

Single studies using the SelfControlledCaseSeries package

Martijn J. Schuemie, Marc A. Suchard and Patrick Ryan

2023-08-18

Contents

1	Introduction	1
2	Installation instructions	2
3	Overview	2
4	Studies with a single drug	3
4.1	Configuring the connection to the server	3
4.2	Preparing the exposure and outcome of interest	3
4.3	Extracting the data from the server	3
4.4	Creating the study population	5
4.5	Defining a simple model	5
4.6	Model fitting	6
4.7	Adding a pre-exposure window	6
4.8	Including seasonality, and calendar time	7
4.9	Removing the COVID blip	11
4.10	Considering event-dependent observation time	13
5	Studies with more than one drug	13
5.1	Adding a class of drugs	14
5.2	Adding all drugs	15
6	Diagnostics	17
6.1	Power calculations	17
6.2	Time from exposure start to event	17
6.3	Increased risk pre-exposure	17
6.4	Ages covered per subject	18
6.5	Dependency between events and observation end	18
6.6	Stability of the outcome over calendar time	19
7	Acknowledgments	20

1 Introduction

This vignette describes how you can use the `SelfControlledCaseSeries` package to perform a single Self-Controlled Case Series (SCCS) study. We will walk through all the steps needed to perform an exemplar study, and we have selected the well-studied topic of the effect of aspirin on epistaxis. `## Terminology`

The following terms are used consistently throughout the package:

- **Case:** a period of continuous observation of a person containing one or more outcomes. One person can contribute to multiple cases, e.g. when the person has multiple observation periods.

- **Observation period:** the time when a person is observed in the database, e.g. the time when enrolled in an insurance plan.
- **Nesting cohort:** a cohort defining when persons are eligible to be included as cases. This is typically the indication of the drug. For example, if the drug treats diabetes, we may want to nest the analysis in the time when people have diabetes to avoid (time-varying) confounding by indication.
- **Study period:** a period of calendar time when persons are eligible to be included as cases. For example, a study period could be January 1, 2020, onward. Unlink nesting cohorts, which could specify different time periods per person, a study period applies to all persons.
- **Naive period:** the first part of a person's observation period (e.g. the first 180 days). This is typically removed from the SCCS model to avoid exposure and incident outcome misclassification. For example, if we observe the outcome on the second day of a person's observation period, we do not know whether the outcome is a new one or just a follow-up for an old one, and whether the patient may have started the exposure just prior to observation start without us knowing about it.
- **Study population:** the set of cases having a specific outcome, that meet certain criteria, such as the naive period and age restrictions.
- **Era:** a data element extracted from the database denoting a time period for a patient (a case). This can be a cohort, but also a drug era.
- **SCCS data:** a data object containing information on cases, observation periods, nesting cohorts, and eras.
- **Covariate:** a time-varying variable used as a predictor for the outcome in the SCCS model. These include splines and era covariates.
- **Era covariate:** a covariate derived from an era. Multiple covariates can be derived from a single era (e.g. a covariate for when on a drug, and a covariate for the pre-exposure time prior to the drug). One covariate can be derived from multiple eras (e.g. one covariate can represent a class of drugs).
- **SCCS interval data:** data of cases chopped into intervals during which all covariates have a constant value. For splines (e.g for season), the effect is assumed to be constant within each calendar month.
- **SCCS model:** a Poisson regression of SCCS interval data, condition on the observation period.

2 Installation instructions

Before installing the `SelfControlledCaseSeries` package make sure you have Java available. For Windows users, RTools is also necessary. See these instructions for properly configuring your R environment.

The `SelfControlledCaseSeries` package is maintained in a Github repository, and can be downloaded and installed from within R using the `remotes` package:

```
install.packages("remotes")
library(remotes)
install_github("ohdsi/SelfControlledCaseSeries")
```

Once installed, you can type `library(SelfControlledCaseSeries)` to load the package.

3 Overview

In the `SelfControlledCaseSeries` package a study requires at least three steps:

1. Loading the necessary data from the database.
2. Transforming the data into a format suitable for an SCCS study. This step can be broken down in multiple tasks:
 - Defining a study population, people having a specific outcome, with possible further restrictions such as age.
 - Creating covariates based on the variables extracted from the database, such as defining risk windows based on exposures.

- Transforming the data into non-overlapping time intervals, with information on the various covariates and outcomes per interval.
3. Fitting the model using conditional Poisson regression.

In the following sections these steps will be demonstrated for increasingly complex studies.

4 Studies with a single drug

4.1 Configuring the connection to the server

We need to tell R how to connect to the server where the data are. `SelfControlledCaseSeries` uses the `DatabaseConnector` package, which provides the `createConnectionDetails` function. Type `?createConnectionDetails` for the specific settings required for the various database management systems (DBMS). For example, one might connect to a PostgreSQL database using this code:

```
connectionDetails <- createConnectionDetails(dbms = "postgresql",
                                             server = "localhost/ohdsi",
                                             user = "joe",
                                             password = "supersecret")

cdmDatabaseSchema <- "my_cdm_data"
cohortDatabaseSchema <- "my_results"
cohortTable <- "my_cohorts"
options(sqlRenderTempEmulationSchema = NULL)
```

The last three lines define the `cdmDatabaseSchema` and `cohortDatabaseSchema` variables, as well as the CDM version. We'll use these later to tell R where the data in CDM format live, where we have stored our cohorts of interest, and what version CDM is used. Note that for Microsoft SQL Server, 'databaseSchemas' need to specify both the database and the schema, so for example `cdmDatabaseSchema <- "my_cdm_data.dbo"`.

4.2 Preparing the exposure and outcome of interest

We need to define the exposure and outcomes for our study. For the exposure, we will directly use the `drug_era` table in the CDM. For the outcome we can use the OHDSI `PhenotypeLibrary` to retrieve a community-approved definition of epistaxis:

```
epistaxis <- 356
cohortDefinitionSet <- PhenotypeLibrary::getPlCohortDefinitionSet(epistaxis)
```

We can use the `CohortGenerator` package to instantiate this cohort:

```
connection <- DatabaseConnector::connect(connectionDetails)
cohortTableNames <- CohortGenerator::getCohortTableNames(cohortTable)
CohortGenerator::createCohortTables(connection = connection,
                                    cohortDatabaseSchema = cohortDatabaseSchema,
                                    cohortTableNames = cohortTableNames)

counts <- CohortGenerator::generateCohortSet(connection = connection,
                                             cdmDatabaseSchema = cdmDatabaseSchema,
                                             cohortDatabaseSchema = cohortDatabaseSchema,
                                             cohortTableNames = cohortTableNames,
                                             cohortDefinitionSet = cohortDefinitionSet)

DatabaseConnector::disconnect(connection)
```

4.3 Extracting the data from the server

Now we can tell `SelfControlledCaseSeries` to extract all necessary data for our analysis:

```
aspirin <- 1112807
```

```
sccsData <- getDbSccsData(connectionDetails = connectionDetails,  
                          cdmDatabaseSchema = cdmDatabaseSchema,  
                          outcomeDatabaseSchema = cohortDatabaseSchema,  
                          outcomeTable = outcomeTable,  
                          outcomeIds = epistaxis,  
                          exposureDatabaseSchema = cdmDatabaseSchema,  
                          exposureTable = "drug_era",  
                          exposureIds = aspirin,  
                          studyStartDates = "20100101",  
                          studyEndDates = "21000101")
```

```
sccsData
```

```
## # SccsData object
```

```
##
```

```
## Exposure cohort ID(s): 1112807
```

```
## Outcome cohort ID(s): 356
```

```
##
```

```
## Inherits from Andromeda:
```

```
## # Andromeda object
```

```
## # Physical location: D:\andromedaTemp\file15bc11a5520f.sqlite
```

```
##
```

```
## Tables:
```

```
## $cases (observationPeriodId, caseId, personId, noninformativeEndCensor, observationPeriodStartDate, ...)
```

```
## $eraRef (eraType, eraId, eraName, minObservedDate, maxObservedDate)
```

```
## $eras (eraType, caseId, eraId, eraValue, eraStartDay, eraEndDay)
```

There are many parameters, but they are all documented in the `SelfControlledCaseSeries` manual. In short, we are pointing the function to the table created earlier and indicating which cohort ID in that table identifies the outcome. Note that it is possible to fetch the data for multiple outcomes at once. We further point the function to the `drug_era` table, and specify the concept ID of our exposure of interest: `aspirin`. Again, note that it is also possible to fetch data for multiple drugs at once. In fact, when we do not specify any exposure IDs the function will retrieve the data for all the drugs found in the `drug_era` table.

All data about the patients, outcomes and exposures are extracted from the server and stored in the `sccsData` object. This object uses the `Andromeda` package to store information in a way that ensures R does not run out of memory, even when the data are large.

We can use the generic `summary()` function to view some more information of the data we extracted:

```
summary(sccsData)
```

```
## SccsData object summary
```

```
##
```

```
## Exposure cohort ID(s): 1112807
```

```
## Outcome cohort ID(s): 356
```

```
##
```

```
## Outcome counts:
```

```
##      Outcome Subjects Outcome Events Outcome Observation Periods
```

```
## 356           99702           157218           1e+05
```

```
##
```

```
## Eras:
```

```
## Number of era types: 2
```

```
## Number of eras: 168013
```

4.3.1 Saving the data to file

Creating the `sccsData` file can take considerable computing time, and it is probably a good idea to save it for future sessions. Because `sccsData` uses `Andromeda`, we cannot use R's regular save function. Instead, we'll have to use the `saveSccsData()` function:

```
saveSccsData(sccsData, "sccsData.zip")
```

We can use the `loadSccsData()` function to load the data in a future session.

4.4 Creating the study population

From the data fetched from the server we can now define the population we wish to study. If we retrieved data for multiple outcomes, we should now select only one, and possibly impose further restrictions:

```
studyPop <- createStudyPopulation(sccsData = sccsData,
                                outcomeId = epistaxis,
                                firstOutcomeOnly = FALSE,
                                naivePeriod = 180)
```

Here we specify we wish to study the outcome with the ID stored in `epistaxis`. Since this was the only outcome for which we fetched the data, we could also have skipped this argument. We furthermore specify that the first 180 days of observation of every person, the so-called 'naive period', will be excluded from the analysis. Note that data in the naive period will be used to determine exposure status at the start of follow-up (after the end of the naive period). We also specify we will use all occurrences of the outcome, not just the first one per person.

We can find out how many people (if any) were removed by any restrictions we imposed:

```
getAttritionTable(studyPop)
```

##	outcomeId	outcomeSubjects	outcomeEvents	outcomeObsPeriods	observedDays	descrip
## 1	356	882260	1386052	908691	3367212134	All outcome occurrences
## 2	356	779787	1221258	797988	2844636018	Outcomes in study period
## 3	356	99702	157218	100000	367899811	Random sample
## 4	356	89269	138585	89470	320619597	Requiring 180 days naive period

4.5 Defining a simple model

Next, we can use the data to define a simple model to fit:

```
covarAspirin <- createEraCovariateSettings(label = "Exposure of interest",
                                           includeEraIds = aspirin,
                                           start = 0,
                                           end = 0,
                                           endAnchor = "era end")
```

```
sccsIntervalData <- createSccsIntervalData(
  studyPopulation = studyPop,
  sccsData = sccsData,
  eraCovariateSettings = covarAspirin
)
```

```
summary(sccsIntervalData)
```

```
## SccsIntervalData object summary
##
## Outcome cohort ID: 356
```

```
##
## Number of cases (observation periods): 3609
## Number of eras (spans of time): 7218
## Number of outcomes: 5808
## Number of covariates: 1
## Number of non-zero covariate values: 3609
```

In this example, we use the `createEraCovariateSettings` to define a single covariate: exposure to aspirin. We specify that the risk window is from start of exposure to the end by setting start and end to 0, and defining the anchor for the end to be the era end, which for drug eras is the end of exposure.

We then use the covariate definition in the `createSccsIntervalData` function to generate the `sccsIntervalData`. This represents the data in non-overlapping time intervals, with information on the various covariates and outcomes per interval.

4.6 Model fitting

The `fitSccsModel` function is used to fit the model:

```
model <- fitSccsModel(sccsIntervalData)
```

We can inspect the resulting model:

```
model

## SccsModel object
##
## Outcome ID: 356
##
## Outcome count:
##      outcomeSubjects outcomeEvents outcomeObsPeriods observedDays
## 356                3609           5808                3609      9511023
##
## Estimates:
## # A tibble: 1 x 7
##   Name                      ID Estimate LB95CI UB95CI LogRr SeLogRr
##   <chr>                   <dbl>    <dbl>  <dbl>  <dbl> <dbl>  <dbl>
## 1 Exposure of interest  1000      1.48   1.38   1.60 0.395  0.0378
```

This tells us what the estimated relative risk (the incidence rate ratio) is during exposure to aspirin compared to non-exposed time.

4.7 Adding a pre-exposure window

The fact that platelet aggregation inhibitors like aspirin can cause epistaxis is well known to doctors, and this knowledge affects prescribing behavior. For example, a patient who has just had a nose bleed is not likely to be prescribed aspirin. This may lead to underestimation of the rate during unexposed time, because the unexposed time includes time just prior to exposure where observing of the outcome is unlikely because of this behavior. One solution to this problem that is often used is to introduce a separate ‘risk window’ just prior to exposure, to separate it from the remaining unexposed time. We can add such a ‘pre-exposure window’ to our analysis:

```
covarPreAspirin <- createEraCovariateSettings(label = "Pre-exposure",
                                              includeEraIds = aspirin,
                                              start = -60,
                                              end = -1,
                                              endAnchor = "era start")
```

```

sccsIntervalData <- createSccsIntervalData(
  studyPopulation = studyPop,
  sccsData = sccsData,
  eraCovariateSettings = list(covarAspirin,
                              covarPreAspirin)
)

model <- fitSccsModel(sccsIntervalData)

```

Here we created a new covariate definition in addition to the first one. We define the risk window to start 60 days prior to exposure, and end on the day just prior to exposure. We combine the two covariate settings in a list for the `createSccsIntervalData` function. Again, we can take a look at the results:

```

model

## SccsModel object
##
## Outcome ID: 356
##
## Outcome count:
##      outcomeSubjects outcomeEvents outcomeObsPeriods observedDays
## 356                3609           5808                3609      9511023
##
## Estimates:
## # A tibble: 2 x 7
##   Name                ID Estimate LB95CI UB95CI LogRr SeLogRr
##   <chr>              <dbl>    <dbl> <dbl> <dbl> <dbl> <dbl>
## 1 Exposure of interest 1000      1.54  1.43  1.66 0.432  0.0385
## 2 Pre-exposure        1001      1.32  1.19  1.46 0.279  0.0531

```

4.8 Including seasonality, and calendar time

Often both the rate of exposure and the outcome change with age, and can even depend on the season or calendar time in general (e.g. rates may be higher in 2021 compared to 2020). This may lead to confounding and may bias our estimates. To correct for this we can include age, season, and/or calendar time into the model.

For computational reasons we assume the effect of age, season, and calendar time are constant within each calendar month. We assume that the rate from one month to the next can be different, but we also assume that subsequent months have somewhat similar rates. This is implemented by using cubic spline functions.

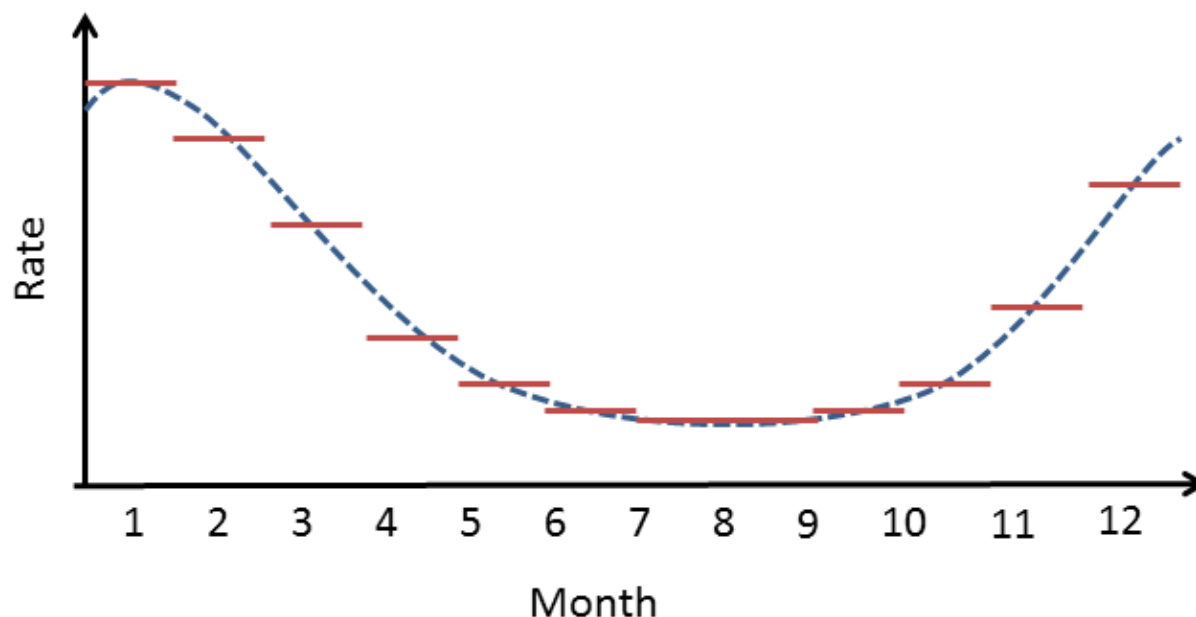


Figure 1. Example of how a spline is used for seasonality: within a month, the risk attributable to seasonality is assumed to be constant, but from month to month the risks are assumed to follow a cyclic cubic spline.

Note that the by default all people that have the outcome will be used to estimate the effect of age, seasonality, and calendar time on the outcome, so not just the people exposed to the drug of interest. Adjusting for age typically only makes sense for small children, where a small difference in age can still make a big difference (remember: we are modeling the effect of age in each person by themselves, not the effect of age across persons). Since age and calendar time are often hard to fit simultaneously, it is often best to only model seasonality, and calendar time, like this:

```
seasonalityCovariateSettings <- createSeasonalityCovariateSettings(seasonKnots = 5)

calendarTimeSettings <- createCalendarTimeCovariateSettings(calendarTimeKnots = 5)

sccsIntervalData <- createSccsIntervalData(
  studyPopulation = studyPop,
  sccsData = sccsData,
  eraCovariateSettings = list(covarAspirin,
                             covarPreAspirin),
  seasonalityCovariateSettings = seasonalityCovariateSettings,
  calendarTimeCovariateSettings = calendarTimeSettings
)

model <- fitSccsModel(sccsIntervalData)
```

Again, we can inspect the model:

```
model

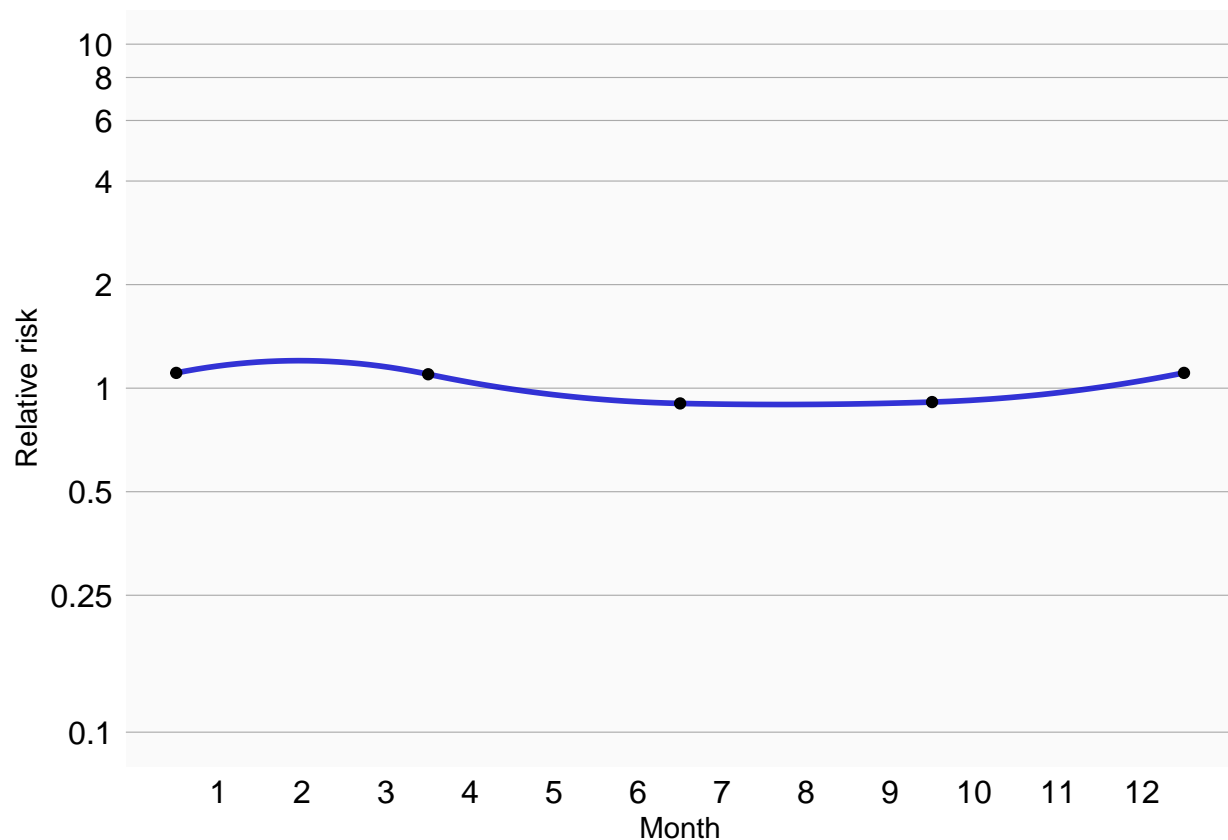
## SccsModel object
##
## Outcome ID: 356
##
## Outcome count:
##   outcomeSubjects outcomeEvents outcomeObsPeriods observedDays
## 356           13242           20596           13244       30396567
```



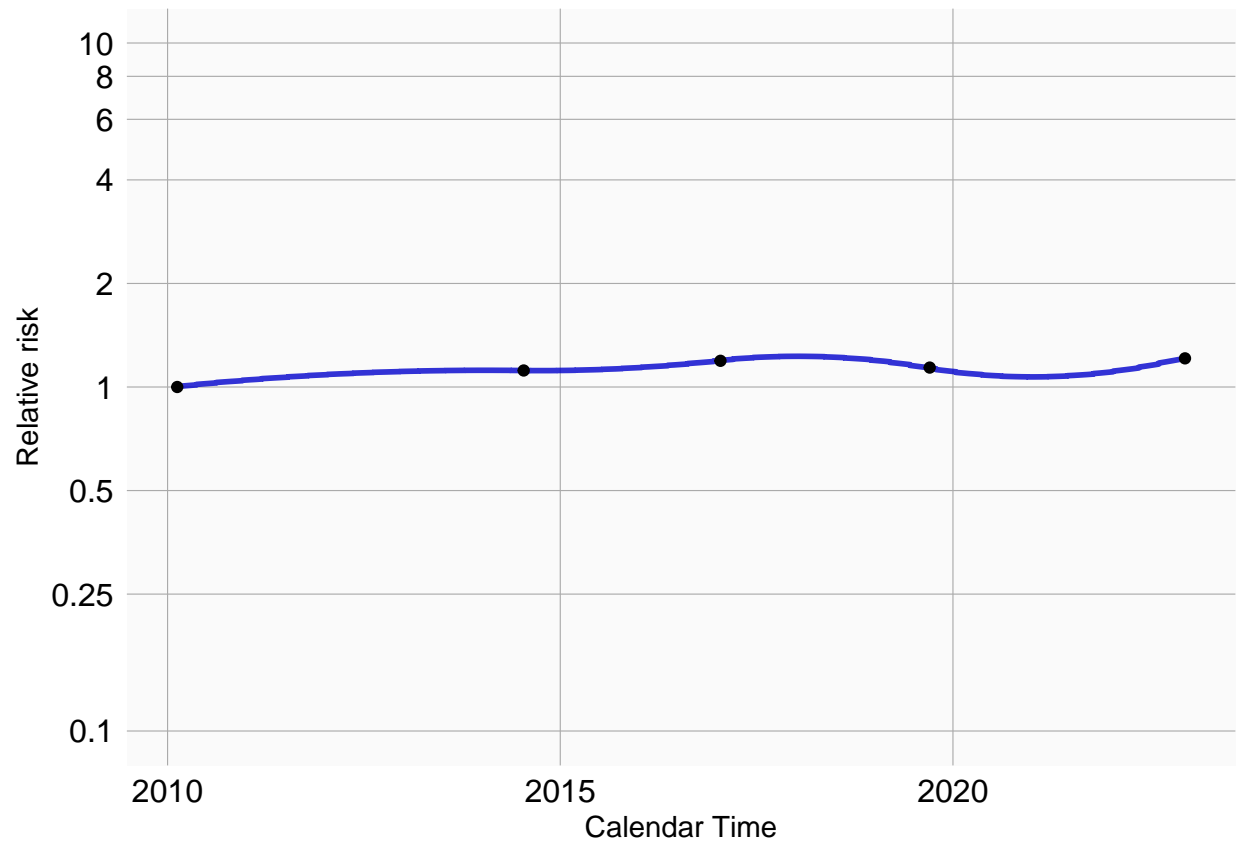
```
##
## Estimates:
## # A tibble: 11 x 7
##   Name                                ID Estimate LB95CI UB95CI   LogRr SeLogRr
##   <chr>                                <dbl>   <dbl>   <dbl>   <dbl>   <dbl>   <dbl>
## 1 Seasonality spline component 1      200    1.02    NA      NA     0.0228    NA
## 2 Seasonality spline component 2      201    1.43    NA      NA     0.359     NA
## 3 Seasonality spline component 3      202    1.00    NA      NA     0.00464   NA
## 4 Seasonality spline component 4      203    0.969   NA      NA    -0.0319   NA
## 5 Calendar time spline component 1     300    1.13    NA      NA     0.125     NA
## 6 Calendar time spline component 2     301    1.11    NA      NA     0.102     NA
## 7 Calendar time spline component 3     302    1.29    NA      NA     0.253     NA
## 8 Calendar time spline component 4     303    0.980   NA      NA    -0.0205   NA
## 9 Calendar time spline component 5     304    1.21    NA      NA     0.192     NA
## 10 Exposure of interest               1000    1.54    1.43    1.66    0.431     0.0385
## 11 Pre-exposure                       1001    1.32    1.19    1.46    0.278     0.0531
```

We see that our estimates for exposed and pre-exposure time have not changes much. We can plot the spline curves for season, and calendar time to learn more:

```
plotSeasonality(model)
```

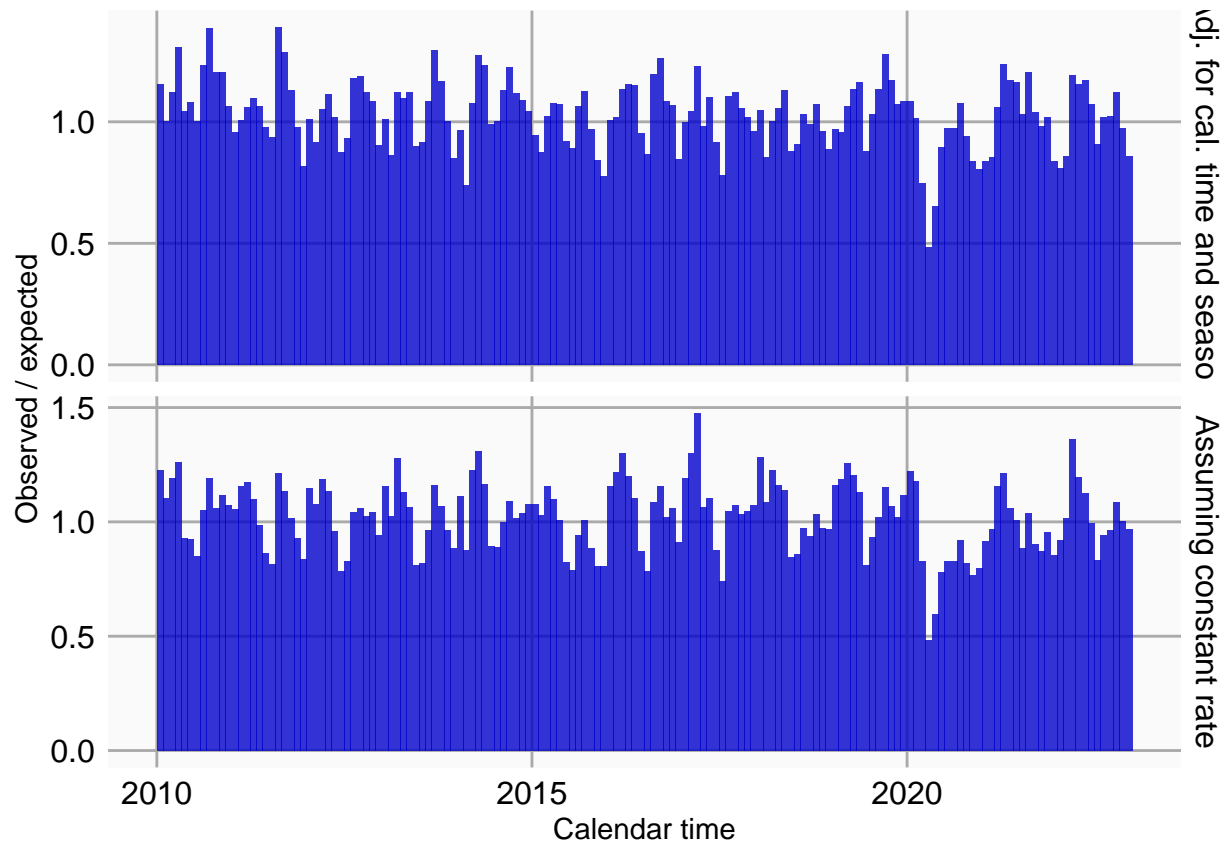


```
plotCalendarTimeEffect(model)
```



We see some effect for season: epistaxis tends to be more prevalent during winter. We should verify if our model accounts for time trends, by plotting the rate of the outcome across time, before and after adjustment:

```
plotEventToCalendarTime(studyPopulation = studyPop,  
                        sccsModel = model)
```



Here we see that after adjustment, in general, the rate of outcome appears fairly stable across time, with the exception of some months in 2020. This is what we refer to as the ‘COVID-blip’, because during the time of the COVID-19 pandemic regular healthcare was put on hold. Our time trend diagnostic confirms the rate of the outcome was significantly lower during these months:

```
stability <- computeTimeStability(studyPopulation = studyPop,
                                  sccsModel = model)
stability %>%
  filter(!stable)
```

```
## # A tibble: 1 x 3
##   ratio    p stable
##   <dbl> <dbl> <lgl>
## 1  1.12     1 TRUE
```

4.9 Removing the COVID blip

The discontinuity in the rate of the outcome during the COVID pandemic could cause bias. For this reason it is probably best to remove this time from the analysis altogether. We can achieve this by defining two separate study periods:

```
sccsData <- getDbSccsData(connectionDetails = connectionDetails,
                          cdmDatabaseSchema = cdmDatabaseSchema,
                          outcomeDatabaseSchema = cohortDatabaseSchema,
                          outcomeTable = cohortTable,
                          outcomeIds = epistaxis,
                          exposureDatabaseSchema = cdmDatabaseSchema,
                          exposureTable = "drug_era",
```

```

      exposureIds = aspirin,
      studyStartDates = c("19000101", "20220101"),
      studyEndDates = c("20191231", "21001231"))
studyPop <- createStudyPopulation(sccsData = sccsData,
                                outcomeId = epistaxis,
                                firstOutcomeOnly = FALSE,
                                naivePeriod = 180)
sccsIntervalData <- createSccsIntervalData(
  studyPopulation = studyPop,
  sccsData = sccsData,
  eraCovariateSettings = list(covarAspirin,
                             covarPreAspirin),
  seasonalityCovariateSettings = seasonalityCovariateSettings,
  calendarTimeCovariateSettings = calendarTimeSettings
)
model <- fitSccsModel(sccsIntervalData)

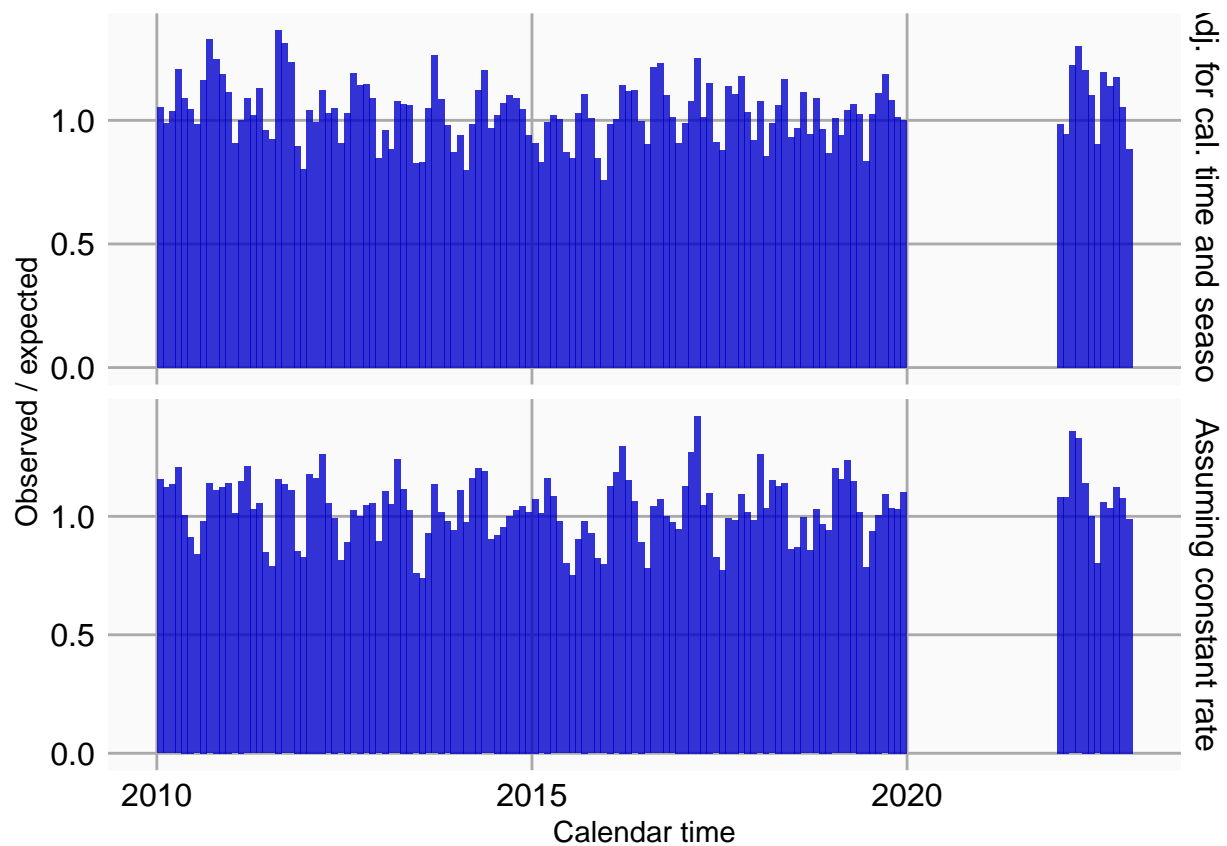
```

If we plot the outcomes over time, we see the entire time of the COVID pandemic has been removed:

```

plotEventToCalendarTime(studyPopulation = studyPop,
                        sccsModel = model)

```



We can verify if this improved temporal stability:

```

stability <- computeTimeStability(studyPopulation = studyPop,
                                sccsModel = model)
stability %>%

```

```

filter(!stable)

## # A tibble: 1 x 3
##   ratio      p stable
##   <dbl> <dbl> <lgl>
## 1  1.10      1 TRUE

```

4.10 Considering event-dependent observation time

The SCCS method requires that observation periods are independent of outcome times. This requirement is violated when outcomes increase the mortality rate, since censoring of the observation periods is then event-dependent. A modification to the SCCS has been proposed that attempts to correct for this. First, several models are fitted to estimate the amount and shape of the event-dependent censoring, and the best fitting model is selected. Next, this model is used to reweigh various parts of the observation time. This approach is also implemented in this package, and can be turned on using the `eventDependentObservation` argument of the `createSccsIntervalData` function. However, this method has proven to be somewhat unstable in combinations with other corrections, so for now it is best to keep the model simple:

```

sccsIntervalData <- createSccsIntervalData(
  studyPopulation = studyPop,
  sccsData = sccsData,
  eraCovariateSettings = list(covarAspirin,
                              covarPreAspirin),
  eventDependentObservation = TRUE
)

model <- fitSccsModel(sccsIntervalData)

```

Again, we can inspect the model:

```

model

## SccsModel object
##
## Outcome ID: 356
##
## Outcome count:
##   outcomeSubjects outcomeEvents outcomeObsPeriods observedDays
## 356             2910           4504             2923       2167829
##
## Estimates:
## # A tibble: 2 x 7
##   Name                ID Estimate LB95CI UB95CI LogRr SeLogRr
##   <chr>              <dbl>    <dbl> <dbl> <dbl> <dbl>  <dbl>
## 1 Exposure of interest 1000      1.47  1.35  1.60 0.387  0.0434
## 2 Pre-exposure         1001      1.42  1.27  1.58 0.348  0.0557

```

5 Studies with more than one drug

Although we are usually interested in the effect of a single drug or drug class, it could be beneficial to add exposure to other drugs to the analysis if we believe those drugs represent time-varying confounders that we wish to correct for.

5.1 Adding a class of drugs

For example, SSRIs might also cause epistaxis, and if aspirin is co-prescribed with SSRIs we don't want the effect of SSRIs attributed to aspirin. We would like our estimate to represent just the effect of the aspirin, so we need to keep the effect of the SSRIs separate. First we have to retrieve the information on SSRI exposure from the database:

```
ssris <- c(715939, 722031, 739138, 751412, 755695, 797617, 40799195)
sccsData <- getDbSccsData(connectionDetails = connectionDetails,
  cdmDatabaseSchema = cdmDatabaseSchema,
  outcomeDatabaseSchema = cohortDatabaseSchema,
  outcomeTable = cohortTable,
  outcomeIds = epistaxis,
  maxCasesPerOutcome = 100000,
  exposureDatabaseSchema = cdmDatabaseSchema,
  exposureTable = "drug_era",
  exposureIds = c(aspirin, ssris),
  studyStartDates = c("19000101", "20220101"),
  studyEndDates = c("20191231", "21001231"))
```

sccsData

```
## # SccsData object
##
## Exposure cohort ID(s): 1112807,715939,722031,739138,751412,755695,797617,40799195
## Outcome cohort ID(s): 356
##
## Inherits from Andromeda:
## # Andromeda object
## # Physical location: D:\andromedaTemp\file15bc79745009.sqlite
##
## Tables:
## $cases (observationPeriodId, caseId, personId, noninformativeEndCensor, observationPeriodStartDate, ...)
## $eraRef (eraType, eraId, eraName, minObservedDate, maxObservedDate)
## $eras (eraType, caseId, eraId, eraValue, eraStartDay, eraEndDay)
```

Once retrieved, we can use the data to build and fit our model:

```
studyPop <- createStudyPopulation(sccsData = sccsData,
  outcomeId = epistaxis,
  firstOutcomeOnly = FALSE,
  naivePeriod = 180)
covarSsris <- createEraCovariateSettings(label = "SSRIs",
  includeEraIds = ssris,
  stratifyById = FALSE,
  start = 1,
  end = 0,
  endAnchor = "era end")
sccsIntervalData <- createSccsIntervalData(
  studyPopulation = studyPop,
  sccsData = sccsData,
  eraCovariateSettings = list(covarAspirin,
    covarPreAspirin,
    covarSsris),
  seasonalityCovariateSettings = seasonalityCovariateSettings,
  calendarTimeCovariateSettings = calendarTimeSettings
)
```

```
model <- fitSccsModel(sccsIntervalData)
```

Here, we added a new covariate based on the list of concept IDs for the various SSRIs. In this example we set `stratifyById` to `FALSE`, meaning that we will estimate a single incidence rate ratio for all SSRIs, so one estimate for the entire class of drugs. Note that duplicates will be removed: if a person is exposed to two SSRIs on the same day, this will be counted only once when fitting the model. Again, we can inspect the model:

```
model

## SccsModel object
##
## Outcome ID: 356
##
## Outcome count:
##      outcomeSubjects outcomeEvents outcomeObsPeriods observedDays
## 356              21185             31914             21293       39699113
##
## Estimates:
## # A tibble: 13 x 7
##   Name                                     ID Estimate LB95CI UB95CI   LogRr SeLogRr
##   <chr>                                <dbl>   <dbl>   <dbl>   <dbl>   <dbl>   <dbl>
## 1 Seasonality spline component 1         200     1.03    NA      NA     0.0274    NA
## 2 Seasonality spline component 2         201     1.45    NA      NA     0.372     NA
## 3 Seasonality spline component 3         202     1.02    NA      NA     0.0203    NA
## 4 Seasonality spline component 4         203     0.956   NA      NA    -0.0445    NA
## 5 Calendar time spline component 1        300     1.04    NA      NA     0.0345    NA
## 6 Calendar time spline component 2        301     0.904   NA      NA    -0.101     NA
## 7 Calendar time spline component 3        302     0.986   NA      NA    -0.0142    NA
## 8 Calendar time spline component 4        303     0.942   NA      NA    -0.0593    NA
## 9 Calendar time spline component 5        304     0.996   NA      NA    -0.00365   NA
## 10 Calendar time spline component 6       305     1.01    NA      NA     0.00886   NA
## 11 Exposure of interest                   1000     1.58     1.46    1.72    0.459     0.0420
## 12 Pre-exposure                           1001     1.31     1.17    1.46    0.271     0.0564
## 13 SSRIs                                 1002     1.21     1.15    1.26    0.188     0.0227
```

5.2 Adding all drugs

Another approach could be to add all drugs into the model. Again, the first step is to get all the relevant data from the database:

```
sccsData <- getDbSccsData(connectionDetails = connectionDetails,
                           cdmDatabaseSchema = cdmDatabaseSchema,
                           outcomeDatabaseSchema = cohortDatabaseSchema,
                           outcomeTable = outcomeTable,
                           outcomeIds = 1,
                           exposureDatabaseSchema = cdmDatabaseSchema,
                           exposureTable = "drug_era",
                           exposureIds = c(),
                           cdmVersion = cdmVersion)
```

Note that the `exposureIds` argument is left empty. This will cause data for all concepts in the exposure table to be retrieved. Next, we simply create a new set of covariates, and fit the model:

```
studyPop <- createStudyPopulation(sccsData = sccsData,
                                  outcomeId = 1,
```

```

      firstOutcomeOnly = FALSE,
      naivePeriod = 180)

covarAllDrugs <- createEraCovariateSettings(label = "Other exposures",
      excludeEraIds = aspirin,
      stratifyById = TRUE,
      start = 1,
      end = 0,
      endAnchor = "era end",
      allowRegularization = TRUE)

sccsIntervalData <- createSccsIntervalData(
  studyPopulation = studyPop,
  sccsData = sccsData,
  eraCovariateSettings = list(covarAspirin,
                             covarPreAspirin,
                             covarAllDrugs),
  seasonalityCovariateSettings = seasonalityCovariateSettings,
  calendarTimeCovariateSettings = calendarTimeSettings
)

model <- fitSccsModel(sccsIntervalData)

```

The first thing to note is that we have defined the new covariates to be all drugs except aspirin by not specifying the `includeEraIds` and setting the `excludeEraIds` to the concept ID of aspirin. Furthermore, we have specified that `stratifyById` is `TRUE`, meaning an estimate will be produced for each drug.

We have set `allowRegularization` to `TRUE`, meaning we will use regularization for all estimates in this new covariate set. Regularization means we will impose a prior distribution on the effect size, effectively penalizing large estimates. This helps fit the model, for example when some drugs are rare, and when drugs are almost often prescribed together and their individual effects are difficult to untangle.

Because there are now so many estimates, we will export all estimates to a data frame using `getModel()`:

```

estimates <- getModel(model)
estimates[estimates$originalEraId == aspirin, ]

```

```

## # A tibble: 0 x 10
## # i 10 variables: name <chr>, id <dbl>, estimate <dbl>, lb95Ci <dbl>, ub95Ci <dbl>, logRr <dbl>, seLogRr <dbl>, originalEraId <dbl>, originalEraName <chr>

```

Here we see that despite the extensive adjustments that are made in the model, the effect estimates for aspirin have remained nearly the same.

In case we're interested, we can also look at the effect sizes for the SSRIs:

```

estimates[estimates$originalEraId %in% ssris, ]

```

```

## # A tibble: 4 x 10
##   name                                id estimate lb95Ci ub95Ci  logRr seLogRr originalEraId originalEraName
##   <chr>                                <dbl>   <dbl>   <dbl>   <dbl>   <dbl>   <dbl>   <dbl>   <chr>
## 1 Other exposures: citalopram         1148     1.10    NA      NA  0.0929    NA      797617 rx
## 2 Other exposures: sertraline         1470     1.10    NA      NA  0.0961    NA      739138 rx
## 3 Other exposures: escitalopram       1850     1.05    NA      NA  0.0484    NA      715939 rx
## 4 Other exposures: fluoxetine         2128     1.19    NA      NA  0.171     NA      755695 rx

```

Note that because we used regularization, we are not able to compute the confidence intervals for these estimates.

6 Diagnostics

We can perform several diagnostics on the data to verify whether our assumptions underlying the SCCS are met.

6.1 Power calculations

We might be interested to know whether we have sufficient power to detect a particular effect size. It makes sense to perform these power calculations once the study population has been fully defined, so taking into account loss to the various inclusion and exclusion criteria. This means we will use the `sccsIntervalData` object we've just created as the basis for our power calculations. Since the sample size is fixed in retrospective studies (the data has already been collected), and the true effect size is unknown, the `SelfControlledCaseSeries` package provides a function to compute the minimum detectable relative risk (MDRR) instead:

```
computeMdrd(sccsIntervalData,
             exposureCovariateId = 1000,
             alpha = 0.05,
             power = 0.8,
             twoSided = TRUE,
             method = "binomial")
```

```
## # A tibble: 1 x 5
##   timeExposed timeTotal propTimeExposed events  mdrd
##       <dbl>      <int>          <dbl> <int> <dbl>
## 1    2038397  11069291            0.184   5808  1.10
```

Note that we have to provide the covariate ID of the exposure of interest, which we learned by calling `summary` on `sccsIntervalData` earlier. This is because we may have many covariates in our model, but will likely only be interested in the MDRR of one.

6.2 Time from exposure start to event

To gain a better understanding of when the event occurs relative to the start of exposure, we can plot their relationship. Note that we specify the naive period, so this can be applied to the data prior to showing the plot. This will make the plot better in line with the data we ended up fitting:

```
plotExposureCentered(studyPop, sccsData, exposureEraId = aspirin)
```

```
## Warning in computeTimeToEvent(studyPopulation, sccsData, exposureEraId): No exposures found with era
## Warning: There were 2 warnings in `summarise()`.
## The first warning was:
## i In argument: `eraStartDay = min(.data$eraStartDay, na.rm = TRUE)` .
## Caused by warning in `min()` :
## ! no non-missing arguments to min; returning Inf
## i Run `dplyr::last_dplyr_warnings()` to see the 1 remaining warning.
## NULL
```

This plot suggests an increased rate of events in the first few weeks following the start of exposure, perhaps because of an acute effect.

6.3 Increased risk pre-exposure

If the rate just before exposure is higher compared to during exposure, this indicates there might reverse causality: that the outcome, or some precursor of the outcome, increases the probability of having the exposure. To avoid incorrect causal inference, we want to detect these situations. We can compute a p-value for whether rate is increased pre-exposure:

```
computePreExposureGainP(sccsData, studyPopulation, exposureEraId = aspirin)
```

```
## Warning in computePreExposureGainP(sccsData, studyPop, exposureEraId = aspirin): No exposures found v
## [1] NA
```

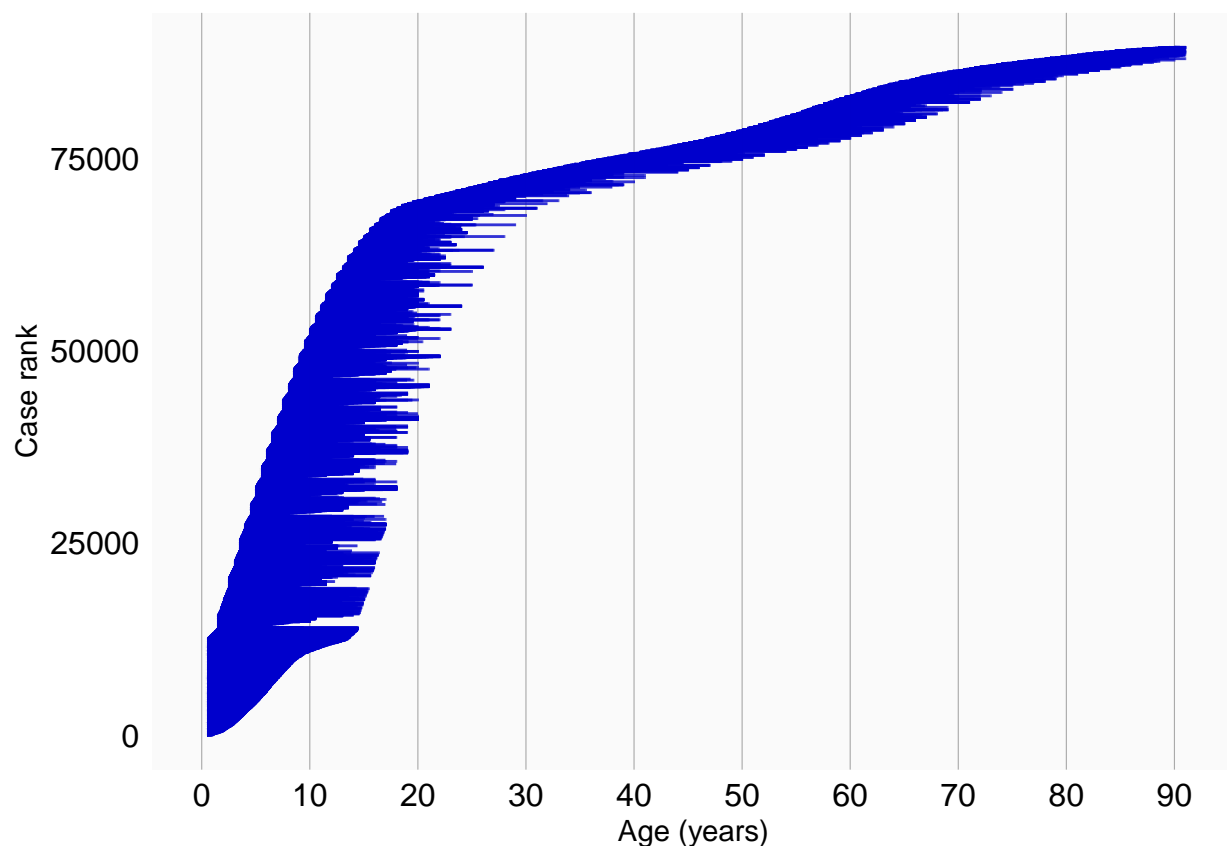
If this p-value is lower than a prespecified threshold (e.g. $p < 0.05$), we can decide to reject the null hypothesis that the rate is not increased, and we probably should not trust our analysis to produce reliable estimates.

6.4 Ages covered per subject

We can visualize which age ranges are covered by each subject's observation time:

```
plotAgeSpans(studyPop)
```

```
## Warning in plotAgeSpans(studyPop): There are 89470 cases. Random sampling 10000 cases.
```

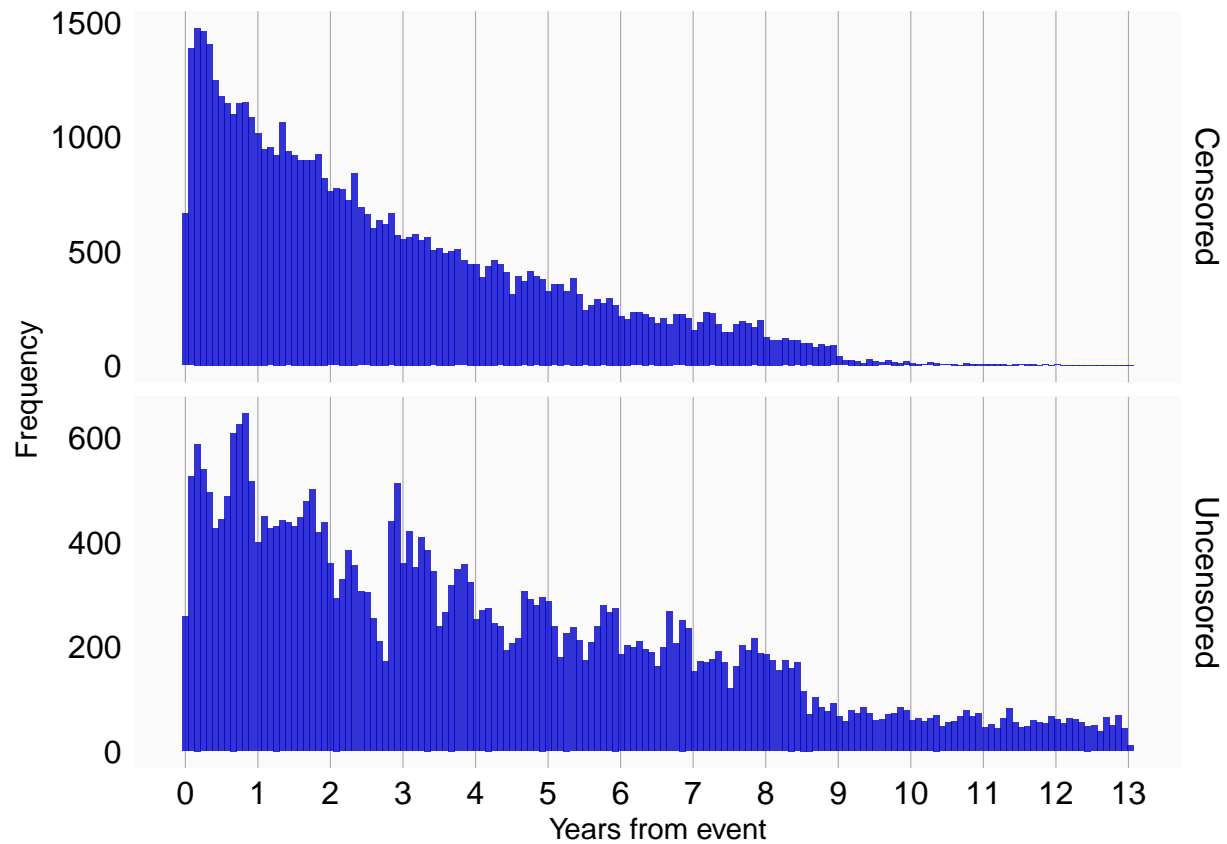


Here we see that most observation periods span only a small age range, making it unlikely that any within-person age-related effect will be large.

6.5 Dependency between events and observation end

To understand whether censoring is dependent on the event, which would violate one of the assumptions of the SCCS, we can plot the difference in distribution between censored and uncensored events. By ‘censored’ we mean periods that end before we would normally expect. Here, we define periods to be uncensored if they end at either the study end date (if specified), database end date (i.e. the date after which no data is captured in the database), or maximum age (if specified). All other periods are assumed to be censored.

```
plotEventObservationDependence(studyPop)
```

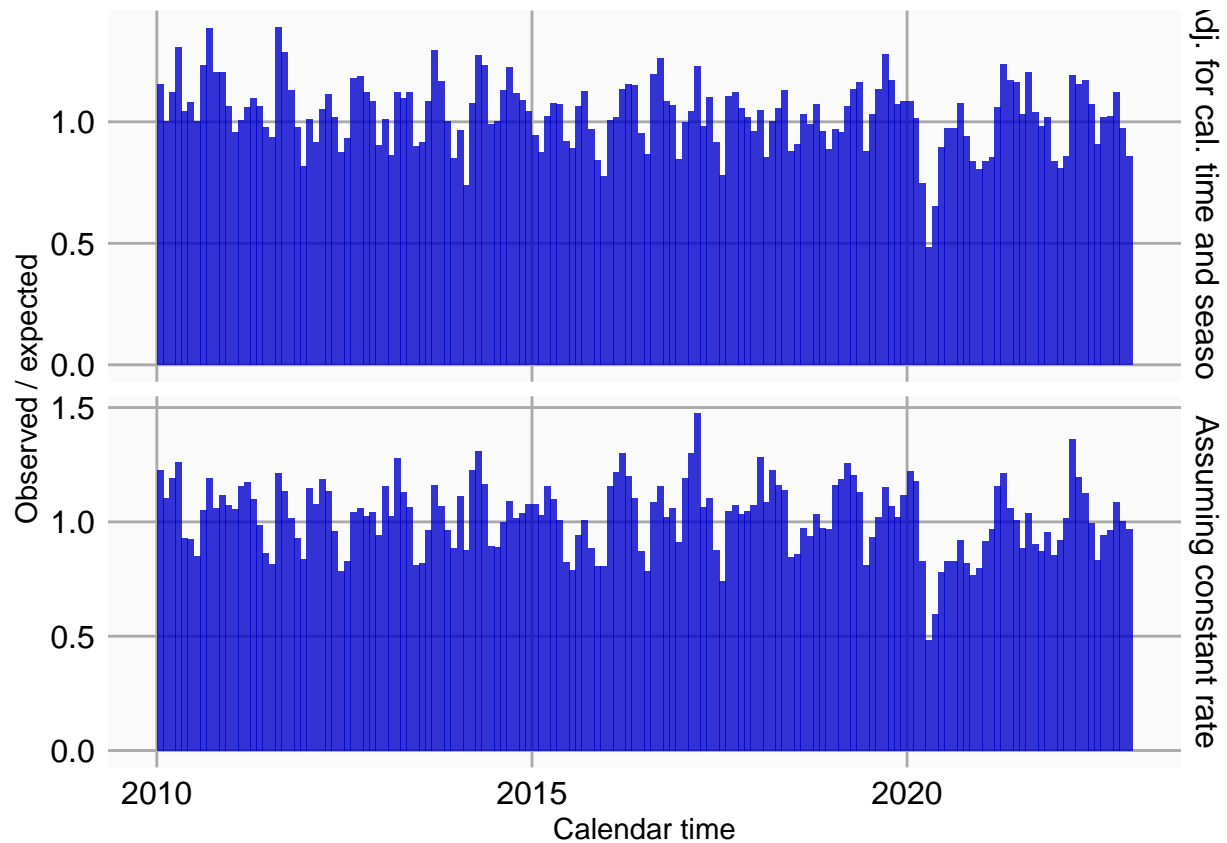


Here we see that overall the two distributions are somewhat similar, with little evidence that censoring tends to lead to shorter times to the end of observation.

6.6 Stability of the outcome over calendar time

If the rate of the outcome changes as a function of calendar time, this could introduce bias. For example, if the outcome is more prevalent during winter, and the exposure also tends to occur in winter, this will create an association between the two that likely doesn't imply causation. We can check for patterns over time:

```
plotEventToCalendarTime(studyPop)
```



In the bottom of the plot, we see the rate of the outcome does change over time.

7 Acknowledgments

Considerable work has been dedicated to provide the `SelfControlledCaseSeries` package.

```
citation("SelfControlledCaseSeries")
```

```
## To cite package 'SelfControlledCaseSeries' in publications use:
```

```
##
```

```
## Schuemie M, Ryan P, Shaddox T, Suchard M (2023). _SelfControlledCaseSeries: Self-Controlled Case S
```

```
## <https://github.com/OHDSI/SelfControlledCaseSeries>.
```

```
##
```

```
## A BibTeX entry for LaTeX users is
```

```
##
```

```
## @Manual{,
```

```
## title = {SelfControlledCaseSeries: Self-Controlled Case Series},
```

```
## author = {Martijn Schuemie and Patrick Ryan and Trevor Shaddox and Marc Suchard},
```

```
## year = {2023},
```

```
## note = {R package version 5.0.0},
```

```
## url = {https://github.com/OHDSI/SelfControlledCaseSeries},
```

```
## }
```

Furthermore, `SelfControlledCaseSeries` makes extensive use of the `Cyclops` package.

```
citation("Cyclops")
```

```
## To cite Cyclops in publications use:
```

```

##
## Suchard MA, Simpson SE, Zorych I, Ryan P, Madigan D (2013). "Massive parallelization of serial inference algorithms for complex generalized linear models". ACM Transactions on Modeling and Computer Simulation, 23(1), 10. <https://dl.acm.org/doi/10.1145/2414416.2414791>
##
## A BibTeX entry for LaTeX users is
##
## @Article{
##   author = {M. A. Suchard and S. E. Simpson and I. Zorych and P. Ryan and D. Madigan},
##   title = {Massive parallelization of serial inference algorithms for complex generalized linear models},
##   journal = {ACM Transactions on Modeling and Computer Simulation},
##   volume = {23},
##   pages = {10},
##   year = {2013},
##   url = {https://dl.acm.org/doi/10.1145/2414416.2414791},
## }

```

Part of the code (related to event-dependent observation periods) is based on the SCCS package by Yonas Ghebremichael-Weldeselassie, Heather Whitaker, and Paddy Farrington.

This work is supported in part through the National Science Foundation grant IIS 1251151.