

SBCNY

NIGMS funded Center

Experimental Methods in Systems Biology

Part of the Coursera Certificate in Systems Biology

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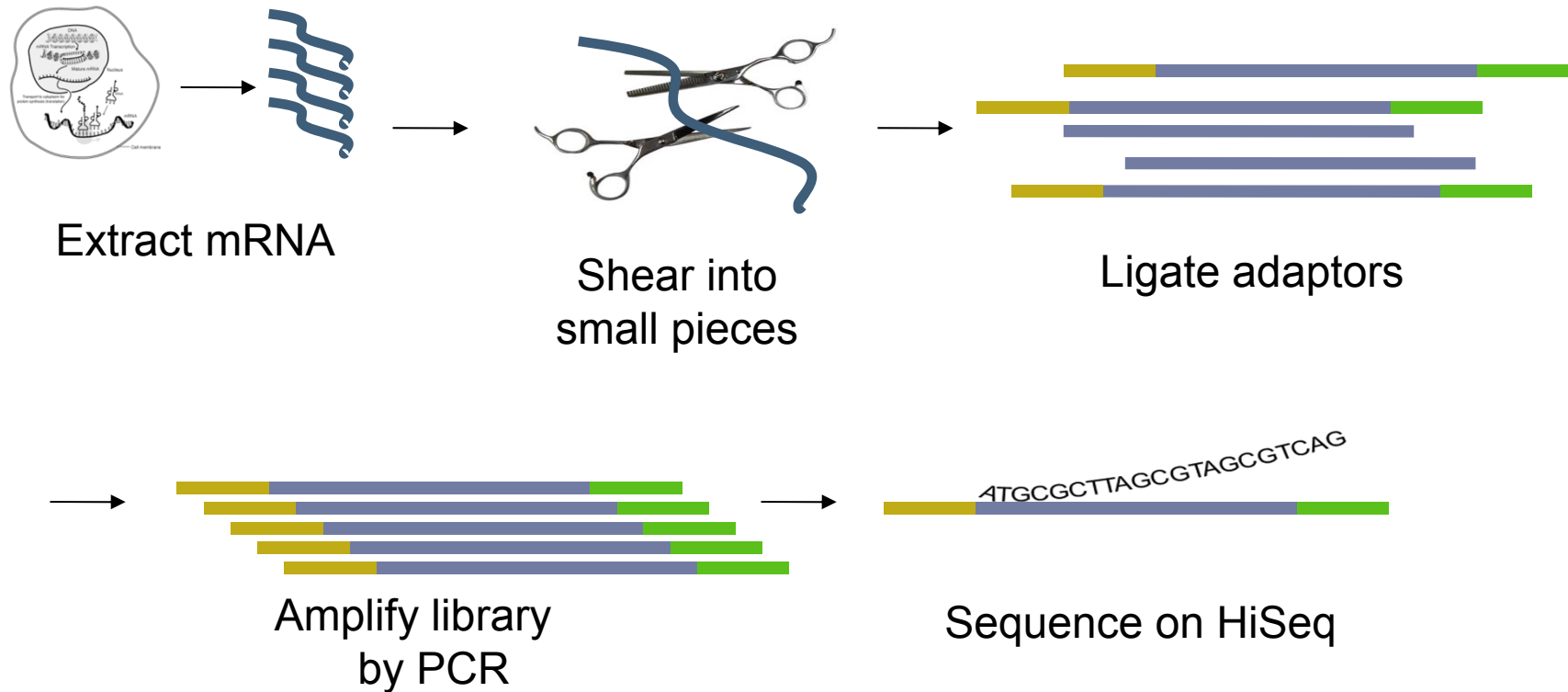
Fall 2014, Week 2, Deep mRNA Sequencing



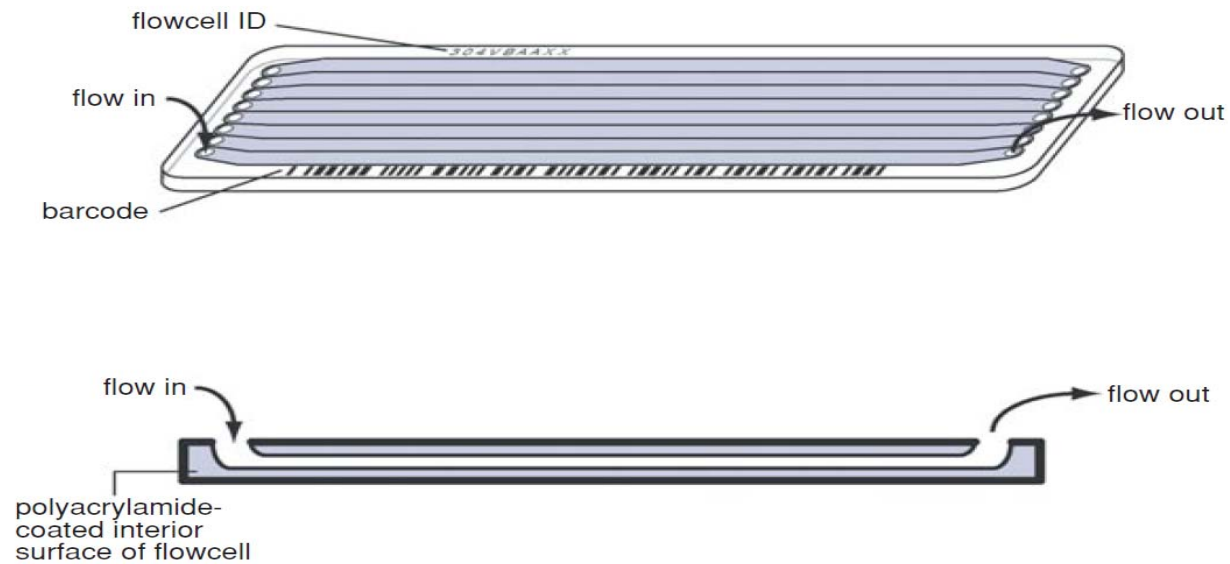
Icahn School
of Medicine at
**Mount
Sinai**

The Illumina Proccotol

A typical Illumina mRNA sequencing process

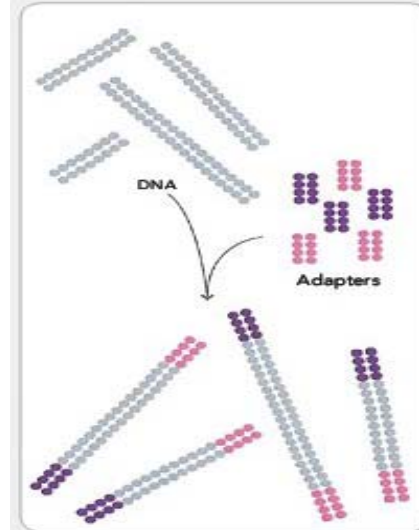


Illumina sequencing

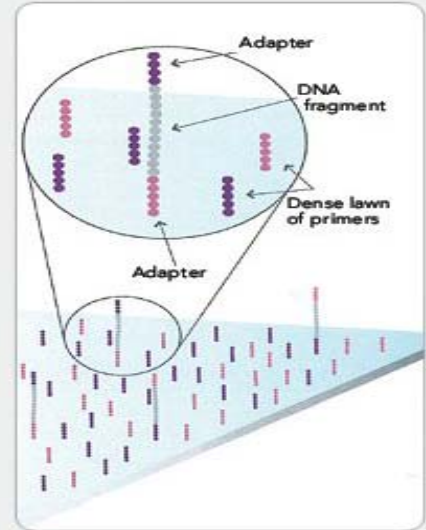


Layering Seq library on the chip (flowcell) surface

Prepare sample

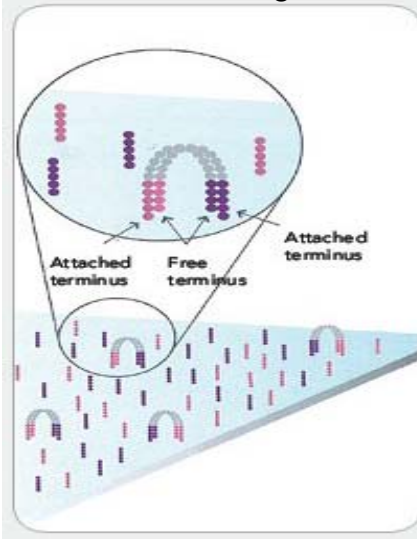


Attach ssDNA to surface

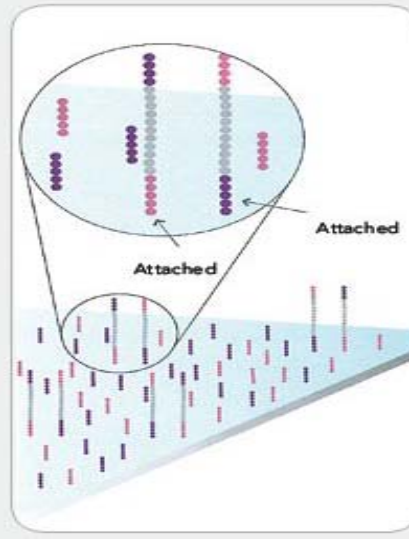


Preparation of DNA on flowcell

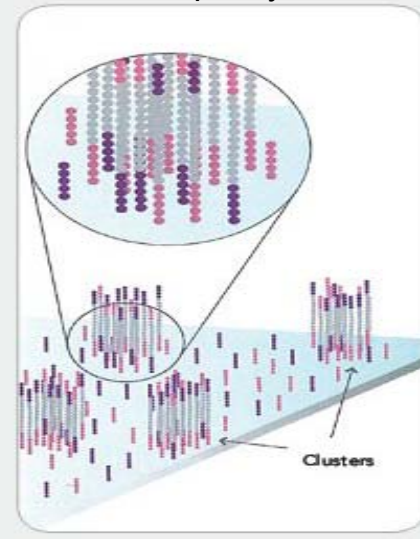
Double-strand fragments



Denature

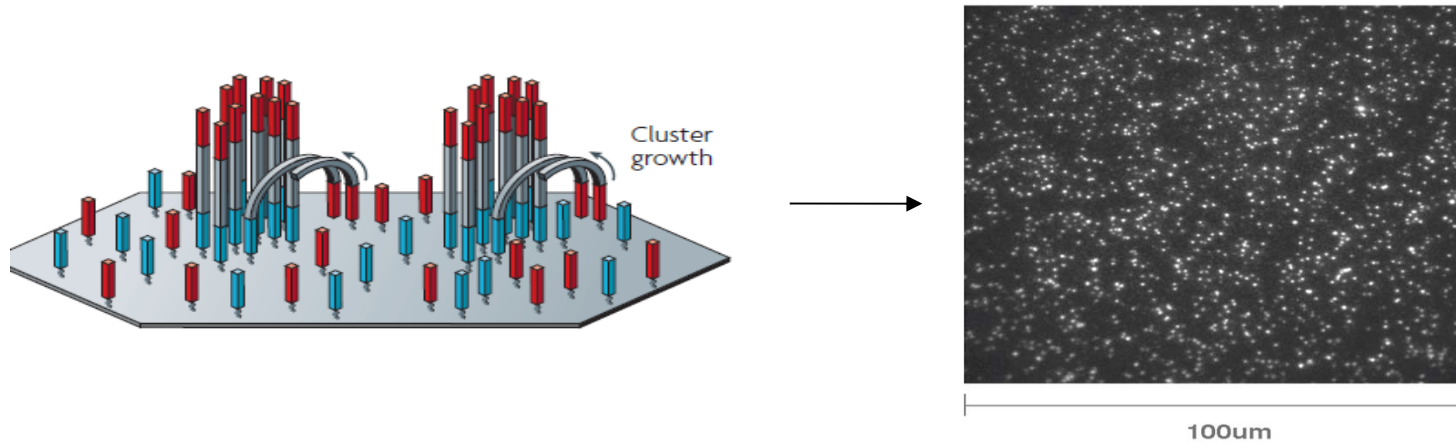


After multiple cycles



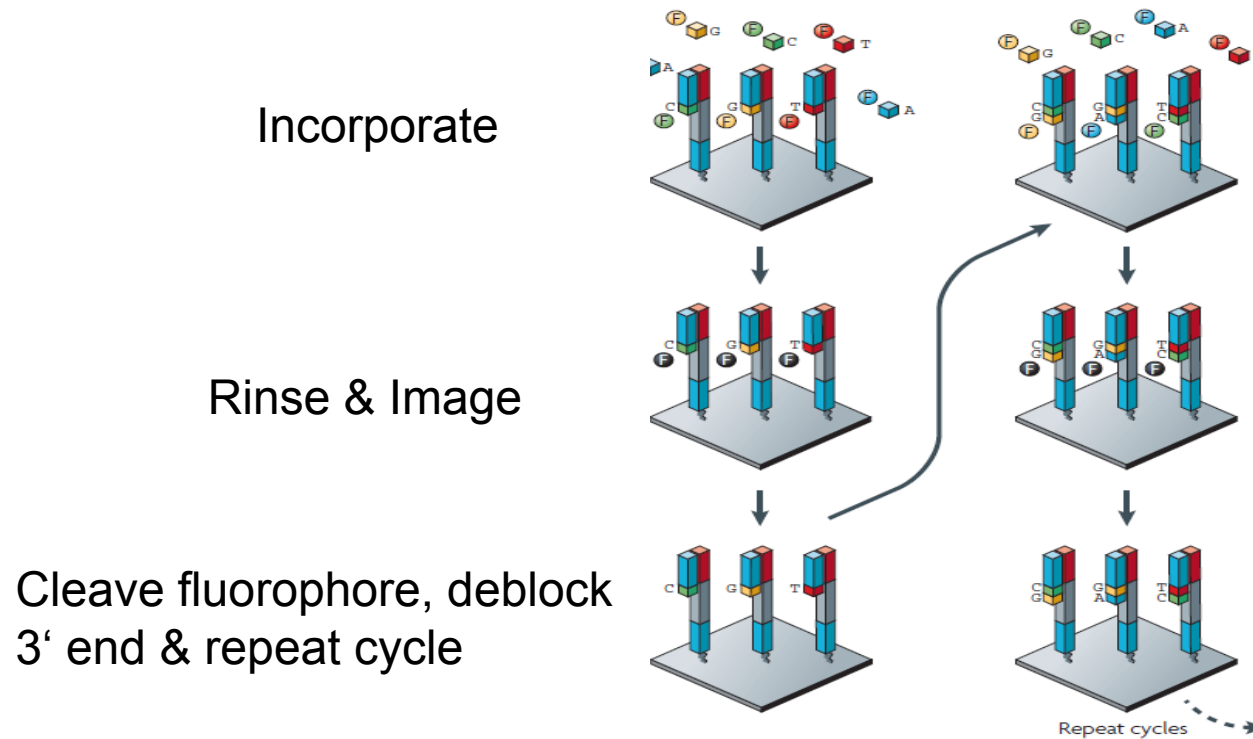
Isothermal “Bridge PCR” with immobilized primers and chemical denaturation

Visualization of clusters



- Position of initial strand is random
- Must avoid overlapping bridges
- Length of library strand matters

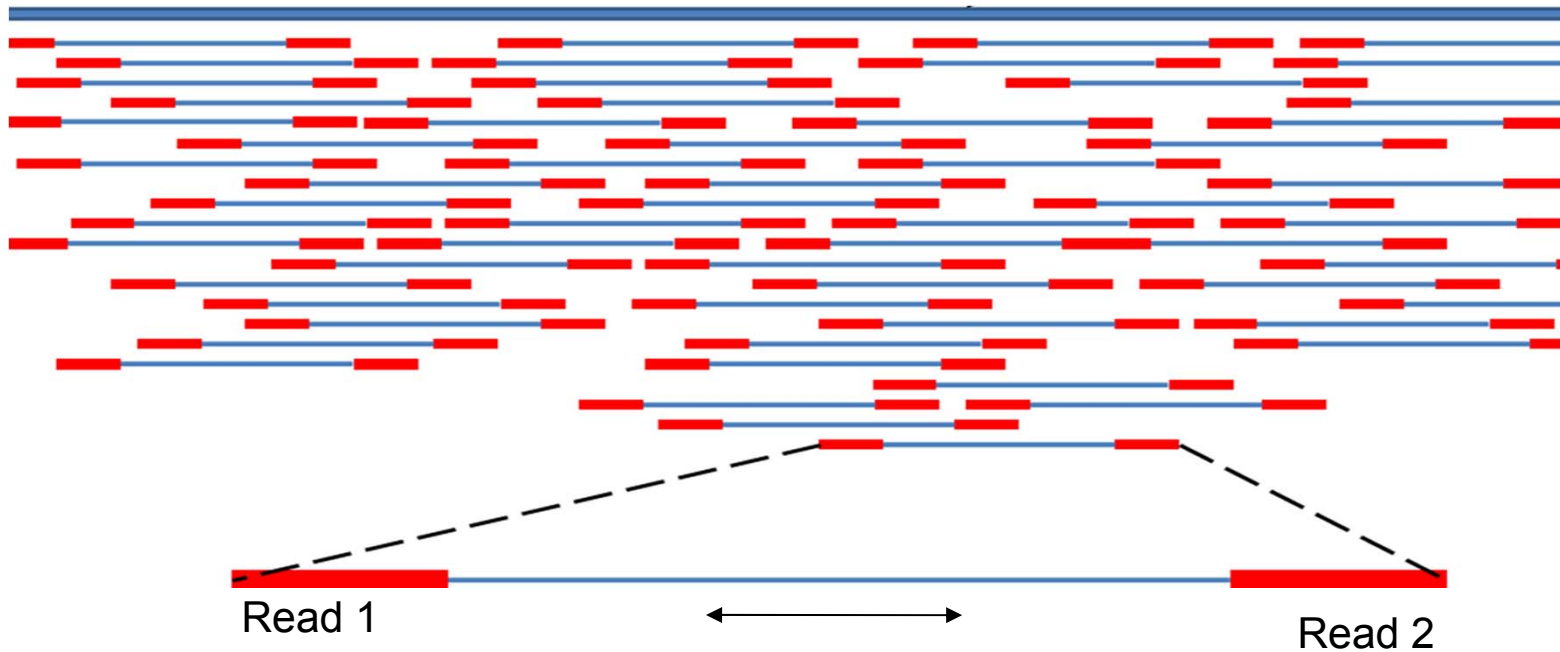
Sequencing reaction



Read length is the number of repeats. The whole fragment is usually not sequenced (fragment length > read length)

Single-end vs. Paired-end

Reference genome

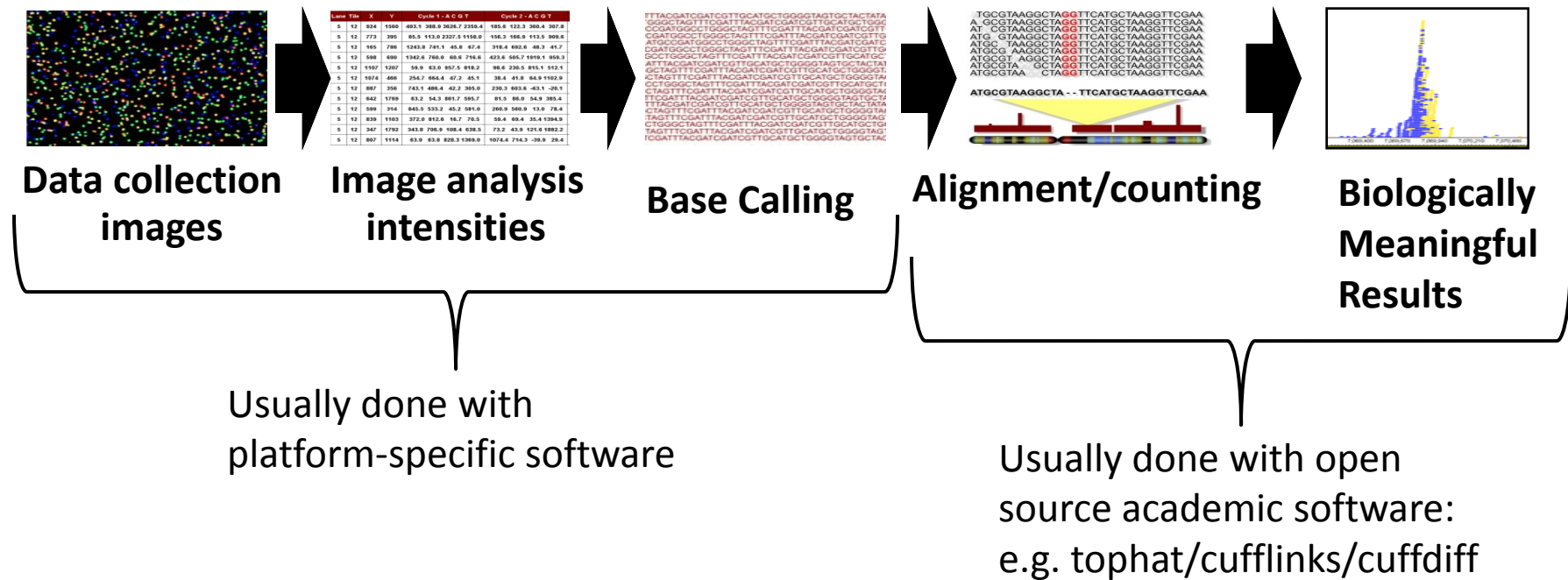


Paired end gives a constraint on length & position

For mRNAseq gives great advantage for alternative splicing analysis

However it doubles the cost and running time

Illumina Seq Data analysis: Overview



mRNA Seq Quantification

- Counts
 - Simply count the number of reads that align to a transcript
 - Biased by transcript length
 - Not comparable between experiments
 - Different sequencing depth (number of clusters identified, sequenced and mapped)
- RPKM—Reads per kilobase of transcript length per million mapped reads
 - Addresses the above problems with pure count data
- FPKM—Fragments per kilobase of transcript length per million mapped reads
 - Applicable for paired end experiments
 - Sometimes only one end of a read is uniquely assignable; FPKM corrects for this
- Cufflinks software will calculate transcript abundances with uncertainties
 - Uncertainties arise from randomness of where reads fall in relation to splicing junctions
 - The mathematical model underlying cufflinks calculations is well-built and an excellent read but is outside the scope of this course (buried in the supplement of Trapnell et al., Nat Biotech, 2008)
 - They have updated it to attempt to also correct for biases in the PCR steps and capture steps
- There are many statistical nuances of differential expression analysis which are also outside the scope of this course but Cuffdiff software addresses many of them.