Project 1

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1

a)

Let $Y \sim Poisson(\lambda)$ where

$$y_i \sim_{iid.} Poisson(\lambda_i)$$

The Canonical choice of link function for the Poisson distribution is

$$\eta_i = g(\lambda_i) \\
= log(\lambda_i) \\
= X_i^T \beta$$

Log-likelihood:

The likelihood is given by:

$$\begin{split} L(\lambda) &= P(Y_1 = y_1, ..., Y_n = y_n) \\ &= [y_i\text{'s are indep.}] \\ &= P(Y_1 = y_1) \cdot ... \cdot P(Y_n = y_n) \\ &= \frac{\lambda_i^{-y_1}}{y_1!} e^{-\lambda_i} \cdot ... \cdot \frac{\lambda_i^{-y_n}}{y_n!} e^{-\lambda_i} \\ &= \prod_{i=1}^{\infty} \frac{\lambda_i^{-y_i}}{y_i!} e^{-\lambda_i} \end{split}$$

Taking the log of the likelihood gives:

$$\begin{split} l(\beta) &= ln(L(\beta)) \\ &= ln(\prod_{i=1}^{n} \frac{\lambda_i^{-y_i}}{y_i!} e^{-\lambda_i}) \\ &= \sum_{i=1}^{n} (y_i ln(\lambda_i) - \lambda_i - ln(y_i!)) \\ &= \sum_{i=1}^{n} (y_i ln(e^{\eta_i}) - e^{\eta_i} - ln(y_i!)) \\ &= \sum_{i=1}^{n} (y_i ln(e^{X_i^T \beta}) - e^{X_i^T \beta} - ln(y_i!)) \end{split}$$

Because y_i do not depend on β_i , we can omit $ln(y_i!)$ when finding the score function.

Score function:

$$s(\beta) = \frac{\partial}{\partial \beta} l(\beta)$$

$$= \frac{\partial}{\partial \beta} \left(\sum_{i=1}^{n} (y_i ln(e^{x_i^T \beta}) - e^{x_i^T \beta}) \right)$$

$$= \sum_{i=1}^{n} (y_i x_i - x_i e^{x_i^T \beta})$$

$$= \sum_{i=1}^{n} (y_i - e^{x_i^T \beta}) x_i$$

Observed Fisher information:

$$H(\beta) = -\frac{\partial^2}{\partial \beta \partial \beta^T} l(\beta)$$

$$= -\frac{\partial}{\partial \beta} s^T(\beta)$$

$$= -\sum_{i=1}^n \frac{\partial}{\partial \beta} ((y_i - e^{x_i^T \beta}) x_i^T)$$

$$= \sum_{i=1}^n x_i x_i^T e^{x_i^T \beta}$$

$$= X^T W X$$

where $W = diag(e^{X_1^T\beta},...,e^{X_n^T\beta}) = diag(e^{\eta_1},...,e^{\eta_n})$

Expected Fisher information:

$$F(\beta) = Var[s(\beta)]$$

$$= Var[\sum_{i=1}^{n} (y_i - e^{x_i^T \beta})x_i]$$

$$= \sum_{i=1}^{n} x_i Var[y_i - e^{x_i^T \beta}]x_i^T$$

$$= \sum_{i=1}^{n} x_i Var[y_i]x_i^T$$

$$= \sum_{i=1}^{n} x_i \lambda_i x_i^T$$

$$= X^T W X$$

where W is the same as in the observed fisher information

b)

```
myglm <- function(formula, data, start = 0) {</pre>
   # Defining y and X:
   y <- data$y
   X <- model.matrix(formula, data)</pre>
   # Defining beta:
   beta_0 <- rep(start, ncol(X))</pre>
   beta <- solve(t(X) %*% X) %*% t(X) %*% y
   # Fisher scoring algorithm:
   while (TRUE) {
     beta_0 <- beta
     eta <- as.vector((X %*% beta))</pre>
     scoreF <- t(X) %*% (y - exp(eta))</pre>
     A <- diag(exp(eta))
     fisherInf <- t(X) %*% A %*% X
     if (all(abs(scoreF) < 1e-04)) {</pre>
       break}
     beta <- beta + solve(fisherInf) %*% scoreF
   # Deviance:
   eta <- as.vector((X %*% beta))</pre>
   yhat <- exp(eta)</pre>
   deviance <- 2 * sum(dpois(y, y, TRUE) - dpois(y, yhat, TRUE))</pre>
   # Estimated variance matrix:
   vcov <- solve(fisherInf)</pre>
   # Coefficients
   coefficients <- matrix(, nrow = 3, ncol = 2)</pre>
   colnames(coefficients) <- c("Estimate", "Std. Error")</pre>
   rownames(coefficients) <- c("(Intercept)", "t^2", "t")</pre>
   coefficients[,1] <- beta</pre>
   coefficients[,2] <- sqrt(diag(vcov))</pre>
   return (list(coefficients = coefficients, deviance = deviance, vcov = vcov))
}
```

c)

We now compare the "myglm" function to the built in "glm" and "vcov" functions in R: This will be done with the simulated data obtained from the "rpois" function.

Data:

```
set.seed(999) t1 \leftarrow c(16,15,17,18,15,14,16,17,18,15) \# t^2 t2 \leftarrow c(16,14,19,18,15,14,16,17,18,15) \# t testData \leftarrow data.frame(y = rpois(10, 10),t1,t2)
```

Our function:

```
testOur <- myglm(y ~ ., testData)</pre>
testOur
## $coefficients
                    Estimate Std. Error
## (Intercept) 3.3556208985 1.3908842
              -0.0730511905 0.2089459
               0.0001284135 0.1631588
## t
##
## $deviance
## [1] 8.166999
##
## $vcov
##
               (Intercept)
## (Intercept) 1.93455873 -0.19419883 0.07346046
               -0.19419883 0.04365840 -0.03134958
## t1
## t2
               0.07346046 -0.03134958 0.02662080
```

Built in functions:

```
testBuilt <- glm(y ~ ., family = poisson(link = log), data = testData)
summary(testBuilt)$coefficients</pre>
```

```
## Estimate Std. Error z value Pr(>|z|)
## (Intercept) 3.3556200137 1.3908835 2.4125816552 0.01583999
## t1 -0.0730511496 0.2089459 -0.3496175622 0.72662573
## t2 0.0001284232 0.1631588 0.0007871056 0.99937198
```

```
summary(testBuilt)$deviance
```

```
## [1] 8.166999
```

```
vcov(testBuilt)
```

```
## (Intercept) t1 t2
## (Intercept) 1.93455691 -0.19419866 0.07346041
## t1 -0.19419866 0.04365838 -0.03134957
## t2 0.07346041 -0.03134957 0.02662079
```

We get the same results.

 $\mathbf{2}$

We are given a Poisson distribution with $\lambda_i = \lambda_0 e^{\frac{-(t_i - \theta)^2}{2\omega^2}}$. λ_i are the number of fledglings at time i (number of days after April 1)

a)

When $\theta = t_i$, the exponent of the e is zero, so $\lambda_i = \lambda_0$. This means that λ_0 is the maximum expected number of fledglings leaving the nest when $t_i = \theta$.

The ω determine the growth of the function. If it is large the function will decrease slower.

The function reaches its maximum value when the exponent is 0, i.e $t_i = \theta$. This means that the θ is the optimal time for breeding.

b)

This is a GLM because it is Poisson distributed.

Know that $y \sim Poisson(\lambda_0 e^{\frac{-(t_i - \theta)^2}{2\omega^2}})$

$$\lambda_i = \lambda_0 e^{\frac{-(t_i - \theta)^2}{2\omega^2}}$$
$$= e^{\eta}$$
$$= e^{x_i^T \beta}$$

Taking ln on both sides:

$$x_i^T \beta = \ln(\lambda_0) - \frac{(t_i - \theta)^2}{2\omega^2}$$

$$= \ln(\lambda_0) - \frac{1}{2} \left(\frac{t_i^2}{\omega^2} - \frac{2t_i \theta}{\omega^2} + \frac{\theta^2}{\omega^2}\right)$$

$$= \ln(\lambda_0) - \frac{\theta^2}{2\omega^2} + \frac{\theta}{\omega^2} t_i - \frac{1}{2\omega^2} t_i^2$$

$$= \beta_0 + \beta_1 t + \beta_2 t^2$$

So

$$\beta_0 = \ln(\lambda_0) - \frac{\theta^2}{2\omega^2}$$
$$\beta_1 = \frac{\theta}{\omega^2}$$
$$\beta_2 = \frac{1}{2\omega^2}$$

c)

```
load(url("https://www.math.ntnu.no/emner/TMA4315/2022h/hoge-veluwe.Rdata")) # Defaults to "data"
testOur <- myglm(y ~ I(t^2) + t, data)
testOur
## $coefficients
                  Estimate Std. Error
## (Intercept) 1.420130462 0.282434733
              -0.003298608 0.001019464
## t^2
               0.085183057 0.034053955
## t
##
## $deviance
## [1] 277.4613
##
## $vcov
##
                (Intercept)
                                  I(t^2)
## (Intercept) 0.0797693783 2.550195e-04 -9.308596e-03
## I(t^2)
               ## t
              -0.0093085957 -3.369024e-05 1.159672e-03
# Just to check if it works on this data aswell
# testBuilt \leftarrow glm(y \sim I(t^2) + t, family = poisson(link = log), data)
# summary(testBuilt)
# vcov(testBuilt)
```

We see that:

$$\hat{\beta}_0 = 1.42, \qquad \hat{\beta}_1 = 0.085, \qquad \hat{\beta}_2 = -0.0033$$

d)

To test if there is a quadratic effect of t (at a 0.05-level of significance), we test

$$H_0: \hat{\beta}_2 = 0$$
 vs $H_1: \hat{\beta}_2 \neq 0$

using the Wald-test.

The test statistic under H_0 is then: $z = \frac{\hat{\beta}_2}{\hat{\sigma}_{\beta_2}} \sim N(0,1)$, where $\hat{\sigma}_{\beta_2}$ is the estimated standard deviation. At a 0.05-level of significance, H_0 is rejected if |z| > 1.96.

Testing:

```
z = testOur$coefficients[2,1] / (sqrt(testOur$vcov[2,2]))
# z-value from Wald-test
abs(z)
```

```
## [1] 3.235631
```

Here we see that H_0 is rejected, so this indicates that there is in fact a quadratic effect of t (at a 0.05-level of significance)

e)

We have that the deviance for our model is 277.4613. To test if this model is OK, we have the test:

```
H_0: model \ ok \qquad vs \qquad H_1: model \ not \ ok
```

Under H_0 , the deviance is approx. χ^2_{n-p} , where n is the amount of observations and p is the number of parameters.

In our case, n = 135 and p = 3, and H_0 is then rejected if $D > \chi^2_{0.95, n-p}$

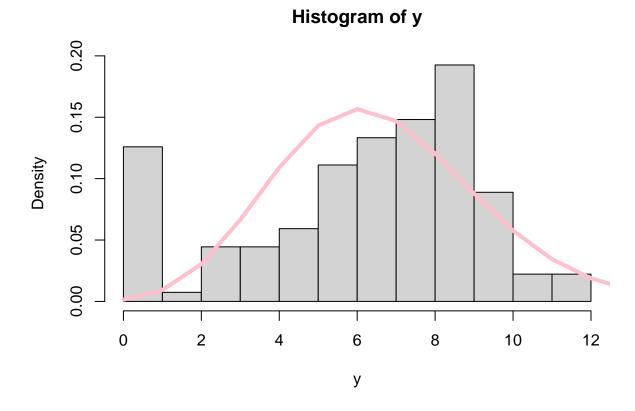
```
criticalValue <- qchisq(0.95, dim(data)[1] - length(testOur$coefficients[,1]))
criticalValue</pre>
```

```
## [1] 159.8135
```

We see that our deviance is larger than the critical value, so our model is probably not the best fit for our data.

Our assumptions for y is that it is Poisson distributed. Looking at the observed density and the poisson distribution given :

```
hist(data[,1], freq=F, main = "Histogram of y", xlab = "y")
lines(x = seq(0,20), dpois(x = seq(0,20), lambda = mean(data$y)), col="pink", lwd = 4)
```



Here the pink line is $Poisson(\lambda = \bar{y})$ and the bar-plot is the density of our data. Looking at this histogram we see that the assumption that y is Poisson distributed does not hold.

f)

We see from 2b) that

$$\hat{\beta_1} = \frac{\theta}{\omega^2}, \qquad \hat{\beta_2} = \frac{1}{2\omega^2}$$

This can be made into:

$$\hat{\omega} = \sqrt{\frac{1}{2\hat{eta}_2}} \qquad and \qquad \hat{\theta} = \hat{eta}_1 \hat{\omega}^2$$

Delta method is the following:

For omegas

$$\begin{split} Var[\hat{\omega}] &= (\frac{\partial \hat{\omega}}{\partial \hat{\beta}_2})^2 \ Var[\hat{\beta}_2] \\ &= (-\frac{1}{2\sqrt{2\hat{\beta}_2^3}})^2 \ Var[\hat{\beta}_2] \end{split}$$

Using this, the standard error for the omegas is then given by $SE[\hat{\omega}] = \sqrt{Var[\hat{\omega}]}$ For thetas

$$Var[\hat{\theta}] = (\frac{\partial \hat{\theta}}{\partial \hat{\beta}_{1}})^{2} Var[\hat{\beta}_{1}] + (\frac{\partial \hat{\theta}}{\partial \hat{\beta}_{2}})^{2} Var[\hat{\beta}_{2}] + 2 (\frac{\partial \hat{\theta}}{\partial \hat{\beta}_{1}}) (\frac{\partial \hat{\theta}}{\partial \hat{\beta}_{2}}) Cov[\hat{\beta}_{1}, \hat{\beta}_{2}]$$

$$= (\frac{1}{2\hat{\beta}_{2}})^{2} Var[\hat{\beta}_{1}] + (-\frac{\hat{\beta}_{1}}{2\hat{\beta}_{2}^{2}})^{2} Var(\hat{\beta}_{2}) + 2 (\frac{1}{2\hat{\beta}_{2}}) (-\frac{\hat{\beta}_{1}}{2\hat{\beta}_{2}^{2}})^{2} Cov[\hat{\beta}_{1}, \hat{\beta}_{2}]$$

Using this, the standard error for the thetas is then given by $SE[\hat{\theta}] = \sqrt{Var[\hat{\theta}]}$

```
omegaHat <- sqrt(abs(1/(2*testOur$coefficients[2,1])))
thetaHat <- omegaHat^2 * testOur$coefficients[3,1]
cat("omega hat:", omegaHat, "theta hat:", thetaHat)</pre>
```

omega hat: 12.31175 theta hat: 12.91197

```
omegaHat_var <- (-1/(2 * sqrt(abs(2 * testOur$coefficients[2,1]^3))))^2 * testOur$vcov[2,2]
omegaHat_se <- sqrt(omegaHat_var)

thetaHat_var <- (1/(2*testOur$coefficients[2,1]))^2*testOur$vcov[3,3] + ((testOur$coefficients[3,1])/(2
thetaHat_se <- sqrt(thetaHat_var)

cat("SE of omega:", omegaHat_se, "SE of theta:", thetaHat_se)</pre>
```

SE of omega: 1.902526 SE of theta: 1.609386

We get

$$\hat{\omega} = 12.31175$$
 and $\hat{\theta} = 12.91197$

and

$$SE[\hat{\omega}] = 1.902526$$
 and $SE[\hat{\theta}] = 1.609386$

 \mathbf{g}

From point 2.f we know that the estimate of the optimal breeding time $\hat{\theta}$ is 12.91. Instead, looking at the data we can easily compute the mean value of the breeding dates (i.e. by extracting data\$t) that is equal to 15.937. This difference can be interpreted as the result of a faster globally environmental change on the weather and a faster increase in temperature in the spring months compared to the evolutionary response. According to this intuitive idea, we can make a test in order to verify if the mean value of t is significantly different from the estimated optimal date based on the fitted model. So, we define a new variable $\hat{z} = \mu - \hat{\theta}$, where μ is the mean value of the column representing the breeding times and $\hat{\theta}$ is the same item as above. We can proceed with the test: $H_0: \hat{z} = 0$ vs $H_1: \hat{z} \neq 0$ Taking into account the assumption of gaussianity of \hat{z} we can use the Z-test under H_0 . We then have

$$Z = \frac{\hat{z}}{\hat{\sigma}_z}$$

with $Z \sim N(0,1)$. As regards the computation of $\hat{\sigma}_z$ we need an extra hypothesis, we assume that μ and $\hat{\theta}$ are independent, so from basic theory of statistic we know that:

$$\hat{\sigma}_z = \sqrt{Var[\mu] + Var[\hat{\theta}]}$$

From this equation we can find the value of our statistic:

```
mu = mean(data$t)
n = nrow(data)
var_mu = var(data$t)/n
std_err = sqrt(var_mu + thetaHat_var)
Z_val = (mu - thetaHat) / std_err
Z_val
```

[1] 1.821029

That is Z=1.821. It is known that for a test with 0.05 significance level the null hypothesis is rejected for a value of |Z|>1.96. Hence we do not have evidence to reject H_0 and to assert that there exist a significant difference between μ and $\hat{\theta}$.

h)

```
set.seed(999)
lambda <- function(t, thetaHat, c){</pre>
   \exp(\text{testOur}\text{scoefficients}[1,1] + (\text{thetaHat}^2)/(2 * \text{omegaHat}^2)) * \exp(-((t - \text{thetaHat})^2)/(2 * \text{omegaHat}^2))
}
B = 1000
bootpred <- rep(NA, B)
beta0s <- rep(NA,B)</pre>
beta1s <- rep(NA,B)
beta2s <- rep(NA,B)
for (i in 1:B){
   lambdab <- mapply(lambda, data$t, thetaHat, omegaHat)</pre>
   y <- rpois(nrow(data), lambdab)
   data_new <- data.frame(y,data$t)</pre>
   names(data_new)[2] <- "t"</pre>
   testOur_new <- myglm(y ~ I(t^2) + t, data_new)</pre>
   beta0s[i] <- testOur_new$coefficients[1,1]</pre>
   beta1s[i] <- testOur_new$coefficients[3,1]</pre>
   beta2s[i] <- testOur_new$coefficients[2,1]</pre>
}
betabootstrap <- t(cbind(var(beta0s), var(beta2s), var(beta1s)))</pre>
colnames(betabootstrap) <- c("Variance")</pre>
rownames(betabootstrap) <- c("(intercept) bootstrap", "I(t^2) bootstrap", "t bootstrap")
betabootstrap
##
                                Variance
## (intercept) bootstrap 7.982436e-02
## I(t^2) bootstrap 1.042069e-06
## t bootstrap
                           1.151188e-03
Comparing this result to the variances obtained from 2c):
betaOur0 <- testOur$vcov[1,1]</pre>
betaOur2 <- testOur$vcov[2,2]</pre>
betaOur1 <- testOur$vcov[3,3]</pre>
betaOur <- t(cbind(betaOur0, betaOur2, betaOur1))</pre>
colnames(betaOur) <- c("Variance")</pre>
rownames(betaOur) <- c("(intercept)", "I(t^2)", "t")</pre>
beta0ur
                     Variance
## (intercept) 7.976938e-02
## I(t^2) 1.039306e-06
                1.159672e-03
## t
```

Calculating the differences:

```
diff <- abs(betaOur - betabootstrap)
rownames(diff) <- c("diff (intercept)", "diff I(t^2)", "diff t")
colnames(diff) <- c("Difference")
diff</pre>
```

```
## Difference
## diff (intercept) 5.498462e-05
## diff I(t^2) 2.762294e-09
## diff t 8.483787e-06
```

We see that we get pretty similar results.