

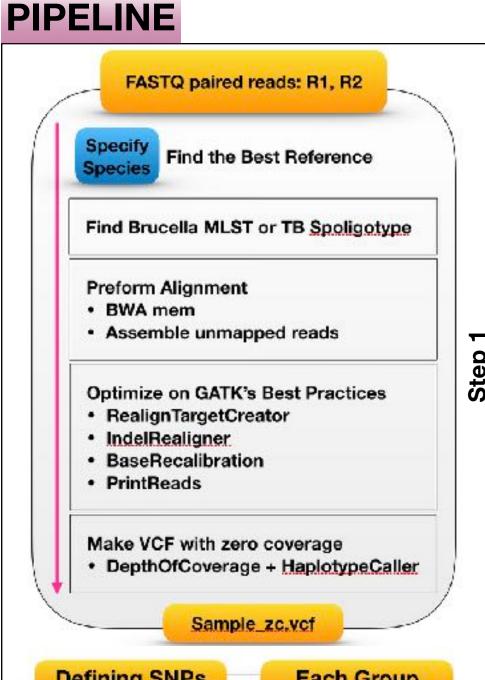
### vSNP: Bacterial validation SNP genotyping tool utilizing Mycobacterium and Brucella species

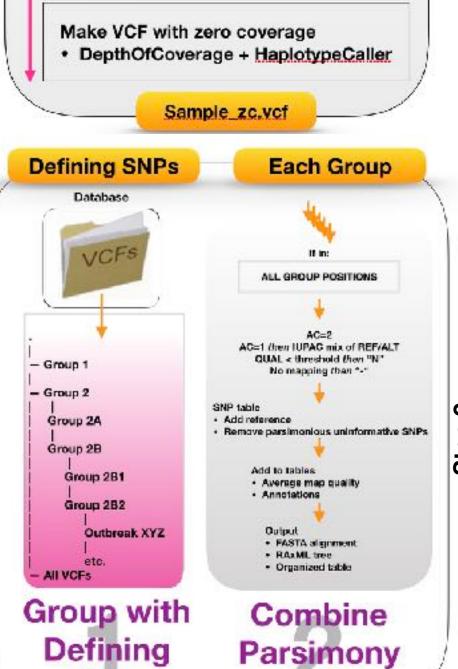
Barcode

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**Output** 





SNPs

**SNPs** 

### **Overview**

Single nucleotide polymorphism (SNP) analysis of high-throughput sequencing (HTS) data is increasingly the preferred method to genotype bacterial outbreaks. A specific tool was needed to rapidly call, validate and compare SNPs from FASTQ files in a timely manner while utilizing large datasets. vSNP was developed to address these challenges. It is particular well suited for outbreaks or clonal organisms such as Mycobacterium tuberculosis and Brucella species.

vSNP is publicly available at: https://usda-vs.github.io/snp\_analysis/

### Method

vSNP runs on macOS and Linux systems. It is written in Python 3, utilizing Python packages and system programs available from Anaconda package manager. The program is run from the command-line, requiring minimal experience to setup and execute.

vSNP provides a high-resolution SNP analysis from Illumina pair FASTQ files. vSNP is implemented in a two-step process. Step 1 chooses the best reference, and outputs alignments and SNP calls into VCF files. Step 2 takes in a collection of VCF files and outputs SNP alignments and phylogenetic trees. Predetermined SNP positions are used to create groups. Grouping samples on defining SNPs provides manageable datasets and allows results to be quickly and thoroughly validated. Poor SNP positions are easily targeted for further visual validation using Excel formatted SNP alignment tables. Phylogenetic trees are created from validated tables. SNP tables provide the average map quality and annotation at each position.

### Conclusion

To achieve high resolution genotyping, it is necessary to validate SNPs within an outbreak. This must be accomplished quickly to minimize personnel time and to not delay actionable results. We have found it best to validate using SNP tables with fewer than 50 SNP positions. Defining SNPs are used to group similar samples into smaller, more manageable groups. This use of a defining SNP to group closely related isolates is a key element that differentiates vSNP from other pipelines. It allows both a perspective against other isolates and allows the validation necessary to make confident, actionable decisions.

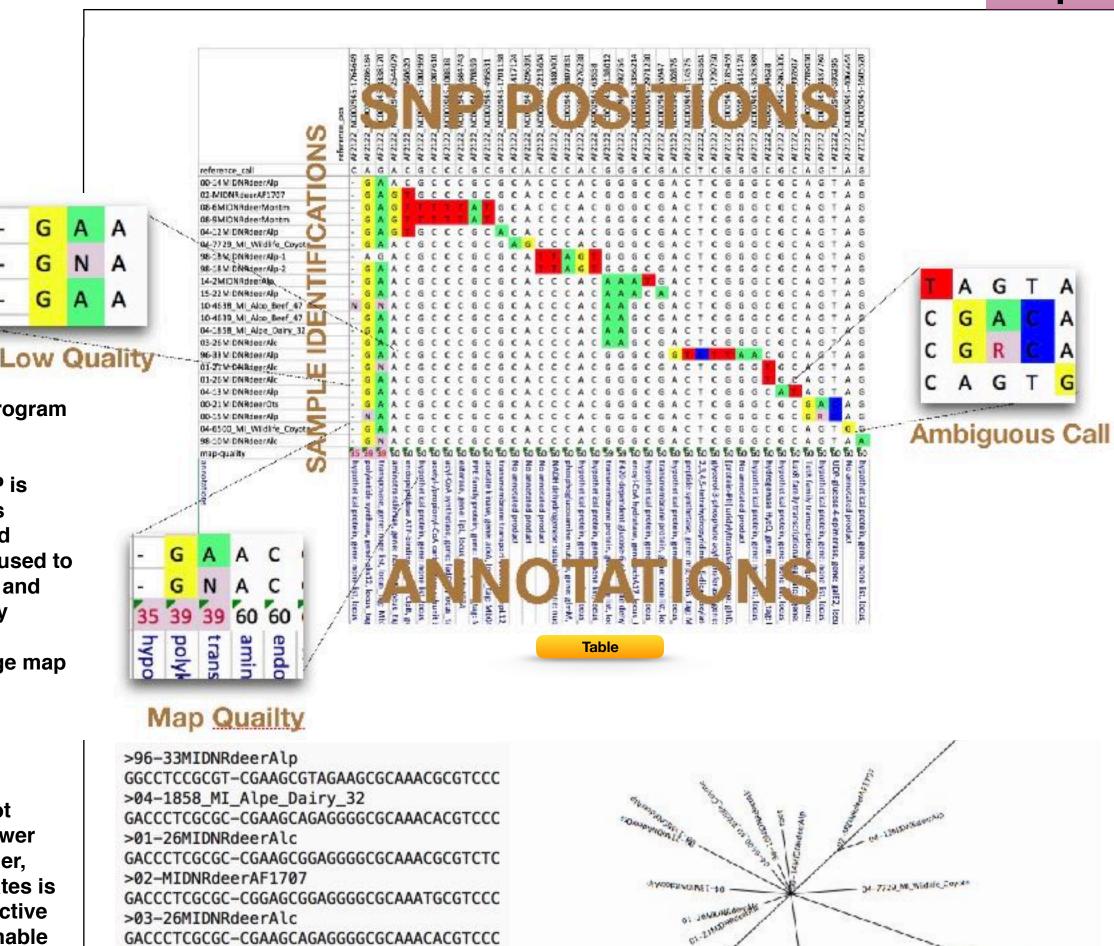
Other clonal organisms, or bacteria or even viruses within an outbreak situation, can also be incorporated into vSNP to provide the same high-level resolution.

### **Pros:**

- Proven pipeline workflow
- Maintain sense of data ownership
- High resolution, validated genotyping

### Cons:

- Command-line drive
- Unable to run open source tools (GATK) in Windows



# >03-26MIDNRdeerAlc GACCCTCGCGC-CGAAGCAGAGGGGGCGCAAACACGTCCC >08-9MIDNRdeerMontm TATTCTCGTGC-CGGAGCGGAGGGGCACAAATGTGTCCC >04-13MIDNRdeerAlp GACCCTCGCGC-CGAAACGGAGGGGCGCAAACGCGTCCT

>01-21MIDNRdeerAlc GACCCTCGCGC-CGAAGCGGNGGGGCGCAAACGCGTCTC >15-22MIDNRdeerAlp GACCCTCGCGC-CGAAGCAGAAGGCCGCAAACACATCCC

Alignment

Mbovis-01D2

### FASTQ paired reads: R1, R2

## Specify Species

### Find the Best Reference

### Find Brucella MLST or TB Spoligotype

### **Preform Alignment**

- BWA mem
- Assemble unmapped reads

### **Optimize on GATK's Best Practices**

- RealignTargetCreator
- IndelRealigner
- BaseRecalibration
- PrintReads

### Make VCF with zero coverage

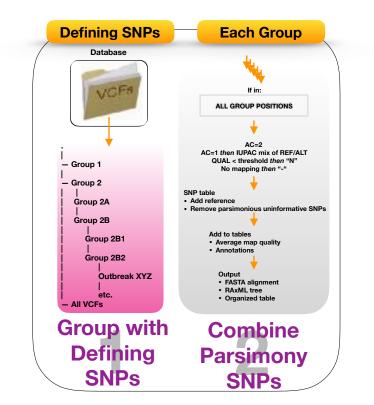
• DepthOfCoverage + HaplotypeCaller

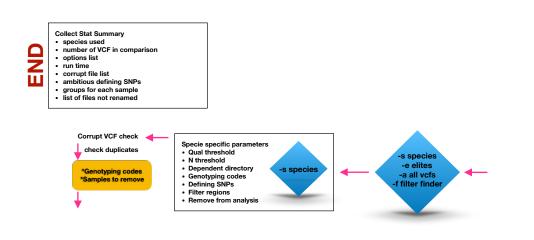
Sample\_zc.vcf

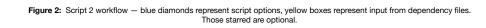
FASTQ paired reads: R1, R2

START









Table



Find Quality SNPs in each VCF

### Mbovis-01D2

