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# Experimental Methods in Systems Biology

Part of the Coursera Certificate in Systems Biology

Marc Birtwistle, PhD

Department of Pharmacology & Systems Therapeutics

Fall 2014, Week 1, Overview



Icahn School  
of Medicine at  
**Mount  
Sinai**

# Outline

- Introductions
- Course Scope
  - What we will cover in depth
  - What we will cover briefly (only in Week 1 intro)
  - What we will not cover
- The “Systems Biology Loop”
- Distinctive Features of Systems Biology Experiments
- Important Features of Any Experiment

# Introductions

- About me—Marc Birtwistle
- B.S. and Ph.D in Chemical Engineering
- Postdoctoral Work in Cell and Molecular Biology
- Became Assistant Professor at the Icahn School of Medicine at Mount Sinai Sept. 2012
- Our lab focuses on integrating experiments and computation in the context of cancer systems pharmacology of glioblastoma multiforme, a deadly brain tumor
- We are particularly interested in cellular heterogeneity and how that impacts disease progression and treatment response

# Introductions

- I assume you have a basic knowledge of cell and molecular biology, but I review relevant concepts to the extent possible.
- I also assume you have a knowledge of basic statistics.
- Although systems biology can be mathematics and computer science heavy, this course does not go into much depth in that regard.

# Introductions

- As you can guess from my training and lab focus, I am familiar with many types of experiments and how they are used in computational modeling.
- However it's impossible for anyone to be an expert in a wide array of techniques, therefore I draw on the help of many people in creating this course.
- It's also not possible to cover every technique in reasonable depth.
- I've chosen a few key techniques to focus on that will give you reasonable coverage of important items, and briefly mention many others.

# Scope—In Depth Coverage

- mRNA sequencing (Week 2)
  - Uses “next-generation” massively parallel DNA sequencing to quantify gene expression in an “omic” (genome-scale) fashion
  - Mortazavi et al., 2008, Nat Meth; Wang, Gerstein, Snyder, 2009, Nat Rev Genetics
- Mass spectrometry-based proteomics (Week 3)
  - Uses state-of-the-art mass spectrometry to quantify the levels of proteins (or protein states) in an “omic” fashion
  - Schwanhauser et al., 2011, Nature

# Scope—In Depth Coverage

- The previous two techniques are omic-scale, and typically applied to cell populations rather than single cells
  - However single cell mRNA sequencing is becoming more feasible—e.g. Patel et al, Science, 2014 (Regev lab), Treutlein et al., Nature, 2014;
- Things need not be omic-scale to be systems biology.
- Many systems biology experiments
  - concern single-cell behavior and dynamics/time courses, which are difficult to probe on a omic-scale
  - are focused on small subsystems, e.g. a kinase signaling cascade

# Scope—In Depth Coverage

- Flow and Mass Cytometry (Week 5)
  - Uses labeled antibodies and other specific labeled reagents to stain single cells in suspension, then measures labeling intensities in these single cells relatively rapidly (often easily 100s of cells/second)
  - Allows observation of a handful of analytes in many single cells of a population at fixed time points
  - Bendall et al., Science, 2011
- Live-cell Imaging (Week 6)
  - Uses (predominantly) fluorescence microscopy to observe processes in living cells in real time.
  - Allows observation of one or a few analytes in a handful of cells with dense temporal sampling
  - Regot et al., Cell, 2014



# Scope—Brief Coverage

- Nucleic acid measurements
  - Quantitative polymerase chain reaction (qPCR)
  - Microarrays
  - Whole genome sequencing
  - Exome sequencing
  - Bisulfite (methylation) sequencing
  - ChIP sequencing (epigenetics)
  - Fluorescence in situ hybridization (FISH)

# Scope—Brief Coverage

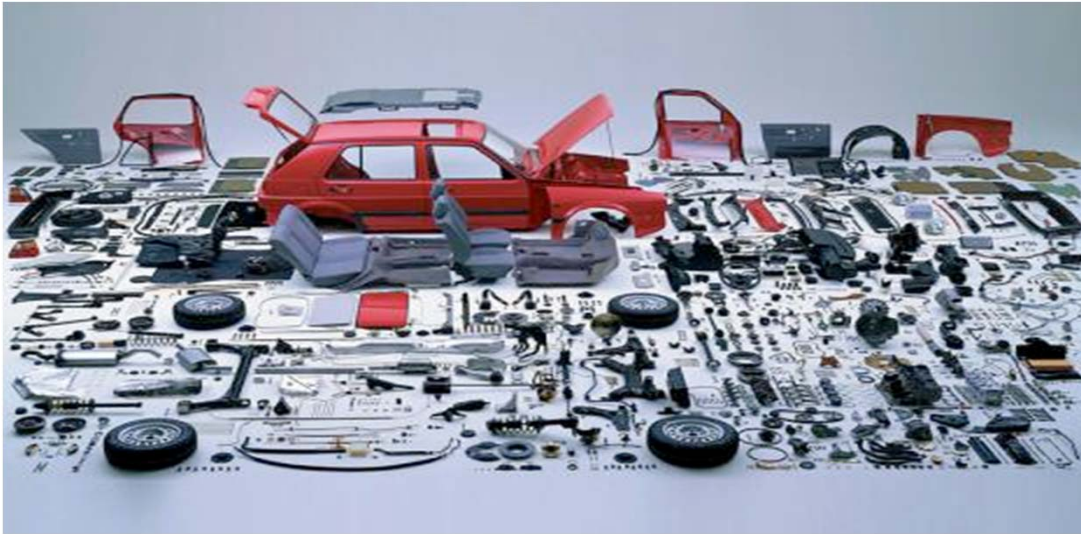
- Protein and Protein State Measurements
  - Western blot/microwestern
  - Reverse-phase Protein Array
  - Immunofluorescence

# Scope—No Coverage

- Microfluidics
  - All kinds of clever devices built by microfabrication techniques that allow precise perturbation and/or capture of cells for unique measurements
  - It's relevant and interesting but we don't have time to do it justice
- Metabolomics
  - Mainly mass spec-based approaches to measure metabolites present in cells on an omic-scale
  - Again, it's quite relevant in many areas of systems biology but we just don't have time to give it proper coverage
  - We do cover mass spectrometry so its not a large stretch to consider it also applies to metabolites
- High-throughput Techniques
  - Many of the techniques we will describe have been optimized to be done in a high-throughput fashion
  - We describe the principals of the technique rather than its scaling
- Formal Design of Experiments
  - There are established statistical fields devoted to design of experiments
  - There is much ongoing work in systems biology to use quantitative computational models as part of this
  - We rather focus on the experimental techniques themselves

# The Systems Biology Loop

# What is Systems Biology?

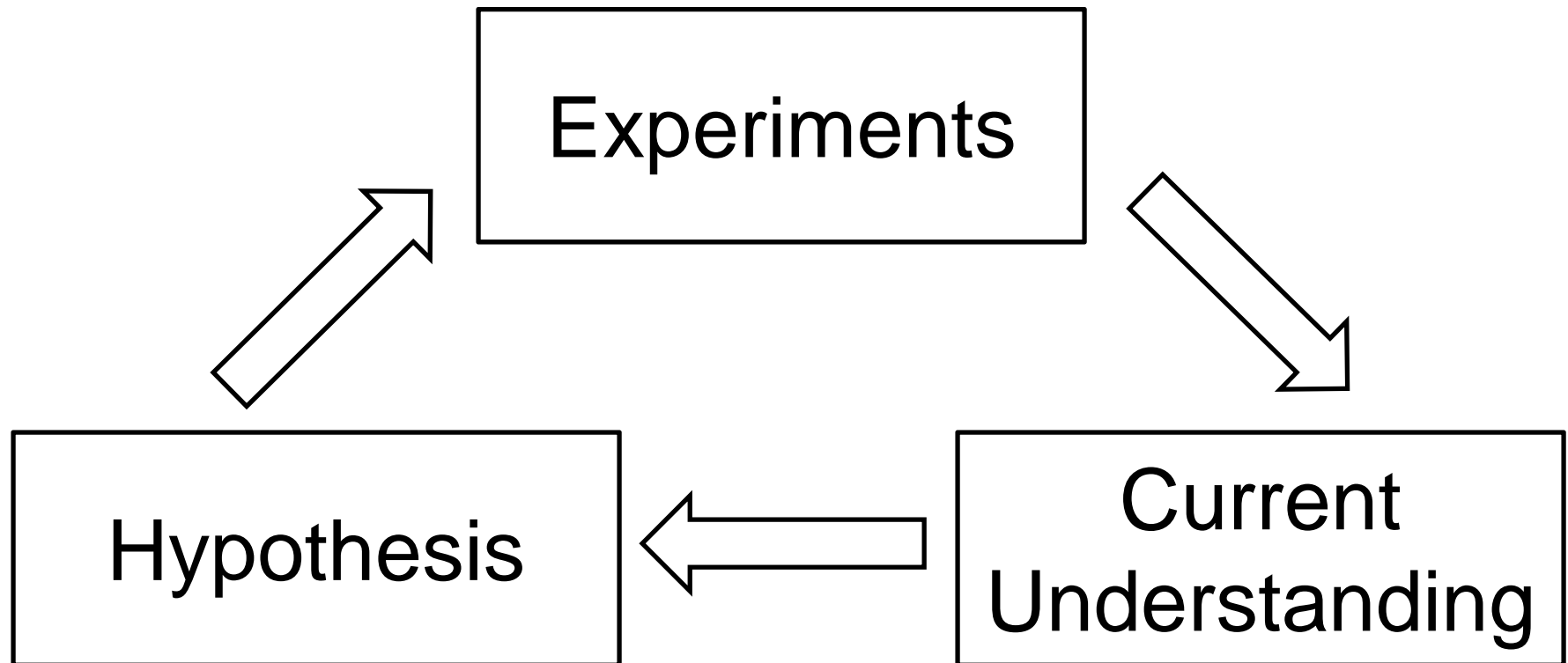


*We know all the parts, so we should be able to understand how it works, right?*

**Biologists typically try to understand one part.**

**Systems biologists try to understand how the interactions between many parts give rise to function. Some say it is physiology re-invented (they're probably correct, although there are some differences).**

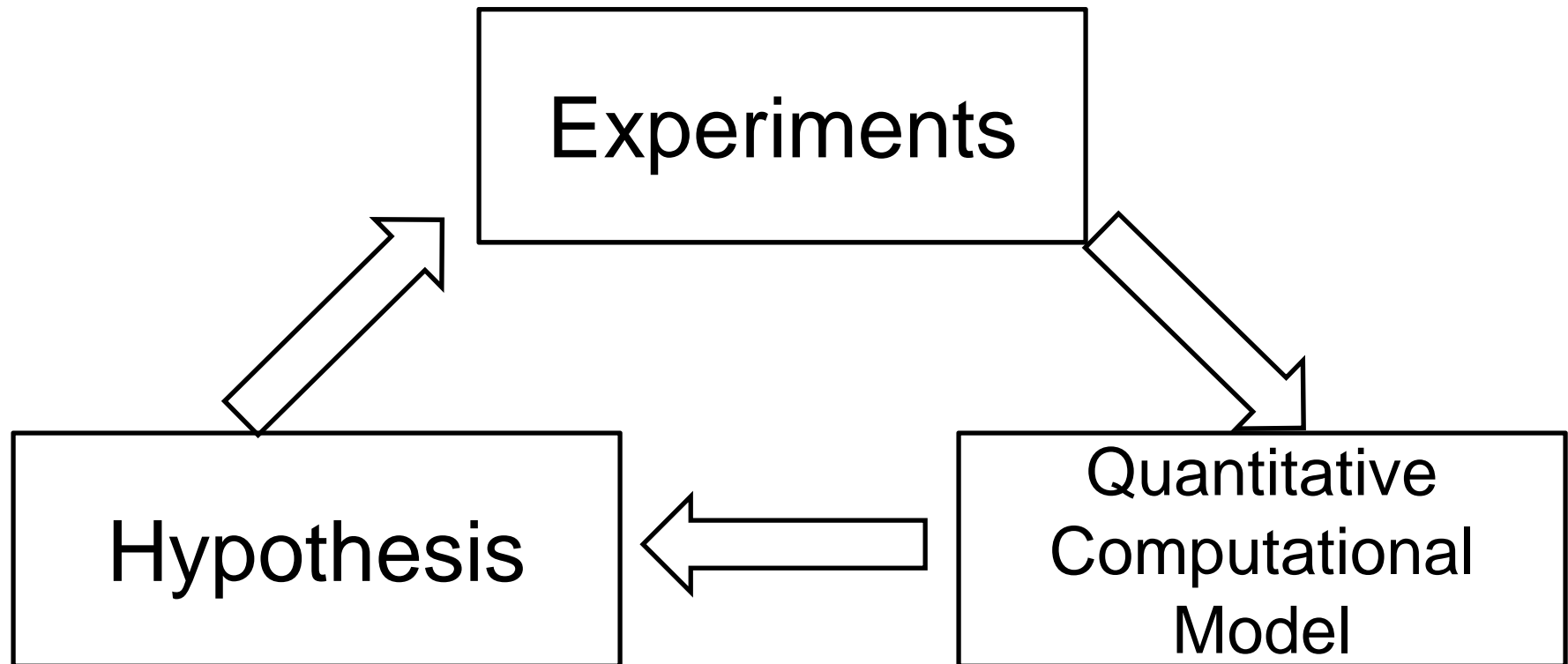
# Common Scientific Workflow



# Common Scientific Workflow

- In practice, one can enter this loop anywhere.
- The loop need not be completed by one group.
  - In fact, many great discoveries have been made when the loop arrows are completed by different groups sometimes long separated in time.
    - E.g. Watson and Crick with the DNA double helix structure
    - Recent discovery of the Higgs Boson in particle physics

# How is Systems Biology Different?

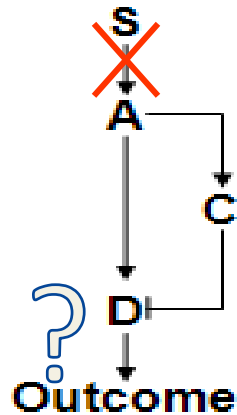




# What Difference Does That Make?

- As opposed to many other scientific disciplines, “Current Understanding” in biology is often semantic or cartoon-based.
- Even though much can and has been learned in this way, it can be imprecise, even in simple cases.
- Moreover, biology can be remarkably complex.
  - This makes semantic or cartoon-based understanding even more imprecise, and sometimes even impossible.
- Many engineered systems are also remarkably complex, but the difference in biology is that we didn’t build it, so we have limited knowledge of what is in there!

# Issues with Cartoon-Based Understanding



- **Need to know:**
  - **Magnitude of Effects on D**
    - A strong; C weak  $\rightarrow$  D up
    - A weak; C strong  $\rightarrow$  D down
  - **Dynamics of Interactions with D**
    - A slow; C fast  $\rightarrow$  D down then up
    - A fast; C slow  $\rightarrow$  D up then down
  - **Localization with D**
    - A local; C distant  $\rightarrow$  D up
    - A distant; C local  $\rightarrow$  D down

**Quantitative Computational Models Allow Us To Keep Track of These Kinds of Properties in Complex Systems**

# Distinctive Features of Systems Biology Experiments

- Because systems biology experiments usually support development or refinement of a quantitative computational model, they tend to have the following properties
  - Quantitative
    - Many biological experiments tend not to be quantitative, either because the technique inherently disallows it, or because a great deal of extra effort is required to be quantitative.
    - We will give strong focus to how one makes experiments quantitative.
  - Dynamic
    - Often, critical information from a systems biology experiment is how the system responds to a perturbation over time, which supports development of a computational model and can support inference of causality.
    - Many important phenomena in biology are time-dependent, e.g. circadian clocks, cell cycle, developmental processes, drug responses, action potentials, etc.
- These two features transcend most systems biology experiments

# Distinctive Features of Systems Biology Experiments

- Large-scale vs. small-scale systems biology
  - Although omic-level measurements are often associated with systems biology, one needn't have omic-level measurements to do systems biology
  - In fact, sometimes much is learned by “simple models” and experiments
    - E.g. Shen-Orr et al., Nat Genetics, 2002; Chang and Ferrell, Nature, 2013; Lim, Nat. Rev. Mol Cell Biol, 2010
  - Hence why we spend half the course on more “low-throughput” methods
- Spanning across systems and scales
  - Whether small or large scale, usually systems biology experiments aim to answer questions that go beyond traditional boundaries of biological disciplines, e.g.
    - Instead of looking at single proteins in a signaling pathways, understanding how coordinate regulation of many proteins within a pathway give rise to observed biological behavior and context-dependent phenotype
    - How does tissue level function (such as shape) arise from molecular level phenomena in individual cells?
  - This focus on molecular mechanisms is one way in which systems biology seems to be different from traditional physiology (the jury is still out...).

# Important Features of Any Experiment

- Experiments are expensive and time consuming
- A very important part of any experiment is having a clear and precise question (hypothesis) that the experiment is intended to address.
  - If I answer this question, will the answer be significant?
  - Will the answer lead to new knowledge that drives the field forward or accomplishes an important business goal?
- There are many possible questions, very few will be worthwhile to investigate!

# Important Features of Any Experiment

- Given a specific question, one can then come up with answers to the following three key properties of the experiment:
  1. What biological system?
    - E.g. Do I look at human cell lines, a mouse, yeast, etc.?
  2. What perturbation/treatment conditions?
    - E.g. What compounds should I apply to the system to elicit a relevant response?
  3. What measurements?
    - E.g. What transcripts do I need to look at, and/or do I need to look at protein levels instead?
- Often (but not always), if you can't design an experiment that only has a handful of conditions and measurements, results may be difficult to interpret
  - Usually the question is too complex or not significant
  - Exceptions are sometimes screening-based studies, but those also typically have a specific question of interest

# Important Features of Any Experiment

- Positive and Negative Controls
  - You never know when your experiment will fail, but at some point it **will** happen.
  - You will want to know why it failed when it does.
  - Therefore always include positive and negative controls, no matter how simple the experiment.
- Replicates
  - Surprisingly, many studies in biology do not perform a sufficient number of replicates.
  - Especially with quantitative data, one needs AT LEAST three replicates, to estimate the mean and standard deviation of the measurement, and do proper statistical inference or hypothesis testing
  - If the experiment is too expensive to do replicates, but still worthwhile to do, then careful consideration is needed about caveats.
    - E.g. How we will determine significance of the result?

# Next Time—Biological Systems