# class 09

### Yinuo Song

#### **PDB** statistics

To read the csv file downloaded from PDB, we are going to use the command "read.csv"

```
pdb_stats <- read.csv('Data Export Summary.csv',row.names=1)</pre>
```

Q1 What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy? I need to sum all the elements of the x-ray column

```
as.numeric(gsub(",","",pdb_stats$X.ray))
```

[1] 154766 9083 8110 2664 163 11

```
n_xray <- sum(as.numeric(gsub(",","",pdb_stats$X.ray)))
n_em <- sum(as.numeric(gsub(",","",pdb_stats$EM)))
n_total <- sum(as.numeric(gsub(",","",pdb_stats$Total)))

(n_xray+n_em)/n_total</pre>
```

[1] 0.9299297

92.99 percentage of structures in the PDB are solved by X-Ray and Electron Microscopy?

Q2 What proportion of structures in the PDB are protein?

```
total_protein <- as.numeric(gsub(",","",pdb_stats[1,7]))</pre>
```

Protein

#### [1] 0.8681246

86.81 percentage of structures in the PDB are protein.

Q3 Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

There are 5 structures in the current PDB.

# 2. Visualizing the HIV-1 protease structure

figure 1

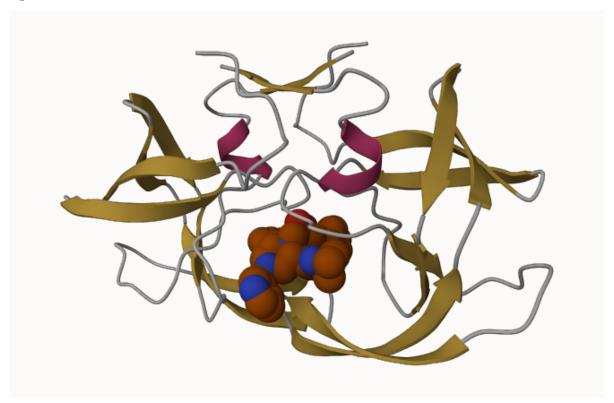
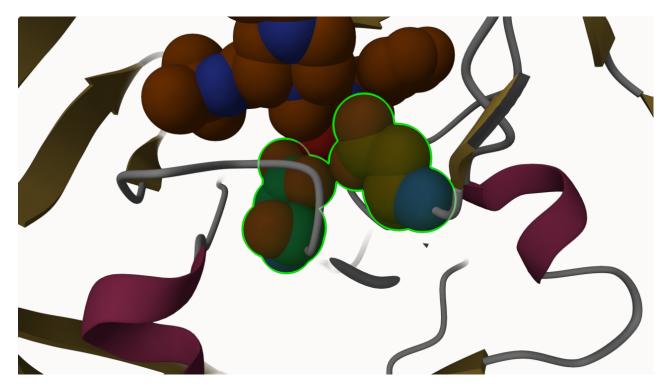
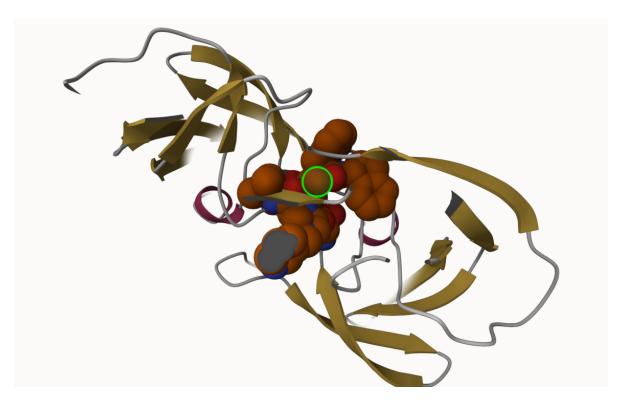


figure 2( Asp 25)



**Q6**: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend " $Ball\ \mathcal{E}\ Stick$ " for these side-chains). Add this figure to your Quarto document.

Figure 3



**Q4**: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

Because the structure of a water molecule compared to an amino acid structure is too small.

**Q5**: There is a critical "conserved" water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

The water molecule highlighted in figure 3 is HOH 308.

## 3. Introduction to Bio3D in R

```
library(bio3d)

pdb <- read.pdb("1hsg")

Note: Accessing on-line PDB file

attributes(pdb)</pre>
```

```
$names
[1] "atom" "xyz"
                       "seqres" "helix" "sheet" "calpha" "remark" "call"
$class
[1] "pdb" "sse"
  pdb
 Call: read.pdb(file = "1hsg")
   Total Models#: 1
     Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
     Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
     Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
     Non-protein/nucleic Atoms#: 172 (residues: 128)
     Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
   Protein sequence:
      PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
      QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
      \verb|ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP|
      VNIIGRNLLTQIGCTLNF
+ attr: atom, xyz, seqres, helix, sheet,
        calpha, remark, call
Q7: How many amino acid residues are there in this pdb object?
198
Q8: Name one of the two non-protein residues?
nucleic Atoms
Q9: How many protein chains are in this structure?
2
  head(pdb$atom)
```

```
z o
 type eleno elety alt resid chain resno insert
                                                   Х
                                                          У
1 ATOM
          1
                N < NA >
                        PRO
                                Α
                                     1
                                         <NA> 29.361 39.686 5.862 1 38.10
2 ATOM
          2
               CA <NA>
                        PRO
                                      1
                                         <NA> 30.307 38.663 5.319 1 40.62
                                Α
3 ATOM
          3
              C <NA>
                        PRO
                                    1 <NA> 29.760 38.071 4.022 1 42.64
                                Α
4 ATOM
          4
                O <NA>
                        PRO
                               Α
                                    1 <NA> 28.600 38.302 3.676 1 43.40
                                      1 <NA> 30.508 37.541 6.342 1 37.87
5 ATOM
          5
                        PRO
               CB <NA>
                                Α
6 ATOM
          6
              CG <NA>
                        PRO
                             A 1 <NA> 29.296 37.591 7.162 1 38.40
 segid elesy charge
1 <NA>
          N
              <NA>
2 <NA>
           С
              <NA>
3 <NA>
           C <NA>
4 <NA>
           O <NA>
           C <NA>
5 <NA>
           C
               <NA>
6 <NA>
```

### Predicting functional motions of a single structure by NMA

```
adk <- read.pdb('6s36')

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE

adk

Call: read.pdb(file = "6s36")

Total Models#: 1
   Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
   Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)
   Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

Protein sequence:
   MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLVT
   DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI</pre>
```

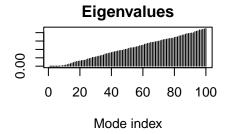
### VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG YYSKEAEAGNTKYAKVDGTKPVAEVRADLEKILG

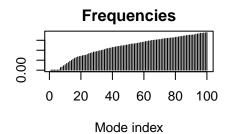
+ attr: atom, xyz, seqres, helix, sheet, calpha, remark, call

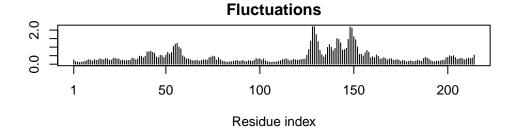
m <- nma(adk)

Building Hessian... Done in 0.029 seconds. Diagonalizing Hessian... Done in 0.347 seconds.

plot(m)







mktrj(m,file="adk\_m7.pdb")

View the file in Mol Viewer

