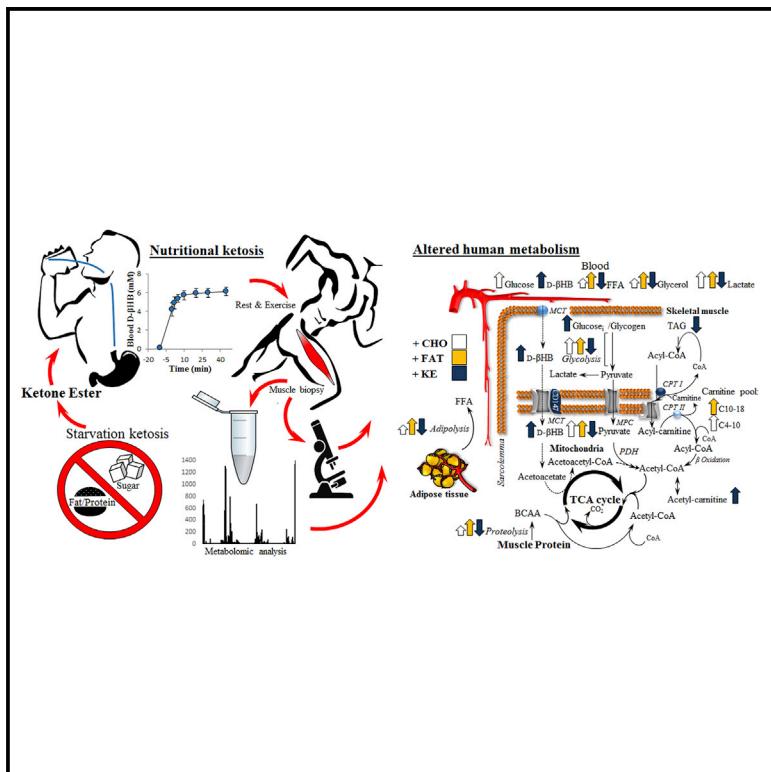


# Cell Metabolism

Clinical and Translational Report

## Nutritional Ketosis Alters Fuel Preference and Thereby Endurance Performance in Athletes

### Graphical Abstract



### Authors

Pete J. Cox, Tom Kirk,  
Tom Ashmore, ..., Richard L. Veech,  
Julian L. Griffin, Kieran Clarke

### Correspondence

petejcox456@gmail.com

### In Brief

Cox et al. show the metabolic benefit of ketone metabolism through the administration of a ketone ester-based drink to athletes during exercise. The physiological alterations achieved by acute nutritional ketosis may improve human physical performance in some athletes as indicated by initial endurance test results.

### Highlights

- Nutritional ketone bodies can promote the advantageous aspects to starvation ketosis
- Nutritional ketosis alters the hierarchy of substrate competition for respiration in exercise
- Ketosis increases metabolic flexibility during exercise, reducing glycolysis and increasing muscle fat oxidation
- Improved performance during cycling time trial suggests ketosis during exercise may be beneficial for some athletes

# Nutritional Ketosis Alters Fuel Preference and Thereby Endurance Performance in Athletes

Pete J. Cox,<sup>1,2,\*</sup> Tom Kirk,<sup>1</sup> Tom Ashmore,<sup>3</sup> Kristof Willerton,<sup>1</sup> Rhys Evans,<sup>1</sup> Alan Smith,<sup>4</sup> Andrew J. Murray,<sup>5</sup> Brianna Stubbs,<sup>1</sup> James West,<sup>3</sup> Stewart W. McLure,<sup>1</sup> M. Todd King,<sup>6</sup> Michael S. Dodd,<sup>1</sup> Cameron Holloway,<sup>1,2</sup> Stefan Neubauer,<sup>2</sup> Scott Drawer,<sup>4</sup> Richard L. Veech,<sup>6</sup> Julian L. Griffin,<sup>3</sup> and Kieran Clarke<sup>1</sup>

<sup>1</sup>Department of Physiology, Anatomy and Genetics, University of Oxford, Oxford OX1 3PT, UK

<sup>2</sup>Department of Cardiovascular Medicine, University of Oxford, Oxford OX3 9DU, UK

<sup>3</sup>Department of Biochemistry & Cambridge Systems Biology Centre, University of Cambridge & MRC Human Nutrition Research, Cambridge CB1 9NL, UK

<sup>4</sup>UK Sport, 40 Bernard Street, London WC1N 1ST, UK

<sup>5</sup>Department of Physiology, Development and Neuroscience, University of Cambridge, Cambridge CB2 3EG, UK

<sup>6</sup>Laboratory of Metabolic Control, NIAAA/NIH, Rockville, MD 20852, USA

\*Correspondence: petejcox456@gmail.com

<http://dx.doi.org/10.1016/j.cmet.2016.07.010>

## SUMMARY

Ketosis, the metabolic response to energy crisis, is a mechanism to sustain life by altering oxidative fuel selection. Often overlooked for its metabolic potential, ketosis is poorly understood outside of starvation or diabetic crisis. Thus, we studied the biochemical advantages of ketosis in humans using a ketone ester-based form of nutrition without the unwanted milieu of endogenous ketone body production by caloric or carbohydrate restriction. In five separate studies of 39 high-performance athletes, we show how this unique metabolic state improves physical endurance by altering fuel competition for oxidative respiration. Ketosis decreased muscle glycolysis and plasma lactate concentrations, while providing an alternative substrate for oxidative phosphorylation. Ketosis increased intramuscular triacylglycerol oxidation during exercise, even in the presence of normal muscle glycogen, co-ingested carbohydrate and elevated insulin. These findings may hold clues to greater human potential and a better understanding of fuel metabolism in health and disease.

## INTRODUCTION

Ketone body metabolism is a survival trait conserved in higher organisms to prolong life during an energy deficit or metabolic crisis. The advantages of ketone body metabolism during starvation are clear; providing an oxidizable carbon source to conserve precious glucose/gluconeogenic reserves while simultaneously satisfying the specific fuel demands of the brain. Ketone bodies, when present, act not only as respiratory fuels to power oxidative phosphorylation but as signals regulating the preferential oxidation and mobilization of fuel substrates (Robinson and Williamson, 1980). The conservation of CHO reserves in the form of glycogen and gluconeogenic skeletal muscle protein is a hallmark of starvation induced ketosis (Cahill, 1970), dramatically

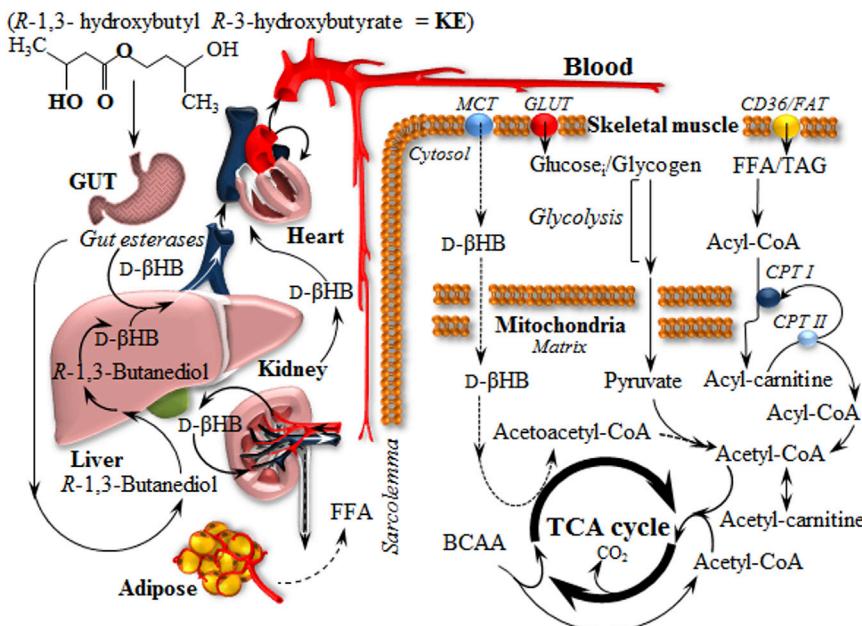
increasing survival duration (Cahill and Owen, 1968; Felig et al., 1969). Ketosis may also provide thermodynamic advantages over other carbon substrates by increasing the free energy conserved in ATP ( $\Delta G_{ATP}$ ) by the oxidation of ketones during mitochondrial oxidative phosphorylation (Sato et al., 1995). The combination of improved energetic efficiency and fuel sparing is vitally important not only during famine, but could also provide clues to new methods of sustaining human performance, or restoring dysregulated substrate metabolism.

Produced continuously under normal physiological conditions, a significant increase in the ketone bodies, D-β-hydroxybutyrate (D-βHB) and acetoacetate (AcAc), rarely manifests in concentrations above 1 mM (Robinson and Williamson 1980). However, the production of ketone bodies increases rapidly in response to calorie deprivation or energy deficit such as starvation, prolonged exercise, and as part of the clinical manifestations of diseases, such as uncontrolled diabetes (Robinson and Williamson, 1980).

As a fuel source, ketone bodies are readily oxidized by most body tissues (Robinson and Williamson, 1980), the major exception being the liver due to its lack of the enzyme succinyl-CoA:3-ketoacid CoA transferase, which permits oxidative disposal of ketones in the TCA cycle. The favorable thermodynamic characteristics of ketone body oxidation and their regulatory role controlling the preferential use and release of other substrates, such as fat (FAT) and glucose, may also have therapeutic utility for the treatment of disease (Veech, 2004; Keene, 2006).

Achieving ketosis by feeding D-βHB in an acid or salt form is not advisable due to the accompanying acid/salt load. To circumvent this, and the unwanted dietary restriction of adhering to a ketogenic diet, we generated an edible form of a ketone body by transesterifying ethyl (R)-3-hydroxybutyrate with (R)-1,3-butanediol using lipase (Figure 1; Table S1). Previously we have shown the nutritional ingestion of this (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone ester (KE) is a safe and effective way of elevating blood ketone levels (Clarke et al., 2012; Shivva et al., 2016) and provides a means of investigating human ketone metabolism independent of caloric or CHO deficit.

In some ways, the metabolic demands of prolonged exercise parallel (albeit on a more rapid scale) the metabolic conditions important to survival in starvation; it being well known that



**Figure 1. Proposed Ketone Ester Metabolism**

Ingested KE is hydrolysed in the small intestine by nonspecific gut esterases (Van Gelder et al., 2000), which cleave it into D-βHB and (R)-1,3-butanediol. Both metabolites are absorbed into the portal circulation, with butanediol undergoing first-pass metabolism in the liver to form D-βHB (Desrochers et al., 1992). D-βHB is subsequently released into the circulating blood, to be transported into muscle cytosol and mitochondria via the monocarboxylate transporters (MCTs) (Halestrap and Meredith 2004). Glucose is transported across the sarcolemma by GLUTs, and FFAs are transported by the FAT/CD36 transporters. Once inside the mitochondrial matrix, all substrates are metabolized to acetyl-CoA and oxidized in the TCA cycle.

skeletal muscle fuel selection shifts as exercise intensity rises, placing a premium on CHO reserves, resulting in an almost exclusive reliance on glycogen and blood glucose for its energy requirements (Romijn et al., 1993; van Loon et al., 2001). We reasoned that the combination of improved energetic efficiency and fuel sparing induced by ketosis is vitally important not just in famine, and that harnessing the metabolic actions of ketosis in nutritional form may provide a method of sustaining human physical performance (Cox and Clarke 2014).

Therefore, we sought to determine the mechanisms governing skeletal muscle substrate metabolism during acute nutritional ketosis in exercising humans, as well as their effects on endurance performance in this unique metabolic state.

## RESULTS

### Exercise Intensity Alters the Metabolism of Nutritional Ketosis (Study 1)

To determine whether exercise intensity altered the metabolism of diet-derived ketones, we examined the effects of steady-state exercise on the clearance of blood and urinary D-βHB in six male endurance athletes (Table S2A). An identical amount of KE was consumed by athletes at rest, and during 45 min of cycling exercise (40% and 75% of  $W_{\text{Max}}$ ) in a randomized crossover design (Figure 2A). Ingestion of a drink containing 573 mg/kg body weight of KE resulted in a rapid rise in circulating D-βHB from overnight fasted levels (0.1 mM) to ~3 mM after 10 min of rest. After the onset of exercise, D-βHB concentrations were divergent, reaching new steady-state concentrations after approximately 10 min, with high-intensity (75%  $W_{\text{Max}}$ ) exercise reducing D-βHB concentrations by  $1.05 \pm 0.2$  mM compared to workloads of 40%  $W_{\text{Max}}$ , and by  $3.1 \pm 0.4$  mM compared with resting conditions (Figure 2B). D-βHB area under curve (AUC) during 45 min of rest or exercise was significantly decreased with increasing exercise intensity (Figure 2C) and correlated closely with increasing oxygen consumption (Figure 2D). Indirect calorimetry

equations were adjusted for ketone oxidation (Frayn, 1983) (Supplemental Experimental Procedures) and used to calculate relative contributions of each substrate to total oxygen consumption during exercise at 40% and 75%  $W_{\text{Max}}$  (Figure 2E). D-βHB oxidation was estimated to account for 16%–18% of total oxygen consumption during exercise.

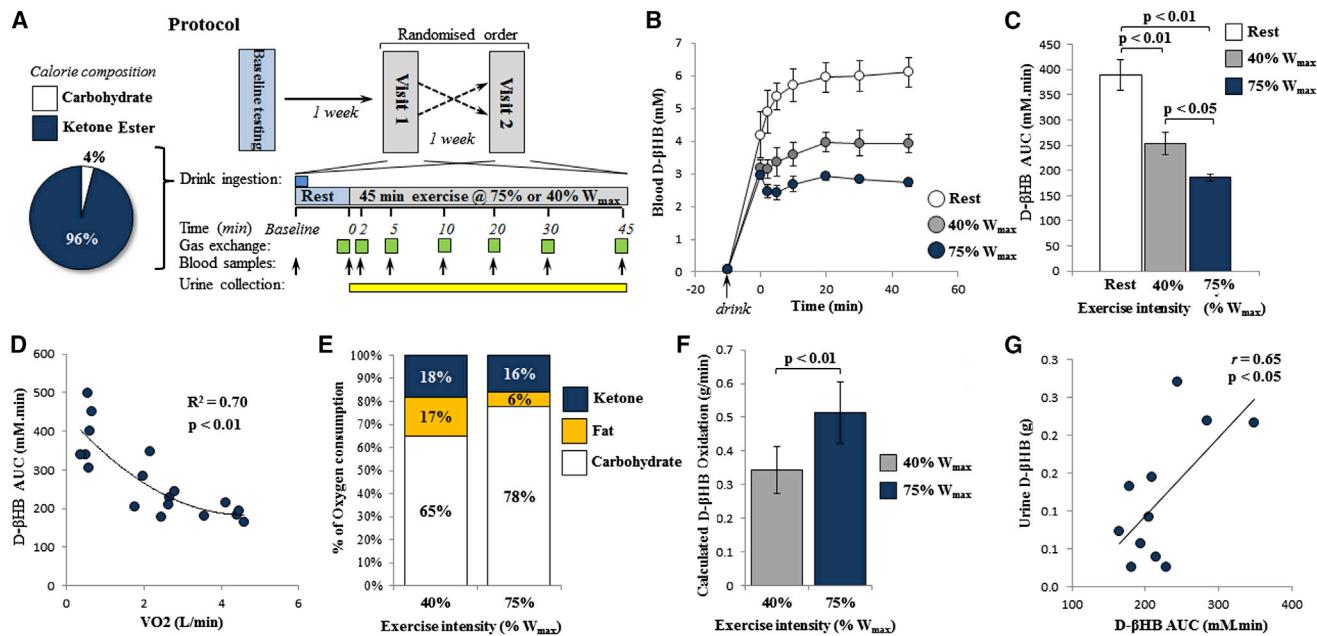
Estimated D-βHB oxidation during steady state exercise increased from 0.35 g/min at 40%  $W_{\text{Max}}$  to ~0.5 g/min at 75% intensity (Figure 2F). Urinary elimination of D-βHB during exercise was negligible, ranging from 0.05 to 0.3 g (~0.2% of total ingested KE) over the entirety of the protocol, although it correlated positively with D-βHB AUC (Figure 2G).

### The Metabolic Effects of Nutritional Substrate Alteration during Exercise (Study 2)

Each athlete ( $n = 10$ , Table S2B) completed three experimental trials consisting of 1 hr of constant load cycling at 75% of  $W_{\text{Max}}$  in a randomized, single-blind, cross-over design (Figure 3A). Isocaloric drinks contained a minimum of 96% of their calories from the one substrate (Figure 3A; Supplemental information). Subjects ingested 573 mg/kg BW of KE, isocaloric CHO, or FAT 15 min prior to the start of exercise, and 191 mg/kg BW KE 45 min into each 1 hr trial. Resting blood ketone body kinetic profiles, using an identical protocol, were determined on a separate (non-exercising) study day.

Ingestion of a drink containing 573 mg/kg body weight of KE resulted in a rapid rise in circulating D-βHB from overnight fasted levels of  $0.13 \pm 0.1$  mM to  $3.5 \pm 0.3$  mM during 10 min of rest, where they remained throughout 1 hr of exercise (Figure 3B). When no exercise was performed, plasma D-βHB concentrations increased to  $>5$  mM.

Lactate concentrations were the same at baseline for all conditions (Figure 3C). However, after the onset of exercise, blood lactate concentrations were significantly lower on KE, resulting in average exercise lactate concentrations ~2–3 mM (~50%) lower than CHO, and lower than FAT at 30 and 45 min.



**Figure 2. Effects of Exercise on D-βHB Metabolism (Study 1)**

Incremental exercise intensity increases the clearance of blood ketosis (lowering blood ketone concentrations) following the consumption of identical quantities of KE.

(A) Study protocol and interventions.

(B) D-βHB concentrations at rest and exercise at 40% and 75% W<sub>Max</sub>.

(C) D-βHB AUC during rest and exercise at 40% and 75% W<sub>Max</sub>.

(D) D-βHB AUC versus oxygen consumption.

(E) Calculated contribution of D-βHB oxidation to total O<sub>2</sub> consumption in exercise.

(F) Calculated oxidation of D-βHB (g/min) at 40% and 75% W<sub>Max</sub>.

(G) D-βHB AUC versus urinary elimination during exercise.

All data are means ± SEM.

FFA concentrations were significantly higher at baseline on FAT after 24 hr of high-FAT low-CHO meals (Figure 3D), remaining elevated throughout exercise compared with CHO or KE, reaching 0.85 mM at the end of exercise. FFA concentrations were lower than FAT at baseline before and fell after CHO or KE ingestion. Ketosis suppressed the rise in FFA seen after 25 min of exercise compared with FAT and, to a lesser extent, CHO. Exercise caused significant increases in plasma glycerol following both CHO and FAT ingestion (Figure 3E), but not after KE.

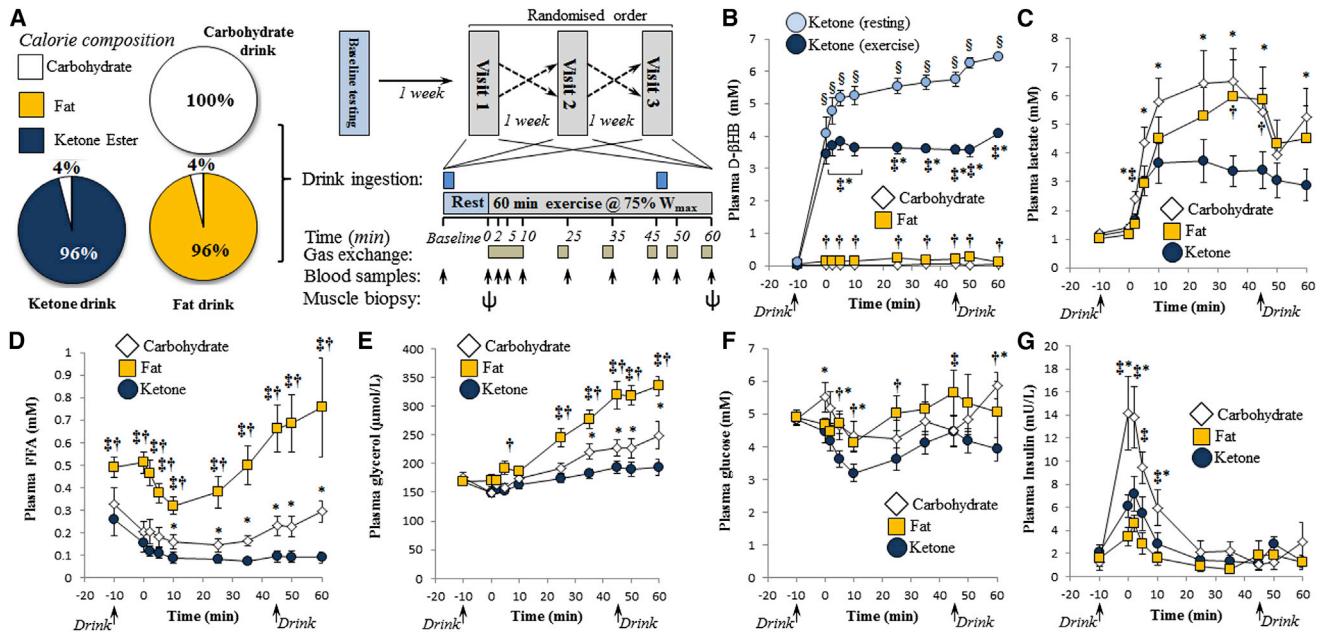
Plasma glucose concentrations were similar for all athletes at baseline but increased significantly after consuming CHO (Figure 3F). Glucose fell during the first 10 min of exercise after CHO or KE and was significantly lower after KE than either FAT or CHO intake within 5 min of exercise, remaining lower than FAT for much of the exercise protocol.

Plasma insulin concentrations were significantly elevated following CHO compared with FAT and KE (Figure 3G). Insulin concentrations peaked 10 min after the CHO drink and fell to baseline levels after 25 min of exercise. There were no significant differences in insulin after FAT and KE intake. Gas exchange (RER) was higher on CHO, with values consistently close to unity on all arms (Table S3). There were no significant differences between FAT and KE.

### Ketosis Altered Skeletal Muscle Metabolism at Rest and during Exercise

D-βHB and other metabolites were measured in skeletal muscle biopsies before and after bicycle ergometer exercise ([Supplemental Information](#)). At rest, after KE intake, intramuscular concentrations of D-βHB were ~3-fold higher than after the ingestion of CHO or FAT (Figure 4A) and remained double the concentrations following either FAT or CHO after 1 hr of exercise. Intramuscular glucose was increased pre-exercise following CHO versus FAT and KE, but was significantly greater at the end of exercise on KE (Figure 4B). Pre-exercise muscle concentrations of the glycolytic intermediates, glyceraldehyde-3-phosphate, 2&3-phosphoglycerate, and pyruvate, were significantly lower following KE consumption compared with CHO and FAT. Fructose-1,6-bisphosphate and 1,3-bisphosphoglycerate were similar at rest in all subjects (Figures 4C–4G and S1).

Following exercise, concentrations of all measured muscle glycolytic intermediates were significantly lower after KE versus CHO and FAT. The sum of glycolytic intermediates also decreased proportionately with increased intramuscular D-βHB concentration (Figure 4H). Taken together, these findings suggest that ketosis suppressed skeletal muscle glycolysis, explaining the lower blood lactate concentration described previously. Glycolytic intermediates were not different following FAT and CHO.



**Figure 3. Effects of Dietary Substrates on Plasma Metabolites during Exercise (Study 2)**

Acute nutritional substrate provision before exercise resulted in significant alterations in circulating metabolite concentrations, with KE ingestion increasing blood ketone levels, while reducing blood lactate, and circulating fats.

(A) Study protocol and interventions.

(B) D- $\beta$ Hb concentrations.

(C) Plasma lactate concentrations.

(D) Plasma FFA concentrations.

(E) Plasma glycerol concentrations.

(F) Plasma glucose concentrations.

(G) Plasma insulin concentrations.

All data are means  $\pm$  SEM.  $^{\circ}p < 0.05$  exercise versus resting,  $\dagger p < 0.05$  KE versus FAT,  $*p < 0.05$  KE versus CHO,  $\ddagger p < 0.05$  CHO versus FAT.

Drink ingestion did not change free carnitine and acyl-carnitine concentrations before exercise, but after 1 hr of high-intensity exercise, free carnitine was lower and acetyl- and short-chain C3-carnitines were higher following KE versus CHO and FAT (Figures 4I and S1), with a positive relationship between acetyl-carnitine/free carnitine ratio and D- $\beta$ Hb (Figure 4J). C8 and C10 carnitine derivatives were higher following the CHO drinks, whereas C16 and C18 longer chain acyl-carnitines were increased following FAT intake (Figures 4K, 4L, and S1). The pool of TCA intermediates remained largely unaffected by substrate provision, both at rest and following exercise, albeit expanding ~2-fold with exercise. With the exception of increased oxaloacetate concentrations following FAT, and lower malate concentrations after CHO (Figures 4M and 4N), TCA metabolites were unchanged by the type of nutritional substrate (Figure S1).

Branched-chain amino acids (BCAAs), leucine, isoleucine, and valine, are mobilized during exercise as muscle energetic and anaplerotic demands increase (van Hall et al., 1995). At rest, skeletal muscle BCAAs were significantly higher after FAT than CHO or KE (Figure S2A). During exercise, leucine + isoleucine increased, but were 50% lower following the ketone drink than CHO or FAT. The exercise-induced demand for anaplerotic substrates was reflected in the strong positive relationship between muscle leucine + isoleucine and muscle pyruvate (Figure S2B). Reducing glycolytic demand during exercise by increasing intra-

muscular D- $\beta$ Hb proportionately decreased leucine + isoleucine, and pyruvate (Figures S2C and S2D).

### The Effects of Synergistic CHO and Ketone Delivery on Human Substrate Metabolism (Study 3)

The provision of CHO with high ketone levels would never usually co-exist with an intact insulin axis and is unique to this form of ketosis. In order to determine the metabolic effects of synergistic nutritional provision of KE and CHO during exercise, each athlete ( $n = 8$ , Table S2C) completed three experimental trials consisting of 1 hr of constant load cycling at 75%  $W_{\text{Max}}$  in a randomized, single-blind, cross-over design (Figure S3A). Alterations in plasma metabolites were highly reproducible, with ingestion of KE increasing D- $\beta$ Hb levels versus CHO and B3 (Figure S3B) similar to Studies 1 and 2. To mimic the effects of ketone agonism of the nicotinic acid receptor (Taggart et al., 2005), but without the oxidizable carbon source, nicotinic acid (B3) was ingested as a control. Blood lactate concentrations were significantly decreased during exercise after KE+CHO versus CHO and B3 (Figure S3C), with no differences observed between the latter. Plasma FFA concentration fell on all arms after administration of study drinks or B3. During exercise on CHO, FFA concentration rose in identical fashion to Study 2, significantly higher than KE+CHO or B3 (Figure S3D) after ~30 min, as would be expected.

Plasma glucose remained virtually unaltered by vitamin B3 consumption; however, CHO and KE+CHO conditions resulted in transient decreases in plasma glucose on initiation of exercise, which returned to pre-exercising concentrations after 35–45 min (Figure S3E). Alterations in plasma glucose can be explained by the increases in plasma insulin following CHO-containing drinks on KE+CHO and CHO (Figure S3F). No changes in plasma insulin were observed after B3 ingestion, which remained low throughout exercise. No differences in plasma insulin concentration were observed between KE+CHO and CHO conditions. Gas exchange (RER) was similar between all three arms, with values consistently around unity (Table S4).

### Synergistic Substrate Delivery Alters Human Skeletal Muscle Metabolism

At rest, following KE+CHO ingestion, intramuscular concentrations of d-βHB were ~7-fold higher than after the ingestion of CHO or vitamin B3 (Figure 5A), and >5-fold at the end of exercise. Consumption of drinks containing CHO resulted in significant increases in intramuscular total hexose (CHO) concentration at rest. However, following 1 hr of exercise at 75%  $W_{Max}$ , hexose concentrations were significantly higher on KE+CHO versus CHO or B3 reflecting preserved intramuscular CHO stores (Figure 5B). Average plasma lactate concentration during exercise negatively correlated with end exercise intramuscular hexose (Figure 5C), while intramuscular hexose concentrations at the end of exercise correlated positively with free carnitine (Figure 5D). Intramuscular glutamine concentrations were increased on KE+CHO versus B3 and CHO (Figure 5E). No correlation was found between blood d-βHB and intramuscular d-βHB (Figure S4A), in keeping with selective trans-sarcolemmal transport by monocarboxylate transporters (MCT) (Halestrap and Meredith, 2004). A strongly positive correlation ( $r = 0.72$ ,  $p < 0.05$ ) was found between intramuscular d-βHB concentration and intramuscular hexose at the end of exercise on KE+CHO (Figure S4B).

### Alterations in Carnitine Metabolism

Free carnitine concentrations were elevated on KE+CHO versus B3 at rest and significantly greater than both CHO and B3 after exercise at 75%  $W_{Max}$  for 60 min (Figure 5F). Acetyl-carnitine/free carnitine ratio was elevated on KE+CHO versus CHO or B3 at rest, likely reflecting alterations in acetyl-CoA/CoA ratio. After exercise, however, the reverse was observed with a pronounced increase in ratio on CHO and B3, but not on KE+CHO, where a decrease occurred (Figure 5G). Commensurate with these changes, an increase in acetyl-carnitine was observed on KE+CHO at rest versus CHO and B3. However, after exercise no differences in acetyl-carnitine were observed between nutritional conditions (Figure 5H). Considerable increases in C<sub>4</sub>-OH carnitine (“keto-carnitine”) levels were observed following KE+CHO both at rest and after exercise (Figure 5I), likely reflecting buffered intra-mitochondrial ketone, and a strongly positive relationship ( $r = 0.93$ ,  $p < 0.01$ ) was observed between C<sub>4</sub>-OH-carnitine concentration and acetyl-carnitine on KE+CHO (Figure 5J).

### The Effect of Nutritional Ketosis on Intramuscular Fat and Glycogen Stores in Prolonged Exercise (Study 4)

Having demonstrated the actions of acute nutritional ketosis on skeletal muscle energy metabolism, we sought to determine

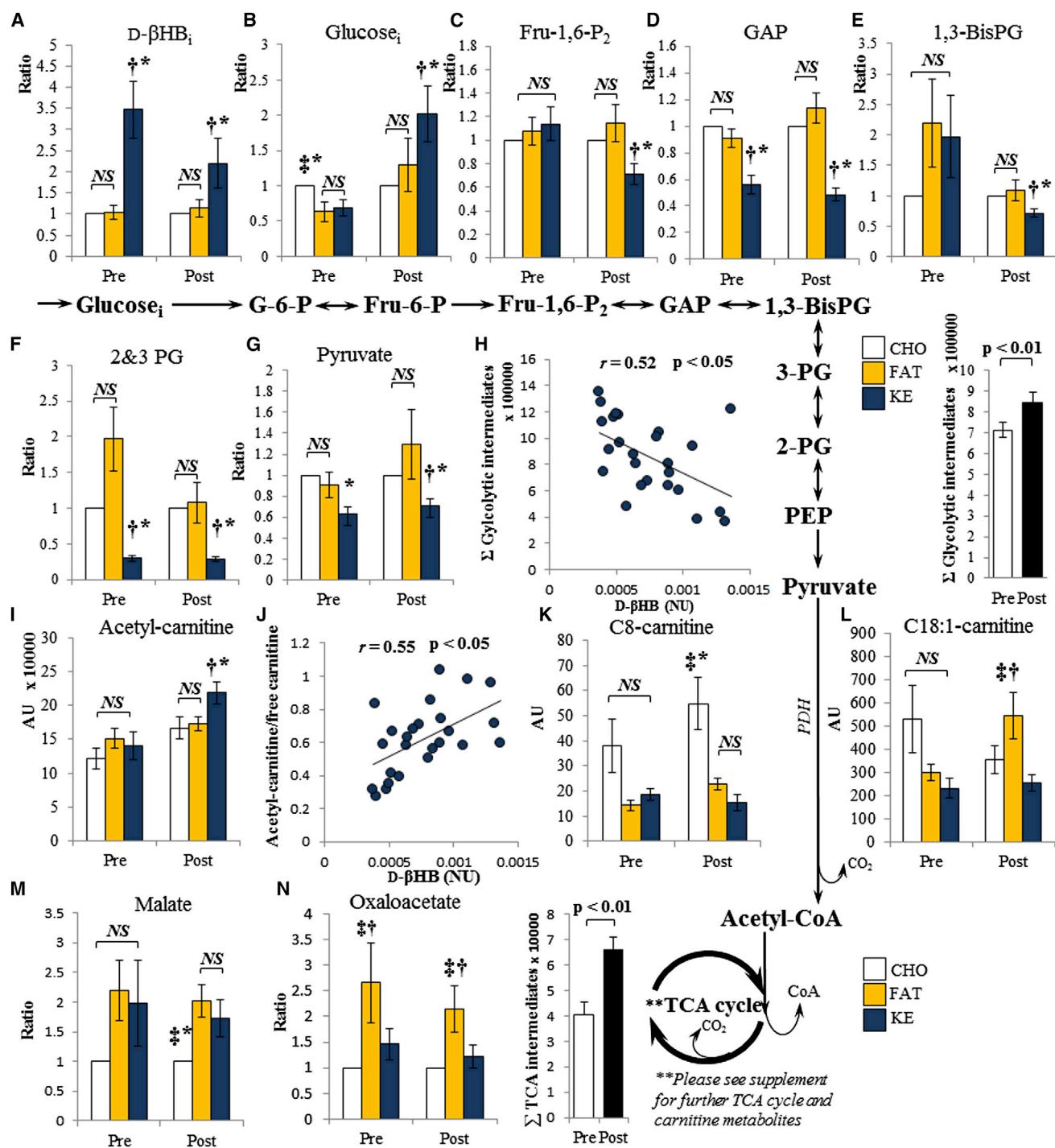
whether these changes resulted in altered intramuscular stores of FAT and glycogen during prolonged (2 hr) exercise (Figure 6A).

Alterations in plasma metabolites were highly reproducible between study participants ( $n = 7$ , Table S2D) and comparable with those in Studies 1–3. Ingestion of ketone ester increased d-βHB levels from 0.1 mM after an overnight fast to 2.2 mM ( $p < 0.01$ ) following KE+CHO ingestion (Figure 6B). Blood d-βHB concentration continued to slowly increase throughout exercise with regular ingestion of drinks, reaching  $3.2 \pm 0.2$  mM after 2 hr of exercise. Similar profiles in blood AcAc were observed (Figure S5A). d-βHB concentration remained unchanged on CHO throughout exercise ( $0.1 \pm 0.05$  mM,  $p < 0.01$  versus KE+CHO). Blood lactate concentrations were significantly decreased during exercise on KE+CHO versus CHO (Figure 6C). Plasma glucose concentrations were, on average ~1–2 mM higher on CHO following ingestion of high-CHO-containing drinks (Figure 6D). Plasma FFA concentration fell progressively on KE+CHO over the course of the study. Similar alterations were observed on CHO at rest. However, during exercise, FFA concentration was significantly higher than KE+CHO after 2 hr. (Figure 6E). No significant differences were observed in plasma insulin or cortisol (Figures S5B and S5C). In contrast to the previous studies involving shorter and higher intensity exercise, respiratory exchange ratios were consistently lower for much of the 2 hr exercise duration on KE+CHO studies versus CHO (Table S5) suggesting greater lipid oxidation.

Intramuscular triacylglycerol (IMTG) content was not significantly different between nutritional conditions at baseline. However, after 2 hr of exercise at 70%  $VO_2 Max$ , intramuscular lipids fell by 24% during KE+CHO, but only 1% on CHO ( $p < 0.01$ ) (Figure 6F). Intramuscular glycogen content was not significantly different between nutritional conditions, with all athletes demonstrating a high level (dark staining) of intramuscular glycogen before exercise (Figure 6G). As expected after 2 hr of exercise, glycogen concentrations fell on both arms with reductions in dark PAS staining and proportionate increases in moderate and light staining intensities. The degree of change was most marked on CHO, where significantly more glycogen deposits appeared moderate or light, or were no longer visible versus KE+CHO ( $p < 0.05$ ).

### The Effect of Nutritional Ketosis on Endurance Exercise Performance (Study 5)

Finally, to determine whether exercise performance could be altered by the metabolic changes arising from nutritional provision of CHO and KE, we examined the effects of steady-state exercise and time trial performance in ( $n = 8$ ) highly trained endurance athletes (Table S2E). Following an overnight fast, study participants completed two blinded bicycle exercise trials consisting of 1 hr steady-state workload at 75%  $W_{Max}$  followed by a blinded 30 min time trial (TT) for maximum distance (Figure 7A; Supplemental Information). Ingestion of a drink containing 573 mg/kg body weight of KE resulted in a rapid rise in circulating d-βHB from overnight fasted levels to ~2 mM after 20 min. Ketone concentrations remained elevated throughout subsequent exercise with a fall in concentration on initiation of exercise at 75%  $W_{Max}$  workload, after which blood concentration rose reaching a new approximate steady state after 30 min, where they remained for the rest of the protocol.



**Figure 4. Metabolic Effects of Dietary Substrates on Human Skeletal Muscle Metabolism before and after Exercise (Study 2)**

The effects of CHO, FAT, and KE ingestion on skeletal muscle metabolism pre- (Pre) and post (Post)-cycling exercise for 1 hr at 75%  $W_{Max}$ . Glycolytic and TCA cycle intermediates are expressed relative to CHO.

#### (A) D- $\beta$ HB concentrations.

**(B) Intramuscular glucose concentrations.**

(C) Fru-1,6-P2: fructose-1,6-bisphosphate concentrations.

(D) GAP: glyceraldehyde-3-phosphate concentrations

(F) 1,3-BisPG: 1,3-bisphosphoglycerate concentrations

(E) 1,3-BisPG: 1,3-bisphosphoglycerate concentrations.  
 (F) 2&3-PG: 2- and 3-phosphoglycerate concentrations

(F) 2&3-PG, 2- and 3-phospho  
(G) Pyruvate concentrations

(legend continued on next page)

In almost identical fashion to studies 3 and 4, blood lactate concentrations increased during exercise, but were ~1.5–2 mM lower on KE+CHO versus CHO (Figure 7C). Blood glucose was raised following ingestion of both drinks at rest, but fell during the first 10 min of exercise (Figure 7D). Glucose concentrations were lower on KE+CHO versus CHO during the first 25 min but were similar by 1 hr. KE+CHO significantly suppressed the exercise induced rise in FFA seen after 25 min of exercise versus CHO (Figure 7E). No significant differences in gas exchange parameters were detected during the 1 hr constant load exercise (Table S6).

Time trial performance following 1 hr of high-intensity exercise was significantly improved in KE+CHO versus CHO conditions. Athletes cycled on average  $411 \pm 162$  m further ( $p < 0.05$ ) over 30 min on KE+CHO versus CHO equating to a mean performance improvement of 2%. Pooled and individual TT performances are shown in Figures 7F and 7G. The metabolic changes arising from altered nutritional substrate provision during exercise are summarized in Figure 7H.

## DISCUSSION

In common with many disease conditions, the possible range of oxidizable carbon sources to power exercise becomes highly selective, favoring glucose as energetic demands increase (Romijn et al., 1993; van Loon et al., 2001). Here we show how a nutritional source of ketone bodies alters conventional muscle fuel metabolism and physical performance, alone and in combination with nutritional CHOs. This physiological state operates in contrast to that of endogenous ketosis, where replete glucose reserves, an intact insulin axis, and elevated ketone bodies would never usually coexist.

### Ketosis Alters the Hierarchy of Skeletal Muscle Substrate Metabolism

Substrate metabolism in the normal human body is flexible: our bodies having evolved to utilize different fuel sources depending on their availability (Randle et al., 1963). During exercise, energy expenditure increases dramatically above resting levels, with rapid turnover of mobilized fuels required to keep pace with ATP demand (Spratt and Peters, 1998). Usually, as exercise intensity increases, mitochondrial oxidation of fatty acids reaches a ceiling, shifting the burden of energy provision to CHO so that glycolytic supply of pyruvate is the major carbon source for oxidation during heavy exercise (Romijn et al., 1993; van Loon et al., 2001). Despite the stimulation of sustained exercise here, the elevated circulating ketone concentrations significantly decreased human skeletal muscle glycolytic intermediates, including pyruvate. Remarkably, this suppression of glycolysis

occurred despite physical workloads that would normally be highly glycolytic (~75%  $W_{Max}$ ).

Conversely, the same exercise overrode inhibition of glycolysis by fatty acids, in agreement with evidence suggesting that the glucose-FFA cycle (Randle et al., 1963) does not operate during intense exercise. Rather, we suggest that ketone metabolism may hold hierarchical preference over CHO and FAT metabolism, even during conditions that strongly favor CHO oxidation, such as heavy exercise. In essence, ketosis allows substrate competition for respiration during exercise that is not observed in their absence. In support of this theory, intramuscular d- $\beta$ Hb and acetyl-carnitine levels were raised ~3- to 7-fold by KE ingestion, while glycolytic intermediates were decreased without altering the pool of TCA cycle metabolites during exercise. While direct calculation of metabolic flux is not possible from single time point measurements, these data suggest that ketones and FAs were oxidized as an alternative to pyruvate, easing the reliance on glycolysis to provide acetyl-CoA to the TCA cycle. Furthermore, ketosis reduced intramuscular BCAA concentrations, supporting previous evidence that ketosis tightly regulates glycolysis (and therefore pyruvate), ultimately reducing the requirement for BCAA deamination (Thompson and Wu, 1991). Such metabolic effects may have sound survival advantages, limiting the catabolism of CHOs and skeletal muscle protein for gluconeogenesis in starvation.

Taken together, these findings support a mechanism whereby ketosis alters substrate signaling, oxidation, and energy transduction in working muscle, free of the confounding effects of elevated FFA, and reduced CHO reserves that occur with endogenous ketosis (Phinney et al., 1983a, 1983b).

### Ketosis in a Glycogen-Replete State

We have shown here how nutritional ketosis enables comparable physiological function to that of glucose, but via very different metabolic actions. Preservation of physiological function is very much in keeping with survival metabolism, where maintenance of homeostasis during conditions of altered fuel availability is vital (Cahill and Owen, 1968). Ample evidence during starvation (Hagenfeldt and Wahren, 1971; Féry and Balasse, 1983), and during high-FAT diets (Phinney et al., 1983b), suggests that ketone oxidation by skeletal muscle is minimal following the transition from fed to starvation states (Féry and Balasse, 1983)—conditions where glycogen is exhausted and FFA oxidation predominates. The observations that “starved” skeletal muscle does not utilize significant quantities of ketone bodies are in contrast to our findings in this post-absorptive (glycogen replete) state that ketone body oxidation may account for ~10%–18% of the total oxygen consumption during exercise; values in close agreement with radio-isotope studies of

(H)  $\Sigma$  Glycolytic intermediates versus d- $\beta$ Hb concentrations.

(I) Acetyl carnitine concentrations.

(J) Acetyl carnitine/free carnitine ratio versus d- $\beta$ Hb concentrations.

(K) C8-carnitine concentrations.

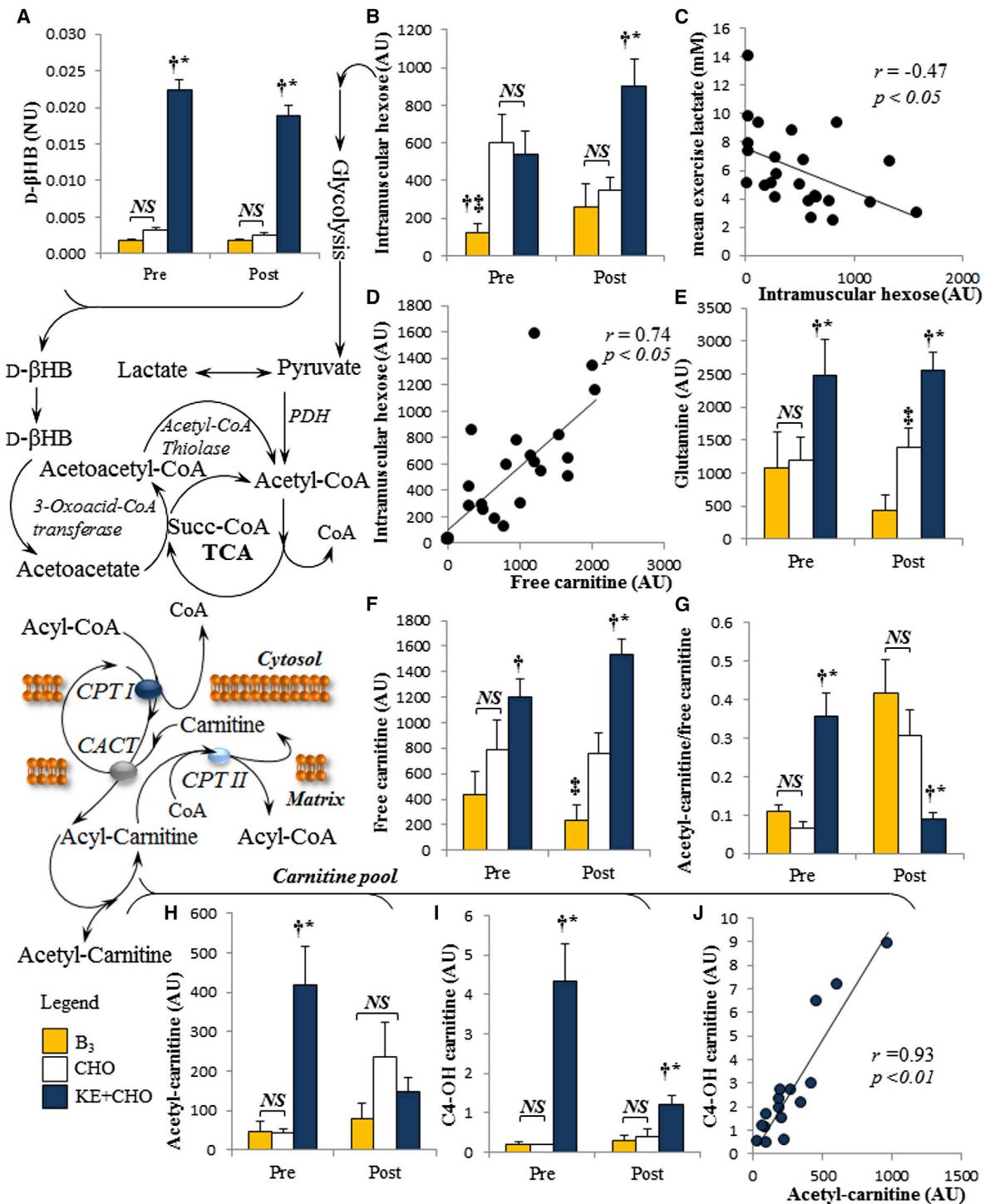
(L) C18:1 Carnitine concentrations.

(M) Malate concentrations.

(N) Oxaloacetate concentrations.

$\dagger p < 0.05$  KE versus FAT.  $*p < 0.05$  KE versus CHO,  $\ddagger p < 0.05$  FAT versus CHO. All data are means  $\pm$  SEM.

G-6-P: glucose-6-phosphate; Fru-6-P: fructose-6-phosphate; PDH: Pyruvate dehydrogenase; PEP: phosphoenolpyruvate. Arbitrary units: AU; normalized units, NU.



**Figure 5. Metabolic Effects of Dietary Substrates on Human Skeletal Muscle Metabolism before and after Exercise (Study 3)**

Nutritional substrate provision significantly altered the major pathways of muscular energy transduction, with KE ingestion increasing total CHO levels, and shifting the carnitine axis.

(A) Intramuscular D- $\beta$ Hb concentrations.

(B) Intramuscular Hexose concentrations.

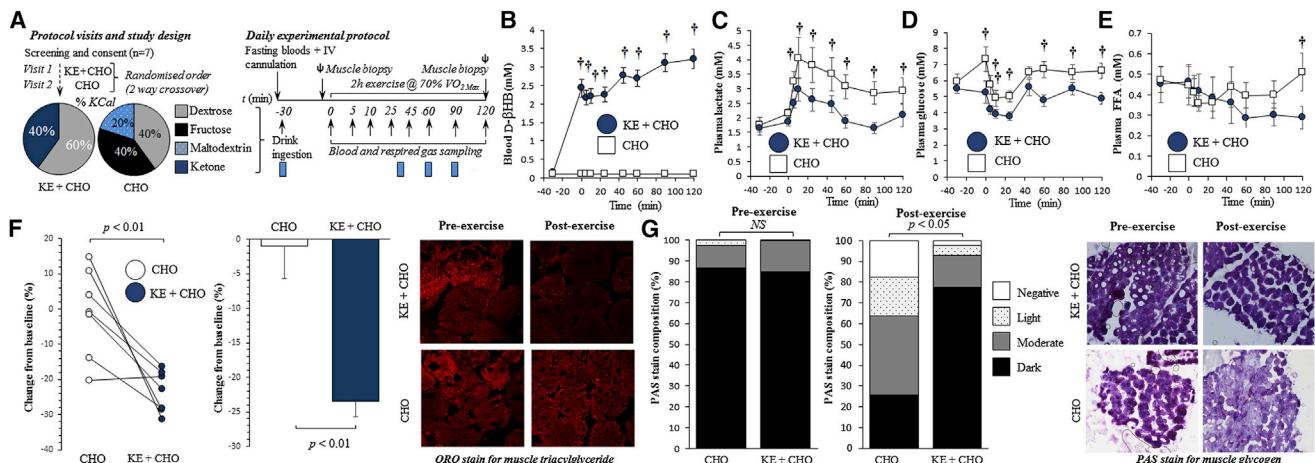
(C) End exercise intramuscular hexose versus mean plasma lactate during exercise.

(D) End exercise intramuscular hexose versus free carnitine.

(E) Intramuscular glutamine concentrations.

(F) Intramuscular free carnitine concentrations.

(legend continued on next page)



**Figure 6. The Effects of Ketosis on Intramuscular Fat and CHO Fuel Reserves during Prolonged Exercise (Study 4)**

Combined provision of nutritional ketosis with CHO ingestion increased intramuscular triacylglycerol breakdown while preserving muscle glycogen during sustained endurance exercise.

- (A) Study protocol, and interventions.
- (B) Plasma D- $\beta$ HB concentrations.
- (C) Plasma lactate concentrations.
- (D) Plasma glucose concentrations.
- (E) Plasma FFA concentrations.
- (F) Intramuscular triacylglycerol (IMTAG) levels (expressed as a % change during exercise).
- (G) Intramuscular glycogen (PAS stain intensity).

All data are means  $\pm$  SEM.  $\dagger p < 0.05$  KE+CHO versus CHO.

exercising man (Balasse et al., 1978). Furthermore, the permissive link between the supply of CHO to sustain anaplerosis, and thus TCA flux, during rat heart perfusion with acetoacetate is well known (Russell and Taegtmeyer 1991a, 1991b). As such, it seems that ketone bodies may “burn in the flame of CHOs,” whereupon the ensuing cataplerosis (and exhaustion of muscle glycogen) in ‘starved’ muscle may limit ketone body oxidation to preserve a circulating substrate for the brain.

It should also be emphasized that the coincident oxidation of ketone bodies as fuel substrates alongside FAT and glucose confounds the conventional interpretation of RER to determine fuel oxidation by indirect calorimetry (Frayn, 1983). The stoichiometry of ketone body oxidation yields RQ values of 1.0 and 0.89 for acetoacetate and  $\beta$ Hb, respectively, making isolated inferences of FAT and CHO metabolism based on RER inaccurate during ketosis (Frayn, 1983).

### Substrate Competition for Respiratory Oxidation

We have demonstrated how ketosis alters the hierarchy of fuel selection, restoring substrate competition for respiration where fatty acid oxidation cannot conventionally keep pace with TCA flux. In support of previous work in rodents (Sato et al., 1995; Kawashiyama et al., 1997; Ruderman et al., 1999), we have provided

evidence of a combined action between CHOs and ketone bodies, accentuating vital elements of the major fuel pathways known to influence muscular energy transduction. In comparison to CHO consumption alone (CHO), nutritional ketosis from KE+CHO consumption significantly increased human skeletal muscle IMTAG oxidation. Remarkably, this occurred despite highly glycolytic workloads and with increased concentrations of glucose and insulin, as both drinks contained significant quantities of CHO. Conversely, feeding isocaloric CHOs during the same exercise demonstrated no appreciable change in IMTAG levels after 2 hr of cycling. Inhibition of lipolysis via nicotinic acid receptor agonism (Taggart et al., 2005) could conceivably reduce circulating FFA availability, thus increasing IMTAG oxidation. However, this seems unlikely, as providing nicotinic acid, with no oxidizable carbon source, only increased the reliance on glycolysis for energy provision in Study 3, similar to the findings of Bergström et al. (1969). Furthermore, FFA levels following CHO and KE ingestion are suppressed, and any small ( $<0.1$  mM) differences in circulating FFA cannot account for the magnitude of change in the intramuscular lipids observed. It is tempting to suggest that a greater capacity to oxidize fatty-acid-derived carbon moieties during ketosis could power improvements in exercise capacity (as shown here) where exhaustion of glycogen

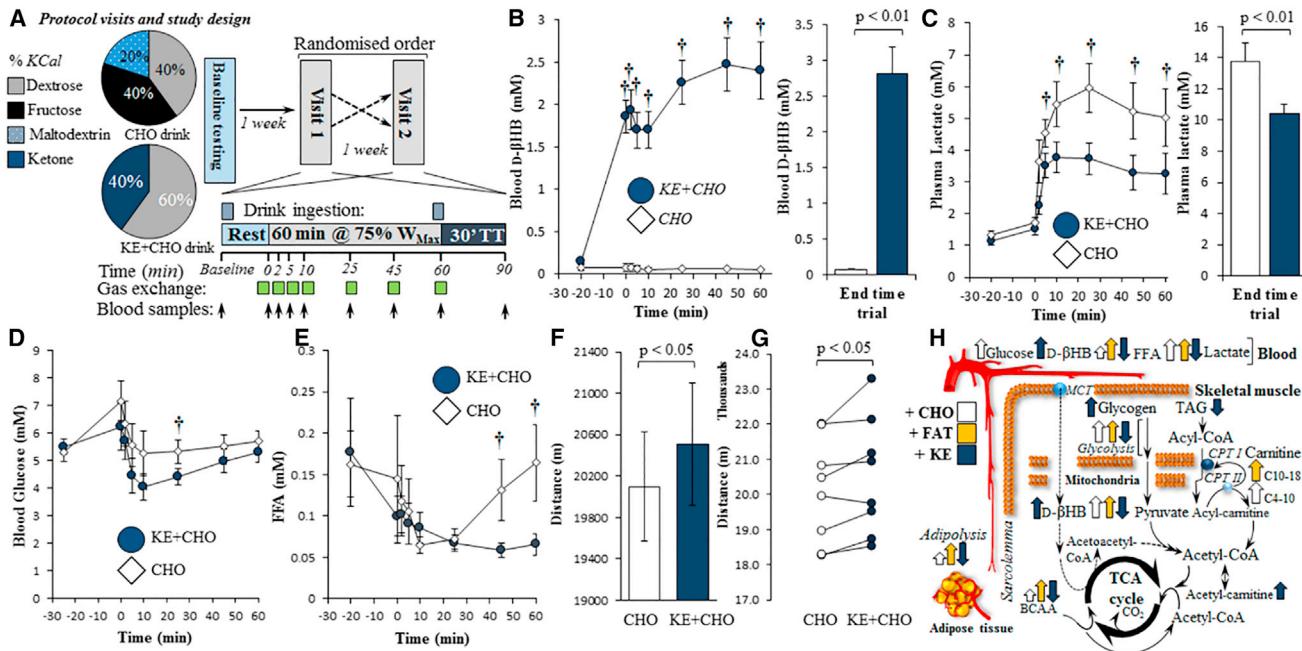
(G) Intramuscular acetyl-carnitine/free carnitine ratio.

(H) Intramuscular acetyl-carnitine concentrations.

(I) Intramuscular C<sub>4</sub>-OH carnitine concentrations.

(J) Intramuscular C<sub>4</sub>-OH carnitine versus acetyl-carnitine.

All data are means  $\pm$  SEM.  $\dagger p < 0.05$  KET versus B3,  $*p < 0.05$  KET versus CHO,  $\ddagger p < 0.05$  B3 versus CHO. TCA: tri-carboxylic acid cycle; PDH: pyruvate dehydrogenase; Succ-CoA: succinyl-CoA; CACT: carnitine-acylcarnitine translocase; CPT I and II: carnitine palmitoyltransferase; normalized units, NU; arbitrary units, AU.



**Figure 7. The Effects of Altered Fuel Metabolism on Human Physical Performance**

Combined provision of nutritional ketosis with CHO ingestion to fuel exercise altered fuel metabolism, and increased bicycle time-trial performance after 1 hr of fatiguing exercise.

(A) Study protocol, and interventions.

(B) Plasma d- $\beta$ Hb concentrations.

(C) Plasma lactate concentrations.

(D) Plasma glucose concentrations.

(E) Plasma FFA concentrations.

(F) Time trial (TT) performance (expressed as % versus CHO).

(G) Bland Altman plot of TT performance.

(H) Summary of the changes in the major fuel pathways involved in skeletal muscle energy transduction during exercise following nutritional ingestion of FAT, CHO, and KE.

All data are means  $\pm$  SEM.  $\dagger p < 0.05$  KE+CHO versus CHO.

reserves limit physical endurance. However, further work is required to confirm this. Skeletal muscle adaptions to exercise training may have influenced the changes in substrate preference observed here, and it remains to be seen whether similar changes occur in untrained individuals.

### Randle Cycle Revisited?

The promotion of intramuscular lipid oxidation during ketosis is in effect signaling an “energetic crisis” in the organism (Robinson and Williamson, 1980; Newman and Verdin, 2014), conserving glucose by forcing skeletal muscle to shift substrate oxidation to more ample fat reserves. Unlike glucose or FAT, acetyl group production from ketone bodies is independent of both PDH and CPT transporters (Halestrap and Meredith, 2004), with the increased acetyl-carnitine concentrations observed during ketosis representing an increase in acetyl-CoA production from ketones or FAT, rather than glycolysis (Sato et al., 1995; Kashiyama et al., 1997). As was proposed by Randle (Randle et al., 1963; Randle, 1998), feedback inhibition of glycolysis by a high acetyl-CoA/CoA ratio or NADH/NAD<sup>+</sup> ratio during ketosis could account for the observed decrease in glycolytic intermediates and preserved intramuscular CHO stores, as has been reported

in rodent muscle (Maizels et al., 1977) and heart (Williamson and Krebs, 1961; Sato et al., 1995). Ketones may have improved the efficiency of either the carnitine transport of acyl-CoA or  $\beta$ -oxidation, resulting in greater acyl-group oxidation. KE ingestion resulted in profound differences in carnitine species, increasing free carnitine during exercise when fed with CHO. As suggested by Wall et al. (Stephens et al., 2007; Wall et al., 2011) (who observed an improvement in physical performance with greater free carnitine availability), the matching of TCA flux with acetyl-CoA supply may have been improved, rendering oxidative ATP production more efficient. Ketosis may also augment (or mimic) the physiological actions of CHO and insulin (Kashiyama et al., 1997), increasing ketone body disposal in preference to glucose or FAT. Such metabolic actions suggest a plausible mechanism to allow the rapid clearance of ketone bodies on re-feeding following starvation, thus restoring conventional fuel metabolism. Similar findings have been shown during hyperinsulinemic clamp and ketone salt infusions in man (Keller et al., 1988). However, the exact mechanism of how ketones promoted skeletal muscle fatty acid oxidation during conditions in which glucose is conventionally preferred, and in the presence of an intact insulin axis, is unknown.

## Altered Athletic Performance

In some ways, the demands of endurance exercise parallel (albeit more rapidly) the metabolic constraints pertinent to survival in starvation, placing a premium on glucose reserves and effective oxidative respiration. We have shown here the benefit of inducing ketosis and how the combination of metabolic alterations achieved by nutritional ketosis may create a potentially advantageous physiological state, distinctly different from that of endogenous ketosis (Cahill, 1970). Athletic adaptions to harness greater circulating fuels for combustion (including ketones) are well known (Johnson and Walton, 1972; Winder et al., 1974), making athletes ideally placed to capitalize on altered substrate provision. However, it remains unclear whether similar changes to those shown here can occur in untrained individuals.

In study 5, bicycle ergometer time trial performance was ~2% greater following KE+CHO versus CHO, representing a modest increase in physical capacity in these highly trained athletes, despite significant changes in muscular metabolism. These findings suggest that the ceiling for human performance is not purely constrained by muscular energetics (Noakes, 2011). However, ketosis may not be advantageous in physiological conditions that rely almost solely on anaerobic glycolysis, or extremely high glycolytic flux for ATP production, such as sprint or short-duration exercise. Furthermore, highly glycolytic exercise may even be impaired if ketone body oxidation restricts glycolysis by negative feedback, either by an increase in NADH/NAD<sup>+</sup> or acetyl-CoA/CoA ratio. Therefore, the utility of nutritional ketosis appears more suited to metabolic conditions where dysregulated substrate selection exists, and where incremental improvements in energy transduction, or CHO preservation, may translate to significant increases in muscular endurance. Further work to determine the factors influencing human performance in ketosis, including the role of cerebral metabolism, is already underway.

## CONCLUSION

We have demonstrated the metabolic effects of elevated circulating ketone bodies as a fuel and biological signal to create a unique physiological condition. Ketosis may alter substrate competition for respiration, while improving oxidative energy transduction under certain conditions, such as endurance exercise. Consequently, nutritional ketosis may help to unlock greater human metabolic potential.

## EXPERIMENTAL PROCEDURES

### Subjects and Screening

Endurance athletes ( $n = 39$ ) participated in a series of studies investigating the effects of nutritional ketosis on resting and exercise metabolism (see Figure S6 for diagrammatic overview). Ethics approvals for all human studies were granted in accordance with Oxfordshire Regional Ethics Committee (OXREC) and NHS national research ethics service (NRES) requirements. The trials were deemed not to constitute clinical trials and were not registered as such. Written informed consent was obtained from all participants following an explanation of the risks associated with participation, and all testing conformed to the standards of ethical practice as outlined in the declaration of Helsinki. Prior to their inclusion, athletes completed a confidential medical questionnaire and received a resting ECG before exercise testing. Participants were asked not to perform strenuous exercise within 48 hr of each test, to refrain from alcohol and caffeine for 24 hr, and to consume an identical pre-testing meal the night

before every test. Water intake was provided ad libitum to each participant. In all studies comparing the effects of nutritional substrates, drink allocation was concealed and the trials were conducted in a randomized, single-blind, cross-over fashion. A double randomization method was used; the order of drink allocation was determined using a random number generator, and the order of participation was determined by participant enrolment.

### General Study Design (Study 1)

To determine whether exercise intensity altered the metabolism of diet-derived ketosis, we examined the effects of steady-state exercise on the clearance of blood and urinary D-βHB in six male endurance athletes (Table S2A). An identical amount of KE (573 mg/kg BW) was consumed by athletes at rest, and during 45 min of cycling exercise 40% and 75% of  $W_{Max}$  in a randomized crossover designed trial (Figure 2A) with 1 week between trials.

### General Study Design (Study 2)

In order to compare the metabolic alterations arising from the provision of ketones as an alternative fuel during the same physical workload, male athletes ( $n = 10$ ) (Table S2B) undertook a three-way crossover study of fixed intensity cycling at 75%  $W_{Max}$  for 1 hr. Before each test, athletes consumed a taste-matched, isocaloric flavored beverage containing ≥96% of calories from CHO (dextrose = CHO), KE (573 mg/kg BW), or FAT. Blood and respiratory gas samples were collected at regular intervals throughout exercise (Figure 3A). Muscle biopsy was performed before and after exercise on all participants.

### General Study Design (Study 3)

In order to investigate the metabolic alterations arising from the synergistic combination of fuel substrates at rest and during the same physical workload, male athletes ( $n = 8$ ) (Table S2C) undertook a three-way crossover study of fixed intensity cycling at 75%  $W_{Max}$  for 1 hr. Before each test, athletes consumed a taste-matched, isocaloric flavored beverage containing 60% of calories from CHO (dextrose), and KE (573 mg/kg BW) = KE+CHO, or a mixture of carbohydrates (CHO). On the third arm, no calories were provided in the beverage, and vitamin B3 (1,000 mg = B3) was ingested as a control to mimic the effects of ketone agonism of the nicotinic acid receptor, but without the oxidizable carbon source. Blood and respiratory gas samples were collected at regular intervals throughout exercise (Figure 3A). Muscle biopsy was performed before and after exercise on all participants.

### General Study Design (Study 4)

To investigate the effects of ketosis on intramuscular fuel reserves during prolonged exercise a further study of  $n = 7$  male athletes (Table S2D) undertook a two-way crossover study of fixed intensity bicycle ergometry at 70%  $VO_2 \text{ Max}$  for 2 hr. All participants consumed a taste-matched, isocaloric flavored beverage containing 60% of calories from CHO (dextrose) and 40% from KE = KE+CHO, or a mixture of CHOs. 50% of the total KE (573 mg/kg BW) was ingested at baseline, with the remaining 50% ingested as equal aliquots at 30 min, 1 hr, and 90 min during exercise. Blood and respiratory gas samples were collected at regular intervals throughout exercise (Figure 6A). Muscle biopsy was performed before and after exercise on all participants.

### General Study Design (Study 5)

To determine the effect of altered substrate metabolism on human physical performance, study participants ( $n = 6$  male,  $n = 2$  female) (Table S2E) completed two blinded exercise trials following an overnight fast, consisting of 1 hr steady-state workload at 75%  $W_{Max}$  followed by a blinded 30 min time trial for maximum distance. Before each test, athletes consumed a drink containing either ketone and dextrose, or CHOs alone, in randomized order (Figure 7). Athletes completed all trials on identical bike set up dimensions (SRM training systems, Germany), with no external stimuli. Blood and pulmonary gas measurements were collected during the first 1 hr fixed workload period; however, athletes were left free of distractions throughout the time trial, with a blood sample obtained immediately after the completion of the time trial. Athletes were blinded to work output, heart rate, and cadence during the 30 min time trial, and only elapsed time was visible to athletes.

### Baseline Testing and Workload Prescription

All participants undertook a stepped (25 W/3 min) incremental exercise test to exhaustion on an electronically braked bicycle ergometer (Ergoline, Germany) for the determination of  $\dot{V}CO_2$  Max (Cortex Biophysik, Germany) and  $W_{Max}$  at least 1 week prior to the start of each trial (Supplemental Information). The same ergometer was used for subsequent exercise tests.

### Substrate Drinks

In Studies 1 and 2, participants ingested drinks containing >96% of total calories from a single dietary fuel substrate as CHO, KE, or long-chain FAT (Supplemental Information). In Studies 3 and 4, participants' ingested drinks containing isocaloric quantities of CHO+KE, or 1:1:2 mixtures of dextrose, fructose, and maltodextrin. In both latter studies, a minimum of 1.2 g/min of CHO supply was ensured during exercise trials to allow comparisons according to evidence based "optimal CHO feeding strategy" (Jeukendrup and Jentjens, 2000; Jentjens et al., 2004). In Studies 3–5, drinks were prepared that contained KE as 40% of calories, with the remainder made up from CHO (dextrose). The dose response, determined previously (Clarke et al., 2012; Shiva et al., 2016), showed that 500 mg of KE/kg body weight produced blood  $\alpha$ - $\beta$ Hb concentrations of ~3 mM after 30–60 min. All drinks were taste, color, and volume matched (Supplemental Information).

### Pulmonary Gas Exchange and Blood Sampling

Respiratory gas collections (Cortex Biophysik, Germany) were obtained at identical times during exercise as blood was sampled (Supplemental Information). Blood samples (2 ml) were obtained via a venous catheter inserted percutaneously into an antecubital vein (Supplemental Information). Samples were immediately stored on ice, centrifuged (3,600 rpm for 10 min), and stored at  $-80^{\circ}\text{C}$  until further analysis. Glucose, FFA, triglycerides,  $\alpha$ - $\beta$ Hb, and lactate were assayed using a commercial automated bench-top analyzer (ABX Pentra, France). Glycerol and insulin assays were performed using ELISA kits (Mercodia, Sweden). Acetoacetate was assayed using enzymatic methods (Bergmeyer and Gawehn, 1974).

### Muscle Biopsy

Muscle tissue was collected using percutaneous needle biopsies from the lower third of the vastus lateralis muscle (Bard Monoply, USA). Samples were obtained from new incisions at rest and immediately following exercise. Tissue was frozen immediately in liquid nitrogen and stored at  $-80^{\circ}\text{C}$  until further analysis.

### Metabolite Extraction from Skeletal Muscle

Metabolites were extracted from approximately 100 mg tissue using a modified Folch method (Le Belle et al., 2002). The aqueous and organic fractions were separated and split into two identical volumes to allow multiple analyses (Supplemental Information). Histological analyses were performed using staining and confocal microscopy methods described previously (Gollnick et al., 1973; Halkjaer-Kristensen and Ingemann-Hansen, 1979; Koopman et al., 2001) (Supplemental Information).

### $^1\text{H-NMR}$ Analysis of Aqueous Metabolites

Half of the aqueous fraction (~25 mg wet weight tissue) was dried under nitrogen and resuspended in 600  $\mu\text{l}$   $\text{D}_2\text{O}$  containing 0.09% w/v NaCl (Sigma), 0.01% w/v  $\text{NaN}_3$  (Sigma), and 0.25 mM deuterated sodium-3-trimethylsilylpropionate (NaTMS-2,2,3,3-D4, Cambridge Isotope Laboratories, USA) as a chemical shift reference. Samples were analyzed on a Bruker NMR spectrometer interfaced with an 11.8 Tesla superconducting magnet at 310K using a  $^1\text{H}$ -NOESY 1D pulse sequence with 128 scans. Data were integrated using fixed integral sizes of 0.02 ppm within 1D Spec Manager (v12, Advanced Chemistry Development, Canada).

### Carnitine Analysis

Half the aqueous fractions were combined with half the organic fraction, and 200  $\mu\text{l}$  acyl-carnitine standard containing eight deuterated species was added (Cambridge Isotope Laboratories, Inc.). Samples were dried under nitrogen and butyliated with 3 M butanolic-HCl (Sigma). Samples were resuspended in 200  $\mu\text{l}$  of 4:1 acetonitrile:water containing 0.1% v/v formic acid (Sigma) and analyzed using multiple reaction monitoring on a Waters Quattro Premier

XE triple quadrupolar mass spectrometer. Chromatograms were integrated using QuanLynx v4.1 (Waters Ltd, UK).

### Statistics

Results are expressed as means  $\pm$  SEM and significance was established a priori at  $p < 0.05$ . All clinical and laboratory data were analyzed for all subjects (Supplemental Information). Statistical analysis was performed using SPSS (V21, USA). For the human trials containing paired data with three arms, repeated-measures ANOVA was performed following initial tests to ensure sphericity assumptions were not violated, and then corrected with additional post hoc Tukey corrections for multiple comparisons where appropriate (Supplemental Information). Cycling performance results were paired comparisons containing two arms, with comparisons performed using a two tailed paired t test. Correlations were tested using a two-tailed Pearson's test.

### SUPPLEMENTAL INFORMATION

Supplemental Information includes six figures, ten tables, and Supplemental Experimental Procedures and can be found with this article online at <http://dx.doi.org/10.1016/j.cmet.2016.07.010>.

### AUTHOR CONTRIBUTIONS

Study design: P.J.C., T.K., K.C. Conducting studies: P.J.C., T.K., A.S., S.W.M., B.S., S.D., C.H., S.N., R.L.V., M.T.K. Analysis: P.J.C., T.A., T.K., C.H., J.W., B.S., J.L.G., A.J.M., M.S.D., S.W.M., R.E. Manuscript preparation: P.J.C., K.C. Manuscript editing: All authors.

### CONFLICTS OF INTEREST

The intellectual property and patents covering the uses of ketone bodies and esters are owned by BTG Ltd, The University of Oxford, the NIH and TdeltaS Ltd. Should royalties ever accrue from these patents, R.L.V., K.C., A.J.M., M.T.K., and P.J.C. as named inventors may receive a share of royalties as determined by the terms of the respective institutions. K.C. is director of TdeltaS, a spin out company of the University of Oxford, to develop and commercialize products based on the ketone ester. B.S., T.K., and S.W.M. are employees of TdeltaS Ltd.

### ACKNOWLEDGMENTS

The authors thank Dr. R. Stillion, Dr. O. Faull, E. Carter, Dr. N Böehlke, and Y Green for their excellent assistance with these studies. The authors thank the Defence Advanced Research Projects Agency (DARPA) and UK Sport for funding this work.

Received: January 7, 2016

Revised: April 18, 2016

Accepted: July 17, 2016

Published: July 27, 2016

### REFERENCES

- Balasse, E.O., Fery, F., and Neef, M.A. (1978). Changes induced by exercise in rates of turnover and oxidation of ketone bodies in fasting man. *J. Appl. Physiol.* 44, 5–11.
- Bergmeyer, H.U., and Gawehn, K. (1974). *Methods of Enzymatic Analysis, Volume 3* (Academic Press).
- Bergström, J., Hultman, E., Jorfeldt, L., Pernow, B., and Wahren, J. (1969). Effect of nicotinic acid on physical working capacity and on metabolism of muscle glycogen in man. *J. Appl. Physiol.* 26, 170–176.
- Cahill, G.F., Jr. (1970). Starvation in man. *N. Engl. J. Med.* 282, 668–675.
- Cahill, G.F., Jr., and Owen, O.E. (1968). Starvation and survival. *Trans. Am. Clin. Climatol. Assoc.* 79, 13–20.
- Clarke, K., Tchabanenko, K., Pawlosky, R., Carter, E., Todd King, M., Musa-Veloso, K., Ho, M., Roberts, A., Robertson, J., Vanitallie, T.B., and Veech, R.L. (2012). Ketone bodies: physiological effects and therapeutic potential. *Adv. Nutr.* 3, 110–125.
- Gollnick, P.D., Lindstrom, C., and Simon, R. (1973). Ultrastructural changes in skeletal muscle during prolonged exercise. *Am. J. Pathol.* 73, 333–346.
- Halkjaer-Kristensen, J., and Ingemann-Hansen, T. (1979). Histological and histochemical changes in rat skeletal muscle during prolonged exercise. *Scand. J. Clin. Invest.* 39, 267–272.
- Jeukendrup, A.E., and Jentjens, R. (2000). Carbohydrate feeding strategies for endurance exercise. *Br. J. Sports Med.* 34, 365–373.
- Jentjens, R., Jeukendrup, A.E., and Wagenmakers, A. (2004). Carbohydrate feeding during exercise. *Br. J. Sports Med.* 38, 105–112.
- Shiva, A., Clarke, K., Tchabanenko, K., and Veech, R.L. (2016). Ketone bodies: metabolic effects and therapeutic potential. *Adv. Nutr.* 7, 10–20.

- R.L. (2012). Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate in healthy adult subjects. *Regul. Toxicol. Pharmacol.* 63, 401–408.
- Cox, P.J., and Clarke, K. (2014). Acute nutritional ketosis: implications for exercise performance and metabolism. *Extrem. Physiol. Med.* 3, 17.
- Desrochers, S., David, F., Garneau, M., Jetté, M., and Brunengraber, H. (1992). Metabolism of R- and S-1,3-butanediol in perfused livers from meal-fed and starved rats. *Biochem. J.* 285, 647–653.
- Felig, P., Owen, O.E., Wahren, J., and Cahill, G.F., Jr. (1969). Amino acid metabolism during prolonged starvation. *J. Clin. Invest.* 48, 584–594.
- Féry, F., and Balasse, E.O. (1983). Ketone body turnover during and after exercise in overnight-fasted and starved humans. *Am. J. Physiol.* 245, E318–E325.
- Frayn, K.N. (1983). Calculation of substrate oxidation rates in vivo from gaseous exchange. *J. Appl. Physiol.* 55, 628–634.
- Gollnick, P.D., Armstrong, R.B., Saubert, C.W., 4th, Sembrowich, W.L., Shepherd, R.E., and Saltin, B. (1973). Glycogen depletion patterns in human skeletal muscle fibers during prolonged work. *Pflugers Arch.* 344, 1–12.
- Hagenfeldt, L., and Wahren, J. (1971). Human forearm muscle metabolism during exercise. VI. Substrate utilization in prolonged fasting. *Scand. J. Clin. Lab. Invest.* 27, 299–306.
- Halestrap, A.P., and Meredith, D. (2004). The SLC16 gene family—from monocarboxylate transporters (MCTs) to aromatic amino acid transporters and beyond. *Pflugers Arch.* 447, 619–628.
- Halkjaer-Kristensen, J., and Ingemann-Hansen, T. (1979). Microphotometric determination of glycogen in single fibres of human quadriceps muscle. *Histochem. J.* 11, 629–638.
- Jentjens, R.L., Achten, J., and Jeukendrup, A.E. (2004). High oxidation rates from a mixture of glucose, sucrose and fructose ingested during prolonged exercise. *Med. Sci. Sports Exerc.* 36, S19–S20.
- Jeukendrup, A.E., and Jentjens, R. (2000). Oxidation of carbohydrate feedings during prolonged exercise: current thoughts, guidelines and directions for future research. *Sports Med.* 29, 407–424.
- Johnson, R.H., and Walton, J.L. (1972). The effect of exercise upon acetoacetate metabolism in athletes and non-athletes. *Q. J. Exp. Physiol. Cogn. Med. Sci.* 57, 73–79.
- Kashiwaya, Y., King, M.T., and Veech, R.L. (1997). Substrate signaling by insulin: a ketone bodies ratio mimics insulin action in heart. *Am. J. Cardiol.* 80 (3A), 50A–64A.
- Keene, D.L. (2006). A systematic review of the use of the ketogenic diet in childhood epilepsy. *Pediatr. Neurol.* 35, 1–5.
- Keller, U., Lustenberger, M., and Stauffacher, W. (1988). Effect of insulin on ketone body clearance studied by a ketone body “clamp” technique in normal man. *Diabetologia* 31, 24–29.
- Koopman, R., Schaart, G., and Hesselink, M.K. (2001). Optimisation of oil red O staining permits combination with immunofluorescence and automated quantification of lipids. *Histochem. Cell Biol.* 116, 63–68.
- Le Belle, J.E., Harris, N.G., Williams, S.R., and Bhakoo, K.K. (2002). A comparison of cell and tissue extraction techniques using high-resolution 1H-NMR spectroscopy. *NMR Biomed.* 15, 37–44.
- Maizels, E.Z., Ruderman, N.B., Goodman, M.N., and Lau, D. (1977). Effect of acetoacetate on glucose metabolism in the soleus and extensor digitorum longus muscles of the rat. *Biochem. J.* 162, 557–568.
- Newman, J.C., and Verdin, E. (2014). Ketone bodies as signaling metabolites. *Trends Endocrinol. Metab.* 25, 42–52.
- Noakes, T.D. (2011). Time to move beyond a brainless exercise physiology: the evidence for complex regulation of human exercise performance. *Appl. Physiol. Nutr. Metab.* 36, 23–35.
- Phinney, S.D., Bistrian, B.R., Evans, W.J., Gervino, E., and Blackburn, G.L. (1983a). The human metabolic response to chronic ketosis without caloric restriction: preservation of submaximal exercise capability with reduced carbohydrate oxidation. *Metabolism* 32, 769–776.
- Phinney, S.D., Bistrian, B.R., Wolfe, R.R., and Blackburn, G.L. (1983b). The human metabolic response to chronic ketosis without caloric restriction: physical and biochemical adaptation. *Metabolism* 32, 757–768.
- Randle, P.J. (1998). Regulatory interactions between lipids and carbohydrates: the glucose fatty acid cycle after 35 years. *Diabetes Metab. Rev.* 14, 263–283.
- Randle, P.J., Garland, P.B., Hales, C.N., and Newsholme, E.A. (1963). The glucose fatty-acid cycle. Its role in insulin sensitivity and the metabolic disturbances of diabetes mellitus. *Lancet* 1, 785–789.
- Robinson, A.M., and Williamson, D.H. (1980). Physiological roles of ketone bodies as substrates and signals in mammalian tissues. *Physiol. Rev.* 60, 143–187.
- Romijn, J.A., Coyle, E.F., Sidossis, L.S., Gastaldelli, A., Horowitz, J.F., Endert, E., and Wolfe, R.R. (1993). Regulation of endogenous fat and carbohydrate metabolism in relation to exercise intensity and duration. *Am. J. Physiol.* 265, E380–E391.
- Ruderman, N.B., Saha, A.K., Vavvas, D., and Witters, L.A. (1999). Malonyl-CoA, fuel sensing, and insulin resistance. *Am. J. Physiol.* 276, E1–E18.
- Russell, R.R., 3rd, and Taegtmeyer, H. (1991a). Changes in citric acid cycle flux and anaplerosis antedate the functional decline in isolated rat hearts utilizing acetoacetate. *J. Clin. Invest.* 87, 384–390.
- Russell, R.R., 3rd, and Taegtmeyer, H. (1991b). Pyruvate carboxylation prevents the decline in contractile function of rat hearts oxidizing acetoacetate. *Am. J. Physiol.* 261, H1756–H1762.
- Sato, K., Kashiwaya, Y., Keon, C.A., Tsuchiya, N., King, M.T., Radda, G.K., Chance, B., Clarke, K., and Veech, R.L. (1995). Insulin, ketone bodies, and mitochondrial energy transduction. *FASEB J.* 9, 651–658.
- Shivva, V., Cox, P.J., Clarke, K., Veech, R.L., Tucker, I.G., and Duffull, S.B. (2016). The Population Pharmacokinetics of D-beta-Hydroxybutyrate Following Administration of (R)-3-Hydroxybutyl (R)-3-Hydroxybutyrate. *AAPS J.* 18, 678–688.
- Spriest, L.L., and Peters, S.J. (1998). Influence of diet on the metabolic responses to exercise. *Proc. Nutr. Soc.* 57, 25–33.
- Stephens, F.B., Constantin-Teodosiu, D., and Greenhaff, P.L. (2007). New insights concerning the role of carnitine in the regulation of fuel metabolism in skeletal muscle. *J. Physiol.* 581, 431–444.
- Taggart, A.K., Kero, J., Gan, X., Cai, T.Q., Cheng, K., Ippolito, M., Ren, N., Kaplan, R., Wu, K., Wu, T.J., et al. (2005). (D)-beta-Hydroxybutyrate inhibits adipocyte lipolysis via the nicotinic acid receptor PUMA-G. *J. Biol. Chem.* 280, 26649–26652.
- Thompson, J.R., and Wu, G. (1991). The effect of ketone bodies on nitrogen metabolism in skeletal muscle. *Comp. Biochem. Physiol. B* 100, 209–216.
- Van Gelder, J., Shafiee, M., De Clercq, E., Penninckx, F., Van den Mooter, G., Kinget, R., and Augustijns, P. (2000). Species-dependent and site-specific intestinal metabolism of ester prodrugs. *Int. J. Pharm.* 205, 93–100.
- van Hall, G., van der Vusse, G.J., Söderlund, K., and Wagenmakers, A.J. (1995). Deamination of amino acids as a source for ammonia production in human skeletal muscle during prolonged exercise. *J. Physiol.* 489, 251–261.
- van Loon, L.J., Greenhaff, P.L., Constantin-Teodosiu, D., Saris, W.H., and Wagenmakers, A.J. (2001). The effects of increasing exercise intensity on muscle fuel utilisation in humans. *J. Physiol.* 536, 295–304.
- Veech, R.L. (2004). The therapeutic implications of ketone bodies: the effects of ketone bodies in pathological conditions: ketosis, ketogenic diet, redox states, insulin resistance, and mitochondrial metabolism. *Prostaglandins Leukot. Essent. Fatty Acids* 70, 309–319.
- Wall, B.T., Stephens, F.B., Constantin-Teodosiu, D., Marimuthu, K., Macdonald, I.A., and Greenhaff, P.L. (2011). Chronic oral ingestion of L-carnitine and carbohydrate increases muscle carnitine content and alters muscle fuel metabolism during exercise in humans. *J. Physiol.* 589, 963–973.
- Williamson, J.R., and Krebs, H.A. (1961). Acetoacetate as fuel of respiration in the perfused rat heart. *Biochem. J.* 80, 540–547.
- Winder, W.W., Baldwin, K.M., and Holloszy, J.O. (1974). Enzymes involved in ketone utilization in different types of muscle: adaptation to exercise. *Eur. J. Biochem.* 47, 461–467.