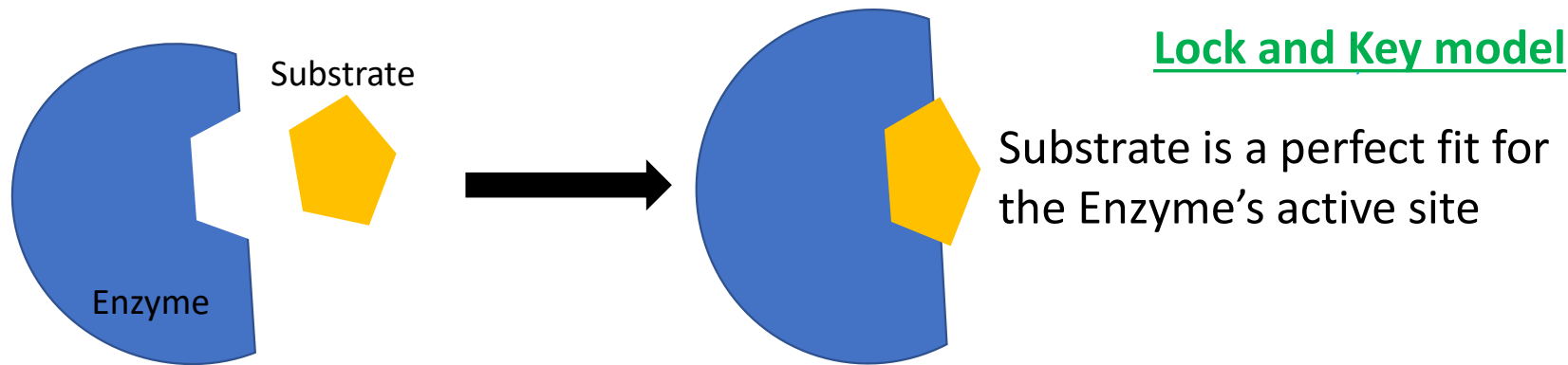
- The active site excludes water except for **catalytic water**
- Can have **metal ions (cofactors)** that are bound to the enzyme and participate in the catalytic process
- Substrate binding and stabilization of the transition state involve **non-covalent bonding between enzyme and substrate**-hydrogen bonding, Vander Waals forces and the hydrophobic effect



The active site of Human Carbonic anhydrase II. The zinc ion is tetrahedrally coordinated by 3 histidine's (His-94, His-96, and His-119) and catalytic water (Wat-263).

Björn Sjöblom et al. PNAS 2009;106:10609-10613

# Substrate Binding



# Substrate specificity

- Enzymes catalyze specific type of reactions and products are defined-no side reactions

➤ Specific substrate or a group of substrates

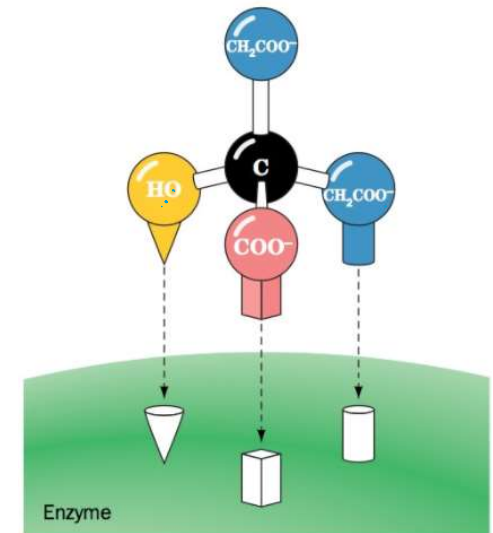
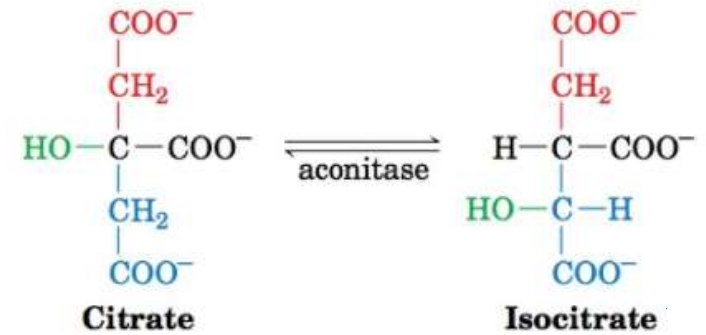
➤ Stereospecificity- Specific for one kind of **chiral molecule**

➤ Enzymes involved in glucose metabolism are specific for D-glucose

➤ Trypsin will only hydrolyze L-amino acids

➤ Geometric specificity- specific for **type of chemical groups** in their substrates

Yeast alcohol dehydrogenase shows a strong specificity for ethanol



# Coenzyme and Cofactors

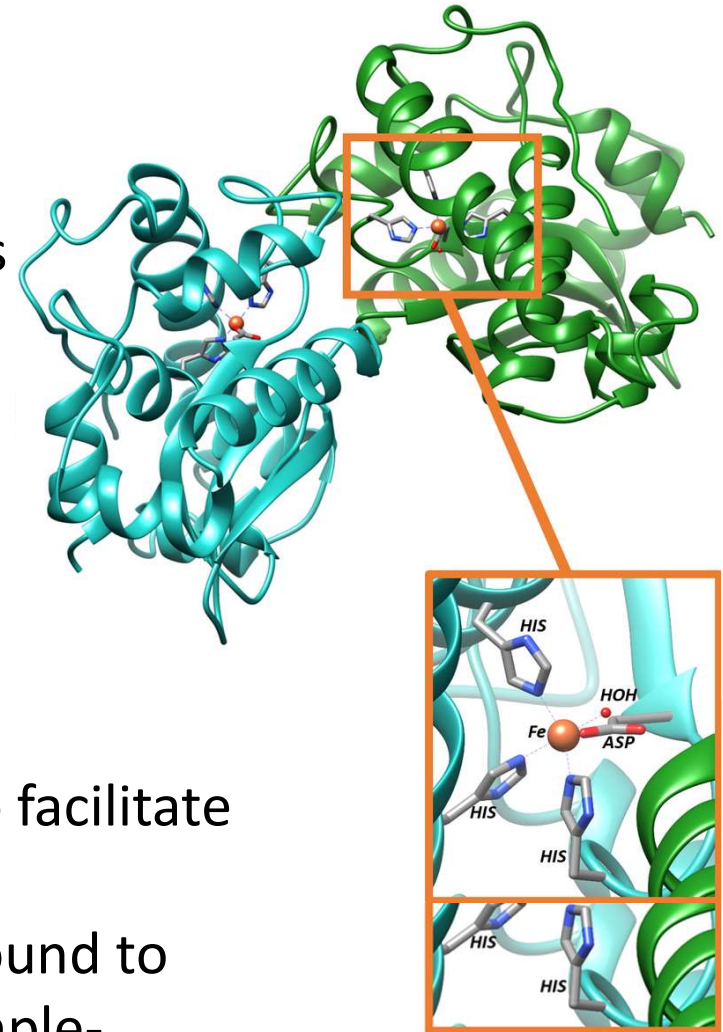
In addition to the amino acid sidechains, enzymes use **Cofactors**-inorganic and organic molecules to aid in catalysis.

Cofactors can be **metal ions** as in the case of **Carbonic anhydrase (CA)** and **Superoxide dismutase(SOD)** and are called **Metalloenzymes**

**Coenzymes** are organic cofactors.

Often they transiently interact in the active site to facilitate catalysis (**co-substrates**)

Some enzymes have an organic cofactor that is bound to the enzyme and is called a **prosthetic group**. Example- Horseradish peroxidase(Hrp) has **Heme**



**Fe-SOD**

Coenzymes can **carry groups or electrons** that are **transferred** to the substrate.

See a list of some common coenzymes

The coenzyme is regenerated after the reaction takes place, sometimes by another reaction.

coenzyme	abbreviation	entity transferred
nicotine adenine dinucleotide	<u>NAD</u> - partly composed of niacin	electron (hydrogen atom)
nicotine adenine dinucleotide phosphate	NADP -Partly composed of niacin	electron (hydrogen atom)
flavine adenine dinucleotide	<u>FAD</u> - Partly composed of riboflavin (vit. B2)	electron (hydrogen atom)
coenzyme A	CoA	Acyl groups $\text{CH}_3 - \overset{\text{O}}{\underset{\text{  }}{\text{C}}}$
coenzymeQ	<u>CoQ</u>	electrons (hydrogen atom)
thiamine pyrophosphate	thiamine (vit. B1)	aldehydes
pyridoxal phosphate	pyridoxine (vit B6)	amino groups
biotin	biotin	carbon dioxide
carbamide coenzymes	vit. B12	alkyl groups

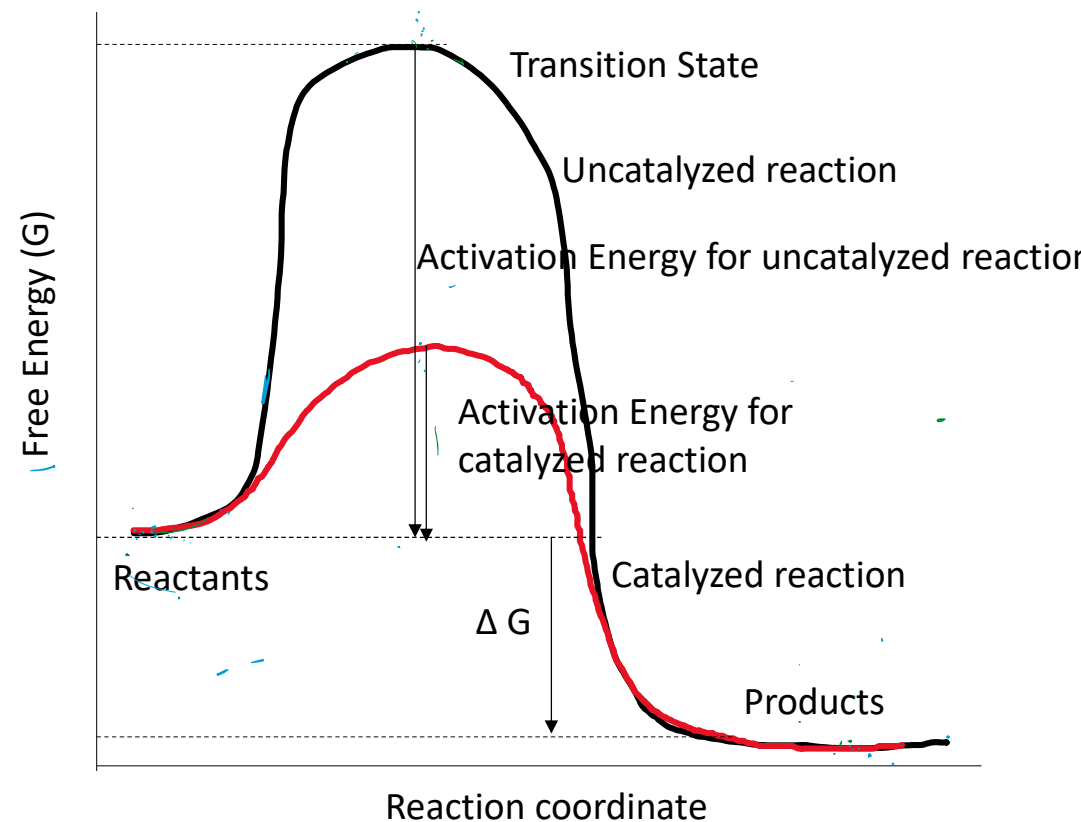
# Mechanism of Enzyme Catalysis

- Stabilization of the transition state
- Proximity and Orientation Effects
- Acid Base Catalysis
- Covalent Catalysis
- Metal ion Catalysis



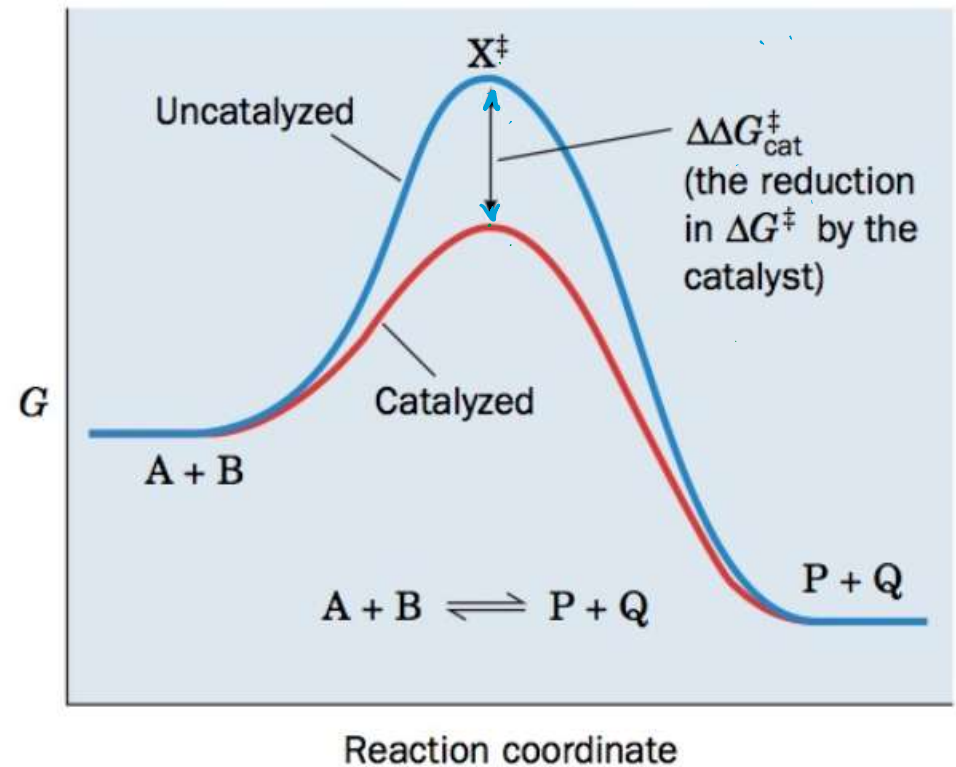
# Thermodynamics of Enzyme action

- $A+B \rightleftharpoons P+Q$
- $A+B \longrightarrow X^\# \longrightarrow P+Q$
- Where the  $X^\#$  is called the **Transition State**
- $\Delta G$  is **negative** –reaction is favorable
- The difference in energy between the transition state and the reactants is called the **Activation Energy –  $G^\#$**





- The uncatalyzed reaction occurs at a fairly low rate because of the high activation energy
- The enzyme increases the rate of this reaction by lowering the activation energy-  $G^\ddagger$
- The enzyme lowers the activation energy by stabilizing the transition state
- The  $\Delta \Delta G^\ddagger_{\text{cat}}$  is a measure of the enzymes efficiency
- The rate enhancement of a reaction by an enzyme is calculated by
- Rate enhancement =  $e^{\Delta \Delta G^\ddagger_{\text{cat}} / RT}$



### SAMPLE CALCULATION 11-1

By how much must an enzyme reduce the activation energy of a reaction at 37°C for the reaction to occur 50 times faster than in the absence of the enzyme?

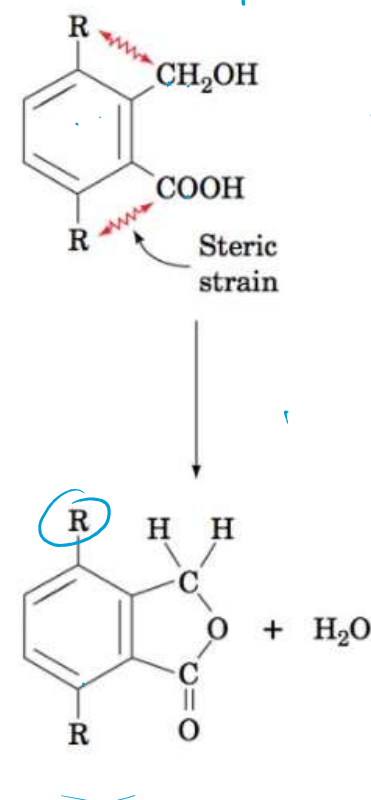
Rate enhancement = 50 =  $e^{\Delta\Delta G_{\text{cat}}^{\ddagger}/RT}$ . Rearranging the equation gives

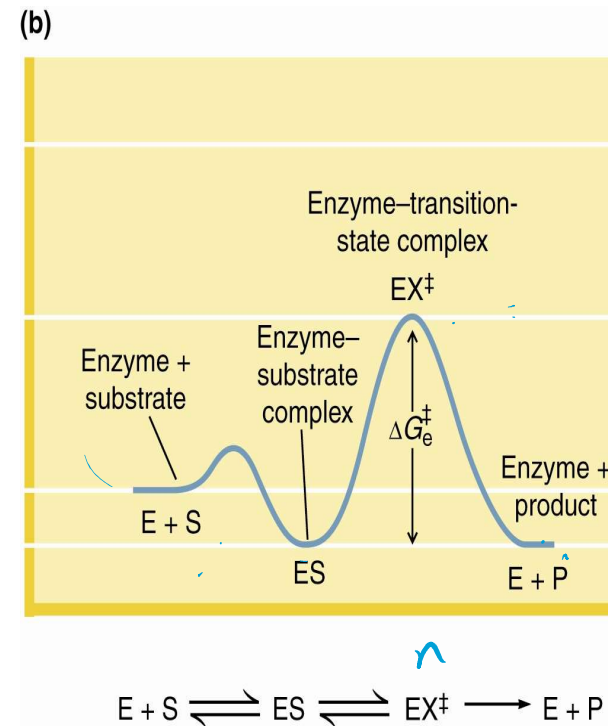
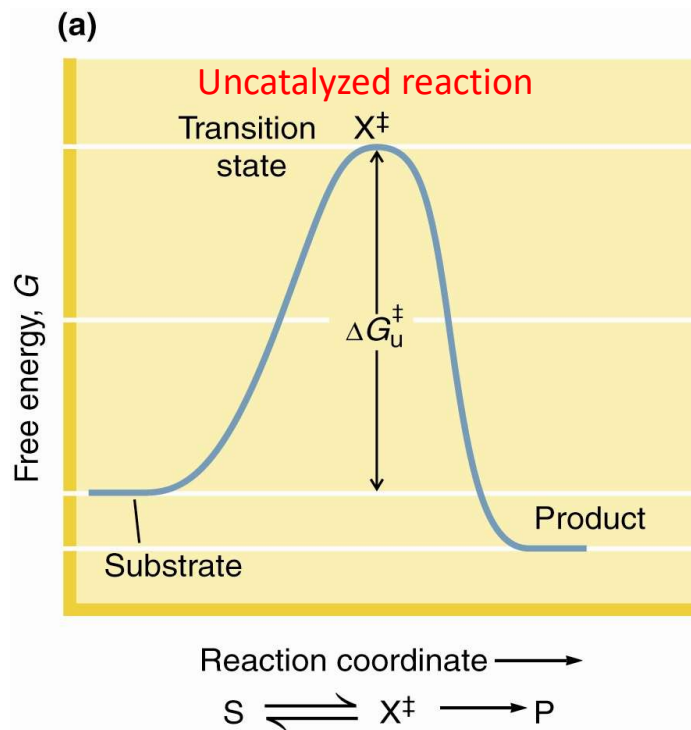
$$\begin{aligned}\ln 50 &= \Delta\Delta G_{\text{cat}}^{\ddagger}/RT \\ \Delta\Delta G_{\text{cat}}^{\ddagger} &= RT \ln 50 \\ &= (8.3145 \text{ J} \cdot \text{K}^{-1} \cdot \text{mol}^{-1}) (37 + 273)\text{K} \ln 50 \\ &= 10,000 \text{ J} \cdot \text{mol}^{-1} = 10 \text{ kJ} \cdot \text{mol}^{-1}\end{aligned}$$

**WPLS** See Sample Calculation Videos.

# Transition state

- The rate of the reaction shown here is 315 times faster if the R group is a  $\text{CH}_3$  molecule than if it's a H molecule.
- The transition state closely resembles the strained reactant than the unstrained reactant.
- The transition state is a state where both bond breaking and bond formation are happening.
- The bonds that are forming and those that are breaking are weak bonds.
- The transition state for most enzymes is a fleeting state occurring in the femtosecond time scale

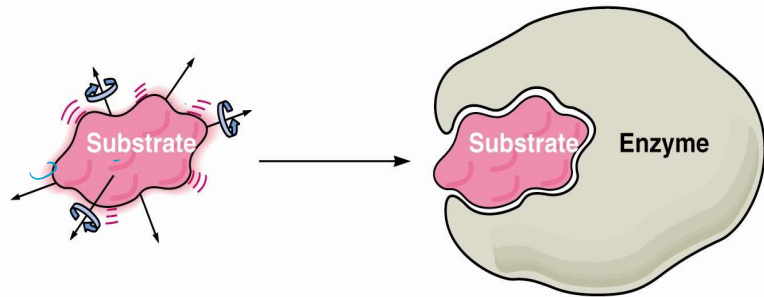




The enzyme has to **bind the transition state (T) more tightly** than the substrate or reactants to enhance the rate of the reaction.

## Destabilization of the ES complex

(a)



Substrate (and enzyme) are free to undergo translational motion. A disordered, high-entropy situation

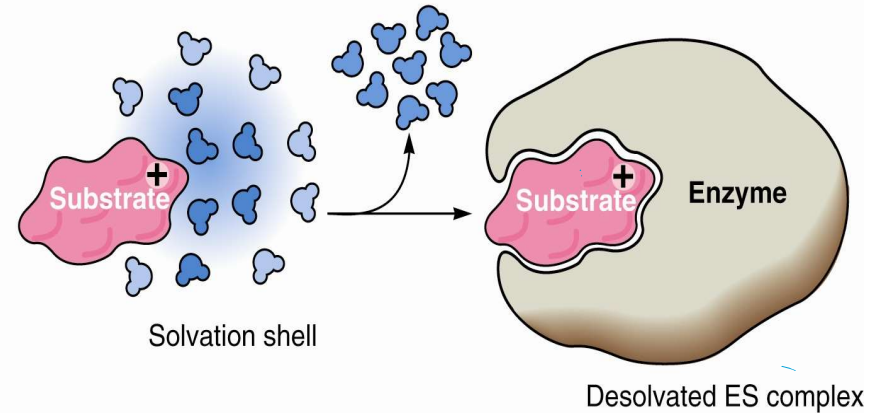
The highly ordered, low-entropy complex

The favorable binding energy of substrate and Enzyme is offset by destabilizing effects such as

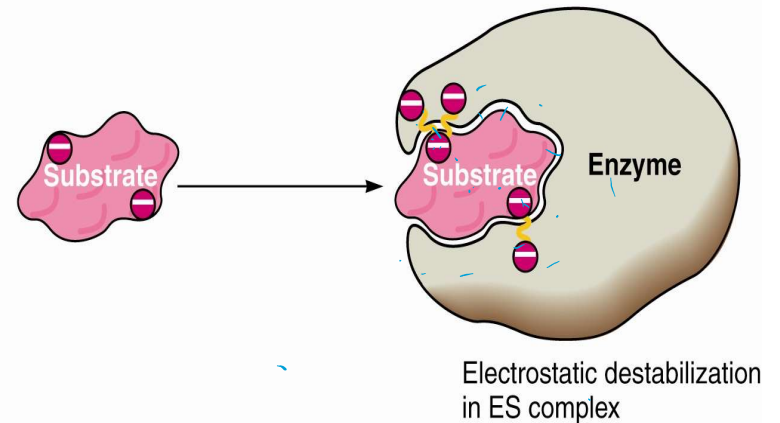
- 1) **Loss of entropy** upon binding
- 2) Destabilization of the Enzyme substrate complex by **desolvation** of charged groups on the substrate
- 3) **Electrostatic destabilization** of the ES complex by charged groups in the active site

Note: These factors actually are favorable in the case of the transition state

(b)



(c)



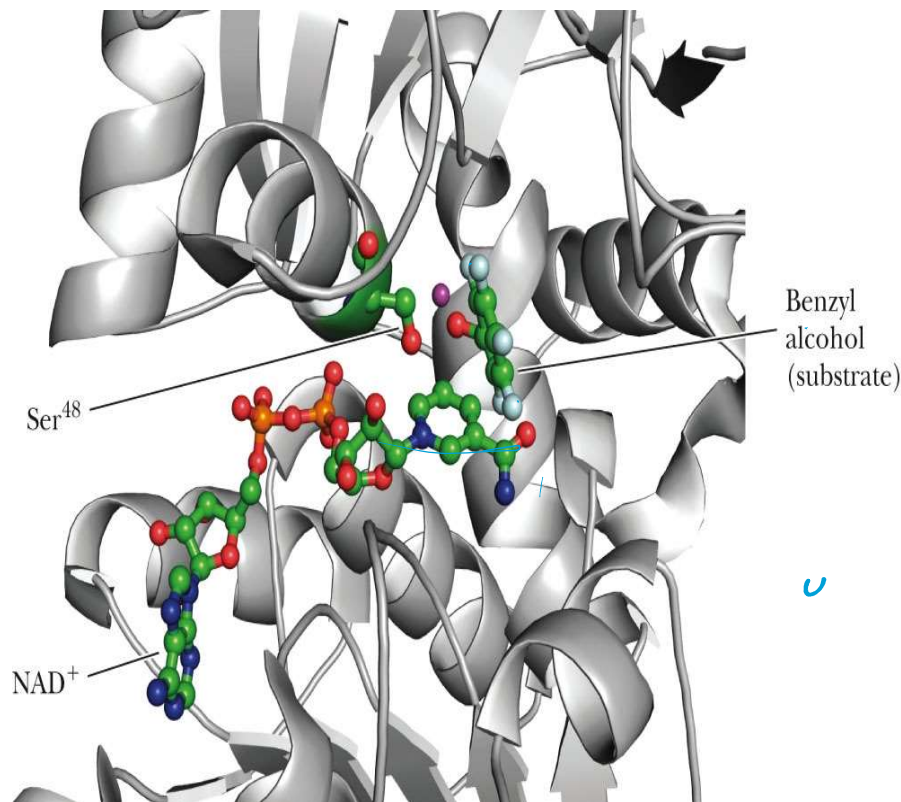
# Proximity and Orientation effects

- Enzyme bring substrates in close proximity with one another
- Enzymes orient the molecules in such a way that it tremendously increases the likelihood of a successful reaction occurring
- Charged groups in the enzymes active site can stabilize the transition state and this phenomenon is called electrostatic catalysis which is enhanced by the expulsion of water from the active site
- Enzyme active sites freeze the motions within the transition state to significantly enhance the rate of the reaction.

# Near attack conformation

- Enzyme active sites are referred to as “Preorganized” meaning the residues in the active site are placed in such a way that the reacting atoms are properly oriented and in vanderwaals contact and are in an angle that will resemble the bonds formed in the transition state.
- This is called the Near attack conformation.
- Thomas Bruice has proposed that near-attack conformations are precursors to transition states
- In the absence of an enzyme, potential reactant molecules adopt a NAC only about 0.0001% of the time
- On the other hand, NACs have been shown to form in enzyme active sites from 1% to 70% of the time

# The active site of liver alcohol dehydrogenase – a near-attack conformation.



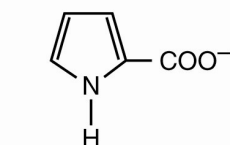
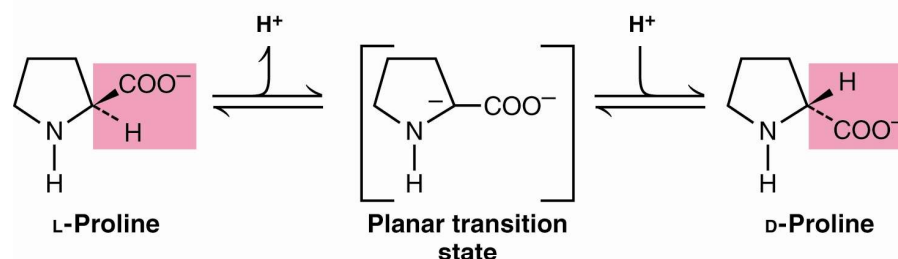
- The Benzyl alcohol is oriented in such a way that
- It is oriented in such a way that the Serine can abstract a proton from the Benzyl alcohol, and the NAD can accept the hydrogen from a pro-R position on the alcohol.
- There is a Zn<sup>2+</sup> ion in the active site which stabilizes the negative charges formed on the substrate during the reaction
- <https://pdb101.rcsb.org/motm/13>



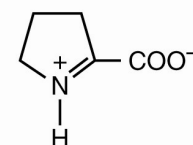
# Transition State Analogs

- The binding constant of the enzyme for the transition state may be  $10^{-20}$  to  $10^{-26} M$
- Transition state analogs (TSAs) are stable molecules that are chemically and structurally similar to the transition state

Proline racemase reaction



Pyrrole-2-carboxylate

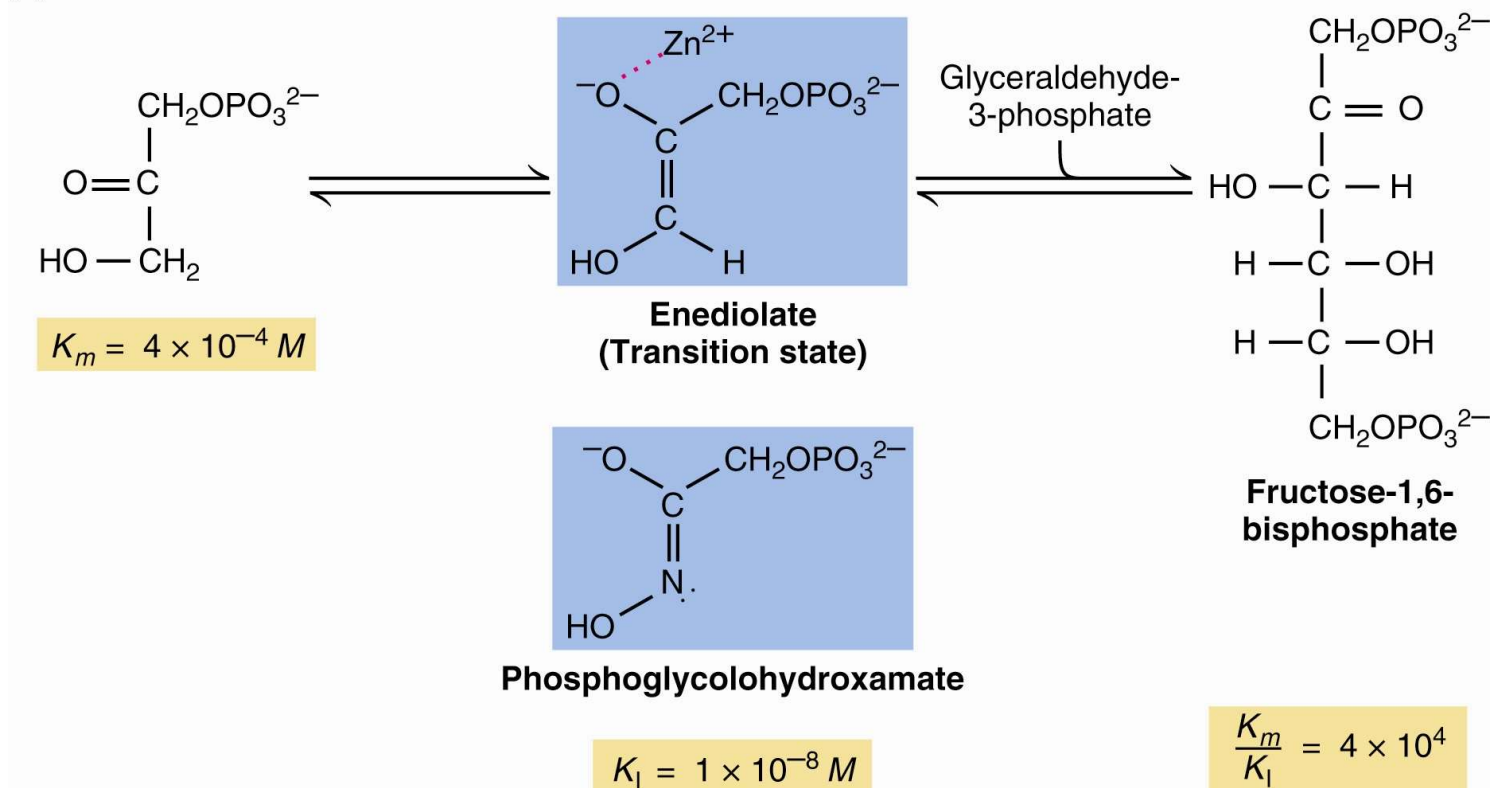


$\Delta$ -1-Pyrroline-2-carboxylate

Transition state analogs

# How Tightly Do Transition-State Analogs Bind to the Active Site?

(a) Yeast aldolase reaction

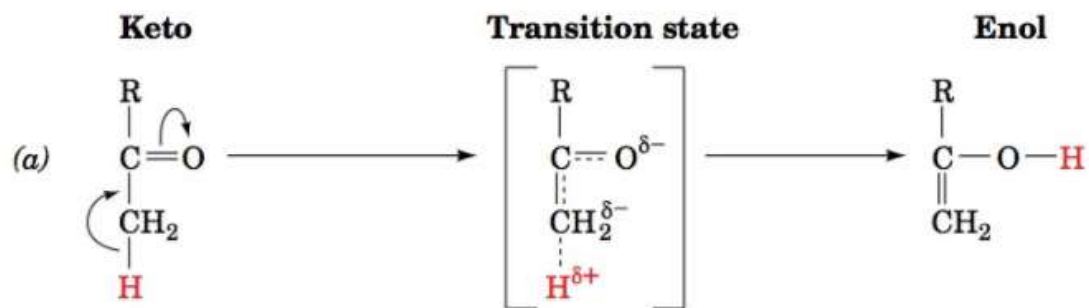


Phosphoglycolohydroxamate is an **analog of the enediolate transition state** of the yeast aldolase reaction.

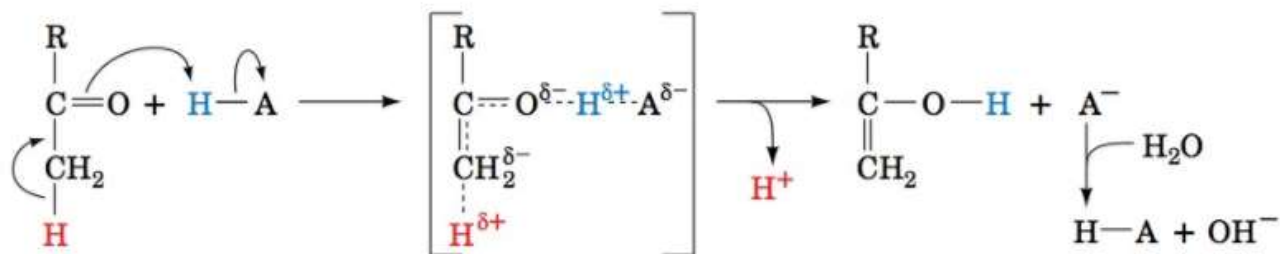
# Transition State Analogs as therapeutics

- Transition-state analogs often make ideal enzyme inhibitors
- **Statins** such as Lipitor are powerful cholesterol-lowering drugs, because they are transition-state analog inhibitors of HMG-CoA reductase, a key enzyme in the biosynthetic pathway for cholesterol.
- **Protease inhibitors** are AIDS drugs. Invirase (saquinavir) by Roche and similar “protease inhibitor” drugs are transition-state analogs for the HIV-1 protease.
- **Tamiflu is a neuraminidase inhibitor** and antiviral agent based on the transition state of the neuraminidase reaction

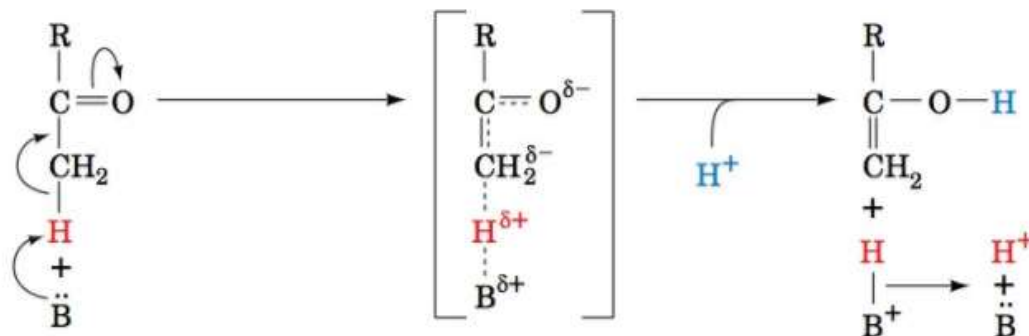
# Acid Base Catalysis

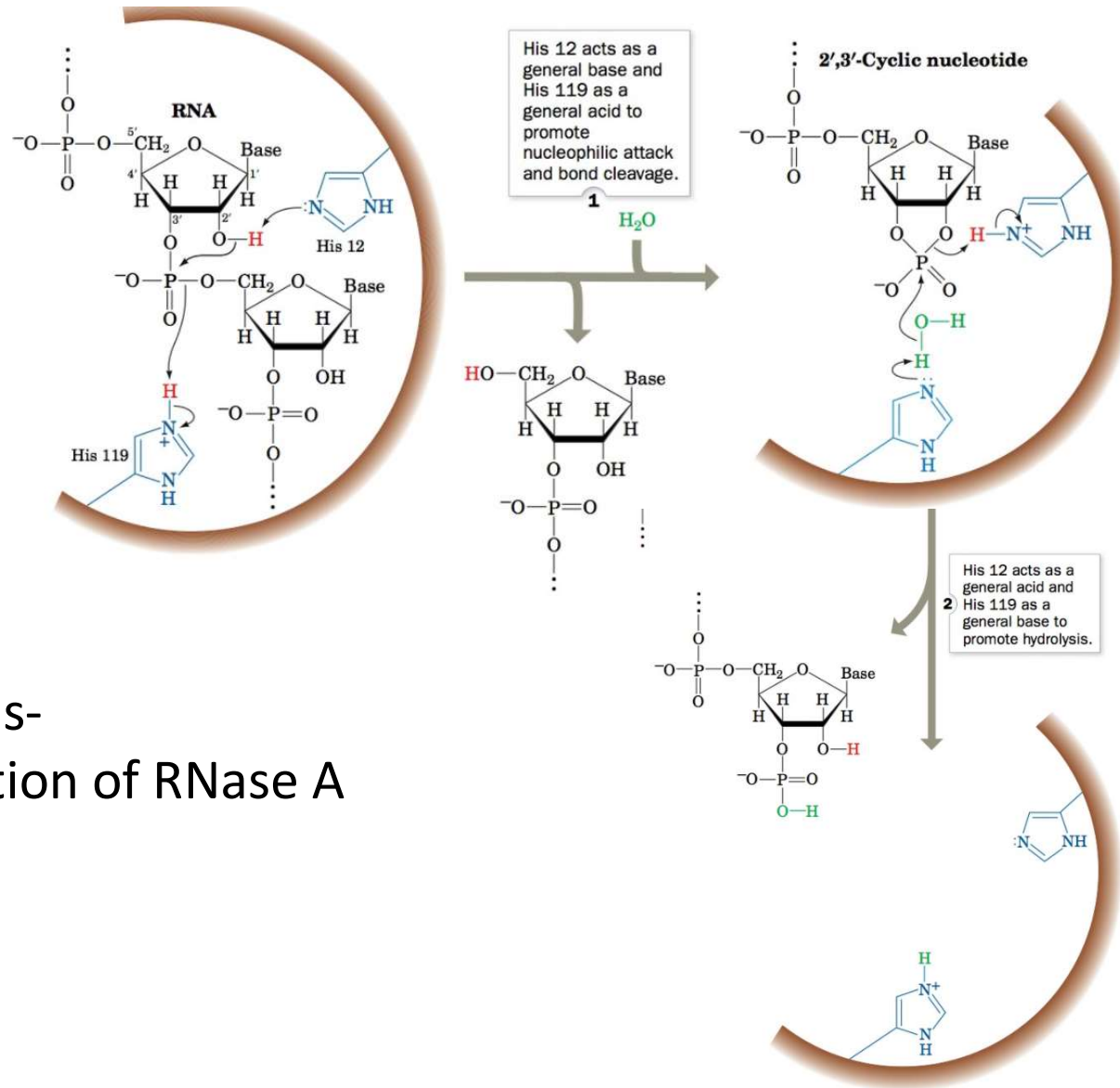


General Acid Catalyzed reaction



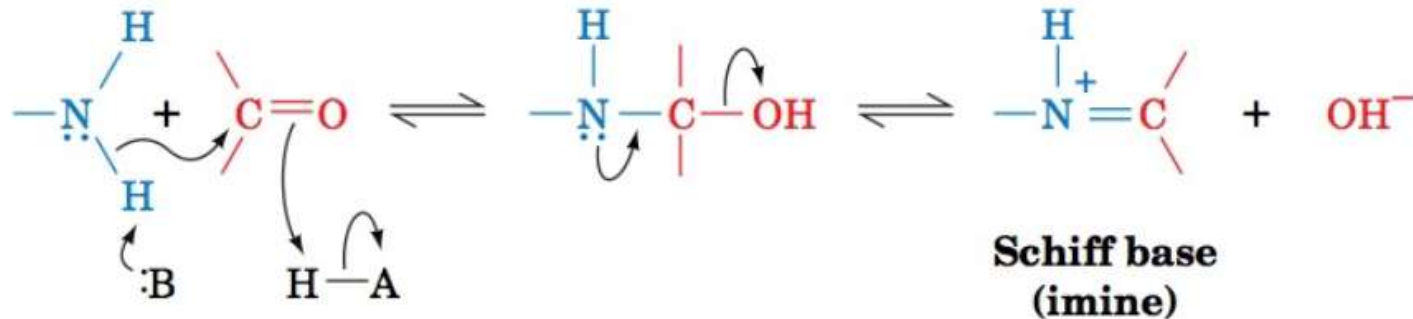
General Base Catalyzed reaction





## Acid base Catalysis- Mechanism of action of RNase A

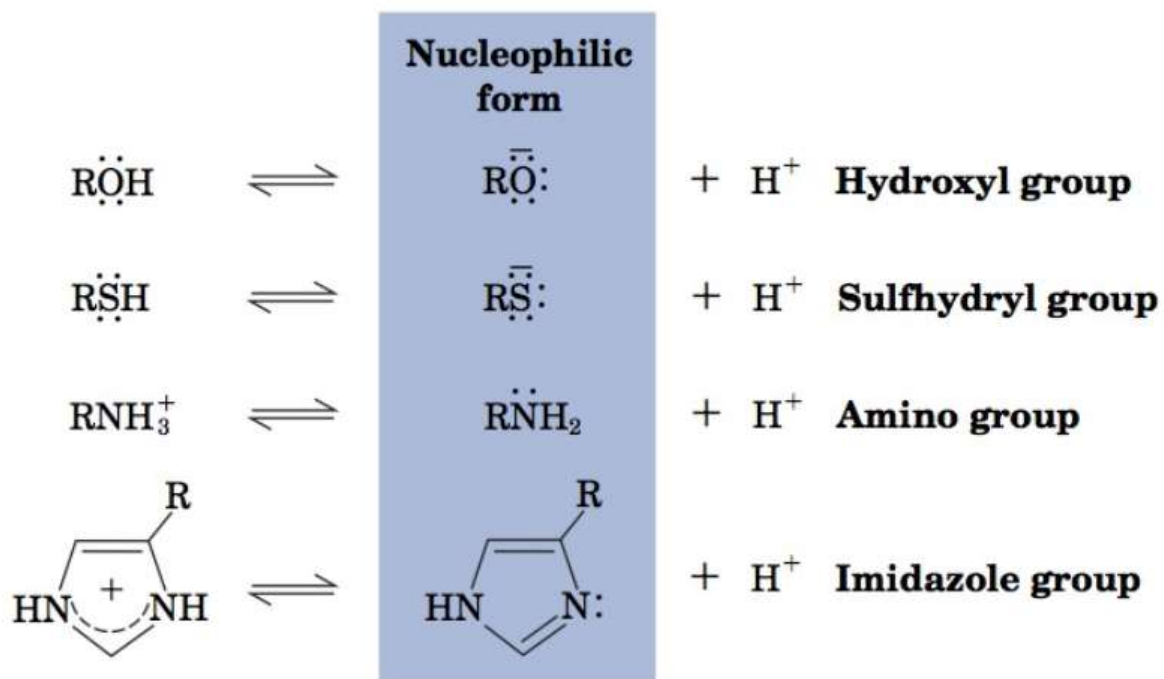
# Covalent Catalysis



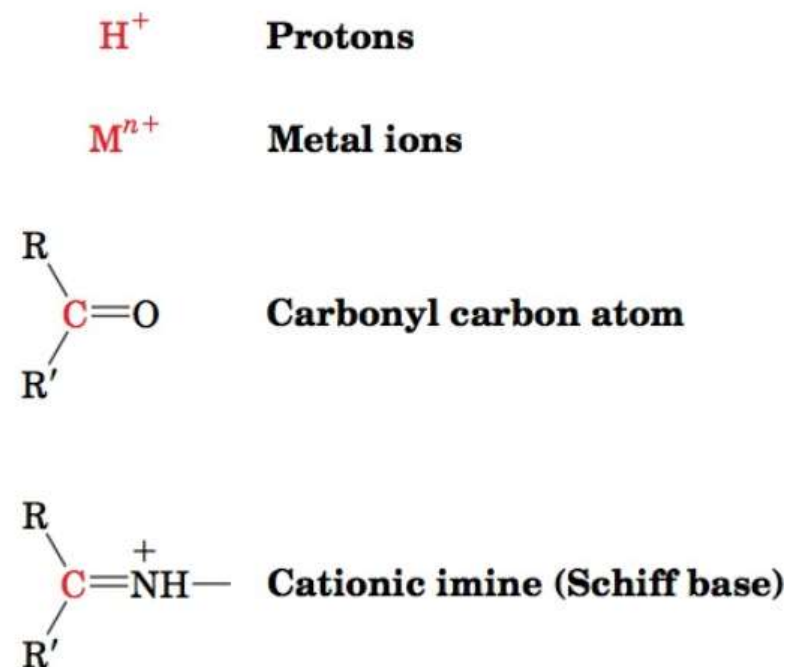
- Some enzymes derive much of their **rate acceleration from formation of covalent bonds between enzyme and substrate**
- The side chains of amino acids in proteins offer a variety of **nucleophilic** centers for catalysis
- These groups **readily attack electrophilic centers** of substrates, forming covalent enzyme-substrate complexes
- The covalent intermediate can be attacked in a second step by water or by a second substrate, forming the desired product

# Nucleophiles and Electrophiles

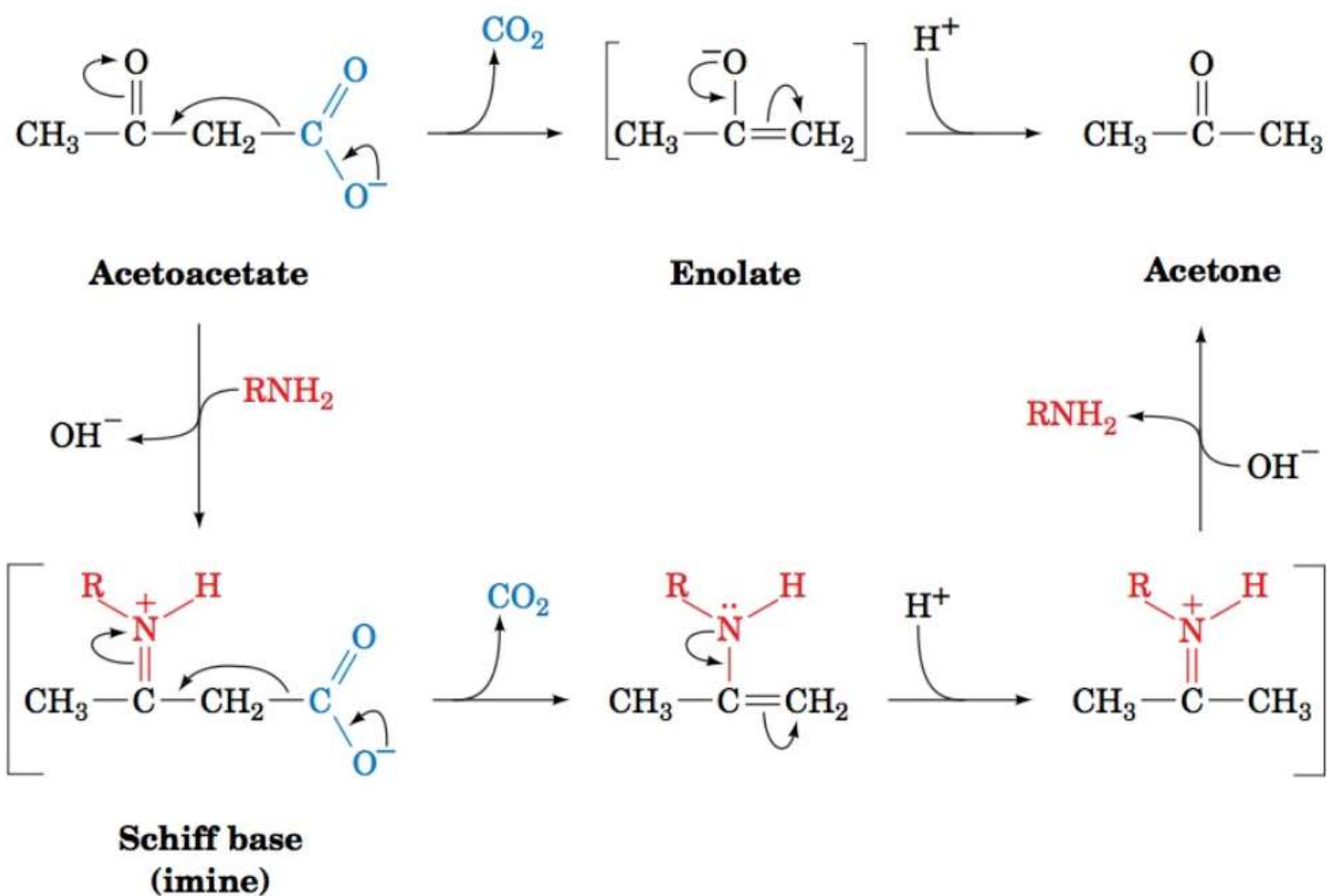
## (a) Nucleophiles



## (b) Electrophiles

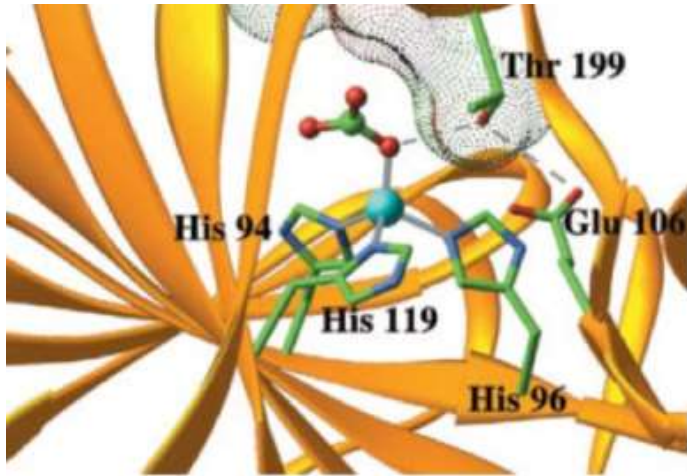


# Covalent Catalysis of Acetoactate to Acetone

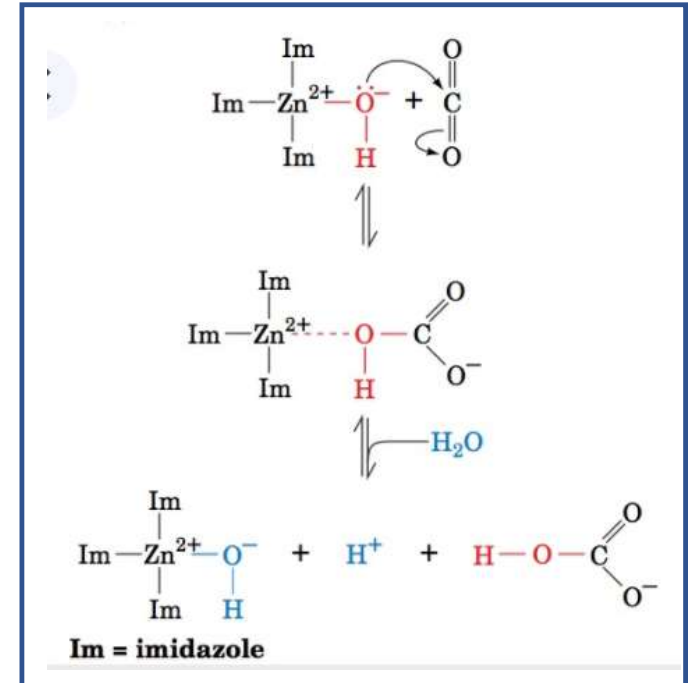




# Metal ion catalysis



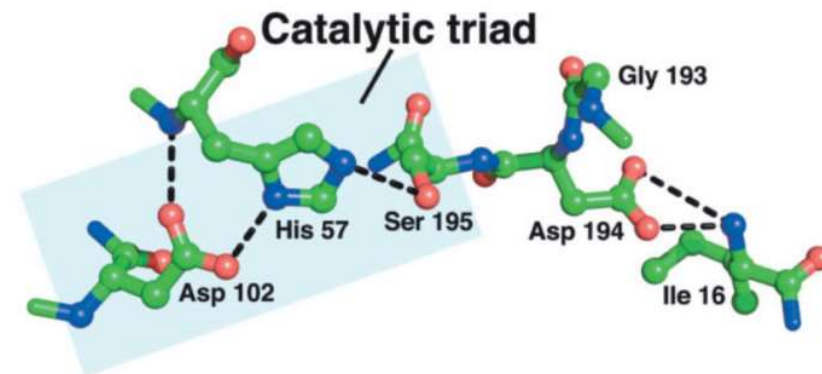
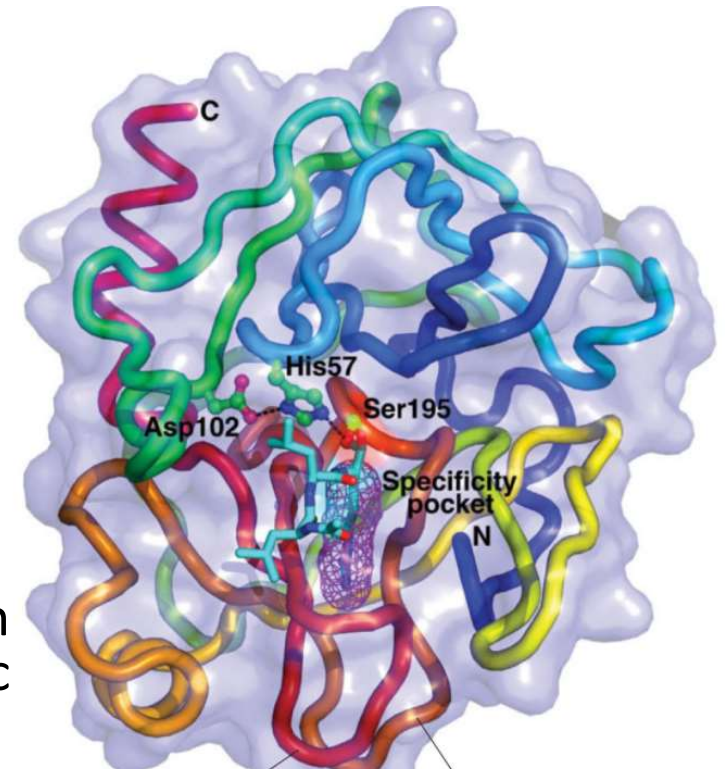
Carbonic anhydrase with Zn<sup>2+</sup>  
in its active site



- The Zn<sup>2+</sup> ion polarizes a water molecule (not shown) so it ionizes to form OH<sup>-</sup>, which nucleophilically attacks the substrate CO<sub>2</sub> to yield HCO<sub>3</sub><sup>-</sup>
- Metal ions will help **orient the substrate in the active site**
- Metal ions can participate in **oxidation reduction reactions** by changes in the ionization state of the metal
- Metal ions electrostatically **shield the negative charges** and hence stabilize transition state

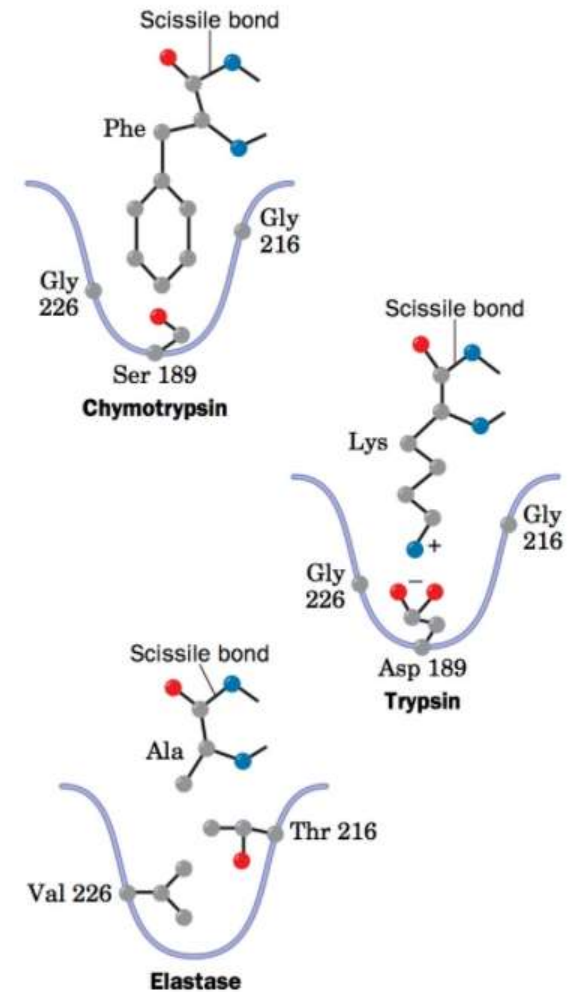
# Serine Proteases

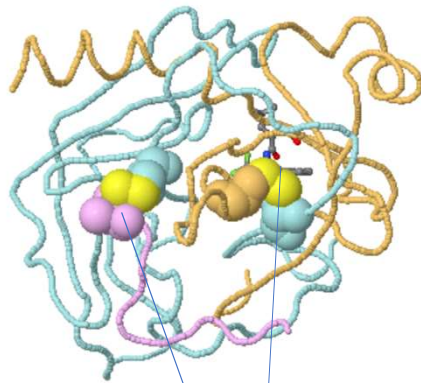
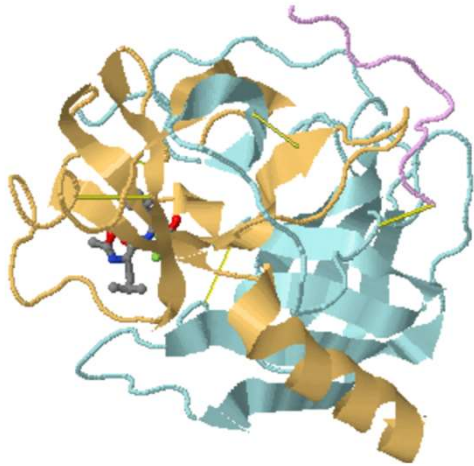
- Serine Proteases are a group of proteolytic enzymes which include digestive enzymes and the enzymes of the coagulation cascade
- E.g: Trypsin, Chymotrypsin and Elastase
- Shown here is the Active site of Bovine trypsin with 3 key residues which are referred to as the catalytic triad.
- The catalytic triad is found in all Serine proteases with serine always being one of the residues.
- Significant sequence identity and structural similarity



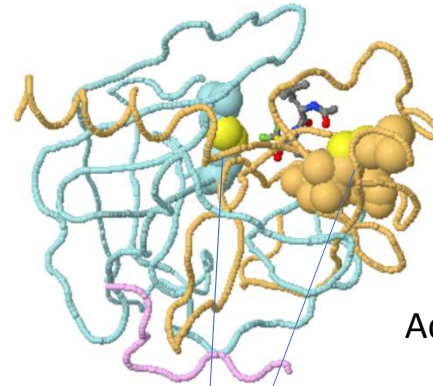
# Substrate specificity

- The specificity pocket shown in the fig on right defines the choice of substrate for each of these enzymes
- However mutating these residues of the specificity pocket alone is not sufficient to interchange substrate specificity

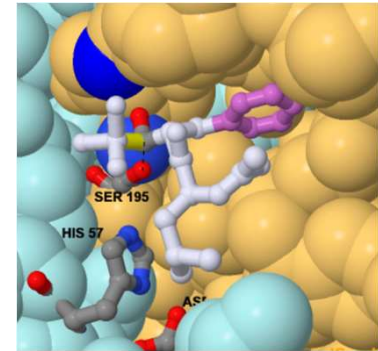




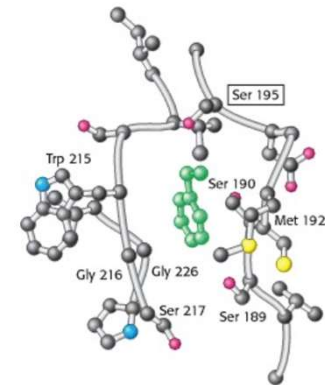
Inter-chain disulfide bonds



Intra-chain disulfide bonds

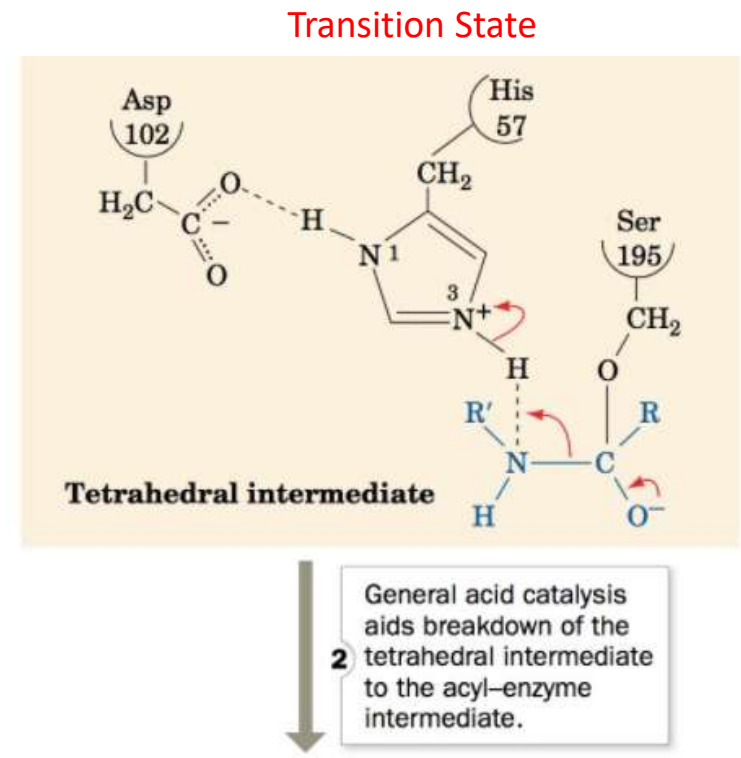
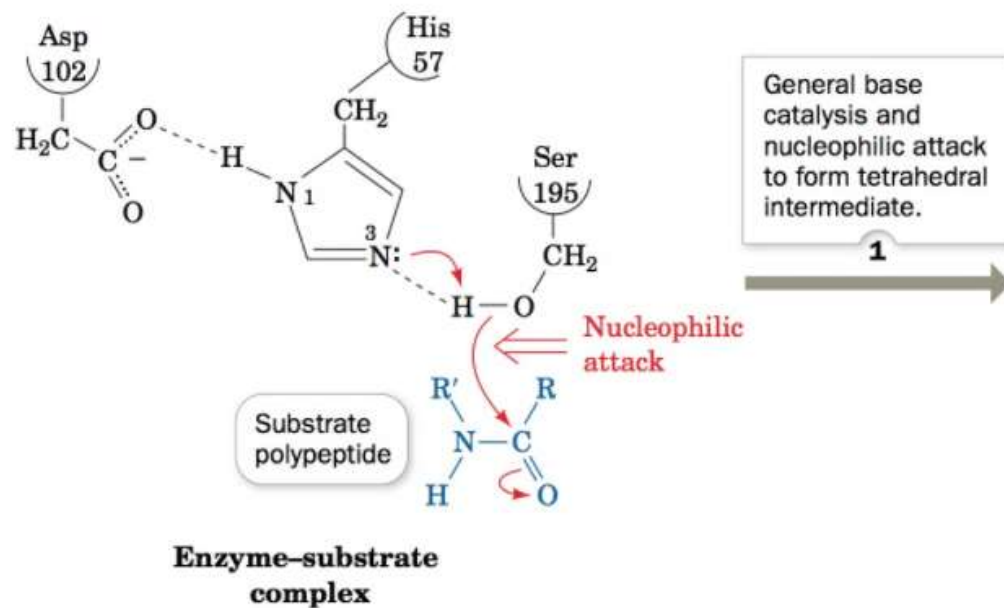


Active site and substrate binding pocket



Chymotrypsin is made up of three polypeptide chains indicated in blue, beige and pink. These three polypeptide chains are linked by 2 interchain disulfide bonds. There are also three intrachain disulfide bonds. Chymotrypsin structure has 2 beta barrel domains and one alpha helix has its primary secondary structural units.

# Mechanism of Serine Proteases (e.g Chymotrypsin)





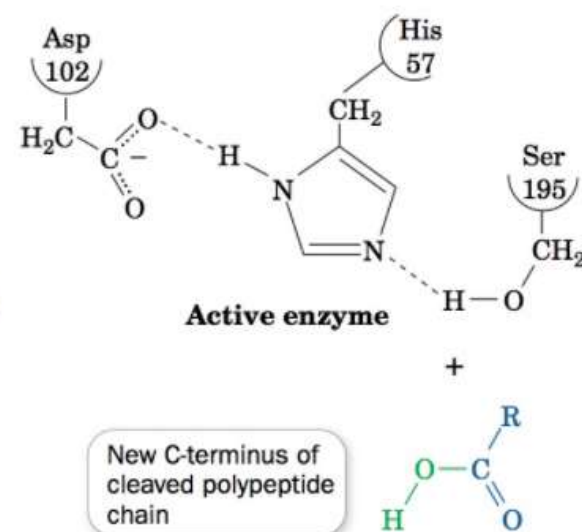
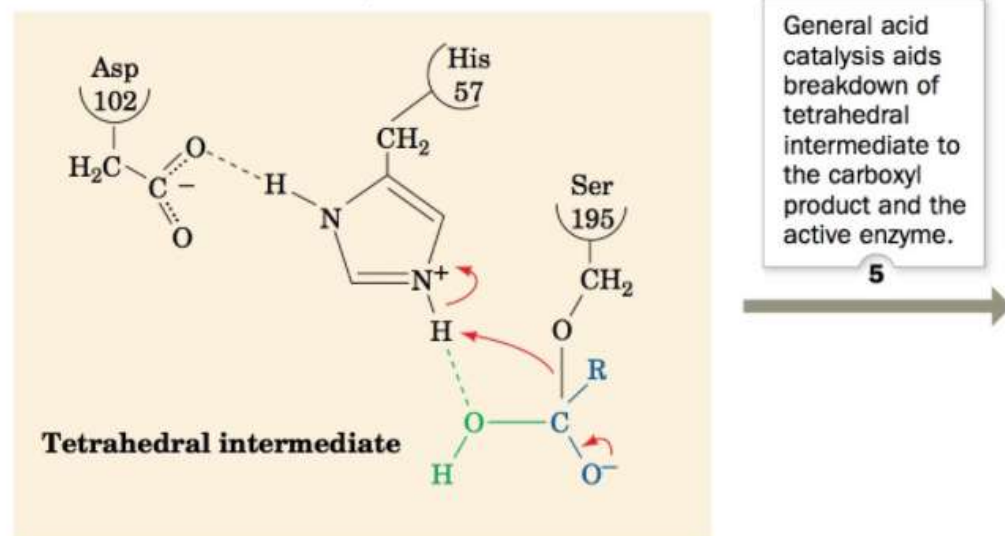
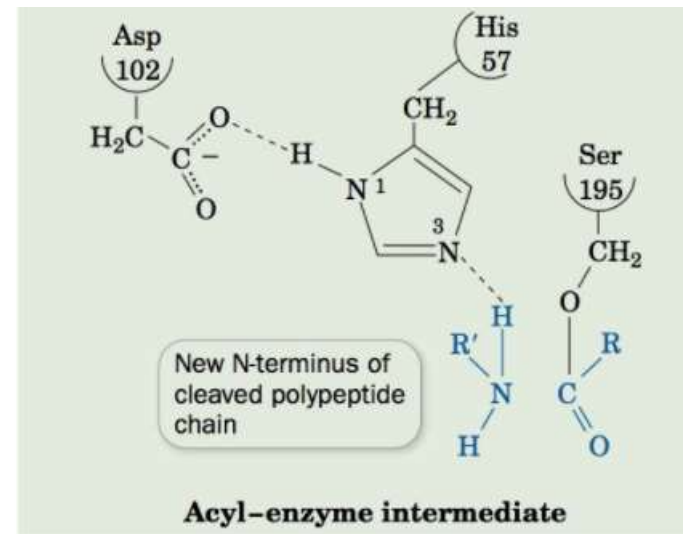
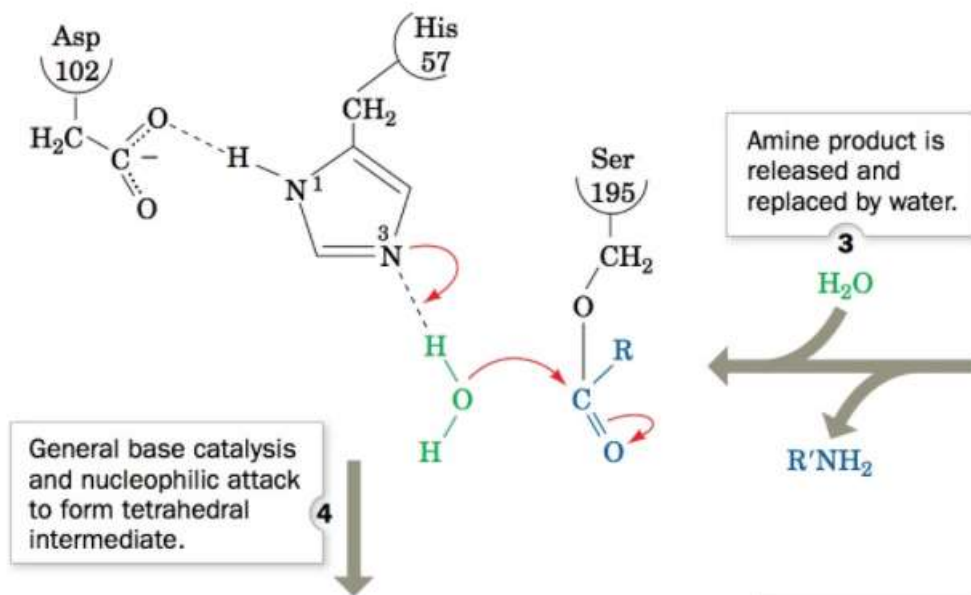


Figure 1 consists of two diagrams, (a) and (b), illustrating the catalytic mechanism of the catalytic triad in the active site of a protein. The active site is represented by a tan-colored cavity with residues Ser 195, His 57, and Asp 102 highlighted in blue. Gly 193 is shown in purple. The substrate is shown in red.

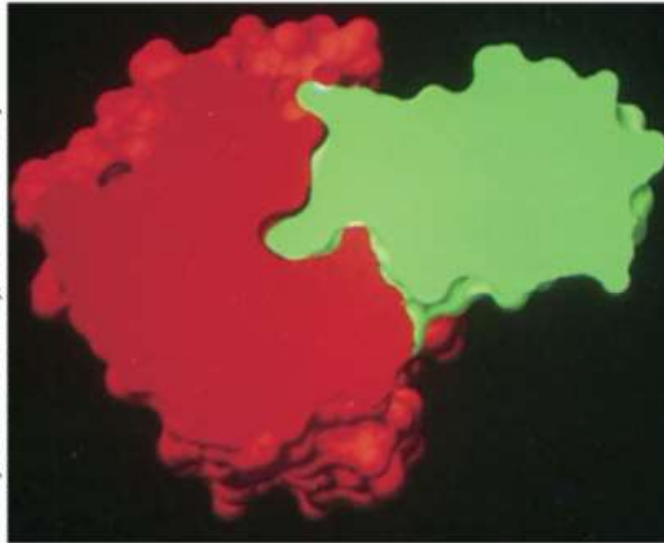
(a) Transition state of the reaction. Ser 195 is covalently bonded to the substrate. His 57 is hydrogen-bonded to the Ser 195 residue, and Asp 102 is hydrogen-bonded to His 57. The substrate is shown in red, and the catalytic triad residues are shown in blue. An "Oxyanion hole" is indicated by a green circle. The Ser 195 residue is shown with its side chain (NH) and backbone (NH, C=O). The His 57 residue is shown with its side chain (NH) and backbone (NH, C=O). The Asp 102 residue is shown with its side chain (NH) and backbone (NH, C=O). The substrate is shown with its side chain (NH) and backbone (NH, C=O). The Ser 195 residue is covalently bonded to the substrate's C $\alpha$  atom. The His 57 residue is hydrogen-bonded to the Ser 195 residue's side chain. The Asp 102 residue is hydrogen-bonded to the His 57 residue's side chain. The substrate's C $\beta$  atom is shown with a red circle. The Ser 195 residue's side chain is shown with a green circle. The His 57 residue's side chain is shown with a blue circle. The Asp 102 residue's side chain is shown with a red circle. The Gly 193 residue is shown with its side chain (NH) and backbone (NH, C=O). The Ser 195 residue's backbone is shown with a blue circle. The His 57 residue's backbone is shown with a blue circle. The Asp 102 residue's backbone is shown with a blue circle. The substrate's backbone is shown with a red circle. The Ser 195 residue's side chain is covalently bonded to the substrate's C $\alpha$  atom. The His 57 residue's side chain is hydrogen-bonded to the Ser 195 residue's side chain. The Asp 102 residue's side chain is hydrogen-bonded to the His 57 residue's side chain. The substrate's C $\beta$  atom is shown with a red circle. The Ser 195 residue's side chain is shown with a green circle. The His 57 residue's side chain is shown with a blue circle. The Asp 102 residue's side chain is shown with a red circle. The Gly 193 residue is shown with its side chain (NH) and backbone (NH, C=O). The Ser 195 residue's backbone is shown with a blue circle. The His 57 residue's backbone is shown with a blue circle. The Asp 102 residue's backbone is shown with a blue circle. The substrate's backbone is shown with a red circle.

(b) Ground state of the reaction. Ser 195 is covalently bonded to the substrate. His 57 is hydrogen-bonded to the Ser 195 residue, and Asp 102 is hydrogen-bonded to His 57. The substrate is shown in red, and the catalytic triad residues are shown in blue. The Ser 195 residue is shown with its side chain (NH) and backbone (NH, C=O). The His 57 residue is shown with its side chain (NH) and backbone (NH, C=O). The Asp 102 residue is shown with its side chain (NH) and backbone (NH, C=O). The substrate is shown with its side chain (NH) and backbone (NH, C=O). The Ser 195 residue is covalently bonded to the substrate's C $\alpha$  atom. The His 57 residue is hydrogen-bonded to the Ser 195 residue's side chain. The Asp 102 residue is hydrogen-bonded to the His 57 residue's side chain. The substrate's C $\beta$  atom is shown with a red circle. The Ser 195 residue's side chain is shown with a green circle. The His 57 residue's side chain is shown with a blue circle. The Asp 102 residue's side chain is shown with a red circle. The Gly 193 residue is shown with its side chain (NH) and backbone (NH, C=O). The Ser 195 residue's backbone is shown with a blue circle. The His 57 residue's backbone is shown with a blue circle. The Asp 102 residue's backbone is shown with a blue circle. The substrate's backbone is shown with a red circle.

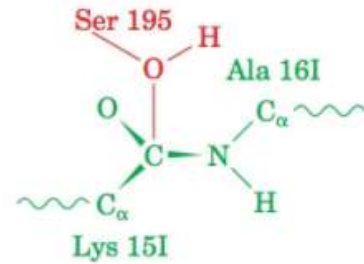
# Proteolytic Inhibitors are Transition state analogs

- Bovine trypsin inhibitor is an proteolytic enzyme inhibitor that naturally exists
- The inhibitor bound to the enzyme (fig) resembles the transition state.
- The Ser 195 forms a bond with the scissile peptide and the distortion in this case is such that the cleaved group is constrained the active site and cannot leave and water is unable to enter the active site

(a)  
Courtesy of Michael Connolly, New York University



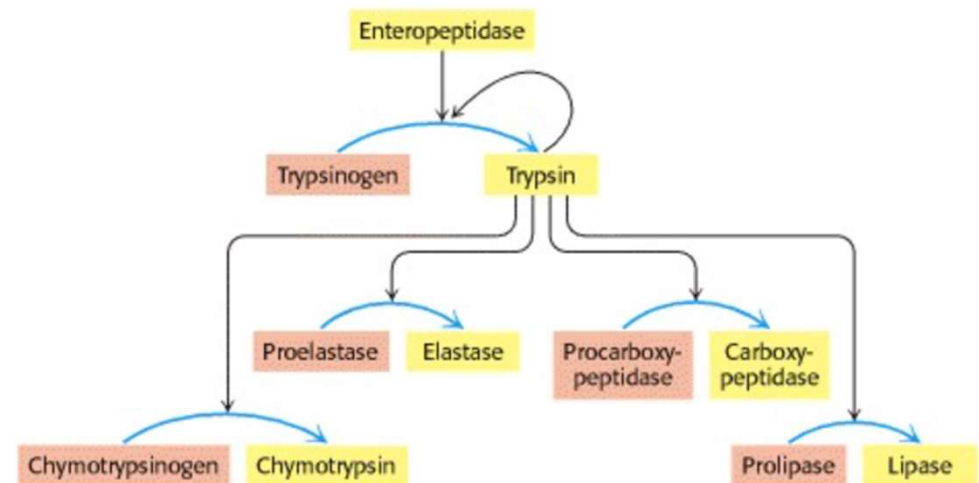
(b)





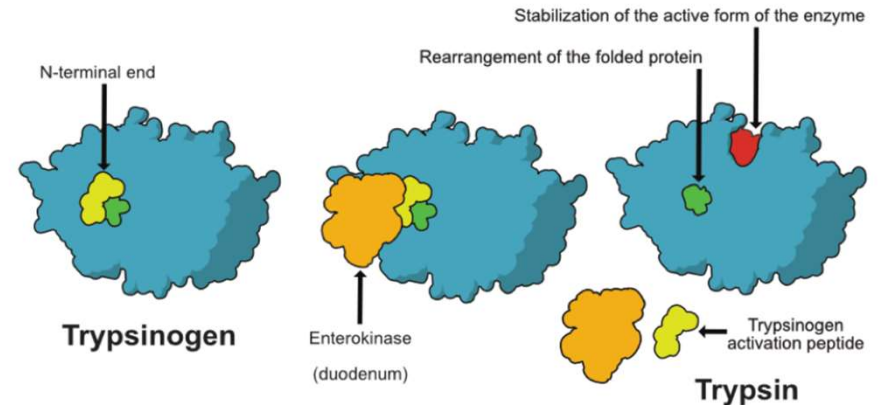
# Zymogens

- Serine Proteases exist in an **inactive precursor state called proenzymes or Zymogens**
- This is to prevent the enzymes from digesting tissues
- Eg is **Trypsinogen** is the precursor of Trypsin
- It is activated by the enzyme Enteropeptidase which is under hormonal control
- Trypsin then autocatalytically cleaves more Trypsinogen
- Trypsin also activates **Chymotrypsinogen, proelastase** and other proteolytic enzymes

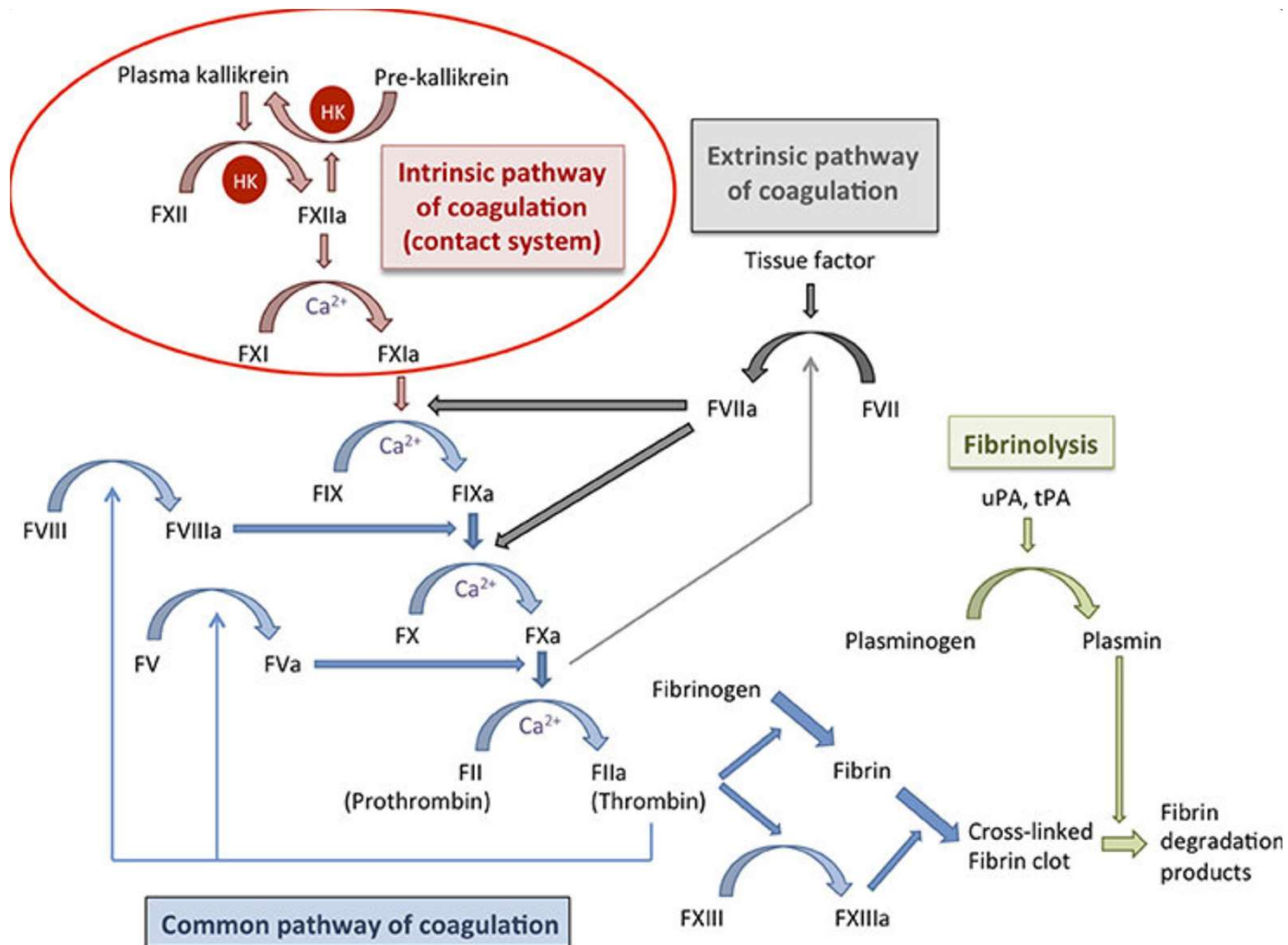


# Zymogen activation

- Zymogens have **distorted active sites** and upon losing the small peptide that distorts the active site, the enzyme changes conformation and the active site is restored
- The enzymes of the **coagulation cascade are Zymogens** and are activated in a cascade fashion



# Coagulation Cascade



# Activation of Chymotrypsinogen

Chymotrypsinogen is a single polypeptide consisting of 245 aminoacids. It is converted into a fully active form called  $\pi$  Chymotrypsin by cleaving between **Arg 15 and Ile 16**.

$\pi$  Chymotrypsin further cleaves out two dipeptides **Ser 14-Arg 15 and Thr 147-Asn 148**.

The newly formed amino-terminal group of Ile 16 turns inward and forms an ionic bond with Asp 194 in the interior of the chymotrypsin molecule.

The formation of the ionic bond triggers a series of conformational changes that results in the creation of the substrate specificity pocket. The changes also completes the formation of the oxyanion hole.

This form of Chymotrypsin is called  **$\alpha$ -Chymotrypsin**.

