

SBCNY

NIGMS funded Center

Experimental Methods in Systems Biology

Part of the Coursera Certificate in Systems Biology

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Fall 2014, Week 1, Experimental Perturbations



Icahn School
of Medicine at
**Mount
Sinai**

Important Features of Any Experiment

- Given a specific question, one can then come up with answers to the following three key properties of the experiment:
 1. What biological system?
 - E.g. Do I look at human cell lines, a mouse, or yeast?
 2. What perturbation/treatment conditions?
 - E.g. What compounds should I apply to the system to elicit a relevant response?
 3. What measurements?
 - E.g. What transcripts do I need to look at, and/or do I need to look at protein levels instead?
- Often (but not always), if you can't design an experiment that only has a handful of conditions and measurements, results may be difficult to interpret
 - Usually the question is too complex or not significant
 - Exceptions are screening based studies, but those also typically have a specific question of interest

Outline

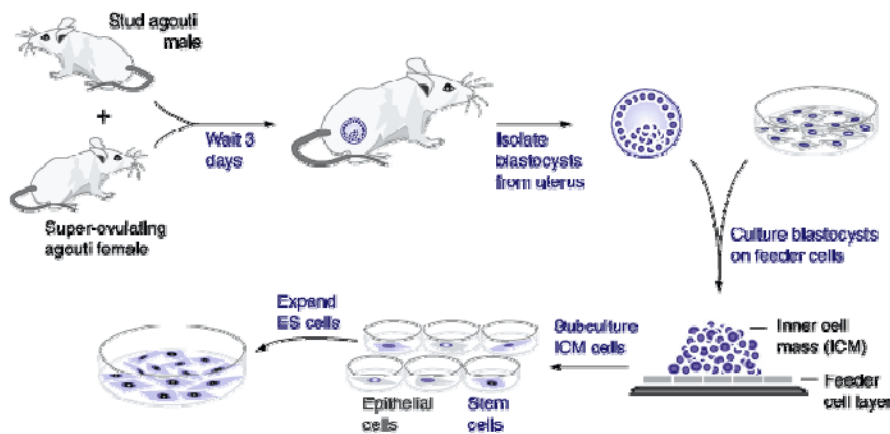
- “Basal” Perturbations
 - Genetic
 - Non-genetic
- “Acute” Perturbations
 - Compound-based (pharmacological and biological)
 - Light-based
- We focus on mammalian cell line and cell culture studies, but many of the techniques we describe can be applied to other model systems (with reasonable alteration)
 - There are lots of techniques in mouse genetics that I won't cover much here

Basal vs. Acute Perturbations

- Basal Perturbation: An alteration that is permanently present (or at least can be considered permanent relative to the time scale of the experiment), and for which one does not measure dynamic response.
 - Often followed by an acute perturbation
 - Can also be thought of as “chronic”
- Acute Perturbation: An alteration to the biological system that induces rapid changes (relative to the time scale of the experiment)
 - One is often interested in a specific time point or an entire time course of many time points post-perturbation

Basal Perturbations

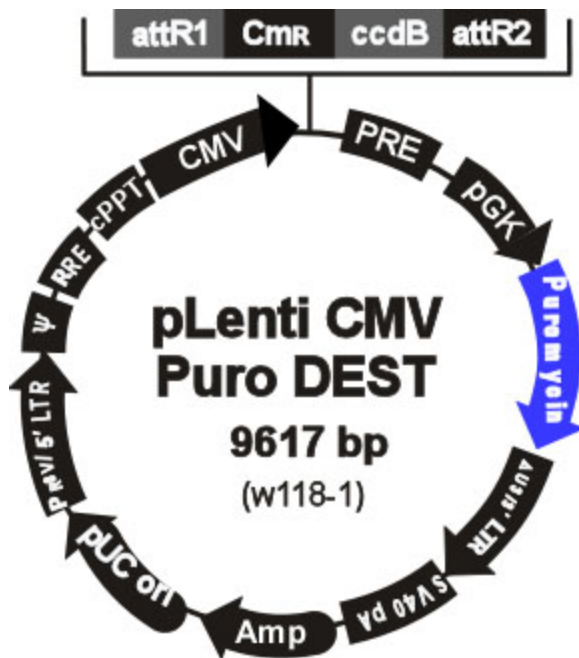
Genetic—Mouse Embryonic Fibroblasts (MEFs) or Embryonic Stem (ES) Cells



<http://www.biochem.arizona.edu/miesfeld/teaching/Bioc471-2/pages/Lecture19/Lecture19.html>

- Wide variety of mouse genetic knockouts are available
- Many knockouts are embryonic lethal, but one can usually at least obtain fibroblasts or ES cells from the embryo
 - They can be grown in culture
- These can be useful for exploring the molecular functions of particular gene products
- They can also be useful for negative and positive controls for other types of experiments
- ES cells can be differentiated into a variety of other cell types (growing field)
 - Induced pluripotent stem (iPS) cells are similar but can be derived for adult human cells (e.g. fibroblasts)

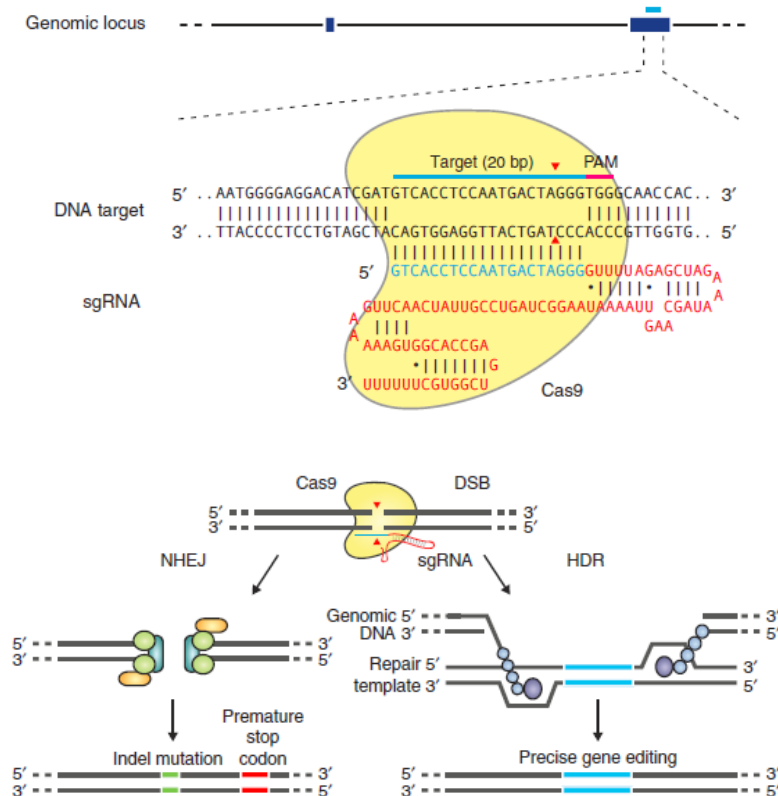
Genetic—Plasmid-Based Selection



<https://www.addgene.org/static/data/61/90/2513a6d2-af62-11e0-90fe-003048dd6500.jpeg>

- Plasmids can contain “resistance genes”
 - Code for a protein that metabolizes a toxic drug
 - Puromycin, G418 (similar to neomycin), blasticidin, zeocin, hygromycin B
- By transfecting the plasmid and treating cells with the drug, only cells that incorporated the plasmid’s resistance gene will survive
 - The plasmid gets randomly incorporated into the genome
 - Linearizing the plasmid by making a single restriction enzyme cut improves efficiency
- Incorporating the plasmid into a lentivirus can improve efficiency
- One can also use fluorescent proteins combined with flow sorting as an alternative (although there is no active selection pressure)

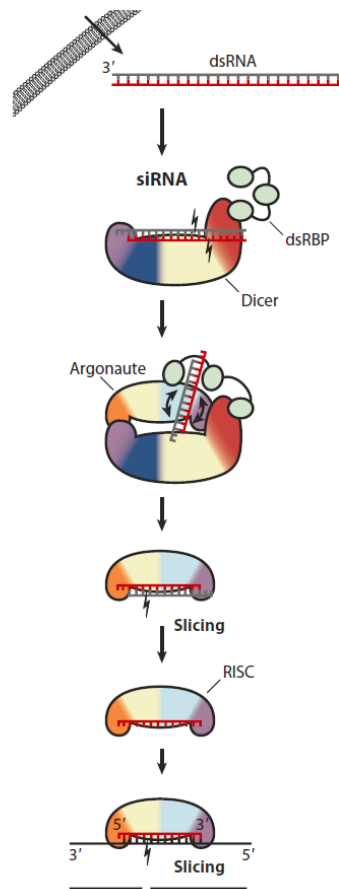
Genetic—Genome Engineering



Ran et al., Nat Protocols, 2013

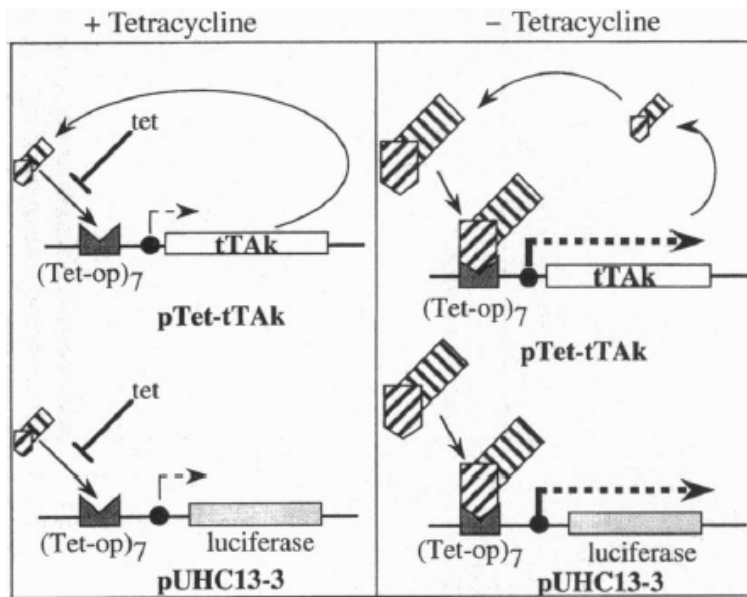
- Genome engineering: directed modification of particular loci
- General strategy—induce DNA double strand breaks and then rely on
 - non-homologous end joining for repair, which typically disrupts gene function
 - Homologous recombination, which places a provided double stranded DNA template where the break occurred
- Recombination enzymes
 - Flp recombinase and CRE/Lox systems—these recombination enzymes
- TALENs and ZFNs
 - Requires protein engineering to achieve DNA sequence specificity of ~12-24 bp and do not always cut in the same location
 - Can only reasonably target one locus at a time.
 - Thus they are somewhat difficult to use and target
- Latest—CRISPR/Cas9 (Ran et al., Nat Protocols, 2013 and References therein)
 - Uses a 20 bp “guide RNA” (gRNA) to bring the Cas9 nuclease to DNA which is complementary to the guide RNA
 - Allows simple and cheap targeting by oligo synthesis
 - Can be targeted almost anywhere in the genome quite specifically (20 bp can be reasonably unique)
 - Tools at www.genomeengineering.org
 - Wild type Cas9 vs. “nickase” Cas9
 - Wild type catalyzes a full double strand break where the gRNA dictates
 - Nickase Cas9 catalyzes a single strand break, therefore one needs two “nicks” to make the double strand break

Non-Genetic—RNA interference



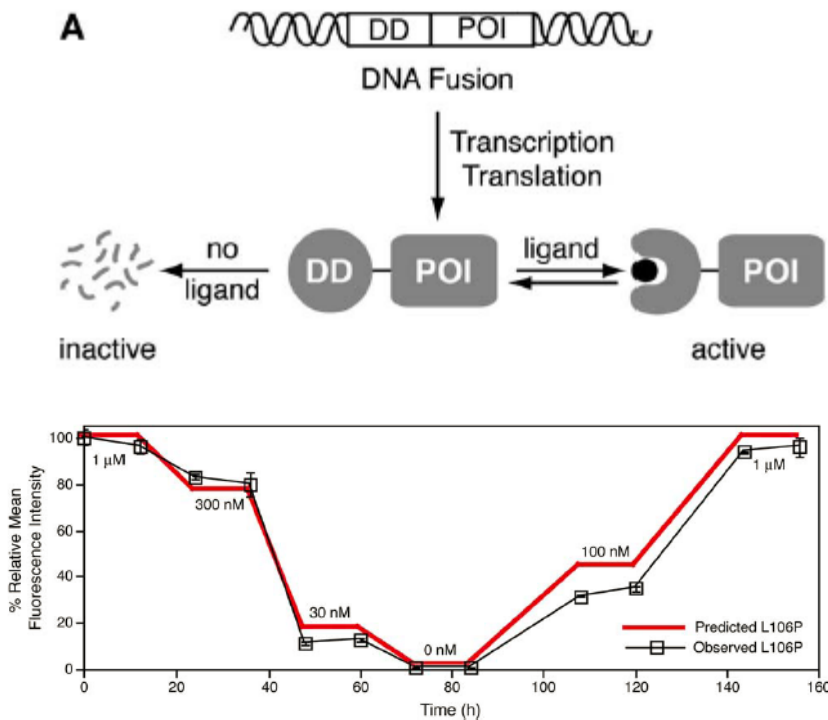
- Discovered in 1998 by Fire and Mello (Fire et al., Nature, 1998)—awarded Nobel prize in 2006
 - Notably rapid progression from discovery to Nobel
- A short (~20 bp) duplex RNA is directed to complementary transcripts, which are then degraded
 - Duplex can be small interfering RNA (siRNA)—two independent strands, or short hairpin (shRNA)—one strand that hybridizes to itself with a stem loop at one end (looks like a hairpin!)
- This “knocks down” expression of the corresponding genes
 - Similar pathways process micro RNAs (miRNA) that are produced endogenously and can inhibit translation and degrade mRNAs
- Efficiency of knockdown widely varies across genes and cell types
- Need to use two independent sequences to control for off-target effects
 - Gold standard is a “rescue” experiment, where an RNAi resistant transcript is ectopically expressed from a plasmid (resistant through engineered silent mutations)
- Can be done in a transient or stable manner
- Can be inducible (see next slide)
- Collection of Reviews: <http://www.nature.com/focus/rnai/index.html>

Non-Genetic—Transcription Rate



- A variety of systems exist to control transcription rate with small molecules
 - Too many to cover here, but they all work similarly to a certain extent
- One common system is with tetracycline (or more commonly its more stable and higher affinity analog doxycycline)
- Tet-on system:
 - Cells expressing the tet repressor, and with the gene of interest downstream of the tet repressor binding sites
 - The tet-repressor only binds DNA when it is not bound to tetracycline
- There are also “Tet-off” systems
- Can control expression of genes and shRNAs

Non-Genetic—Protein Half-Life



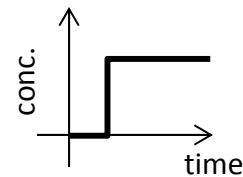
- A few systems allow modulation of protein half life with small molecules
- They all involve some kind of alteration to the protein-of-interest
- Examples
 - DD domain/Shield1 (Banaszynski et al., Cell, 2006; 126(5): 995-1004)
 - Shield1 enters live cells, binds to the DD domain, and increases protein stability.
 - auxin/AID (Holland et al., PNAS, 2012; 109(49))
 - Auxin (IAA) works similar to Shield1 but decreases protein stability
 - However it requires ectopic expression of TIR1

Banaszynski et al., Cell, 2006; 126(5): 995-1004

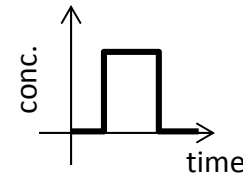
Acute—Pharmacological or Biological Compounds—Pipette-Based

- Pharmacological: a compound that is not naturally occurring in the system of interest but interacts with the model system of interest
 - E.g. a drug
- Biological: a compound that normally interacts with the system of interest in a biological context
 - E.g. a growth factor

Step Input: Treatment with a compound at a certain constant concentration.



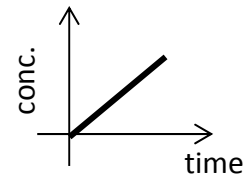
Pulse-Chase Input: Treatment with a compound at a certain concentration for a specified time period, followed by a second concentration (usually zero).



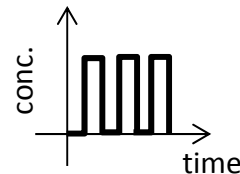
Acute—Pharmacological or Biological Compounds—Pump-Based

- Although it is not common or very simple, one can use perfusion or pump-based systems to create complex time dependent inputs
- Microfluidic designs can give enormous flexibility here

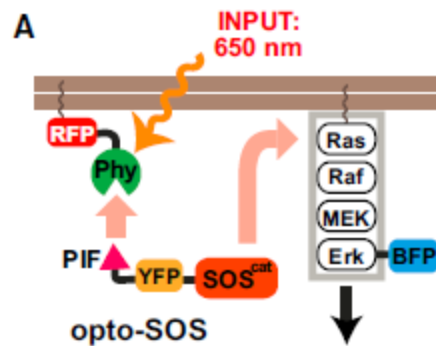
Ramp: Treatment concentration linearly ramps up or down with time (e.g. Sasagawa et al., Nat. Cell Biol., 7, 2005)



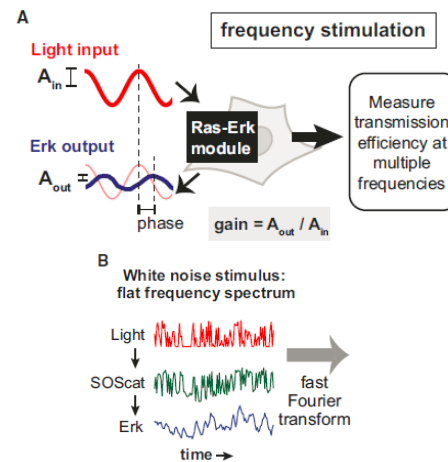
Wave: Treatment concentration oscillates in time with a fixed amplitude and period (e.g. Mettetal et al., Science, 2008, 319 (5862))



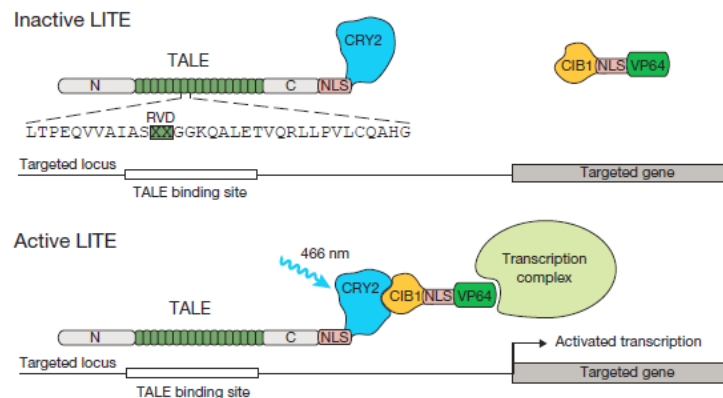
Acute—Light-Based



Toettcher et al., Cell, 2013



- Optogenetics: Using light-sensitive proteins to control biochemical activities.
- First application was ion channels in neuroscience (reviewed in Zhang et al., Cell, 2011, 147(7))
- Has been extended to various light-sensitive protein-protein interactions (e.g. Levskaya et al., Nature, 2009—Phy-PIF; Kennedy et al., Nat. Methods, 2010—CIB-CRY) and transcription (Konermann et al., Nature, 2013)



Konermann et al., Nature, 2013

Next Time—Nucleic Acid Measurements