

# **PROTOCOL EXCHANGE | COMMUNITY CONTRIBUTED** Isolation of extracellular vesicle RNA from cell culture supernatant

**Irene Yan & Tushar Patel**

**Patel Lab, Mayo Clinic**

## **Abstract**

The study of extracellular RNA has been recently reported as an important mechanism of intercellular signaling. Extracellular vesicles which contain protein, mRNA, and non-coding RNA can be isolated from cell culture supernatant. Extracellular vesicle RNA isolated from tumor cells in culture can be a useful tool for analyzing the role of either extracellular vesicles or extracellular vesicle RNA. This protocol presents a method of isolating extracellular vesicle RNA from human cholangiocarcinoma cells in culture.

**Subject terms:** Isolation, Purification and Separation Cell biology Cell culture  
Nucleic acid based molecular biology Tissue culture

**Keywords:** Extracellular vesicles Tumor Cells Isolation RNA

## **Reagents**

Reagents:

One of the following commercial kits:

- miCURY RNA Isolation Kit –Biofluids (Exiqon, #300112)
- miRNeasy Mini Kit (Qiagen, #217004)
- SeraMir Exosome RNA Purification Kit (System Biosciences, #RA806TC-1)
- Isopropanol
- Absolute ethanol
- Nuclease free H<sub>2</sub>O

## **Equipment**

Equipment:

- Bench top centrifuge
- Vortex Mixer

- Micropipettes with sterile tips 20-1000
- Sterile collection bottle
- Microcentrifuge tubes
- 0.22  $\mu$ m PES vacuum filter

## Procedure

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1. Grow KMBC cholangiocarcinoma cells on 32×10 cm tissue culture plates. Use Extracellular vesicle (EV)-cleared media for cultures (total 400 ml). Incubate on cells for 48 hr.
2. Collect supernatant and filter through a 0.22  $\mu$ M PES filter
3. Aliquot into ultracentrifugation tubes, and centrifuge for 70 min at 100,000 x g at 4°C.
4. Discard supernatant, and resuspend pellet from each tube with 1 ml PBS.
5. Combine washed pellets into one ultracentrifuge tube, and centrifuge for 70 min at 100,000 x g at 4°C.
6. Discard supernatant, resuspend final EV pellet with 600  $\mu$ l PBS. Store samples at -70°C until ready for use.
7. RNA Isolation can be performed from one of the following options.

### A. miCURY RNA Isolation Kit, Exiqon

- i. Transfer 200  $\mu$ l of EV into 1.5 ml microcentrifuge tube.
- ii. Add 60  $\mu$ l Lysis Solution BF, vortex 5 sec, incubate for 3 min at room temperature.
- iii. Add 20  $\mu$ l Protein Precipitation Solution BF, vortex 5 sec, incubate for 1 min at room temperature.
- iv. Centrifuge at 11,000g for 3 min at room temperature.
- v. Transfer clear supernatant to new microcentrifuge tube.
- vi. Add 270  $\mu$ l Isopropanol, vortex 5 sec.
- vii. Assemble microRNA Mini Spin Column BF in a new collection tube and load entire sample onto column.
- viii. Centrifuge at 11,000g for 30 sec at room temperature. Discard flow-through and return column to collection tube.
- ix. Add 700  $\mu$ l Wash Solution 2 BF to column and centrifuge at 11,000g for 30 sec at room temperature. Discard flow-through and return column to collection tube.
- x. Add 250  $\mu$ l Wash Solution 2 BF to column and centrifuge at 11,000g for 2 min at room temperature. Discard flow-through and return column to collection tube.
- xi. Add 50  $\mu$ l rDNase directly onto membrane of spin column.
- xii. Close lid and incubate for 15 min at room temperature
- xiii. Add 100  $\mu$ l Wash Solution 1 BF to column and centrifuge at 11,000g for 30 sec at room temperature. Discard flow-through and return column to collection tube.
- xiv. Add 700  $\mu$ l Wash Solution 2 BF to column and centrifuge at 11,000g for 30 sec at room

temperature. Discard flow-through and return column to collection tube.

xv. Add 250 µl Wash Solution 2 BF to column and centrifuge at 11,000g for 2 min at room temperature.

xvi. Transfer spin column into a new collection tube.

xvii. Add 50 µl RNase free H<sub>2</sub>O directly onto the membrane of spin column.

xviii. Close lid and incubate for 1 min at room temperature.

xix. Centrifuge at 11,000g for 1 min at room temperature.

xx. Discard spin column and store purified RNA at -70°C.

## B. miRNeasy Mini Kit

i. Transfer 200 µl of EV into 1.5 ml microcentrifuge tube.

ii. Add 700 µl QIAzol Lysis Reagent, vortex 5 sec, incubate for 5 min at room temperature.

iii. Add 140 µl chloroform, shake vigorously for 15 sec, incubate for 3 min at room temperature.

iv. Centrifuge sample at 12,000g for 15 min at 4°C.

v. Transfer the upper aqueous phase to a new microcentrifuge tube.

vi. Add 500 µl 100% Ethanol, mix by pipetting.

vii. Assemble RNeasy Mini Column in a new collection tube and load entire sample onto column.

viii. Centrifuge sample at 8,000g for 15 sec at room temperature. Discard flow-through and return column to collection tube.

ix. Add 700 µl Buffer RPE to column and centrifuge at 8,000g for 15 sec at room temperature.

Discard flow-through and return column to collection tube.

x. Add 500 µl Buffer RPE to column and centrifuge at 8,000g for 2 min at room temperature.

Discard flow-through and return column to collection tube.

xi. Centrifuge column at 8,000 x g for 1 min at room temperature to dry membrane.

xii. Transfer spin column into a new collection tube.

xiii. Add 30 µl RNase free H<sub>2</sub>O directly onto the membrane of spin column.

xiv. Close lid and centrifuge for 1 min at 8,000 x g at room temperature.

xv. Discard spin column and store purified RNA at -70°C.

## C. SeraMir Exosome RNA Purification Kit (Since EV is already isolated by ultracentrifugation, omit ExoQuick-TC precipitation step)

i. Transfer 200 µl of EV into 1.5 ml microcentrifuge tube.

ii. Add 140 µl Lysis Buffer to EV, vortex 15 sec, incubate for 5 min at room temperature.

iii. Add 80 µl 100% Ethanol, vortex 10 sec.

iv. Assemble Spin Column in a new collection tube and load entire sample onto column.

v. Centrifuge sample at 13,000 rpm for 1 min at room temperature. Discard flow-through and return column to collection tube.

vi. Add 400 µl Wash Buffer to column and centrifuge at 13,000 rpm for 1 min at room temperature.

Discard flow-through and return column to collection tube.

- vii. Add 400 µl Wash Buffer to column and centrifuge at 13,000 rpm for 1 min at room temperature. Discard flow-through and return column to collection tube.
- viii. Centrifuge column at 13,000 rpm for 2 min at room temperature to dry membrane.
- ix. Transfer spin column into a new collection tube.
- x. Add 30 µl Elution Buffer directly onto the membrane of spin column.
- xi. Close lid and centrifuge at 2,000 rpm for 2 min at room temperature.
- xii. Increase speed and centrifuge at 13,000 rpm for 1 min at room temperature.
- xiii. Discard spin column and store purified RNA at -70°C.

## Associated Publications

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This protocol is related to the following articles:

- Extracellular vesicle-mediated transfer of long non-coding RNA ROR modulates chemosensitivity in human hepatocellular cancer  
Kenji Takahashi, Irene K. Yan, Takayuki Kogure, Hiroaki Haga, and Tushar Patel
- Involvement of Extracellular Vesicle Long Noncoding RNA (linc-VLDLR) in Tumor Cell Responses to Chemotherapy  
K. Takahashi, I. K. Yan, J. Wood, H. Haga, and T. Patel
- Modulation of hypoxia-signaling pathways by extracellular linc-RoR  
K. Takahashi, I. K. Yan, H. Haga, and T. Patel
- Intercellular nanovesicle-mediated microRNA transfer: A mechanism of environmental modulation of hepatocellular cancer cell growth  
Takayuki Kogure, Wen-Lang Lin, Irene K. Yan, Chiara Braconi, and Tushar Patel

## Author information

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### Affiliations

#### Patel Lab, Mayo Clinic

Irene Yan & Tushar Patel

### Competing financial interests

None

### Corresponding author

Correspondence to: Tushar Patel (patel.tushar@mayo.edu)

### Readers' Comments

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