

# Quantification

# Quantifying Fluorescence Intensity

Table II. Protocol for quantitation of fluorescence intensity values

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## 1. Acquire optical images

- Set up specimen and imaging system for optimal signal detection, low background, and low noise (Table I)

## 2. Acquire digital images

- Use software to monitor intensity values in the image to choose the best acquisition settings<sup>a</sup>
- Use full dynamic range of the camera for fixed specimens<sup>a</sup>
- For live-cell work, it is often necessary to sacrifice SNR to minimize specimen exposure to light and maintain cell health and viability<sup>a</sup>
- Consider binning to increase SNR<sup>a</sup>
- Avoid high camera gain when a large dynamic range is needed<sup>a</sup>
- Avoid saturating pixels in the image<sup>a</sup>
- Eliminate or minimize exposure of specimen to fluorescence excitation light prior to image acquisition<sup>a</sup>
- Focus carefully, preferably with phase or DIC<sup>b</sup>

## 3. Store images

- Always save the raw images<sup>c</sup>
- Use either no compression or lossless compression<sup>c</sup>

## 4. Process images

- Use flat-field correction to correct for uneven illumination<sup>d</sup>
- Be sure any other image processing used prior to quantitation preserves relative intensity values<sup>c,d</sup>

## 5. Analyze images

- Subtract local background value from intensity measurements<sup>e</sup>
- Do not measure intensity values on compressed or pseudo-colored images<sup>c</sup>
- Validate image segmentation and analysis method<sup>f</sup>
- Calculate and report the error in your measurements<sup>d,g</sup>

Waters, JCB, 2009 7(185)

# Quantifying Intermolecular FRET— Sensitized Emission

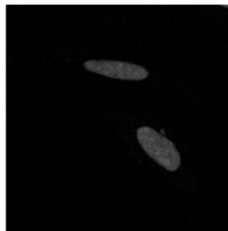
Predetermined factors with pure samples of donor and acceptor:

Donor cross-talk :  $R_D$

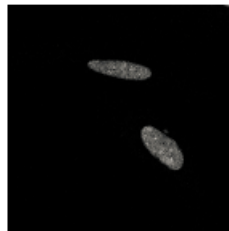
Acceptor cross-excitation:  $R_E$

Required images:

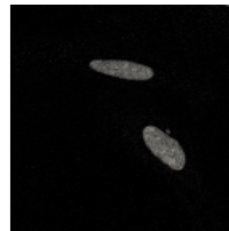
Donor channel  
Donor excitation  
 $F^D$



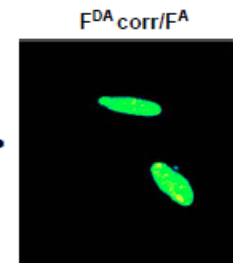
Acceptor channel  
Donor excitation  
 $F^{DA}$



Acceptor channel  
Acceptor excitation  
 $F^A$



=>

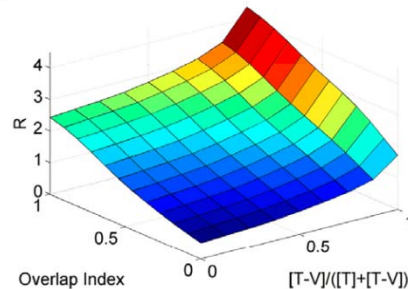


$$E_a = \frac{F^{DA} - \overset{\text{Donor cross-talk correction}}{F^D \cdot R_D} - \overset{\text{Acceptor cross-excitation correction}}{F^A \cdot R_E}}{F^A} = C \cdot E \cdot \alpha_A$$

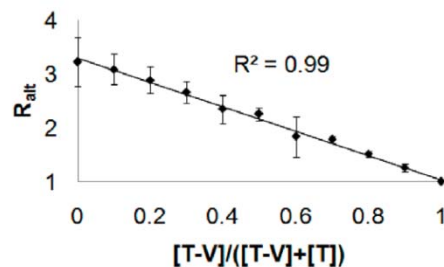
$$\Rightarrow E_a = \frac{F^{DA}_{corr}}{F^A}$$

# Quantifying Intramolecular FRET

C



D



Birtwistle et al., PLoS One, 2011

- Because the stoichiometry is known and fixed (typically 1 to 1) a simple ratio can be used
- Typically, it is the Donor Ex/Acceptor Em channel divided by the Donor Ex/Donor Em channel (denoted  $R$ )
- However this can have a non-linear relationship to FRET efficiency (Birtwistle et al., 2011)
- Something which may solve that is using Donor Ex/Donor Em channel divided by Acceptor Ex/Acceptor Em channel (denoted  $R_{alt}$ )
- Another way to quantify FRET, either intra or inter, is by fluorescence lifetime (FLIM)...we won't cover that here