

## Keyence BZ-X800 Fluorescence Microscope Instructions

- Fix cover slips to the slide; you can use nail polish
- Imperative to use oil with 100x
- Incubator (live cells). Regulate temp, CO<sub>2</sub>, humidity
  - Prolonged studies
- Lift condenser before loading samples
- Load slides outside of the microscope
- Have cover slip face down; bottom right corner of slide less than 90°
- Writing on bottom and you should see the text (place on top)
- BZ-X800- viewer pics
  - Analyzer- counts and image processing
- ★ Probably won't use other 2 software's
- Resolution sensitivity- binning- combine in single pixel
  - High resolution: 100x very fine structures
- Longer exposure- risk to photobleaching
- High sensitivity- higher
- Standard- 90% of the time sufficient
- Manual Focus
  - Scroll mouse- change Z (move lens up or down) (fine focus)
  - Hold control and scroll (coarse focus)
- ★ 100x manual and NOT autofocusing
- Navigation- find sample and creates map
- Correction collar- adjust lens (only for 20 and 40x)
  - Adjusts beam to compensate for thickness of the vessel
  - Ring with numbers on the lens
  - Will have to change for multi well plates
- Low photo bleach- excitation light not continuously on to preserve samples
- Multiple markers- multi color box on the top left of the screen
  - Channel cube
  - Camera setting auto
  - Layers different color images
- Image processing- per individual channel, then merge
- ★ 3 Main Options
  1. Black Balance
    - Set square over background
    - Eliminates background, low level fluorescence
    - Keep sample parameters consistent- save conditions
    - Load conditions and pull up next file
    - Important for quantitative analysis

2. Lookup Table
    - Shadow similar to black balance
    - Highlight- brighter
    - Leave gamma
  3. Haze Reduction
    - Looks for localized peaks
    - More crisp and clear images
    - Usually preview 2-3 gets the job done
    - Works well on fine structures; fluorescence
    - Won't work well with brightfield
- Overlay images
  - Insert scale bar, numeric display, calibration setting micron per pixel
  - Click "line image" after capturing photo to unfreeze microscope/screen
  - X40-x100 (best frame of reference)
    - Drop immersion oil (gets through around 2 cover slips)
    - Lens paper to clean it
    - Pipette dropper to dab oil
    - Do not suck up air bubbles
    - White through catches extra oil
    - Only need small amount (about as much as a ball point pen)
    - Click yes after adding oil
    - Scroll down and don't do autofocus (it lowers lens and creates an air gap which presses back to the slide and creates an air bubble)
  - Hold control, coarse focus and then go to fine focus
  - Z stacking- scroll up until out of focus (top of sample)
  - Scroll down to find bottom of the sample
    - Images in different planes
    - Z stack puts them together
    - Set upper and lower limit
    - Pitch= step size **0.3 microns**
    - Start capture takes multiple images for Z-stack
    - Analyze images and scroll with yellow bar
    - Load (L) bottom right corner
  - Now we want to squish into one image (full focus, standard image)
  - Z- stacking typically for thick samples, but could be used for cultures too (anywhere between cover slip and slide can be cells within water)
  - Image tab- can do settings before capturing but then that will be the raw image. It also depends on your project requirements
  - Menu- capture setting- save (reproducibility between samples)
    - Can also load settings from image file
  - 100x-4x
    - Click wipe off oil off slide
    - 70% ethanol or sparkle lens cleaner

- NEVER use elm wipe on lens (scratches lens)
- Use optical lens paper. Fold over a couple of time
- Add small bit of lens solution
- Air out so it is not soaking wet
- Clean only in circular motions gently
- Move finger to the spots without oil
- Another sheet of dry lens paper and wipe one more time
- Clean white trough/ring
- Dried oil can scratch lens
- Best practice to just clean in between lenses. 40x highest risk of getting damaged
- Counting software- still cells
- Algae preservation?
- XY measure- count; click cells (> 100 cells)
- 2D measurements, area measurement
- 6mo. To test out software
  - Different pics/areas in slide
  - Number of slides
  - Extrapolations density to whole slide/sample
  - Take standard deviations between samples number of trials to find optimal sweet spot
- Algae- high sensitivity
- Colony counter
  - Food/beverage geared
  - Doesn't do sample prep
  - Separates on morphology
  - Check sheet
  - Count
  - Document
  - Transfer to excel