

UMAP on murine fetal brain 10k cells

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####The clustering and annotation of clusters shown here are only to illustrate how to use the Seurat package as a tool to analyze and visualize single-cell RNA-seq datasets. The analyses presented here are not meant to be used/considered as accurate representations of clustering and cluster annotations of any kind neurological/biological data. The results shown in these analyses are purely for the purpose of exercise and may nor may not have biological relevance.

Loading necessary packages

```
library (dplyr, quietly = TRUE)
```

```
##  
## Attaching package: 'dplyr'
```

```
## The following objects are masked from 'package:stats':  
##  
##      filter, lag
```

```
## The following objects are masked from 'package:base':  
##  
##      intersect, setdiff, setequal, union
```

```
library (Seurat)
```

Direct R to the directory you want to work in

```
setwd ("/Users/nr267/Desktop/All Work/Classes/Spring 2020/Seurat_class_resources/")
```

Load data and carry out QC

```
Neu_10k <- Read10X(data.dir = "../Seurat_class_resources/filtered_feature_bc_matrix_10k_neuron/10k_neuron/")  
Neu_10k <- CreateSeuratObject(counts = Neu_10k, project = "Neu_10k", min.cells = 3, min.features = 200)
```

```
Neu_10k
```

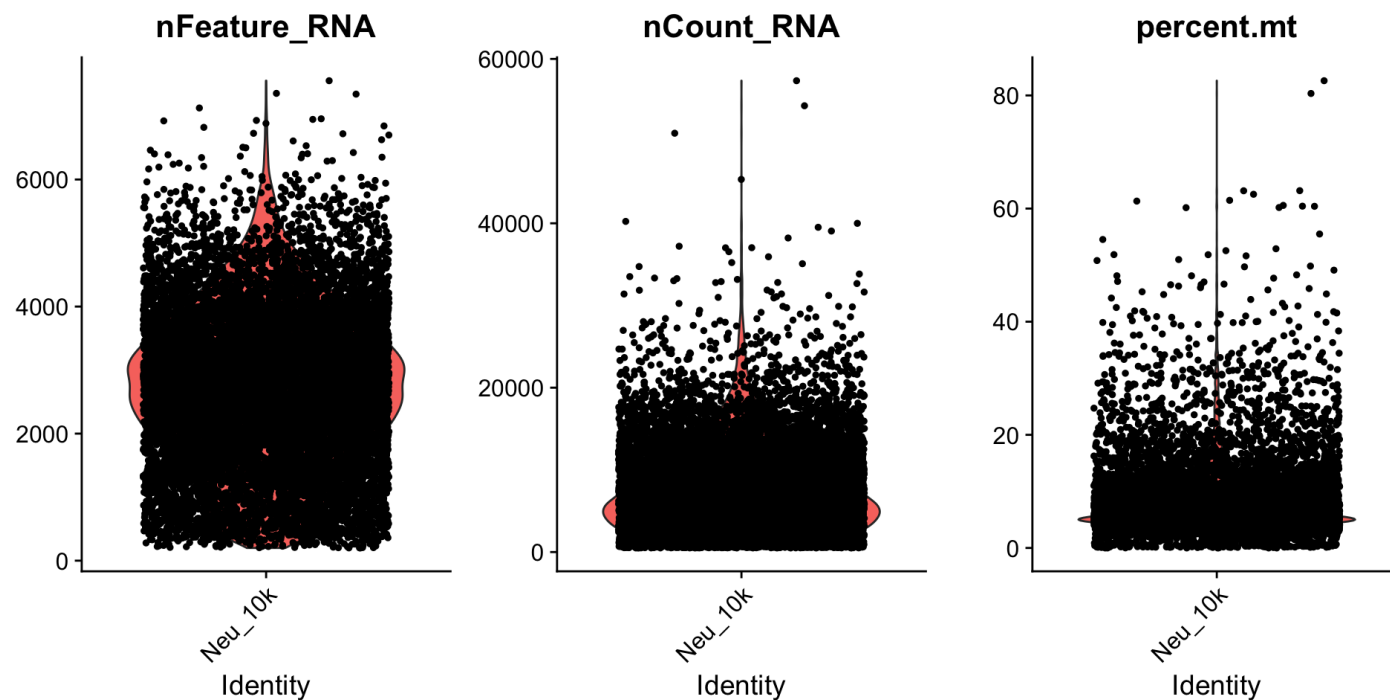
```
## An object of class Seurat
## 18564 features across 11648 samples within 1 assay
## Active assay: RNA (18564 features)
```

```
Neu_10k[["percent.mt"]] <- PercentageFeatureSet(Neu_10k, pattern = "^mt-")

head (Neu_10k@meta.data, 10)
```

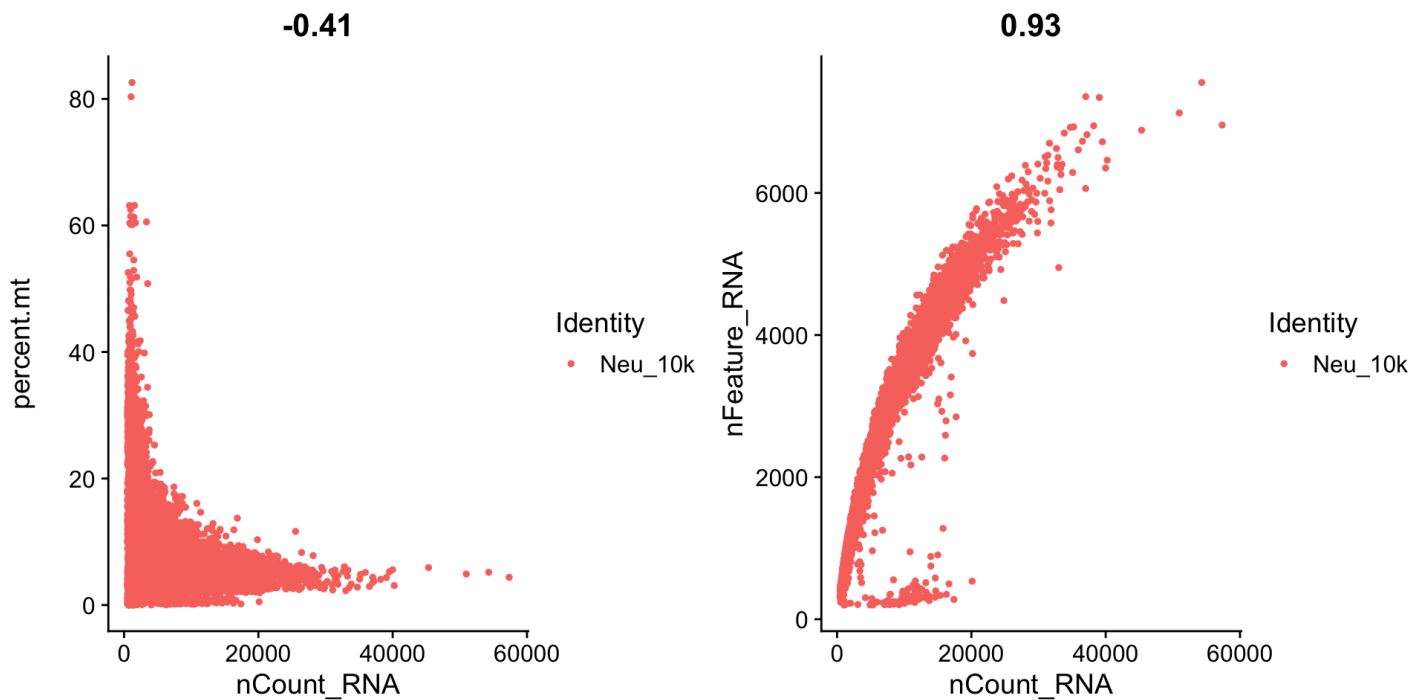
##	orig.ident	nCount_RNA	nFeature_RNA	percent.mt
## AAACCCAAGCAACTCT	Neu_10k	1421	784	0.2111189
## AAACCCACACGCGGTT	Neu_10k	5832	2399	11.9684499
## AAACCCACAGCATACT	Neu_10k	2977	1775	12.1598925
## AAACCCACATACCATG	Neu_10k	3599	1934	6.6129480
## AAACCCAGTCGCACAC	Neu_10k	9152	3473	4.9388112
## AAACCCAGTGCACATT	Neu_10k	3889	1733	12.7796349
## AAACCCAGTGGTAATA	Neu_10k	5206	2314	3.5920092
## AAACCCATCAGGTAATA	Neu_10k	3844	1952	6.1914672
## AAACGAAAGCAATAAC	Neu_10k	5934	2604	3.7074486
## AAACGAAAGGACAAGA	Neu_10k	2536	1410	9.9763407

```
VlnPlot (Neu_10k, features = c("nFeature_RNA", "nCount_RNA", "percent.mt"), ncol = 3)
```



```
plot1 <- FeatureScatter(Neu_10k, feature1 = "nCount_RNA", feature2 = "percent.mt")
plot2 <- FeatureScatter(Neu_10k, feature1 = "nCount_RNA", feature2 = "nFeature_RNA")

CombinePlots(plots = list(plot1, plot2))
```



Pre-process data

```
Neu_10k <- subset(Neu_10k, subset = nFeature_RNA > 200 & nFeature_RNA < 6000 & percent.mt < 10)
```

Check data dimensions after subsetting

Neu_10k

```
## An object of class Seurat
## 18564 features across 9276 samples within 1 assay
## Active assay: RNA (18564 features)
```

LogNormalize Data (normalize count data per cell and transform to log scale)

```
Neu_10k <- NormalizeData(Neu_10k, verbose = TRUE)
```

Pre-process data for PCA

```

Neu_10k <- FindVariableFeatures(Neu_10k, selection.method = "vst", nfeatures = 2000)

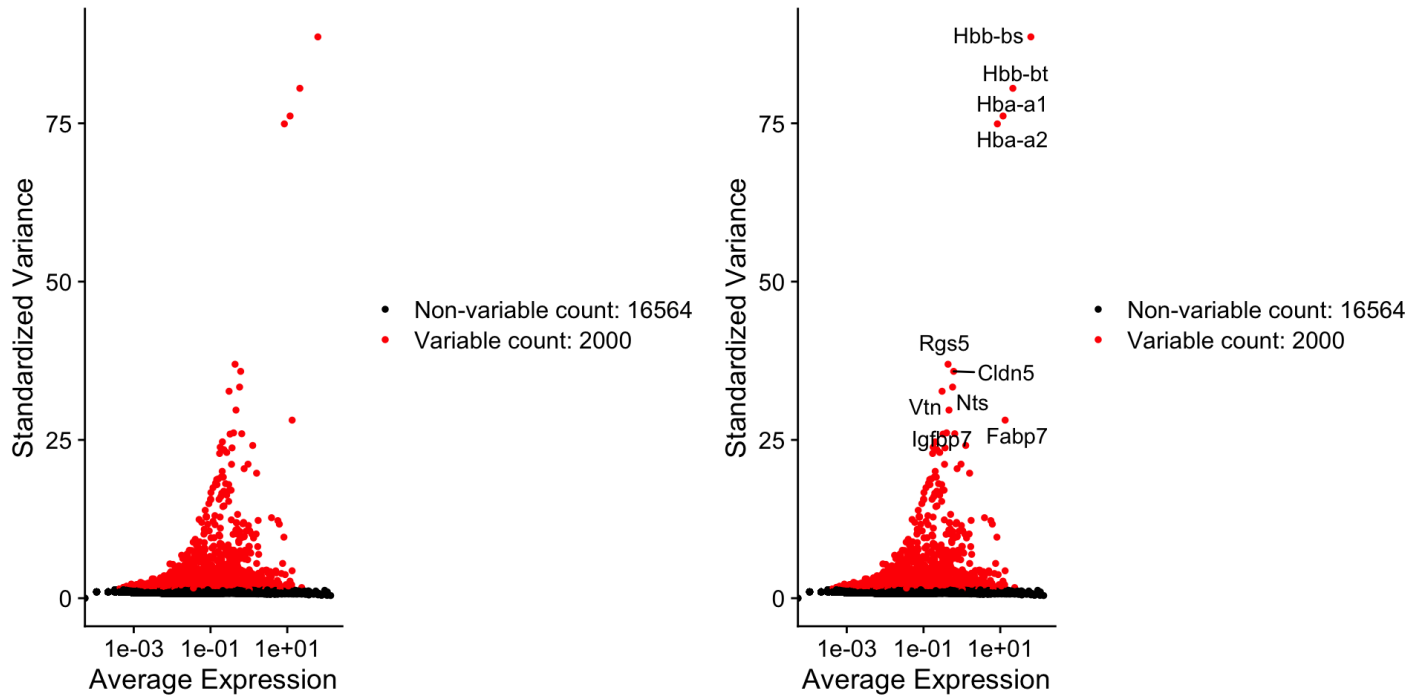
top10 <- head(VariableFeatures(Neu_10k), 10)

plot3 <- VariableFeaturePlot(Neu_10k)

plot4 <- LabelPoints(plot = plot3, points = top10, repel = TRUE)

CombinePlots(plots = list(plot3, plot4))

```



```

all.genes <- rownames(Neu_10k)

Neu_10k <- ScaleData(Neu_10k, features = all.genes)

Neu_10k

```

```

## An object of class Seurat
## 18564 features across 9276 samples within 1 assay
## Active assay: RNA (18564 features)

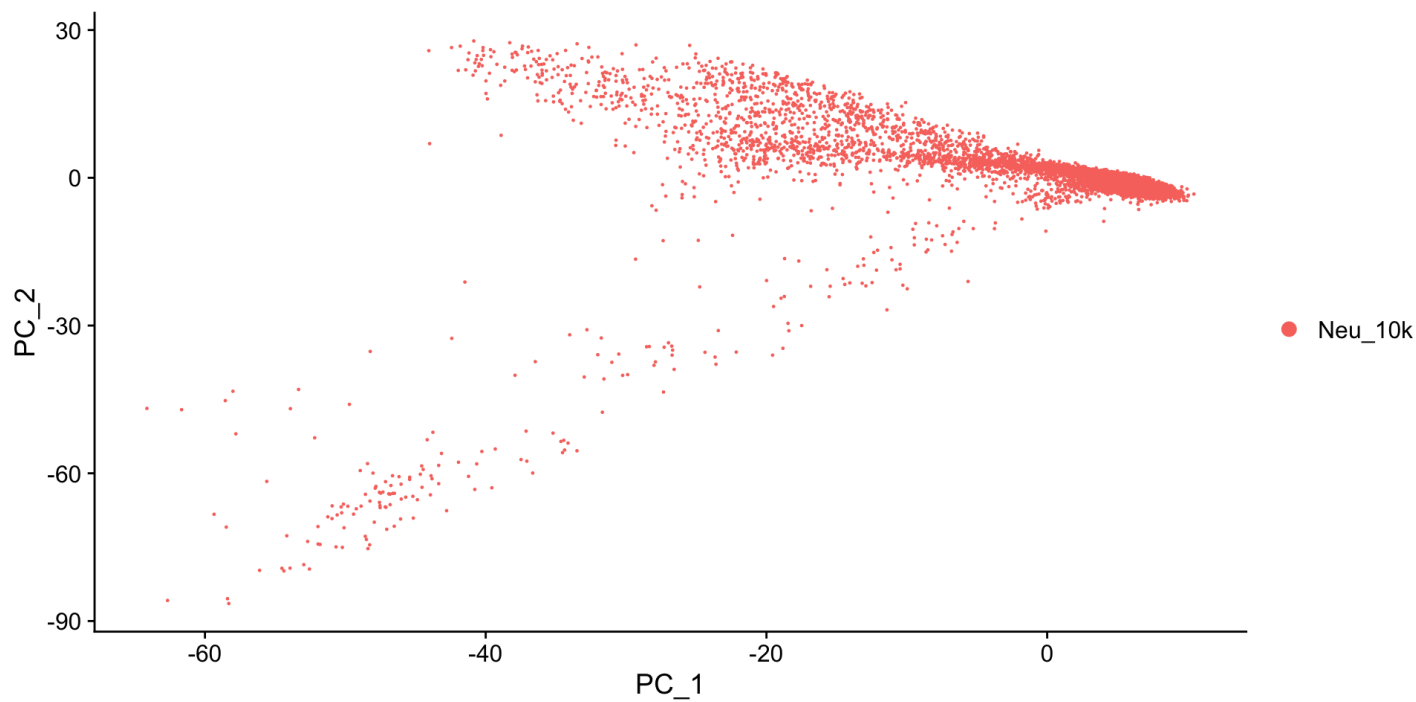
```

```

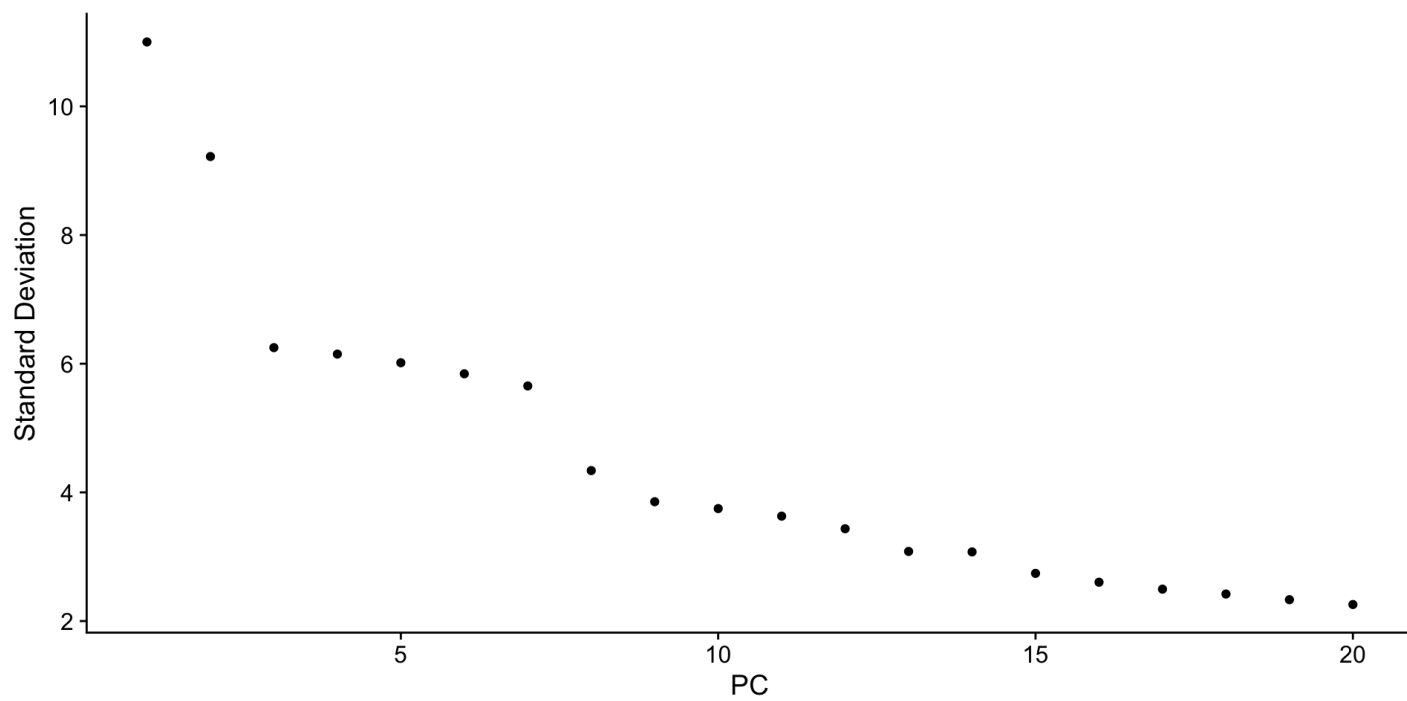
Neu_10k <- RunPCA(Neu_10k, features = VariableFeatures(object = Neu_10k))

DimPlot (Neu_10k, reduction = "pca")

```

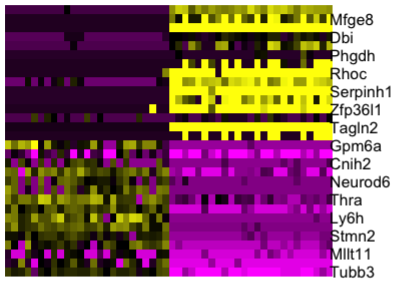


```
ElbowPlot(Neu_10k)
```

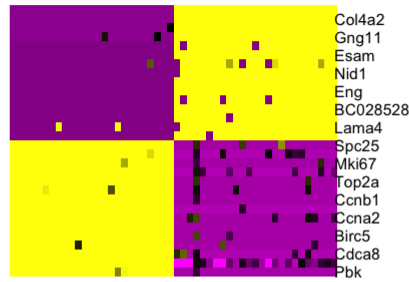


```
DimHeatmap(Neu_10k, dims = 1:15, cells = 50, balanced = TRUE)
```

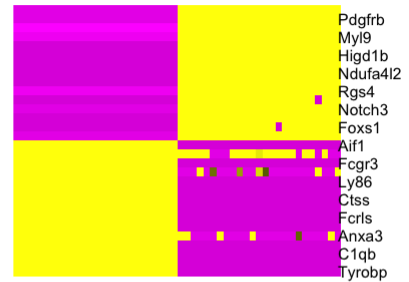
PC_1



PC_2



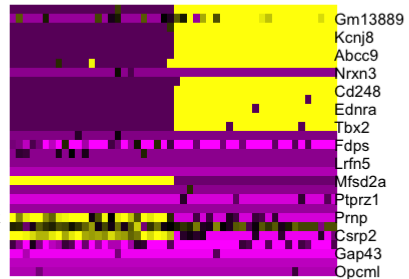
PC_3



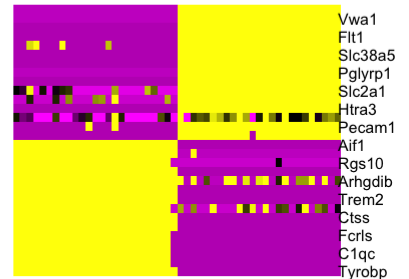
PC_4



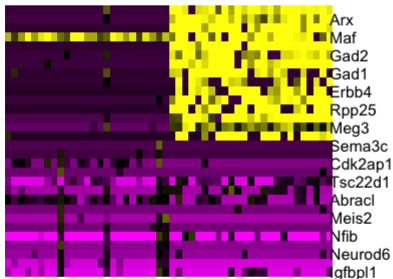
PC_5



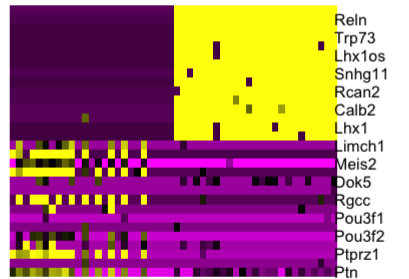
PC_6



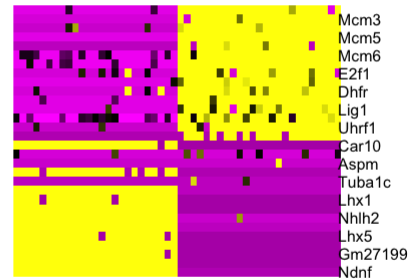
PC_7



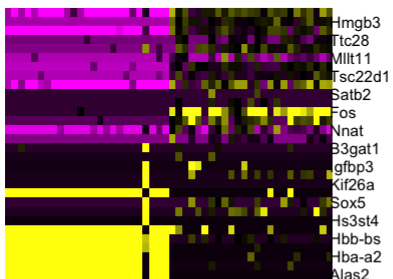
PC_8



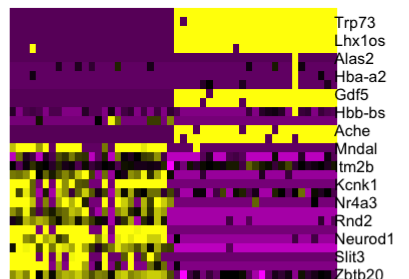
PC_9



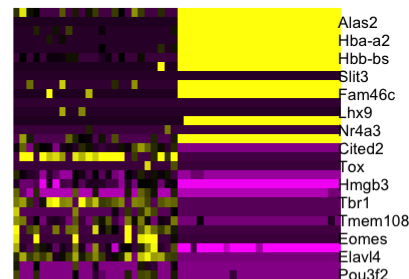
PC_10



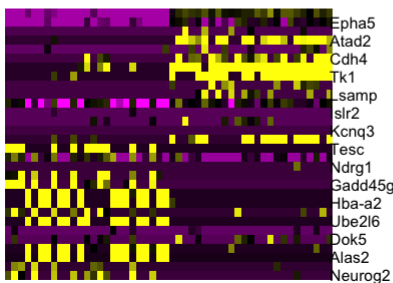
PC_11



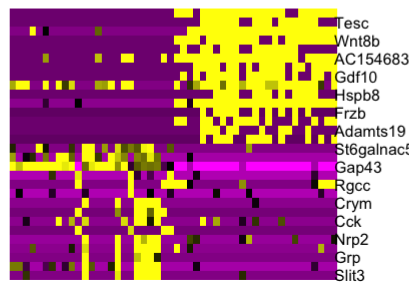
PC_12



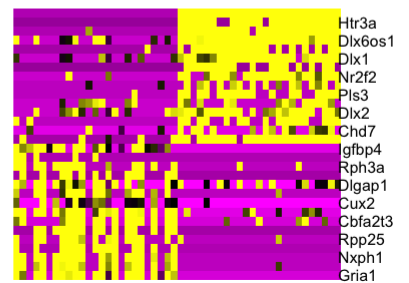
PC_13



PC_14



PC_15



Run UMAP on dataset

```
Neu_10k <- FindNeighbors(Neu_10k, dims = 1:10)
```

```
## Computing nearest neighbor graph
```

```
## Computing SNN
```

```
Neu_10k <- FindClusters (Neu_10k, resolution = 0.5)
```

```
## Modularity Optimizer version 1.3.0 by Ludo Waltman and Nees Jan van Eck
##
## Number of nodes: 9276
## Number of edges: 298365
##
## Running Louvain algorithm...
## Maximum modularity in 10 random starts: 0.9073
## Number of communities: 17
## Elapsed time: 0 seconds
```

```
Neu_10k <- RunUMAP(Neu_10k, dims = 1:10)
```

```
## Warning: The default method for RunUMAP has changed from calling Python UMAP via reti
culate to the R-native UWOT using the cosine metric
## To use Python UMAP via reticulate, set umap.method to 'umap-learn' and metric to 'cor
relation'
## This message will be shown once per session
```

```
## 00:31:00 UMAP embedding parameters a = 0.9922 b = 1.112
```

```
## 00:31:00 Read 9276 rows and found 10 numeric columns
```

```
## 00:31:00 Using Annoy for neighbor search, n_neighbors = 30
```

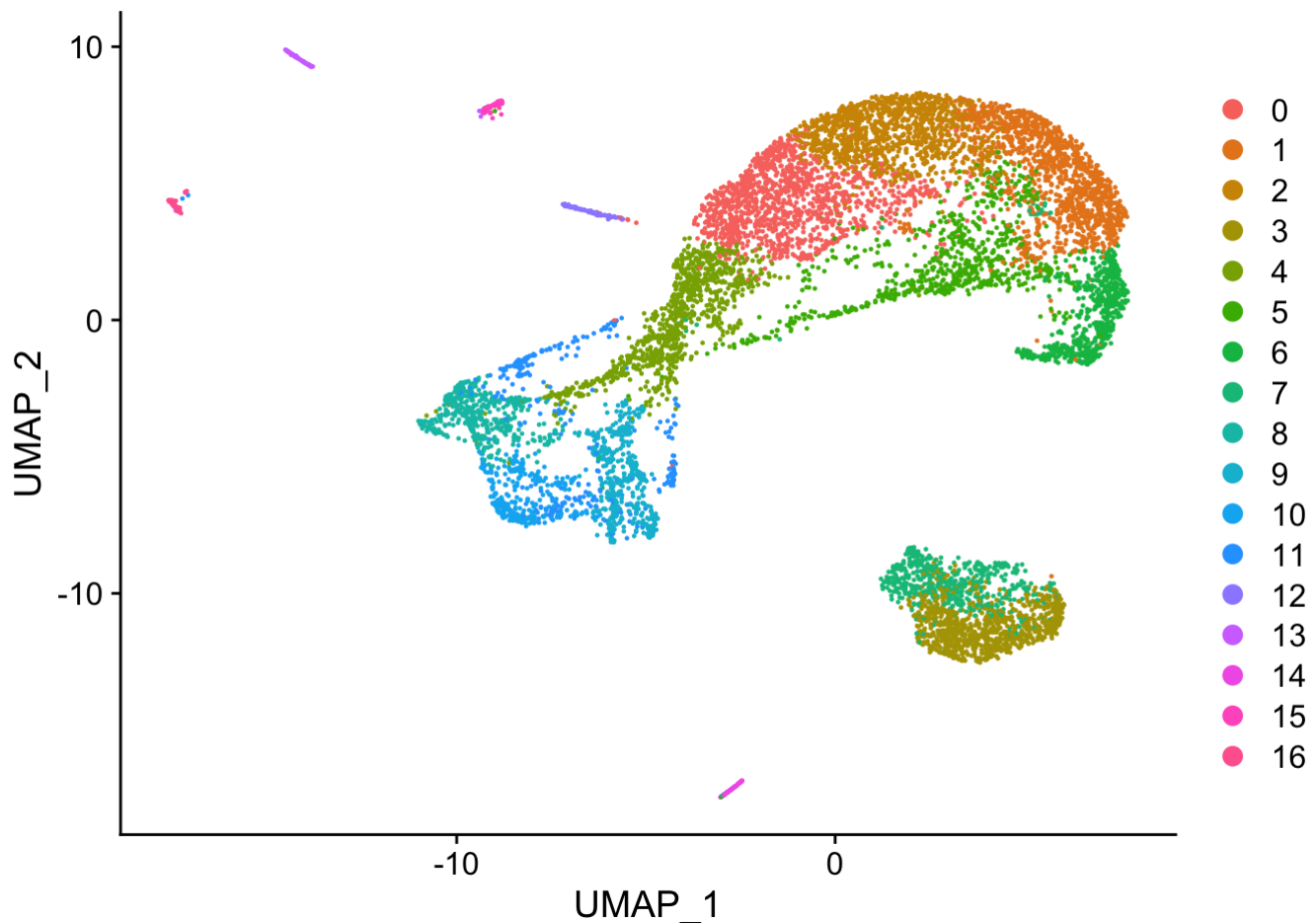
```
## 00:31:00 Building Annoy index with metric = cosine, n_trees = 50
```

```
## 0%    10    20    30    40    50    60    70    80    90   100%
```

```
## [----|----|----|----|----|----|----|----|----|----|
```

```
## *****|
## 00:31:01 Writing NN index file to temp file /var/folders/s9/6k4hffrx25gb0_43dcj132p15
4t8k1/T//Rtmp6NUwtY/file36f026a016b
## 00:31:01 Searching Annoy index using 1 thread, search_k = 3000
## 00:31:03 Annoy recall = 100%
## 00:31:03 Commencing smooth kNN distance calibration using 1 thread
## 00:31:04 Initializing from normalized Laplacian + noise
## 00:31:04 Commencing optimization for 500 epochs, with 374866 positive edges
## 00:31:15 Optimization finished
```

```
DimPlot (Neu_10k, reduction = "umap")
```



Find all markers of each cluster to identify them

```
cluster0.markers <- FindMarkers (Neu_10k, ident.1 = 0, min.pct = 0.5)
cluster1.markers <- FindMarkers (Neu_10k, ident.1 = 1, min.pct = 0.5)
cluster2.markers <- FindMarkers (Neu_10k, ident.1 = 2, min.pct = 0.5)
cluster3.markers <- FindMarkers (Neu_10k, ident.1 = 3, min.pct = 0.5)
cluster4.markers <- FindMarkers (Neu_10k, ident.1 = 4, min.pct = 0.5)
cluster5.markers <- FindMarkers (Neu_10k, ident.1 = 5, min.pct = 0.5)
cluster6.markers <- FindMarkers (Neu_10k, ident.1 = 6, min.pct = 0.5)
```


To view the genes lists on your RStudio console

```
View(cluster0.markers)
```

```
View(cluster1.markers)
```

```
View(cluster2.markers)
```

```
View(cluster3.markers)
```

```
View(cluster4.markers)
```

```
View(cluster5.markers)
```

```
View(cluster6.markers)
```

To “export” the genes lists to your current directory

```
write.csv(cluster0.markers, “cluster0_markers_UMAP”)
```

```
write.csv(cluster1.markers, “cluster1_markers_UMAP”)
```

```
write.csv(cluster2.markers, “cluster2_markers_UMAP”)
```

```
write.csv(cluster3.markers, “cluster3_markers_UMAP”)
```

```
write.csv(cluster4.markers, “cluster4_markers_UMAP”)
```

```
write.csv(cluster5.markers, “cluster5_markers_UMAP”)
```

```
write.csv(cluster6.markers, “cluster6_markers_UMAP”)
```