

Dendritic spiking accounts for rate and phase coding in a biophysical model of a hippocampal place cell

Zsófia Huhn ^{a,*}, Máté Lengyel ^a, Gergő Orbán ^a, Péter Érdi ^{a,b}

^a*Dept. Biophys., KFKI R.I.P.N.P., Hun. Acad. Sci, 29-33 Konkoly Thege M. út, Budapest H-1121, Hungary*

^b*Center for Complex Systems Studies, Kalamazoo College, 1200 Academy street, Kalamazoo, MI 49006-3295, USA*

Abstract

Hippocampal place cells provide prototypical examples of neurons firing jointly phase and rate coded spike trains. We propose a biophysical mechanism accounting for the generation of place cell firing at the single neuron level. An interplay between external theta-modulated excitation impinging the dendrite and intrinsic dendritic spiking as well as between frequency modulated dendritic spiking and periodic somatic hyperpolarization was a key element of the model. Through these interactions robust phase and rate coded firing emerged in the model place cell, reproducing salient experimentally observed properties of place cell firing.

Key words: phase precession, tuning curve, active dendrite

1. Introduction

Hippocampal place cells code the spatial position of the animal both by their firing rate and the precise timing of their firings. A place cell fires only when the rat traverses the particular portion of the environment preferred by the cell, the 'place field' of the cell [1]. Rate coding implies that during a single traversal, firing frequency of the cell increases in the early and decreases in the late portion of the place field [2,3]. Phase coding was demonstrated as a monotonic decrease in the phase of firings relative to the ongoing theta oscillation during a single traversal [2,4,5,3].

In most of the models that have been suggested to account for these two phenomena, hippocampal area CA3, where pyramidal cells are known to be densely interconnected by recurrent collaterals, played a key role [6–8]. However, experimental studies reported that place cell activity could be maintained in CA1 even when input from CA3 was severely impaired [9,10]. Recently, Lengyel et al. [11] has shown both by analytical calculations and numerical simulations of a simplified single neuron model that the doubly coded place cell firing pattern can be generated in a cell receiving periodic perisomatic inhibitory input and dendritic excitatory input from the perforant path without the need for recurrent excitatory connections. Here we present a biologically more realistic version of this model which was extended in two ways. First, a biophysical conductance-based model was adopted,

* Corresponding author.

Email address: zsofi@rmki.kfki.hu (Zsófia Huhn).

and thus the dynamics of intrinsic dendritic membrane potential oscillations were studied in detail. Second, theta modulation of perforant path transmission [12,13] was taken into account. The interaction of these two oscillations in the dendrite generated dendritic spiking that in turn interacted with the periodic hyperpolarization of the soma. Through these interactions robust phase and rate coded firing emerged in the model place cell.

2. Methods

Following our earlier work [11], the present model consisted of a somatic and dendritic compartment, and received two inputs. (1) The somatic compartment received a hyperpolarizing oscillating current (SHOC) with a constant theta frequency, in phase with the ongoing theta field potential oscillation [14–16]. (2) Dendritic postsynaptic current represented perforant path input and was a sum of two terms: (i) a sinusoidal term oscillating with the same frequency but in anti-phase with SHOC, and (ii) a speed-dependent term that was proportional to the speed of the rat and was switched on when the animal was inside the place field of the cell.

The dendrite sustained intrinsic membrane potential oscillations (DMPO) within the place field, that was frequency modulated by the speed-dependent term, and was in anti-phase with SHOC at the beginning of the traversal of the place field due to the oscillatory term. Although entorhinal cells, the source of perforant path input, have position-dependent receptive fields, this ensemble activity was shown to be transformed to velocity-dependent postsynaptic excitation in hippocampal place cells by properly timed theta-modulated inhibition [17]. The net excitation received by the soma of the hippocampal place cell was the sum of SHOC and the current flowing from the dendrite due to DMPO.

The model cell was a modified Pinsky-Rinzel neuron [18] which is a two-compartmental pyramidal cell model possessing voltage-dependent currents. Passive parameters of the model were: $C_m = 1 \mu\text{F}/\text{cm}^2$ membrane capacitance, $g_L =$

$0.3 \text{ mS}/\text{cm}^2$ leak conductance, $V_L = -60 \mu\text{V}$ leak reversal potential, $g_c = 0.01 \text{ mS}/\text{cm}^2$ axial conductance between the soma and the dendrite and $p = 0.2$ soma/cell surface area ratio. Equilibrium potentials (in mV) and maximal channel conductances (in mS/cm^2) of active conductances were $V_{\text{Na}} = 60$, $V_{\text{K}} = -75$, $V_{\text{Ca}} = 80$, and $g_{\text{Na}} = 30$, $g_{\text{K-DR}} = 15$, $g_{\text{Ca}} = 10$, $g_{\text{K-C}} = 15$, $g_{\text{K-AHP}} = 0$. In order to shorten somatic action potentials, the exponents of the m and n gating variables were increased from 2 to 3, and from 1 to 4, respectively.

SHOC represented inhibitory input at theta frequency from hippocampal interneurons [15]: $I_s(t) = A_s/2 \cos(2\pi f_\theta t) + I_{0-s}$, where $f_\theta = 8 \text{ Hz}$, $A_s = 4 \mu\text{A}/\text{cm}^2$ and $I_{0-s} = -0.5 \mu\text{A}/\text{cm}^2$. The only input to the dendrite arrived from entorhinal cortex, and it had two components: $I_d(t) = I_\theta(t) + I_v[v(t)]$. I_θ was a sinusoid depolarizing current in antiphase with SHOC: $I_\theta(t) = A_d \cos(\pi + 2\pi f_\theta t) + I_{0-d}$, where $A_d = 0.5 \mu\text{A}/\text{cm}^2$ and $I_{0-d} = 1.9 \mu\text{A}/\text{cm}^2$. Velocity-dependent current arose only when the rat was within the afferent place field of the cell (i.e., the area covered by all the place fields of its presynaptic entorhinal partners), and there it was a linear function of the rat's running speed: $I_v[v(t)] = kv(t) \cdot \mathcal{H}[x(t) - x_{\text{in}}] \cdot \mathcal{H}[x_{\text{out}} - x(t)]$, where $k = 0.01 (\mu\text{A}/\text{cm}^2) / (\text{cm}/\text{sec})$, $x(t)$ is the position of the rat, and x_{in} and $x_{\text{out}} = x_{\text{in}} + l$ are the entry and exit points of the afferent place field, l is the length of the afferent place field, and \mathcal{H} is the Heaviside function.

Phase response curves (PRCs) of the separated dendrite were calculated in response to pulse and periodic perturbations. Dendritic spiking was elicited by $I_{0-d} = 1.82 \mu\text{A}/\text{cm}^2$ current injection. Perturbations were timed at different phases (τ) of the spiking cycle, and consisted of current pulses of 0.5 msec duration and variable amplitude (Fig. 5) starting at τ , or one cycle of sinusoid current injection of $A_d = 0.5 \mu\text{A}/\text{cm}^2$ amplitude and f_θ frequency starting with its peak at $-\tau$. Phase response was measured as a phase advancement of the dendritic spiking oscillation in response to the perturbation compared to the unperturbed case: positive or negative values showed shortening or lengthening of the spiking cycle, respectively. As a control, PRCs for periodic perturbation were

also calculated from pulse PRCs by integrating the product of the pulse PRC with the sinusoid perturbation.

3. Results

First we analyzed the behavior of the separated dendrite ($g_c = 0$) in response to constant current injection (data not shown). Depending on the level of depolarization, the dendrite either settled to a resting membrane potential (stable fixed point) or sustained an intrinsic membrane potential oscillation. Amplitude of oscillation decreased while frequency increased with increasing external current amplitude. In the relevant frequency regime, that is close to the 8 Hz theta frequency, these oscillations were high amplitude dendritic calcium spikes (see Fig. 2B).

The interaction of a theta frequency (8 Hz) sinusoid external current oscillating around a given mean value ('offset') with intrinsic dynamics was also studied in the separated dendrite, so that the offset of the external oscillation was changed while its amplitude (minimum to maximum depth) was kept constant (Fig. 1A). At lowest current intensities ($< 1.4 \mu\text{A}/\text{cm}^2$) no spikes or subthreshold oscillations were generated. After a short transition of dendritic oscillations at half theta frequency, dendritic spikes appeared and were driven by the external oscillation at 8 Hz in a wide range of medium current intensities (between $1.5 - 1.9 \mu\text{A}/\text{cm}^2$). At higher levels of injected current the offset component of external depolarization dominated, eliciting intrinsic membrane potential oscillations in the dendrite at frequencies corresponding to the offset level (frequency modulation). Due to interaction with the sinusoid component of the external current these oscillations were not perfectly regular: interspike intervals (ISIs) varied, but this variation was small enough not to mask the frequency modulation effect.

In the complete model with soma and dendrite coupled through an axial conductance, the amount of current received by the dendrite during a traversal of the environment was in the medium to high ranges. When the rat was outside the

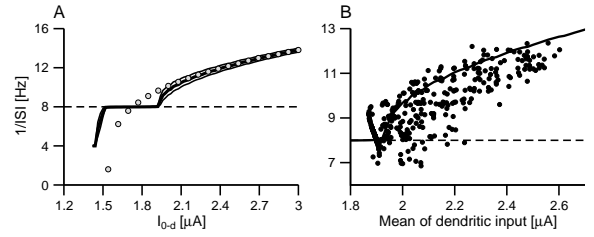


Fig. 1. Frequency modulation of dendritic spiking in response to sinusoid current injection. Offset level of external current was changed, while its peak-to-trough amplitude was kept constant. A. Firing frequency tuning curve of the separated dendrite at different offset levels of external current (I_{0-d}). ISIs were transformed to instantaneous firing frequency values. Thick solid line shows average firing frequency, thin solid lines show range of ± 1 standard deviation. Open circles show frequency-current tuning curve in response to constant current injection for comparison. Dashed line shows frequency of external stimulation at 8 Hz. B. Dendritic firing frequency with soma attached. Offset level was continuously changing in time. Dots show instantaneous firing frequencies calculated from individual ISIs as a function of average external current entering the dendrite during the corresponding ISI. Solid line shows average ISIs expected from simulations of the separated dendrite (solid line in A) for reference. Dashed line shows 8 Hz theta frequency.

place field, offset of dendritic current injection was low, thus its relative depth of modulation (peak-to-trough amplitude relative to peak level) was high, effectively driving DMPO at theta frequency (Fig. 1B). As the rat crossed the boundary of the place field, the velocity-dependent component of dendritic excitation was switched on, increasing its offset to a level where it induced intrinsic dendritic oscillations and also reducing its relative depth of modulation. Consequently, intrinsic dynamics of the dendrite shaped the frequency of DMPO similarly to that seen in the case of the separated dendrite (Fig. 1B). Note, that the dendritic frequency-current tuning curve in the complete model was slightly shifted towards higher current intensities compared to the separated dendrite. This can be explained as a weak shunting of dendritic membrane potential by the somatic compartment through the axial conductance. Positive axial current to the dendrite flowed only during somatic action potential peaks, but these were too short to have any measurable effect on DMPO (data not shown).

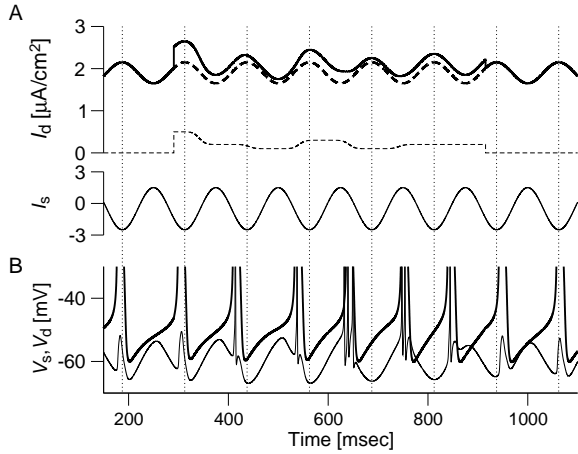


Fig. 2. A typical traversal of the place field. A. External inputs to the somatic (lower trace) and dendritic (upper traces) compartments. Net dendritic transmembrane excitation (thick solid line) consisted of a persistent theta frequency (thick dashed line) and a place field-specific velocity-dependent (thin dashed line) component. Current injected to the soma (thin solid line) was in antiphase with dendritic periodic input. Vertical dotted lines show troughs of SHOC for reference. B. Somatic (lower thin line) and dendritic (upper thick line) membrane potential. Note the somatic depolarization bumps caused by dendritic spikes not reaching firing threshold in the first and last cycles shown, but giving rise to a graded number of action potentials in intermediate cycles. Dendritic and somatic spikes were clipped for better visibility of subthreshold events.

As intradendritic firing frequencies inside the afferent place field were higher than theta frequency phase of dendritic spiking gradually phase precessed relative to SHOC reflecting field theta oscillation (Fig. 2). The effect of dendritic spikes on somatic firing depended on the phase of SHOC at which they were fired (Fig. 2B). Dendritic spikes fired near the troughs of SHOC did not cause somatic firing, because somatic hyperpolarization was strong enough to prevent them reaching firing threshold in the soma. However, as dendritic spikes appeared at increasingly higher levels of somatic depolarization, they were able to trigger bursts consisting of an increasing number of somatic action potentials. This mechanism together with the phase precession effect ensured that firing frequency as a function of position formed a unimodal curve (Fig. 3A).

Earlier work on a simplified neuron [11] pre-

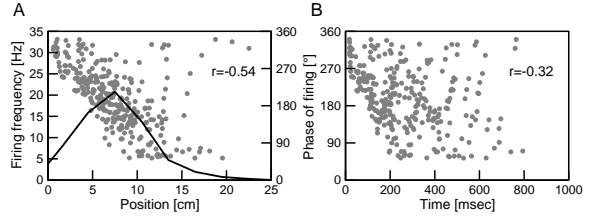


Fig. 3. Firing phase and rate during multiple traversals. A. Firing phase was monotonically decreasing (dots), while average firing rate was a unimodal function of position (solid line). Firing phase was better correlated with position (A) than with time spent in the place field (B). Each dot represent a single spike.

dicted that, in line with experiments [2], correlation between phase of somatic spikes and position would be higher than between somatic spikes and time, as observed in experiments. To test this prediction we simulated 50 traversals of the place field and calculated respective correlation coefficients. We found that firing phase correlated significantly stronger with the position of the animal than with time spent in the place field (Fig. 3A and B).

Place cells often show nearly 360° phase precession [2,4]. In order to reproduce this in the model (Figs. 2,3) two parameters were carefully tuned: l , the length of afferent place field, i.e. the area containing place fields of all presynaptic entorhinal cells, and k , the current/velocity conversion factor being proportional to the synaptic strength of perforant path synapses. Both of these parameters were proportional to the cumulative velocity-dependent excitation received by the dendrite during a traversal of the place field, and thus to the phase at which dendritic spikes were emitted on exit from the place field (see also Methods). Furthermore, if dendritic spiking phase had not been in antiphase with SHOC on exit, it would also have corrupted the firing frequency tuning curve of the cell, not just the pattern of phase precession, as firing would have continued on indefinitely.

Contrary to these ominous expectations, the model showed robust behavior even when k and l were set to non-optimal values: phase of dendritic firing precessed or recessed until it got in antiphase with SHOC (Fig. 4). This was achieved through the following ‘branching behavior’: if dendritic spiking phase on exit did not reach a $\sim 180^\circ$

threshold value then dendritic spiking recessed (reversed precession), otherwise, if place field-specific input was able to cause sufficient precession to get past this threshold phase, then dendritic spiking finished precession up to a full theta cycle independent of the place field-specific term of the entorhinal input. By analysing phase response curves of the dendritic compartment, the source of this branching behavior became apparent: DMPO was only sensitive to perturbations in the part of the cycle immediately preceding spiking (Fig. 5A). As a consequence, in response to periodic excitation, dendritic spiking had one stable fixed point at 330° (approximately in phase with dendritic excitation, thus in antiphase with SHOC), and an unstable fixed point at 220° (Fig. 5B). Because the length of the animal's trajectory within the afferent place field may never be exactly the same, especially in two-dimensional environments, it seems inevitable that on some traversals phase recession instead of precession will occur in the late portion of the place field. We predict that this branching behavior may be the source of the decreased correlation of firing phase with position in the second half of the place field as observed in experiments [4,19,20].

4. Discussion

Place cells provide prototypical examples of neurons firing jointly phase and rate coded spike trains [2,5,19,3]. Many theories have tried to explain this phenomenon, but most of these models used only abstract neuron [2,19,11], relied on implausible assumptions about the connectivity of the network encompassing place cells [6–8,21,22], or failed to reproduce both the phase and rate code appropriately [6–8,21,23,5,19].

In this study, we have proposed a biophysical mechanism accounting for the generation of place cell firing at the single neuron level. An interplay between external theta-modulated excitation impinging the dendrite and intrinsic dendritic spiking (Fig. 1) as well as between dendritic spiking and periodic somatic hyperpolarization (Fig. 2) was a key element of the model. Indeed, hippocampal pyramidal cells were shown to be capable of sus-

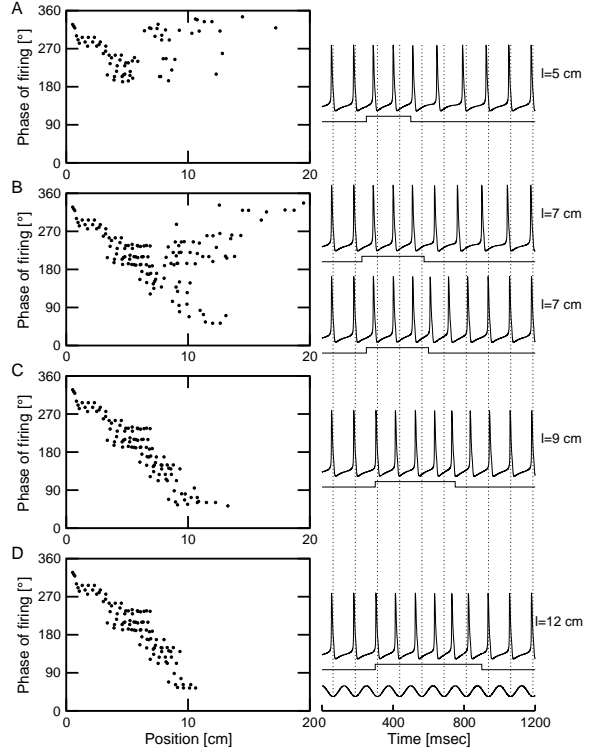


Fig. 4. The effect of changing the length of the afferent place field, l , while keeping the current/velocity conversion factor, k , constant. If the afferent place field is too short and dendritic spiking on exit does not reach a $\sim 220^\circ$ threshold value, phase recession follows the break off of place field-specific excitation (A). If the afferent place field is longer and thus phase precession continues longer in it, past the threshold value, precession after exit continues ‘automatically’ until reaching 360° (C,D). For intermediate afferent place field lengths, some traversals may result in recession while in others a full cycle of precession is produced depending on the exact phase on exit from the place field (B). Panels on the right show dendritic membrane potential traces during individual traversals (upper lines) with binary traces showing when the animal was within the afferent place field (lower lines), and vertical dotted lines showing troughs of SHOC. Panels on the left show phase-position plots of multiple traversals (with different running velocities and entry phases), each dot corresponding to a single spike.

taining rhythmic firing of dendritic calcium spikes close to theta frequency [16]. Moreover, extracellular currents in the somatic and dendritic layer of the hippocampus proper are theta modulated and in antiphase with each other [16], just as assumed by the model.

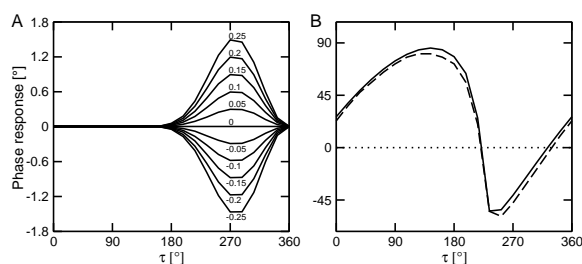


Fig. 5. Phase response curves of the dendritic compartment. A. The separated dendrite received constant current that triggered repetitive dendritic firing with approximately theta frequency. The effect of the injection of a short extra current pulse depended on the phase of the intrinsic oscillation at which this perturbing current was applied ($0 = 360^\circ$: peak of dendritic spike without perturbation). In the first half of the cycle ($0-180^\circ$) perturbations had no effect; in the second half ($180-360^\circ$) the sensitivity of the dendritic oscillation increased until about 270° and decreased afterwards. Numbers on different phase response curves denote the amplitudes of perturbations in $\mu\text{A}/\text{cm}^2$. Negative current caused lengthening of the cycle (dendritic spike was delayed), whereas positive current accelerated the cycle (dendritic spike occurred earlier). The extent of this decelerating/accelerating effect increased nearly linearly with the amplitude of the applied current. B. The separated dendrite received one cycle sinusoid current, with the same offset value as the baseline excitation in A. Solid line shows the result of simulations with sinusoid excitation, dashed line shows prediction from instantaneous phase response curves in A for comparison (see Methods for details). The horizontal axis shows phase lag of dendritic spiking to the periodic excitation. Intersections with the horizontal axis show stable values of this phase lag in response to continued sinusoid excitation: intersection with negative slope at 220° is unstable, intersection with positive slope at 330° is stable.

The model produced spike trains conforming experimental result (Fig. 3), in which firing rate was found to be a graded, unimodal function of position [24,25,2,3], phase precession was a monotonically decreasing function of position [2,4,3] and phase was better correlated with position than with time [2,3].

References

- [1] J. O'Keefe, J. Dostrovsky, The hippocampus as a spatial map. Preliminary evidence from unit activity in the freely moving rat, *Brain Res* 34 (1971) 171–175.
- [2] J. O'Keefe, M. L. Recce, Phase relationship between hippocampal place units and the EEG theta rhythm,

Hippocampus 3 (1993) 317–330.

- [3] J. Huxter, N. Burgess, J. O'Keefe, Independent rate and temporal coding in hippocampal pyramidal cells, *Nature* 425 (2003) 828–832.
- [4] W. E. Skaggs, B. L. McNaughton, M. A. Wilson, C. A. Barnes, Theta phase precession in hippocampal neuronal populations and the compression of temporal sequences, *Hippocampus* 6 (1996) 149–172.
- [5] K. D. Harris, D. A. Henze, H. Hirase, X. Leinekugel, G. Dragoi, A. Czurkó, G. Buzsáki, Spike train dynamics predicts theta-related phase precession in hippocampal pyramidal cells, *Nature* 417 (6890) (2002) 738–741.
- [6] O. Jensen, J. E. Lisman, Hippocampal CA3 region predicts memory sequences: accounting for the phase precession of place cells, *Learn Mem* 3 (1996) 279–287.
- [7] M. V. Tsodyks, W. E. Skaggs, T. J. Sejnowski, B. L. McNaughton, Population dynamics and theta phase precession of hippocampal place cell firing: a spiking neuron model, *Hippocampus* 6 (1996) 271–280.
- [8] G. V. Wallenstein, M. E. Hasselmo, GABAergic modulation of hippocampal activity: sequence learning, place field development, and the phase precession effect, *J Neurophysiol* 78 (1997) 393–408.
- [9] V. H. Brun, M. K. Otnass, S. Molden, H. A. Steffenach, M. P. Witter, M. B. Moser, E. I. Moser, Place cells and place recognition maintained by direct entorhinal-hippocampal circuitry, *Science* 296 (5576) (2002) 2243–2246.
- [10] K. Nakazawa, L. D. Sun, M. C. Quirk, L. Rondi-Reig, M. A. Wilson, S. Tonegawa, Hippocampal CA3 NMDA receptors are crucial for memory acquisition of one-time experience, *Neuron* 38 (2) (2003) 305–315.
- [11] M. Lengyel, Z. Szatmáry, P. Érdi, Dynamically detuned oscillations account for the coupled rate and temporal code of place cell firing, *Hippocampus* 13 (6) (2003) 700–714.
- [12] J. J. Chrobak, A. Lőrincz, G. Buzsáki, Physiological patterns in the hippocampo-entorhinal cortex system, *Hippocampus* 10 (4) (2000) 457–465.
- [13] G. Buzsáki, Theta oscillations in the hippocampus, *Neuron* 33 (3) (2002) 325–340.
- [14] J. Green, A. Arduini, Hippocampal electrical activity in arousal, *J Neurophysiol* 17 (1954) 533–557.
- [15] L. S. Leung, C. Y. Yim, Intracellular records of theta rhythm in hippocampal CA1 cells of the rat, *Brain Res* 367 (1-2) (1986) 323–327.
- [16] A. Kamondi, L. Acsády, X.-J. Wang, G. Buzsáki, Theta oscillation in somata and dendrites of hippocampal pyramidal cells in vivo: activity-dependent phase-precession of action potentials, *Hippocampus* 8 (1998) 244–261.

- [17] M. Lengyel, P. Érdi, Theta modulated feed-forward network generates rate and phase coded firing in the entorhino-hippocampal system, *IEEE Trans Neur Netw* in press.
- [18] P. F. Pinsky, J. Rinzel, Intrinsic and network rythmogenesis in a reduced Traub model for CA3 neurons, *J Comput Neurosci* 1 (1994) 39–60.
- [19] M. R. Mehta, A. K. Lee, M. A. Wilson, Role of experience and oscillations in transforming a rate code into a temporal code, *Nature* 417 (6890) (2002) 741–746.
- [20] Y. Yamaguchi, Y. Aota, B. L. McNaughton, P. Lipa, Bimodality of theta phase precession in hippocampal place cells in freely running rats, *J Neurophysiol* 87 (2002) 2639–2642.
- [21] A. Bose, V. Booth, M. Recce, A temporal mechanism for generating the phase precession of hippocampal place cells, *J Comput Neurosci* 9 (2000) 5–30.
- [22] V. Booth, A. Bose, Neural mechanisms for generating rate and temporal codes in model CA3 pyramidal cells, *J Neurophysiol* 85 (6) (2001) 2432–2445.
- [23] A. Bose, M. Recce, Phase precession and phase-locking of hippocampal pyramidal cells, *Hippocampus* 11 (3) (2001) 204–215.
- [24] J. O’Keefe, Place units in the hippocampus of the freely moving rat, *Exp Neurol* 51 (1976) 78–109.
- [25] B. L. McNaughton, C. A. Barnes, J. O’Keefe, The contributions of position, direction, and velocity to single unit activity in the hippocampus of freely-moving rats, *Exp Brain Res* 52 (1983) 41–49.

Biosketches



Zsófia Huhn (born in 1981, Szeged, Hungary) is studying Human-, and Neurobiology at Eötvös University of Sciences, Budapest. She has been working in Prof. Érdi’s CNS group since 2001. She is interested in modeling phase precession and other place cell related phenomena.



Gergő Orbán (born in 1977, Budapest, Hungary) received his M.Sc. degree in Molecular- and Biophysics in 2000 and is finishing his Ph.D. course in 2004 at Eötvös University of Sciences, Budapest, Hungary. He has been working in Prof. Érdi’s CNS group since 1998. He is interested in understanding physiological phenomena by means of developing suitable and biologically plausible models.



Máté Lengyel (born in 1975, Budapest, Hungary) received his Ph.D. in Neurobiology in 2003 at Eötvös University of Sciences, Budapest. He has been working in Prof. Érdi’s CNS group since 1995. His main motivation is to understand how the brain represents and stores abstract features of the external world or correlates of behavior, such as autobiographic memory or allocentric space.



Péter Érdi (born in 1946, Budapest, Hungary) received his Ph.D. in Chemistry in 1981. He is the head of Department of Biophysics of the KFKI Research Institute for Particle and Nuclear Physics of the Hungarian Academy of Sciences, and Henry R. Luce Professor at the Center for Complex Systems Studies, Kalamazoo College, MI. His main scientific interest is computational modeling of the functional organization of the nervous system and other complex systems.