

Aucubin, a naturally occurring iridoid glycoside inhibits TNF- α -induced inflammatory responses through suppression of NF- κ B activation in 3T3-L1 adipocytes

Kyoung Sik Park*

Department of Genetic Engineering, College of Natural Science and Engineering, Cheongju University, Chungbuk, Republic of Korea

ARTICLE INFO

Article history:

Received 10 December 2012

Received in revised form 20 March 2013

Accepted 1 April 2013

Available online 20 April 2013

Keywords:

Iridoid glycoside

Herbal medicine

Adipocyte

Inflammatory response

ABSTRACT

Obesity is closely associated with a state of chronic, low-grade inflammation characterized by abnormal cytokine production and activation of inflammatory signaling pathways in adipose tissue. Tumor necrosis factor (TNF)- α is chronically elevated in adipose tissues of obese rodents and humans. Increased levels of TNF- α are implicated in the induction of atherogenic adipokines, such as plasminogen activator inhibitor (PAI)-1, adipose-tissue-derived monocyte chemoattractant protein (MCP)-1, and interleukin (IL)-6. Aucubin, an iridoid glycoside existing in medicinal plants, has been reported to show an anti-inflammatory activity by suppression of TNF- α production in murine macrophages. The present study is aimed to investigate the effects of aucubin on TNF- α -induced atherogenic changes of the adipokines in differentiated 3T3-L1 cells. Aucubin significantly inhibited TNF- α -induced secretion and mRNA synthesis of the atherogenic adipokines including PAI-1, MCP-1, and IL-6. Further investigation of the molecular mechanism revealed that pretreatment with aucubin suppressed extracellular signal-regulated kinase (ERK) activation, inhibitory kappa B α (I κ B α) degradation, and subsequent nuclear factor kappa B (NF- κ B) activation. These findings suggest that aucubin may improve obesity-induced atherosclerosis by attenuating TNF- α -induced inflammatory responses.

© 2013 Elsevier Ltd. All rights reserved.

1. Introduction

Obesity, which is characterized by excessive accumulation of abdominal fat, is casually associated with the premature development of atherosclerosis, increased risk of stroke, and development of congestive heart failure [1]. Recent studies have indicated that the adipocyte secretes a variety of adipokines involved in energy metabolism, inflammation, and cardiovascular functions [2]. The cellular mechanisms linking obesity and atherosclerosis are complex and have not been fully elucidated. However, increasing evidences suggest that the changes of adipokines including PAI-1, MCP-1, and IL-6 due to excess adipose tissue may be a cause of atherosclerosis [3].

Plasminogen activator inhibitor (PAI)-1 is the primary inhibitor of plasminogen activation. Plasma levels of PAI-1 are markedly elevated in obese individuals as well as in patients with insulin resistance, type 2 diabetes, and cardiovascular disease (CVD) [4,5]. PAI-1 is thought to be the link between obesity and increased risk for CVDs [6]. Although several tissues are known to produce PAI-1,

adipose tissue appears to be the major contributor to elevated PAI-1 levels observed in cases of obesity [7,8]. Monocyte chemoattractant protein (MCP)-1 chemotactically recruits monocytes to sites of inflammation. Although this protein is traditionally thought to be expressed mainly endothelial cells and macrophages, it has been shown to be primarily expressed by adipose tissues [9,10]. Adipocyte-derived MCP-1 induces macrophage infiltration into adipose tissues and thus secretes inflammatory cytokines including tumor necrosis factor (TNF)- α , which in turn leads to the dysfunction of adipocytes [11]. In addition, MCP-1 inhibits insulin-dependent glucose uptake and the expression of adipogenic genes [10].

Aucubin is a natural constituent with a monoterpene cyclic ring system found in a wide range of some insects and higher plants such as *Aucuba japonica* and *Plantago asiatica* [12,13]. In recent years, a variety of bio-activities of aucubin have been reported; liver-protective activities against hepato-toxicants, stimulation of bile acid excretion, anti-microbial activities, anti-tumor activities, antidotal activities for noxious amanita mushroom poisoning, anti-viral activities against hepatitis B virus, and anti-inflammatory activities [14–22]. Interestingly, a number of plants containing aucubin have been used long as medicinal herbs for anti-inflammatics and anti-rheumatics in Oriental and Occidental hemispheres. As for the molecular mechanism for anti-inflammatory

* Address: Department of Genetic Engineering, College of Natural Science and Engineering, Cheongju University, 586 Daesung-ro, Sangdang-gu, Cheongju-si, Chungbuk 360-764, Republic of Korea. Tel.: +82 43 229 8566; fax: +82 43 229 8559.

E-mail address: parkks@cju.ac.kr

activities of aucubin, it was reported that aucubin inhibits the production of TNF- α resulting from the both I κ B α degradation and the nuclear translocation of NF- κ B in RAW 264.7 cells [21]. However, the inhibitory effects of aucubin on TNF- α -induced inflammatory responses in adipocytes have not been reported previously.

The present study was designed to determine whether it attenuates TNF- α -induced secretion and mRNA production of the atherogenic adipokines including PAI-1, MCP-1, and IL-6 in differentiated 3T3-L1 adipocytes. In addition, the possible mechanisms for inhibitory effects of aucubin on obesity-related inflammatory responses were examined.

2. Materials and methods

2.1. Reagents

Aucubin with 99.5% purity was purchased from Wako Pure Chemical Industries Ltd., (Osaka, Japan). Recombinant murine tumor necrosis factor (TNF)- α was from R&D Systems (Minneapolis, MN, USA). Insulin, dexamethasone, and 3-isobutyl-1-methylxanthine were from Sigma (St. Louis, MO, USA). All tissue culture materials were from Gibco-BRL (Rockville, MD, USA). Primary anti-mouse antibodies for extracellular signal-regulated kinases (ERK), phospho-ERK, I κ B α , phospho-I κ B α , and β -actin were obtained from Cell Signaling Technology (Beverly, MA, USA). Poly (ADP-ribose) polymerase (PARP) and p65 monoclonal antibodies were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA).

2.2. 3T3-L1 cell culture and treatment

3T3-L1 preadipocytes (American Type Culture Collection, Rockville, MD) were cultured in Dulbecco's Modified Eagle Medium (DMEM) with 10% (v/v) fetal bovine serum (FBS), 1% (v/v) penicillin-streptomycin (10,000 U/mL penicillin and 10,000 cg/mL streptomycin in 0.85% saline), and 1% (v/v) 100 μ M pyruvate at 37 °C in 95% air 10% CO₂. Differentiation of 2-day postconfluent preadipocytes (designated as day 0) was initiated with 0.5 mM 3-isobutyl-1-methylxanthine, 1 μ M dexamethasone, and 1 μ g/mL insulin in DMEM supplemented with 10% fetal bovine serum. After 48 h (day 2), the culture medium was replaced with DMEM supplemented with 10% fetal bovine serum and 1 μ g/mL insulin, and the cells were then fed every other day with DMEM containing 10% fetal bovine serum. Aucubin was reconstituted, filter sterilized, and stored at -20 °C. For each experiment, cells received aucubin premixed with culture medium. TNF- α treatment was carried out after a 6-h pretreatment with aucubin. Aucubin was dissolved in dimethyl sulfoxide (DMSO) before it was added to culture medium. The final concentration of DMSO in culture medium was 0.5% (v/v).

2.3. Measurement of adipokines by ELISA

The conditioned culture medium from 3T3-L1 adipocytes was collected from each sample. The concentrations of MCP-1, PAI-1, and IL-6 were assayed using a mouse MCP-1 ELISA kit (R&D Systems, Minneapolis, MN, USA), a mouse PAI-1 ELISA kit (Molecular Innovations Inc., Southfield, MI, USA), and a mouse IL-6 ELISA kit (Biosource Inc., Camarillo, CA, USA), respectively. All assays were performed according to the manufacturer's instructions.

2.4. Evaluation of gene expression levels by quantitative real-time RT-PCR

For real-time RT-PCR, cDNA was prepared from 1 μ g total RNA in a final reaction volume of 15 μ L. Real-time RT-PCR was performed using 2 μ L cDNA diluted with each of the gene-specific pri-

mer sets (Bioneer Inc., Korea). Each primer set was used at a concentration of 150 nM in a final reaction volume of 20 μ L and reactions were performed on the Lightcycler 480 SYBR Green I Master (Roche Applied Science, Germany). The sequences for primers are: MCP-1 (5'-GCCCACTACCTGCTGCTACT-3' and 5'-CCTGCTGCTGGTGATCCTCTTGT-3'), PAI-1 (5'-AGGATCGAGGTAAACGAGAGC-3' and 5'-GCGGGCTGAGATGACAAA-3'), and IL-6 (5'-GCTACCAAAGTGGATATAATCAGGA-3' and 5'-CCAGGTAGCTATGGTACTCCAGAA-3'). Levels of mRNA, expressed as relative mRNA levels compared with control, were calculated after normalization to β -actin.

2.5. Nuclear extraction

Nuclear fractions of cells were prepared as previously described [23]. Briefly, cells were scraped and suspended in 400 μ L of cold buffer A (HEPES 10 mM; pH 7.9; KCl 10 mM; EDTA 0.1 mM; EGTA 0.1 mM; dithiothreitol 1 mM; phenylmethylsulfonyl fluoride 0.5 mM; pepstatin A 1 μ g/mL; leupeptin 10 μ g/mL; aprotinin

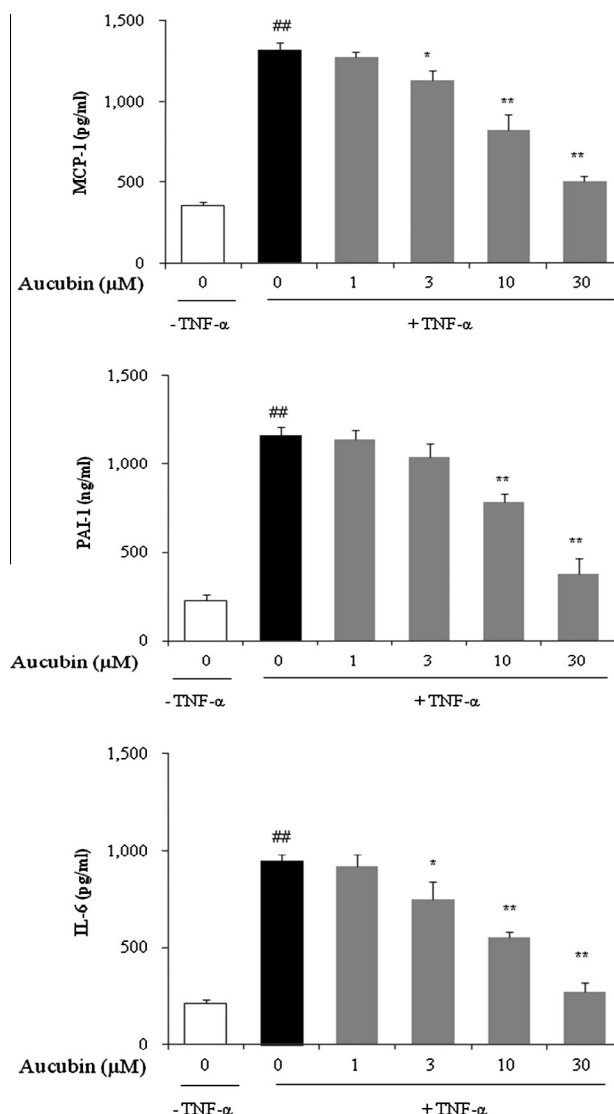


Fig. 1. Effects of aucubin on TNF- α -stimulated adipokine production in 3T3-L1 adipocytes. 3T3-L1 adipocytes were exposed or not to increasing concentrations of aucubin (1, 3, 10, and 30 μ M) for 6 h and then stimulated for 24 h by the addition of 10 ng/mL TNF- α . The secreted antigens of MCP-1, PAI-1, and IL-6 in conditioned medium were measured as described in Section 2. Results represent the mean \pm SD from three independent experiments. ## p < 0.01 versus untreated control; * p < 0.05, ** p < 0.01 versus TNF- α -stimulated control.

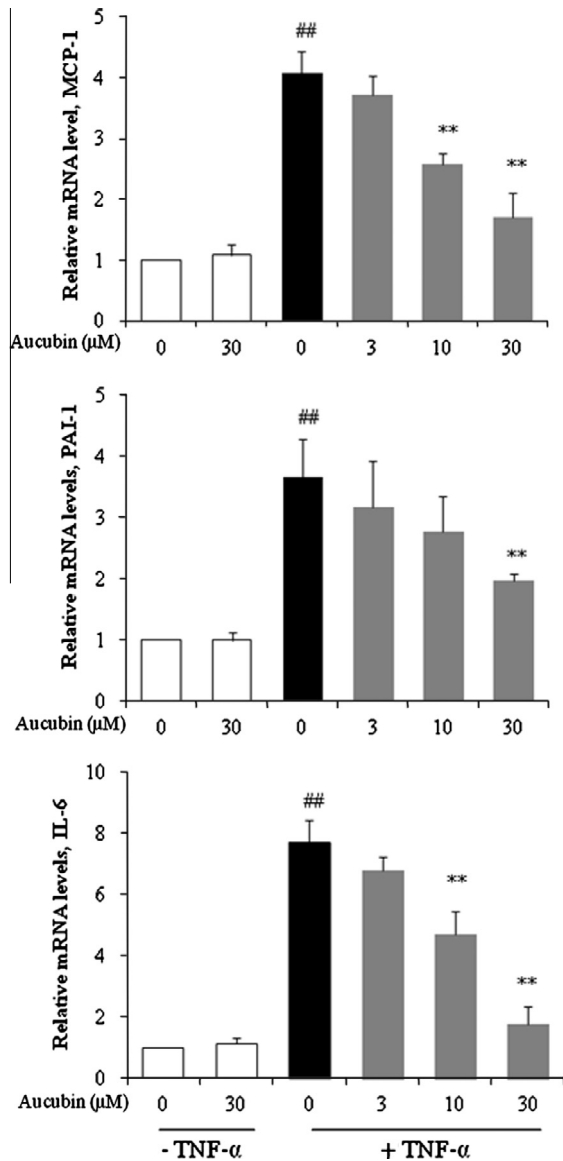


Fig. 2. Effects of aucubin on TNF- α -induced adipokine mRNA synthesis in 3T3-L1 adipocytes. After exposure to various concentrations of aucubin (1, 3, 10, and 30 μ M) for 6 h, 3T3-L1 adipocytes were stimulated with 10 ng/mL TNF- α for 24 h. Then cells were harvested, and total RNA was extracted for measuring the mRNA levels of MCP-1, PAI-1, and IL-6 by quantitative RT-PCR. Values are normalized to β -actin RNA expression levels and expressed relative to untreated control cells. Results represent the mean \pm SD from three independent experiments. ## p < 0.01 versus untreated control; ** p < 0.01 versus TNF- α -stimulated control.

10 μ g/mL) in ice bath under the presence of 25 μ L 1% IGEPALCA-630 (Sigma). Then, the above samples were vortexed and centrifuged for 1 min at 10,000g. The precipitated cell pellet were used as the nuclear fraction for immunoblot assay. And the precipitated cell pellet was re-suspended with shaking in 100 μ L of buffer B (HEPES 20 mM; pH 7.9; NaCl 400 mM; EDTA 1 mM; EGTA 1 mM; dithiothreitol 1 mM; phenylmethylsulfonyl fluoride 0.5 mM; pepstatin A 1 μ g/mL; leupeptin 10 μ g/mL; aprotinin 10 μ g/mL), and centrifuged for 15 min at 10,000g. The supernatant aliquots (70 μ L) were used for assay.

2.6. Western blot analysis

Cell lysates were collected, and then protein concentrations were determined with bicinchoninic acid (BCA) protein assay re-

agents (Thermo Fisher Scientific, Rockford, IL, USA). Equal amounts of cell extracts were separated by electrophoresis using a 10% SDS-polyacrylamide gel and transferred to nitrocellulose membranes (Whatman Protran, Whatman GmbH, Dassel, Germany). After being blocked at room temperature in 5% non-fat dry milk with Tris-buffered saline Tween-20 (TBST) buffer for 2 h, the blots were probed with the primary antibody against each target protein at a concentration of 1:500–1:1000 in 5% non-fat dry milk in TBST overnight at 4 $^{\circ}$ C. Bound antibodies were detected by horseradish peroxidase-conjugated secondary antibodies and signals were detected and quantified using a chemiluminescent detection system, LAS-3000 (Fujifilm, Japan).

2.7. Statistics

Data expressed as mean \pm SD were obtained from three separate experiments. Statistical analysis of the data was performed by one-way analysis of variance followed by Dunnett's test. P < 0.05 was considered to have significant difference.

3. Results

3.1. Effects of aucubin on the TNF- α -induced secretion of adipokines in 3T3-L1 adipocytes

To examine the effects of aucubin on TNF- α -induced secretion of adipokines, 3T3-L1 adipocytes were pretreated with various concentrations of aucubin for 6 h and then incubated with 10 ng/mL TNF- α for 24 h. After incubation, conditioned medium was collected for ELISA assay. Treatment with aucubin inhibited TNF- α -induced increase in the secretion of MCP-1, PAI-1, and IL-6 in dose-dependent manner with IC₅₀ of 8.20, 9.94, and 6.85 μ M, respectively (Fig. 1).

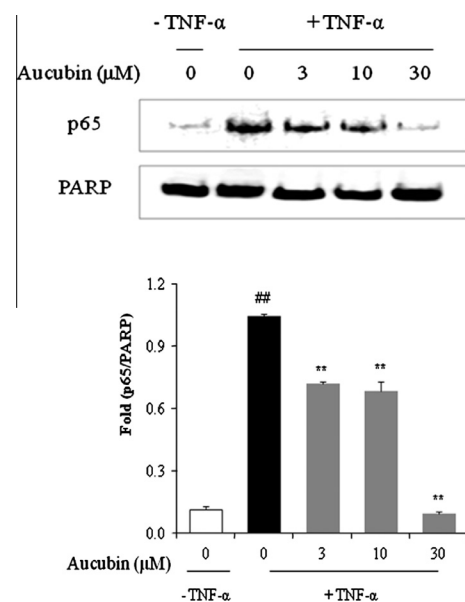


Fig. 3. Effects of aucubin on TNF- α -induced nuclear translocation of NF- κ B. Nuclear extracts, prepared from control or pretreated with different concentrations (1, 3, 10, and 30 μ M) of aucubin for 6 h, then TNF- α (10 ng/mL) for 24 h, were prepared for the Western blotting of p65 of NF- κ B using specific anti-p65 monoclonal antibody. PARP was used as internal controls. Density ratios versus PARP were determined by densitometry. The data shown are representative of three independent experiments. The values shown are means \pm SD. ## p < 0.01 versus untreated control; ** p < 0.01 versus TNF- α -only treated control.

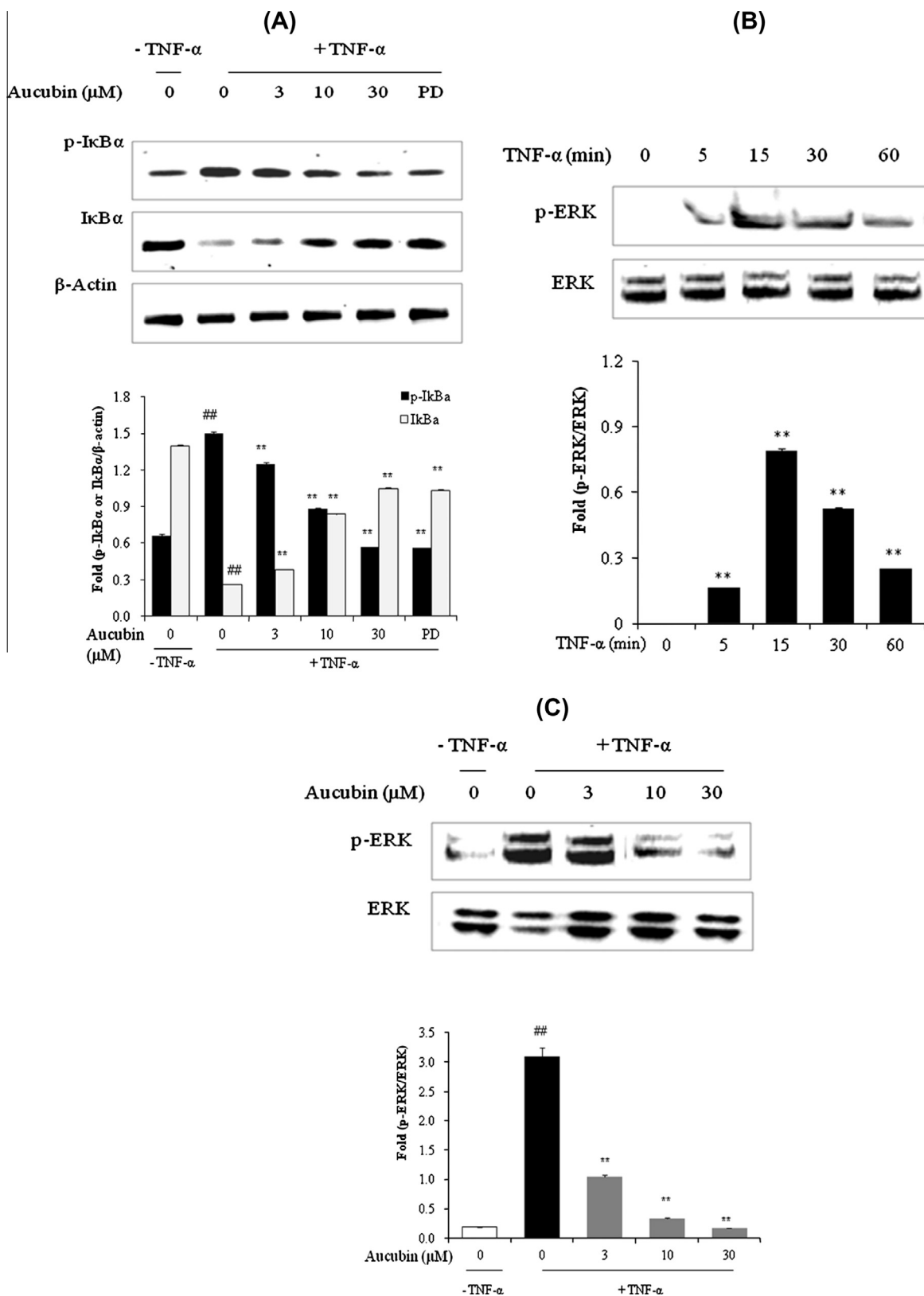


Fig. 4. Effects of aucubin on TNF- α -induced I κ B α degradation and ERK phosphorylation. (A) Effects of aucubin on I κ B α protein phosphorylation and degradation. Following pretreatment with aucubin (1, 3, 10, and 30 μ M) for 6 h, cells were treated with TNF- α (10 ng/mL) for 15 min. Total proteins were prepared and Western blotting was performed using specific p-I κ B α and I κ B α antibodies. PD98059 (PD, 20 μ M) was used as an ERK1/2-specific inhibitor. (B) Time-dependent changes of TNF- α -induced ERK phosphorylation. After stimulation with TNF- α (10 ng/mL), whole cell lysates prepared at different times (5, 15, 30, 60 min) were analyzed by Western blotting using specific p-ERK and ERK antibodies. (C) Effects of aucubin on TNF- α -induced ERK phosphorylation. Following pretreatment with aucubin (1, 3, 10, and 30 μ M) for 6 h, cells were treated with TNF- α (10 ng/mL) for 15 min. Whole cell lysates were prepared and Western blotting was performed using specific p-ERK and ERK antibodies. Density ratios versus β -actin or ERK were determined by densitometry. The data shown are representative of three independent experiments. The values are represented as means \pm SD. $^{##}p < 0.01$ versus untreated control; $^{**}p < 0.01$ versus TNF- α -only treated control.

3.2. Effects of aucubin on the TNF- α -induced expression of adipokine genes in 3T3-L1 adipocytes

To investigate whether aucubin inhibited TNF- α -induced secretion of adipokines by altering the gene expression, real-time quantitative RT-PCR was carried out with total RNA extracted from 3T3-L1 adipocytes of each group. The gene expression of adipokines was measured after 3T3-L1 adipocytes were exposed to aucubin for 6 h prior to TNF- α (10 ng/mL) treatment for 24 h. The enhanced production of MCP-1, PAI-1, and IL-6 mRNA by TNF- α was suppressed by aucubin pretreatment (Fig. 2). At a aucubin concentration of 10 μ M, the mRNA production of MCP-1, PAI-1, and IL-6 was suppressed by 36.5%, 24.3%, and 39.0%, respectively.

3.3. Effects of aucubin on the TNF- α -induced nuclear translocation of NF- κ B

Since NF- κ B is a major transcription factor that modulates the expression of pro-inflammatory cytokines, it was investigated whether aucubin prevents the translocation of the p65 subunit of NF- κ B from cytosol into the nucleus using Western blotting. It was found that aucubin prior to TNF- α significantly attenuated p65 level in the nuclear fraction (Fig. 3).

3.4. Effects of aucubin on the TNF- α -induced phosphorylation of I κ B α and ERK

In unstimulated adipocytes, NF- κ B is sequestered in the cytosol by its inhibitor I κ B α , and when cells are stimulated with TNF- α , I κ B is phosphorylated, ubiquitinated, and rapidly degraded via 26S proteasome, which results in the release of NF- κ B. Therefore, the effects of aucubin on the TNF- α -induced phosphorylation and degradation of I κ B α was examined using Western blotting. Aucubin significantly reduced the TNF- α -induced I κ B α phosphorylation in a dose-dependent manner. Furthermore, aucubin pretreatment blocked TNF- α -induced I κ B α degradation (Fig. 4A). Next, the effects of ERK1/2-specific inhibitor PD98059 (PD) on TNF- α -induced I κ B α phosphorylation and degradation were investigated. In parallel with the effects of aucubin, pretreatment with 20 μ M PD98059 reduced the TNF- α -induced I κ B α phosphorylation and degradation in adipocytes (Fig. 4A). These observations suggest that the phosphorylation and degradation of I κ B α is affected by ERK1/2 signal pathway. In fact, it has been known that activation of I κ B α degradation is a downstream event following the activation of the ERK1/2 pathway [24]. Thus, it was investigated whether aucubin regulates the phosphorylation of ERK using Western blotting. Challenge with TNF- α brought about the ERK phosphorylation within 15 min; then p-ERK band was gradually restored to basal level (Fig. 4B). Therefore, Western blotting for ERK was carried out after treating with TNF- α (10 ng/mL) for 15 min in the presence or absence of aucubin. 3T3-L1 adipocytes were pretreated with aucubin in the indicated concentrations for 6 h and then stimulated with TNF- α (10 ng/mL) for 15 min. Total cell lysates were then probed with phosphospecific antibodies for ERK1/2. The phosphorylation of ERK1/2 was increased in cells treated with TNF- α alone. However, aucubin significantly inhibited phosphorylated ERK1/2 levels in TNF- α -induced 3T3-L1 adipocytes in a concentration-dependent manner (Fig. 4C).

4. Discussions

Aucubin [1,4a,5,7a-tetra-5-hydroxy-7-(hydroxymethyl)cyclopenta(c)pyran-1-yl- β -D-glucopyranoside] is a common iridoid glycoside, found in a wide range of plants, which are used in folk medicine and traditional Chinese medicine. Those medicinal plants

containing aucubin show a variety of biological activities as aforementioned. Of pharmaceutical products as well as dietary supplements containing such medicinal plant materials, agents for treating inflammatory ailments including rheumatic inflammation have been marketed and used for long time. As for the molecular mechanism for anti-inflammatory activities of aucubin, it was reported that aucubin inhibits the production of TNF- α resulting from the both I κ B α degradation and the nuclear translocation of NF- κ B in RAW 264.7 cells [21].

It was previously reported that the increased levels of TNF- α associated with obesity may significantly contribute to elevated plasma and adipose tissue expression levels of adipokines [25]. This study demonstrated that aucubin effectively inhibited TNF- α -stimulated increases in the secretion of MCP-1, PAI-1, and IL-6 in 3T3-L1 adipocytes (Fig. 1). The mRNA production of each adipokines was suppressed in parallel with its secretion regulated by aucubin (Fig. 2).

Transcription factor NF- κ B has been evidenced to play a major role in the TNF- α -induced expression of adipokines in adipocytes [26]. NF- κ B is associated and controlled by I κ B α , which presents an inactive form in the cytoplasm. Upon stimulation, I κ B α becomes phosphorylated, undergoes degradation, and allows NF- κ B translocation into the nucleus, where it activates proinflammatory genes and participates in the inflammatory response [27]. In the present study, aucubin suppressed the nuclear translocation of NF- κ B (Fig. 3), which suggests that aucubin may decrease NF- κ B activity by inhibition of I κ B α degradation. Furthermore, it revealed that ERK1/2-specific inhibitor PD98059 blocked degradation of I κ B α upon TNF- α stimulation, implying that I κ B α degradation is a downstream event following the activation of the ERK1/2 pathway (Fig. 4A). Therefore, to confirm the inhibitory mechanisms of I κ B α degradation by aucubin, the effects of aucubin on TNF- α -induced ERK phosphorylation were investigated. It was found that aucubin exhibited significant inhibition on TNF- α -stimulated ERK1/2 phosphorylation in a concentration-dependent fashion (Fig. 4C).

In conclusion, aucubin can attenuate TNF- α -stimulated increases in gene expression and secretion of adipokines in 3T3-L1 adipocytes. The effects of aucubin were mediated by suppression of ERK phosphorylation, I κ B α degradation, and subsequent NF- κ B activation. The present study may contribute to the understanding of anti-inflammatory effects of aucubin and provide a novel mechanism of aucubin in preventing obesity-related pathologies.

References

- [1] Douketis JD, Sharma AM. Obesity and cardiovascular disease: pathogenic mechanisms and potential benefits of weight reduction. *Semin Vasc Med* 2005;5:25–33.
- [2] Trayhurn P, Wood IS. Adipokines: inflammation and the pleiotropic role of white adipose tissue. *Br J Nutr* 2004;92:347–55.
- [3] Grundy SM. Obesity, metabolic syndrome, and cardiovascular diseases. *J Clin Endocrinol Metab* 2004;89:2595–600.
- [4] Lyon CJ, Hsueh WA. Effect of plasminogen activator inhibitor-1 in diabetes mellitus and cardiovascular disease. *Am J Med* 2003;115:S62–8.
- [5] Shurk T, Hauner H. Obesity and impaired fibrinolysis: role of adipose production of plasminogen activator inhibitor-1. *Int J Obes Relat Metab Disord* 2004;28:1357–64.
- [6] Juhan-Vague I, Alessi MC, Mavri A, Morange PE. Plasminogen activator inhibitor-1, inflammation, obesity, insulin resistance, and vascular risk. *J Thromb Haemost* 2003;1:1575–9.
- [7] Morange PE, Alessi MC, Verdier M, Casanova S, Magalon G, Juhan-Vague I. PAI-1 produced ex vivo by human adipose tissue is relevant to PAI-1 blood level. *Arterioscler Thromb Vasc Biol* 1999;19:1361–5.
- [8] Shimomura I, Funahashi T, Takahashi M, Maeda K, Kotani K, Nakamura T, et al. Enhanced expression of PAI-1 in visceral fat: possible contributor to vascular disease in obesity. *Nat Med* 1996;2:800–3.
- [9] Gehardt CC, Romero IA, Cancelli R, Camoin L, Strosberg AD. Chemokines control fat accumulation and leptin secretion by cultured human adipocytes. *Mol Cell Endocrinol* 2001;175:81–92.

- [10] Sartipy P, Loskutoff DJ. Monocyte chemoattractant protein 1 in obesity and insulin resistance. *Proc Natl Acad Sci* 2003;100:7265–70.
- [11] De Taeye BM, Novitskaya T, McGuinness OP, Gleaves L, Medda M, Covington JW, et al. Macrophage TNF- α contributes to insulin resistance and hepatic steatosis in diet-induced obesity. *Am J Physiol Endocrinol Metab* 2007;293:E713–25.
- [12] Bernini R, Iavarone C, Trogolo C. 1-O- β -D-glucopyranosyleucommiol, an iridoid glucoside from *Aucuba japonica*. *Phytochemistry* 1984;23:1431–3.
- [13] Choi SY, Jung SH, Lee HS, Park KW, Yun BS, Lee KW. Plants with liver-protective activities (II). Glycation inhibitory activity and the identification of an active compound in *Plantago asiatica* extract. *Phytother Res* 2008;22:323–9.
- [14] Ho JN, Lee YH, Park JS, Jun WJ, Kim HK, Hong BS, et al. Protective effects of aucubin isolated from *Eucommia ulmoides* against UVB-induced oxidative stress in human skin fibroblasts. *Biol Pharm Bull* 2005;28:1244–8.
- [15] Miyagoshi M, Amagaya S, Ogihara Y. Choleretic actions of iridoid compounds. *Pharmacobiodynamics* 1988;11:186–90.
- [16] Davini E, Iavarone C, Trogolo C, Aureli P, Pasolini B. The quantitative isolation and antimicrobial activity of the aglycone of aucubin. *Phytochemistry* 1986;25:2420–2.
- [17] Konoshima T, Takasaki M, Tokuda H, Nishino H. Cancer chemopreventive activity of an iridoid glycoside, 8-acetylharpagide, from *Ajuga decumbens*. *Cancer Lett* 2000;157:87–92.
- [18] Chang IM, Yamaura Y. Aucubin: a new antidote for poisonous *Amanita* mushrooms. *Phytother Res* 1993;7:53–6.
- [19] Chang IM. Antiviral activity of aucubin against Hepatitis B virus replication. *Phytother Res* 1997;11:189–92.
- [20] Recio MC, Giner RM, Manez S, Rios JL. Structural considerations on the iridoids as anti-inflammatory agents. *Planta Med* 1994;60:232–4.
- [21] Park KS, Chang IM. Anti-inflammatory activity of aucubin by inhibition of tumor necrosis factor- α production in RAW 264.7 cells. *Planta Med* 2004;70:778–9.
- [22] Kim BH, Park KS, Chang IM. Elucidation of anti-inflammatory potencies of *Eucommia ulmoides* bark and *Plantago asiatica* seeds. *J Med Food* 2009;12:764–9.
- [23] Hasko G, Nemeth ZH, Szabo C, Zsilla G, Salzman AL, Vizi ES. Isoproterenol inhibits IL-10, TNF- α , and nitric oxide production in RAW 264.7 macrophages. *Brain Res Bull* 1998;45:183–7.
- [24] Shao Q, Shen LH, Hu LH, Pu J, Jing Q, He B. Atorvastatin suppresses inflammatory response induced by oxLDL through inhibition of ERK phosphorylation, I κ B α degradation, and COX-2 expression in murine macrophages. *J Cell Biochem* 2012;113:611–8.
- [25] Samad F, Loskutoff DJ, Pandey M. Molecular mechanisms of tumor necrosis factor- α -mediated plasminogen activator inhibitor-1 expression in adipocytes. *FASEB J* 2005;19:1317–9.
- [26] Jain RG, Phelps KD, Pekala PH. Tumor necrosis factor- α -initiated signal transduction in 3T3-L1 adipocytes. *J Cell Physiol* 1999;179:58–66.
- [27] May MJ, Acquisto FD, Madge LA, Glockner J, Pober JS, Ghosh S. Selective inhibition of NF- κ B activation by a peptide that blocks the interaction of NEMO with the I- κ B kinase complex. *Science* 2000;289:1550–4.