# Building and evaluation of a PBPK model for inulin in rats

Version	1.0-OSP12.0
based on Model Snapshot and Evaluation Plan	https://github.com/Open-Systems-Pharmacology/Inulin-Model/releases/tag/v1.0
OSP Version	12.0
Qualification Framework Version	3.3

This evaluation report and the corresponding PK-Sim project file are filed at:

https://github.com/Open-Systems-Pharmacology/OSP-PBPK-Model-Library/

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# 1 Introduction

Inulin is a highly hydrophilic polysaccharide which does not distribute into cells and is cleared via glomerular filtration.

Inulin has a considerably smaller solute radius than the proteins which had been used to develop the generic large molecule physiologically based pharmacokinetic (PBPK) model in PK-Sim (Niederalt 2018).

The herein presented evaluation report evaluates the performance of the PBPK model for inulin in rats using the large molecule model in PK-Sim.

The presented inulin PBPK model as well as the respective evaluation plan and evaluation report are provided open-source (https://github.com/Open-Systems-Pharmacology/Inulin-Model).

## 2 Methods

## 2.1 Modeling Strategy

The development of the large molecule PBPK model in PK-Sim® has previously been described by Niederalt et al. (Niederalt 2018). In short, the model was built as an extension of the PK-Sim® model for small molecules incorporating (i) the two-pore formalism for drug extravasation from blood plasma to interstitial space, (ii) lymph flow, (iii) endosomal clearance and (iv) protection from endosomal clearance by neonatal Fc receptor (FcRn) mediated recycling.

For model development and evaluation, PK data were used from compounds with a wide range of solute radii and from different species. The PK data used for parameter estimation were from the following compounds: antibody–drug conjugate BAY 79-4620 in mice (Bayer in house data), antibody 7E3 in wild-type and FcRn knockout mice (Garg 2007, Garg2009), domain antibody dAb2 in mice (Sepp 2015), antibodies MEDI-524 and MEDI-524-YTE in monkeys (Dall'Acqua 2006), and antibody CDA1 in humans (Taylor 2008). The PK data used for model evaluation were from inulin in rats (Tsuji1983) and tefibazumab in humans (Reilly 2005).

The PBPK model including the estimated physiological parameters as described by Niederalt et al. (Niederalt 2018) is available in the Open Systems Pharmacology Suite from version 7.1 onwards.

This evaluation report focuses on the PBPK model for inulin.

Details about input data (physicochemical, in vitro and PK) can be found in Section 2.2.

Details about the structural model and its parameters can be found in Section 2.3.

#### 2.2 Data

## 2.2.1 In vitro / physico-chemical Data

A literature search was performed to collect available information on physicochemical properties of Inulin. The obtained information from literature is summarized in the table below.

Parameter	Unit	Value	Source	Description
MW	g/mol	5000- 5500	Ohno 1978	Molecular weight
r	nm	1.39	Ghandehari 1997	Hydrodynamic solute radius
logP	μМ	< -10	Dubbelboer 2022	Lipophilicity (octanol/water partition coefficient). Inulin is highly hydrophilic. A logP = -10 is insensitively small in the PBPK model.
Kd (FcRn)	μM	999,999		Dissociation constant for binding to FcRn. High value representing no FcRn binding.

#### 2.2.2 PK Data

Published plasma and tissue PK data on inulin in rats were used.

Publication	Description
Tsuji 1983	Plasma and tissue concentrations after i.v. application of 20 and 200 mg/kg inulin in rats (for 200 mg/kg plasma only).

## 2.3 Model Parameters and Assumptions

#### 2.3.1 Absorption

There is no absorption process since inulin was administered intravenously

#### 2.3.2 Distribution

The standard vascular properties of the different tissues (hydraulic conductivity, pore radii, fraction of flow via large pores) and standard lymph and fluid recirculation flow rates from PK-Sim were used (Niederalt 2018).

#### 2.3.3 Metabolism and Elimination

Inulin is renally excreted via glomerular filtration. The standard glomerular filtration rate from the PK-Sim library was used (GFR fraction = 1).

#### 2.3.4 Tissue Concentrations

For the comparison with experimental data the parameters <code>Fraction of blood for sampling</code> used in the Observer for the tissue concentrations were set for all organs to 0.18. This value is based on the parameter identification for different compounds reported in Ref. (Niederalt 2018) for comparison with tissue dissection data. (The parameter <code>Fraction of blood for sampling</code> specifies residual blood in tissue as ratio of blood volume contributing to the measured tissue concentration to the total in vivo capillary blood volume.)

Model Parameter	Value	Unit
Fraction of blood for sampling (all organs)	0.18	

Experimentally, gut concentrations (from duodenum to the cecum) were measured (Tsuji 1983). In the present evaluation report, the experimental gut concentrations were compared to simulated organ concentrations for small and large intestine separately in the goodness of fit plots as well as in the concentration-time profile plot.

#### 2.3.5 Automated Parameter Identification

No drug specific parameters were fitted.

# 3 Results and Discussion

The PBPK model for inulin was evaluated with plasma and tissue PK data from rats.

The next sections show:

- 1. the final model parameters for the building blocks: Section 3.1.
- 2. the overall goodness of fit: Section 3.2.
- 3. simulated vs. observed concentration-time profiles for the clinical studies used for model building and for model verification: Section 3.3.

# 3.1 Final input parameters

The compound parameter values of the final PBPK model are illustrated below.

### **Compound: Inulin**

#### **Parameters**

Name	Value	Value Origin	Alternative	Default
Solubility at reference pH	9999 mg/l	Other-/Dummy value not used in the simulation	Measurement	True
Reference pH	7	Other-/Dummy value not used in the simulation	Measurement	True
Lipophilicity	-10 Log Units	Other-Highly hydrophilic	Measurement	True
Fraction unbound (plasma, reference value)	1	Other-Assumption	Measurement	True
Is small molecule	Yes			
Molecular weight	5500 g/mol	Publication-Ohno1978		
Plasma protein binding partner	Unknown			
Radius (solute)	0.00139 μm	Publication-Ghandehari1997		

#### **Calculation methods**

Name	Value	
Partition coefficients	PK-Sim Standard	
Cellular permeabilities	PK-Sim Standard	

#### **Processes**

Systemic Process: Glomerular Filtration-GFR

Species: Human

**Parameters** 

Name	Value	Value Origin
GFR fraction	1	Publication-Other-Tsuji1983 (DOI: 10.1002/jps.2600721103)

# 3.2 Diagnostics Plots

Below you find the goodness-of-fit visual diagnostic plots for the PBPK model performance of all data used presented in Section 2.2.2.

The first plot shows observed versus simulated plasma concentration, the second weighted residuals versus time.

Table 3-1: GMFE for Goodness of fit plot for concentration in plasma and tissues

Group	GMFE
Plasma concentrations (20 & 200 mg/kg dose)	1.38
Tissue concentrations (20 mg/kg dose)	2.20
All	1.79

- Plasma concentrations (20 & 200 mg/kg dose)
- Tissue concentrations (20 mg/kg dose)

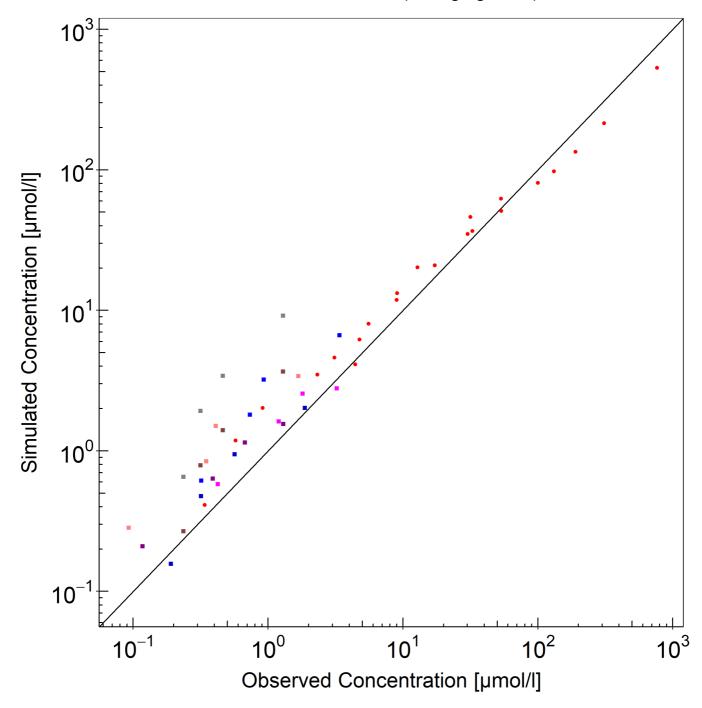


Figure 3-1: Goodness of fit plot for concentration in plasma and tissues

- Plasma concentrations (20 & 200 mg/kg dose)
- Tissue concentrations (20 mg/kg dose)

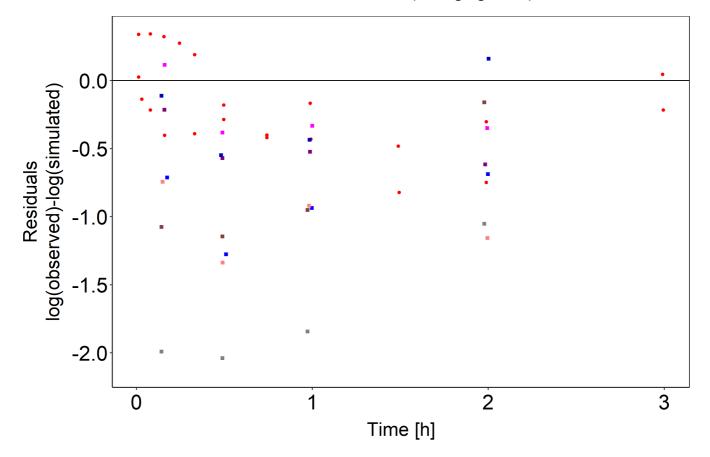


Figure 3-2: Goodness of fit plot for concentration in plasma and tissues

## 3.3 Concentration-Time Profiles

Simulated versus observed concentration-time profiles of all data listed in Section 2.2.2 are presented below.

- Inulin-Venous Blood-Plasma-Concentration
- Tsuji1983.VenousBlood.200-Inulin-VenousBlood-Plasma-Measurement

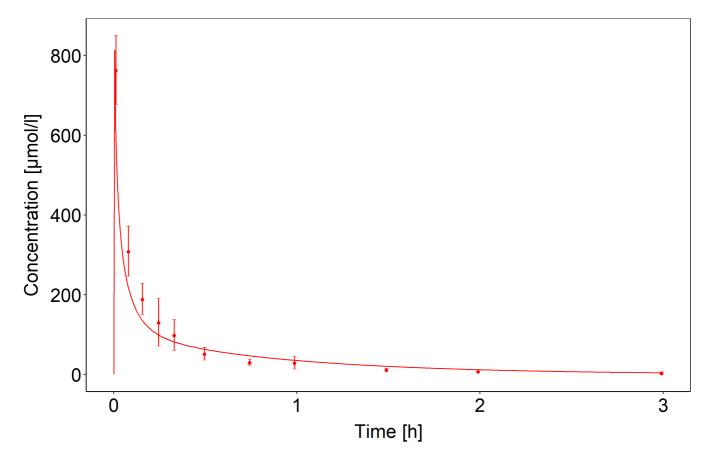


Figure 3-3: Plasma concentration (linear scale)

- Inulin-Venous Blood-Plasma-Concentration
- Tsuji1983.VenousBlood.200-Inulin-VenousBlood-Plasma-Measurement

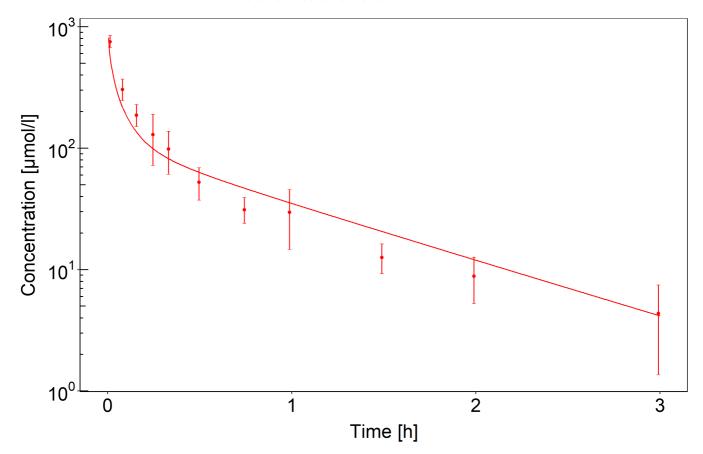


Figure 3-4: Plasma concentration (log scale)

- Inulin-Venous Blood-Plasma-Concentration
- Tsuji1983.VenousBlood.20-Inulin-VenousBlood-Plasma-Measurement

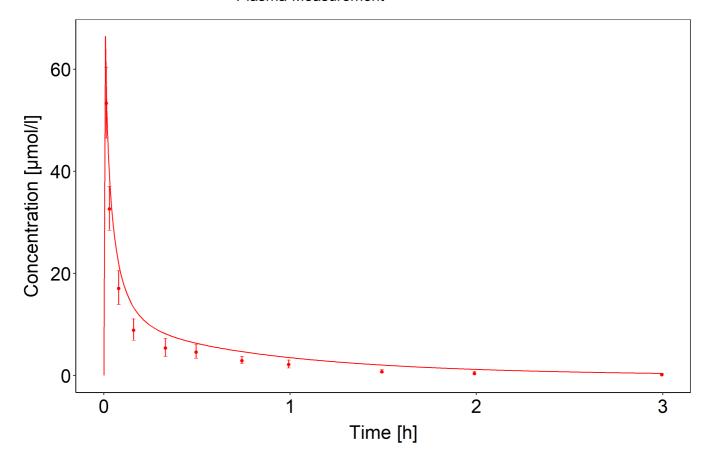


Figure 3-5: Plasma (linear scale)

- Inulin-Venous Blood-Plasma-Concentration
- Tsuji1983.VenousBlood.20-Inulin-VenousBlood-Plasma-Measurement

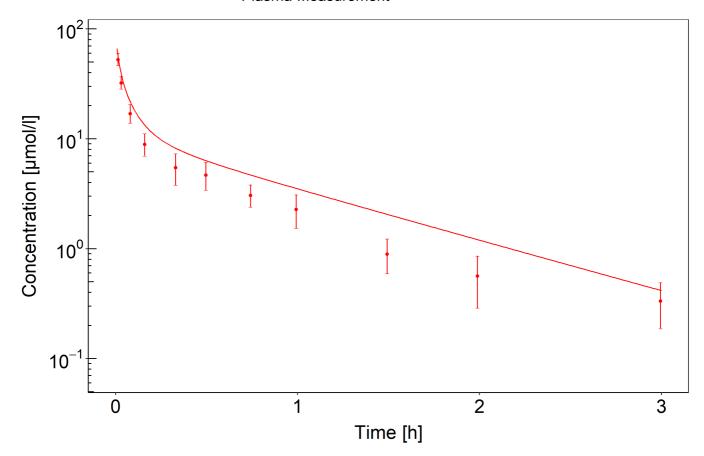


Figure 3-6: Plasma (log scale)

- Inulin-Lung-Whole Organ-Concentration
- Tsuji1983.Lung.20-Inulin-Lung-Tissue-Measurement

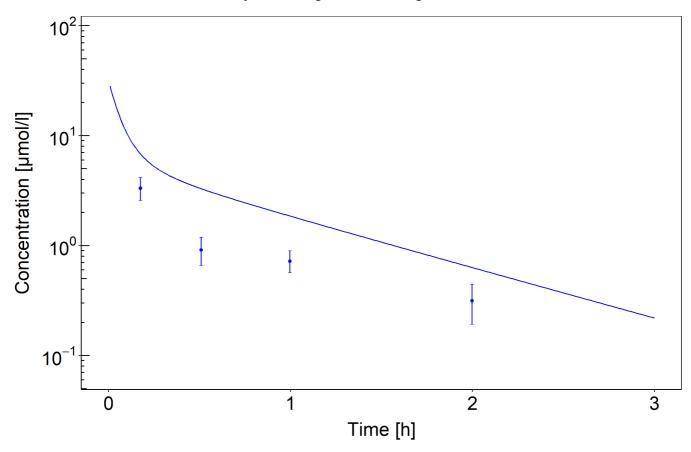


Figure 3-7: Lung

- Inulin-Muscle-Whole Organ-Concentration
- Tsuji1983.Muscle.20-Inulin-Muscle-Tissue-Measurement

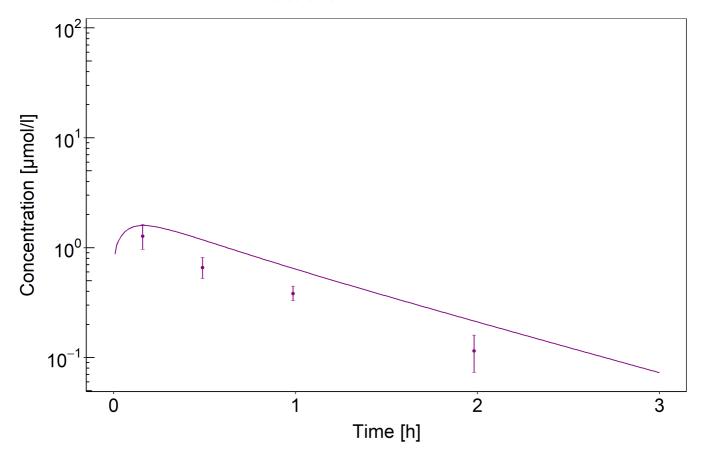


Figure 3-8: Muscle

- Inulin-Bone-Whole Organ-Concentration
- Tsuji1983.Bone.20-Inulin-Bone-Tissue-Measurement

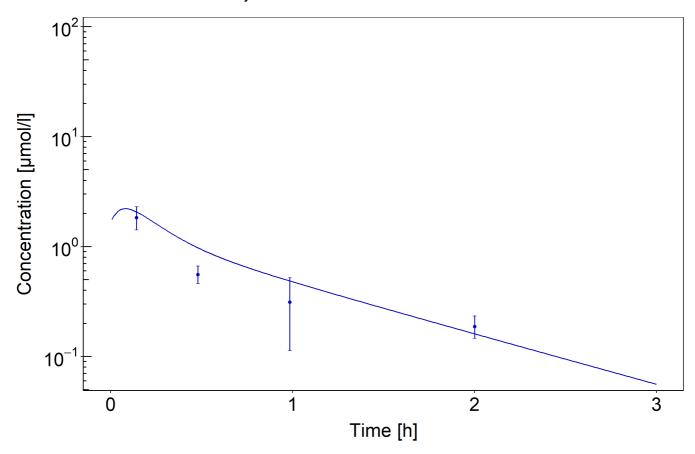


Figure 3-9: Bone

- Inulin-Heart-Whole Organ-Concentration
- Tsuji1983.Heart.20-Inulin-Heart-Tissue-Measurement

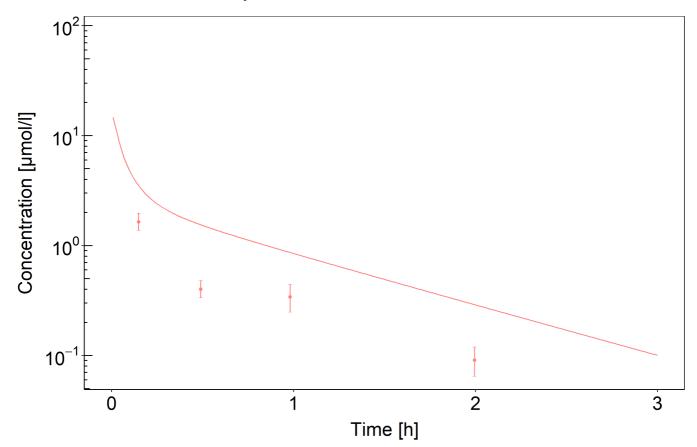


Figure 3-10: Heart

- Inulin-Skin-Whole Organ-Concentration
- Tsuji1983.Skin.20-Inulin-Skin-Tissue-Measurement

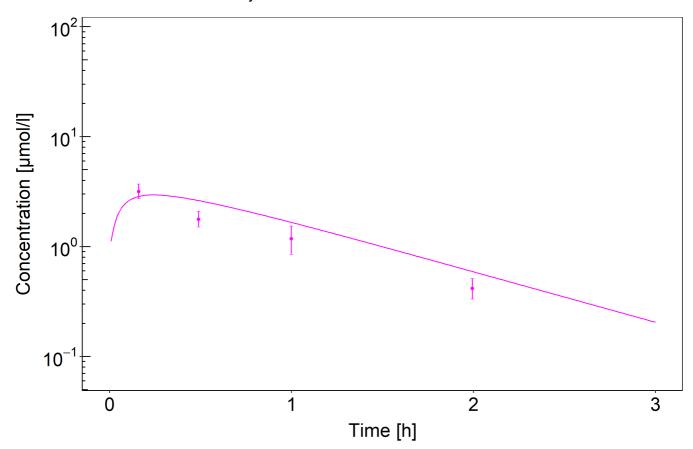


Figure 3-11: Skin

- Inulin-Small Intestine-Whole Organ-Concentration
- Inulin-Large Intestine-Whole Organ-Concentration
- Tsuji1983.Gut.20-Inulin-Gut-Tissue-Measurement

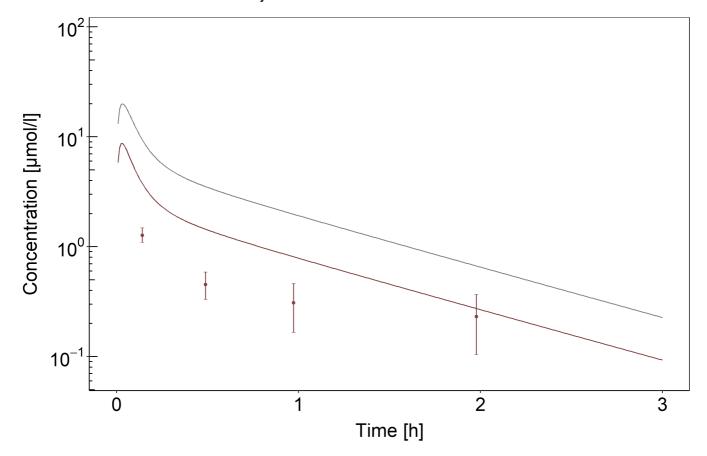


Figure 3-12: Gut

# 4 Conclusion

The herein presented PBPK model overall adequately describes the plasma pharmacokinetics of inulin in rats. Tissue concentrations tend to be overestimated by the model, the largest deviations being observed for lung, gut, and heart concentrations. Simulations are predictions without adjusting any compound specific parameter (i.e., using a literature value for the solute radius of inulin and the default physiological parameters in PK-Sim especially for the glomerular filtration rate, vascular properties and compartment volumes).

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